

TREATED CLINOPTILOLITE-MODIFIED GRAPHITE FELT  
BIOANODE MICROBIAL FUEL CELLS FOR POWER  
GENERATION AND DYE DECOLOURISATION

SEYEDEH NAZANIN KARDI

UNIVERSITI TEKNOLOGI MALAYSIA

TREATED CLINOPTILOLITE-MODIFIED GRAPHITE FELT  
BIOANODE MICROBIAL FUEL CELLS FOR POWER  
GENERATION AND DYE DECOLOURISATION

SEYEDEH NAZANIN KARDI

A thesis submitted in fulfilment of the  
requirements for the award of the degree of  
Doctor of Philosophy (Bioscience)

Faculty of Biosciences and Medical Engineering  
Universiti Teknologi Malaysia

NOVEMBER 2017

*Specially dedicated to my beloved Dad and Mom,  
Reza Kardi and Maria Hadighi*

*and*

*To my adorable husband*

*Nima*

## ACKNOWLEDGEMENT

Acknowledgments for the completion of this thesis must be extended to many people who provided me with precious time and invaluable advice. My gratitude to the Almighty God, due to all His blessings and grace, this thesis finally came to an end.

I wish to express my sincerest appreciations to Dr. Norahim Bin Ibrahim for his valuable comments, genuine encouragement, constructive advice, permanent support, and patient guidance during the formulation of this thesis. It is from his I understood the meaning of patient. Thank you for the opportunity you granted to me. I am also very thankful to my co-supervisor Prof. Dr. Nooraini Abdul Rashid for her solid support, continuing feedback, valuable advices, and motivation. Sincerest appreciation to my external co-supervisor Prof. Dr. Ghasem Najafpour for all his limitless encouragement, precious time and professional guidance which gave me the strength to complete my project.

My sincere gratitude also goes to all laboratory technicians in the Bionanotechnology, Nanomaterial, and Central laboratories for their genuine help in carrying out the laboratory tests and physical modeling works throughout the study.

Last but not least, my utmost appreciations go to my beloved parents for their eternal support, unconditional love, sacrifice, and encouragement. I am nothing without you both (Reza and Maria). My special thanks go to my adorable husband, Nima for all his support and tolerance throughout this research journey. Words really fail to appreciate his for everything.

## ABSTRACT

One important factor in microbial fuel cells (MFCs) study is the anode. In MFCs, the anode acts as the key component in the generation of bioelectricity and power. Despite the fact that there have been some improvements in the electrochemical performance of MFCs in recent years, their low power generation is still deemed a major drawback. The effects of surface modifications of the anode as biofilm carrier on the performance of MFCs were investigated. This research focused on the role of the novel fabricated anode as support material for the adhesion of bacterial consortium (NAR-2) consisted of *Citrobacter* sp. A1, *Enterobacter* sp. L17 and *Enterococcus* sp. C1 were used in MFCs reactor for the decolourisation of Acid Red 27 (AR27) and the simultaneous generation of electricity. The performance of a modified anode fabricated using surfactant-treated clinoptilolite (S-TC) with common type of carbon-based material, namely treated clinoptilolite-modified graphite felt (TC-MGF) anode was evaluated with different MFCs constructions. Prior to the MFCs experiments, the modification of anode was successfully verified using different spectroscopic and microscopic techniques such as EDX, FESEM, ATR-FTIR and BET analysis. In addition, screening of parameters for the adhesion of bacterial consortium NAR-2 onto TC-MGF anode (NAR-2-bioanode) was accomplished. The newly-developed TC-MGF bioanode was implemented in the dual-chamber (H-type) of the MFC. The performance of TC-MGF bioanode was compared to the results obtained using non-modified graphite felt (BGF) bioanode. Maximum power densities for BGF and TC-MGF bioanodes were  $458.8 \pm 5.0$  and  $940.3 \pm 4.2$   $\text{mWm}^{-2}$ , respectively. In the following experimental, a small MFC reactor was fabricated with TC-MGF bioanode to compare the performance of the MFC with commonly used fuel cell membranes, Nafion (N-117 and N-115), which were examined along with the N-212 membrane in a single-chamber cubic di-air cathode (S-CCD-AC) design. The power density and columbic efficiency of N-115 membrane ( $1022.5 \text{ mWm}^{-2}$  - 35.4%) were significantly higher than the values obtained for the N-117 ( $592 \text{ mWm}^{-2}$  - 15.6%) and N-212 ( $493 \text{ mWm}^{-2}$  - 12.3%) membranes. A novel MFC reactor with TC-MGF bioanode novel design (Conch shell) using the N-115 membrane having an air-cathode upflow (A-CU) MFC, as a combination of upflow and MFC technologies was used to compare the presence and absence of a membrane design. The A-CUMFC with membrane-less at flow rate  $0.6 \text{ mL min}^{-1}$ , anode distance of 0.5 cm and a concentration of AR27 at  $900 \text{ mg L}^{-1}$ , high decolourisation rate (98%) achieved in a 60-day operation, was 40% higher than that of the membrane-MFC. The average maximum power density obtained ( $1250 \text{ mWm}^{-2}$ ) using the membrane-less MFC was higher than that of the membrane-MFC ( $1108 \text{ mWm}^{-2}$ ) during the 80-day operation with TC-MGF bioanode.

## ABSTRAK

Satu faktor penting di bawah kajian sel bahan bakar mikrob (MFCs) ialah anod. Di MFC, anod bertindak sebagai komponen utama dalam penjanaan bio-elektrik dan kuasa. Walaupun terdapat beberapa peningkatan dalam prestasi elektrokimia MFC dalam beberapa tahun kebelakangan ini, penjanaan kuasa rendah mereka masih dianggap sebagai kelemahan utama. Kesan pengubahsuaian permukaan anod sebagai pembawa biofilem terhadap prestasi MFC telah dikaji. Kajian ini menumpukan kepada peranan anod yang baru dibuat bagi melekatkan filem mikrob sebagai bahan sokongan untuk menawarkan tapak pelekat konsortium bakteria (NAR-2) yang terdiri daripada *Citrobacter* sp. A1, *Enterobacter* sp. L17 dan *Enterococcus* sp. C1 digunakan dalam reaktor MFC, dari segi penyahbauan Acid Red 27 (AR27), dan penjanaan elektrik serentak. Prestasi anod yang diubahsuai yang dibuat menggunakan klinoptilolit (S-TC) yang dirawat dengan terapi surfaktan dengan bahan yang berasaskan karbon yang biasa, iaitu anoda grafit dan klinoptilolit yang diubahsuai (TC-MGF) dirawat dengan pembinaan MFC yang berbeza. Sebelum kajian MFC, pengubahsuaian anod berjaya disahkan menggunakan teknik spektroskopi dan mikroskopik yang berbeza seperti analisis EDX, FESEM, ATR-FTIR dan BET. Di samping itu, pemeriksaan parameter untuk pelekat konsortium bakteria NAR-2 ke anod TC-MGF (NAR-2-bioanode) telah dicapai. Bioanod TC-MGF yang baru dibangunkan telah dilaksanakan di ruang dobel (H-jenis) MFC. Prestasi bioanod TC-MGF dibandingkan dengan hasil yang diperoleh menggunakan bioanod grafit (BGF) yang tidak diubah suai. Keupayaan kuasa maksimum untuk bioanod BGF dan TC-MGF masing-masing adalah  $458.8 \pm 5.0$  and  $940.3 \pm 4.2$   $\text{mWm}^{-2}$ . Dalam kajian seterusnya, reaktor MFC kecil dibuat dengan bioanod TC-MGF untuk membandingkan prestasi MFC dengan membran sel bahan bakar yang biasa digunakan, Nafion (N-117 dan N-115), yang dikaji bersama dengan N-212 Membran dalam reka bentuk katod di-udara kubus tunggal-ruang (S-CCD-AC). Ketumpatan kuasa dan kecekapan *columbic* ( $1022.5 \text{ mWm}^{-2}$  - 35.4%) membran N-115 jauh lebih tinggi dan daripada nilai yang diperoleh untuk N-117 ( $592 \text{ mWm}^{-2}$  - 15.6%) dan membran N-212 ( $493 \text{ mWm}^{-2}$  - 12.3%). Reka bentuk kelompok MFC dengan reka bentuk kelompok bioanod TC-MGF (Conch shell) menggunakan membran N-115 yang mempunyai MFC aliran udara-katod (A-CU) MFC, sebagai gabungan aliran dan teknologi MFC digunakan untuk membandingkan kewujudan dan ketiadaan reka bentuk membran. A-CUMFC dengan membran yang kurang pada kadar aliran  $0.6 \text{ mL min}^{-1}$ , jarak anod  $0.5 \text{ cm}$  dan kepekatan AR27 pada  $900 \text{ mg L}^{-1}$ , kadar penyahairan yang tinggi (98%) dicapai dalam operasi 60 hari, adalah 40% lebih tinggi daripada membran-MFC. Ketumpatan kuasa maksimum purata yang diperoleh ( $1250 \text{ mWm}^{-2}$ ) dengan menggunakan MFC yang tidak membran adalah lebih tinggi daripada membran-MFC ( $1108 \text{ mWm}^{-2}$ ) dalam operasi 80 hari dengan bioanod TC-MGF.

## TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	<b>DECLARATION</b>	ii
	<b>DEDICATION</b>	iii
	<b>ACKNOWLEDGEMENT</b>	iv
	<b>ABSTRACT</b>	v
	<b>ABSTRAK</b>	vi
	<b>TABLE OF CONTENTS</b>	vii
	<b>LIST OF TABLES</b>	xvi
	<b>LIST OF FIGURES</b>	xvii
	<b>LIST OF ABBREVIATIONS</b>	xxv
	<b>LIST OF SYMBOLS</b>	xxviii
	<b>LIST OF APPENDICES</b>	xxix
<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
	1.1 Study Background	1
	1.2 Problem Statement	6
	1.3 Objectives of this study	7
	1.4 Scope of the Study	8
	1.5 Significance of this Study	9
	1.6 Thesis Organisation	10
<b>2</b>	<b>LITERATURE REVIEW</b>	<b>12</b>
	2.1 Classification of Dyes	12
	2.1.1 Azo Dyes	16
	2.1.1.1 Acid Red 27(AR27)/Amaranth	18
	2.2 Toxicity of Azo Dyes	19

2.3	Different Treatments for Azo Dye Effluents	19
2.3.1	Physical Methods	20
2.3.2	Chemical Methods	21
2.3.3	Biological Methods	21
2.4	Azo Dye Decolourisation	24
2.4.1	Acid Red 27 (AR27) Decolourisation Mechanism	26
2.5	Decolourisation of Dye Using Pure Bacterial Cultures versus Mixed Cultures or Co-cultures	27
2.5.1	Dye Degradation Ability of the NAR-2 Microbial Consortium	29
2.6	Biodegradation of Dye Intermediates	31
2.6.1	Biodegradation Pathway of Acid Red 27 (AR27)	32
2.7	Factors Affecting Dye Biodegradation	34
2.8	Detection and Analysis of Degradation Products of Azo Dyes	34
2.9	Zeolite (Clinoptilolite)	36
2.10	Surfactants	38
2.10.1	Surfactant-Modified Clinoptilolite (S-MC)	40
2.10.2	Applications of the Surfactant-Modified Clinoptilolite (S-MC)	44
2.11	Biological Fuel Cells	45
2.12	Basic Principles of an Microbial Fuel Cell (MFC)	46
2.12.1	Anode	48
2.12.1.1	Modified Anode Carbon-Based Supports	49
2.12.2	Cathode	55
2.12.2.1	Conditions in a Cathode Chamber	56
2.12.3	Membrane	56
2.12.4	Active Biocatalysts	59
2.12.4.1	Microbial Ecology in the Anode Compartment (Bioanode)	60
2.12.4.2	Biofilms and the Exopolysaccharide (EPS) Characterisation on the Surface of the Anode (Bioanode)	63
2.13	Electrochemistry in Microbial Fuel Cells (MFCs)	66



2.13.1	Activation Loss	67
2.13.2	Ohmic Loss	68
2.13.3	Transport Loss	68
2.14	Electron Transfer in Microbial Fuel Cells (MFCs)	69
2.14.1	Presence of Nanowires	71
2.14.2	Use of Mediators for Electron Transport	72
2.14.2.1	Use of Artificial Mediators	73
2.14.2.2	Self-producing Electron Mediators Generated by Microbes	73
2.15	Microbial Fuel Cell (MFC) Design	74
2.15.1	Mediator and Mediator-less MFC Designs	74
2.15.2	MFC Component Design	75
2.15.2.1	Two-compartment MFC System	76
2.15.2.2	One-compartment MFC System	78
2.16	Upflow Mode MFC System	80
2.17	MFC Applications	82
2.17.1	Electricity Generation	82
2.17.2	Biosensors	83
2.17.3	Biological Hydrogen Production	83
2.17.4	Wastewater Treatment	84
<b>3</b>	<b>MATERIALS AND METHODS</b>	<b>86</b>
3.1	Introduction	86
3.2	Microorganisms	88
3.3	Chemicals	88
3.4	Preparation of the Stock Solutions	88
3.5	Growth Media	90
3.5.1	Nutrient Agar	90
3.5.2	P5 and the Modified P5 Media	90
3.5.3	Preparation of the Growth Medium (starter culture-subculture)	92
3.6	Preparation and Characterisation of Treated Clinoptilolite with Surfactant	93
3.6.1	Preparation of Clinoptilolite (ZeoChem®)	93
3.6.2	Preparation of Hexadecyltrimethylammonium bromide (HDTMA-Br) Surfactant Solution	93

3.6.3	Preparation of the Surfactant-Treated Clinoptilolite (S-TC)	94
3.6.4	Spectrophotometric Determination of Surfactant (HDTM-Br)	94
3.6.4.1	Determination of HDTMA Concentration Adsorbed on Surfactant-Treated Clinoptilolite (S-TC)	95
3.6.5	Characterisation of Surfactant-Treated Clinoptilolite (S-TC)	96
3.6.5.1	Field Emission Scanning Electron Microscopy (FESEM)	96
3.6.5.2	X-Ray Diffraction (XRD) Analysis	96
3.6.5.3	Attenuated Total Reflection Fourier Transform Infrared (ATR-FTIR) Spectroscopy	96
3.6.5.4	Energy Dispersive X-ray (EDX) Spectroscopy	97
3.6.5.5	Cyclic Voltammetry (CV) Analysis Experiments	97
3.7	Preparation and Characterisation of Modified Anode	100
3.7.1	Treated Clinoptilolite-Modified Graphite Felt (TC-MGF) Anode Preparation	100
3.7.2	Characterisation of Treated Clinoptilolite-modified graphite felt (TC-MGF) Anode	102
3.7.2.1	Surface Area Analysis	103
3.8	Immobilisation of the Bacterial Consortium NAR-2, on the TC-MGF Anode	104
3.8.1	Preparation of Bacteria at Exponential Phase for Adsorption onto TC-MGF anode	104
3.8.1.1	Plate Count Method	104
3.8.1.2	Estimation of Cell Dry Weight and Optical Density (OD)	105
3.8.2	Optimisation Studies for the Immobilisation of Bacterial Cells on TC-MGF Anode	106
3.8.2.1	Immobilisation of the NAR-2 Bacterial Consortium into Upflow Closed Loop Reactor	107
3.9	Dual-Chamber MFC (H-Type)	111
3.9.1	Introduction	111

3.9.2	Dual-Chamber MFC (H-Type) Construction and Operation	113
3.9.2.1	Operation of Dual-Chamber (H-type) MFC	113
3.9.3	Analytical and Calculation Methods	115
3.9.3.1	Dye decolourisation	115
3.9.3.2	Electrical Parameters and Measurements	115
3.9.4	Characterisation of Bioanode	116
3.9.4.1	Cyclic Voltammetry analysis	117
3.9.5	Determination of the Degradation Products	117
3.9.5.1	UV-Vis Spectrophotometer	117
3.9.5.2	Gas Chromatography-Mass Spectrometry (GC-MS) Analysis	118
3.10	Cube-Shaped Single-Chamber Di-Air Cathode	118
3.10.1	Introduction	118
3.10.2	Nafion Membranes	119
3.10.2.1	Identification of the Membrane Characteristics	119
3.10.2.2	Measurement of Dissolved Oxygen Concentration and Dye Diffusion Coefficient	120
3.10.3	Cube-Shaped Single-Chamber Di-Air Cathode Construction	121
3.10.3.1	Preparation of Membrane-Cathode	121
3.10.3.2	Cube-Shaped Single-Chamber Di-Air Cathode Construction	122
3.10.3.3	Operation of Cube-Shaped Single-Chamber Di-Air Cathode	124
3.10.4	Analysis	124
3.10.4.1	Electrochemical Measurements	124
3.10.4.2	Coulombic Efficiency (CE)	125
3.11	Single Column Upflow MFC	126
3.11.1	Introduction	126
3.11.2	Construction and Operation of Single Column Upflow MFC	127
3.11.3	Experimental Setup	129

3.11.3.1	Setup for the Immobilisation of the NAR-2 Bacterial Consortium in an Upflow Closed Loop MFC	129
3.11.3.2	Operation of Single Column Upflow MFC for decolourisation and power generation	132
3.11.3.3	Optimisation the Dye Medium in an Upflow MFC for Decolourisation and Bioelectricity Generation	132
3.11.3.4	Operation Upflow MFC for Optimised Power (cathode and anode distance)	133
3.11.4	Determination of Glucose Concentration by Dinitrosalicylic (DNS) Method	134
3.11.5	Chemical Analytical Techniques	135
3.11.5.1	Determination of Total Aromatic Amines (TAA)	136
3.11.5.2	TNT Persulphate Digestion Technique	138
3.11.5.2.1	Accuracy Checks	138
3.11.5.3	Chemical Oxygen Demand (COD)	140
3.11.5.3.1	Standard Potassium Dichromate Reagent-Digestion Solution	141
3.11.5.3.2	Sulphuric Acid Reagent-Catalyst Solution	141
3.11.5.3.3	Standard Ferrous Ammonium Sulphate Solution	142
<b>4</b>	<b>CHARACTERISATION OF TREATED CLINOPTILOLITE AND MODIFIED ANODE (TC-MGF) WITH APPLIED TC-MGF BIOANODE IN DUAL-CHAMBER MICROBIAL FUEL CELL</b>	<b>143</b>
4.1	Introduction	143
4.2	Characterisation of Surfactant-Treated Clinoptilolite (S-TC) and Treated Clinoptilolite-Modified Graphite Felt (TC-MGF) Anode	144
4.2.1	Characterisation of Clinoptilolite and Surfactant-Treated Clinoptilolite (S-TC)	144

4.2.1.1	Clinoptilolite Size Range Determination	144
4.2.1.2	Morphological Structure	146
4.2.1.3	Structural Characteristics	148
4.2.1.3.1	X-ray diffraction (XRD)	148
4.2.1.3.2	Attenuated Total Reflection Fourier Transform Infrared Spectroscopy (ATR-FTIR)	149
4.2.1.4	Elemental Characteristics	152
4.2.1.5	Analysis of Quaternary Ammonium Cationic (QAC) Surfactant Adsorbed onto Clinoptilolite	155
4.2.1.6	Cyclic Voltammetry (CV) Analysis	157
4.2.2	Characterisation of Bare Graphite Felt (BGF) and Treated Clinoptilolite-Modified Graphite felt (TC-MGF) Anode	158
4.2.2.1	Optimisation of S-TC Fine Powders on Modified Anodes (CV analysis)	158
4.2.2.2	FESEM and EDX Analysis	160
4.2.2.3	ATR-FTIR and BET analysis	163
4.3	Immobilisation of Bacterial onto TC-MGF anode	164
4.3.1	Initial Bacterial Loading Determination	165
4.3.2	Screening of Optimum HDTMA-Br Surfactant Concentration for A1 Bacterial Cell Immobilisation	168
4.3.3	Optimisation of Bacterial Cells Immobilisation onto Treated Clinoptilolite-Modified Graphite Felt (TC-MGF)	173
4.3.3.1	Initial Cell Concentration ( $\text{cfu mL}^{-1}$ ) as Prerequisite to Cell Immobilisation	174
4.3.3.2	Effect of Contact Time on NAR-2 Immobilisation	175
4.3.3.3	Effect of Agitation Speed (rpm)	176
4.4	Application of Treated Clinoptilolite-Modified (TC-MGF) anode Immobilised Under Pre-optimised	

Condition for Decolourisation of AR27 and Bioelectricity Generation in Dual-Chamber (H-type) MFC	177
4.4.1 Start-up and Open Circuit Voltage (OCV) of the MFC with the TC-MGF	178
4.4.2 Colour Removal AR27 and Bioelectricity Generation in Dual-Chamber (H-type) MFC	180
4.4.2.1 Dye Removal (pre-optimised)	180
4.4.3 Polarisation, Power density, and Internal Resistance (pre-optimised)	186
4.4.4 Characterisation of TC-MGF Bioanodes (EPS)	187
4.4.4.1 Morphological Analysis (FESEM)	187
4.4.4.2 CV, ATR-FTIR and Elemental Mapping Analysis of Bioanode	189
4.4.5 Analysis of Dye Degradation Products	196
4.4.5.1 UV-Vis Spectrophotometer	196
4.4.5.2 Gas Chromatography-Mass Spectrometry (GC-MS) Analysis	197
4.4.6 Colour Removal AR27 and Bioelectricity Generation in Glucose, Mixture AR27-Glucose and AR27 MFCs (TC-MGF MFCs anode material)	198
4.4.6.1 Colour removal and OCV monitoring	198
4.4.6.2 Bacterial Community in the Anode of Three MFCs Reactor (variable carbon source)	200
4.4.6.2.1 FESEM analysis	200
4.4.6.2.2 Quantitative Leached out Analysis of NAR-2 Bacterial Consortium	202
4.5 Summary of Characterisation Anode and Bioanode within Power Generation and Dye Removal	203
<b>5 COMPARISON OF PHYSICAL PROPERTIES OF DIFFERENT NAFION MEMBRANES USED IN CUBIC- MICROBIAL FUEL CELLS WITH TC-MGF ANODE AND DEVELOPED UPFLOW MFC DESIGN</b>	<b>207</b>
5.1 Introduction	207

5.2	Single Cubic Di-Air Cathode MFC	208
5.2.1	Characterisation of the membranes	208
5.2.2	Comparison of Oxygen and Dye mass transfer Coefficients for Different Membranes	213
5.2.3	Effects of Membranes on AR27 Decolourisation with TC-MGF Anode in Cubic MFC	214
5.2.4	Coulombic Efficiency (CE), COD and Power Density in Cubic MFC	216
5.3	Single Column Upflow MFC	217
5.3.1	Effect of Anode and Cathode Spacing on Voltage and Power Generation	218
5.3.2	Effect of the External Resistance	219
5.3.3	Effects of Flow Rate in Decolourisation AR27 and Power Generation	221
5.3.4	Effect of Initial Dye Concentration AR27 on Decolourisation and Bioelectricity Generation	222
5.3.5	Effect of Initial Glucose Concentration on Decolourisation and Cell Leached out NAR-2 Bacterial Consortium	225
5.3.6	Upflow Membrane-Less and Membrane MFC Operation	227
5.3.6.1	Colour Removal, and Coulombic Efficiency Upflow MFC	227
5.3.6.2	TAA and Nitrogen Removal	229
5.3.7	Biofilm Development on Spiral Anode Surface under Membrane-Less and Membrane Operation	231
<b>6</b>	<b>CONCLUSION AND RECOMMENDATIONS</b>	<b>236</b>
6.1	Introduction	236
6.2	Recommendations for Further Research	239
	<b>REFERENCES</b>	<b>241</b>
	Appendices A-P	274-289

## LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Classification of dyes with chromophore structures (Sandhya, 2010)- <i>Part I</i>	13
2.2	Physiochemical structure of Acid Red 27(AR27)/Amaranth	18
2.3	Decolourisation of azo dyes by different microorganisms	23
2.4	List of genes or enzymes bacterial strains A1, C1 and L17 (Chan <i>et al.</i> , 2012b, c; Nasiri <i>et al.</i> , 2014)	31
2.5	Structure of hexadecyltrimethylammonium-bromide (HDTM-Br)	40
2.6	Photographs of anode materials used for MFC (Wei <i>et al.</i> , 2011)	49
2.7	Anode materials and size, inoculation source, and the FESEM images of various bioanode in MFC for the immobilisation of microorganisms- <i>Part I</i>	61
2.8	Electrochemical parameters used in MFCs study (He, 2007)	67
2.9	Two-compartment MFC configuration and inoculation source	77
2.10	One-compartment MFC configuration and inoculation source	79
2.11	Upflow MFC configuration and inoculation source	81
3.1	Medium for the growth of bacteria and decolourisation	91
4.1	Particle size distribution of clinoptilolite after sieving	145
4.2	Bands for HDTMA-(surfactant)/clinoptilolite/surfactant-treated clinoptilolite (S-TC) ATR-FTIR spectra	151
4.3	Elemental composition of clinoptilolite and S-TC from EDX analysis	154
4.4	Compression of the characteristics of the BGF and TC-MGF anodes	164



4.5	Selected initial cell concentrations ( $\text{cfu mL}^{-1}$ ) applied in the immobilisation of bacterial cells onto treated clinoptilolite-modified graphite felt clinoptilolite (TC-MGF)	168
4.6	The bioelectricity generation of TC-MGF bioanode with different substrates	200
5.1	Surface roughness parameters of Nafion 117, Nafion 115, and Nafion 212 membranes resulted from analysing three randomly chosen AFM images	212
5.2	Mass transfer coefficients and diffusivities of oxygen and AR27 on each membrane	213
5.3	Effect of flow rates on COD removal, color removal, CE, OCV, and power density	222
5.4	Colour and COD removal, coulombic efficiency and power density of the membrane-less and membrane MFCs	228

## LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	The general chemical structure of azo dyes (Chacko and Subramaniam, 2011)	17
2.2	Chemical structure of toxic degradation product (Khan and Banerjee, 2010)	19
2.3	Treatment methods for the removal of dyes from wastewater effluents (Saratale <i>et al.</i> , 2011)	20
2.4	The decolourisation mechanism of azo dyes (Sandhya, 2010)	25
2.5	The reduction mechanism of AR27 dye (Wahab <i>et al.</i> , 2012)	26
2.6	General overview of anaerobic-aerobic treatment (Van deer Zee and Villaverde, 2005).	32
2.7	Pathway for the reduction of AR27 by NAR-2 bacterial consortium (adapted from Chan <i>et al.</i> , 2012a).	33
2.8	Binding of primary building units (PBU) and secondary building units (SBU) of zeolite (Margeta <i>et al.</i> , 2013)	37
2.9	Ion exchange mechanisms of HDTMA cationic surfactant with cations on clinoptilolite and electrostatic interaction of HDTMA with OH <sup>-</sup> on clinoptilolite (Kardi, 2013)	41
2.10	(a) Hypothetical transition of HDTMA attachment onto clinoptilolite surface with respect to time as adapted from Li (1999). (b) Adsorption of HDTMA molecules at concentration higher than CMC	42
2.11	Theoretical attachment of dye onto surfactant modified zeolite (Benkli <i>et al.</i> , 2004)	43
2.12	Schematic diagram of a dual-chamber microbial fuel cell (PEM = proton exchange membrane), (Najafpour, 2015).	47

2.13	FESEM images of the anode before and after modification used in MFCs: (a) graphite felt (b) Pani/m-wo <sub>3</sub> (Wang <i>et al.</i> , 2013a); (c) graphite felt (d) Nax (Song <i>et al.</i> , 2015); (e) Staimless Mesh (f) Stainless Mesh-modified graphite (Mardanpour <i>et al.</i> , 2012); (g) Stainless Mesh (300) (h) Stainless Mesh (300)-coated graphite Noori and Najafpour, (2015); (i) graphite felt (j) MCM-41 (Song <i>et al.</i> , 2015); (k) graphite (l) Electropolymerized graphite (EpGr) (Savizi <i>et al.</i> , 2012); and (m) carbon felt (n) Polyaniline-modified carbon (Li <i>et al.</i> , 2011a) -Part I	53
2.14	Mechanism of biofilm formation (Kardi, 2013)	65
2.15	Polarisation curve (He, 2007)	69
2.16	Schematic diagrams of direct electron transfer mechanism via membrane-bound cytochromes (Rinaldi <i>et al.</i> , 2008)	70
2.17	Schematic diagrams of electronically-conducting nanowires electron transfer (pili) (Rinaldi <i>et al.</i> , 2008)	71
2.18	Schematic diagrams of mediated electron transfer mechanism via added (exogenous) or secreted (endogenous) mediators. (Rinaldi <i>et al.</i> , 2008)	72
2.19	Model of components proposed to be the electron transfer from cell to the anode (final electron acceptor) in mediator-less MFCs using metal reducing bacteria ( <i>Geobacter</i> species) (Du <i>et al.</i> , 2007)	75
2.20	A microbial fuel cell supplying for low energy applicant (Najafpour, 2015)	82
3.1	Experimental Design	87
3.2	Photos of (a) p5, and (b) the Modified p5 medium at pH 7.0	92
3.3	Photo of (a) the clinoptilolite (size: 100-500 μm) before and (b) after sieving (size: 44-63 μm)	93
3.4	Standard curve of the absorbance versus HDTMA-Br concentration (mM)	95
3.5	Schematic diagram of a three-electrode electrochemical system	98

3.6	(a) Schematic diagram of preparation treated-clinoptilolite carbon (TC-MC) paste; (b) photo of TC-MC paste as working anode	99
3.7	(a) Bare graphite felt (BGF) and (b) treated clinoptilolite-modified graphite felt (TC-MGF) anodes	101
3.8	Cell dry weight standard curve of A1	106
3.9	Schematic diagram and photo of experimental system. Close loop immobilisation reactor, with fibrous matrix as working electrode. Arrows depict direction of phosphate buffer solution and bacteria	107
3.10	Simplified flow diagrams for optimisation parameters for treated clinoptilolite-modified anode with varying (a) age of bacteria, (b) contact time, and (c) agitation speed	111
3.11	(a) Schematic and (b) picture of the lab-scale of single-chamber di-air cathode MFC auxiliary equipment under OCV test	123
3.12	(a) Schematic upflow MFC and (b) lab-scale picture of cathode-membrane tube	129
3.13	Picture of immobilisation (a) and (b) biofilm at 37 °C in an Upflow closed loop MFC	131
3.14	Glucose standard curve at 540 nm	135
3.15	(a) Calibration curve for reduced AR27 dye, and (b) calibration curve for benzidine	137
4.1	Field emission scanning electron micrographs of clinoptilolite and surfactant-treated clinoptilolite (S-TC). (a) Unmodified Clinoptilolite and (b) treated clinoptilolite with HDTMA coverage at 10K magnification	147
4.2	XRD pattern of (a) clinoptilolite and (b) surfactant-treated clinoptilolite (S-TC) peaks belonging to impurities: (C) celadonite, (P) plagioclase feldspars, (Q) quartz	148
4.3	ATR-FTIR spectra of (a) clinoptilolite, (b) surfactant-treated clinoptilolite and (S-TC), and (c) HDTMA (surfactant)	150
4.4	FTIR spectrum of clinoptilolite (Faghihian <i>et al.</i> , 2010)	152

4.5	EDX spectra of (a) clinoptilolite and (b) surfactant-treated clinoptilolite (S-TC)	153
4.6	Adsorption of HDTMA surfactant onto clinoptilolite surface at varying initial loading concentrations	156
4.7	CVs of S-TC and clinoptilolite at scan rate $0.02 \text{ Vs}^{-1}$ , phosphate buffer 0.05 M, pH 7.0	158
4.8	CVs of different amount of the S-TC fine powders on the graphite felt	160
4.9	FESEM images of (a) BGF and (b) TC-MGF anodes. The area in (a) and (b) (square) are magnified and shown in (c) and (d)	161
4.10	EDX analysis or spectra of (a) BGF and (b) TC-MGF anodes	162
4.11	ATR-FTIR analysis spectra of BGF and TC-MGF anodes	163
4.12	Growth at log phase of A1 reflecting on the general relationship between viable cell count, cell dry weight and optical density (600 nm)	165
4.13	Growth at log phase of L17 reflecting on the general relationship between viable cell count ( $\text{cfu mL}^{-1}$ ) and optical density (600 nm)	166
4.14	Growth at log phase of C1 reflecting on the general relationship between viable cell count ( $\text{cfu mL}^{-1}$ ) and optical density (600 nm)	166
4.15	Effect of HDTMA adsorbed by clinoptilolite ( $\text{mmol kg}^{-1}$ ) towards viability of A1 immobilised TC-MGF ( $\text{cfu mL}^{-1}$ ) in phosphate buffer (pH 7.0) for 9 h agitation at 70 rpm, 37 °C	169
4.16	Effect of high initial loading of HDTMA on the viability of bacteria	170
4.17	(a) Uneven distribution of bacterial colonisation on the TC-MGF anode surface. The red box shows the part of anode which has colonisation by bacteria A1. Other parts of the clinoptilolite surface are only sparsely populated. (b) Patchy bilayer configuration at high concentration of HDTMA. The red box shows detachment of HDTMA that proved deleterious to bacteria A1	173

4.18	The adsorption of A1, L17 and C1 onto TC-MGF with different initial cell concentration (cfu mL <sup>-1</sup> )	175
4.19	The adsorption of A1, L17 and C1 onto TC-MGF anode at varying contact time (minutes)	176
4.20	The adsorption of A1, L17 and C1 onto TC-MGF anode at varying agitation (rpm)	177
4.21	OCV generation in the start-up phase with different anodes. (Each green arrow and green dash represents the beginning of one complete cycle and the maximum OCV, respectively.)	179
4.22	(a) The voltage and green arrow represents the beginning of one complete cycle (external resistance 10 kΩ), (b) colour removal and leached out of the NAR-2 bacterial consortium (pink squares, TC-MGF-bioanode; black triangles, BGF-bioanode; solidline, decolourisation; dotted line, leached out)	181
4.23	Decolourisation of Acid Red 27 (AR27) using bare graphite felt (BGF) and TC-MGF anode that were previously immobilised with bacterial cells at pH 7.0	183
4.24	Illustration showing decolourisation of AR27 by bacterial immobilised onto treated clinoptilolite-modified graphite felt (TC-MGF)	185
4.25	Polarisation curves of the BGF and TC-MGF bioanodes at various resistances (300,000-10 Ω) after 368 hours	186
4.26	FESEM images of the bioanode electrode surfaces after 16 days operation in MFCs (a) TC-MGF bioanode, (b) BGF bioanode. (c, e) The attachment of sessile cells was observed within the EPS on the TC-MGF bioanode (squares) are magnified, and (d) the area on the BGF bioanode (square) is magnified	189
4.27	CVs BGF and TC-MGF bioanodes	190
4.28	ATR-FTIR of BGF and TC-MGF bioanodes surfaces after 16 days operation in MFCs	192
4.29	EDX of (a) BGF and (b) TC-MGF bioanodes showing elemental compositions. (c, d) FESEM elemental mapping of	

	BGF and TC-MGF bioanodes samples showing the location and quantities of individual elements	195
4.30	UV-Vis spectral change of AR27 in BGF and TC-MGF-MFCs at 303 and 368 hours of operating times	197
4.31	Proposed degradation mechanism of AR27 on the BGF and TC-MGF-MFCs	198
4.32	FESEM images of (a) nanowire-like structure on the TCM-GF bioanode, of the AR27-glucose system (8K magnification) and (b) bioanode TC-MGF of the AR27 system (8K magnification)	202
5.1	AFM images of the Nafion 212 membrane with: (a) two and (b) three-dimensional	209
5.2	AFM images of the Nafion 117 membrane with: (a) two and (b) three-dimensional	210
5.3	AFM images of the Nafion 115 membrane with: (a) two and (b) three-dimensional	211
5.4	Decolourisation of AR27 ( $300 \text{ mg L}^{-1}$ ) in MFCs with different membranes using glucose ( $2.5 \text{ g L}^{-1}$ ) as co-substrate at an external load of $10 \Omega$	215
5.5	COD removal and CE of the cubic-MFC systems	216
5.6	Power density graph of the three different MFCs	217
5.7	Effect of spacing between anode and cathode on power generation (glucose $2.5 \text{ g L}^{-1}$ )	219
5.8	Deolourisation rate through the variable external resistance	220
5.9	Influence of initial AR27 concentration ( $\text{mg L}^{-1}$ ) on decolourisation at external load of ( $10 \Omega$ ) and substrate removal (a), (b) voltage generation, electrode potentials (c) in the air-cathode upflow single-chamber. Values of were the mean of duplicate measurements. Bars represent standard errors during the operation period time 400 hours	224
5.10	Effect of glucose concentration on AR27 decolourisation ( $300 \text{ mg L}^{-1}$ )	226
5.11	TAA (a) recovery and (b) removal efficiency in membrane and membrane-less MFC system	230

- 5.12 (a) The bioanode TC-MGF of the membrane-less system, at 2.5K magnification, and (b) partial enlarged view of the bioanode TC-MGF of the membrane-less system at 5K magnification. (c) The bioanode TC-MGF of the membrane system, at 2.5K magnification, and (d) partial enlarged view of the bioanode TC-MGF of the membrane system at 5K magnification



**LIST OF ABBREVIATIONS**

$A_{600\text{ nm}}$	-	Absorbance at the wavelength of 600 nm
A1	-	<i>Citrobacter</i> sp. A1
$\text{Al}^{3+}$	-	Aluminum cation
AFM	-	Atomic Force Microscope
AR27	-	Acid Red 27
APHA	-	American Public Health Association
ATR-FTIR	-	Attenuated total reflection Fourier transform infrared
BET	-	Brunauer emmett and teller
BFC	-	Biological fuel cell
BS	-	British standard
C1	-	<i>Enterococcus</i> sp. C1
Ca	-	Calcium
$\text{Ca}^{2+}$	-	Calcium cation
CC	-	Carbon cloth
CE	-	Coulombic efficiency
CEC	-	Cation exchange capacity
CEM	-	Cation exchange membrane
CMC	-	Critical micelle concentration
COD	-	Chemical Oxygen Demand
CT	-	Contact time
CV	-	Cyclic Voltammetry
$e$	-	Electronic charge
EDX	-	Energy dispersive x-ray analysis
EPSs	-	Exopolysaccharides
FESEM	-	Field emission scanning electron microscopy
BGF	-	Bare Graphite felt
GC-MS	-	Gas Chromatography-Mass Spectrophotometry
$\text{H}^+$	-	Hydrogen cation

H <sub>2</sub> O	-	Water
H <sub>2</sub> SO <sub>4</sub>	-	Sulphuric acid
HDTM-BR	-	Hexadecyltrimethylammonium bromide
HPLC	-	High Performance Liquid Chromatography
I	-	Current
K	-	Potassium
K <sup>+</sup>	-	Potassium cation
K <sub>2</sub> HPO <sub>4</sub>	-	Dipotassium hydrogen phosphate
KH <sub>2</sub> PO <sub>4</sub>	-	Potassium dihydrogen phosphate
K <sub>2</sub> Cr <sub>2</sub> O <sub>7</sub>	-	Potassium dichromate
KBr	-	Potassium bromide
L17	-	<i>Enterobacter</i> sp. L17
MCM-41	-	Mobile Composition of Matter No. 41
MFC	-	Microbial fuel cell
Mg	-	Magnesium
Mg <sup>+2</sup>	-	Magnesium cation
NA	-	Nutrient agar
Na	-	Sodium
Na <sup>+</sup>	-	Sodium cation
NMR	-	Nuclear magnetic resonance
NO <sub>3</sub> <sup>-</sup>	-	Nitrate
O	-	Oxygen
OD	-	Optical Density
(OH) <sup>-</sup>	-	Hydroxide ion
P	-	Power
PBS	-	Phosphate buffer solution
PEM	-	Proton exchange membrane
Pt	-	Platinum
QAC	-	Quarternary Ammonium Cationic Surfactant
R <sub>int</sub>	-	Internal resistance
Rpm	-	Revolution per minute
RP-1	-	Real product-1
RP-2	-	Real product-2
RT	-	Room temperature
S	-	Sulfur

Sec	-	Seconds
SCE	-	Saturated Calomel Electrode
Si	-	Silicon
SiO <sub>2</sub>	-	Silica
SSA	-	Specific surface area
S-TC	-	Surfactant-treated clinoptilolite
SMZ	-	Surfactant modified zeolite
TAA	-	Total Aromatic Amines
TC-MGF	-	Treated clinoptilolite-modified graphite felt
TNT	-	Total Nitrogen Test
UV-Vis	-	Ultraviolet Visible
XRD	-	X-ray diffraction
3-D	-	Three-dimensional

## LIST OF SYMBOLS

$^{\circ}\text{C}$	-	Centigrade
cm	-	centimetre
cfu mL <sup>-1</sup>	-	Colony Forming Unit per millilitre
g L <sup>-1</sup>	-	gram per litre
k $\Omega$	-	Kiloohms
L	-	Litter
mg L <sup>-1</sup>	-	milligram per Litre
mL	-	millilitre
mL min <sup>-1</sup>	-	millilitre per minutes
mM	-	millimolar
M	-	Molar
M $\Omega$	-	Megaohms
$\mu\text{L}$	-	Microliter
$\mu\text{m}$	-	Micrometre
N	-	Normalite
nm	-	nanometer
V	-	Voltage
v	-	Volume of gas adsorbed per unit weight of anode at a pressure
$v_m$	-	Volume of gas adsorbed for monolayer coverage
v/v	-	Volume per volume
w/v	-	Weight per volume
$\Omega$	-	Ohm
$\theta$	-	Critical angle of incidence of the x-ray beam on the crystal plane
$\lambda$	-	Wave-length

## LIST OF APPENDICES

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
A	Characteristics of seven groups of zeolites	274
B	Preparation of phosphate buffer 1M (PH 7.0)	275
C	(a) Field Emission Scanning Electron Microscope and (b) Vacuum sputter coater	276
D	Sample preparation for FESEM test and buffer was used for the fixing process	277
E	The resistance of TC-MGF was measured with a digital multimeter (Snawa-Japan) via 2-point probe method	278
F	(a) Surface area measurement devices and (b) BET plot correlation factor of 0.999 for treated clinoptilolite-modified graphite felt (TC-MGF)	279
G	(a) Settle down and (b) exhausted feed in dual-chamber MFC in see through incubator with temperature control	280
H	Photo of pre-treatment of Nafion using 3% H <sub>2</sub> O <sub>2</sub> and H <sub>2</sub> SO <sub>4</sub> solution, at 80 °C	281
I	Di-air cathode single chamber under (a) OCV and (b) CCV testing	282
J	Membrane-cathode (assembling with hot press machine)	283
K	Calibration of Watson Marlow and Catalyst FH10 Pump	284
L	Square root scans rate measuring	285
M	Square root of 25, 55 and 75% (w/w) S-TC fine powders in the modified anode	286
N	FESEM morphology images of the membrane-electrode surface (Nafion, 115) with (a) top view, (b) cross-section	287

O	FESEM morphology images of the membrane-electrode surface (Nafion 117) with (a) top view, (b) cross-section	288
P	Different images of the cells that have been fabricated at laboratory scale	289

## CHAPTER 1

### INTRODUCTION

#### 1.1 Study Background

Recently, it has been observed that due to a massive increase in population and industrialisation, there has been a need for further knowledge regarding energy resources. Today, scientists are faced with the daunting task of discovering novel and innovative techniques of generating energy from alternative energy sources. To date, the most commonly used source of energy is fossil fuel, which is still able to support energy demands for the next 100 years (Daniel *et al.*, 2009). However, many reports have stated that fossil fuels, especially oil, natural gas, and coal, are being used at a very alarming rate. Additionally, it has also been observed that the use of fossil fuels has led to an increase in global warming and can cause significant variations in climatic conditions. This has resulted in an increasing demand for alternative and cleaner sources of renewable energy (He, 2007).

The demand for energy has tremendously increased over the last few years. Scientists have started focussing on waste streams, including industrial waste, agricultural waste, domestic and food processing wastes, as alternative energy resources and for a cleaner environment due to the presence of biodegradable wastes (He, 2007).

Azo dyes are one of the biggest and the most popular classes of synthetic dyes with wide applications in the paper, textile, cosmetics, food and pharmaceutical

industries (Idel-aouad *et al.*, 2011; Mendes *et al.*, 2011; Jadhav *et al.*, 2013). It has been observed that every year azo dyes are produced approximately  $7 \times 10^5$  metric tons, and approximately 5-10% of these dyes are released into the environment as waste (Rai *et al.*, 2005; Dafale *et al.*, 2010). Azo dyes are very popular due to their cost-effective synthesis process, stability, and availability in many colours. Chemically, azo dyes consist of one or more azo bonds (-N=N-) associated with the aromatic structure in the molecule (Jadhav *et al.*, 2013; Jović *et al.*, 2013; Fang *et al.*, 2015).

Azo dyes itself is hazardous; however, it can be more hazardous when the azo bonds are reduced to give amines which are more carcinogenic than the parent structure (Zouari-Mechichi *et al.*, 2006; Mendes *et al.*, 2011; Costa *et al.*, 2012). Hence, the incomplete treatment of azo dyes leads to the formation of aromatic amines as their breakdown products, which are carcinogenic in nature (Jonstrup *et al.*, 2011). Azo dyes are used extensively in the textile industry, thereby resulting in massive water pollution because the azo dyes are resistant to degradation. One such example is Acid Red 27 (AR27), a naphthylamine sulfonic azo dye which is widely available in the form of atrisodium salt. It is also commonly known as C.I. 16185, C.I. Food Red 9, Amaranth, or Azorubin S (Hong *et al.*, 2007).

The physicochemical methods used for the removal of dyes from wastewater containing azo dyes include coagulation, adsorption, membrane filtration, chemical oxidation, and ozonation (Selcuk, 2005; Dos Santos *et al.*, 2007; Alaton *et al.*, 2002; Forgacs *et al.*, 2004). Many reports have stated that these methods result in massive amounts of sludge and lead to secondary pollution as they use a lot of chemicals. However, ozonation method would not produce solid waste or sludge as in the physical methods, but the energy consuming has turned this method to be less practical (Robinson *et al.*, 2001; Hong *et al.*, 2007; Saratale *et al.*, 2011; Dafale *et al.*, 2010). Additionally, the physical and chemical methods used for the removal of azo dyes are very expensive and time-consuming (Sarkar *et al.*, 2011; Chengalroyen and Dabbs, 2013). Microbes play very important roles in bringing about the degradation of xenobiotic compounds. Hence, bioremediation using microbes (and their enzymatic



reaction) was developed as an alternative and an environmentally-friendly technique for degrading azo dyes (Saratale *et al.*, 2011; Chengalroyen and Dabbs, 2013).

Biodegradation using a mixed microbial consortium is the most common technique for azo dye degradation. The anaerobic-aerobic treatment of wastewater containing azo dyes is an effective combination method for the biodegradation of azo dyes (Van der Zee and Villaverde, 2005; Jonstrup *et al.*, 2011). The anaerobic treatment procedure removes the colour of azo dyes; however, it also results in the formation of aromatic amines as the decolourisation products (McMullan *et al.*, 2001; Murali *et al.*, 2013; Pandey *et al.*, 2007). These aromatic amines are easily degraded using aerobic processes (Jonstrup *et al.*, 2011) through hydroxylation and ring-fission of the aromatic molecules (Supaka *et al.*, 2004). Earlier, a novel NAR-2 bacterial consortium was developed consisting of the *Citrobacter* sp. A1 (a bacterial strain previously isolated and characterised from the sewage oxidation pond at Universiti Teknologi Malaysia in Johor, Malaysia), *Enterobacter* sp. L17 strain (known as *Enterobacter cloacae* L17) and the *Enterococcus* sp. C1. These were the stock cultures available in the Nanomaterial Lab at UTM. These bacterial strains were given the acronyms A1 for the *Citrobacter* sp. A1, L17 for the *Enterobacter* sp. L17 and C1 for the *Enterococcus* sp. C1 (Chan *et al.*, 2012a). In their study, Chan *et al.* (2012a) reported on the decolourisation of Acid Red 27 using the NAR-2 bacterial consortium. They observed that the C1 culture was a more dominant decolouriser of the dye during the microaerophilic condition, whereas the A1 and the L17 cultures improved the resultant biotransformation of the dye intermediates during the deamination and desulphonation processes.

Microbial fuel cells (MFCs) are novel and ground-breaking technology that can help to reduce the dependence on fossil fuels for energy production. They have been successfully used for the production of energy from biological processes involved in wastewater treatment (Logan *et al.*, 2006; Zuo *et al.*, 2007; Murali *et al.*, 2013; Fang *et al.*, 2015). MFCs use microbes as the biocatalyst. The electrogenic bacteria forming biofilm on the anode surface produce electrons and protons during organic matter degradation under anaerobic conditions. The electrons are transferred to the cathode through an external circuit. Meanwhile, the protons are moved to the cathode through

the electrolyte and the separator. Eventually, the electrons and protons combine with oxygen to form water molecules to complete the reaction scheme (Song *et al.*, 2015). To maintain the electro-neutrality, the cations are transported from the anode to the cathode through a cation exchange membrane (CEM) (Logan *et al.*, 2006; Chen *et al.*, 2010).

Although several earlier reports have observed that microbes are able to produce fuels such as methane, ethane, and even hydrogen, there are very few reports on electrogenic microorganisms with respect to their use in microbial fuel cells (MFCs). Some electrogenic bacteria reported earlier include *Aeromonas*, *Clostridium*, *Citrobacter*, *Geobacter*, *Klebsiella*, *Pseudomonas*, *Rhodospirillum rubrum* and *Shewanella* (He, 2007; Humudat *et al.*, 2015).

In recent years, the azo dye decolourisation technique using MFC has become very interesting for biodegradation purposes. This system helps in a more continuous and flexible decolourisation process that can be used on an industrial scale. Many earlier reports have described the process of developing a mixed microbial consortium for degrading azo dyes by immobilising the culture on some appropriate support materials (Tony *et al.*, 2009). Bacterial immobilisation on the support matrix becomes a useful technique used in bioreactor studies (Hrenović *et al.*, 2005).

The main principle of the MFC revolves around the total number of electric charges which pass through the complete circuit every second. Hence, the electron transfer passage route should be as simple as possible, the distance between both electrodes has to be optimised to improve the mass transfer occurring between the electrodes (Cheng *et al.*, 2006b). Furthermore, the external resistance effect contributes greatly to the change in current density, which highlights the maximum probable power density. An earlier study reported that to produce a maximum power output when the internal resistance is equal to the external resistance of the MFC systems (Logan *et al.*, 2006).

Additionally, other studies have also noted that for MFCs, the proton exchange membranes (PEMs) play a vital role in separating the anodic and cathodic chambers.

The permeable membranes allow a smooth migration of the protons, produced at the anode, to the cathode. The different pore sizes of the PEMs are able to offer better stability, while the membranes themselves contribute the least internal resistance (Hou *et al.*, 2011a; Leong *et al.*, 2013).

When designing the MFC reactors, the performance of the anodes need to be taken into consideration. Several researchers have tested a wide variety of materials, and many configurations have also been developed in the past few years for improving the performance of MFCs (Liu and Logan, 2004). In recent years, many studies have focused on the use of carbon electrodes and carbon papers for developing MFC electrodes as they are cost-effective, non-corrosive and perfectly biocompatible (Wei *et al.*, 2011). On the other hand, these carbon materials have very low electrocatalytic movements for the anode microbial response and therefore, adjustment on the carbon electrode considered as main method for improving their activity (Park *et al.*, 2013).

In order to improve the efficiency of MFCs, investigation on modification of conventional carbon-based materials, such as graphite felt, carbon paper, and carbon cloth for the fabrication of the anode were conducted (Wang *et al.*, 2013a; Wang *et al.*, 2013b; Cheng and Logan 2007). Modification of anode may lead to a more efficient MFC; with specific properties of anodes, including biocompatibility, electronic conductivity, surface wettability and surface area were investigated (Park *et al.*, 2013; Chen *et al.*, 2013; Wei *et al.*, 2011). The anode property is strongly linked to the electrochemical reaction that affects the performance of the cell adhesion (Ginsburg and Karamanev 2007; Wu *et al.*, 2015).

Additionally, changes to the surface of the anode are known to enhance the bacterial adhesion and, at the same time, to improve the electron transfer between the microbes and the surface of the electrode. Many researchers have implemented a variety of physical and chemical modification processes, such as ammonia treatment, acid oxidation, and electrochemical oxidation, for improving the transfer of electrons (Du *et al.*, 2007; Wei *et al.*, 2011). An earlier study indicated that enhancing the electron transfer through an anode biofilm would result in an improvement of power output in the MFCs (Xie *et al.*, 2012; Fiset and Puig, 2015).

In their study, Park *et al.* (2013) conducted a magnetite/multiwall carbon nanotube (MWCNT) for coating anode, and it was observed that the reception of electrons by the anodic electrode was significantly enhanced by the use of *E. coli* biofilm in powered MFCs. Reports have stated that the anodic component and the arrangement can greatly affect the electron transmission, bacterial supplement, and substrate oxidation (Logan *et al.*, 2006; Lanas *et al.*, 2014). Recent studies have also revealed that positively-charged modifications and the use of natural surfaces, such as sand and quartz, improve the adhesion of the negatively-charged microbial culture due to the electrostatic force of attraction between the cells and the surface. Cheng and Logan (2007) studied the ammonium treatment and their results indicated that the positive charges on the carbon surfaces were greatly enhanced due to the ammonia gas which decreased the process and improved the power produced in the MFC with a microbial consortium. This approach proved to be effective in allowing the bacterial nanowire to adhere to the carbon cloth electrodes. In addition, Wu *et al.* (2015) observed that the use of different varieties of zeolite, namely NaX and Mobil Catalytic Material Number 41 (MCM-41), as a coating for the electrodes improves the performance of the MFC. This is due to an increase in the surface area in the range of 6.1-11 m<sup>2</sup> g<sup>-1</sup>, which favours the formation of a thick biofilm.

## 1.2 Problem Statement

Over the years, it has become very important to find novel and cleaner energy sources. Nowadays, many industries around the world contribute to water and environmental pollution. Many reports have stated that the microbial fuel cell (MFC) system is a very good technique for energy production, while several researchers have suggested that the MFC is capable of bringing about the treatment of azo dyes from wastewater and simultaneously producing electricity (Li *et al.*, 2010; Hou *et al.*, 2011a). Despite the fact that MFC being a novel technology for wastewater treatment and energy production, there are also some major drawbacks which limit the actual use of this technique, for example, the unsustainability of biocatalysts present on the anode, the configuration of the MFC, which limits the power output mainly during the scale-up, and the impractical batch-operation type of MFC systems as the microbes

would easily wash out from the system. Therefore, the anode compartment needs a stable biofilm to be formed on a suitable and large anode surface. Furthermore, the characteristics of the anode need to be adapted to the biofilm and the used application in the MFC system.

Thus, the power output from MFCs must be improved by optimising the reactor configuration and operating conditions, while deploying proper anode materials with biocompatibility and large specific surface area. Nevertheless, no research has demonstrated the use of the treated clinoptilolite-modified graphite felt (TC-MGF) anode as a support material for the adhesion of the NAR-2 bacterial consortium and the effect of TC-MGF on MFC performance. It is therefore important to fully understand the properties of this modified anode and its effect on the performance of the MFC system for simultaneously Acid Red 27 (AR27) azo dye decolourisation and electricity generation.

### **1.3 Objectives of this study**

In view of the current understanding and limited research to date, this study was conducted to determine the effect of a modified anode involved in the immobilisation and performance of a developed MFC on dye degradation and electricity generation. Hence, the following objectives were established to achieve the aim of the research:

1. To characterise treated clinoptilolite with surfactant hexadecyltrimethylammonium bromide (HDTMA-Br) followed by graphite felt anode modification using treated clinoptilolite (TC-MGF).
2. To determine the optimum conditions for the immobilisation of NAR-2 consortium onto TC-MGF anode using conventional method and characterise the TC-MGF bioanode.
3. To evaluate the effectiveness of TC-MGF bioanode for the generation of electricity from Acid Red 27 (AR27) dye in a dual-chamber (H-type)

MFC and its impact on the biodegraded products in comparison to graphite felt (BGF) bioanode.

4. To characterise and evaluate the performances of the different Nafion membranes for the decolourisation of the AR27 dye and the generation of electric power using a single cube-chamber di-air cathode MFC with TC-MGF anode.
5. To assess the generation of electric power and decolourisation efficiency in a continuous upflow single column air cathode MFC system by a TC-MGF spiral design along membrane-less and membrane operations followed by analysis of the resultant degraded products.

#### **1.4 Scope of the Study**

This study has focused primarily on investigating the improvement of anode using modification technique to improve bacterial adhesion, that would then be incorporated in the different types of the MFCs system. Hence, the performance of the MFCs were evaluated at their optimised conditions.

The scope of this research was to study the modification effect of the anode surface on the performance of the MFC for simultaneous azo dye decolourisation of the Acid Red 27 (AR27) and electricity generation, using bacterial consortium NAR-2. An initial attempt was made using a dual-chamber MFC utilising modified anode, namely treated clinoptilolite-modified graphite felt (TC-MGF). This study introduced a modification process into the MFC design using an inert, inorganic and robust material known as surfactant-treated clinoptilolite (S-TC). The TC-MGF anode was then further improved by optimisation studies in a single-chamber air cathode MFC system with the evaluation of the physical properties, the decolourisation and power output performance of the variable Nafion membranes (N-115, N-117 and N-212) in the MFC. Furthermore, in order to improve the MFC system for the biodegradation process and power generation, further modifications were made to the MFC design

using a spiral anode with membrane-less operation with respect to the chemical analysis in a continuous upflow single column air cathode MFC reactor.

### **1.5 Significance of this Study**

Earlier studies focused on the treatment of azo dyes using MFCs system for power generation. However, this study used a modified anode for immobilising the bacterial consortium, thereby improving the decolourisation and degradation of the model azo dye AR27, and also the resultant electricity production. The main idea behind this study was based on the ionic or hydrophobic interactions which take place between microbes and the modified anodes, based on the actual design. The anode was modified to be positively charged, so that it would then attract the negatively charged bacteria cell wall and the azo dye molecules. Hence, interaction between the dye molecules, nutrients and bacteria present would result in a desired decolourisation efficacy, especially if the system is continuous in operation. Therefore, the modified clinoptilolite anodes were installed to form an MFC system and were immobilised using the bacterial consortium, NAR-2. This system was used to carry out dye decolourisation and production of electricity. This was an initial attempt to construct an MFC system to generate electricity by decolourising the AR27 mono azo dye using the NAR-2 bacterial consortium. In several earlier studies, it was noted that when the bacteria were immobilised, they were able to be more resistant to the shock loads of the dye compared to the suspended bacterial culture (Ab.llah, 2012). The use of an upflow continuous MFC system along TC-MGF spiral anode (Conch Shell) with membrane-less design, as reported in this study, was demonstrated to be very significant and noteworthy as it brought more effective degradation of the azo dye within improved power generation and therefore, can be applied in the future investigation for the treatment of real wastewater containing different types of azo dyes for the generation of bioelectricity.

## 1.6 Thesis Organisation

The entire thesis consists of six chapters. Chapter 1 presents a concise introduction to the role of the MFC system in the degradation of recalcitrant chemical pollutants and also in the production of electric power using an active biofilm that is formed on a modified anode. Moreover, the chapter also exploits the use of the modified anode as a site for the adhesion of bacterial colonies. Furthermore, the chapter covers the problem statement, and the objectives, scope and significance of the study.

In Chapter 2, the basics of the modified anodes and the assessment of their role in the bacterial and anode interactions will be explained. The chapter also discusses the various anodic procedures and the MFC design, and also hypothesises about the reaction format mechanism. Furthermore, the chapter also includes earlier researches into the performance of the MFC system on the degradation of pollutants along with the subsequent power generation. Thereafter, the chapter presents the research framework, depending on the current understanding of the available studies.

In Chapter 3, an in-depth analysis of the methods used, along with the research methodology which would be applied in the study, is presented. Furthermore, some laboratory experiments were conducted to determine the performance of the MFC based on the electricity calculations. Also, the chapter discusses the characterisation studies which were carried out using the modified anodes with the help of various microscopic and spectroscopic techniques along with optimisation (conventional method) studies. All the results of these experiments are presented and discussed in further detail in Chapters 4 and 5.

Chapter 4 focuses on the fabrication and the behaviour of the modified anodes in immobilising the bacterial consortium and performance on MFC. In Chapter 5, the optimisation parameters and the evaluation of the physical properties, the decolourisation and power output performance of the variable Nafion membranes in the MFC are clarified. The optimised parameters were then developed into continuous MFC systems with/without a membrane for carrying out biodegradation and electricity



production. One of these systems was subsequently further explored for lower degradation sensitivity.

Lastly, Chapter 6 revolves around the conclusion of the whole study and highlights the major contributions of this work. Furthermore, the chapter also presents some recommendations for future studies.

## REFERENCES

- Abbona, F., Christensson, F., Angela, M. F., and Madsen, H. L. (1993). Crystal habit and growth conditions of brushite,  $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$ . *Journal of crystal growth*, 131(3), 331-346.
- Ab.llah. N. (2012). *Surfactant Modified Clinoptilolite Immobilised With Bacterial Consortium For Efficient and Ecofriendly Decolourisation of Acid Red 27*. Master Thesis. Universiti Teknologi Malaysia, Skudai.
- Aelterman, P., Rabaey, K., Pham, H. T., Boon, N., and Verstraete, W. (2006). Continuous electricity generation at high voltages and currents using stacked microbial fuel cells. *Environmental science and technology*, 40(10), 3388-3394.
- Agnew, D. K. (2006). *Evaluation of Surfactant-Modified Zeolite for the Removal of Giardia Lamblia from Contaminated Waters* (Doctoral dissertation, New Mexico Institute of Mining and Technology).
- Alaton, I. A., Balcioglu, I. A. and Bahnemann, D.W. (2002). Advanced oxidation of a reactive dye bath effluent: comparison of  $\text{O}_3$ ,  $\text{H}_2\text{O}_2/\text{UV-C}$  and  $\text{TiO}_2/\text{UV-A}$  processes. *Water. Res.* 36: 1143-1154.
- Alam, J., Dass, L. A., Alhoshan, M. S., Ghasemi, M., and Mohammad, A.W. (2012). Development of polyaniline-modified polysulfone nanocomposite membrane. *Applied Water Science*, 2(1), 37-46.
- Ali, H., Ahmad, W., and Haq, T. (2009). Decolorization and degradation of malachite green by *Aspergillus flavus* and *Alternaria solani*. *African Journal of Biotechnology*, 8(8).
- Allen, J. B., and Larry, R. F. (2001). *Electrochemical methods: fundamentals and applications*. Department of Chemistry and Biochemistry University of Texas at Austin, John Wiley and Sons, Inc, 156-176.

- Allen, M. S., Welch, K. T., Prebyl, B. S., Baker, D. C., Meyers, A. J., and Sayler, G. S. (2004). Analysis and glycosyl composition of the exopolysaccharide isolated from the floc-forming wastewater bacterium *Thauera* sp. MZ1T. *Environmental microbiology*, 6(8), 780-790.
- Allen, R. M., and Bennetto, H. P. (1993). Microbial fuel-cells. *Applied biochemistry and biotechnology*, 39(1), 27-40.
- Alves, C. F., Melo, L. F., and Vieira, M. J. (2002). Influence of medium composition on the characteristics of a denitrifying biofilm formed by *Alcaligenes denitrificans* in a fluidised bed reactor. *Process Biochemistry*, 37(8), 837-845.
- Angenent, L. T., Karim, K., Al-Dahhan, M. H., Wrenn, B. A., and Domínguez-Espinosa, R. (2004). Production of bioenergy and biochemicals from industrial and agricultural wastewater. *Trends in Biotechnology*, 22(9), 477-485.
- Armağan, B., Özdemir, O., Turan, M., and Celik, M. S. (2003). The removal of reactive azo dyes by natural and modified zeolites. *Journal of Chemical Technology and Biotechnology*, 78(7), 725-732.
- Armbruster, T. (2001). Clinoptilolite-heulandite: applications and basic research. *Studies in Surface Sciences and Catalysis*, 135, 13-27.
- Baerlocher, C., McCusker, L. B., and Olson, D. H. (2007). *Atlas of zeolite framework types*. Elsevier.UK.
- Banat, I.M., Nigam, P., Singh, D. and Marchant, R. (1996). Microbial decolorization of textile-dyecontaining effluents: A review. *Bioresource Technology*, 58, 217-227.
- Beech, I. B., Cheung, C. S., Johnson, D. B., and Smith, J. R. (1996). Comparative studies of bacterial biofilms on steel surfaces using atomic force microscopy and environmental scanning electron microscopy. *Biofouling*, 10(1-3), 65-77.
- Bennetto, H. P., Delaney, G. M., Mason, J. R., Roller, S. D., Stirling, J. L., and Thurston, C. F. (1985). The sucrose fuel cell: efficient biomass conversion using a microbial catalyst. *Biotechnology letters*, 7(10), 699-704.
- Bergel, A., Féron, D., and Mollica, A. (2005). Catalysis of oxygen reduction in PEM fuel cell by seawater biofilm. *Electrochemistry Communications*, 7(9), 900-904.
- Bhatt, N., Patel, K. C., Keharia, H., and Madamwar, D. (2005). Decolourisation of Diazo-Dye Reactive Blue 172 by *Pseudomonas aeruginosa* NBAR12. *Journal of Basic Microbiology*, 45(6), 407-418.

- Bogdanov, B., Georgiev, D., Angelova, K., and Yaneva, K. (2009). Natural Zeolites: Clinoptilolite Review. *International Science Conference*. 4<sup>th</sup>-5<sup>th</sup> June 2009. Bulgaria, 6-11.
- Bond, D. R., and Lovley, D. R. (2003). Electricity production by *Geobacter sulfurreducens* attached to electrodes. *Applied and environmental microbiology*, 69(3), 1548-1555.
- Bond, D. R., and Lovley, D. R. (2005). Evidence for involvement of an electron shuttle in electricity generation by *Geothrix fermentans*. *Applied and environmental microbiology*, 71(4), 2186-2189.
- Bowman, R. S. (2003). Applications of surfactant-modified zeolites to environmental remediation. *Microporous and Mesoporous Materials*, 61(1), 43-56.
- Braissant, O., Decho, A. W., Dupraz, C., Glunk, C., Przekop, K. M., and Visscher, P. T. (2007). Exopolymeric substances of sulfate-reducing bacteria: interactions with calcium at alkaline pH and implication for formation of carbonate minerals. *Geobiology*, 5(4), 401-411.
- Braissant, O., Decho, A. W., Przekop, K. M., Gallagher, K. L., Glunk, C., Dupraz, C., and Visscher, P. T. (2009). Characteristics and turnover of exopolymeric substances in a hypersaline microbial mat. *FEMS Microbiology Ecology*, 67(2), 293-307.
- Bullen, R. A., Arnot, T. C., Lakeman, J. B., and Walsh, F. C. (2006). Biofuel cells and their development. *Biosensors and Bioelectronics*, 21(11), 2015-2045.
- Cao, Y., Hu, Y., Sun, J., and Hou, B. (2010). Explore various co-substrates for simultaneous electricity generation and Congo red degradation in air-cathode single-chamber microbial fuel cell. *Bioelectrochemistry*, 79(1), 71-76.
- Chacko, J. T., and Subramaniam, K. (2011). Enzymatic degradation of azo dyes-a review. *International Journal of Environmental Sciences*. 1(6), 1250-1260.
- Chakraborty, J.N. (2011). *An overview of dye fastness testing*. Clark, M (ED.) *Handbook of Textile and Industrial Dyeing: Principles, Processes and Types of Dyes* 1. (446). USA: Woodhead Publishing.
- Chan, G. F. (2004). *Molecular And Enzymatic Studies on the Decolourization of Azo Dyes by Citrobacter Freundii A1*. PhD Thesis, Universiti Teknologi Malaysia, Skudai.

- Chan, G. F., Rashid, N. A., Koay, L. L., Chang, S. Y., and Tan, W. L. (2011). Identification and optimization of novel NAR-1 bacterial consortium for the biodegradation of Orange II. *Insight Biotechnol*, 1(1), 7-16.
- Chan, G. F., Rashid, N. A. A., Chua, L. S., Nasiri, R., and Ikubar, M. R. M. (2012a). Communal microaerophilic–aerobic biodegradation of Amaranth by novel NAR-2 bacterial consortium. *Bioresource technology*, 105, 48-59.
- Chan, G. F., Gan, H. M., and Rashid, N. A. A. (2012b). Genome sequence of *Citrobacter* sp. strain A1, a dye-degrading bacterium. *Journal of bacteriology*, 194(19), 5485-5486.
- Chan, G. F., Gan, H. M., and Rashid, N. A. A. (2012c). Genome sequence of *Enterococcus* sp. strain C1, an Azo dye decolorizer. *Journal of bacteriology*, 194(20), 5716-5717.
- Chang, J. S., and Kuo, T. S. (2000a). Kinetics of bacterial decolorization of azo dye with *Escherichia coli* NO3. *Bioresource Technology*, 75(2), 107-111.
- Chang, J. S., and Lin, Y. C. (2000b). Fed- Batch Bioreactor Strategies for Microbial Decolorization of Azo Dye Using a *Pseudomonas luteola* Strain. *Biotechnology Progress*, 16(6), 979-985.
- Chang, J. S., Chou, C., and Chen, S. Y. (2001). Decolorization of azo dyes with immobilized *Pseudomonas luteola*. *Process Biochemistry*, 36(8), 757-763.
- Chang, J. S., Chen, B. Y., and Lin, Y. S. (2004). Stimulation of bacterial decolorization of an azo dye by extracellular metabolites from *Escherichia coli* strain NO3. *Bioresource technology*, 91(3), 243-248.
- Chaudhuri, S. K., and Lovley, D. R. (2003). Electricity generation by direct oxidation of glucose in mediatorless microbial fuel cells. *Nature biotechnology*, 21(10), 1229-1232.
- Chen, B. Y., and Chang, J. S. (2007). Assessment upon species evolution of mixed consortia for azo dye decolorization. *Journal of the Chinese Institute of Chemical Engineers*, 38(3), 259-266.
- Chen, B.Y., Zhang, M.M., Chang, C.T., Ding, Y., Lin, K.L., Chiou, C.S., Hsueh, C.C., Xu, H. (2010). Assessment upon azo dye decolorization and bioelectricity generation by *Proteus hauseri*. *Bioresource Technology* 101(12), 4737-4741.
- Chen, B. Y., Wang, Y. M., and Ng, I. S. (2011a). Understanding interactive characteristics of bioelectricity generation and reductive decolorization using *Proteus hauseri*. *Bioresource technology*, 102(2), 1159-1165.

- Chen, J. Y., Li, N., and Zhao, L. (2014). Three-dimensional electrode microbial fuel cell for hydrogen peroxide synthesis coupled to wastewater treatment. *Journal of Power Sources*, 254, 316-322.
- Chen, K. C., Huang, W. T., Wu, J. Y., and Hwang, J. Y. (1999). Microbial decolorization of azo dyes by *Proteus mirabilis*. *Journal of Industrial Microbiology and Biotechnology*, 23(1), 686-690.
- Chen, K. C., Wu, J. Y., Liou, D. J., and Hwang, S. C. J. (2003). Decolorization of the textile dyes by newly isolated bacterial strains. *Journal of Biotechnology*, 101(1), 57-68.
- Chen, S., He, G., Carmona-Martinez, A. A., Agarwal, S., Greiner, A., Hou, H., and Schröder, U. (2011b). Electrospun carbon fiber mat with layered architecture for anode in microbial fuel cells. *Electrochemistry Communications*, 13(10), 1026-1029.
- Chen, Y.M., Wang, C.T., Yang, Y.C., Chen, W.J. (2013). Application of aluminum-alloy mesh composite carbon cloth for the design of anode/cathode electrodes in *Escherichia coli* microbial fuel cell. *International Journal of Hydrogen Energy*, 38(25), 11131-11137.
- Cheng, S., Liu, H., and Logan, B. E. (2006a). Power densities using different cathode catalysts (Pt and CoTMPP) and polymer binders (Nafion and PTFE) in single chamber microbial fuel cells. *Environmental science and technology*, 40(1), 364-369.
- Cheng, S., Liu, H., and Logan, B. E. (2006b). Increased power generation in a continuous flow MFC with advective flow through the porous anode and reduced electrode spacing. *Environmental science and technology*, 40(7), 2426-2432.
- Cheng, S., Liu, H., and Logan, B. E. (2006c). Increased performance of single-chamber microbial fuel cells using an improved cathode structure. *Electrochemistry Communications*, 8(3), 489-494.
- Cheng, S., and Logan, B. E. (2007). Ammonia treatment of carbon cloth anodes to enhance power generation of microbial fuel cells. *Electrochemistry Communications*, 9(3), 492-496.
- Chengalroyen, M. D., and Dabbs, E. R. (2013). The microbial degradation of azo dyes: minireview. *World Journal of Microbiology and Biotechnology*, 29(3), 389-399.

- Choi, Y., Kim, N., Kim, S., and Jung, S. (2003). Dynamic behaviors of redox mediators within the hydrophobic layers as an important factor for effective microbial fuel cell operation. *Bulletin-Korean Chemical Society*, 24(4), 437-440.
- Choi, Y., Jung, E., Park, H., Paik, S. R., Jung, S., and Kim, S. (2004). Construction of microbial fuel cells using thermophilic microorganisms, *Bacillus licheniformis* and *Bacillus thermoglucosidasius*. *Bulletin-Korean Chemical Society*, 25(6), 813-818.
- Costa, M. C., Mota, F. S. B., Santos, A. B. D., Mendonça, G. L. F., and Nascimento, R. F. D. (2012). Effect of dye structure and redox mediators on anaerobic azo and anthraquinone dye reduction. *Quimica Nova*, 35(3), 482-486.
- Coughlin, M. F., Kinkle, B. K., and Bishop, P. L. (2002). Degradation of acid orange 7 in an aerobic biofilm. *Chemosphere*, 46(1), 11-19.
- Cui, D., Li, G., Zhao, D., Gu, X., Wang, C., and Zhao, M. (2012). Microbial community structures in mixed bacterial consortia for azo dye treatment under aerobic and anaerobic conditions. *Journal of hazardous materials*, 221, 185-192.
- Dafale, N., Agrawal, L., Kapley, A., Meshram, S., Purohit, H., and Wate, S. (2010). Selection of indicator bacteria based on screening of 16S rDNA metagenomic library from a two-stage anoxic-oxic bioreactor system degrading azo dyes. *Bioresource Technology*, 101, 476-484.
- Dalgaard, P., and Koutsoumanis, K. (2001). Comparison of maximum specific growth rates and lag times estimated from absorbance and viable count data by different mathematical models. *Journal of Microbiological Methods*, 43(3), 183-196.
- Daneshvar, N., Rabbani, M., Modirshahla, N., and Behnajady, M. A. (2004). Critical effect of hydrogen peroxide concentration in photochemical oxidative degradation of CI Acid Red 27 (AR27). *Chemosphere*, 56(10), 895-900.
- Daniel, D. K., Das Mankidy, B., Ambarish, K., and Manogari, R. (2009). Construction and operation of a microbial fuel cell for electricity generation from wastewater. *International journal of hydrogen energy*, 34(17), 7555-7560.
- Dawkar, V. V., Jadhav, U. U., Ghodake, G. S., and Govindwar, S. P. (2009). Effect of inducers on the decolorization and biodegradation of textile azo dye Navy blue 2GL by *Bacillus* sp. VUS. *Biodegradation*, 20(6), 777-787.

- Devanathan, R. (2008). Recent developments in proton exchange membranes for fuel cells. *Energy and Environmental Science*, 1(1), 101-119.
- Deng, Q., Li, X., Zuo, J., Ling, A., and Logan, B. E. (2010). Power generation using an activated carbon fiber felt cathode in an upflow microbial fuel cell. *Journal of Power Sources*, 195(4), 1130-1135.
- Dignac, M. F., Urbain, V., Rybacki, D., Bruchet, A., Snidaro, D., and Scribe, P. (1998). Chemical description of extracellular polymers: implication on activated sludge floc structure. *Water Science and Technology*, 38(8-9), 45-53.
- Domenek, S., Feuilloley, P., Gratraud, J., Morel, M. H., and Guilbert, S. (2004). Biodegradability of wheat gluten based bioplastics. *Chemosphere*, 54(4), 551-559.
- Donlan, R. M. (2002). Biofilms: microbial life on surfaces. *Emerging Infectious Diseases*, 8(9).
- Dos Santos, A. B., Traverse, J., Cervantes, F. J., and Van Lier, J. B. (2005). Enhancing the electron transfer capacity and subsequent color removal in bioreactors by applying thermophilic anaerobic treatment and redox mediators. *Biotechnology and Bioengineering*, 89(1), 42-52.
- Dos Santos, A. B., Marta, P., De Bok, F. A., Stams, A. J., Van Lier, J. B., and Cervantes, F. J. (2006). The contribution of fermentative bacteria and methanogenic archaea to azo dye reduction by a thermophilic anaerobic consortium. *Enzyme and microbial technology*, 39(1), 38-46.
- Dos Santos, A. B., Cervantes-Carillo, F. J. and van Lier, J. B. (2007). Review paper on current technologies for decolourisation of textile wastewaters: perspectives for anaerobic biotechnology. *Bioresource Technology*, 98, 2369-2385.
- Du, Z., Li, H., and Gu, T. (2007). A state of the art review on microbial fuel cells: a promising technology for wastewater treatment and bioenergy. *Biotechnology advances*, 25(5), 464-482.
- Dufrêne, Y. F., and Rouxhet, P. G. (1996). X-ray photoelectron spectroscopy analysis of the surface composition of *Azospirillum brasilense* in relation to growth conditions. *Colloids and Surfaces B: Biointerfaces*, 7(5), 271-279.
- Durham, D. R., Marshall, L. C., Miller, J. G. and Chmurn, A. B. (1994). New Composite Biocarriers Engineered To Contain Adsorptive and Ion-Exchange Properties Improve Immobilized-Cell Bioreactor Process Dependability. *Appl. Environ. Microbiol.* November; 60(11), 4178-4181.



- Eaktasang, N., Kang, C. S., Ryu, S. J., Suma, Y., and Kim, H. S. (2013). Enhanced current production by electroactive biofilm of sulfate-reducing bacteria in the microbial fuel cell. *Environmental Engineering Research*, 18(4), 277-281.
- Ersoy, B., and Çelik, M. S. (2003). Effect of hydrocarbon chain length on adsorption of cationic surfactants onto clinoptilolite. *Clays and Clay Minerals*, 51(2), 172-180.
- Esparza-Soto, M., and Westerhoff, P. K. (2001). Fluorescence spectroscopy and molecular weight distribution of extracellular polymers from full-scale activated sludge biomass. *Water Science and Technology*, 43(6), 87-95.
- Faghihian, H., Kabiri-Tadi, M., and Ahmadi, S. J. (2010). Adsorption of <sup>103</sup>Ru from aqueous solutions by clinoptilolite. *Journal of radioanalytical and nuclear chemistry*, 285(3), 499-504.
- Fang, Z., Song, H. L., Cang, N., and Li, X. N. (2015). Electricity production from Azo dye wastewater using a microbial fuel cell coupled constructed wetland operating under different operating conditions. *Biosensors and Bioelectronics*, 68, 135-141.
- Farías, T., De Menorval, L. C., Zajac, J., and Rivera, A. (2011). Benzalkonium chloride and sulfamethoxazole adsorption onto natural clinoptilolite: Effect of time, ionic strength, pH and temperature. *Journal of colloid and interface science*, 363(2), 465-475.
- Feng, Y. J., and Li, X. Y. (2003). Electro-catalytic oxidation of phenol on several metal-oxide electrodes in aqueous solution. *Water Research*, 37(10), 2399-2407.
- Fiset, E., and Puig, S. (2015). Modified carbon electrodes: a new approach for bioelectrochemical systems. *J. Bioremediation Biodegrad*, 6, 1-2.
- Fletcher, M. (1977). The effects of culture concentration and age, time, and temperature on bacterial attachment to polystyrene. *Canadian journal of microbiology*, 23(1), 1-6.
- Forgacs, E., Cserháti, T., Oros, G. (2004). Removal of synthetic dyes from wastewaters: a review. *Environment International*, 30, 953–971.
- Francois, K., Devlieghere, F., Standaert, A.R., Geeraerd, A.H., Cools, I., Van Impe, J.F. and Debevere, J. (2005). Environmental factors influencing the relationship between optical density and cell count for *Listeria monocytogenes*. *Journal of Appl. Microbiol.*, 99(6), 1503–1515

- Fricke, K., Harnisch, F., and Schröder, U. (2008). On the use of cyclic voltammetry for the study of anodic electron transfer in microbial fuel cells. *Energy and Environmental Science*, 1(1), 144-147.
- Galarneau, A., Di Renzo, F., Fajula, F., and Vedrine, J. eds. (2001). *Zeolites and Mesoporous Materials at the Dawn of the 21st Century*. Elsevier Science.
- Gedikoglu, Y., Gedikoglu, G., Berkin, G., Ceyhan, T., and Altinoz, M. A. (2012). Employing volcanic tuff minerals in interior architecture design to reduce microbial contaminants and airborne fungal carcinogens of indoor environments. *Toxicology and industrial health*, 28(8),708-719.
- Ghasemi, M., Daud, W. R. W., Ismail, M., Rahimnejad, M., Ismail, A. F., Leong, J. X., Miskan, M., and Liew, K. B. (2013a). Effect of pre-treatment and biofouling of proton exchange membrane on microbial fuel cell performance. *International journal of hydrogen energy*, 38(13), 5480-5484.
- Ghasemi, M., Rahimnejad, M., Esmaili, C., Daud, W. R., Alam, J., and Alhoshan, M. (2013b). Polysulfone composed of polyaniline nanoparticles as nanocomposite proton exchange membrane in microbial fuel cell. *American Journal of Biochemistry and Biotechnology*, 4(1), 19-27.
- Ghiaci, M., Abbaspur, A., Kia, R., and Seyedeyn-Azad, F. (2004). Equilibrium isotherm studies for the sorption of benzene, toluene, and phenol onto organo-zeolites and as-synthesized MCM-41. *Separation and purification technology*, 40(3), 217-229.
- Gil, G. C., Chang, I. S., Kim, B. H., Kim, M., Jang, J. K., Park, H. S., and Kim, H. J. (2003). Operational parameters affecting the performance of a mediator-less microbial fuel cell. *Biosensors and Bioelectronics*, 18(4), 327-334.
- Giltner, C. L., Van Schaik, E. J., Audette, G. F., Kao, D., Hodges, R. S., Hassett, D. J., and Irvin, R. T. (2006). The *Pseudomonas aeruginosa* type IV pilin receptor binding domain functions as an adhesin for both biotic and abiotic surfaces. *Molecular microbiology*, 59(4), 1083-1096.
- Ginsburg, M.A., and Karamanev, D. (2007). Experimental study of the immobilization of *Acidithiobacillus ferrooxidans* on carbon based supports. *Biochemical Engineering Journal*, 36(3), 294-300.
- Gorby, Y. A., Yanina, S., McLean, J. S., Rosso, K. M., Moyles, D., Dohnalkova, A., Beveridge, T. J., Chang, I. S., Kim, B. H., Kim, K. S., Culley, D. E., Reed, S. B., Romine, M. F., Saffarini, D.A., Hill, E. A., Shi, L., Elias, D. A., Kennedy,

- D. W., Pinchuk, G., Watanabe, K., Ishii, S., Logan, B., Neals, K. H., Culley, D. E. (2006). Electrically conductive bacterial nanowires produced by *Shewanella oneidensis* strain MR-1 and other microorganisms. *Proceedings of the National Academy of Sciences*, 103(30), 11358-11363.
- Gouda, M. K. (2002). Decolorization of some textile dyes by *Aspergillus terreus*. *African Journal of Mycology and Biotechnology*, 10(1), 1-14.
- Habermann, W., and Pommer, E. H. (1991). Biological fuel cells with sulphide storage capacity. *Applied microbiology and biotechnology*, 35(1), 128-133.
- Hart, J. P., Abass, A. K., Honeychurch, K. C., Pemberton, R. M., Ryan, S. L., and Wedge, R. (2003). Sensors/biosensors, based on screen-printing technology for biomedical applications. *Indian Journal of Chemistry Section A*, 42(4), 709-718.
- Hashemi, J., and Samimi, A. (2012). Steady state electric power generation in up-flow microbial fuel cell using the estimated time span method for bacteria growth domestic wastewater. *Biomass and Bioenergy*, 45, 65-76.
- Haug, W., Schmidt, A., Nortemann, B., Hempel, D.C., Stolz, A. and Knackmuss, H.J. (1991). "Mineralization of the sulfonated azo dye Mordant Yellow 3 by a 6-aminonaphthalene-2-sulfonate-degrading bacterial consortium". *Applied Environmental Microbiology*, 57, 3144-3149.
- He, Z. (2007). *Microbial fuel cells: their application and microbiology*. PhD Thesis, Washington University, Saint Louis.
- He, Z., Minteer, S. D., and Angenent, L. T. (2005). Electricity generation from artificial wastewater using an upflow microbial fuel cell. *Environmental science and technology*, 39(14), 5262-5267.
- Helder, M., Strik, D. P. B. T. B., Hamelers, H. V. M., Kuyken, R. C. P., and Buisman, C. J. N. (2012). New plant-growth medium for increased power output of the plant-microbial fuel cell. *Bioresour. Technol.*, 104, 417-423.
- Higgins, S. R., Lau, C., Atanassov, P., Minteer, S. D., and Cooney, M. J. (2011). Hybrid biofuel cell: Microbial fuel cell with an enzymatic air-breathing cathode. *ACS Catalysis*, 1(9), 994-997.
- Hrenović, J., Tibljaš D., Orhan, Y. and Büyükgüngör, H. (2005). Immobilisation of *Acinetobacter calcoaceticus* using natural carriers. *Water SA*. 31(2), 261-266.

- Hrenovic, J., Rozic, M., Sekovanic, L., and Anic-Vucinic, A. (2008). Interaction of surfactant-modified zeolites and phosphate accumulating bacteria. *Journal of hazardous materials*, 156(1), 576-582.
- Hrenovic, J., Rozic, M., Ivankovic, T., and Farkas, A. (2009). Biosorption of phosphate from synthetic wastewater by biosolids. *Open Life Sciences*, 4(3), 397-403
- Hrenovic, J., Tibljas, D., Ivankovic, T., Kovacevic, D., and Sekovanic, L. (2010). Sepiolite as carrier of the phosphate-accumulating bacteria *Acinetobacter junii*. *Applied clay science*, 50(4), 582-587.
- Hrenovic, J., Kovacevic, D., Ivankovic, T. and Tibljas, D. (2011). Selective immobilization of *Acinetobacter junii* on the natural zeolitized tuff in municipal wastewater. *Colloids and Surfaces B: Biointerfaces*, 88(1), 208-214.
- Hong, Y., Guo, J., Xu, Z., Mo, C., Xu, M. and Sun, G. (2007). Reduction and partial degradation mechanisms of naphthylaminesulfonic azo dye Amaranth by *Shewanella decolorationis* S12. *Applied Microbiology and Biotechnology*, 42, 44-55.
- Hou, B., Sun, J., and Hu, Y. Y. (2011a). Simultaneous Congo red decolorization and electricity generation in air-cathode single-chamber microbial fuel cell with different microfiltration, ultrafiltration and proton exchange membranes. *Bioresource technology*, 102(6), 4433-4438.
- Hou, B., Sun, J., and Hu, Y. (2011b). Effect of enrichment procedures on performance and microbial diversity of microbial fuel cell for Congo red decolorization and electricity generation. *Applied microbiology and biotechnology*, 90(4), 1563-1572.
- Huang, L., Zeng, R. J., and Angelidaki, I. (2008). Electricity production from xylose using a mediator-less microbial fuel cell. *Bioresource technology*, 99(10), 4178-4184.
- Huang, T. S., Tzeng, Y., Liu, Y. K., Chen, Y. C., Walker, K. R., Guntupalli, R., and Liu, C. (2004). Immobilization of antibodies and bacterial binding on nanodiamond and carbon nanotubes for biosensor applications. *Diamond and related materials*, 13(4), 1098-1102.
- Humudat, Y. R., Al-Naseri, S. K., and Kadhim, S. A. (2015). Using Bacteria in Microbial Fuel Cells. *Int. J. Pure App. Biosci*, 3(5), 21-25.

- Ibrahim, N. (2012). *Sensor Innovations Based on Modified Carbon Electrodes*. PhD Thesis, University of Bath, Bath.
- Idel-aouad, R., Valiente, M., Yaacoubi, A., Tanouti, B. and Lopez-Mesas, M. (2011). Rapid decolourization and mineralization of the azo dye C.I. Acid Red 14 by heterogeneous Fenton reaction. *Journal of Hazardous Materials*, 186,745-750.
- Ikubar, M. R. M. (2010). *Decolourisanon of amaranth by bacterial consortium immobilised onto modified zeolite nax*. PhD Thesis, Universiti Teknologi Malaysia, Skudai.
- Jadhav, S. B., Patil, N. S., Watharkar, A. D., Apine, O. A., Jadhav, J. P. (2013). Batch and continuous biodegradation of Amaranth in plain distilled water by *P. aeruginosa* BCH and toxicological scrutiny using oxidative stress studies. *Environ. Sci. Pollut. Res.* 20(5), 2854-2866.
- Jadhav, U. U., Dawkar, V. V., Ghodake, G. S., and Govindwar, S. P. (2008). Biodegradation of Direct Red 5B, a textile dye by newly isolated *Comamonas* sp. UVS. *Journal of Hazardous Materials*, 158(2), 507-516.
- Jang, J. K., Pham, T. H., Chang, I. S., Kang, K. H., Moon, H., Cho, K. S., and Kim, B. H. (2004). Construction and operation of a novel mediator-and membrane-less microbial fuel cell. *Process Biochemistry*, 39(8), 1007-1012.
- Jiang, D., Huang, Q., Cai, P., Rong, X., and Chen, W. (2007). Adsorption of *Pseudomonas putida* on clay minerals and iron oxide. *Colloids and Surfaces B: Biointerfaces*, 54(2), 217-221.
- Jong, B. C., Kim, B. H., Chang, I. S., Liew, P. W. Y., Choo, Y. F., and Kang, G. S. (2006). Enrichment, performance, and microbial diversity of a thermophilic mediatorless microbial fuel cell. *Environmental Science and Technology*, 40(20), 6449-6454.
- Jonstrup, M., Punzi, M. and Mattiasson, B. (2011). Comparison of anaerobic pretreatment and aerobic post-treatment coupled to photo-Fenton oxidation for degradation of azo dyes. *Journal of Photochemistry and Photobiology A: Chemistry*, 224(1), 55-61.
- Joshi, P., Rayalu, S., Bansiwala, A., and Juwarkar, A. A. (2007). Surface modified zeolite, a novel carrier material for *Azotobacter chroococcum*. *Plant and soil*, 296(1-2), 151-158.

- Joshi, T., Iyengar, L., Singh, K., Garg, S. (2008). Isolation, identification and application of novel bacterial consortium TJ-1 for the decolorization of structurally different azo dyes. *Bioresour. Technol.* 99, 7115-7121.
- Jung, S., and Regan, J. M. (2007). Comparison of anode bacterial communities and performance in microbial fuel cells with different electron donors. *Applied microbiology and biotechnology*, 77(2), 393-402.
- Jović, M., Stanković, D., Manojlović, D., Anđelković, I., Milić, A., Dojčinović, B., and Roglić, G. (2013). Study of the electrochemical oxidation of reactive textile dyes using platinum electrode. *Int. J. Electrochem. Sci.*, 8(1), 168-183.
- Kalyani, D. C., Telke, A. A., Dhanve, R. S., and Jadhav, J. P. (2009). Ecofriendly biodegradation and detoxification of Reactive Red 2 textile dye by newly isolated *Pseudomonas* sp. SUK1. *Journal of Hazardous Materials*, 163(2), 735-742.
- Kang, J., Kim, T., Tak, Y., Lee, J. H., and Yoon, J. (2012). Cyclic voltammetry for monitoring bacterial attachment and biofilm formation. *Journal of Industrial and Engineering Chemistry*, 18(2), 800-807.
- Karadag, D. (2007). Modeling the mechanism, equilibrium and kinetics for the adsorption of Acid Orange 8 onto surfactant-modified clinoptilolite: The application of nonlinear regression analysis. *Dyes and Pigments*, 74(3), 659-664.
- Kardi, S. N. *Biodegradation of Remazol Black B in sequential microaerophilic-aerobic operations by NAR-2 bacterial consortium.* (2013) Master Thesis, Universiti Teknologi Malaysia, Skudai.
- Karkmaz, M., Puzenat, E., Guillard, C., Herrmann, J.M., (2004). "Photocatalytic degradation of the alimentary azo dye amaranth, Mineralization of the azo group to nitrogen." *Applied Catalysis B: Environmental* 51, 183-194.
- Karube, I., Matsunaga, T., Mitsuda, S., and Suzuki, S. (1977). Microbial electrode BOD sensors. *Biotechnology and bioengineering*, 19(10), 1535-1547.
- Kasinath, A., Novotný, Č., Svobodová, K., Patel, K. C., and Šásek, V. (2003). Decolorization of synthetic dyes by *Irpex lacteus* in liquid cultures and packed-bed bioreactor. *Enzyme and Microbial Technology*, 32(1), 167-173.
- Ke, C., Yang, W. L., Ni, Z., Wang, Y. J., Tang, Y., Gu, Y., and Gao, Z. (2001). Electrophoretic assembly of nanozeolites: zeolite coated fibers and hollow zeolite fibers. *Chemical Communications*, (8), 783-784.

- Khan, R., and Banerjee, U. C. (2010). Decolorization of azo dyes by immobilized bacteria. *Biodegradation of Azo Dyes Springer*. 73-84.
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S., Chimni, S.S. (2005). Comparative studies on potential of consortium and constituent pure bacterial isolates to decolorize azo dyes. *Water Res.* 39, 5135–5141.
- Khehra, M. S., Saini, H. S., Sharma, D. K., Chadha, B. S., and Chimni, S. S. (2006). Biodegradation of azo dye CI Acid Red 88 by an anoxic–aerobic sequential bioreactor. *Dyes and Pigments*, 70(1), 1-7.
- Khodakov, A. Y., Kustov, L. M., Kazansky, V. B., and Williams, C. (1993). Infrared spectroscopic study of the interaction of cations in zeolites with simple molecular probes. Part 3.—Adsorption and polarization of methane and ethane on cationic forms of high-silica zeolites. *Journal of the Chemical Society, Faraday Transactions*, 89(9), 1393-1395.
- Kim, B. H., Chang, I. S., Gil, G. C., Park, H. S., and Kim, H. J. (2003). Novel BOD (biological oxygen demand) sensor using mediator-less microbial fuel cell. *Biotechnology letters*, 25(7), 541-545.
- Kim, B. H., Park, H. S., Kim, H. J., Kim, G. T., Chang, I. S., Lee, J., and Phung, N. T. (2004). Enrichment of microbial community generating electricity using a fuel-cell-type electrochemical cell. *Applied microbiology and biotechnology*, 63(6), 672-681.
- Kim, H. J., Hyun, M. S., Chang, I. S., and Kim, B. H. (1999). A microbial fuel cell type lactate biosensor using a metal-reducing bacterium, *Shewanella putrefaciens*. *Journal of Microbiology and Biotechnology*, 9(3), 365-367.
- Kim, J. R., Cheng, S., Oh, S. E., and Logan, B. E. (2007). Power generation using different cation, anion, and ultrafiltration membranes in microbial fuel cells. *Environmental science and technology*, 41(3), 1004-1009.
- Kim, N. J., Choe, Y. J., Jeong, S. H., and Kim, S. H. (2000). Development of microbial fuel cells using *Proteus vulgaris*. *Bulletin of the Korean Chemical Society*, 21(1), 44-48.
- Kirby, N., Marchant, R., and McMullan, G. (2000). Decolourisation of synthetic textile dyes by *Phlebia tremellosa*. *FEMS Microbiology Letters*, 188(1), 93-96.
- Knapp, J. S., and Newby, P. S. (1995). The microbiological decolorization of an industrial effluent containing a diazo-linked chromophore. *Water research*, 29(7), 1807-1809.

- Koay, L. L. (2004). Optimization of Orange II Biodegradation and Detection of Products of Biodegradation by Bacterium C1 and L17. Undergraduate Thesis, Universiti Teknologi Malaysia, Skudai.
- Kulla, G. H., Klausener, F., Meyer, U., Ludeke, B. and Leisinger, T. (1983). Interference of aromatic sulfo groups in the microbial degradation of the azo dyes Orange. *Archives of Microbiology*. 135, 1-7.
- Kumar, K., Devi, S.S., Krishnamurthi, K., Dutta, D., Chakrabarti, T. (2007). Decolorisation and detoxification of Direct Blue-15 by a bacterial consortium. *Bioresource Technology*, 98, 3168-3171.
- Kyotani, T., Sato, K., Mizuno, T., Kakui, S., Aizawa, M., Saito, J., Ikeda, S., Ichikawa, S., and NAKANE, T. (2005). Characterization of zeolite NaA membrane by FTIR-ATR and its application to the rapid evaluation of dehydration performance. *Analytical sciences*, 21(3), 321-325.
- Lade, H. S., Waghmode, T. R., Kadam, A. A., and Govindwar, S. P. (2012). Enhanced biodegradation and detoxification of disperse azo dye Rubine GFL and textile industry effluent by defined fungal-bacterial consortium. *International Biodeterioration and Biodegradation*, 72, 94-107.
- Lanas, V., Ahn, Y., and Logan, B. E. (2014). Effects of carbon brush anode size and loading on microbial fuel cell performance in batch and continuous mode. *Journal of Power Sources*, 247, 228-234.
- Larrosa-Guerrero, A., Scott, K., Katuri, K. P., Godinez, C., Head, I. M., and Curtis, T. (2010). Open circuit versus closed circuit enrichment of anodic biofilms in MFC: effect on performance and anodic communities. *Applied microbiology and biotechnology*, 87(5), 1699-1713.
- Lattner, D., Flemming, H. C., and Mayer, C. (2003). <sup>13</sup>C-NMR study of the interaction of bacterial alginate with bivalent cations. *International journal of biological macromolecules*, 33(1), 81-88.
- Lebedynets, M., Sprynskyy, M., Sakhnyuk, I., Zbytniewski, R., Golembiewski, R., and Buszewski, B. (2004). Adsorption of ammonium ions onto a natural zeolite: transcarpathian clinoptilolite. *Adsorption Science and Technology*, 22(9), 731-741.
- Lee, S. Y., Cho, W. J., Kim, K. J., Ahn, J. H., and Lee, M. (2005). Interaction between cationic surfactants and montmorillonites under nonequilibrium condition. *Journal of colloid and interface science*, 284(2), 667-673.



- Lefebvre, O., Tan, Z., Shen, Y., and Ng, H. Y. (2013). Optimization of a microbial fuel cell for wastewater treatment using recycled scrap metals as a cost-effective cathode material. *Bioresource technology*, 127, 158-164.
- Lembre, P., Lorentz, C., Di Martino, P., and Di Martino, P. (2012). *Exopolysaccharides of the biofilm matrix: A complex biophysical world*. INTECH Open Access Publisher.
- Leong, J. X., Daud, W. R. W., Ghasemi, M., Liew, K. B., and Ismail, M. (2013). Ion exchange membranes as separators in microbial fuel cells for bioenergy conversion: a comprehensive review. *Renewable and Sustainable Energy Reviews*, 28, 575-587.
- Leung, S., Barrington, S., Wan, Y., Zhao, X., and El-Husseini, B. (2007). Zeolite (clinoptilolite) as feed additive to reduce manure mineral content. *Bioresource technology*, 98(17), 3309-3316.
- Li, C., Zhang, L., Ding, L., Ren, H., and Cui, H. (2011a). Effect of conductive polymers coated anode on the performance of microbial fuel cells (MFCs) and its biodiversity analysis. *Biosensors and Bioelectronics*, 26(10), 4169-4176.
- Li, D. H., and Ganczarczyk, J. J. (1990). Structure of activated sludge flocs. *Biotechnology and bioengineering*, 35(1), 57-65.
- Li, T., Fang, Z., Yu, R., Cao, X., Song, H., and Li, X. (2016). The performance of the microbial fuel cell-coupled constructed wetland system and the influence of the anode bacterial community. *Environmental technology*, 37(13), 1683-1692.
- Li, W. W., Sheng, G. P., Liu, X. W., and Yu, H. Q. (2011b). Recent advances in the separators for microbial fuel cells. *Bioresource technology*, 102(1), 244-252.
- Li, X., and Logan, B. E. (2004). Analysis of bacterial adhesion using a gradient force analysis method and colloid probe atomic force microscopy. *Langmuir*, 20(20), 8817-8822.
- Li, Z. (1999). Sorption kinetics of hexadecyltrimethylammonium on natural clinoptilolite. *Langmuir*, 15(19), 6438-6445.
- Li, Z., Alessi, D., and Allen, L. (2002). Influence of quaternary ammonium on sorption of selected metal cations onto clinoptilolite zeolite. *Journal of environmental quality*, 31(4), 1106-1114.

- Li, Z., Willms, C., Roy, S., and Bowman, R. S. (2003). Desorption of hexadecyltrimethylammonium from charged mineral surfaces. *Environmental Geosciences*, 10(1), 37-45.
- Li, Z., Zhang, X., Lin, J., Han, S., and Lei, L. (2010). Azo dye treatment with simultaneous electricity production in an anaerobic–aerobic sequential reactor and microbial fuel cell coupled system. *Bioresource technology*, 101(12), 4440-4445.
- Lin, J., Zhang, X., Li, Z., and Lei, L. (2010). Biodegradation of Reactive blue 13 in a two-stage anaerobic/aerobic fluidized beds system with a *Pseudomonas* sp. isolate. *Bioresource technology*, 101(1), 34-40.
- Liu, H., Ramnarayanan, R., and Logan, B. E. (2004). Production of electricity during wastewater treatment using a single chamber microbial fuel cell. *Environmental science & technology*, 38(7), 2281-2285.
- Liu, H., and Logan, B. E. (2004). Electricity generation using an air-cathode single chamber microbial fuel cell in the presence and absence of a proton exchange membrane. *Environmental science and technology*, 38(14), 4040-4046.
- Liu, H., Cheng, S., and Logan, B. E. (2005). Production of electricity from acetate or butyrate using a single-chamber microbial fuel cell. *Environmental science and technology*, 39(2), 658-662.
- Liu, Y., and Fang, H. H. (2003). Influences of extracellular polymeric substances (EPS) on flocculation, settling, and dewatering of activated sludge.
- Logan, B. E., Hamelers, B., Rozendal, R., Schröder, U., Keller, J., Freguia, S., Aelterman, P., Verstraete, W., and Rabaey, K. (2006). Microbial fuel cells: methodology and technology. *Environmental science and technology*, 40(17), 5181-5192.
- Logan, B., Cheng, S., Watson, V., and Estadt, G. (2007). Graphite fiber brush anodes for increased power production in air-cathode microbial fuel cells. *Environmental science and technology*, 41(9), 3341-3346.
- López-Grimau, V., and Gutierrez, M. C. (2006). Decolourisation of simulated reactive dyebath effluents by electrochemical oxidation assisted by UV light. *Chemosphere*, 62(1), 106-112.
- Lovley, D. R., Stolz, J. F., Nord, G. L., and Phillips, E. J. (1987). Anaerobic production of magnetite by a dissimilatory iron-reducing microorganism. *Nature*, 330(6145), 252-254.

- Lovley, D. R. (2006a). Microbial fuel cells: novel microbial physiologies and engineering approaches. *Current opinion in biotechnology*, 17(3), 327-332.
- Lovley, D. R. (2006b). Bug juice: harvesting electricity with microorganisms. *Nature Reviews Microbiology*, 4(7), 497-508.
- Lowy, D. A., Tender, L. M., Zeikus, J. G., Park, D. H., and Lovley, D. R. (2006). Harvesting energy from the marine sediment–water interface II: kinetic activity of anode materials. *Biosensors and Bioelectronics*, 21(11), 2058-2063.
- Luckarift, H. R., Sizemore, S. R., Roy, J., Lau, C., Gupta, G., Atanassov, P., and Johnson, G. R. (2010). Standardized microbial fuel cell anodes of silica-immobilized *Shewanella oneidensis*. *Chemical Communications*, 46(33), 6048-6050.
- Luo, H., Liu, G., Zhang, R., and Jin, S. (2009). Phenol degradation in microbial fuel cells. *Chemical Engineering Journal*, 147(2), 259-264.
- Mardanpour, M. M., Esfahany, M. N., Behzad, T., and Sedaqatvand, R. (2012). Single chamber microbial fuel cell with spiral anode for dairy wastewater treatment. *Biosensors and Bioelectronics*, 38(1), 264-269.
- Margeta, K., Logar, N. Z., Šiljeg, M., and Farkaš, A. (2013). Natural Zeolites in Water Treatment—How Effective is Their Use.
- Mattick, J. S. (2002). Type IV pili and twitching motility. *Annual Reviews in Microbiology*, 56(1), 289-314.
- McMullan, G., Meehan, C., Conneely, A., Kirby, N., Robinson, T., Nigam, P., Banat, I., Marchant, R., and Smyth, W. F. (2001). Microbial decolourisation and degradation of textile dyes. *Applied Microbiology and Biotechnology*, 56(1-2), 81-87.
- Mendes, S., Farinha, A., Ramos, C.G., Leitão, J. H., Viegas, C.A., and Martins, L.O.(2011). Synergistic action of azoreductase and laccase leads to maximal decolourization and detoxification of model dye-containing wastewaters, *Bioresource Technology*, 102, 9852-9859.
- Méndez-Paz, D., Omil, F. and Lema, J. M. (2005). Anaerobic treatment of azo dye Acid Orange 7 under batch conditions. *Enzyme and Microbial Technology*. 36(2-3), 264-272.
- Mendoza-Barrón, J., Jacobo-Azuara, A., Leyva-Ramos, R., Berber-Mendoza, M. S., Guerrero-Coronado, R. M., Fuentes-Rubio, L., and Martínez-Rosales, J. M.

- (2011). Adsorption of arsenic (V) from a water solution onto a surfactant-modified zeolite. *Adsorption*, 17(3), 489-496.
- Miao, Y. (2005). Biological remediation of dyes in textile effluent: a review on current treatment technologies.
- Min, B., and Logan, B. E. (2004). Continuous electricity generation from domestic wastewater and organic substrates in a flat plate microbial fuel cell. *Environmental science and technology*, 38(21), 5809-5814.
- Min, B., Cheng, S., and Logan, B. E. (2005a). Electricity generation using membrane and salt bridge microbial fuel cells. *Water research*, 39(9), 1675-1686.
- Min, B., Kim, J., Oh, S., Regan, J. M., and Logan, B. E. (2005b). Electricity generation from swine wastewater using microbial fuel cells. *Water research*, 39(20), 4961-4968.
- Mi-Na, Z. H. A. N. G., Xue-Pin, L., and Bi, S. (2006). Adsorption of surfactants on chromium leather waste. *J. Soc. Leather Technol. Chem.* 90, (1),1.
- Mittal, A., Kurup, L., and Gupta, V. K. (2005). Use of waste materials—bottom ash and de-oiled soya, as potential adsorbents for the removal of amaranth from aqueous solutions. *Journal of Hazardous Materials*, 117(2), 171-178.
- Mohan, S. V., Saravanan, R., Raghavulu, S. V., Mohanakrishna, G., and Sarma, P. N. (2008a). Bioelectricity production from wastewater treatment in dual chambered microbial fuel cell (MFC) using selectively enriched mixed microflora: effect of catholyte. *Bioresource Technology*, 99(3), 596-603.
- Mohan, S. V., Mohanakrishna, G., Reddy, B. P., Saravanan, R., and Sarma, P. N. (2008b). Bioelectricity generation from chemical wastewater treatment in mediatorless (anode) microbial fuel cell (MFC) using selectively enriched hydrogen producing mixed culture under acidophilic microenvironment. *Biochemical Engineering Journal*, 39(1), 121-130.
- Mohan, S. V., Mohanakrishna, G., Srikanth, S., and Sarma, P. N. (2008c). Harnessing of bioelectricity in microbial fuel cell (MFC) employing aerated cathode through anaerobic treatment of chemical wastewater using selectively enriched hydrogen producing mixed consortia. *Fuel*, 87(12), 2667-2676.
- Mohana, S., Shrivastava, S., Divecha, J., and Madamwar, D. (2008d). Response surface methodology for optimization of medium for decolorization of textile dye Direct Black 22 by a novel bacterial consortium. *Bioresource Technology*, 99(3), 562-569.

- Molin, S., Nielsen, A. T., Christensen, B. B., Andersen, J. B., Licht, T. R., Tolker-Nielsen, T., Sternberg, C., Hansen, M. C., Ramos, C. and Givskov, M. (2000) Molecular ecology of biofilms (ed. J.D. Bryers), *Biofilms II: Process Analysis and Applications*, Wiley Series in Ecological and Applied Microbiology (series ed. R. Mitchell, pp. 89-120, A John Wiley and Sons Inc., New York, USA.
- Moon, H., Chang, I. S., Jang, J. K., and Kim, B. H. (2005). Residence time distribution in microbial fuel cell and its influence on COD removal with electricity generation. *Biochemical Engineering Journal*, 27(1), 59-65.
- Moosvi, S., Kher, X., and Madamwar, D. (2007). Isolation, characterization and decolorization of textile dyes by a mixed bacterial consortium JW-2. *Dyes and pigments*, 74(3), 723-729.
- Mozgawa, W., Król, M., and Bajda, T. (2011). IR spectra in the studies of anion sorption on natural sorbents. *Journal of Molecular Structure*, 993(1), 109-114.
- Murali, V., Ong, S. A., Ho, L. N., Wong, Y. S., and Hamidin, N. (2013). Comprehensive review and compilation of treatment for azo dyes using microbial fuel cells. *Water Environment Research*, 85(3), 270-277.
- Myers, C. H. A. R. L. E. S. R., and Neelson, K. H. (1988). Bacterial manganese reduction and growth with manganese oxide as the sole electron acceptor. *Science*, 240(4857), 1319-1321.
- Nachiyar, C. V., and Rajakumar, G. S. (2005). Purification and characterization of an oxygen insensitive azoreductase from *Pseudomonas aeruginosa*. *Enzyme and Microbial Technology*, 36(4), 503-509.
- Najafpour, G., Rahimnejad, M., and Ghoreishi, A. (2011). The enhancement of a microbial fuel cell for electrical output using mediators and oxidizing agents. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 33(24), 2239-2248.
- Najafpour, G. (2015). *Biochemical engineering and biotechnology*. (1<sup>st</sup> ed.) Elsevier.
- Nasiri, R., Almaki, J. H., Esfarjani, S. S., Far, C. G., Rashid, A., and Aini, N. (2014). Identification of Biodegradation Related Genes from Bacterial Consortium NAR-2. *Applied Mechanics and Materials*, 735, 220-225.
- Niessen, J., Schröder, U., Rosenbaum, M., and Scholz, F. (2004). Fluorinated polyanilines as superior materials for electrocatalytic anodes in bacterial fuel cells. *Electrochemistry Communications*, 6(6), 571-575.

- Nigam, P., Banat, I. M., Singh, D., and Marchant, R. (1996). Microbial process for the decolorization of textile effluent containing azo, diazo and reactive dyes. *Process biochemistry*, 31(5), 435-442.
- Nizami, A. S., Ouda, O. K. M., Rehan, M., El-Maghraby, A. M. O., Gardy, J., Hassanpour, A., Kumar, S., and Ismail, I. M. I. (2015). The potential of Saudi Arabian natural zeolites in energy recovery technologies. *Energy*, 108, 162-171.
- Noori, P., and Najafpour Darzi, G. (2015). Enhanced power generation in annular single-chamber microbial fuel cell via optimization of electrode spacing using chocolate industry wastewater. *Biotechnology and applied biochemistry*, 63(3), 427-434.
- Oh, S., and Logan, B. E. (2005). Hydrogen and electricity production from a food processing wastewater using fermentation and microbial fuel cell technologies. *Water research*, 39(19), 4673-4682.
- Ong Boon Choon (2001). Pengoptimuman Penyahwarnaan Pewarna Azo Acid Yellow 9 oleh Bakteria. Tesis Ijazah Sarjana Muda. Universiti Teknologi Malaysia, Skudai.
- Omoike, A., and Chorover, J. (2004). Spectroscopic study of extracellular polymeric substances from *Bacillus subtilis*: aqueous chemistry and adsorption effects. *Biomacromolecules*, 5(4), 1219-1230.
- Ortega-Morales, B. O., Santiago-García, J. L., Chan-Bacab, M. J., Moppert, X., Miranda-Tello, E., Fardeau, M. L., Carrero, J. C., Bartolo-Pérez, P., Valadéz-González, A., and Guezennec, J. (2007). Characterization of extracellular polymers synthesized by tropical intertidal biofilm bacteria. *Journal of applied microbiology*, 102(1), 254-264.
- O'Toole, G. A., and Kolter, R. (1998). Flagellar and twitching motility are necessary for *Pseudomonas aeruginosa* biofilm development. *Molecular microbiology*, 30(2), 295-304.
- Pandey, A., Singh, P., and Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *International Biodeterioration and Biodegradation*, 59(2), 73-84.
- Park, D. H., and Zeikus, J. (1999). Utilization of electrically reduced neutral Red by *Actinobacillus succinogenes*: physiological function of neutral Red in membrane-driven fumarate reduction and energy conservation. *Journal of bacteriology*, 181(8), 2403-2410.

- Park, D. H., and Zeikus, J. G. (2000). Electricity generation in microbial fuel cells using neutral red as an electronophore. *Applied and environmental microbiology*, 66(4), 1292-1297.
- Park, D. H., and Zeikus, J. G. (2003). Improved fuel cell and electrode designs for producing electricity from microbial degradation. *Biotechnology and bioengineering*, 81(3), 348-355.
- Park, H. S., Kim, B. H., Kim, H. S., Kim, H. J., Kim, G. T., Kim, M., Chang, I. S., Park, Y. K., and Chang, H. I. (2001). A novel electrochemically active and Fe (III)-reducing bacterium phylogenetically related to *Clostridium butyricum* isolated from a microbial fuel cell. *Anaerobe*, 7(6), 297-306.
- Park, I. H., Heo, Y. H., Kim, P., and Nahm, K. S. (2013). Direct electron transfer in *E. coli* catalyzed MFC with a magnetite/MWCNT modified anode. *RSC Advances*, 3(37), 16665-16671.
- Pasqualino, J. C., Montane, D., and Salvado, J. (2006). Synergic effects of biodiesel in the biodegradability of fossil-derived fuels. *Biomass and bioenergy*, 30(10), 874-879.
- Pasti-Grigsby, M. B., Paszczynski, A., Goszczynski, S., Crawford, D. L., and Crawford, R. L. (1992). Influence of aromatic substitution patterns on azo dye degradability by *Streptomyces* spp. and *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*, 58(11), 3605-3613.
- Patil, S. A., Surakasi, V. P., Koul, S., Ijmulwar, S., Vivek, A., Shouche, Y. S., and Kapadnis, B. P. (2009). Electricity generation using chocolate industry wastewater and its treatment in activated sludge based microbial fuel cell and analysis of developed microbial community in the anode chamber. *Bioresource technology*, 100(21), 5132-5139.
- Pearce, C.I., Lloyd, J.R., Guthrie, J.T., (2003). The removal of color from textile wastewater using whole bacterial cells: A review. *Dyes Pigm.* 58, 179–196.
- Peixoto, L., Min, B., Martins, G., Brito, A. G., Kroff, P., Parpot, P., Angelidaki, I., and Nogueira, R. (2011). In situ microbial fuel cell-based biosensor for organic carbon. *Bioelectrochemistry*, 81(2), 99-103.
- Pielesz, A. (1999). The process of the reduction of azo dyes used in dyeing textiles on the basis of infrared spectroscopy analysis. *Journal of molecular structure*, 511, 337-344.

- Plumb, J. J., Bell, J., and Stuckey, D. C. (2001). Microbial populations associated with treatment of an industrial dye effluent in an anaerobic baffled reactor. *Applied and environmental microbiology*, 67(7), 3226-3235.
- Rabaey, K., Boon, N., Siciliano, S. D., Verhaege, M., and Verstraete, W. (2004). Biofuel cells select for microbial consortia that self-mediate electron transfer. *Applied and environmental microbiology*, 70(9), 5373-5382.
- Rabaey, K., Clauwaert, P., Aelterman, P., and Verstraete, W. (2005a). Tubular microbial fuel cells for efficient electricity generation. *Environmental science and technology*, 39(20), 8077-8082.
- Rabaey, K., Boon, N., Höfte, M., and Verstraete, W. (2005b). Microbial phenazine production enhances electron transfer in biofuel cells. *Environmental science and technology*, 39(9), 3401-3408.
- Rabaey, K., and Verstraete, W. (2005). Microbial fuel cells: novel biotechnology for energy generation. *Trends in Biotechnology*, 23(6), 291-298.
- Rabaey, K., Rodriguez, J., Blackall, L. L., Keller, J., Gross, P., Batstone, D., Verstraete, W., and Nealson, K. H. (2007). Microbial ecology meets electrochemistry: electricity-driven and driving communities. *The ISME journal*, 1(1), 9-18.
- Rahimnejad, M., Ghasemi, M., Najafpour, G. D., Ismail, M., Mohammad, A. W., Ghoreyshi, A. A., and Hassan, S. H. (2012). Synthesis, characterization and application studies of self-made Fe<sub>3</sub>O<sub>4</sub>/PES nanocomposite membranes in microbial fuel cell. *Electrochimica Acta*, 85, 700-706.
- Rai, H. S., Bhattacharyya, M. S., Singh, J., Bansal, T. K., Vats, P., and Banerjee, U. C. (2005). Removal of dyes from the effluent of textile and dyestuff manufacturing industry: a review of emerging techniques with reference to biological treatment. *Critical reviews in environmental science and technology*, 35(3), 219-238.
- Ramalho, P. A., Cardoso, M. H., Cavaco-Paulo, A., and Ramalho, M. T. (2004). Characterization of azo reduction activity in a novel ascomycete yeast strain. *Applied and environmental microbiology*, 70(4), 2279-2288.
- Reguera, G., McCarthy, K. D., Mehta, T., Nicoll, J. S., Tuominen, M. T., and Lovley, D. R. (2005). Extracellular electron transfer via microbial nanowires. *Nature*, 435(7045), 1098-1101.



- Reguera, G., Nevin, K. P., Nicoll, J. S., Covalla, S. F., Woodard, T. L., and Lovley, D. R. (2006). Biofilm and nanowire production leads to increased current in *Geobacter sulfurreducens* fuel cells. *Applied and environmental microbiology*, 72(11), 7345-7348.
- Reimers, C. E., Tender, L. M., Fertig, S., and Wang, W. (2001). Harvesting energy from the marine sediment-water interface. *Environmental science and technology*, 35(1), 192-195.
- Rhoads, A., Beyenal, H., and Lewandowski, Z. (2005). Microbial fuel cell using anaerobic respiration as an anodic reaction and biomineralized manganese as a cathodic reactant. *Environmental science and technology*, 39(12), 4666-4671.
- Rikame, S. S., Mungray, A. A., and Mungray, A. K. (2012). Electricity generation from acidogenic food waste leachate using dual chamber mediator less microbial fuel cell. *International Biodeterioration and Biodegradation*, 75, 131-137.
- Rinaldi, A., Mecheri, B., Garavaglia, V., Licoccia, S., Di Nardo, P., and Traversa, E. (2008). Engineering materials and biology to boost performance of microbial fuel cells: a critical review. *Energy and Environmental Science*, 1(4), 417-429.
- Ringeisen, B. R., Henderson, E., Wu, P. K., Pietron, J., Ray, R., Little, B., Biffinger, J.C., and Jones-Meehan, J. M. (2006). High power density from a miniature microbial fuel cell using *Shewanella oneidensis* DSP10. *Environmental science and technology*, 40(8), 2629-2634.
- Ringeisen, B. R., Ray, R., and Little, B. (2007). A miniature microbial fuel cell operating with an aerobic anode chamber. *Journal of Power Sources*, 165(2), 591-597.
- Robinson, T., McMullan, G., Marchant, R. and Nigam, P. (2001). Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. *Bioresource Technology*, 77, 247-255.
- Robson, H. (2001). *Verified synthesis of zeolitic materials*. Gulf Professional Publishing.
- Rodrigo, M. A., Canizares, P., Lobato, J., Paz, R., Sáez, C., and Linares, J. J. (2007). Production of electricity from the treatment of urban waste water using a microbial fuel cell. *Journal of Power Sources*, 169(1), 198-204.
- Rosen, M. J., and Kunjappu, J. T. (2012). *Surfactants and interfacial phenomena*. (4<sup>th</sup> ed.) Hoboken, New Jersey: John Wiley and Sons.

- Rozendal, R. A., Hamelers, H. V., and Buisman, C. J. (2006). Effects of membrane cation transport on pH and microbial fuel cell performance. *Environmental science and technology*, 40(17), 5206-5211.
- Rozendal, R. A., Hamelers, H. V., Rabaey, K., Keller, J., and Buisman, C. J. (2008). Towards practical implementation of bioelectrochemical wastewater treatment. *Trends in biotechnology*, 26(8), 450-459.
- Rožić, M., Šipušić, Đ. I., Sekovanić, L., Miljanić, S., Čurković, L., and Hrenović, J. (2009). Sorption phenomena of modification of clinoptilolite tuffs by surfactant cations. *Journal of Colloid and Interface Science*, 331(2), 295-301.
- Sanchez, D. V., Jacobs, D., Gregory, K., Huang, J., Hu, Y., Vidic, R., and Yun, M. (2015). Changes in carbon electrode morphology affect microbial fuel cell performance with *Shewanella oneidensis* MR-1. *Energies*, 8(3), 1817-1829.
- Sandhya, S. (2010). Biodegradation of azo dyes under anaerobic condition: role of azoreductase. *Biodegradation of Azo Dyes*, Springer, 39-57.
- Saratale, R. G., Saratale, G. D., Kalyani, D. C., Chang, J. S., and Govindwar, S. P. (2009a). Enhanced decolorization and biodegradation of textile azo dye Scarlet R by using developed microbial consortium-GR. *Bioresource technology*, 100(9), 2493-2500.
- Saratale, R. G., Saratale, G. D., Chang, J. S., and Govindwar, S. P. (2009b). Decolorization and biodegradation of textile dye Navy blue HER by *Trichosporon beigeli* NCIM-3326. *Journal of Hazardous Materials*, 166(2), 1421-1428.
- Saratale, R. G., Saratale, G. D., Chang, J. S., and Govindwar, S. P. (2011). Bacterial decolorization and degradation of azo dyes: a review. *Journal of the Taiwan. Institue of Chemical Engineers*, 42 (1),138–157.
- Sarayu, K., and Sandhya, S. (2010). Aerobic biodegradation pathway for Remazol Orange by *Pseudomonas aeruginosa*. *Applied biochemistry and biotechnology*, 160(4), 1241-1253.
- Sarkar, P., Fakhruddin, A., Pramanik, M.K. and Al-Mahin, A. (2011). Decolorization of Methyl Red by *Staphylococcus arlettae* PF4 Isolated from Garden Soil. *Int. J. Environ.* 1, 34-39.
- Savizi, I. S. P., Kariminia, H. R., and Bakhshian, S. (2012). Simultaneous decolorization and bioelectricity generation in a dual chamber microbial fuel

- cell using electropolymerized-enzymatic cathode. *Environmental science and technology*, 46(12), 6584-6593.
- Schröder, U., Nießen, J., and Scholz, F. (2003). A generation of microbial fuel cells with current outputs boosted by more than one order of magnitude. *Angewandte Chemie International Edition*, 42(25), 2880-2883.
- Schweizer, F., Jiao, H., Hindsgaul, O., Wong, W. Y., and Irvin, R. T. (1998). Interaction between the pili of *Pseudomonas aeruginosa* PAK and its carbohydrate receptor  $\beta$ -D-GalNAc (1->4)  $\beta$ -D-Gal analogs. *Canadian journal of microbiology*, 44(3), 307-311.
- Scott, G. V. (1968). Spectrophotometric determination of cationic surfactants with Orange II. *Analytical Chemistry*, 40(4), 768-773.
- Selcuk, H. (2005). Decolourization and detoxification of textile wastewater by ozonation and coagulation processes. *Dyes and Pigments*. 64, 217-222.
- Sell, D., Krämer, P., and Kreysa, G. (1989). Use of an oxygen gas diffusion cathode and a three-dimensional packed bed anode in a bioelectrochemical fuel cell. *Applied microbiology and biotechnology*, 31(2), 211-213.
- Shantaram, A., Beyenal, H., Veluchamy, R. R. A., and Lewandowski, Z. (2005). Wireless sensors powered by microbial fuel cells. *Environmental science and technology*, 39(13), 5037-5042.
- Sheng, G. P., Yu, H. Q., and Wang, C. M. (2006). FTIR-spectral analysis of two photosynthetic hydrogen-producing strains and their extracellular polymeric substances. *Applied Microbiology and Biotechnology*, 73(1), 241-241.
- Sheng, G. P., and Yu, H. Q. (2006). Characterization of extracellular polymeric substances of aerobic and anaerobic sludge using three-dimensional excitation and emission matrix fluorescence spectroscopy. *Water Research*, 40(6), 1233-1239.
- Sheng, G. P., Yu, H. Q., and Li, X. Y. (2010). Extracellular polymeric substances (EPS) of microbial aggregates in biological wastewater treatment systems: a review. *Biotechnology advances*, 28(6), 882-894.
- Shukla, A. K., Suresh, P., Sheela, B., and Rajendran, A. (2004). Biological fuel cells and their applications. *Current Science*, 87(4), 455-468.
- Singh, K., and Arora, S. (2011). Removal of synthetic textile dyes from wastewaters: a critical review on present treatment technologies. *Critical Reviews in Environmental Science and Technology*, 41(9), 807-878.

- Solanki, K., Subramanian, S., and Basu, S. (2013). Microbial fuel cells for azo dye treatment with electricity generation: a review. *Bioresource technology*, 131, 564-571.
- Song, T. S., Tan, W. M., Wu, X. Y., and Zhou, C. C. (2012). Effect of graphite felt and activated carbon fiber felt on performance of freshwater sediment microbial fuel cell. *Journal of Chemical Technology and Biotechnology*, 87(10), 1436-1440.
- Song, Y. C., Kim, D. S., Woo, J. H., Subha, B., Jang, S. H., and Sivakumar, S. (2015). Effect of surface modification of anode with surfactant on the performance of microbial fuel cell. *International Journal of Energy Research*, 39(6), 860-868.
- Sponza, D. T., and Işık, M. (2005). Reactor performances and fate of aromatic amines through decolorization of Direct Black 38 dye under anaerobic/aerobic sequential. *Process biochemistry*, 40(1), 35-44.
- Stelting, S., Burns, R. G., Sunna, A., Visnovsky, G., and Bunt, C. R. (2012). Immobilization of *Pseudomonas* sp. strain ADP: a stable inoculant for the bioremediation of atrazine. *Applied Clay Science*, 64, 90-93.
- Su, X., Tian, Y., Sun, Z., Lu, Y., and Li, Z. (2013). Performance of a combined system of microbial fuel cell and membrane bioreactor: wastewater treatment, sludge reduction, energy recovery and membrane fouling. *Biosensors and Bioelectronics*, 49, 92-98.
- Sullivan, E., Carey, J. and Bowman, R., (1998). Thermodynamics of cationic surfactant sorption onto natural clinoptilolite. *Journal of Colloid and Interface Science*, 206 (2), 369-380.
- Sun, J., Hu, Y., Bi, Z., and Cao, Y. (2009a). Improved performance of air-cathode single-chamber microbial fuel cell for wastewater treatment using microfiltration membranes and multiple sludge inoculation. *Journal of Power Sources*, 187(2), 471-479.
- Sun, J., Hu, Y. Y., Bi, Z., & Cao, Y. Q. (2009b). Simultaneous decolorization of azo dye and bioelectricity generation using a microfiltration membrane air-cathode single-chamber microbial fuel cell. *Bioresource Technology*, 100(13), 3185-3192.
- Sun, J., Bi, Z., Hou, B., Cao, Y. Q., and Hu, Y. Y. (2011). Further treatment of decolorization liquid of azo dye coupled with increased power production

- using microbial fuel cell equipped with an aerobic biocathode. *Water research*, 45(1), 283-291.
- Supaka, N., Juntongjin, K., Damronglerd, S., Delia, M. and Strehaiano, P. (2004). Microbial decolorization of reactive azo dyes in a sequential anaerobic–aerobic system. *Chemical Engineering Journal*, 99(2), 169-176.
- Szostak, R. (1989). *Molecular Sieves - Principles of Synthesis and Identification*. (1st ed.). Van Nostrand Reinhold: New York
- Takahashi, S., Okonogi, H., Hagiwara, T., Maekawa, Y. (2008). Preparation of polymer electrolyte membranes consisting of alkyl sulfonic acid for a fuel cell using radiation grafting and subsequent substitution/elimination reactions. *Journal of Membrane Science*, 324(1), 173-180.
- Tan, N. C. G., Prenafeta-Boldu, F. X., Opsteeg, J. L. (1999). Biodegradation of azo dyes in cocultures of anaerobic granular sludge with aerobic aromatic amine degrading enrichment cultures. *Applied Microbiology Biotechnology*, 51, 865-871.
- Tan, N. C. G. (2001). Integrated and Sequential Anaerobic/Aerobic Biodegradation of Azo Dyes. *Environmental Technologies to Treat Sulfur Pollution – Principles and Engineering*. 377-392.
- Tanisho, S., Kamiya, N., and Wakao, N. (1989). Microbial fuel cell using *Enterobacter aerogenes*. *Journal of electroanalytical chemistry and interfacial electrochemistry*, 275(1), 25-32.
- Tartakovsky, B., and Guiot, S. R. (2006). A comparison of air and hydrogen peroxide oxygenated microbial fuel cell reactors. *Biotechnology progress*, 22(1), 241-246.
- Telke, A., Kalyani, D., Jadhav, J., and Govindwar, S. (2008). Kinetics and mechanism of Reactive Red 141 degradation by a bacterial isolate *Rhizobium radiobacter* MTCC 8161. *Acta Chimica Slovenica*, 55(2), 320.
- Telke, A. A., Kalyani, D. C., Dawkar, V. V., and Govindwar, S. P. (2009). Influence of organic and inorganic compounds on oxidoreductive decolorization of sulfonated azo dye CI Reactive Orange 16. *Journal of hazardous materials*, 172(1), 298-309.
- Tender, L. M., Reimers, C. E., Stecher, H. A., Holmes, D. E., Bond, D. R., Lowy, D. A., Pilobello, K., Fertig, S. J., and Lovley, D. R. (2002). Harnessing

- microbially generated power on the seafloor. *Nature biotechnology*, 20(8), 821-825.
- Ter Heijne, A., Hamelers, H. V., De Wilde, V., Rozendal, R. A., and Buisman, C. J. (2006). A bipolar membrane combined with ferric iron reduction as an efficient cathode system in microbial fuel cells. *Environmental science and technology*, 40(17), 5200-5205.
- Thurston, C. F., Bennetto, H. P., Delaney, G. M., Mason, J. R., Roller, S. D., and Stirling, J. L. (1985). Glucose metabolism in a microbial fuel cell. Stoichiometry of product formation in a thionine-mediated *Proteus vulgaris* fuel cell and its relation to coulombic yields. *Microbiology*, 131(6), 1393-1401.
- Tomašević-Čanović, M., Daković, A., Rottinghaus, G., Matijašević, S., and Đuričić, M. (2003). Surfactant modified zeolites—new efficient adsorbents for mycotoxins. *Microporous and mesoporous materials*, 61(1), 173-180.
- Tony, B. D., Goyal, D., and Khanna, S. (2009). Decolorization of Direct Red 28 by mixed bacterial culture in an up-flow immobilized bioreactor. *Journal of industrial microbiology and biotechnology*, 36(7), 955-960.
- Tsai, H. Y., Wu, C. C., Lee, C. Y., and Shih, E. P. (2009). Microbial fuel cell performance of multiwall carbon nanotubes on carbon cloth as electrodes. *Journal of Power Sources*, 194(1), 199-205.
- Vaisman, L., Wagner, H. D., and Marom, G. (2006). The role of surfactants in dispersion of carbon nanotubes. *Advances in colloid and interface science*, 128, 37-46.
- Vandevivere, P.C., Bianchi, R. and Verstraete, W. (1998). Review: Treatment and reuse of wastewater from the textile wet-processing industry: Review of emerging technologies. *Journal of Chemical Technology and Biotechnology*, 72(4), 289-302.
- Van Der Zee, F.P. (2002). *Anaerobic azo dye reduction*. PhD Thesis, Wageningen University.
- Van der Zee, F. P., and Villaverde, S. (2005). Combined anaerobic–aerobic treatment of azo dyes—a short review of bioreactor studies. *Water research*, 39(8), 1425-1440.

- Vega, C. A., and Fernández, I. (1987). Mediating effect of ferric chelate compounds in microbial fuel cells with *Lactobacillus plantarum*, *Streptococcus lactis*, and *Erwinia dissolvens*. *Bioelectrochemistry and Bioenergetics*, 17(2), 217-222.
- Wahab, M. F. A., Rashid, N. A. A., and Mohd, A. R. H. (2006). Degradation of Acid Red 27 by the Recombinant Flavin Reductase. *KUSTEM 5<sup>th</sup> Annual Seminar* 2-3 May.1-5.
- Wahab, M. F.A., Chan, G. F., Yusoff, A. R. M., and Rashid, N. A. A. (2012). Reduction of azo dyes by flavin reductase from *Citrobacter freundii* A1. *Journal of Xenobiotics*, 3(1), 2.
- Wallace, T.H. (2001). *Biological treatment of a synthetic dye water and an industrial textile wastewater containing azo dye compounds*. Master Thesis, Virginia Polytechnic Institute.
- Wang, H., Zheng, X. W., Su, J. Q., Tian, Y., Xiong, X. J., and Zheng, T. L. (2009). Biological decolorization of the reactive dyes Reactive Black 5 by a novel isolated bacterial strain *Enterobacter* sp. EC3. *Journal of Hazardous Materials*, 171(1), 654-659.
- Wang, H. Y., Huang, H. F., and Jiang, J. Q. (2011a). The effect of metal cations on phenol adsorption by hexadecyl-trimethyl-ammonium bromide (hdtma) modified clinoptilolite (Ct.). *Separation and purification technology*, 80(3), 658-662.
- Wang, L., Lin, Y., Yang, L., Yu, P., Xie, Z. and Luo, Y. (2011b). *Candida tropicalis*: characterization of a strain capable of degrading high concentrations of phenol. *Biotechnology Letters*, 33(5), 943-946.
- Wang, L., and Langley, D. (1975). Determining cationic surfactant concentration. *Industrial and Engineering Chemistry Product Research and Development*, 14(3), 210-212.
- Wang, Q., Du, G., and Chen, J. (2004). Aerobic granular sludge cultivated under the selective pressure as a driving force. *Process Biochemistry*, 39(5), 557-563.
- Wang, S., Gong, W., Liu, X., Gao, B., and Yue, Q. (2006). Removal of fulvic acids using the surfactant modified zeolite in a fixed-bed reactor. *Separation and Purification Technology*, 51(3), 367-373.
- Wang, Y. P., Liu, X. W., Li, W. W., Li, F., Wang, Y. K., Sheng, G. P., Zeng, R.J. and Yu, H. Q. (2012). A microbial fuel cell–membrane bioreactor integrated system for cost-effective wastewater treatment. *Applied Energy*, 98, 230-235.

- Wang, Y., Li, B., Zeng, L., Cui, D., Xiang, X., and Li, W. (2013a). Polyaniline/mesoporous tungsten trioxide composite as anode electrocatalyst for high-performance microbial fuel cells. *Biosensors and Bioelectronics*, 41, 582-588.
- Wang, C. T., Huang, R. Y., Lee, Y. C., and Zhang, C. D. (2013b). Electrode material of carbon nanotube/polyaniline carbon paper applied in microbial fuel cells. *J Clean Energy Technol*, 1, 206-210.
- Wang, Z. W., Liang, J. S., and Liang, Y. (2013c). Decolorization of Reactive Black 5 by a newly isolated bacterium *Bacillus* sp. YZU1. *International Biodeterioration and Biodegradation*, 76, 41-48.
- Warchoń, J., Misaelides, P., Petrus, R. and Zamboulis, D. (2006). Preparation and application of organo-modified zeolitic material in the removal of chromates and iodides. *Journal of Hazardous Materials*, 137(3), 1410-1416.
- Watnick, P. I., Fullner, K. J., and Kolter, R. (1999). A role for the mannose-sensitive hemagglutinin in biofilm formation by *Vibrio cholerae* El Tor. *Journal of bacteriology*, 181(11), 3606-3609.
- Watson, V. J., and Logan, B. E. (2010). Power production in MFCs inoculated with *Shewanella oneidensis* MR-1 or mixed cultures. *Biotechnology and bioengineering*, 105(3), 489-498.
- Wei, J., Liang, P., and Huang, X. (2011). Recent progress in electrodes for microbial fuel cells. *Bioresource Technology*, 102(20), 9335-9344.
- Wilkinson, S. (2000). "Gastrobots"- Benefits and Challenges of Microbial Fuel Cells in Food Powered Robot Applications. *Autonomous Robots*, 9(2), 99-111.
- Wingender, J., Neu, T. R., and Flemming, H. C. (1999). What are bacterial extracellular polymeric substances?. In *Microbial extracellular polymeric substances* (pp. 1-19). Springer Berlin Heidelberg.
- Wingender, J., Neu, T. R., and Flemming, H. C. (Eds.). (2012). *Microbial extracellular polymeric substances: characterization, structure and function*. Springer Science and Business Media.
- Wu, X. Y., Tong, F., Song, T. S., Gao, X. Y., Xie, J. J., Zhou, C. C., Zhang, L.X. and Wei, P. (2015). Effect of zeolite-coated anode on the performance of microbial fuel cells. *Journal of Chemical Technology and Biotechnology*, 90(1), 87-92.



- Wuhrmann, K., Mechsner, K. and Kappeler, T. (1980). Investigation On Rate-Determining Factors In The Microbial Reduction Of Azo Dyes. *Eur. J. Applied Microbiology Biotechnology*, 9, 325-338.
- Xie, X., Ye, M., Hu, L., Liu, N., McDonough, J. R., Chen, W., Alshareef, H. N., Criddle, C. S., and Cui, Y. (2012). Carbon nanotube-coated macroporous sponge for microbial fuel cell electrodes. *Energy and Environmental Science*, 5(1), 5265-5270.
- Xu, H., Heinze, T. M., Paine, D. D., Cerniglia, C. E., and Chen, H. (2010). Sudan azo dyes and Para Red degradation by prevalent bacteria of the human gastrointestinal tract. *Anaerobe*, 16(2), 114-119.
- Xu, M., Guo, J., and Sun, G. (2007). Biodegradation of textile azo dye by *Shewanella decolorationis* S12 under microaerophilic conditions. *Applied microbiology and biotechnology*, 76(3), 719-726.
- Xu, S. and Boyd, S. A. (1995). Cationic surfactant sorption to a vermiculitic subsoil via hydrophobic bonding. *Environmental Science and Technology*, 29(2), 312-320.
- Yang, Y., Xiang, Y., Sun, G., Wu, W. M., & Xu, M. (2014). Electron acceptor-dependent respiratory and physiological stratifications in biofilms. *Environmental Science and Technology*, 49(1), 196-202.
- Yener, J., Kopac, T., Dogu, G., and Dogu, T. (2006). Adsorption of Basic Yellow 28 from aqueous solutions with clinoptilolite and amberlite. *Journal of Colloid and Interface Science*, 294(2), 255-264.
- Yong, X. Y., Feng, J., Chen, Y. L., Shi, D. Y., Xu, Y. S., Zhou, J., Wang, S.Y., Xu, L., Yong, Y. C., Sun, Y. M., Shi, C. L., Yang, P. K. O., and Zheng, T. (2014). Enhancement of bioelectricity generation by cofactor manipulation in microbial fuel cell. *Biosensors and Bioelectronics*, 56, 19-25.
- Yoo, E. S., Libra, J., and Adrian, L. (2001). Mechanism of decolorization of azo dyes in anaerobic mixed culture. *Journal of Environmental Engineering*, 127(9), 844-849.
- Yu, J., Wang, X., and Yue, P. L. (2001). Optimal decolorization and kinetic modeling of synthetic dyes by *Pseudomonas* strains. *Water research*, 35(15), 3579-3586.
- Yu, Z., and Wen, X. (2005). Screening and identification of yeasts for decolorizing synthetic dyes in industrial wastewater. *International Biodeterioration and Biodegradation*, 56(2), 109-114.

- Zhang, C. P., Liu, G. L., Zhang, R. D., and Quan, X. (2010a). Power generation from mixed substrates of quinoline and pyridine using microbial fuel cells. *Acta Scientiae Circumstantiae*, 30(7), 1372-1376.
- Zhang, X., Cheng, S., Wang, X., Huang, X., and Logan, B. E. (2009). Separator characteristics for increasing performance of microbial fuel cells. *Environmental Science and Technology*, 43(21), 8456-8461.
- Zhang, L., Zhu, X., Li, J., Liao, Q., and Ye, D. (2011). Biofilm formation and electricity generation of a microbial fuel cell started up under different external resistances. *Journal of Power Sources*, 196(15), 6029-6035.
- Zhang, M. M., Chen, W. M., Chen, B. Y., Chang, C. T., Hsueh, C. C., Ding, Y., Lin, K. L., and Xu, H. (2010b). Comparative study on characteristics of azo dye decolorization by indigenous decolorizers. *Bioresource technology*, 101(8), 2651-2656.
- Zhang, T., and Fang, H. H. (2001). Quantification of extracellular polymeric substances in biofilms by confocal laser scanning microscopy. *Biotechnology letters*, 23(5), 405-409.
- Zhang, T., Gannon, S. M., Nevin, K. P., Franks, A. E., and Lovley, D. R. (2010c). Stimulating the anaerobic degradation of aromatic hydrocarbons in contaminated sediments by providing an electrode as the electron acceptor. *Environmental microbiology*, 12(4), 1011-1020.
- Zhao, F., Harnisch, F., Schröder, U., Scholz, F., Bogdanoff, P., and Herrmann, I. (2005). Application of pyrolysed iron (II) phthalocyanine and CoTMPP based oxygen reduction catalysts as cathode materials in microbial fuel cells. *Electrochemistry Communications*, 7(12), 1405-1410.
- Zollinger, H. (2003). Color chemistry: syntheses, properties, and applications of organic dyes and pigments. 3<sup>rd</sup> edition Wiley-Vch.
- Zouari-Mechichi, H., Mechichi, T., Dhoui, A., Sayad, S., Martínez, A. T., and Martínez, M. J. (2006). Laccase purification and characterization from *Trametes troglis* isolated in Tunisia: decolorization of textile dyes by the purified enzyme. *Enzyme and Microbial Technology*, 39, 141-148
- Zuo, Y., Cheng, S., Call, D., and Logan, B. E. (2007). Tubular membrane cathodes for scalable power generation in microbial fuel cells. *Environmental science and technology*, 41(9), 3347-3353.