



UNIVERSITI PUTRA MALAYSIA

***PERFORMANCE OF CROSSES AND DEVELOPMENT OF DOUBLED
HAPLOID LINES IN WHEAT (*Triticum aestivum* L.) FOR SALT
TOLERANCE***

ABBAS LATEEF ABDULRAHMAN

FP 2018 20



**PERFORMANCE OF CROSSES AND DEVELOPMENT OF DOUBLED
HAPLOID LINES IN WHEAT (*Triticum aestivum* L.) FOR SALT
TOLERANCE**

By

ABBAS LATEEF ABDULRAHMAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

February 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

PERFORMANCE OF CROSSES AND DEVELOPMENT OF DOUBLED HAPLOID LINES IN WHEAT (*Triticum aestivum* L.) FOR SALT TOLERANCE

By

ABBAS LATEEF ABDULRAHMAN

February 2018

Chairman : Professor Siti Nor Akmar Abdullah, PhD
Faculty : Agriculture

Wheat (*Triticum aestivum* L.) is an important staple food crop for the world population. Despite its importance, the crop productivity is often affected by salinity stress. This study aimed to develop sodium chloride (NaCl) tolerant doubled haploid wheat using anther culture technique. The effect of different concentrations of NaCl (0, 50, 100, 150 and 250 mM) on seed germination and selected seedling growth traits of eight wheat genotypes was evaluated. The G1, G2, G6, G8, and G10 were classified as salt tolerant while Abo- Graib, Forat and, Dijla genotypes were salt susceptible. Increasing concentration of NaCl solution led to the gradual reduction in the seed germination percentage, and seedling growth traits. The three salt susceptible and one high salt tolerant genotypes (G8) were selected for crossing to obtain a total of six crosses. The agronomic performance of the six F₁ crosses was investigated under 0, 50, 100, 150 mM of NaCl. For all, grain yield exhibited positive correlation with spikes number/plant, grains number per spike and weight of 1000 grain. Abo-Graib×G8, Forat×G8, and Dijla×G8 showed superiority in grain yield (7.3 g/plant, 7.4 g/plant and 6 g/plant, respectively) under high concentration of NaCl (100 mM) related to the spikes number/plant, grains number per spike and the 1000 grain weight. Investigation on callus induction from anther culture of the F₁ progenies was performed. All F₁ produced a high percentage of callus (47.9%) when subjected to 33°C + chemical pre-treatment for 48 hours followed by culturing on CHB₃ induction medium containing Myo-inositol (300 mg/L), L-glutamine (993.5 mg/L) and 2 mg/L of both 2, 4-D and kinetin. The F₁ produced good percentage (45.6%) of green plants and the lowest percentage (6.8%) of albino plants after growing on R9 regeneration medium containing 0.076 mg/L CuSO₄·5H₂O and 90000 mg/L maltose following the pre-treatment of the anther with heat + chemical and culturing on CHB₃ callus induction medium. Abo-Graib×G8 displayed the highest ability for callus induction and green plants production (82.3% and 81.3%, respectively) and the lowest

percentage of albino plants (2%). The 883 regenerated plants from the F₁, 646 were green and 237 were albino plants. Among the green plants, 281 grew to maturity. Thirty-nine lines were fertile and their seeds were grown for developing salt tolerant doubled haploid lines. Haploid plant's chromosomes were doubled under colchicine treatment (0.2%) with dimethyl sulfoxide (DMSO) (2%) and 2-3 drops of Tween-20 for 4 hours at room temperature. The doubling of chromosomes was validated using a fluorescent microscope. Thirty-nine doubled haploid lines were evaluated under 0, 50, 100 and 150 mM of NaCl in the greenhouse. The doubled haploid lines DH6 obtained from (Abo-Graib×G8), DH8 (Forat×G8) and DH6 (Dijla×G8) showed a high performance due to their superiority in leaves K⁺ content, K⁺/Na⁺ ratio, the spikes number, grains number, and the decrease in leaves Na⁺ content. Taken together, it can be concluded that through anther culture, doubled haploid lines that displayed high yielding trait were developed for future exploitation.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PRESTASI KACUKAN DAN PEMBANGUNAN TITISAN HAPLOID
GANDA DUA RINTANG NATRIUM KLORIDA DALAM GANDUM
(*Triticum aestivum* L.)**

Oleh

ABBAS LATEEF ABDULRAHMAN

Februari 2018

Pengerusi : Profesor Siti Nor Akmar Abdullah, PhD
Fakulti : Pertanian

Gandum (*Triticum aestivum* L.) merupakan makanan ruji utama kepada populasi dunia. Walaupun penting, produktiviti tanaman sering dipengaruhi oleh tekanan kemasinan. Penyelidikan ini telah dilakukan untuk menghasilkan jenis gandum yang toleran pada natrium klorida melalui teknik kultur anter. Kesan perbezaan kepekatan NaCl (0, 50, 100, 150, 200, dan 250 mM) pada percambahan biji benih dan pada sebahagian ciri-ciri pertumbuhan anak benih dari lapan genotip gandum telah dikaji. Genotip G1, G2, G6, G10 telah dikelasifikasikan sebagai toleran garam tahap sederhana manakala Abo- Graib, Forat, dan Dijla sensitif garam. Peningkatan kepekatan larutan NaCl membawa kepada pengurangan secara beransur-ansur pengambilan air, peratus percambahan biji benih dan ciri-ciri pertumbuhan anak benih. Tiga varieti gandum sensitif garam dan satu kultivar toleran garam (G8) telah dipilih untuk kacukan untuk mendapatkan sejumlah 6 kacukan. Pencapaian agronomik enam kacukan telah dikaji pada tiga kepekatan (0, 50, 100, 150 mM) NaCl. Kesemuanya menunjukkan korelasi positif dengan bilangan/tumbuhan, bilangan bijian setiap spike dan berat 1000 bijian. Abo-Graib×G8, Forat×G8, dan Dijla×G8 menunjukkan keunggulan di dalam hasil bijian (7.3 g/tumbuhan, 7.4 g/tumbuhan dan 6 g/tumbuhan) di bawah kepekatan tinggi NaCl (100 mM) berkaitan dengan bilangan spike/tumbuhan, bilangan bijian pada setiap spike dan berat 1000 bijian. Kajian mengenai induksi kalus dari kultur anter progeni F₁ telah dijalankan. Semua kacukan F₁ yang dikaji telah menghasilkan peratusan kalus yang tinggi (47.9%) apabila dikenakan suhu 33°C + pra-rawatan kimia selama 48 jam diikuti dengan pengkulturan pada media induksi kalus CHB₃ yang mengandungi Myo-inositol (300 mg/L), L - glutamine (993,5 mg/L) dan 2 mg/L kedua-dua 2, 4-D dan kinetin. F₁ telah menghasilkan peratusan tumbuhan hijau yang baik (45.6%) dan peratus terendah (6.8%) tumbuhan albino selepas tumbuh di media regenerasi R9 yang mengandungi 0.076 mg/L CuSO₄·5H₂O dan 90000 mg/L maltosa diikuti dengan pra-rawatan anter

dengan haba + kimia dan pengkulturan pada media induksi kalus CHB₃. Abo-Graib×G8 mempamerkan keupayaan tertinggi bagi induksi kalus dan penghasilan tumbuhan hijau (masing masing 82.3% dan 81.3%) serta peratusan terendah tumbuhan albino (2.0%). Daripada 883 tumbuhan yang tergenerasi dari F₁, 646 adalah tumbuhan hijau dan 237 adalah tumbuhan albino. Di antara tumbuhan hijau, 281 tumbuhan sahaja yang membesar hingga matang. Tiga puluh sembilan tumbuhan didapati subur dan menghasilkan anak benih titisan haploid ganda dua tahan garam. Kromosom tumbuhan haploid ini telah meningkat dua kali ganda di bawah rawatan colchicine (0.2%), DMSO (2%) dan 2-3 titis tween-20 selama 4 jam pada suhu bilik. Pengesanan jumlah kromosom titisan haploid ganda dua ditentukan dengan mengira bilangan kromosom di bawah mikroskop fluoresen. Tiga puluh sembilan DH telah dinilai di bawah 0, 50, 100 dan 150 mM NaCl di dalam rumah hijau. Titisan DH6 (Abo-Graib×G8), DH8 (Forat×G8) dan DH6 (Dijla×G8) menunjukkan prestasi yang tinggi dan melampaui batas untuk kandungan K⁺ pada daun, nisbah K⁺/Na⁺, bilangan spike setiap tumbuhan, jumlah bijirin setiap spike, dan penurunan dalam kandungan Na⁺ pada daun. Secara keseluruhannya, ia boleh disimpulkan bahawa melalui kultur anter, titisan haploid ganda dua yang memaparkan hasil yang tinggi telah berjaya dihasilkan untuk dieksploitasi dimasa hadapan.

ACKNOWLEDGEMENTS

First of all, I would like to express my greatest gratitude to merciful Allah SWT for his grace and for giving me the opportunity to complete my research and thesis writing. I thank the Ministry of Higher Education and Scientific Research, and Ministry of Agriculture, Iraq for giving me the opportunity to complete my post-graduate studies. I wish to express my greatest gratitude and respect to my supervisor, Prof. Datin Dr. Siti Nor Akmar Abdullah for her unlimited support, fruitful advice and valuable guidance during the period of my Ph.D. study. I also wish to express my thanks to the members of the supervisory committee, Prof. Dr. Mohd Rafii Yusop and Prof. Dr. Mohd Razi Ismail for their kindness and support. Regards to Associate Prof. Dr. Maheran Abdul Aziz who was my first supervisor but has recently retired. My deepest appreciation to all the lecturers and staff in UPM particularly those who shared their knowledge and experience with me. I also express my appreciation to the following Iraqi people who have helped me throughout the period of my study: Dr. Maged Shae, Medhat Elshahookie, Dr. Emad Khalaf, Dr. Hussien Hadi, Dr. Mohammed Omar, Dr. Husham Sarhan, Dr. Khdhayer Abbas, Dr. Hatem Jabbar, Dr. Ali Asseel and Dr. Nadir Alzubair. I wish to express my warm gratitude to my father, mother, brothers, and sisters for their support. I am very grateful to my wife Wahran and daughters Nabba, Msra and my son Anas who have been very understanding and supported me by their continuous encouragement, especially for traveling from Iraq and staying with me in Malaysia during my study.

I certify that a Thesis Examination Committee has met on 30 January 2018 to conduct the final examination of Abbas Lateef Abdulrahman on his thesis entitled "Performance of Crosses and Development of Doubled Haploid Lines in Wheat (*Triticum aestivum* L.) for Salt Tolerance" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Janna Ong binti Abdullah, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Uma Rani a/p Sinniah, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Ghizan bin Saleh, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Festo Massawe, PhD

Professor
University of Nottingham Campus Malaysia
Malaysia
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 28 March 2018

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Siti Nor Akmar Abdullah, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Mohd Rafii Yusop, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Mohd Razi Ismail, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software


Signature: _____ Date: _____

Name and Matric No: Abbas Lateef Abdulrahman, GS39683


Declaration by Members of Supervisory Committee

This is to confirm that:


- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: 
Name of
Chairman of
Supervisory
Committee: Prof. Datin Dr. Siti Nor Akmar Abdullah

PROF. DATIN DR. SITI NOR AKMAR ABDULLAH
Pengarah
Institut Kajian Perladangan
Universiti Putra Malaysia
43400 UPM Serdang, Selangor Darul Ehsan

Signature: 
Name of
Member of
Supervisory
Committee: Prof. Dr. Mohd Rafii Yusop

PROF. DR. MOHD RAFII YUSOP
Head
Laboratory of Climate-Smart Food Crop Production
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
43400 UPM Serdang, Selangor

Signature: 
Name of
Member of
Supervisory
Committee: Prof. Dr. Mohd Razi Ismail

PROF. DR. MOHD RAZI ISMAIL
Deputy Director
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
43400 UPM Serdang, Selangor

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF APPENDICES	xvi
LIST OF ABBREVIATIONS	xviii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	3
2.1 Developmental history (Phylogeny) of cultivated wheat	3
2.1.1 Diploid wheat	4
2.1.2 Tetraploid wheat	4
2.1.3 Hexaploid wheat	5
2.2 Significance of wheat as a food crop	5
2.3 Soil salinity	5
2.3.1 Processes used for studying the osmotic vs. salt-specific effects on the plant growth	6
2.3.2 Relationship between the time and salt specific effects	7
2.3.2.1 Salt specific effects on plant growth weeks after salt absorbtion	9
2.3.2.2 Salt specific effects on plant growth months after salt absorbtion	9
2.3.3 Soil salinity as a limiting factor of wheat production in Iraq	10
2.3.4 Salinity tolerance mechanisms	10
2.3.5 Effect of salinity on seed germination	13
2.3.6 Effect of salinity on seedling growth	14
2.3.7 Salinity effect on yield of grain	14
2.3.8 Approaches for improving the wheat plants for salt tolerance	15
3 SEED GERMINATION AND SEEDLINGS GROWTH OF HEXAPLOID WHEAT (<i>Triticum aestivum</i> L.) GENOTYPES AS INFLUENCED BY DIFFERENT LEVELS OF SODIUM CHLORIDE	24
3.1 Introduction	24
3.2 Materials and Methods	25

3.2.1	Plant Materials	25
3.2.2	Determination of water uptake by seeds	25
3.2.3	Determination of germination percent	26
3.2.4	Determination of plumule and radicle length (cm)	26
3.2.5	Strong seed index	26
3.2.6	Salt tolerance index	27
3.2.7	Statistical analysis	27
3.3	Results and Discussion	27
3.3.1	Water uptake	27
3.3.2	Seed germination percentage	30
3.3.3	Plumule and radicle length	31
3.3.4	Strong seed index	33
3.3.5	Correlation	35
3.4	Conclusion	35
4	DEVELOPMENT OF SALT TOLERANT F₁ WHEAT CROSSES AND EVALUATION OF THEIR GRAIN YIELD AND ITS COMPONENTS UNDER DIFFERENT CONCENTRATIONS OF SODIUM CHLORIDE	
		36
4.1	Introduction	36
4.2	Materials and Methods	36
4.2.1	Plant height (cm)	37
4.2.2	Number of spikes per plant	37
4.2.3	Weight of 1000 grain (g)	37
4.2.4	Number of grain per spike	38
4.2.5	Grain yield per plant (g)	38
4.2.6	Statistical Analysis	38
4.3	Results and Discussion	38
4.3.1	Grain yield and its components	38
4.3.2	Correlation	43
4.4	Conclusion	43
5	DEVELOPMENT OF DOUBLED HAPLOID WHEAT LINES FROM F₁ PROGENIES	
		44
5.1	Introduction	44
5.2	Materials and Methods	45
5.2.1	Growth of Donor Plants	45
5.2.2	Pre-treatment of Spikes	45
5.2.3	Culture of isolated anthers	47
5.2.4	Development of doubled haploid plants	48
5.2.5	Statistical analysis	50
5.3	Results and Discussion	50
5.3.1	Anther culture response of the F ₁ progenies	50
5.3.2	The effects of pre-treatment on anther culture	51
5.3.3	Effect of callus induction and regeneration media on studied traits	52

5.3.4	Effect of interaction of genotypes and pre-treatments on callus induction percentage	54
5.3.5	Effect of interaction between genotypes and media on percentage of callus induction, green and albino plants production	55
5.3.6	Effect of interaction among genotypes, pre-treatments, and media on percentage of callus induction	56
5.3.7	Effect of interaction among genotypes and media on percentage of green and albino plantlets production	56
5.4	Conclusion	59
6	AGRONOMIC PERFORMANCE OF DOUBLED HAPLOID WHEAT LINES UNDER SODIUM CHLORIDE STRESS	60
6.1	Introduction	60
6.2	Materials and Methods	60
6.2.1	Validation of haploid and doubled haploid lines	60
6.2.2	Factors used in experiment	63
6.2.3	Sampling strategy	63
6.3	Results and Discussions	64
6.4	Conclusion	73
7	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	74
7.1	Summary	74
7.2	General Conclusion	76
7.3	Recommendation for Future Research	76
	REFERENCES	77
	APPENDICES	103
	BIODATA OF STUDENT	130
	LIST OF PUBLICATIONS	131

LIST OF TABLES

Table		Page
1	Mean comparison of five traits studied by LSD of five concentrations of NaCl	28
2	Mean comparison of five traits studied by LSD of eight wheat genotypes	28
3	Salt tolerance traits indices (STTIs) of four traits studied in eight wheat genotypes	30
4	Salt tolerant categories of eight wheat genotypes according to salt tolerance index (STI) under five concentrations of NaCl	34
5	Correlation among five traits in eight varieties of wheat under various concentrations of NaCl	35
6	F ₁ progenies derived from crossing between their parents	37
7	Mean comparison of five traits studied by LSD of four concentrations of NaCl	39
8	Mean effect of traits studied in four six F ₁ progenies and their parents	40
9	Five traits mean comparison by LSD of four genotypes and their six F ₁ crosses under 0, 50 and 100 mM of NaCl	41
10	Correlation among five traits in six F ₁ crosses and their parents under various concentrations of NaCl	43
11	Mean comparison of callus induction by LSD of six F ₁ progenies	50
12	Mean comparison of green and albino plantlets by LSD of six F ₁ progenies	51
13	Mean comparison of callus induction by LSD at two pretreatments	52
14	Mean comparison of callus induction by LSD on three media	53
15	Mean comparison of green and albino plantlets by LSD of three regeneration media	53
16	Mean comparison of callus induction by LSD of six F ₁ progenies and two pretreatments of anther	54

17	Mean comparison of callus induction by LSD of interaction of six F ₁ progenies and three callus induction media	55
18	Mean comparison of green and albino plantlets by LSD of interaction of six F ₁ progenies and three regeneration media	55
19	Mean comparison of callus induction by LSD of interaction six F ₁ progenies, two pretreatments of anther followed by cultured on three callus induction media	56
20	Mean comparison of green and albino plantlets by LSD of interaction of six F ₁ progenies and three regeneration media	57
21	Eight traits mean comparison by LSD of three doubled haploid lines derived from Abo-Graib×Dijla cross	64
22	Eight traits mean comparison by LSD of five doubled haploid lines derived from Abo-Graib×Forat cross	65
23	Eight traits mean comparison by LSD of ten doubled haploid lines derived from Abo-Graib×G8 cross	65
24	Eight traits mean comparison by LSD of seven doubled haploid lines derived from Forat×Dijla cross	65
25	Eight traits mean comparison by LSD of eight doubled haploid lines derived from the Forat×G8 cross	66
26	Eight traits mean comparison by LSD of six doubled haploid lines derived from the Dijla×G8 cross	66
27	Eight traits mean comparison by LSD of three doubled haploid lines derived from the Abo-Graib×Dijla cross	67
28	Eight traits mean comparison by LSD of five doubled haploid lines derived from the Abo-Graib×Forat cross	68
29	Eight traits mean comparison by LSD of ten doubled haploid lines derived from the Abo-Graib×G8 cross	69
30	Eight traits mean comparison by LSD of seven doubled haploid lines derived from the Forat×Dijla cross	70
31	Eight traits mean comparison by LSD of eight doubled haploid lines derived from the Forat×G8 cross	71
32	Eight traits mean comparison by LSD of six doubled haploid lines derived from the Dijla×G8 cross	72

LIST OF FIGURES

Figure		Page
1	An overview of the Triticum wheat diversification	3
2	The evaluation of the <i>T. turgidum</i> - <i>T. aestivum</i> complex	4
3	The trio of basic processes describing salinity tolerance in crop plants	11
4	Comparison between traditional breeding method and anther culture technique	20
5	Comparison between traditional and doubled haploid wheat variety development	22
6	Effect of interaction between concentration of NaCl and genotypes on water uptake%	29
7	Effect of interaction between NaCl concentration and genotypes on plumule length	32
8	Effect of interaction between NaCl concentration and genotypes on radicle length	33
9	Effect of interaction between NaCl concentration and genotypes on strong seed index	34
10	Suitable developmental stages of uninucleate microspore for anther culture	46
11	Steps in anther culture procedure for doubled haploid wheat lines production	49
12	Validation of chromosomes number of haploid and doubled haploid wheat plants	62

LIST OF APPENDICES

Appendix		Page
1	Types of wheat and their species, genome constitution, and common names	102
2	Response of plant to salinity at different time scales	103
3	Mean square of variance analysis on germination percentage and growth of seedling characteristics under different concentrations of NaCl	104
4	Salt tolerance traits indices (STTIs) mean square of four traits studied in eight wheat genotypes under five concentrations of NaCl	104
5	A pedigree of wheat parents	104
6	Chemical and physical properties of soil	105
7	Mean square of variance analysis on five traits studied in four genotypes and their six F ₁ crosses under different concentrations of NaCl	105
8	Composition of callus induction media	106
9	Composition of regeneration media	107
10	Mean square of analysis of variance of callus induction in six F ₁ crosses	108
11	Mean square of analysis of variance of green and albino plantlets in six F ₁ crosses	108
12	Mean square of analysis of variance of traits studied for the Abo-Graib×Dijla cross	109
13	Mean square of analysis of variance of traits studied for the Abo-Graib×Forat cross	109
14	Mean square of analysis of variance of traits studied for the Abo-Graib×G8 cross	110
15	Mean square of analysis of variance of traits studied for the Forat×Dijla cross	110

16	Mean square of analysis of variance of traits studied for the Forat×G8 cross	111
17	Mean square of analysis of variance of traits studied for the Dijla×G8 cross	111
18	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Abo-Graib×Dijla cross	112
19	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Abo-Graib×Dijla cross	113
20	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Abo-Graib×Forat cross	115
21	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Abo-Graib×Forat cross	116
22	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Abo-Graib×G8 cross	118
23	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Abo-Graib×G8 cross	119
24	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Forat×Dijla cross	121
25	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Forat×Dijla cross	122
26	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Forat×G8 cross	124
27	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Forat×G8 cross	125
28	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Dijla×G8 cross	127
29	Mean performance for the studied traits of doubled haploid (DH) lines derived from the Dijla×G8 cross	128

LIST OF ABBREVIATIONS

ANOVA	Analyses of Variance
BAP	Benzylaminopurine
bp	base pair
C1	The initial Conc. (mM) of stock solution
C2	Final concentration
CHB3	Callus induction medium
2, 4-D	2, 4-Dichlorophenoxyacetic acid
DAPI	4'-6-diamidino-2-phenylindole HCl staining
df	Degree of freedom
DH	Doubled haploid
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
EC	Electrical Conductivity
ETOH	Ethyl Alcohol
FDA	Fluorescein diacetate
FGP	Final germination percentage of seeds
GY	Grain yield
HIR	High haploid induction rate
2-HNA	2-Hydroxynicotinic acid
K ⁺	Potassium ion
K ⁺ /Na ⁺	Ratio of Potassium ion to Sodium ion
LSD	Least significant difference
mg/L	Milligrams per liter

mM	Mill molar
Mpa	Megapascal
MS	Murashige and Skoog medium
N	Total number of seeds
Na ⁺	Sodium ion
NAA	Naphthalene acetic acid
NaCl	Sodium Chloride
Ni	Number of germinated seed
NG	Number of grain
NS	Number of spikes
PCR	Polymorphism chain reaction
PH	Plant height
pH	Negative logarithm of the hydrogen ion concentration
QTL	Quantitative trait locus
RCBD	Randomized Complete Block Design
S.O.V	Source of variance
STTIs	Salt tolerance traits indices
TBE	Tris/Borate/EDTA buffer
V1	Initial Volume
V2	Final volume
W1	Initial weight of seed
W2	Weight of seed after water uptake at a particular time
WG	Weight of grain

CHAPTER 1

INTRODUCTION

In the developing countries, around 2.5 billion people depend on wheat (*T. aestivum*) as their food source (FAOSTAT, 2010). For them, wheat is a staple commodity whose production involves around 200 million farmers and their families. Production of wheat and its yield directly impact human existence in developing nations and quality of life in industrial economies. From 1995 to 2013, a 1.1% (Dixon et al., 2009) to 0.9% (Ray et al., 2013) yearly rise in productivity was noted in around 85% of 20 wheat producing nations. Projections based on this growth rate indicated that wheat productivity would rise 17% by 2025, which is lower than the required 25% to feed the projected populace at that time (William et al., 2008). More improvements in wheat productivity can be accomplished by utilizing the land resources that are at present unfeasible to produce crops such as wheat because of the surplus of salts and other abiotic stresses.

It is projected that around 60% rise in cereal production (Rosegrant and Cline, 2003; Long et al., 2015) and 38% in wheat production (Ray et al., 2013) is necessary to meet the global demand by the year 2050. This situation highlights that food security is still the most fundamental concern despite advancement and deployment of advanced technologies. Selection of varieties using conventional technique needs many generations of selection which may take more than ten years to produce a variety (Mansouri et al., 2005; Breseghello and Coelho, 2013). However, through the procedure of *in vitro* culture, the time required for the creation of new lines can be significantly shortened by taking benefits of existing genetic variation (Maheswary and Mak, 1993; Tadesse et al., 2012). In addition to conventional breeding techniques used for wheat development (Su et al., 1992; Wilkinson et al., 2012), the anther culture technique could be used as an untraditional supplementary technique (Santra et al., 2012). *In vitro* anther culture exhibits a fast technique for production of true breeding lines in the next generation from any segregated species (Marassi et al., 1993; Yan et al., 2017) and promoting homozygosity (Smith and Drew, 1990; Asif et al., 2014). Breeding researchers showed that the development of doubled haploid (Mohiuddin et al., 2006; Ma'arup et al., 2012) to achieve homozygosity allows the possibility for crop improvement in producing the wanted trait at a faster time through minimizing the breeding processes (Hassawi et al., 2005; El-Hennawy et al., 2011).

For a successful breeding programme using the microspores for an increased doubled haploid production, it is important to use genotypes having higher regeneration capacity. Several studies stated that the anther culture is heritable through crossing (Grauda et al., 2010; Nielsen et al., 2015). However, since the varieties are derived from several genetic sources, it is possible to find good genotypes in unselected crosses or genetic lines (Govindaraj et al., 2015).

Stress treatment for inducing embryogenesis is a prerequisite for the success of anther culture. Different stresses have been applied including physical, physiological and chemical stresses of excised spikes, anthers or microspores for inducing androgenesis (Zorinants et al., 2005; Echavarri and Cistue, 2016). In general, it seems that stress performances by applying the pretreatment lead to the change from the gametophytic pathway to the saprophytic pathway (Rakha et al., 2012).

Production of albino plants often limits the capability of anther culture from cereals such as wheat (Lantos et al., 2013). Therefore, it is important to provide a suitable environment which is necessary for the development and proliferation of the embryogenic microspores and production of embryos. Auxiliary factors during this stage may include improved physical conditions and appropriate elements in the media particularly glutamine (amino acid), myo-inositol, sulfate pentahydrate and maltose (Ekhveh et al., 2013). For the development of high grain yielding-salt tolerant wheat genotypes, it is hypothesized there would be tolerant genotypes for different concentrations of sodium chloride for selection. With some chosen wheat genotypes known to be high grain yield- or salt tolerant, it is hypothesized that F₁ crosses that are both high yielding and tolerance to salinity could be developed by crossing among these selected genotypes. Development of pure lines using self-pollinating crops needs many generations after making a cross to obtain homozygosity and uniformity to be able to evaluate of inherited traits such as yield and quality. However, doubled haploids can acquire uniformity in short generations. Therefore, it is hypothesized that high grain yielding-salt tolerant doubled haploid lines of wheat using anther culture technique can be developed from F₁ crosses. The objectives of this study were to

- 1- Study the effect of salinity on seed germination percentage and seedling growth traits of eight bread wheat genotypes (*Triticum aestivum* L.) and select the best genotypes for the development of sodium chloride tolerant progenies
- 2- Obtain F₁ crosses by crossing among genotypes based on their high yielding and salt tolerant traits and evaluate their growth and grain yield performance under different concentrations of NaCl
- 3- Develop high grain yielding-salt tolerant homozygous pure lines derived from segregating populations of heterozygous F₁'s and evaluate their agronomic performance under salt stress conditions to select the superior salt tolerant high grain yielding lines

REFERENCES

- Abbas, M. K., Ali, A. S., Hasan, H. H. and Ghal, R. H. 2012. Salt tolerance study of six cultivars of rice (*Oryza sativa* L.) during germination and early seedling growth. *Journal of Agricultural Science* 5(1): 250.
- Abdelkader, S., Ramzi, C., Mustapha, R., Houcine, B., M'barek, B. N., Inagaki, M. N. and Abdallah, B. 2015. Effect of salt stress on germination and biological growth of 50 genotypes of durum wheat (*Triticum durum* Desf). *Pakistan Journal of Nutrition* 14(12): 957.
- Abdul-Baki, A. A. and Anderson, J. D. 1970. Viability and leaching of sugars from germinating barley. *Crop Science* 10(1): 31–34.
- Ahmad, M. 2002. Assessment of genomic diversity among wheat genotypes as determined by simple sequence repeats. *Genome* 45(4): 646–651.
- Ahmad, M., Munir, M., Ahmad, I. and Yousuf, M. 2011. Evaluation of bread wheat genotypes for salinity tolerance under saline field conditions. *African Journal of Biotechnology* 10(20): 4086–4092.
- Ahmad, M., Shahzad, A., Iqbal, M., Asif, M. and Hirani, A.H. 2013. Morphological and molecular genetic variation in wheat for salinity tolerance at germination and early seedling stage. *Australian Journal of Crop Science* 7(1): 66.
- Akbari, G., Sanavy, S. A. and Yousefzadeh, S. 2007. Effect of auxin and salt stress (NaCl) on seed germination of wheat cultivars (*Triticum aestivum* L.). *Pakistan journal of Biological Sciences: PJBS* 10(15): 2557–2561.
- Al-Ashkar, I. M. 2013. Anther culture response and salt tolerance in some wheat genotypes. *Annals of Agricultural Sciences* 58(2): 139–145.
- Ali, Z., Khan, A. S. and Asad, M. A. 2002. Salt tolerance in bread wheat: Genetic variation and heritability for growth and ion relation. *Asian Journal of Plant Science* 1: 420–422.
- Ali, Z., Salam, A., Azhar, F. M. and Khan, I.A. 2007. Genotypic variation in salinity tolerance among spring and winter wheat (*Triticum aestivum* L.) accessions. *South African Journal of Botany* 73(1): 70–75.
- Ali, Z., Salam, A., Azhar, F. M., Khan, I. A., Khan, A. A., Bahadur, S., Mahmood, T., Ahmad, A. and Trethowan, R. 2012. The response of genetically distinct bread wheat genotypes to salinity stress. *Plant Breeding* 131(6): 707–715.
- Amor, F. M., Martinez, V. and Cerda, A. 2001. Salt tolerance of tomato plants as affected by stage of plant development. *Horticulture Science* 36(7): 1260–1263.
- Araki, H., Hossain, M. A. and Takahashi, T. 2012. Waterlogging and hypoxia have permanent effects on wheat root growth and respiration. *Journal of Agronomy and Crop Science* 198(4): 264–275.

- Aronson, J. A. and Whitehead, E. E. 1989. *HALOPH*: a data base of salt tolerant plants of the world.
- Ashraf, M. and Foolad, M. R. 2013. Crop breeding for salt tolerance in the era of molecular markers and marker-assisted selection. *Plant Breeding* 132(1): 10–20.
- Asif, M., Eudes, F., Randhawa, H., Amundsen, E. and Spaner, D. 2014. Phytosulfokine alpha enhances microspore embryogenesis in both triticale and wheat. *Plant Cell, Tissue and Organ Culture (PCTOC)* 116(1): 125–130.
- Asouti, E., Fuller, D. Q., Barker, G., Finlayson, B., Matthews, R., Fazeli Nashli, H., McCorrison, J., Riehl, S., Rosen, A. M. and Asouti, E. 2013. A contextual approach to the emergence of agriculture in Southwest Asia: reconstructing Early Neolithic plant-food production. *Current Anthropology* 54(3): 0.
- Assadian, N. W. and Miyamoto, S. 1987. Salt effects on alfalfa seedling emergence. *Agronomy Journal* 79(4): 710–714.
- Assaha, D. V. M., Ueda, A., Saneoka, H., Al-Yahyai, R. and Yaish, M. W. 2017. The role of Na⁺ and K⁺ transporters in salt stress adaptation in glycophytes. *Frontiers in Physiology* 8: 1-19.
- Ayed-Slama, O., Ayed, S. and Slim-Amara, H. 2015. Selection of tolerant lines for salinity derived from durum wheat (*Triticum durum* Desf.) *in vitro* culture. *Agricultural Sciences* 6(7): 699–706.
- Ayed, O. S., Buyser, J. D., Picard, E., Trifa, Y. and Amara, H. S. 2010. Effect of pretreatment on isolated microspores culture ability in durum wheat (*Triticum turgidum* subsp. durum Desf.). *Journal of Plant Breeding and Crop Sciences* 2: 30–38.
- Bahrani, H. and Razmjoo, J. 2012. Effect of salinity stress (NaCl) on germination and early seedling growth of ten sesame cultivars (*Sesamum indicum* L.). *International Journal of AgriScience* 2(6): 529–537.
- Bahrani, A. and Haghjoo, M. 2012. Response of some wheat (*Triticum aestivum* L.) genotypes to salinity at germination and early seedling growth stages. *World Applied Sciences Journal* 16(4): 599–609.
- Barakat, M. N., Al-Doss, A. A., Elshafei, A. A., Moustafa, K. A. and Ahmed, E. I. 2012. Anther culture response in wheat (*Triticum aestivum* L.) genotypes with HMW alleles. *Cereal Research Communications* 40(4): 583–591.
- Barakat, M. N., Al-Doss, A. A., Ghazy, A. I., Moustafa, K. A., Elshafei, A. A. and Ahmed, E. I. 2017. Doubled haploid wheat lines with high molecular weight glutenin alleles derived from microspore cultures. *New Zealand Journal of Crop and Horticultural Science* 45: 1–14.
- Barkley, A., Chumley, F. G. and Innovations, H. P. 2012. A doubled haploid laboratory for Kansas wheat breeding: an economic analysis of biotechnology adoption. *International Food and Agribusiness Management Review* 15: 99–

- Barragan, V., Leidi, E. O., Andres, Z., Rubio, L., De Luca, A., Fernandez, J.A., Cubero, B. and Pardo, J.M. 2012. Ion exchangers NHX1 and NHX2 mediate active potassium uptake into vacuoles to regulate cell turgor and stomatal function in Arabidopsis. *The Plant Cell* 24(3): 1127–1142.
- Barrett-Lennard, E. G. 2003. The interaction between waterlogging and salinity in higher plants: causes, consequences and implications. *Plant and Soil* 253(1): 35–54.
- Bennett, E., Roberts, J. A. and Wagstaff, C. 2012. Manipulating resource allocation in plants. *Journal of Experimental Botany* 63(9): 3391–3400.
- Bernhardt, N. 2015. Taxonomic treatments of Triticeae and the wheat genus Triticum (PP. 1–19). In *Alien Introgression in Wheat*. Springer.
- Botia, P., Carvajal, M., Cerdá, A. and Martinez, V. 1998. Response of eight Cucumis melo cultivars to salinity during germination and early vegetative growth. *Agronomie* 18(8–9): 503–513.
- Breseghello, F. and Coelho, A. S. G. 2013. Traditional and modern plant breeding methods with examples in rice (*Oryza sativa* L.). *Journal of Agricultural and Food Chemistry* 61(35): 8277–8286.
- Brew-Appiah, R. A. T., Ankrah, N., Liu, W., Konzak, C. F., Von Wettstein, D. and Rustgi, S. 2013. Generation of doubled haploid transgenic wheat lines by microspore transformation. *PLoS One* 8(11): e80155.
- Bustos, D. V, Hasan, A. K., Reynolds, M. P. and Calderini, D. F. 2013. Combining high grain number and weight through a DH-population to improve grain yield potential of wheat in high-yielding environments. *Field Crops Research* 145: 106–115.
- Cabot, C., V Sibole, J., Barceló, J. and Poschenrieder, C. 2014. Lessons from crop plants struggling with salinity. *Plant Science* 226: 2–13.
- Castillo, A.M., Nielsen, N.H., Jensen, A. and Valles, M.P. 2014. Effects of n-butanol on barley microspore embryogenesis. *Plant Cell, Tissue and Organ Culture (PCTOC)* 117(3): 411–418.
- Catalán, L., Balzarini, M., Taleisnik, E., Sereno, R. and Karlin, U. 1994. Effects of salinity on germination and seedling growth of *Prosopis flexuosa* (DC). *Forest Ecology and Management* 63(2–3): 347–357.
- Ceccoli, G., Ramos, J., Pilatti, V., Dellaferrera, I., Tivano, J. C., Taleisnik, E. and Vegetti, A. C. 2015. Salt glands in the Poaceae family and their relationship to salinity tolerance. *The Botanical Review* 81(2): 162–178.
- Chakraborty, K., Singh, A. L., Bhaduri, D. and Sairam, R. K. 2013. Mechanism of salinity stress tolerance in crop plants and recent developments. *Advances in Plant Physiology* 14: 466–496.

- Chandra, C. R., Kishor, S. N., Mithilesh, K. and Rajeev, K. 2014. Wheat genotypes (*Triticum aestivum* L.) vary widely in their responses of fertility traits to high temperature at anthesis. *International Research Journal of Biological Sciences* 3(7): 54–60.
- Chandra, R., Takeuchi, H., Hasegawa, T. and Kumar, R. 2012. Improving biodegradability and biogas production of wheat straw substrates using sodium hydroxide and hydrothermal pretreatments. *Energy* 43(1): 273–282.
- Chapman, V.J. 2013. *Salt marshes and salt desert of the world*: 3–19.
- Chartzoulakis, K. and Klapaki, G. 2000. Response of two greenhouse pepper hybrids to NaCl salinity during different growth stages. *Scientia Horticulturae* 86(3): 247–260.
- Chauhan, H. and Khurana, P. 2011. Use of doubled haploid technology for the development of stable drought tolerant bread wheat (*Triticum aestivum* L.) transgenics. *Plant Biotechnology Journal* 9(3): 408–417.
- Chauhan, B. S., Mahajan, G., Sardana, V., Timsina, J. and Jat, M. L. 2012. Productivity and sustainability of the rice-wheat cropping system in the Indo-Gangetic Plains of the Indian subcontinent: problems, opportunities, and strategies. *Advances in Agronomy* 117(1): 315–369.
- Chazen, O., Hartung, W. and Neumann, P. M. 1995. The different effects of PEG 6000 and NaCl on leaf development are associated with differential inhibition of root water transport. *Plant, Cell and Environment* 18(7): 727–735.
- Cheeseman, J. 2016. Food security in the face of salinity, drought, climate change, and population growth. *Halophytes for Food Security in Dry Lands*: 111–123.
- Chu, C. C. 1978. *Proceedings of Symposium on Plant Tissue Culture*, Beijing, Science Press, pp 43-50.
- Çifci, E.A. 2012. Estimate of heterosis, correlation and path analysis for grain yield per spike and some agronomic traits on durum wheat (*Triticum durum* Desf.). *The Journal of Animal and Plant Sciences*, 22 (3): 747–752.
- Cistue, L., Romagosa, I., Batlle, F. and Echavarri, B. 2009. Improvements in the production of doubled haploids in durum wheat (*Triticum turgidum* L.) through isolated microspore culture. *Plant Cell Reports* 28(5): 727–735.
- Civávn, P., Ivaničová, Z. and Brown, T. A. 2013. Reticulated origin of domesticated emmer wheat supports a dynamic model for the emergence of agriculture in the fertile crescent. *PLoS One* 8(11): e81955.
- Colmer, T. D. 2006. Use of wild relatives to improve salt tolerance in wheat. *Journal of Experimental Botany* 57(5): 1059–1078..
- Colmer, T. D., Epstein, E. and Dvorak, J. 1995. Differential solute regulation in leaf blades of various ages in salt-sensitive wheat and a salt-tolerant wheat×*Lophopyrum elongatum* (Host) A. Love amphiploid. *Plant Physiology*

108(4): 1715–1724.

- Colmer, T. D., Munns, R. and Flowers, T. J. 2006. Improving salt tolerance of wheat and barley: future prospects. *Animal Production Science* 45(11): 1425–1443.
- Cramer, G. R. and Bowman, D. C. 1991. Kinetics of maize leaf elongation I. Increased yield threshold limits short-term, Steady-state elongation rates after exposure to salinity. *Journal of Experimental Botany* 42(11): 1417–1426.
- Curtis, T. and Halford, N. G. 2014. Food security: the challenge of increasing wheat yield and the importance of not compromising food safety. *Annals of Applied Biology* 164(3): 354–372.
- Datta, J. K., Nag, S., Banerjee, A. and Mondai, N. K. 2009. Impact of salt stress on five varieties of Wheat (*Triticum aestivum* L.) cultivars under laboratory condition. *Journal of Applied Sciences and Environmental Management* 13(3): 93–97.
- Davenport, R., James, R. A., Zakrisson-Plogander, A., Tester, M. and Munns, R. 2005. Control of sodium transport in durum wheat. *Plant Physiology* 137(3): 807–818.
- Debez, A., Saadaoui, D., Ramani, B., Ouerghi, Z., Koyro, H.W., Huchzermeyer, B. and Abdelly, C. 2006. Leaf H⁺ ATPase activity and photosynthetic capacity of *Cakile maritima* under increasing salinity. *Environmental and Experimental Botany* 57(3): 285–295.
- Deinlein, U., Stephan, A. B., Horie, T., Luo, W., Xu, G. and Schroeder, J. I. 2014. Plant salt-tolerance mechanisms. *Trends in Plant Science* 19: 371–379.
- Deoliveira, A. B., Alencar, N. L.M. and Gomes-Filho, E. 2013. Comparison between the water and salt stress effects on plant growth and development (PP. 67–94). In *Responses of Organisms to Water Stress*. Intech.
- DePauw, R. M., Knox, R. E., Humphreys, D. G., Thomas, J. B., Fox, S. L., Brown, P. D., Singh, A. K., Pozniak, C., Randhawa, H. S. and Fowler, D. B. 2011. New breeding tools impact Canadian commercial farmer fields. *Czech Journal of Genetics and Plant Breeding* 47: 28-34.
- Devaux, P. and Pickering, R. 2005. Haploids in the improvement of Poaceae (PP. 215–242). In *Haploids in Crop Improvement II*. Springer.
- Dixon, J., Braun, H. J. and Crouch, J. H. 2009. Overview: transitioning wheat research to serve the future needs of the developing world. *Wheat Facts and Futures* 2009 (23): 1–25.
- Echavarri, B. and Cistue, L. 2016. Enhancement in androgenesis efficiency in barley (*Hordeum vulgare* L.) and bread wheat (*Triticum aestivum* L.) by the addition of dimethyl sulfoxide to the mannitol pretreatment medium. *Plant Cell, Tissue and Organ Culture (PCTOC)* 125(1): 11–22.

- Ekhveh, M. J., Moieni, A. and Jalali Javaran, M. 2013. The evaluation of response to isolated microspores culture in some Iranian hexaploid wheat (*Triticum aestivum* L.) cultivars. *Journal of Agricultural Science* 5(7): 206–216.
- El-Hendawy, S. E., Hu, Y., Sakagami, J. I. and Schmidhalter, U. 2012. Screening Egyptian wheat genotypes for salt tolerance at early growth stages. *International Journal of Plant Production* 5(3): 283–298.
- El-Hendawy, S. E., Hu, Y., Yakout, G. M., Awad, A. M., Hafiz, S. E. and Schmidhalter, U. 2005. Evaluating salt tolerance of wheat genotypes using multiple parameters. *European Journal of Agronomy* 22(3): 243–253.
- El-Hendawy, S. E., Ruan, Y., Hu, Y. and Schmidhalter, U. 2009. A comparison of screening criteria for salt tolerance in wheat under field and controlled environmental conditions. *Journal of Agronomy and Crop Science* 195(5): 356–367.
- El-Hennawy, M. A., Abdalla, A. F., Shafey, S. A. and Al-Ashkar, I. M. 2011. Production of doubled haploid wheat lines (*Triticum aestivum* L.) using anther culture technique. *Annals of Agricultural Sciences* 56(2): 63–72.
- Eskandari, H. and Kazemi, K. 2011. Germination and Seedling Properties of Different Wheat Cultivars under Salinity Conditions. *Notulae Scientia Biologicae* 3(3): 130–134.
- Fahramand, M., Mahmoodi, M., Keykha, A., Noori, M. and Rigi, K. 2014. Influence of abiotic stress on proline, photosynthetic enzymes and growth. *International Research Journal of Applied and Basic Sciences* 8(3): 257–265.
- FAOSTAT, F.A.O. 2010. FAO Statistical Database. Acessado em setembro.
- FAOSTAT. 2015. FAO Statistical Database.
- Faris, J. D. 2014. Wheat domestication: Key to agricultural revolutions past and future. (PP. 439–464). In *Genomics of Plant Genetic Resources*. Springer.
- Fauziah, C. 2010. Temporal changes in chemical properties of acid soil profiles treated with magnesium limestone and gypsum. *Pertanika Journal of Tropical Agricultural Science* 33(2): 277–295.
- Fazeli-nasab, B., Omid, M. and Amiritokaldani, M. 2012. Callus induction and plant regeneration of wheat mature embryos under abscisic acid treatment. *International Journal of Agriculture and Crop Sciences* 4: 17–23.
- Fercha, A., Capriotti, A. L., Caruso, G., Cavaliere, C., Samperi, R. S. Stampacchiacchiere, R. and Laganà, A. 2014. Comparative analysis of metabolic proteome variation in ascorbate-primed and unprimed wheat seeds during germination under salt stress. *Journal of proteomics* 108: 238–257.
- Filiault, D. L., Seymour, D. K., Maruthachalam, R. and Maloof, J. N. 2017. The generation of doubled haploid Lines for QTL mapping. *plant genomics: Methods and Protocols*: 39–57.

- Flowers, T. J. and Colmer, T. D. 2015. Plant salt tolerance: adaptations in halophytes. *Annals of botany* 115(3): 327–331.
- Flowers, T. J. and Hajibagheri, M. A. 2001. Salinity tolerance in *Hordeum vulgare*: ion concentrations in root cells of cultivars differing in salt tolerance. *Plant and Soil* 231(1): 1–9.
- Flowers, T. J., Hajibagherp, M. A. and Yeo, A. R. 1991. Ion accumulation in the cell walls of rice plants growing under saline conditions: evidence for the Oertli hypothesis. *Plant, Cell and Environment* 14(3): 319–325.
- Forni, C., Duca, D. and Glick, B. R. 2017. Mechanisms of plant response to salt and drought stress and their alteration by rhizobacteria. *Plant and soil* 410(1–2): 335–356.
- Frauen, M. 1994. Use of biotechnology in breeding new varieties of winter oilseed rape in Germany. *Bul. GCIRC* 10: 16–29.
- Frensch, J. and Hsiao, T. C. 1994. Transient responses of cell turgor and growth of maize roots as affected by changes in water potential. *Plant Physiology* 104(1): 247–254.
- Fricke, W., Akhiyarova, G., Wei, W., Alexandersson, E., Miller, A., Kjellbom, P. O., Richardson, A., Wojciechowski, T., Schreiber, L. and Veselov, D. 2006. The short-term growth response to the salt of the developing barley leaf. *Journal of Experimental Botany* 57(5): 1079–1095.
- Fricke, W., Bijanzadeh, E., Emam, Y. and Knipfer, T. 2014. Root hydraulics in salt-stressed wheat. *Functional Plant Biology* 41(4): 366–378.
- Gallandt, E. R., Dofing, S. M., Reisenauer, P. E. and Donaldson, E. 2001. Diallel analysis of cultivar mixtures in winter wheat. *Crop Science* 41(3): 792–796.
- Garcia, G. A., Hasan, A. K., Puhl L. E., Reynolds, M. P., Calderini, D. F. and Miralles, D. J. 2013. Grain yield potential strategies in an elite wheat double-haploid population grown in contrasting environments. *Crop Science* 53(6): 2577–2587.
- Garcia Morales, S., Trejo-Téllez, L. I., Gómez Merino, F. C., Caldana, C., Espinosa-Victoria, D. and Herrera Cabrera, B. E. 2012. Growth, photosynthetic activity, and potassium and sodium concentration in rice plants under salt stress. *Acta Scientiarum. Agronomy* 34(3): 317–324.
- Garthwaite, A. J., von Bothmer, R. and Colmer, T. D. 2005. Salt tolerance in wild *Hordeum* species is associated with restricted entry of Na⁺ and Cl⁻ into the shoots. *Journal of Experimental Botany* 56(419): 2365–2378.
- Gaut, B. S. 2002. Evolutionary dynamics of grass genomes. *New phytologist* 154(1): 15–28.
- Genc, Y., Oldach, K., Verbyla, A. P., Lott, G., Hassan, M., Tester, M., Wallwork, H. and Mcdonald, G.K. 2010. Sodium exclusion QTL associated with improved

- seedling growth in bread wheat under salinity stress. *Theoretical and Applied Genetics* 121(5): 877–894.
- George, L. Y. and Williams, W. A. 1964. Germination and respiration of barley, strawberry clover, and ladino clover seeds in salt solutions. *Crop Science* 4(5): 450–452.
- Germana, M. A. 2011. Anther culture for haploid and doubled haploid production. *Plant Cell, Tissue and Organ Culture* (PCTOC) 104(3): 283–300.
- Gilliham, M., Able, J. A. and Roy, S. J. 2017. Translating knowledge about abiotic stress tolerance to breeding programmes. *The Plant Journal* 90(5): 898–917.
- Goertz, S. H. and Coons, J. M. 1991. Tolerance of tepary and navy beans to NaCl during germination and emergence. *Horticulture Science* 26(3): 246–249.
- Gomez-Pando, L. R., Jimenez-Davalos, J., Labarra, A., Aguilar-Castellanos, E., Falconi-Palomino, J., Ibañez-Tremolada, M., Varela, M. and Lorenzo, J. C. 2009. Field performance of new in vitro androgenesis-derived double haploids of barley. *Euphytica* 166(2): 269–276.
- Govindaraj, M., Vetriventhan, M. and Srinivasan, M. 2015. Importance of genetic diversity assessment in crop plants and its recent advances: an overview of its analytical perspectives. *Genetics Research International* 2015: 1–14.
- Grauda, D., Lepse, N., Strazdicna, V., Kokina, I., Lapicna, L., Mickelson, A., Clubinskis, L. and Rashal, I. 2010. Obtaining of doubled haploid lines by anther culture method for the Latvian wheat breeding. *Agronomy Research* 8(3): 545–552.
- Grzesiak, M., Filek, M., Barbasz, A., Kreczmer, B. and Hartikainen, H. 2013. Relationships between polyamines, ethylene, osmoprotectants and antioxidant enzymes activities in wheat seedlings after short-term PEG and NaCl induced stresses. *Plant Growth Regulation* 69(2): 177–189.
- Guasmi, F., Elfalleh, W., Hannachi, H., Fères, K., Touil, L., Marzougui, N., Triki, T. and Ferchichi, A. 2013. Influence of various physical parameters on anther culture of barley. *Journal of Plant Nutrition* 36(5): 836–847.
- Gupta, B. and Huang, B. 2014. Mechanism of salinity tolerance in plants: physiological, biochemical, and molecular characterization. *International Journal of Genomics* 2014.
- Hallajian, M. T. 2016. Mutation breeding and drought stress tolerance in plants (PP. 359–383). In *Drought Stress Tolerance in Plants, Vol 2*. Springer.
- Hampson, C. R. and Simpson, G. M. 1990. Effects of temperature, salt, and osmotic potential on early growth of wheat (*Triticum aestivum*). I. Germination. *Canadian Journal of Botany* 68(3): 524–528.
- Hancock, J. F. 2012. *Plant evolution and the origin of crop species*. CABI.

- Hanif, Z., Naeem, M., Ali, H. H., Tanveer, A., Javaid, M. M. Peerzada, A. M. and Chauhan, B. S. 2017. Effect of environmental factors on germination of *Salsola foetida*: potential species for rehabilitation of degraded rangelands. *Rangeland Ecology and Management* 70(5): 638–643.
- Hanumantharao, B., Nair, R. M. and Nayyar, H. 2016. Salinity and high temperature tolerance in mungbean [*Vigna radiata* (L.) Wilczek] from a physiological perspective. *Frontiers in Plant Science* 7(957): 1–20.
- Haq, T. U., Akhtar, J., Steele, K. A., Munns, R. and Gorham, J. 2014. Reliability of ion accumulation and growth components for selecting salt tolerant lines in large populations of rice. *Functional Plant Biology* 41(4): 379–390.
- Hasanuzzaman, M., Nahar, K., Alam, M. M., Bhowmik, P.C., Hossain, M. A., Rahman, M. M., Prasad, M. N.V., Ozturk, M. and Fujita, M. 2014. Potential use of halophytes to remediate saline soils. *BioMed research International* 2014.
- Hassawi, D. S., Qrunfleh, I. and Dradkah, N. 2005. Production of doubled haploids from some Jordanian wheat cultivars via anther culture technique. *Journal of Food Agriculture and Environment* 3(1): 161–164.
- Hazman, M., Hause, B., Eiche, E., Riemann, M. and Nick, P. 2016. Different forms of osmotic stress evoke qualitatively different responses in rice. *Journal of Plant Physiology* 202: 45–56.
- Hazarika, R. R., Mishra, V. K. and Chaturvedi, R. 2013. In Vitro Haploid Production. A Fast and Reliable Approach for Crop Improvement.(PP.171–212). In *Crop Improvement Under Adverse Conditions*. Springer.
- Herzog, M., Striker, G. G., Colmer, T. D. and Pedersen, O. 2016. Mechanisms of waterlogging tolerance in a wheat—a review of root and shoot physiology. *Plant, Cell and Environment* 39(5): 1068–1086.
- Hetherington, A. M. and Brownlee, C. 2004. The generation of Ca²⁺ signals in plants. *Annual Review of Plant Biology*. 55: 401–427.
- Hollington, P. A. 1998. Technological breakthroughs in screening/breeding wheat varieties for salt tolerance (PP. 2–5). In *Proceedings of the National Conference on Salinity Management in Agriculture*.
- Homayoun, H. 2011. Effect of NaCl salinity on wheat (*Triticum aestivum* L.) cultivars at germination stage. *Advances in Environmental Biology*: 1716–1721.
- Horie, T., Karahara, I. and Katsuhara, M. 2012. Salinity tolerance mechanisms in glycophytes: an overview with the central focus on rice plants. *Rice* 5(1): 11.
- Hsiao, T. C. and Xu, L. K. 2000. Sensitivity of growth of roots versus leaves to water stress: biophysical analysis and relation to water transport. *Journal of Experimental Botany* 51(350): 1595–1616.
- Hu, Y., Burucs, Z., Von Tucher, S. and Schmidhalter, U. 2007. Short-term effects of

drought and salinity on mineral nutrient distribution along growing leaves of maize seedlings. *Environmental and Experimental Botany* 60(2): 268–275.

- Hu, T. and Kasha, K. J. 1999. A cytological study of pretreatments used to improve isolated microspore cultures of wheat (*Triticum aestivum* L.) cv. Chris. *Genome* 42(3): 432–441.
- Huang, J. and Redmann, R. E. 1995. Salt tolerance of *Hordeum* and *Brassica* species during germination and early seedling growth. *Canadian Journal of Plant Science* 75(4): 815–819.
- Hu, Y. and Schmidhalter, U. 1998. Spatial distributions and net deposition rates of mineral elements in the elongating wheat (*Triticum aestivum* L.) leaf under saline soil conditions. *Planta* 204(2): 212–219.
- Humphreys, D. G. and Knox, R. E. 2015. Doubled haploid breeding in cereals (PP. 241–290). In *Advances in Plant Breeding Strategies: Breeding, Biotechnology and Molecular Tools*. Springer.
- Hussain, B., Khan, M. A., Ali, Q. and Shaukat, S. 2012. Double haploid production is the best method for genetic improvement and genetic studies of wheat. *International Journal for Agro Veterinary and Medical Sciences* 6(4): 216–228.
- Islam, S. M. S. and Tuteja, N. 2012. Enhancement of androgenesis by abiotic stress and other pretreatments in major crop species. *Plant Science* 182: 134–144.
- Ismail, A. M. 2003. Effect of salinity on the physiological responses of selected lines/variety of wheat. *Acta Agronomica Hungarica* 51(1): 1–9.
- Iyamabo, O. E. and M. Hayes, P. 1995. Effects of selection and opportunities for recombination in doubled-haploid populations of barley (*Hordeum vulgare* L.). *Plant Breeding* 114(2): 131–136.
- Jafar, M. Z., Farooq, M., Cheema, M. A., Afzal, I., Basra, S. M. A., Wahid, M. A., Aziz, T. and Shahid, M. 2012. Improving the performance of wheat by seed priming under saline conditions. *Journal of Agronomy and Crop Science* 198(1): 38–45.
- Jajoo, A. 2013. Changes in photosystem II in response to salt stress (PP.149–168). In *Ecophysiology and Responses of Plants under Salt Stress*. Springer.
- James, R. A., Blake, C., Byrt, C. S. and Munns, R. 2011. Major genes for Na⁺ exclusion, Nax1 and Nax2 (wheat HKT1; 4 and HKT1; 5), decrease Na⁺ accumulation in bread wheat leaves under saline and waterlogged conditions. *Journal of Experimental Botany* 62(8): 2939–2947.
- James, R. A., Von Caemmerer, S., Condon, A. G. T., Zwart, A. B. and Munns, R. 2008. Genetic variation in tolerance to the osmotic stress component of salinity stress in durum wheat. *Functional Plant Biology* 35(2): 111–123.
- Jaradat, A. A. 2003. Agriculture in Iraq: Resources, potentials, constraints, and

- research needs and priorities. *Food, Agriculture and Environment* 1(2): 160–166.
- Jaradat, A. A. 2013. Wheat landraces: a mini review. *Emirates Journal of Food and Agriculture* 25(1): 20.
- Jat, N. K. and Sharma, V. 2006. The interactive effect of salinity and PGR on certain biochemical parameters in wheat seedlings. *American Journal of Plant Physiology* 1(2): 132–141.
- Jauhar, P. P., Almouslem, A. B., Peterson, T. S. and Joppa, L.R. 1999. Inter and intragenomic chromosome pairing in haploids of durum wheat. *Journal of Heredity* 90(4): 437–445.
- Jeschke, W. D., Aslam, Z. and Greenway, H. 1986. Effects of NaCl on ion relations and carbohydrate status of roots and on osmotic regulation of roots and shoots of *Atriplex amnicola*. *Plant, Cell and Environment* 9(7): 559–569.
- Jouyban, Z. 2012. The effects of salt stress on plant growth. *Technical Journal of Engineering and Applied Sciences* 2(1): 7–10.
- Kanai, M., Higuchi, K., Hagihara, T., Konishi, T., Ishii, T., Fujita, N., Nakamura, Y., Maeda, Y., Yoshiba, M. and Tadano, T. 2007. Common reed produces starch granules at the shoot base in response to salt stress. *New Phytologist* 176(3): 572–580.
- Kasha, K. J., Hu, T. C., Oro, R., Simion, E. and Shim, Y. S. 2001. Nuclear fusion leads to chromosome doubling during mannitol pretreatment of barley (*Hordeum vulgare* L.) microspores. *Journal of Experimental Botany* 52(359): 1227–1238.
- Kaushal, L., Balachandran, S. M., Ulaganathan, K., Singh, A. K., Priyadarshi, R. and Shenoy, V. 2015. Auxin to improve green plant regeneration of rice anther culture. *International Journal of Agriculture and Crop Sciences* 8(1): 15.
- Kayani, S. A. and Gul S., 2000. Combined effects of temperature and salinity stress on corn cv. Sunahry. *Pakistan Journal of Biological Sciences (Pakistan)* 3(9): 1459–1463.
- Kayani, S. A., Naqvi, H. H. and Ting, I. P. 1990. Salinity effects on germination and mobilization of reserves in jojoba seed. *Crop Science* 30(3): 704–708.
- Kayani, S. A. and Rahman, M. 1987. Salt tolerance in corn (*Zea Mays* L) at the germination stage. *Pakistan Journal of Botany* 19(1): 9–15.
- Kedzony, M., Forster, B. P., Żur, I., Golemiec, E., Szechyńska-Hebda, M., Dubas, E. and Gotkebiowska, G. 2009. *Progress in doubled haploid technology in higher plants* (pp. 1–33). In *Advances in Haploid Production in Higher Plants*. Springer.
- Kerber, E. R. and Rowland, G. G. 1974. Origin of the free threshing character in hexaploid wheat. *Canadian Journal of Genetics and Cytology* 16(1): 145–154.

- Khaled, A. B., Hayek, T., Mansour, E. and Ferchichi, A. 2016. Comparative effect of salinity on ion accumulation, grain yield and stability salt tolerance degree of barley (*Hordeum vulgare* L.) in the different growth stage. *Journal of Agriculture and Environment for International Development (JAEID)* 110(2): 249–266.
- Khan, M. A., Shirazi, M. U., Mahboob, W., Mujtaba, S. M., Khan, M. A., Mumtaz, S. and Shereen, A. 2014. Morpho-physiological adaptations of wheat genotypes to salinity stress. *Pakistan Journal of Botany* 46(6): 1981–1985.
- Khatun, M., Hafiz, M. H. R., Hasan, M. A., Hakim, M. A. and Siddiqui, M. N. 2013. Responses of wheat genotypes to salt stress in relation to germination and seedling growth. *International Journal of Bio-resource and Stress Management* 4(4): 635–640.
- Khatun, R., Islam, S. M., Ara, I., Tuteja, N. and Bari, M. A. 2012. Effect of cold pretreatment and different media in improving anther culture response in rice (*Oryza sativa* L.) in Bangladesh. *Indian Journal of Biotechnology* 11: 458–463.
- Khayatnezhad, Zaefizadeh, M., M., Gholamin, R. and Jamaatiesomarin, S. 2010. Study of genetic diversity and path analysis for yield in durum wheat genotypes under water and dry conditions. *World Applied Sciences Journal* 9(6): 655–665.
- Kiani, A. R. and Mirlatifi, S. M. 2012. Effect of different quantities of supplemental irrigation and its salinity on yield and water use of winter wheat (*Triticum Aestivum*). *Irrigation and Drainage* 61(1): 89–98.
- Kilic, H. and Yagbasanlar, T. 2010. The effect of drought stress on grain yield, yield components and some quality traits of durum wheat (*Triticum turgidum* ssp. durum) cultivars. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 38(1): 164.
- King, I. P., Forster, B. P., Law, C. C., Cant, K. A., Orford, S. E., Gorham, J., Reader, S. and Miller, T.E. 1997. Introgression of salt tolerance genes from *Thinopyrum bessarabicum* into wheat. *New Phytologist* 137(1): 75–81.
- Koyro, H. W. 2006. Effect of salinity on growth, photosynthesis, water relations and solute composition of the potential cash crop halophyte *Plantago coronopus* (L.). *Environmental and Experimental Botany* 56(2): 136–146.
- Krasensky, J. and Jonak, C. 2012. Drought, salt, and temperature stress induced metabolic rearrangements and regulatory networks. *Journal of Experimental Botany* 63(4): 1593–1608.
- Kumari, A., Das, P., Parida, A. K. and Agarwal, P. K. 2015. Proteomics, metabolomics, and ionomics perspectives of salinity tolerance in halophytes. *Frontiers in Plant Science* 6(537): 1–20.

- Kwon, S. H. and Torrie, J. H. 1964. Heritability and interrelationship among traits of two soybean populations. *Crop Science* 4(2): 196–198.
- Labrani, Z., De Buyser, J. and Picard, E. 2007. Effect of mannitol pretreatment to improve green plant regeneration on isolated microspore culture in *Triticum turgidum* ssp. durum cv. Jennah Khetifa. *Plant Breeding* 126(6): 565–568.
- Lantos, C., Jenes, B., Bóna, L., Cserhádi, M. and Pauk, J. 2016. High frequency of doubled haploid plant production in spelt wheat. *Acta Biologica Cracoviensia Series Botanica* 58(2): 107–112.
- Lantos, C., Páricsi, S., Zofajova, A., Weyen, J. and Pauk, J. 2006. Isolated microspore culture of wheat (*Triticum aestivum* L.) with Hungarian cultivars. *Acta Biologica Szegediensis* 50(1–2): 31–35.
- Lantos, C., Weyen, J., Orsini, J. M., Gnad, H., Schlieter, B., Lein, V., Kontowski, S., Jacobi, A., Mihaly, R. and Broughton, S. 2013. Efficient application of *in vitro* anther culture for different European winter wheat (*Triticum aestivum* L.) breeding programmes. *Plant Breeding* 132(2): 149–154.
- Läuchli, A. and Grattan, S. R. 2007. *Plant growth and development under salinity stress* (pp. 1–32). In *Advances in Molecular Breeding Toward Drought and Salt Tolerant Crops*. Springer.
- Lauteri, M., Haworth, M., Serraj, R., Monteverdi, M.C. and Centritto, M. 2014. Photosynthetic diffusional constraints affect yield in drought stressed rice cultivars during flowering. *PloS One* 9(10): e109054.
- Lazaridou, T. B., Lithourgidis, A. S., Kotzamanidis, S. T. and Roupakias, D. G. 2005. Anther culture response of barley genotypes to cold pretreatments and culture media. *Russian Journal of Plant Physiology* 52(5): 696–699.
- Lazaridou, T., Pankou, C., Xynias, I. and Roupakias, D. 2016. Effect of D genome on wheat anther culture response after cold and mannitol pretreatment. *Acta Biologica Cracoviensia s. Botanica* 58(1): 95–102.
- Li, J., Ye, X., An, B., Du, L. and Xu, H. 2012. Genetic transformation of wheat: current status and future prospects. *Plant Biotechnology Reports* 6(3): 183–193.
- Lin, J., Li, X., Zhang, Z., Yu, X., Gao, Z., Wang, Y., Wang, J., Li, Z. and Mu, C. 2012. Salinity-alkalinity tolerance in wheat: Seed germination, early seedling growth, ion relations and solute accumulation. *African Journal of Agricultural Research* 7(3): 467–474.
- Liu, W., Zheng, M.Y., Polle, E. A. and Konzak, C. F. 2002. Highly efficient doubled haploid production in wheat (L.) via induced microspore embryogenesis. *Crop Science* 42(3): 686–692.
- Long, S. P., Marshallcolon, A. and Zhu, X. G. 2015. Meeting the global food demand of the future by engineering crop photosynthesis and yield potential. *Cell* 161(1): 56–66.

- Luo, M. C., Yang, Z. L. and Dvovrák, J. 2000. The Q locus of Iranian and European spelt wheat. *Theoretical and Applied Genetics* 100(3–4): 602–606.
- Ma'arup, R., Aziz, M. A. and Osman, M. 2012. Development of a procedure for the production of haploid plants through microspore culture of roselle (*Hibiscus sabdariffa* L.). *Scientia Horticulturae* 145: 52–61.
- Maas, E. V. and Grieve, C. M. 1990. Spike and leaf development of salad stressed wheat. *Crop Science* 30(6): 1309–1313.
- Maas, E. V. and Hoffman, G.J. 1977. Crop salt tolerance-current assessment. *Journal of the Irrigation and Drainage Division* 103(2): 115–134.
- Maathuis, F. J. M., Ahmad, I. and Patishtan, J. 2014. Regulation of Na⁺ fluxes in plants. *Frontiers in Plant Science* 5: 1–9.
- Maheswary, V. and Mak, C. 1993. The influence of genotypes and environments on induction of pollen plants for anther culture of *Capsicum annuum* L. *Asia-Pacific Journal of Molecular Biology and Biotechnology* 1(1): 43–50.
- Mahmood, A. and Baenziger, P. 2008. Creation of salt tolerant wheat doubled haploid lines from wheat×maize crosses. *Cereal Research Communications* 36(3): 361–371.
- Mahmood, I., Razzaq, A., Khan, Z. U. D., Hafiz, I. A. and Kaleem, S. 2012. Evaluation of tissue culture responses of promising wheat (*Triticum aestivum* L.) cultivars and development of efficient regeneration system. *Pakistan Journal of Botony* 44(1): 277–284.
- Makowska, K. and Oleszczuk, S. 2014. Albinism in barley androgenesis. *Plant Cell Reports* 33(3): 385–392.
- Maluszynski, M. 2003. *Doubled haploid production in crop plants: a manual*. Springer Science and Business Media.
- Maluszynski, M., Szarejko, I., Barriga, P. and Balcerzyk, A. 2002. Heterosis in crop mutant crosses and production of high yielding lines using doubled haploid systems (PP. 55–73). In *Mutations, In Vitro and Molecular Techniques for Environmentally Sustainable Crop Improvement*. Springer.
- Mansouri F., Sonia, Kobaissi, A. and Nziengui, H. 2005. In vitro gynogenesis in some varieties of durum wheat of Maghreb and Middle East (*Triticum durum* L.) as a tool for regenerating plants from salt tolerant callus lines. *International journal of tropical geology, geography and ecology* 29(29): 77–88
- Marassi, M. A., Bovo, O. A., Lavia, G. L. and Mroginski, L. A. 1993. Regeneration of rice double haploids using a onestep culture procedure. *Journal of Plant Physiology* 141(5): 610–614.
- Marcussen, T., Sandve, S. R., Heier, L., Spannagl, M., Pfeifer, M., Jakobsen, K. S., H. Wulff, B. B., Steuernagel, B., Mayer, K. F. X. and Olsen, O. A. 2014. Ancient hybridizations among the ancestral genomes of bread wheat. *Science*

345(6194): 1250092.

- Masood, S. A., Ahmad, S., Kashif, M. and Ali, Q. 2014a. Role of combining the ability to develop higher yielding wheat (*Triticum aestivum* L.) genotypes: An overview. *Natural Science* 12(11): 155–161.
- Masood, S. A., Ahmad, S., Kashif, M. and Ali, Q. 2014b. Correlation analysis for grain and its contributing traits in wheat (*Triticum aestivum* L.). *Natural Science* 12(11): 168–176.
- Masood, S. A., Ali, Q. and Abbas, H. 2014c. Estimation of general and specific combining ability for grain yield traits in *Triticum aestivum*. *Natural Science* 12(11): 191–198.
- Matsuoka, Y. 2011. Evolution of polyploid *Triticum* wheats under cultivation: the role of domestication, natural hybridization and allopolyploid speciation in their diversification. *Plant and Cell Physiology* 52(5): 750–764.
- Meiri, A. and Poljakoff-Mayber, A. 1970. Effect of various salinity regimes on growth, leaf expansion and transpiration rate of bean plants. *Soil Science* 109(1): 26–34.
- Mer, R. K., Prajith, P. K., Pandya, D. H. and Pandey, A. N. 2000. Effect of salts on germination of seeds and growth of young plants of *Hordeum vulgare*, *Triticum aestivum*, *Cicer arietinum* and *Brassica juncea*. *Journal of Agronomy and Crop Science* 185(4): 209–217.
- Mhaimed, A. S. 2013. Soil Resources of Iraq. *Soil Resources of Mediterranean and Caucasus Countries*: 98.
- Mirzaei, M., Soltani, N., Sarhadi, E., Pascovici, D., Keighley, T., Salekdeh, G. H., Haynes, P. A. and Atwell, B. J. 2011. Shotgun proteomic analysis of long-distance drought signaling in rice roots. *Journal of Proteome Research* 11(1): 348–358.
- Mishra, V. K. and Gowswami, R. 2014. Haploid production in the higher plant. *International Journal of Chemical and Biological Sciences* 1: 26–45.
- Mochida, K., Tsujimoto, H. and Sasakuma, T. 2004. Confocal analysis of chromosome behavior in wheat × maize zygotes. *Genome* 47(1): 199–205.
- Modhej, A., Naderi, A., Emam, Y., Aynehband, A. and Normohamadi, G. 2012. Effects of post-anthesis heat stress and nitrogen levels on grain yield in wheat (*T. durum* and *T. aestivum*) genotypes. *International Journal of Plant Production* 2(3): 257–268.
- Mohammed, L. S. 2013. Comparison of salt tolerance among some bread and durum wheat cultivars in germination and seedling development stages. *Journal of Tikrit University for Agricultural Sciences* 13(1): 135–142.
- Mohiuddin, A. K. M., Sultana, S. and Ferdous, J. 2006. Increased regeneration efficiency in seed derived callus of rice (*Oryza sativa* L.). *Plant Tissue Culture*

and Biotechnology 16(1): 45–52.

- Moien, A. and Sarrafiu, A. 1995. Genetic analysis for haploid regenerations responses of the hexaploid wheat anther cultures. *Plant Breeding* 114(3): 247–249.
- Monneveux, P., Jing, R. and Misra, S. C. 2012. Phenotyping for drought adaptation in wheat using physiological traits. *Frontiers in Physiology* 3(429): 1–12.
- Mühling, K. H. and Läuchli, A. 2002. Effect of salt stress on growth and cation compartmentation in leaves of two plant species differing in salt tolerance. *Journal of Plant Physiology* 159(2): 137–146.
- Munns, R. 1993. Physiological processes limiting plant growth in saline soils: some dogmas and hypotheses. *Plant, Cell and Environment* 16(1): 15–24.
- Munns, R. 2002. Comparative physiology of salt and water stress. *Plant, Cell and Environment* 25(2): 239–250.
- Munns, R. 2005. Genes and salt tolerance: bringing them together. *New Phytologist* 167(3): 645–663.
- Munns, R. 2006. Approaches to increasing the salt tolerance of wheat and other cereals. *Journal of Experimental Botany* 57(5): 1025–1043.
- Munns, R. 2011. Plant adaptations to salt and water stress: differences and commonalities. *Advances in Botanical Research* 57: 1–32.
- Munns, R. and Gilliham, M. 2015. Salinity tolerance of crops-what is the cost? *New Phytologist* 208(3): 668–673.
- Munns, R., Guo, J., Passioura, J. B. and Cramer, G. R. 2000. Leaf water status controls day-time but not daily rates of leaf expansion in salt-treated barley. *Functional Plant Biology* 27(10): 949–957.
- Munns, R., Hare, R. A., James, R. A. and Rebetzke, G. J. 1999. Genetic variation for improving the salt tolerance of durum wheat. *Crop and Pasture Science* 51(1): 69–74.
- Munns, R. and James, R. A. 2003. Screening methods for salinity tolerance: a case study with tetraploid wheat. *Plant and Soil* 253(1): 201–218.
- Munns, R., James, R. A., Gilliham, M., Flowers, T. J. and Colmer, T. D. 2016. Tissue tolerance: an essential but elusive trait for salt-tolerant crops. *Functional Plant Biology* 43(12): 1103–1113.
- Munns, R., R. James, A., Xu, B., Athman, A., Conn, S. J., Jordans, C., Byrt, C.S., Hare, R. A., Tyerman, S. D. and Tester, M. 2012. Wheat grain yield on saline soils is improved by an ancestral Na⁺ transporter gene. *Nature Biotechnology* 30(4): 360–364.
- Munns, R. and Rawson, H. M. 1999. Effect of salinity on salt accumulation and reproductive development in the apical meristem of wheat and barley.

- Functional Plant Biology* 26(5): 459–464.
- Munns, R., Schachtman, D. P. and Condon, A. G. 1995. The significance of a two phase growth response to salinity in wheat and barley. *Functional Plant Biology* 22(4): 561–569.
- Munns, R. and Termaat, A. 1986. Whole-plant responses to salinity. *Functional Plant Biology* 13(1): 143–160.
- Munns, R. and Tester, M. 2008. Mechanisms of salinity tolerance. *Annual Review of Plant Biology* 59: 651–681.
- Murashige, T. and Skoog, F. 1962. A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum* 15(3): 473–497.
- Muscolo, A., Sidari, M., Panuccio, M.R., Santonoceto, C., Orsini, F. and Depascale, S. 2011. Plant responses in saline and arid environments: an overview. Proceedings of the European cost action FAO901. *European Journal of Plant Science and Biotechnology* 5: 1–11.
- My, Z., Liu, W., Weng, Y., Polle, E. and Konzak, C. F. 2013. Production of doubled haploids in wheat (*Triticum aestivum* L.) through microspore embryogenesis triggered by inducer chemicals. *Doubled Haploid Production in Crop Plants: A Manual*: 83–94.
- Nandal, M. and Hooda, R. 2013. Salt tolerance and physiological response of plants to salinity: A Review. *International Journal of Scientific and Engineering Research* 4(10): 44–67.
- Nielsen, N. H., Andersen, S. U., Stougaard, J., Jensen, A., Backes, G. and Jahoor, A. 2015. Chromosomal regions associated with the in vitro culture response of wheat (*Triticum aestivum* L.) microspores. *Plant Breeding* 134(3): 255–263.
- Niemirowicz-Szczytt, K. 1997. Excessive homozygosity in doubled haploids. advantages and disadvantages for plant breeding and fundamental research. *Acta Physiologiae Plantarum* 19(2): 155–167.
- Niu, Z., Jiang, A., Abu Hammad, W., Oladzadabbasabadi, A., Xu, S. S., Mergoum, M. and Elias, E. M. 2014. Review of doubled haploid production in durum and common wheat through wheat×maize hybridization. *Plant Breeding* 133(3): 313–320.
- Nouri, H., Borujeni, S.C., Nirola, R., Hassanli, A., Beecham, S., Alaghmand, S., Saint, C. and Mulcahy, D. 2017. Application of green remediation on soil salinity treatment; a review on halophytoremediation. *Process Safety and Environmental Protection* 107: 94-107 .
- Obata, T. and Fernie, A. R. 2012. The use of metabolomics to dissect plant responses to abiotic stresses. *Cellular and Molecular Life Sciences* 69(19): 3225–3243.
- Orshinsky, B. R. and Sadasivaiah, R. S. 1997. Effect of plant growth conditions, plating density, and genotype on the anther culture response of soft white

- spring wheat hybrids. *Plant Cell Reports* 16(11): 758–762.
- Osei, M. K., Asante, M. D., Agyeman, A., Adebayo, M. A. and Adu-Dapaah, H. 2014. Plant breeding: A tool for achieving food sufficiency (PP. 253–274). In *Sustainable Horticultural Systems*. Springer.
- Oyiga, B. C., Sharma, R. C., Baum, M., Ogbonnaya, F. C., Leon, J. and Ballvora, A. 2017. Allelic variations and differential expressions detected at quantitative trait loci for salt stress tolerance in wheat. *Plant, Cell and Environment*: 1–17.
- Ozhan, N. and Hajibabaei, M. 2015. Studies on germination properties of spring wheat with treatment by poly amines under salinity stress. *International Journal of Advanced Biological and Biomedical Research* 3(1): 19–23.
- Pandey, A. N. and Thakarar, N. K. 1997. Effect of chloride salinity on survival and growth of prosopis chilensis seedlings. *Tropical Ecology* 38(1): 145–148.
- Parihar, P., Singh, S., Singh, R., Singh, V. P. and Prasad, S. M. 2015. Effect of salinity stress on plants and its tolerance strategies: a review. *Environmental Science and Pollution Research* 22(6): 4056–4075.
- Pauk, J., R. Mihály and Puolimatka, M. 2003. Protocol for wheat (*Triticum aestivum* L.) anther culture (PP. 59–64). In *Doubled Haploid Production in Crop Plants*. Springer.
- Pearce, S. C. and Moore, C. S. 1976. Reduction of experimental error in perennial crops, using adjustment by neighbouring plots [tea, peaches, pineapples, England]. *Experimental Agriculture* (UK) 12(3): 267–272.
- Perveen, S., Shahbaz, M. and Ashraf, M. 2013. Influence of foliar applied triacontanol on growth, gas exchange characteristics, and chlorophyll fluorescence at different growth stages in wheat under saline conditions. *Photosynthetica* 51(4): 541–551.
- Picard, E. and Debuyser, J. 1973. Obtainment of haploid plantlets of *Triticum aestivum* L. from anther cultures *in vitro*. [Wheat]. *Sciences Naturelles*: 1463–1466.
- Pirasteh-Anosheh, H., Ranjbar, G., Pakniyat, H. and Emam, Y. 2016. Physiological mechanisms of salt stress tolerance in plants: An overview. Plant environment interaction: *Responses and Approaches to Mitigate Stress* 1(1): 141–160.
- Platten, J. D., Egdane, J.A. and Ismail, A. M. 2013. Salinity tolerance, Na⁺ exclusion and allele mining of HKT1; 5 in *Oryza sativa* and *O. glaberrima*: many sources, many genes, one mechanism? *BMC Plant Biology* 13(1): 32.
- Poland, J. A., Brown, P. J., Sorrells, M. E. and Jannink, J. L. 2012. Development of high density genetic maps for barley and wheat using a novel two enzyme genotyping by sequencing approach. *PloS One* 7(2): e32253.
- Poustini, K. and Siosemardeh, A. 2004. Ion distribution in wheat cultivars in response to salinity stress. *Field Crops Research* 85(2): 125–133.

- Pratap, A., Sethi, G. S. and Chaudhary, H. K. 2006. Relative efficiency of anther culture and chromosome elimination techniques for haploid induction in triticale×wheat and triticale×triticale hybrids. *Euphytica* 150(3): 339–345.
- Prigge, V. and Melchinger, A. E. 2012. Production of haploids and doubled haploids in maize. *Plant Cell Culture Protocols* 877: 161–172.
- Pugnaire, F. I., Serrano, L. and Pardos, J. 1999. Constraints by water stress on plant growth. *Handbook of Plant and Crop Stress 2*: 271–283.
- Radi, A. A., Farghaly, F. A., and Hamada, A. M. 2013. Physiological and biochemical responses of salt tolerant and salt sensitive wheat and bean cultivars to salinity. *Journal of Biology and Earth Sciences* 3(1): 72–88.
- Rahdari, P. and Hoseini, S. M. 2015. Evaluation of germination percentage and some physiologic factors under salinity stress and gibberellic acid hormone (GA3) treatments in wheat (*Triticum aestivum* L.). *The AYER* 2: 395–405.
- Rahman, M., Soomro, U. A., Haq, M. Z. and Gul, S. 2008. Effects of NaCl salinity on wheat (*Triticum aestivum* L.) cultivars. *World Journal of Agricultural Sciences* 4(3): 398–403.
- Rajendran, K., Tester, M. and Roy, S. J. 2009. Quantifying the three main components of salinity tolerance in cereals. *Plant, cell and environment* 32(3): 237–249.
- Rakha, M. T., Metwally, E. I., Moustafa, S. A., Etman, A. A. and Dewir, Y. H. 2012. Evaluation of regenerated strains from six 'Cucurbita' interspecific hybrids obtained through anther and ovule 'in vitro' cultures. *Australian Journal of Crop Science* 6(1): 23.
- Ramirez, J. I. S. and Maiti, R. 2016. Research trends in abiotic stress resistance of crops (PP. 131–163). In *Bioresource and Stress Management*. Springer.
- Rao, A., Ahmad, S. D., Sabir, S. M., Awan, S. I., Shah, A. H., Abbas, S. R., Shafique, S., Khan, F. and Chaudhary, A. 2013. Potential antioxidant activities improve salt tolerance in ten varieties of wheat (*Triticum aestivum* L.). *American Journal of Plant Sciences* 4(6): 69.
- Rauf, M. 2005. *Screening of wheat genotypes for better germination ability under low moisture levels in the laboratory*. M. Sc.(Hons) Dissertation, University of Arid Agriculture, Rawalpindi, Pakistan.
- Rawlins, S. L. 2012. Principles of salinity control in irrigated agriculture. *Advances in Food-Producing Systems For Arid and Semiarid Lands* 1: 391.
- Rawson, H. M., Long, M. J. and Munns, R. 1988. Growth and development in NaCl treated plants. I. Leaf Na⁺ and Cl⁻ concentrations do not determine gas exchange of leaf blades in barley. *Functional Plant Biology* 15(4): 519–527.
- Ray, D. K., Mueller, N. D., West, P. C. and Foley, J. A. 2013. Yield trends are insufficient to double global crop production by 2050. *PloS One* 8(6): e66428.

- Rengasamy, P. 2002. Transient salinity and subsoil constraints to dryland farming in Australian sodic soils: an overview. *Animal Production Science* 42(3): 351–361.
- Rodriguez, H. G., Roberts, J. K.M., Jordan, W. R. and Drew, M. C. 1997. Growth, water relations, and accumulation of organic and inorganic solutes in roots of maize seedlings during salt stress. *Plant Physiology* 113(3): 881–893.
- Romero-Aranda, M. R., Jurado, O. and Cuartero, J. 2006. Silicon alleviates the deleterious salt effect on tomato plant growth by improving plant water status. *Journal of Plant Physiology* 163(8): 847–855.
- Rosegrant and Cline. 2003. Global food security: challenges and opportunities. *Science* 302: 1917–1919.
- Rout, P., Naik, N., Ngangkham, U., Verma, R. L., Katara, J. L., Singh, O. N. and Samantaray, S. 2016. Doubled haploids generated through anther culture from an elite long duration rice hybrid, CRHR32: Method optimization and molecular characterization. *Plant Biotechnology* 33(3): 177–186.
- Roy, S. J., Negrão, S. and Tester, M. 2014. Salt resistant crop plants. *Current Opinion in Biotechnology* 26: 115–124.
- Roychoudhury, A., and M. Chakraborty. 2013. Biochemical and molecular basis of the varietal difference in plant salt tolerance. *Annual Review and Research in Biology* 3(4): 422–454.
- Royo, A. and Abiό, D. 2003. Salt tolerance in durum wheat cultivars. *Spanish Journal of Agricultural Research* 1(3): 27–35.
- Royo, A., Playán, E. and Ortiz, R. 2000. Salinity-grain yield response functions of barley cultivars assessed with a drip injection irrigation system. *Soil Science Society of America Journal* 64(1): 359–365.
- Rozema, J. and Schat, H. 2013. Salt tolerance of halophytes, research questions reviewed in the perspective of saline agriculture. *Environmental and Experimental Botany* 92: 83–95.
- Saboora, A., Kiarostami, K., Behroozbayati, F. and Hajhashemi, S. 2006. Salinity (NaCl) tolerance of wheat genotypes at germination and early seedling growth. *Pakistan Journal of Biological Sciences* 9(11): 2009–2021.
- Santra, M., Ankrah, N., Santra, D. K. and Kidwell, K. K. 2012. An improved wheat microspore culture technique for the production of doubled haploid plants. *Crop Science* 52(5): 2314–2320.
- Saqib, Z. A., Akhtar, J., Haq, M. A. and Ahmad, I. 2012a. Salt induced changes in leaf phenology of wheat plants are regulated by accumulation and distribution pattern of Na ion. *Pakistan Journal of Agricultural Sciences* 49: 141–148.
- Saqib, Z. A., Akhtar, J., Ulhaq, M. A., Ahmad, I. and Bakhat, H. F. 2012b. Rationality of using various physiological and yield related traits in determining salt

- tolerance in wheat. *African Journal of Biotechnology* 11(15): 3558–3568.
- Saulescu, N. N., Ittu, G., Giura, A., Mustatea, P. and Ittu, M. 2012. Results of using Zea method for doubled haploid production in wheat breeding at NARDI Fundulea Romania. *Romanian Agricultural Research* (29): 3–8.
- Scagliusi, S. M. 2014. Establishing isolated microspore culture to produce doubled haploid plants in Brazilian wheat (*Triticum aestivum* L.). *Australian Journal of Crop Science* 8(6): 887–894.
- Schachtman, D. P., R. Munns and Whitecross, M. I. 1991. Variation in sodium exclusion and salt tolerance in *Triticum tauschii*. *Crop Science* 31(4): 992–997.
- Setter, T. L., Waters, I., Stefanova, K., Munns, R. and Barrettlennard, E. G. 2016. Salt tolerance, date of flowering and rain affect the productivity of wheat and barley on rainfed saline land. *Field Crops Research* 194: 31–42.
- Shahzad, A., Ahmad, M., Iqbal, M., Ahmed, I. and Ali, G. M. 2012. Evaluation of wheat landrace genotypes for salinity tolerance at the vegetative stage by using morphological and molecular markers. *Genetics and Molecular Research* 11(1): 679–692.
- Shahzad, A., Iqbal, M., Asif, M., Hirani, A. H. and Goyal, A. 2013. Growing wheat on saline lands: Can a dream come true? *Australian Journal of Crop Science* 7(4): 515–524.
- Sharma, A. K. and Sharma, A. 2014. *Chromosome techniques: theory and practice*. Butterworth-Heinemann.
- Shiferaw, B., Smale, M., Braun, H. J., Duveiller, E., Reynolds, M. and Muricho, G. 2013. Crops that feed the world 10. Past successes and future challenges to the role played by wheat in global food security. *Food Security* 5(3): 291–317.
- Shirdelmoghanloo, H., Moieni, A. and Mousavi, A. 2009. Effects of embryo induction media and pretreatments in isolated microspore culture of hexaploid wheat (*Triticum aestivum* L. cv. Falat). *African Journal of Biotechnology* 8(22): 6134–6140.
- Shrewry, P. R. 2001. *The world wheat book. A history of wheat breeding* (pp. 1131). Paris: Larosier Publishing and Andover: Intercept. (hardback).
- Singh, A. 2016. Varied responses and tolerant mechanisms towards salinity stress in Piant. *International Journal of Plant and Soil Science* 11(5): 1–13.
- Singh, U. M., Sareen, P., Sengar, R. S. and Kumar, A. 2013. Plant ionomics: a newer approach to study mineral transport and its regulation. *Acta Physiologiae Plantarum* 35(9): 2641–2653.
- Singla, R. and Garg, N. 2005. Influence of salinity on growth and yield attributes in chickpea cultivars. *Turkish journal of Agriculture and Forestry* 29(4): 231–235.

- Siti Aishah, H., Saberi, A. R., Halim, R.A. and Zanhara, A. R. 2010. Salinity effects on germination of forage sorghumes. *Journal of Agronomy* 9(4): 169–174.
- Ślusarkiewicz-Jarzina, A., Pudelska, H., Woźna, J. and Pniewski, T. 2017. Improved production of doubled haploids of winter and spring triticale hybrids via a combination of colchicine treatments on anthers and regenerated plants. *Journal of Applied Genetics*: 1–9.
- Smith, M. K. and Drew, R. A. 1990. Current applications of tissue culture in plant propagation and improvement. *Functional Plant Biology* 17(3): 267–289.
- Snape, J. W. and Simpson, E. 1981. The genetical expectations of doubled haploid lines derived from different filial generations. *Theoretical and Applied Genetics* 60(2): 123–128.
- Soriano, M., Li, H. and Boutilier, K. 2013. Microspore embryogenesis: Establishment of embryo identity and pattern in culture. *Plant Reproduction* 26(3): 181–196.
- Steel, R. G. D. and Torrie, J. H. 1980. Analysis of covariance. Principles and procedures of statistics: *A Biometrical Approach*: 401–437.
- Steppuhn, H., Van Genuchten, M. T. and Grieve, C. M. 2005. Root zone salinity. *Crop Science* 45(1): 221–232.
- Su, R. C., Rudert, M. L. and Hodges, T. K. 1992. Fertile indica and japonica rice plants regenerated from protoplasts isolated from embryogenic haploid suspension cultures. *Plant Cell Reports* 12(1): 45–49.
- Subira, J., Álvaro, F., Dlmoral, L. F. G. and Royo, C. 2015. Breeding effects on the cultivar×times×environment interaction of durum wheat yield. *European Journal of Agronomy* 68: 78–88.
- Szopko, D., Darko, E., Molnar, I., Kruppa, K., Halo, B., Vojtko, A., Molnar-Lang, M. and Dulai, S. 2017. Photosynthetic responses of a wheat (Asakaze) barley (Manas) 7H addition line to salt stress. *Photosynthetica* 55(2): 317–328.
- Tadesse, W., Inagaki, M., Tawkaz, S., Baum, M. and Van Ginkel, M. 2012. Recent advances and application of doubled haploids in wheat breeding. *African Journal of Biotechnology* 11(89): 15484–15492.
- Tadesse, W., Tawkaz, S., Inagaki, M. N., Picard, E. and Baum, M. 2013. Methods and applications of doubled haploid technology in wheat breeding. *A Technical Manual. Tools and Guidelines*: 36.
- Tarchoune, I., Deglinnocenti, E., Kaddour, R., Guidi, L., Lachaâl, M., Navarilzzo, F. and Ouerghi, Z. 2012. Effects of NaCl or Na₂SO₄ salinity on plant growth, ion content and photosynthetic activity in *Ocimum basilicum* L. *Acta Physiologiae Plantarum* 34(2): 607–615.
- Termaat, A., Passioura, J. B. and Munns, R. 1985. Shoot turgor does not limit shoot growth of NaCl affected wheat and barley. *Plant Physiology* 77(4): 869–872.

- Tezara, W., Martinez, D., Rengifo, E. and Herrera, A. N. A. 2003. Photosynthetic responses of the tropical spiny shrub *Lycium nodosum* (Solanaceae) to drought, soil salinity and saline spray. *Annals of Botany* 92(6): 757–765.
- Touraev, A., Indrianto, A., Wratschko, I., Vicente, O. and Heberle-Bors, E. 1996. Efficient microspore embryogenesis in wheat (*Triticum aestivum* L.) induced by starvation at high temperature. *Sexual Plant Reproduction* 9(4): 209–215.
- Troeh, F. R. and Thompson, L. M. 2005. *Soils and soil fertility*. Blackwell Iowa.
- Trottier, M. C., Collin, J. and Comeau, A. 1993. Comparison of media for their aptitude in wheat anther culture. *Plant Cell, Tissue and Organic Culture* 35(1): 59–67.
- Tsuchiya, T. 1971. An improved acetocarmine squash method, with special reference to the modified Rattenbury's method of making a preparation permanent. *Barley Genetic Newsl:* 71–72.
- Turesson, S., Ljungberg, A., Johansson, N., Karlsson, K. E., Suijs, L.W. and Josset, J.P. 2000. Large scale production of wheat and triticale double haploids through the use of a single anther culture method. *Plant Breeding* 119(6): 455–459.
- Wajid, A., Hussain, A., Ahmed, A., Rafiq, M., Goheer, A. R. and Ibrahim, M. 2004. Effect of sowing date and plant density on growth, light interception and yield of wheat under semi arid condition. *International Journal of Agriculture and Biology* 6: 1119–1123.
- Wan, Y., Petolino, J. F. and Widholm, J. M. 1989. Efficient production of doubled haploid plants through colchicine treatment of anther derived maize callus. *Theoretical and Applied Genetics* 77(6): 889–892.
- Wang, M., Zheng, Q., Shen, Q. and Guo, S. 2013. The critical role of potassium in plant stress response. *International journal of Molecular Sciences* 14(4): 7370–7390.
- Wilkinson, P. A., Winfield, M. O., Barker, G. L. A., Allen, A. M., Burridge, A., Coghill, J. A. and Edwards, K. J. 2012. Cereals DB 2.0: an integrated resource for plant breeders and scientists. *BMC Bioinformatics* 13(1): 219.
- William, M., Langridge, P., Trethowan, R., Dreisigacker, S. and Crouch, J. 2008. Genomics of wheat, the basis of our daily bread (PP. 515–548). In *Genomics of Tropical Crop Plants*. Springer.
- Wu, W., Al-Shafie, W. M., Mhaimed, A. S., Ziadat, F., Nangia, V. and Payne, W. B. 2014. Soil salinity mapping by multiscale remote sensing in Mesopotamia, Iraq. *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing* 7(11): 4442–4452.
- Wu, X., Chang, X. and Jing, R. 2012. Genetic insight into yield associated traits of wheat grown in multiple rain-fed environments. *PloS One* 7(2): e31249.

- Wu, H., Shabala, L., Liu, X., Azzarello, E., Zhou, M., Pandolfi, C., Chen, Z. H., Bose, J., Mancuso, S. and Shabala, S. 2015. Linking salinity stress tolerance with tissue specific Na⁺ sequestration in wheat roots. *Frontiers in Plant Science* 6(71): 1–31.
- Xie, J. H., Gao, M. W., Liang, Z. Q., Shu, Q.Y., Cheng, X.Y. and Xue, Q. Z. 1997. The effect of cool-pretreatment on the isolated microspore culture and the free amino acid change of anthers in Japonica rice (*Oryza sativa* L). *Journal of Plant Physiology* 151(1): 79–82.
- Xuehua, W., Xiaoren, L. U. O. and Li, D. A. I. 2016. Research progress on the biological basis of cereal pollen culture. *Agricultural Science and Technology* 17(3).
- Yadav, S., Irfan, M., Ahmad, A. and Hayat, S. 2011. Causes of salinity and plant manifestations to salt stress: a review. *Journal of Environmental Biology* 32(5): 667.
- Yan, G., Liu, H., Wang, H., Lu, Z., Wang, Y., Mullan, D., Hamblin, J. and Liu, C. 2017. Accelerated generation of selfed pure line plants for gene identification and crop breeding. *Frontiers in Plant Science*. 8: 1-14.
- Yang, C., Zhao, L., Zhang, H., Yang, Z., Wang, H., Wen, S., Zhang, C., Rustgi, S., Von Wettstein, D. and Liu, B. 2014. Evolution of physiological responses to salt stress in hexaploid wheat. *Proceedings of the National Academy of Sciences* 111(32): 11882–11887.
- Yaycili, O. and Alikamanoglu, S. 2012. Induction of salt tolerant potato (*Solanum tuberosum* L.) mutants with gamma irradiation and characterization of genetic variations via RAPD-PCR analysis. *Turkish Journal of Biology* 36(4): 405–412.
- Yeo, A. R., Izard, P., Boursier, P. J. and Flowers, T. J. 1991. Short and long term effects of salinity on leaf growth in rice (*Oryza sativa* L.). *Journal of Experimental Botany* 42(7): 881–889.
- Yeticsir, H. and Uygur, V. 2009. Plant growth and mineral element content of different gourd species and watermelon under salinity stress. *Turkish Journal of Agriculture and Forestry* 33(1): 65–77.
- Yonezawa, K., Nomura, T. and Sasaki, Y. 1987. Conditions favouring doubled haploid breeding over conventional breeding of self fertilizing crops. *Euphytica* 36(2): 441–453.
- Younis, A., Hwang, Y. J. and Lim, K. B. 2014. Exploitation of induced 2n gametes for plant breeding. *Plant Cell Reportseports* 33(2): 215–223.
- Yu, S., Wang, W. and Wang, B. 2012. Recent progress of salinity tolerance research in plants. *Russian Journal of Genetics* 48(5): 497–505.
- Zafarnaderi, N., Aharizad, S. and Mohammadi, S. A. 2013. Relationship between grain yield and related agronomic traits in bread wheat recombinant inbred

- lines under water deficit condition. *Annals of Biological Research* 4(4): 7–11.
- Zaharieva, M. and Monneveux, P. 2014. Cultivated einkorn wheat (*Triticum monococcum* L. subsp. *monococcum*): the long life of a founder crop of agriculture. *Genetic Resources and Crop Evolution* 61(3): 677–706.
- Zapata, F. J., Alejar, M. S., Torrizo, L. B., Novero, A. U., Singh, V. P. and Senadhira, D. 1991. Field performance of anther culture derived lines from F₁ crosses of indica rices under saline and nonsaline conditions. *Theoretical and Applied Genetics* 83(1): 6–11.
- Zare, M., Mehrabi, O. A. A. and Sharafzadeh, S. H. 2007. Investigation of GA₃ and kinetin effects on seed germination and seedling growth of wheat under salinity stress. *Journal of Agricultural Science* 12(4): 855–865.
- Zhang, H., Bian, Y., Gou, X., Dong, Y., Rustgi, S., Zhang, B., Xu, C., Li, N., Qi, B. and Han, F. 2013. Intrinsic karyotype stability and gene copy number variations may have laid the foundation for tetraploid wheat formation. *Proceedings of the National Academy of Sciences* 110(48): 19466–19471.
- Zhao, X., Wang, W., Zhang, F., Deng, J., Li, Z. and Fu, B. 2014. Comparative metabolite profiling of two rice genotypes with contrasting salt stress tolerance at the seedling stage. *PloS One* 9(9): e108020.
- Zheng, M., Liu, W., Weng, Y., Polle, E. and Konzak, C. 2001. Culture of freshly isolated wheat (*Triticum aestivum* L.) microspores treated with inducer chemicals. *Plant Cell Reports* 20(8): 685–690.
- Zheng, M. Y., Liu, W., Weng, Y., Polle, E. and Konzak, C. F. 2003. Production of doubled haploids in wheat (*Triticum aestivum* L.) through microspore embryogenesis triggered by inducer chemicals (PP. 83–94). In *Doubled Haploid production in crop plants*. Springer.
- Zhou, G., Johnson, P., Ryan, P. R., Delhaize, E. and Zhou, M. 2012. Quantitative trait loci for salinity tolerance in barley (*Hordeum vulgare* L.). *Molecular Breeding* 29(2): 427–436.
- Zhu, G. Y., Kinet, J. M. and Lutts, S. 2004. Characterisation of rice (*Oryza sativa*) F₃ populations selected for salt resistance. 2. Relationships between yield-related parameters and physiological properties. *Animal Production Science* 44(3): 333–342.
- Zlatev, Z. and Lidon, F. C. 2012. An overview on drought induced changes in plant growth, water relations and photosynthesis. *Emirates Journal of Food and Agriculture* 24(1): 57.
- Zohary, D., Hopf, M. and Weiss, E. 2012. *Domestication of plants in the old world: The origin and spread of domesticated plants in southwest Asia, Europe, and the mediterranean basin*. Oxford University Press on Demand.

Zorinants, S., Tashpulatov, A. S., Heberle-Bors, E. and Touraev, A. 2005. The role of stress in the induction of haploid microspore embryogenesis (PP. 35–52). In *Haploids in Crop Improvement II*. Springer.

Zur, I., Dubas, E., Krzewska, M., Janowiak, F., Hura, K., Pocięcha, E., Bączek-Kwinta, R. and Płazek, A. 2014. Antioxidant activity and ROS tolerance in triticale (*×Triticosecale* Wittm.) anthers affect the efficiency of microspore embryogenesis. *Plant Cell Tiss Organ Cult.* 19(1): 79–94.



LIST OF PUBLICATIONS

Abbas Lateef Abdulrahman, Siti Nor Akmar Abdullah, Mohd Rafii Yusop, Mohd Razi Ismail and Mahdi Moradpour. 2016. Seed germination and seedling growth of hexaploid wheat (*Triticum aestivum* L.) varieties as influenced by different levels of sodium chloride. *Res. on Crops* 17 (3): 445-453 (2016).

Abbas Lateef Abdulrahman, Siti Nor Akmar Abdullah, Mohd Rafii Yusop and Mohd Razi Ismail. 2016. Role of wheat germplasm, anther pre-treatments, and medium on the production of doubled haploid plants. *Transactions of Persatuan Genetik of Malaysia*, No 3, pp. 113-122, 2016.





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT :

PERFORMANCE OF CROSSES AND DEVELOPMENT OF DOUBLED HAPLOID LINES IN WHEAT (*Triticum aestivum* L.) FOR SALT TOLERANCE

NAME OF STUDENT: ABBAS LATEEF ABDULRAHMAN

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (v)

- CONFIDENTIAL (Contain confidential information under Official Secret Act 1972).
RESTRICTED (Contains restricted information as specified by the organization/institution where research was done).
OPEN ACCESS I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT Embargo from _____ until _____ (date) (date)

(Signature of Student) New IC No/ Passport No.: A7509378

Approved by: PROF. DATIN DR. SITI NOR AKMAR ABDULLAH Pengarah Institut Kajian Perladangan Universiti Putra Malaysia (Signature of Chairman of Supervisory Committee) Name: 400 UPM Serdang, Selangor Darul Ehsan

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted.]