

Glycyrrhizic acid: extraction, screening and evaluation of anti-inflammatory property

Ácido glicirrónico: extracción, cribado y evaluación de la propiedad anti-inflamatoria

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ABSTRACT

Objective: Glycyrrhizic acid is a widely used medicinal component as an anti-inflammatory agent, anti ulcer agent, anti-allergy agent and anti-psoriatic agent. The present investigation deals with the extraction of glycyrrhizic acid from licorice roots and evaluating its *in-vitro* anti inflammatory activity.

Methods: Glycyrrhizic acid was extracted using the procedure of maceration. The extract was evaluated for its physicochemical property, biochemical tests and phytochemical properties. The *li vitro* anti-inflammatory activity was evaluated by albumin denaturation technique

Results: The results showed that the ash value and the extractive values for the extract were found to be in the limit as given by Ayurvedic Pharmacopoeia of India. Presence of flavonoids, saponins and triterpenoids were identified in the extract from phytochemical parameters. Thin layer chromatographic technique showed a retention value of 0.5 cm. The percentage inhibition showed that the extract is having some potential of healing inflammation.

Conclusion: Glycyrrhizic acid was successfully extracted from licorice roots. The evaluation parameters showed the presence of less impurity in the extract, with the potential of having anti-inflammatory property.

Keywords: Glycyrrhizic acid, Licorice roots, Anti-inflammatory activity, Albumin denaturation technique

RESUMEN

Objetivo: El ácido glicirrónico es un componente medicinal ampliamente utilizado como agente antiinflamatorio, agente antiulceroso, agente antialérgico y agente anti-psoriásico. La presente investigación trata de la extracción de ácido glicirrónico a partir de raíces de regaliz y la evaluación de su actividad antiinflamatoria *in vitro*.

Métodos: el ácido glicirrónico fue extraído usando el procedimiento de la maceración. El extracto fue evaluado por su propiedad fisicoquímica, pruebas bioquímicas y propiedades fitoquímicas. La actividad antiinflamatoria *in vitro* fue evaluada por la técnica de desnaturalización de albúmina

Resultados: los resultados demostraron que el valor de la ceniza y los valores extractivos para el extracto se encontraron en el valor límite según lo dado por la farmacopea de Ayurveda de la India. La presencia de flavonoides, de saponinas y de triterpenoides fue identificada en el extracto mediante parámetros fitoquímico. La técnica cromatográfica en capa delgada demostró un valor de retención de 0,5 centímetros. La inhibición porcentual mostró que el extracto tiene algún potencial de curación de la inflamación.

Conclusión: el ácido glicirrónico fue extraído con éxito de las raíces de regaliz. Los parámetros de evaluación mostraron la presencia de menos impureza en el extracto, con el potencial de tener propiedades antiinflamatorias.



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Palabras clave: ácido glicirrónico, raíces de regaliz, actividad anti-inflamatoria, técnica de desnaturalización de la albúmina.

INTRODUCTION

Licorice or Liquorice is the dried peeled or unpeeled roots and stolons of *Glycyrrhiza glabra* Linn, Family: *Fabaceae*. This plant has been used for its medicinal property for more than 4000 years.

Licorice has proved to be effective in the treatment of disease like gastric ulcers, arthritis, allergy, inflammation, leukemia, cancer, psoriasis, atopic dermatitis and in hepatotoxicity¹. Basically, licorice comprises of two components i.e. glycone and aglycone which are responsible for its medicinal properties. Glycone is glycyrrhizic acid (GA) and aglycone is glycyrrhetic acid, Out of these, glycone part i.e. GA is an important compound responsible for the pharmacological and biological properties of licorice. GA is a triterpenoid, diasaccharide glycoside proved to attain anti-inflammatory property, anti-diabetic, anti allergic and many more properties^{2,3,4}.

GA exhibit the mechanism of curing inflammation by inhibiting the generation of reactive oxygen species produced by neutrophils^{5,6}.

GA has molecular weight of 822.93 g/mol and molecular formula $C_{42}H_{62}O_{16}$, having a physical appearance as a yellow to orange color powder. The objective of the present investigation was to extract GA from licorice roots. The extracted GA was validated by physicochemical parameters, thin layer chromatographic analysis (High-performance liquid chromatography). *In vitro* technique was opted for evaluating its anti-inflammatory property^{7,8,9,10}.

MATERIALS AND METHODS

Materials

Licorice was purchased from the local market and authenticated from Pharmacopoeial Laboratory of Indian Medicine (PLIM); Standard glycyrrhizic acid was procured from sigma Aldrich. All the solvents used were of analytical grade.

METHODOLOGY

Physicochemical analysis

Total ash: Total ash was calculated by incinerating the fine powder of crude drug (2g) in a tarred silica crucible at the temperature of 450°C such that there is complete removal of carbon. After that the ash obtained was allowed to cooled and weighed. The percentage of total ash was calculated using the weighed value of ash and powdered crude drug.

Acid insoluble ash: The ash value was determined for detecting the undesirable or harmful or earthy matter which can be present in the crude drug. For estimating acid insoluble value, the ash obtained from the above method was poured in 25 ml of dil. HCl kept on heating mantle. The mixture was filtered using the ash filter paper, washed with hot water, ignited and weighed.

Water soluble ash: For determining the water soluble ash value the ash obtained from the total ash procedure was used and mixed with 25 ml of water. The mixture was filtered and the mixture obtained on the filter paper was collected and weighed. This weighed quantity of insoluble matter was subtracted from the weighed of ash for obtaining water soluble ash value. This weighed quantity was used for calculating the percentage of water soluble ash value.

Determination of Extractive values

Alcohol and water extractive values were determined using the same procedure except the use of alcohol to determine alcohol extractive value and the use of water in case of calculating water soluble extractive value. To determine the extractive value, powdered crude drug was macerated with the respective solution (alcohol for alcohol extractive value and water for water extractive value) in a closed flask for 24h (shaking frequently for the starting six hours). The solution was filtered and 25 ml of the filtrate was evaporated to dryness in tarred petri plate. The solution was kept at 105°C and the finally the rest amount was weighed.^{9,11}

Extraction of glycyrrhizic acid from licorice root

GA was extracted by using the method of maceration with slight modification in the method described in literature. For this purpose, drug (licorice roots) powders was macerated with the solvent mixture of acetone and dilute nitric acid for 2 h. The contents were filtered and additional 20 ml of acetone was added to the marc and warmed gently. The contents were filtered and filtrate was obtained. To this filtrate sufficient volume of dilute ammonia solution was added till precipitation of ammonium glycyrrhizinate is completed. The precipitate was collected and washed with 5 ml of acetone, dried and collected¹².

Phytochemical screening of GA

Phytochemical screening of GA was performed for the identification of phytoconstituents present.

Test for saponins (foam test)

The extract was dissolved with 20 ml of distilled water and stirred for 15 minutes. The formation of 1cm layer of foam for a period of time showed the presence of saponins.

Tests for flavonoids

With sodium hydroxide: Extract was mixed with 1 ml of sodium hydroxide solution. Blue to violet color indicates the presence of anthocyanins, yellow to orange color shows the presence of flavonones and yellow color indicates flavones.

With concentrated sulphuric acid: Extract was mixed with concentrated sulphuric acid. Yellow orange color indicates the presence of anthocyanins, orange to red color indicates the presence of flavones.

Shinoda test: For performing shinoda's test, extract was dissolved in ethanol, to which magnesium turnings were added. To this mixture, Conc. Hydrochloric acid was added. Turning of magenta to purple color indicates the presence of flavonoids.

Test for terpenoids

Lieberman's test: Acetic acid was added to the extract kept on the hot plate. To this mixture, concentrated sulphuric acid was added. Presence of pink color indicates the presence of triterpenoids in the extract.

Trichloroacetic acid test: trichloroacetic acid was added to the extract. Formation of yellow color indicates the presence of terpenoids.

Fehling's test

On the water bath, Extract was kept. To which Fehling solution A and B were mixed. Brick red precipitate showed the presence of reducing sugars.^{9,13,14}

Loss on drying

Loss on drying was calculating by the mentioned procedure. Weighed quantity of extract was poured onto a weighed petri plate. The petri plate was kept in oven and weighed at different time interval at 105°C, till two consecutive weighing didn't differ by more than 0.25mg which indicates the final loss of moisture present in the drug. Percentage loss on drying was calculated using the below mention formula^{9,15,16}.

$$\text{LOD}(\%) = \frac{\text{weight of porcelain dish with drug at time 0} - \text{weight of porcelain dish after 6 h}}{\text{weight of porcelain dish at time 0} - \text{weight of empty porcelain dish}}$$

pH determination

The extract was dissolve in 10 ml of distilled water for evaluating the pH. The pH was determined using digital pH meter. The pH was measured in triplicate.

Melting point range

The melting point was determined by capillary technique. The drug was filled in the capillary tube, sealed to the one end. The capillary was introduced into digital melting point apparatus. The temperature, at which the drug melts, signifies the melting point of drug.

UV spectral analysis

A UV spectrum of the extracted GA was obtained by scanning the extract solution in the range of 200-800nm using UV spectrophotometer. For this, stock solution of 100µg/ml of GA solution was prepared¹⁷.

Analysis using Thin Layer Chromatography (TLC)

For TLC test solution, standard solution and developing solvent system were prepared. For preparing test solution, alcohol and water (7:3) was mixed to licorice extract. The solution was heated using water bath for 5 minutes, cooled and filtered. Standard solution was prepared by dissolving 5 gm of standard glycyrrhizic acid in 1ml of a mixture of alcohol and water (7:3). Developing solvent system contains the mixture of butyl alcohol, water and glacial acetic acid (7:2:1). TLC plates were prepared using silica gel solution and the retention value (Rf.) was calculated. The plates were kept in developing solvent system and examined under UV light at 254nm¹⁸.

In vitro anti inflammatory activity

The *In Vitro* anti inflammatory activity of GA was evaluated by albumin denaturation technique as given by Mizushima and kobayashi with slight modification. Extract was mixed with 1% aqueous solution of fetal bovine albumin. pH of the mixture was adjusted using 0.1NHCl. The solution was kept in a incubator at 37°C for 20 min^{19,20,21}. Afterwards, denaturation was induced by keeping the reaction mixture at 60±1°C in water bath for 10 min. The mixture was cooled and the turbidity was measured using UV spectrophotometer. Percentage inhibition was calculated using the following equation. For this, acetyl salicylic acid was considered as a standard and the solution containing no drug was considered as control^{22,23,24}.

% Inhibition = (Absorbance of control – Absorbance of sample) × 100 / Absorbance of control

RESULTS

Physicochemical analysis

Physicochemical parameters like ash value, acid insoluble, water soluble and alcohol soluble value were carried out.

The results showed that all the values obtained were in the limit as given in Ayurvedic Pharmacopoeia of India (API). The total ash value was found to be 3.75%. The values of acid insoluble, water soluble extractive and alcohol soluble extractive value were found to be 1.93%, 3.51% and 2.18% respectively as given in Table 1, indicating the presence of water soluble components than alcohol soluble components.

Table 1. Compiled results for total ash, acid insoluble ash, water soluble extractives and alcohol soluble extractive values as compared to standard values

S. No.	TESTS	OBSERVATIONS (%)	STANDARD % (API)
1	Total ash	3.75	Not more than 10
2	Acid-insoluble ash	1.93	Not more than 2.5
3	Water soluble extractives	3.51	Not less than 20
4	Alcohol soluble extractives	2.18	Not less than 10

Phytochemical screening of GA

The tests performed for Phytochemical screening showed the presence of flavonoids, saponins and triterpenoids in the extract (GA) as shown in Table 2.

Table 2 Observation found via phytochemical tests

S. No.	TESTS	OBSERVATIONS	RESULT	
1	Fehling test	Brick red precipitate	+	
2	Test for Flavonoids	With sodium hydroxide	Yellow color	+
		With sulphuric acid	Orange to red color	
		Shinoda test	Magenta to purple color	
3	Test for Saponins Foam Test	Persistence of foam for a period of time	+	
4	Test for terpinoids	Lieberman's test	Pink color	+
		Trichloroacetic acid test	yello Yellow color w color	
NOTE: '+' Indicates the presence of the compound.				

The extract was evaluated for the parameters like pH, melting point and loss on drying. The loss on drying after 6 hours was found to be 6.5%. According to the monograph, the extract should not lose more than 12.0% of its weight. The value for loss on drying has been summarized in Table 3. The pH of the extract was evaluated in triplicate and the average was found to be 5.53 ± 0.05 as depicted in Table no.4. The melting point was found to be -240°C , which is comparable to the reported melting point of GA i.e. -220°C .

Table 3. Loss on drying observations at different time intervals

Time (h)	Weight (g)
0	64.75
1	64.38
2	64.21
3	64.15
4	64.08
5	64.10
6	64.10

Table 4 pH values of extract

S. No.	pH	Mean \pm S.D
1	5.5	5.53 ± 0.05
2	5.6	
3	5.5	

UV spectral analysis

Spectral analysis was evaluated using UV spectrophotometer. The highest peak was attained at the wavelength of 230nm. As reported, the standard wavelength of GA is 254nm. The graph obtained via UV spectrophotometer has been depicted in Figure 1.

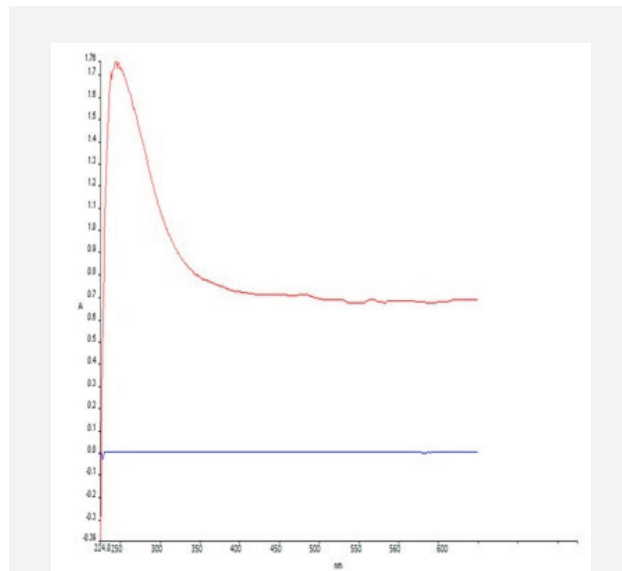


Figure 1. UV spectra of Licorice root Extract showing peak at 230nm.

Thin layer chromatographic analysis

R_f value was obtained through TLC analysis by dividing the distance travelled by solute from the distance travelled by solvent. The R_f value was found to be 0.5 cm. The TLC plate showed purple color when seen through UV fluorescent light.

$R_f = \text{Distance travelled by solute} / \text{Distance travelled by solvent}$

$$= 3.5 / 7.0$$

$$= 0.5 \text{ cm}$$

In vitro Anti-inflammatory activity

In vitro anti inflammatory studies were carried to relate anti inflammatory activity of GA with standard drug, salicylic acid (most commonly used as anti inflammatory agent).

Percentage inhibition was calculated using the absorbance and it was found that the extract and standard showed 27.11 % inhibition and 49.15 % inhibition respectively. The results have been compiled in Table 5.

Table 5. Data for the percentage inhibition of albumin denaturation

Sample	Absorbance	Mean absorbance \pm SD	Inhibition (%)
Control	0.059 0.061 0.057	0.059 \pm 0.002	-
Extract	0.043 0.047 0.041	0.043 \pm 0.003	27.11
Standard	0.027 0.031 0.033	0.030 \pm 0.003	49.15

DISCUSSION

Results obtained from physicochemical analysis (as shown in Table 1) showed that there was significant difference found between total ash and acid insoluble ash, which indicates that the ash contains a considerable amount of inorganic radicals like calcium oxalate which are acid soluble. Phytochemical screening revealed the presence of flavonoids, saponins and triterpenoids in the extract. The *in vitro* anti-inflammatory evaluation showed the inhibition of 27.11% from which it can be concluded that the extract has less inflammatory property than standard, but is able to show a significant percentage of inflammatory property.

CONCLUSION

Glycyrrhizic acid was extracted from licorice roots and evaluated for physicochemical, phytochemical analysis, extractive value. The calculated R_f value of the extract was found to be 0.5 cm. From the results obtained from evaluation parameters it can be concluded that the extracted glycyrrhizic acid contains less and permissible amount of impurities. Glycyrrhizic acid was also evaluated for its anti-inflammatory activity using *in vitro* analysis, which proved its ability on counteracting on inflammatory activity.

REFERENCES

- Deng S, May BH, Zhang AL, Lu C, Xue CCL. Topical herbal medicine combined with pharmacotherapy for psoriasis: a systematic review and meta-analysis. *Archives of Dermatological Research*. 2013; 305(3):179-89.
- Li J, Cao H, Liu P, Cheng G, Sun M. Glycyrrhizic Acid in the Treatment of Liver Diseases: Literature Review. *BioMed Research International*. 2014:872139.
- Darvishi B, Manoochehri S, Kamalinia G, *et al.*. Preparation and Antibacterial Activity Evaluation of 18- β -glycyrrhetic Acid Loaded PLGA Nanoparticles. *IJPR*. 2015; 14(2):373-383.
- Ram HNA, Lachake P, Kaushik U, Shreedhara CS. Formulation and evaluation of floating tablets of liquorice extract. *Pharmacognosy Research*. 2010; 2(5):304-308.
- Račková L, Jančinová V, Petříková M, Drábíková K, Nosál R, Štefek M, *et al.*. Mechanism of anti-inflammatory action of liquorice extract and glycyrrhizin. *Natural Product Research*. 2007; 21(14):1234-41.
- Yang R, Yuan B-C, Ma Y-S, Zhou S, Liu Y. The anti-inflammatory activity of licorice, a widely used Chinese herb. *Pharmaceutical Biology*. 2016; 55(1):5-18.
- Zhao H, Zhao M, Wang Y, Li F, Zhang Z. Glycyrrhizic Acid Attenuates Sepsis-Induced Acute Kidney Injury by Inhibiting

- NF- κ B Signaling Pathway. Evidence-based Complementary and Alternative Medicine : eCAM. 2016;8219287.
8. Wang L, Yang R, Yuan B, Liu Y, Liu C. The antiviral and antimicrobial activities of licorice, a widely-used Chinese herb. *Acta Pharmaceutica Sinica B*. 2015; 5(4):310-315.
 9. Bysani S, Babu PS, Karthikeyan R. Proximate, powder microscopic, liquid chromatographic and in-vitro anti-inflammatory activity of marketed athimadhuram churnas. *Journal of Medicinal Plants Studies*. 2017; 5(3): 373-383.
 10. Li J, Cao H, Liu P, Cheng G, Sun M. Glycyrrhizic Acid in the Treatment of Liver Diseases: Literature Review. *BioMed Research International*. 2014;872139.
 11. The Ayurvedic Pharmacopoeia of India. Part I, vol I, 168-169.
 12. Tian M, Yan H, Row KH. Extraction of Glycyrrhizic Acid and Glabridin from Licorice. *International Journal of Molecular Sciences*. 2008; 9(4):571-577.
 13. Hemraj Vashist, Diksha Sharma. Pharmacognostical Aspects of Glycyrrhiza Glabra. *Asian J Pharm Clin Res*. 2013; 6(4): 55-59.
 14. D'Sousa' Costa CO, Ribeiro PR, Loureiro MB, Simões RC, de Castro RD, Fernandez LG. Phytochemical screening, antioxidant and antibacterial activities of extracts prepared from different tissues of *Schinus terebinthifolius* Raddi that occurs in the coast of Bahia, Brazil. *Pharmacognosy Magazine*. 2015; 11(43):607-614.
 15. . Purkait K, Das S, Maity T, Chakraborty P. Preliminary Phytochemical screening of *Abelmoschus esculentus* Linn. *Journal of Biomedical and Pharmaceutical Research*. 2017; 21-34.
 16. U.S. Pharmacopoeia-National Formulary (USP 39 NF 34). Vol I, 1445-1468.
 17. Raja MS, Khan I, Perumal P, Srikakolapu SR, Gotteti SD, Quantitative analysis of glycyrrhizic acid in crude drug and its herbal formulation by UV spectroscopy, *Archives of Applied Science and Research*, 2(2). 2010; 184-189.
 18. http://www.drugfuture.com/pharmacopoeia/usp32/pub/data/v32270/usp32nf27s0_m45050.html.
 19. Mizushima Y, Kobayashi M. Interaction of anti-inflammatory drugs with serum proteins, especially with some biologically active proteins. *Journal of Pharmacy and Pharmacology*. 1968; 20(3):169-73.
 20. Hossain MS, Chowdhury MEH, Das S, Chowdhury IU. In-vitro thrombolytic and anti-inflammatory activity of *Swerthia chirata* ethanolic extract. *J. Pharmacog. And Phytochem*. 2012; 1(4):99-104.
 21. Sakat SS, Archana RJ, Gambhire MN. In vitro antioxidant and inflammatory activity of methanol extract of *Oxalis corniculata* Linn. *Int J Pharmacy and Pharm Sci* 2010; 2 Suppl 1: 146-155.
 22. Leelaprakash G, Mohan Dass S. In-vitro anti-inflammatory activity of methanol extract of *Enicostemma axillare*. *Int J Drug Dev Res*. 2011; 3(3):189-196.
 23. Pemiah, Brindha; Reshma, K.P, Arun. In vitro anti-inflammatory, antioxidant and nephroprotective studies on leaves of *Aegle marmelos* and *Ocimum sanctum*. *Asian Journal of Pharmaceutical and Clinical Research*. 2014; 121-129.
 24. Chowdhury A, Azam S, Jainul MA, Faruq KO, Islam A. Antibacterial Activities and In Vitro Anti-Inflammatory (Membrane Stability) Properties of Methanolic Extracts of *Gardenia coronaria* Leaves. *International Journal of Microbiology*. 2014; 1