PERFORMANCE OF MICROWAVE ASSISTED AQUEOUS ENZYMATIC TECHNIQUE FOR *ELATERIOSPERMUM TAPOS* SEED OIL EXTRACTION

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To my beloved husband, family and friends

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ABSTRACT

Green, sustainable and effective extraction method is currently in demand for extraction of valuable compounds from natural resources. In this study, microwave assisted aqueous enzymatic extraction (MAAEE) which uses an electromagnetic wave as heating medium incorporated with non-toxic enzymatic aqueous solution as solvent was applied to extract omega-3 (ω -3) rich oil from *Elateriospermum tapos* seed. The effect of parameters in the MAAEE process including optimization, oil characterization, and mass transfer models relationship were investigated. In this study, experiments were carried out based on the design of experiment by central composite design using the design expert software. The optimization of extraction was analyzed by the response surface methodology and mass transfer models which were correlated with the models of the modified Fick's law, Patricelli's model and mass balance based on broken and intact cells. The results revealed that low microwave power (110 W) and small particle size (0.5 mm) gave significant effect on the extraction yield whereas increasing concentration of enzyme cocktail from 1 to 3% significantly increased the concentration of extracted ω -3 fatty acid. The optimum conditions were determined to be 110 W microwave power, 30 second extraction time, 1% enzyme cocktail concentration and 0.5 mm particle size, resulted in 46.12 \pm 1.48% recovery of extraction. Meanwhile, the optimum ω -3 fatty acid concentration was achieved at microwave power of 550 W, extraction time of 75 second, enzyme cocktail concentration of 3% and particle size of 0.5 mm resulted into 348.96 \pm 24.88 mg ω -3/g extracted oil compared to 106.57 \pm 4.32 mg ω -3/g of oil from Soxhlet extraction. The modeling study indicated mass balance model based on broken and intact cells as the best fitted model which gave the highest value of R^2 (0.9932) and lower value of absolute average relative deviation (3.7983) at 550 W of microwave power. Thus, it was proven that MAAEE was able to accelerate the extraction process and provided high quality of ω -3 rich oil extract at the same time.

ABSTRAK

Teknologi pengekstrakan hijau yang lestari dan berkesan kini menjadi pilihan untuk mengekstrak sebatian berharga daripada sumber alam semula jadi. Dalam kajian ini, pengekstrakan akueus enzimatik berbantukan gelombang mikro (MAAEE) menggunakan gelombang elektromagnet sebagai media pemanasan yang mana digabungkan dengan larutan enzim yang merupakan pelarut bukan toksik telah digunakan untuk mengekstrak minyak yang kaya dengan omega-3 (ω -3) daripada biji benih Elateriospermum tapos. Kesan parameter terhadap proses MAAEE; iaitu pengoptimuman, pencirian minyak, dan hubungan model pemindahan jisim telah Penyelidikan telah dijalankan berdasarkan reka bentuk eksperimen dikaji. menggunakan reka bentuk komposit pusat oleh perisian design expert. Pengoptimuman pengekstrakan dianalisis oleh kaedah gerak balas permukaan dan model pemindahan jisim yang telah dikolerasikan dengan model-model daripada hukum Fick terubah suai; model Patricelli dan imbangan jisim berdasarkan sel pecah dan sel tak terusik. Hasil kajian menunjukkan bahawa gelombang mikro yang berkuasa rendah (110 W) dan saiz zarah yang kecil (0.5 mm) memberikan kesan yang ketara ke atas hasil pengeluaran minyak sebaliknya peningkatan kepekatan enzim koktel dari 1 hingga 3% memberi kesan yang ketara kepada peningkatan jumlah kepekatan ω -3. Keadaan optimum telah ditentukan pada kuasa gelombang mikro 110 W, masa pengekstrakan 30 saat, kepekatan koktel enzim 1% dan saiz zarah 0.5 mm dengan memberi $46.12 \pm 1.48\%$ daripada jumlah hasil pengekstrakan. Kepekatan optimum ω -3 diperoleh pada keadaan kuasa gelombang mikro 550 W, masa pengekstrakan 75 saat, kepekatan enzim koktel 3% dan saiz zarah 0.5 mm dengan memberikan 348.96 \pm 24.88 mg ω -3/g minyak yang dikeluarkan berbanding dengan pengekstrakan soxhlet 106.57 \pm 4.32 mg ω -3/g minyak. Untuk kajian pemodelan, model imbangan jisim berdasarkan sel pecah and sel tak terusik merupakan model paling sepadan yang memberi nilai R^2 (0.9932) yang lebih tinggi dan nilai sisihan bandingan purata mutlak (3.7988) yang lebih rendah pada kuasa gelombang mikro 550 W. Oleh itu, MAAEE terbukti mampu mempercepatkan proses pengekstrakan dan pada masa yang sama dapat menghasilkan minyak yang kaya ω -3 yang berkualiti tinggi.

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LIST OF ABBREVIATIONS

AARD	-	Absolute Average Relative Deviation
AEE	-	Aqueous Enzymatic Extraction
AHA	-	American Health Association
ALA	-	Alpha Linolenic Acid
ANOVA	-	Analysis of The Variance
ATR	-	Attenuated Total Reflectance
BIC	-	Broken and Intact Cells
BFRs	-	Brominated Flame Retardants
CCD	-	Central Composite Design
CH ₃	-	Methyl
CHD	-	Coronary Heart Disease
COOH	-	Carboxyl Group
CV	-	Cardiovascular
CVD	-	Cardiovascular Disease
DHA	-	Docosapentaenoic Acid
DOE	-	Design of Experiment
DTGS	-	Deuterated Triglycine Sulphate
DW	-	Dry Weight
EPA	-	Eicosapentanoic Acid
ET	-	Elateriospermum Tapos
ETS	-	Elateriospermum Tapos Seed
FAO	-	Food And Agriculture Organization
FAMEs	-	Fatty Acids Methyl Ester
FDA	-	Food and Drug Administration
FTIR	-	Fourier Transform Infrared
GAE	-	Gallic Acid Equivalents

GC	-	Gas Chromatography			
GHz	-	Gigahertz			
GRAS	-	Generally Recognized As Save			
HCN	-	Hydrogen Cyanide			
HDL-	-	High Density Lipoprotein Cholesterol to Low Density			
C:LDL-C		Lipoprotein Cholesterol			
HIPS	-	High-Impact Polystyrene			
HPLC	-	High Performance Liquid Chromatogram			
ISM	-	Industrial, Scientific And Medical			
L	-	Linoleic			
La	-	Lauric			
LA	-	Linoleic Acid			
LDL	-	Low Density Lipoprotein			
LLL	-	Linoleic-Linoleic			
LLLn	-	Linoleic-Linolenic			
Ln	-	Linolenic			
LnOO	-	Linolenic-Oleic-Oleic			
Μ	-	Myristic			
MAE	-	Microwave Assisted Extraction			
MAAEE	-	Microwave Assisted Aqueous Enzymatic Extraction			
MUFA	-	Monounsaturated Fatty Acids			
NA	-	Not Applicable			
ND	-	Non-Detectable			
0	-	Oleic			
OLL	-	Oleic-Linoleic			
OLLn	-	Oleic-Linolenic			
OOL	-	Oleic-Oleic-Linoleic			
000	-	Oleic-Oleic			
Р	-	Palmatic			
PLL	-	Palmatic-Linoleic-Linoleic			
PLL-MOL	-	Palmatic-Linoleic-Linoleic-Myristic-Oleic-Linoleic			
Ро	-	Palmitoleic			
POL	-	Palmatic-Oleic-Linoleic			

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POO	-	Palmatic-Oleic-Oleic
PPL	-	Palmatic-Palmatic-Oleic
PoPoPo	-	Palmitoleic-Palmitoleic-Palmitoleic
PCB	-	Polychlorinated Biphenyl
PUFAs	-	Polyunsaturated Fatty Acids
PSE	-	Pressurized Solvent Extraction
RSM	-	Response Surface Methodology
S	-	Steric
SAFA	-	Saturated Fatty Acids
SE	-	Soxhlet Extraction
SEM	-	Scanning Electron Microscope
SFE	-	Supercritical Fluid Extraction
SSE	-	Sum of Squares Regression
SSR	-	Sum of Squares Error
SST	-	Sum of Squares Total
TAG	-	Triglycerol
TAGs	-	Triglycerides
TC	-	Serum Cholesterol
TPC	-	Total Phenolic Content
UAE	-	Ultrasonic-Assisted Extraction
USA	-	United States of America
UV	-	Ultraviolet
VOCs	-	Volatile Organic Compounds
WHO	-	World Health Organization

LIST OF SYMBOLS

3D	-	three dimensional
А	-	microwave power parameter in RSM
А	-	cellulase parameter in crossed mixture design
b	-	coefficients for extraction kinetics in washing step
В	-	extraction time parameter in RSM
В	-	pectinase parameter in crossed mixture design
С	-	concentration solute in the solid matrix
С	-	enzyme cocktail concentration
С	-	proteinase parameter in crossed mixture design
С	-	concentration of omega 3 fatty acid or amygdalin obtained from
		calibration curve
cm	-	centimeter
CV	-	coefficient of variation
d	-	Particle Size
df	-	degree of freedom
D	-	diffusion coefficient of the solute
D	-	particle size parameter in RSM
D	-	temperature parameter in crossed mixture design
Ε	-	field strength
E	-	incubation time parameter in crossed mixture design
f	-	frequency
g/d	-	gram per day
Н	-	extraction bed length
J	-	flux of solute
k	-	constant
k	-	coefficients for diffusion step

n-meternh-solvent flow rateng-milligramng/g-milligram per gramng/kg-milligram per kilogramng/kg-milligram per kilogramng/kg-milligram per kilogramng/kg-minoreng/mL-minoreng/mL-minutenL/min-milliliternL/min-milliliter per gramnd/g-milliliter per gramnfm(w)-milliliter per gramnfm(w)-milliliter per gramnfm-milliliter per gramnfm-millimeterM(w)-mass of the extractM(w)-mass flux of soluten-mometernfm-potential of HydrogenPHpHpHsecond-S/F-secondS/F-secondS/F-super to solid (feed)t-super to solid (feed)t/u-super ficial fluid velocityV/wwatt-wattW/w-wattW/w-wattW/w-watt	kV	-	kilo voltage
mg-miligrammg/g-miligram per grammg/kg-microgram per litegmg/L-microgram per millilitermg/L-microgram per millilitermin-minutemL-milliliter per minutemL/min-milliliter per grammL/g-milliliter per grammM(x)-milliliter per grammM(x)-mass of the extractM(mo)-mass of the extractM(mo)-mass flux of soluten-nanometerpH-potential of HydrogenP-radius of the particleR-solvent to solid (feed)r-solvent to solid (feed)t/m-solvent to solid (feed)t/m-solven	m	-	meter
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V-volumev/v-volume per volumeW-wattW/W-weight per weight	Т	-	temperature
v/v-volume per volumeW-wattW/W-weight per weight	и	-	superficial fluid velocity
W-wattW/W-weight per weight	V	-	volume
W/W - weight per weight	v/v	-	volume per volume
	W	-	watt
<i>x</i> - distance inside the porous part of the solid matrix	W/W	-	weight per weight
1	x	-	distance inside the porous part of the solid matrix

r		mass fraction in solid phase
X V	-	-
Y	-	mass fraction in fluid phase
У	-	output peak area of the absorbance
Z.	-	axial co-ordinate
8	-	bed void fraction
μL	-	microliter
μm	-	micrometer
%	-	percentage
>	-	greater than
<	-	lower than
°C	-	degree Celsius
tan δ	-	loss tangent
ω-3	-	omega-3
\mathbf{R}^2	-	correlation coefficient
Y^*	-	equilibrium fluid phase mass fraction
ε'	-	dielectric constant
ε"	-	dielectric loss
Co	-	initial concentration of solute in sample particle
C_{amyg}	-	concentration of the extracted amygdalin
Ci	-	concentration of solute at the interface of sample particle
C_t	-	concentration of solute extracted as function of time
C_1	-	amount of solute equilibrium yield at washing step
C_2	-	amount of solute equilibrium yield at diffusion step
\mathcal{C}_{∞}	-	concentration of solute in the extraction solvent after infinite
		time
$C_{\omega-3}$	-	concentration of the extracted omega-3
C_{exp}	-	experimental yield of omega-3 concentration
C_{model}	-	predicted yield of omega-3 concentration
k_1	-	mass transfer coefficient during washing step
k_2	-	mass transfer coefficient during diffusion step
k_{f}	-	fluid phase mass transfer coefficient
k _s	_	solid phase mass transfer coefficient
Ms	_	mean square
		· · · · · · · · · · · · · ·

P _{diss}	-	microwave power dissipation per unit volume
x_0	-	initial mass fraction of solute in solid phase
x_k	-	easily accessible solute in solid phase
W_{oil}	-	weight of the extracted oil
X_i and X_j	-	independent parameters level
Y_{l}	-	total oil yields
Y_2	-	concentration of omega-3
eta_{0}	-	constant
eta_i	-	Linear coefficient
β_{ii}	-	quadratic coefficient
eta_{ij}	-	interactive coefficient
$ ho_a$	-	apparent density
$ ho_f$	-	solvent density
$ ho_s$	-	solid density
$ ho_s$	-	real density

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CHAPTER 1

INTRODUCTION

1.1 Background

Nowadays, researches on therapeutical compounds from natural resources gain major interest from academic researchers. Omega-3 fatty acid is believed to be one of the therapeutical compounds that are currently attracting a great deal of attention. There are increasing amount of evidences citing omega-3 fatty acid ability to be used in the treatment and prevention of chronic diseases especially cardiovascular disease (CVD) known as the leading cause of death among people around the world. Omega-3 fatty acid is an essential fatty acid that need to be consumed through supplements or food products due to the incapability of human body to generate the compound naturally by itself (Poudyal et al., 2011; Kapoor and Patil, 2011). Omega-3 fatty acid is frequently found in marine animals and plants. However, due to several safety concerns, marine animals are no longer a remarkable source of omega-3 fatty acid as most of the researchers nowadays focus on expanding the discovery of omega-3 from plant as an alternative source. One underutilized local plant seed which is known as Perah or scientifically known as *Elateriospermum tapos* seed (ETS) had recently been reported to be rich of omega-3 fatty acid. A research by Yong and Salimon (2006) claimed that 17.4% of alpha linolenic acid (ALA), an omega-3 fatty acid is contained in the ETS. However, there is no further literature for the quantification of the specific amount of omega-3 fatty acid concentration in the ETS.

The primary method in extracting valuable active compounds from plants is the Soxhlet extraction (SE). This method has been comprehensively used as a standard reference to other methods of extraction due to its >99% extraction recovery (Pradhan et al., 2010). The microwave assisted aqueous enzymatic extraction (MAAEE) method is a promising new extraction method that is green, fast, efficient This method eliminates the disadvantages of conventional and energy saving. solvent extraction method which is the undesirable effect on oil quality due to the Water and aqueous based solvent system offer an organic solvent usage. increasingly crucial choice for the replacement of conventional organic solvent (Gai et al., 2013). MAAEE had been applied for oil extraction from seed crops such as pumpkin seed (Jiao et al., 2014), Isatis indigotica seed (Gai et al., 2013), yellow horn (Li et al., 2013), and Forsythia suspense seed (Gai et al., 2013). Microwave uses electromagnetic wave which penetrates into certain materials to provide volumetric heating through ionic conduction and dipole rotation (Chan et al., 2014). Treatment of ETS with enzymes enhance the extraction of oil yield due to its hydrolyzed structural polysaccharide of the cell walls and proteins associated with the lipid bodies (Jiao et al., 2014). Hence, a novel combination of MAAEE and enzymatic treatment will create effective synergy in enhancing the oil extraction process. To our knowledge, the combined use of enzymes and microwave assisted extraction of omega-3 fatty acid from ETS has not been previously reported.

The performance and efficiency of MAAEE depend upon many factors including microwave power, temperature, extraction time, solid to solvent ratio, and particle size. Some of these factors should be considered for the optimization of extraction condition. According to Baş *et al.* (2007), the optimization process can be carried out effectively using Response Surface Methodology (RSM) in which has become a powerful tool to determine the effect of the factors and their interaction. This method is the preferred experimental design technique for fitting polynomial model to analyze the response of multi-factor combination.

Mathematical modeling is useful in improving, optimizing, stimulating and scaling up a process design of the extraction process. It must be considered as a fundamental step during the operation of industrial process (Franco-vega *et al.*, 2016;

Xavier *et al.*, 2011). Mass transfer model of MAAEE is developed for the purpose of explaining the physical mechanism of extraction process based on mass transfers fundamentals.

Therefore, the aims of this research work are to study the interaction of different operating parameters on the extraction of ETS in order to attain the maximum yield of oil and omega-3 fatty acid concentration from ETS as well as the optimization purposes. Meanwhile, performances of the MAAEE are further characterized by physicochemical properties of ETS oil and compared with microwave assisted extraction (MAE) and SE methods. Furthermore, with the intention of evaluating the kinetic behavior (mass transfer coefficient) of data from the experiments; a mathematical model was also developed to study the relationship and correlate the best fit with three different mass transfer models.

1.2 Problem Statement

In Malaysia, Elateriospermum tapos seed (ETS) is a local seed that is in abundance but underutilized, which had been found to contain high valuable omega-3 fatty acid. Up to date, non-specific amounts of omega-3 fatty acid concentrations had been found in the seed according to previous studies. In addition, ETS also contains an antinutritional compounds known as amygdalin which is one of the cyanogenic glycoside compound that causes dizziness when consume in a large quantity. Thus, it is risky for consumers to consume omega-3 fatty acid directly from perah seed oil due to the presence of amygdalin in the seed. Hence, it is necessary to remove the amygdalin compound in order to obtain high quality omega-3 fatty acid. Amygdalin can be removed in sufficient amount using conventional methods such as fermentation, roasting and boiling. A study by Ngamriabsakul and Kommen (2009) reported that the amount of amygdalin was reduced from 660 mg/L to 100 mg/L and 25 mg/L after the seeds had been cooked and fermented, However, these methods are time consuming and the bioactive respectively. compounds present in ETS oil might be thermally oxidized. Thus, green and

effective methods are desired in order to extract and separate both omega-3 fatty acid and amygdalin simultaneously so that pure and high quality ETS oil can be attained.

There are various techniques of extraction that can be employed for the recovery of therapeutic compounds from plants. Recently, the MAAEE technique had been successfully applied in the extraction of oil from various seeds due to its special heating mechanism, fast extraction time and non-tolerable with organic solvent. However, this technique is considered as a new combination extraction technique which deals with enzymes and microwave assisted extraction. In MAAEE, it is crucial to study useful data for optimization such as ratio of the enzyme cocktail and operating parameters such as microwave power, extraction time, enzymes concentration, and particle size. These data need to be further studied as the extraction conditions available in the literature are only applicable for specific microwave system where different instrumental setup of microwave system would results to different performance if applied with the same operating conditions. In other words, the optimum extraction conditions reported in the literature are valid only as guides and references for new extraction microwave system. Therefore, optimization and response of interactions of the operating parameters need to be determined in order to maximize the yield of omega-3 fatty acid.

In addition, scarce availability of mass transfer data for ETS oil extracted by MAAEE reported also contributes to the problem of this study. The data is essential for further use as reference, for scaling up the production and for predicting the extraction behavior. Hence, mass transfer modeling of MAAEE needs to be in correlation with appropriated models.

1.3 Objectives of the Study

The main objective of this study is to investigate the potential of MAAEE as extraction method of seed oil by determining the most optimum parameters of ETS oil extraction and by modeling the process efficiency via MAAEE technique. Thus, the specific objectives of this research are:

- i) To identify and quantify the omega-3 fatty acid and amygdalin compounds in *Elateriospermum tapos* seed (ETS) oil.
- To investigate the effect of MAAEE operating parameters and to optimize the extraction condition on the ETS oil yield and omega-3 fatty acid concentration using Responses Surface Methodology (RSM).
- iii) To characterize and compare the morphology behavior, chemical structure, fatty acids profile and triglycerides (TAGs) composition among different methods namely MAAEE, microwave assisted extraction (MAE) and Soxhlet extraction (SE).
- iv) To investigate the relationship and best fit model representing mass transfer of extracted oil.

1.4 Scope of the Study

In order to achieve the objectives, the scopes of the study are stated as follows. Identification and determination of omega-3 fatty acid compound were carried out using gas chromatography (GC). This analysis was carried out in order to confirm the presence of omega-3 fatty acid and also to quantify the exact amount of omega-3 concentration in ETS oil. High performance liquid chromatography (HPLC) was used to detect and quantify the exact amount of amygdalin compounds, which is one of cyanogenic compound that is risky for direct consume by human.

In order to minimize the number of experiments and parameters involved for process optimization, some important parameters of MAAEE were set as constant parameters which are solvent to solid ratio, ratio of enzyme cocktail and solvent pH. Solvent to solid ratio needs to be determined in order to know the solvent required to extract the maximum amount of extraction oil. Ratio of the enzyme cocktail concentration is essential for determination in order to know the ratio of each enzyme used for the cell wall hydrolysis. Solvent pH is believed to cause denaturation or aggregation of protein which probably can be utilized for separating oil in extraction process.

In order to determine the optimum condition of MAAEE parameters on extraction yield of ETS oil and omega-3 fatty acid using Response Surface Methodology (RSM), the experiment was carried out at selected conditions at power, P (110-1100 W), extraction time, t (30-120 s), enzyme cocktail concentration, C (1-5%), and particle size, d (0.5-1.5 mm) according to the central composite design (CCD) of experiment using the Design Expert software. The microwave power range was chosen in accordance to low, medium and high level microwave power while the selection of extraction time is not exceeding 120 s or the extraction will no longer be significant. For enzyme cocktail concentration, the amount was selected to be not too high due to the expensive cost of enzymes while the selection of particle size is based on preliminary experiment.

The next scope is to continue the research with the investigation of the effects (P, t, c and d) and their interaction on extracted ETS oil yield and omega-3 fatty acid from Responses Surface Methodology (RSM) and analysis of variance (ANOVA). Low to medium microwave power is expected to favor the extraction efficiency as high power would rapidly increase the temperature. Meanwhile, the extraction yield would increase as extraction time increased until certain level. Increasing amount of enzyme cocktail concentration might speed up the extraction rate while smaller sample particle size would increase the extraction efficiency.

In order to investigate the performance of MAAEE, characterization of extracted ETS oil is compared with other methods (MAE and SE) including extraction yield, the amount of omega-3 concentration, the reduction amount of amygdalin, the properties of the oil including morphology image before and after extraction of ETS using SEM, chemical structure using FTIR, fatty acids analysis by GC and TAGs profile by HPLC.

The last scope of this research covers the investigation of the mass transfer models relationship and best fit model using modified Fick's Law model, Patricelli's model and mass balance model by broken and intact cells.

1.5 Significance of Study

This research looks into the abundant amounts of local seed that were underutilized as a new source of omega-3 fatty acid. Usually, rich omega-3 fatty acid products from plant sources such as flaxseed, linseed, canola, and walnut (Simopoulos, 2002) are imported from other countries which require high production and exportation costs. Thus, this new local source of omega-3 fatty acid from ETS provides huge benefit in term of production cost and less expensive omega-3 fatty acid oil could be produced. Moreover, only few people consume *Elateriospermum tapos* seed (ETS), thus it is not in competition with other food sources as compared to marine life. Hence, a sufficient supply of ETS in Malaysia could contribute to the low cost alternative of omega-3 fatty acid production.

Furthermore, from the aspect of academic contribution, the novel part of this research is the manipulation of microwave assisted extraction condition to remove the undesired amygdalin from ETS in order to produce green, safe and high quality oil. The capability of the MAAEE technique to extract desired concentration of target compounds with fast and low cost extraction setup could be established. Besides, the optimization, influences of operating parameters and mass transfer modeling data are significant to industry as a guideline and references in order to upgrade to industrial scale. A side from that, the quantification method of omega-3 fatty acid concentration using GC conducted in this study could provide consumers with the information of sufficient intake of omega-3 fatty acid. Thus, the extracted ETS oil has huge potential to be further developed either in the food, pharmaceutical or oleo chemical industries.

1.6 Thesis Outline

This thesis consists of five chapters and the content of each chapter are describe as follows:

Chapter 1 introduces the background and objectives of this research. It also discusses problem statement, scope of the study, novelty contribution and thesis outline.

Chapter 2 reviews the properties of omega-3 fatty acid, amygdalin compounds and botanical information of ETS. This chapter also discusses the fundamental and effect of parameters of the microwave assisted extraction (MAE) technique as well association of hydrolytic enzymes in MAAEE which was employed in most plant extracts. The optimization method using response surface methodology (RSM) is also described in this chapter. This chapter also covers the reviews on several relevant mass transfers modeling.

Chapter 3 presents the research methodology for optimization and modeling of MAAEE. It also describes the analysis methods for ETS oil characterization.

Chapter 4 encompasses the results and discussion on the optimization of the ETS oil yield and omega-3 fatty acid concentrations as well as the influences of the extraction parameters. The comparison of MAAEE with the conventional SE as well as MAE is also presented in this chapter which also includes the quantitative amount of omega-3 fatty acid and amygdalin and also the characterizations of the oil. Moreover, the relationship and best fitting of mass transfer models is also evaluated in this chapter.

Chapter 5 concludes all the findings of this research and proposes recommendations for future work.

REFERENCES

- Abraham, K., Buhrke, T., and Lampen, A. (2016). Bioavailability of Cyanide after Consumption of a Single Meal of Foods Containing High Levels of Cyanogenic Glycosides: A Crossover Study in Humans. *Archives of Toxicology*. 90(3), 559– 574.
- Achi, O. K. (2005), Traditional Fermented Protein Condiments in Nigeria. African Jounal of Biotechnology. 4(13), 1612-1621
- Adarme-vega, T. C., Lim, D. K. Y., Timmins, M., Vernen, F., Li, Y., and Schenk, P.
 M. (2012). Microalgal Biofactories: A Promising Approach Towards Sustainable Omega-3 Fatty Acid Production. *Microbial Cell Factories*. 11(1), 96.
- Amarni, F., and Kadi, H. (2010). Kinetics Study of Microwave-Assisted Solvent Extraction of Oil from Olive Cake using Hexane Comparison with the Conventional Extraction. *Innovative Food Science and Emerging Technologies*. 11(2), 322–327.
- Antony, M., Amutha, S., Arasi, G., and Rao, M. G. (2016). Optimization of Microwave-Assisted Extraction of Polysaccharide from *Psidium Guajava* L . Fruits. *International Journal of Biological Macromolecules*. 91, 227–232.
- Armstrong, S. D. (1999). Microwave-Assisted Extraction for the Isolation of Trace Systemic Fungicides from Woody Plant Material. PhD Thesis. Virginia Polytechnic Institute and State University. Blacksburg.
- Arvindekar, A. U., and Laddha, K. S. (2016). An Efficient Microwave-Assisted Extraction of Anthraquinones from Rheum Emodi: Optimisation using RSM, UV and HPLC Analysis and Antioxidant Studies. *Industrial Crops and Products*. 83, 587–595.
- Azmir. J., Zaidul, I. S. M., Rahman., M. M., Sharif, K. M., Mohamed, A., Sahena, F.,Jahurul, M. H. A., Ghafoor, K., Norulaini, N. A. N., Omar, A. K. M. (2013).Techniques for Extraction of Bioactive Compounds from Plant Materials: A

Review. Journal of Food Engineering. 117, 426-436.

- Ballard, T. S. (2008). Optimizing the Extraction of Phenolic Antioxidant Compounds from Peanut Skins. Ph.D Thesis. Virginia Polytechnic Institute and State University. Blacksburg
- Bart, H. J. (2011). Extraction of Natural Product from Plant-An Introduction. Bart,
 H. J.and Pilz, S. (eds). Industrial Scale Natural Products Extraction (pp.1-26).Germany: Wiley-VCH Verlag GmbH & Co. KGaA.
- Baş, D., and Boyacı, İ. H. (2007). Modeling and optimization I: Usability of Response Surface Methodology. *Journal of Food Engineering*. 78(3), 836–845.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., and Escaleira, L. A. (2008). Response Surface Methodology (RSM) as a Tool for Optimization in Analytical Chemistry. *Talanta*. 76(5), 965–977.
- Bolarinwa, I. F., Orfila, C., and Morgan, M. R. a. (2014). Amygdalin Content of Seeds, Kernels and Food Products Commercially-available in the UK. *Food Chemistry*. 152, 133–9.
- Chan, C. (2013). Optimization and Modeling of Microwave-Assisted Extraction of Active Compounds From Cocoa. Ph.D Thesis. University of Malaya. Kuala Lumpur
- Chan, C. H., Yusoff, R., Ngoh, G. C., and Kung, F. W. L. (2011). Microwave-Assisted Extractions of Active Ingredients from Plants. *Journal of Chromatography A*. 1218(37), 6213-6225
- Chan, C., Yusoff, R., and Ngoh, G. (2013). Modeling and Kinetics Study of Conventional and Assisted Batch Solvent Extraction. *Chemical Engineering Research and Design*. 92(6), 1169–1186
- Chen, L., Kang, Y., and Suh, J. (2014). Roasting Processed Oriental Melon (*Cucumis melo* L . var . makuwa Makino) Seed influenced the Triglyceride Pro file and the Inhibitory Potential against Key Enzymes Relevant for Hyperglycemia. Food Research International. 56, 236–242.
- Choonhahirun, A. (2010). Proximate composition and functional properties of pra (Elateriospermun tapos Blume) Seed flour. *African Journal of Biotechnology* 9(36), 5946–5949.
- Chow, Y. L. (1969). Short Communication Biogenetically Related Triterpenes Elateriospermum Tapos Bark. Phytochemistry. 9, 1151-1152
- Chumkiew, S., Srisang, W., Jaroensutasinee, K., Jaroensutasinee, M., and Site, A. S.

(2007). Phenology of the Parah Tree (*Elateriospermum Tapos*) using a GAPS Model. *Word Academy of Science, Engineering and Technology*. 32, 207–211.

- Connor, W. E. (1999). Linolenic Acid in Health and Disease. *The American Journal* of Clinical Nutrition. 69(5), 827–828.
- Cornell, J. A. (1990). *How to Apply Response Surface Methodology*. (8th ed.). American Society for Quality Control Statistics Division.
- Cranbrook, E. and Edwards. D.S. (1994). A Tropical Rainforest, the Nature of Biodiversity in Borneo at Belalong, Brunei. *The Royal Geographical Society* and Sun Tree Publishing. 389
- Crank, J. (1975). The Mathematics of Diffusion. (2nd ed.). Bristol, England: Oxford University Press.
- Cravotto, G., Boffa, L., Mantegna, S., Perego, P., Avogadro, M., and Cintas, P. (2008). Improved Extraction of Vegetable Oils under High-intensity Ultrasound and/or Microwaves. *Ultrasonics Sonochemistry*. 15(5), 898–902.
- Da Porto, C., Decorti, D., & Tubaro, F. (2012). Fatty Acid Composition and Oxidation Stability of Hemp (*Cannabis sativa* L.) Seed Oil Extracted by Supercritical Carbon Dioxide. *Industrial Crops and Products*. 36(1), 401–404.
- Dahmoune, F., Nayak, B., Moussi, K., Remini, H., and Madani, K. (2015). Optimization of Microwave-Assisted Extraction of Polyphenols from *Myrtus Communis* L. leaves. *Food Chemistry*. 166, 585–95.
- Dai, J. (2006) Microwave-Assisted Extraction and Synthesis Studies and the Scale-up Study with the Aid of FDTD Simulation. Ph.D Thesis. McGill University. Canada
- Daley, C. a, Abbott, A., Doyle, P. S., Nader, G. a, and Larson, S. (2010). A Review of Fatty Acid Profiles and Antioxidant Content in Grass-fed and Grain-fed Beef. *Nutrition Journal*. 9:10.
- de la Mata, P., Dominguez-Vidal, A., Bosque-Sendra, J. M., Ruiz-Medina, A., Cuadros-Rodríguez, L., and Ayora-Cañada, M. J. (2012). Olive Oil Assessment in Edible Oil Blends by means of ATR-FTIR and Chemometrics. *Food Control*. 23(2), 449–455.
- Dong, Z., Gu, F., Xu, F., and Wang, Q. (2014). Comparison of Four Kinds of Extraction Techniques and Kinetics of Microwave-Assisted Extraction of Vanillin from Vanilla Planifolia. Food Chemistry. 149, 54–61.

Dutta, R., Sarkar, U., and Mukherjee, A. (2014). Extraction of Oil from Crotalaria

Juncea seeds in a Modified Soxhlet Apparatus: Physical and Chemical characterization of a Prospective Bio-fuel. *Fuel.* 116, 794–802.

- Durmaz, E. (2012). *Microwave Extraction of Phenolic Compounds from Caper and Oleaster*. Master Thesis. Middle East Technical University.
- Eknayake, S., Jansz, E. R., and Nair, B. M. (1999). Proximate Composition, Mineral and Amino Acid Content of *Canavalia Gladiata* Seeds. *Journal of Food Chemistry*. 66, 115–119.
- Follegatti-Romero, L. a., Piantino, C. R., Grimaldi, R., and Cabral, F. a. (2009). Supercritical CO₂ Extraction of Omega-3 Rich oil from Sacha Inchi (*Plukenetia volubilis* L.) seeds. *The Journal of Supercritical Fluids*. 49(3), 323–329.
- Franco-vega, A., Ramírez-corona, N., Palou, E., and Aurelio, L. (2016). Estimation of Mass Transfer Coefficients of the Extraction Process of Essential Oil from Orange Peel using Microwave Assisted Extraction. *Journal of Food Engineering*. 170, 136–143.
- Fu, Y. J., Liu, W., Zu, Y. G., Tong, M. H., Li, S. M., Yan, M. M., Efferth, T., and Luo, H. (2008). Enzyme Assisted Extraction of Luteolin And Apigenin from Pigeonpea [*Cajanus Cajan* (L.) *Millsp.*] Leaves. *Food Chemistry*. 111(2), 508– 512.
- Gable, K. P.(2014). Infrared Spectroscopy: Indentifying Functional Groups.
 Retrieved January 20, 2017, from https://chemistry.oregonstate.edu/courses/ch361-464/ch362/irinterp.htm
- Gai, Q.-Y., Jiao, J., Mu, P.-S., Wang, W., Luo, M., Li, C.-Y., Zu, Y.-G., Fu, Y.-W., and Fu, Y.-J. (2013). Microwave-Assisted Aqueous Enzymatic Extraction of Oil from *Isatis Indigotica* Seeds and its Evaluation of Physicochemical Properties, Fatty Acid Compositions and Antioxidant Activities. *Industrial Crops and Products*. 45, 303–311.
- Gai, Q.-Y., Jiao, J., Wei, F.-Y., Luo, M., Wang, W., Zu, Y.-G., and Fu, Y.-J. (2013). Enzyme-Assisted Aqueous Extraction of Oil from *Forsythia Suspense* Seed and its Physicochemical Property and Antioxidant Activity. *Industrial Crops and Products*. 51, 274–278.
- Ganzler, K.; Salgo, A.; and Valko, K. (1986). Microwave Extraction: A Novel Sample Preparation Method for Chromatography. *Journal of Chromatography*. 371, 299-306.
- Gao, F., Yang, S., and Birch, J. (2016). Physicochemical Characteristics , Fatty Acid

Positional Distribution and Triglyceride Composition in Oil Extracted from Carrot Seeds using Supercritical CO₂. *Journal of Food Composition and Analysis*. 45, 26–33.

- Geller, R. J., Barthold, C., Saiers, J. A., and Hall, A. H. (2006). Pediatric Cyanide Poisoning: Causes, Manifestations, Management, and Unmet needs. *Review Pediatrics*. 118, 2146–2158.
- Gleadow, R. M., and Woodrow, I. E. (2002). Constraints on Effectiveness of Cyanogenic Glycosides in Herbivore Defense. *Journal of Chemical Ecology*. 28, 1301-13.
- Gopalakannan, S., and Senthilvelan, T. (2014). Optimization of Machining Parameters for Edm Operations Based on Central Composite Design and Desirability Approach. *Journal of Mechanical Science and Technology*. 28(3), 1045-1053.
- Grosso, C., Coelho, J. P., Pessoa, F. L. P., Fareleira, J. M. N. A., Barroso, J. G., Urieta, J. S., Palavra, A. F., Sovová, H. (2010). Mathematical Modelling of Supercritical CO₂ Extraction of Volatile Oils from Aromatic Plants. *Chemical Engineering Science*. 65(11), 3579–3590.
- Hamidah, S., Yian, L. N., and Mohd, A. (2011). Comparison of Physico-Chemical Properties and Fatty Acid Compostion of *Elateriospermum Tapos* (Buah Perah), Palm Oil and Soybean Oil. *Word Academy of Science, Engineering and Technology*. 81, 855–858.
- Handa, S. S. (2008). An Overview of Extraction Techniques for Medicinal and Aromatic Plants. Handa, S. S., Khanuja, S. P. S., Longo, G., Rakesh, D.D. (eds). za (pp.21-52). Trieste: ICS-UNIDO
- Harris, W. S., Kris-Etherton, P. M., and Harris, K. a. (2008). Intakes of Long-Chain Omega-3 Fatty Acid Associated with Reduced Risk for Death from Coronary Heart Disease in Healthy Adults. *Current Atherosclerosis Reports*. 10(6), 503– 9.
- Hayta, M., and Alpaslan, M. (2011). Apricot Kernel Flour and its use in Maintaining Health. Preedy, V. R., Waston, R. R., and Patel V. B. (Eds). Flour and Breads and their Fortification in Health and Disease Prevention (pp. 213-221). London: Academic Press
- Hazebroek, H.P., Kassim, A., (2001). *National Parks of Sarawak*. Natural History Publication, Borneo.

- Hu, Z., Cai, M., and Liang, H. H. (2008). Desirability Function Approach for the Optimization of Microwave-Assisted Extraction of Saikosaponins From *Radix Bupleuri*. Separation and Purification Technology. 61(3), 266–275.
- Husin, N., Tan, N. A. H., Idayu, I., and Mohd, N. (2013). Physicochemical and Biochemical Characteristics of the Underutilized. *Jurnal Teknologi*. 64(2), 57– 61.
- Ixtaina, V. Y., Vega, A., Nolasco, S. M., Tomás, M. C., Gimeno, M., Bárzana, E., and Tecante, A. (2010). Supercritical Carbon Dioxide Extraction of Oil from Mexican Chia Seed (*Salvia hispanica* L.): Characterization and Process Optimization. *Journal of Supercritical Fluids*. 55(1), 192–199.
- Jiao, J., Fu, Y.-J., Zu, Y.-G., Luo, M., Wang, W., Zhang, L., and Li, J. (2012). Enzyme-assisted Microwave Hydro-distillation Essential Oil from Fructus Forsythia, Chemical Constituents, and its Antimicrobial and Antioxidant Activities. *Food Chemistry*. 134(1), 235–243.
- Jiao, J., Li, Z., Gai, Q., Li, X., Wei, F., Fu, Y., and Ma, W. (2014). Microwaveassisted Aqueous Enzymatic Extraction of Oil From Pumpkin Seeds and Evaluation of its Physicochemical Properties, Fatty Acid Compositions and Antioxidant Activities. *Food Chemistry*. 147, 17–24.
- Jin, G., Yang, F., Hu, C., Shen, H., & Zhao, Z. K. (2012). Enzyme-assisted Extraction of Lipids Directly from the Culture of the Oleaginous Yeast Rhodosporidium Toruloides. *Bioresource Technology*. 111, 378–82.
- Kanitkar, A. V. (2010). Parameterization of Microwave Assisted Oil Extraction and tts Transesterification to Biodiesel. Master Thesis. Louisiana State University
- Kapoor, R. and Patil, U. K. (2011). Importance and Production of Omega-3 Fatty Acids from Natural Sources. *International Food Research Journal*. 18, 493– 499.
- Kawahito, Y., Kondo, M., Machmudah, S., Sibano, K., Sasaki, M., and Goto, M.(2008). Supercritical CO₂ Extraction of Biological Active Compounds from Loquat Seed. *Separation and Purification Technology*. 61: 130–135
- Khalid, E. K., Babiker, E. E., & EL Tinay, A. H. (2003). Solubility and Functional Properties of Sesame Seed Proteins as Influenced by pH and/or Salt Concentration. *Food Chemistry*. 82(3), 361–366.
- Kim, J.-M., Chang, S.-M., Kim, I.-H., Kim, Y.-E., Hwang, J.-H., Kim, K.-S., and Kim, W.-S. (2007). Design of Optimal Solvent for Extraction of Bio-active

Ingredients from Mulberry Leaves. *Biochemical Engineering Journal*. 37(3), 271–278.

- Kris-etherton, P. M., Harris, W. S., and Appel, L. J. (2002). Fish Consumption, Fish Oil, Omega-3 Fatty Acids, and Cardiovascular Disease Epidemiological and Observational Studies. *American Heart Association*. 106, 2747.
- Latif, S., and Anwar, F. (2011). Aqueous Enzymatic Sesame Oil and Protein Extraction. *Food Chemistry*. 125(2), 679–684.
- Lee, J. H., O'Keefe, J. H., Lavie, C. J., Marchioli, R., and Harris, W. S. (2008). Omega-3 Fatty Acids for Cardioprotection. *Mayo Clinic Proceedings*, 83(3), 324–32.
- Leonelli, C., Veronesi, P., and Cravotto, G. (2013), *Microwave-Assisted Extraction:* An Introduction to Dielectric Heating. Chemat, F., and Cravotto, G. (eds). Microwave-assisted Extraction for Bioactive Compounds. (pp.1-14). London: Springer Science.
- Li, J., Zu, Y.-G., Fu, Y.-J., Yang, Y.-C., Li, S.-M., Li, Z.-N., and Wink, M. (2010). Optimization of Microwave-Assisted Extraction of Triterpene Saponins from Defatted Residue of Yellow Horn (*Xanthoceras Sorbifolia Bunge.*) Kernel and Evaluation of its Antioxidant Activity. *Innovative Food Science and Emerging Technologies.* 11(4), 637–643.
- Li, J., Zu, Y.-G., Luo, M., Gu, C.-B., Zhao, C.-J., Efferth, T., and Fu, Y.-J. (2013). Aqueous Enzymatic Process Assisted by Microwave Extraction of Oil from Yellow Horn (*Xanthoceras Sorbifolia Bunge.*) Seed Kernels and its Quality Evaluation. *Food Chemistry.* 138(4), 2152–8.
- Li, X., Li, Z., Wang, X., Han, J., Zhang, B., and Fu, Y. (2016). Application of Cavitation System to Accelerate Aqueous Enzymatic Extraction of Seed Oil from *Cucurbita Pepo* L. and Evaluation of Hypoglycemic Effect. *Food Chemistry*. 212, 403–410.
- Li, Y., Jiang, L., Sui, X., and Wang, S. (2011). Optimization of the Aqueous Enzymatic Extraction of Pine Kernel Oil by Response Surface Methodology. *Procedia Engineering*. 15, 4641–4652.
- Lim, T. K. (2012). Edible Medicinal And Non-Medicinal Plants. Fruit. 2, 472–475.
- Ling, S. K., Fukumori, S., Tomii, K., Tanaka, T., and Kouno, I. (2006). Isolation, Purification and Identification of Chemical Constituents from *Elateriospermum Tapos*. Journal of Tropical Forest Science. 18(1), 81–85.

- Liu, W., Fu, Y.-J., Zu, Y.-G., Tong, M.-H., Wu, N., Liu, X.-L., and Zhang, S. (2009). Supercritical Carbon Dioxide Extraction of Seed Oil from *Opuntia Dillenii Haw.* and its Antioxidant Activity. *Food Chemistry*. 114(1), 334–339.
- Louli, V., Folas, G., Voutsas, E., and Magoulas, K. (2004). Extraction of Parsley Seed Oil by Supercritical CO₂. *The Journal of Supercritical Fluids*. 30(2), 163– 174. doi:10.1016/j.supflu.2003.07.003
- Luque de Castro, M., and García-Ayuso, L. (1998). Soxhlet Extraction of Solid Materials: an Outdated Technique with a Promising Innovative Future. *Analytica Chimica Acta*. 369(1-2), 1–10.
- Luque de Castro, M. D., and Priego-Capote, F. (2010). Soxhlet Extraction: Past and Present Panacea. *Journal of Chromatography A*. 1217(16), 2383–2389.
- Lv, W., Ding, M., and Zheng, R. (2005). Isolation and Quantitation of Amygdalin in Apricot-kernel and Prunus Tomentosa Thunb by HPLC with Solid-Phase Extraction. *Journal of Chromatographic Science*. 43, 383–387.
- Mandal, V., and Mandal, S. C. (2010). Design and Performance Evaluation of a Microwave Based Low Carbon Yielding Extraction Technique for Naturally Occurring Bioactive Triterpenoid: Oleanolic Acid. *Biochemical Engineering Journal*. 50(1-2), 63–70.
- Masterton, G. S., Plevris, J. N., and Hayes, P. C. (2010). Omega-3 Fatty acids a Promising Novel Therapy for Non-alcoholic Fatty Liver Disease. *Alimentary Pharmacology and Therapeutics*. 31(7), 679–92.
- Mat, M., Gordon, M. H., Ezeh, O., and Niranjan, K. (2016). Aqueous Enzymatic Extraction of Moringa Oleifera Oil. *Food Chemistry*. 211, 400–408.
- Maurer, N. E., Hatta-Sakoda, B., Pascual-Chagman, G., and Rodriguez-Saona, L. E. (2012). Characterization and Authentication of a Novel Vegetable Source of Omega-3 Fatty Acids, Sacha Inchi (*Plukenetia volubilis* L.) Oil. *Food Chemistry*. 134(2), 1173–80.
- Mazlan, H. satirah, Muhamad, I. I., Hassan, N. D., and Tan, N. A. H. (2014). Optimization of Protein Extraction from Fermented and Non Fermented Perah Seed by using Response Surface Methodology. *Jurnal Teknologi*. 68(5), 29–33.
- Mckay, B. D. L., and Sibley, D. (2011). Omega-3 Fatty Acids from Walnuts. Retrieved December 30, 2012, from www.NutritionDimension.com
- Ngamriabsakul, C., and Kommen, H. (2009). The Preliminary Detection of Cyanogenic Glycosides in Pra (*Elateriospermum tapos* Blume) by HPLC.

- Osada, N., Takeda, H., Kawaguchi, H., Furukawa, A., and Awang, M. (2003). Estimation of Crown Characters and Leaf Biomass from Leaf Litter in a Malaysian Canopy Species, *Elateriospermum Tapos* (Euphorbiaceae). *Forest Ecology and Management*. 177(1-3), 379–386.
- Passos, C. P., Coimbra, M. A., Da, F. A., and Silva, C. M. (2010). Modelling the Supercritical Fluid Extraction of Edible Oils and Analysis of the Effect of Enzymatic Pre-treatments of Seed upon Model Parameters. *Chemical Engineering Research and Design.* 89, 1118–1125.
- Patricelli, A., Assogna, A., Casalaina, A., Emmi, E., and Sodini, G. (1979). Fattori che influenzano l'estrazione dei lipidi da semi decorticati di girasole. *Rivista Italiana Delle Sostanze Grasse*. 56, 151–154
- Passos, C. P., Yilmaz, S., Silva, C. M., and Coimbra, M. A. (2009). Enhancement of Grape Seed Oil Extraction using a Cell Wall Degrading Enzyme Cocktail. *Food Chemistry*. 115(1), 48–53.
- Pattamadilok, D., and Suttisri, R. (2008). Seco-terpenoids and other Constituents from *Elateriospermum tapos*. *Journal of Natural Products*. 71(2), 292–4.
- Phongthai, S., Lim, S., and Rawdkuen, S. (2016). Optimization of Microwave-Assisted Extraction of Rice Bran Protein and its Hydrolysates Properties. *Journal of Cereal Science*. 70, 146–154.
- Popoola T. O. S. and Akueshi C. O. (1986). Nutritional Evaluation of Daddawa, A Local Spice Made from Soybean (Glycine Max.). *MIRCEN Journal*. 2, 405-409
- Poudyal, H., Panchal, S. K., Diwan, V., and Brown, L. (2011). Omega-3 Fatty acids and Metabolic Syndrome: Effects and Emerging Mechanisms of Action. *Progress in Lipid Research*. 50(4), 372–87.
- Pradhan, R. C., Meda, V., Rout, P. K., Naik, S., and Dalai, A. K. (2010). Supercritical CO₂ Extraction of Fatty Oil from Flaxseed and Comparison with Screw Press Expression and Solvent Extraction Processes. *Journal of Food Engineering*. 98(4), 393–397.
- Qu, X.-J., Fu, Y.-J., Luo, M., Zhao, C.-J., Zu, Y.-G., Li, C.-Y., Wang, W., Li, J., and Wei, Z.-F. (2013). Acidic pH based Microwave-assisted Aqueous Extraction of Seed Oil from Yellow Horn (*Xanthoceras sorbifolia* Bunge.). *Industrial Crops* and Products. 43, 420–426.

Radovanovi, N., Corovi, M., and Siler-marinkovi, S. (2015). Optimisation of

Microwave-Assisted Extraction Parameters for Antioxidants from Waste *Achillea Millefolium* Dust. *Industrial Crops and Products*. 77, 333–341.

- Rakotondramasy-rabesiaka, L., Havet, J., Porte, C., and Fauduet, H. (2008). Solid liquid Extraction of Protopine from *Fumaria Officinalis* L .— Kinetic Modelling of Influential Parameters. *Industrial Crops and Products*. 29, 516– 523.
- Ranic, M., Nikolic, M., Pavlovic, M., Buntic, A., Siler-Marinkovic, S., and Dimitrijevic-Brankovic, S. (2014). Optimization of Microwave-assisted Extraction of Natural Antioxidants from Spent Espresso Coffee Grounds by Response Surface Methodology. *Journal of Cleaner Production*. 80, 69–79.
- Ratnayake, W. M. N., and Galli, C. (2009). Fat and Fatty Acid Terminology, Methods of Analysis and Fat Digestion and Metabolism. *Annals of Nutrition* and Metabolism. 55(1-3), 8–43.
- Richardson, A. J. (2006). Omega-3 Fatty Acids in ADHD and Related Neurodevelopmental Disorders. *International Review of Psychiatry*, 18(2), 155– 72.
- Rizza, C. S. (2014). Experiments and Modeling of Supercritical CO₂ Extraction of Lipids from Microalgae. Master Thesis. University of Padua
- Rohman, A., and Man, Y. B. C. (2012). The Chemometrics Approach Applied to FTIR Spectral Data for the Analysis of Rice Bran Oil in Extra Virgin Olive Oil. *Chemometrics and Intelligent Laboratory Systems*. 110(1), 129–134.
- Rohman, A., Riyanto, S., Sasi, A. M., and Yusof, F. M. (2014). The use of FTIR Spectroscopy in Combination with Chemometrics for the Authentication of Red Fruit (*Pandanus conoideus* Lam) Oil from Sunflower and Palm oils. *Food Bioscience*. 7, 64–70.
- Rubio-Rodríguez, N., de Diego, S. M., Beltrán, S., Jaime, I., Sanz, M. T., and Rovira, J. (2008). Supercritical Fluid Extraction of the Omega-3 Rich Oil Contained in Hake (*Merluccius capensis–Merluccius paradoxus*) by-products: Study of the Influence of Process Parameters on the Extraction Yield and Oil Quality. *The Journal of Supercritical Fluids*. 47(2), 215–226.
- Sánchez-camargo, A. P., Meireles, M. Â. A., Ferreira, A. L. K., Saito, E., and Cabral, F. A. (2012). Extraction of ω-3 Fatty Acids and Astaxanthin from Brazilian Redspotted Shrimp Waste using Supercritical CO₂ + Ethanol Mixtures. *The Journal of Supercritical Fluids*. 61, 71–77.

- Sánchez-Pérez, R., Arrázola, G., Martín, M. L., Grané, N., and Dicenta, F. (2012). Influence of the Pollinizer in the Amygdalin Content of Almonds. *Scientia Horticulturae*. 139, 62–65
- Saub, R., and Jaafar, N. (2001). A Dental-anthropological Study of Health and Illness Behaviour Among Orang Asli of the Semai Tribe: the Perspective of Traditional Healers. *The Medical Journal of Malaysia*. 56(4), 401–7.
- Schacky, C. von, and Harris, W. S. (2007). Cardiovascular Benefits of Omega-3 Fatty Acids. *Cardiovascular Research*. 73(2), 310–315.
- Setyaningsih, W., Saputro, I. E., Palma, M., and Barroso, C. G. (2015). Optimisation and Validation of the Microwave-assisted Extraction of Phenolic Compounds from Rice Grains. *Food Chemistry*. 169, 141–149.
- Siddhuraju, P., Vijayakumari, K., and Janardhanan, K. (1995). Nutrient and Chemical Evaluation of Raw Seeds of *Xylia Xylocarpa*: An Underutilized Food Source. *Journal of Food Chemistry*. 53, 299–304.
- Simopoulos, A. P. (2002). Omega-3 Fatty Acids in Wild Plants, Nuts and Seeds. Asia Pacific Journal of Clinical Nutrition. 11(S6), S163–S173.
- Simopoulos, A. P. (2007). Omega-3 Fatty Acids and Athletics. Current Sports Medicine Reports. 6(4), 230–6.
- So, G.C., and Macdonald, D.G. (1986). Kinetics of Oil Extraction from Canola (Rapeseed). *Canadian of Journal Chemistry Engineering*. 64, 80-86.
- Sovová, H. (2005). Mathematical Model for Supercritical Fluid Extraction of Natural Products and Extraction Curve Evaluation. *Journal of Supercritical Fluids*. 33(1), 35–52.
- Sovová, H. (2012). Modeling the Supercritical Fluid Extraction of Essential Oils from Plant Materials. *Journal of Chromatography A*. 1250, 27–33.
- Spigno, G., and Faveri, D. M. De. (2009). Microwave-assisted Extraction of Tea Phenols : A Phenomenological Study. *Journal of Food Engineering*. 93(2), 210– 217.
- Spiro, M. (1988). The Rate of Caffeine Infusion from Kenyan Arabica Coffee Beans. 12th Colloque Scientifique International du Cafejä. 260-264.
- Stefanidis, G.D., Munoz, A. N., Sturm, G.S. J., Stankiewicz, A. (2014). A Helicopter View to Microwave Application to Chemical Processes: Reactions, Separations, and Equipment Concepts. Reviews in Chemical Engineering. 30(3), 233-259.

Stoewsand, G. S., Anderson, J. L., and Lamb, R. C. (1975). Cyanide Content of

Apricot Kernels. Journal of Food Science. 40, 1107-1115.

- Swanson, D., Block, R., and Mousa, S. A. (2012). Omega-3 Fatty Acids EPA and DHA : Health Benefits Throughout Life. *An International Review Journal*. 1–7.
- Taghvaei, M., Jafari, S. M., Assadpoor, E., Nowrouzieh, S., and Alishah, O. (2014). Optimization of Microwave-assisted Extraction of Cottonseed Oil and Evaluation of its Oxidative Stability and Physicochemical Properties. *Food Chemistry*. 160, 90–7.
- Takeuchi, T. M., Pereira, C. G., Braga, M. E. M., Maróstica, M. R. Jr., Leal, P. F., Meirele, M. A. A. (2009). Low-Pressure Solvent Extraction (Solid–Liquid Extraction, Microwave-Assisted, and Ultra-Sound-Assisted) from Condimentary Plants. Meireles M. A. A. (ed). Extracting Bio- active Compounds for Food Products (pp. 137–218). Boca Raton : CRC Press/Taylor & Francis.
- Temelli, F., Saldaña, M. D. A., Moquin, P. H. L., and Sun, M. (2008). Supercritical Fluid Extraction of Specialty Oils. Martínez, J. L. (Ed). Supercritical Fluid Extraction of Nutraceuticals and Bioactive Compounds (pp. 52-99). London: CRC Press/Taylor & Francis.
- Thirugnanasambandham, K., Sivakumar, V., and Maran, J. P. (2014). Microwave-Assisted Extraction of Polysaccharides from Mulberry Leaves. *International Journal of Biological Macromolecules*. 72C, 1–5.
- Thirugnanasambandham, K., Sivakumar, V., and Prakash Maran, J. (2014). Process Optimization and Analysis of Microwave Assisted Extraction of Pectin from Dragon Fruit Peel. *Carbohydrate Polymers*. 112, 622–626.
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G., and kaur, H. (2011). Phytochemical Screening and Extraction: A Review. *Internationale Pharmaceutica Sciencia*. 1, 98–106.
- Tamborrino, A., Romaniello, R., Zagaria, R., and Leone, A. (2014). Microwave-Assisted Treatment for Continuous Olive Paste Conditioning: Impact on Olive Oil Quality and Yield. *Biosystems Engineering*. 127,92-102.
- Tuncel, G., and Nout, M. J. R. (1998). Degradation of Cyanogenic Glycosides of Bitter Apricot Seeds (*Prrcnus armeniaca*) by Endogenous and Added Enzymes as Affected by Heat Treatments and Particle Size. *Food Chemistry*. 63(1), 65– 69.
- Tzang, B. S., Yang, S. F., Fu, S. G., Yang, H. C., Sun, H. L., and Chen, Y. C. (2009). Effects of Dietary Flaxseed Oil on Cholesterol Metabolism of Hamsters. *Food*

Chemistry. 114(4), 1450–1455.

- Vadivel, V., and Janardhanan, K. (2001). Diversity in Nutritional Composition of Wild Jack Bean (*Canavalia ensiformis* L. DC) Seeds Collected from South India. *Journal of Food Chemistry*. 74, 507–511.
- Van Sam, H., and Van Welzen, P. C. (2004). Revision of Annesijoa, Elateriospermum and the Introduced Species of Hevea in Malesia (Euphorbiaceae). *Blumea - Biodiversity, Evolution and Biogeography of Plants*. 49(2), 425–440.
- Veggi, P.C., Martinez, J., and Meireles, M. A. A. (2013). Fundamentals of Microwave Extraction. Chemat, F., and Cravotto, G. (eds). Microwave-assisted Extraction for Bioactive Compounds. (pp.15-52). London: Springer Science.
- Vilaplana, F., Ribes-Greus, A., and Karlsson, S. (2009). Microwave-assisted Extraction for Qualitative and Quantitative Determination of Brominated Flame Retardants in Styrenic Plastic Factions from Waste Electrical and Electronic Equipment (WEEE). *Talanta*. 78(1), 33–9.
- Virot, M., Tomao, V., Ginies, C., Visinoni, F., and Chemat, F. (2008). Microwave-Integrated Extraction of Total Fats and Oils. *Journal of Chromatography*. A. 1196-1197, 57–64.
- Vlachos, N., Skopelitis, Y., Psaroudaki, M., Konstantinidou, V., Chatzilazarou, a, and Tegou, E. (2006). Applications of Fourier Transform-infrared Spectroscopy to Edible Oils. *Analytica Chimica Acta*. 573-574, 459–65.
- Vrablík, M., Prusíková, M., Snejdrlová, M., & Zlatohlávek, L. (2009). Omega-3 Fatty Acids and Cardiovascular Disease Risk: Do We Understand the Relationship?. *Physiological Research*. 58(S1), S19–26.
- Wang, Y., You, J., Yu, Y., Qu, C., Zhang, H., Ding, L., Zhang, H., and Li, X. (2008). Analysis of Ginsenosides in Panax Ginseng in High Pressure Microwave-Assisted Extraction. *Food Chemistry*. 110, 161–167.
- Wasserkrug, K. and ElRassi, Z. (1997). High-Performance Liquid Phase Separation of Glycosides. 1. Reversed Phase Chromatography of Cyanogenic Glycosides with UV and Pulsed Amperometric Detection. *Journal Liquid Chromatography Related Technology*. 20: 335–49.
- Wyatt, J.S., (1979). *Pocket Check List of Timber Tree*. Malayan Forest Record No.17. Forest Department Peninsular Malaysia.
- Xavier, V. B., Vargas, R. M. F., Cassel, E., Lucas, A. M., Santos, M. A., Mondin, C.

A., Santarem, E. R., Astarita, L.V., and Sartor, T. (2011). Mathematical Modeling for Extraction of Essential Oil from *Baccharis spp.* by Steam Distillation. *Industrial Crops and Products*. 33(3), 599–604.

- Xu, H., Huang, W., and He, C. (2008). Modeling for Extraction of Isoflavones from Stem of *Pueraria Lobata* (Willd.) Ohwi using n -butanol/Water Two-Phase Solvent System, 62, 590–595.
- Yang, Y., Li, J., Zu, Y., Fu, Y., Luo, M., Wu, N., and Liu, X. (2010). Optimisation of Microwave-Assisted Enzymatic Extraction of Corilagin and Geraniin from Geranium Sibiricum Linne and Evaluation of Antioxidant Activity. *Food Chemistry*. 122(1), 373–380.
- Yanık, D. K. (2017). Alternative to Traditional Olive Pomace Oil Extraction Systems: Microwave-Assisted Solvent Extraction of Oil from Wet Olive Pomace. *Food Science and Technology*. 77, 45-51.
- Yilmaz, E. E., Özvural, E. B., and Vural, H. (2011). Extraction and Identification of Proanthocyanidins from Grape Seed (*Vitis Vinifera*) using Supercritical Carbon Dioxide. *The Journal of Supercritical Fluids*. 55(3), 924–928.
- Yong, O. Y., and Salimon, J. (2006). Characteristics of *Elateriospermum Tapos* Seed Oil as a New Source of Oilseed. *Industrial Crops and Products*. 24(2), 146– 151.
- Zhang, G., Hu, M., He, L., Fu, P., Wang, L., and Zhou, J. (2013). Optimization of Microwave-Assisted Enzymatic Extraction of Polyphenols from Waste Peanut Shells and Evaluation of its Antioxidant and Antibacterial Activities in Vitro. *Food and Bioproducts Processing*. 91(2), 158–168.
- Zhang, Y., Li, S., Yin, C., Jiang, D., Yan, F., and Xu, T. (2012). Response Surface Optimisation of Aqueous Enzymatic Oil Extraction from Bayberry (*Myrica Rubra*) Kernels. *Food Chemistry*. 135(1), 304–308.
- Zlotorzynski, A. (1995). The Application of Microwave Radiation to Analytical and Environmental Chemistry. *Critical Reviews in Analytical Chemistry*. 25, 43-75