

DYNAMIC DEGRADATION OF POROUS MAGNESIUM UNDER  
SIMULATED ENVIRONMENT OF HUMAN CANCELLOUS BONE

AMIR PUTRA BIN MD SAAD

UNIVERSITI TEKNOLOGI MALAYSIA

DYNAMIC DEGRADATION OF POROUS MAGNESIUM UNDER  
SIMULATED ENVIRONMENT OF HUMAN CANCELLOUS BONE

AMIR PUTRA BIN MD SAAD

A thesis submitted in fulfilment of the  
requirement for the award of the degree of  
Doctor of Philosophy (Mechanical Engineering)

Faculty of Mechanical Engineering  
Universiti Teknologi Malaysia

FEBRUARY 2017

This present work is specially dedicated to all my family members, *where life begins and loves never ends.*

To my wife, *Noor Faizah Che Ahmad*, who always supports, loves and taking care our childrens, *Wafa' Safiyyah and Alyaa Safiyyah*, while I'm struggling completing this work.

To my beloved parent, *Md Saad Man and Aishah Itam*, who continuously motivates and taught me to be a man that benefit others.

*Thank you for being in my life.*

## ACKNOWLEDGEMENT

In the name of Allah, the Beneficent, the Merciful.

Praise be to Allah, Lord of the worlds.

Vision, insight, ideas and faith is a huge gift in the completion of this study. I am very grateful to be given the opportunity to study and explore the profound knowledge in this field.

I would like to express my sincere appreciation to Dr Ardiyansyah Syahrom guidance, patience, inspire motivation, support and assistance during the completion of this study. He is a lecturer, researcher, brother and a friend who has taught me to be the man who acts with true knowledge. May Allah make it easy and rewarding to you with all the contributions that have been given.

I would like also to express my gratitude to our mentor, Prof. Dato' Ir. Dr Mohamed Rafiq Bin Dato' Hj. Abdul Kadir for endless supports to attain purpose of this life as well as researcher.

Thanks to all my colleagues, especially on Medical Devices Technology Group (MediTeg) and Sports and Innovation Technology Centre (SITC), and supportive lecturers from the Faculty of Mechanical Engineering for their priceless guidance and encouragement. Finally, I would like to say thanks to all those who have involved and helped me in any aspect to complete the study.

## ABSTRACT

Biodegradable metals have been suggested for bone scaffold applications due to their mechanical properties that are better for load bearing applications. Among biodegradable metals, magnesium and its alloy are the most investigated materials due to their mechanical properties which are closer to the cancellous bone and could prevent complications such as an aseptic loosening of stress shielding effects, and potentially to be used as bone scaffolds. Bone adapts the mechanical loading from the physiological activities that induced the movement of bone marrow passing through the porous structure of cancellous bone due to the pressure differences. The aim of this research is to analyse the degradation behaviour of porous magnesium under dynamic degradation test for bone scaffold applications. Interconnected holes of porous magnesium have been developed with various percentages of porosity (30%, 41% and 55%) and are fabricated using computer numerical control (CNC) machine. Dynamic immersion test rigs are specifically designed to simulate environment of human cancellous bone. There are two types of tests that have been conducted in this study: (1) fluid flow with different flowrates (0.025, 0.4 and 0.8 ml/min) and (2) fluid flow integrated cyclic loading (different cyclic loading (1000, 2000 and 3500  $\mu\epsilon$ ) under constant flowrate of 0.025 ml/min). A dynamic immersion test has been conducted for 24, 48 and 72 hours. The results showed that the specimen with a higher percentage of porosity as well as the exposed surface area degrades faster compared to the others. The degradation product formation and clogging pores phenomenon are influenced by the level of flow rates. The effects of different flow rates towards the mechanical integrity of porous magnesium have shown a huge drop of 95% from their original mechanical properties within 3 days, which have deteriorated in both functions; porosity and degradation time. The variation in flowrates used showed that degradation of the material is seven times higher compared to the static immersion test environment. Furthermore, the influenced of integrating fluid flow and cyclic loading have increased the relative weight loss and degradation rate as high as 61.56% and 93.67%, respectively. Additionally, the mechanical properties have improved and increased from 53% to 87% as compared to dynamic immersion test using the mechanical stimulus of fluid flow only. Therefore, the dynamic immersion test with integrated cyclic loading was more reliable and provides realistic environment for degradation assessment compared to static immersion test for bone scaffold application as this study using the boundary of human cancellous bone environment.

## ABSTRAK

Logam terbiodegradasi dicadangkan untuk aplikasi penggantian tulang disebabkan oleh sifat-sifat mekanik yang lebih baik bagi penggunaan galas beban. Dikalangan logam terbiodegradasi, magnesium dan paduannya adalah yang paling dikaji kerana sifat mekanikal mereka yang lebih dekat dengan tulang kanselus dan boleh mencegah komplikasi seperti aseptik yang merenggangkan kesan perisaian tekanan, dan ianya berpotensi untuk digunakan sebagai penggantian tulang. Tulang dapat menyesuaikan bebanan mekanikal daripada aktiviti fisiologi yang mengaruhkan pergerakan sum-sum tulang melalui struktur poros tulang kanselus kerana perbezaan tekanan. Tujuan kajian ini adalah untuk menganalisis kelakuan degradasi magnesium berliang di bawah ujian degradasi dinamik untuk aplikasi penggantian tulang. Lubang saling magnesium berliang telah dibangunkan dengan pelbagai peratusan keliangan (30%, 41% dan 55%) dan direka menggunakan mesin kawalan berangka terkomputer (CNC). Pelantar ujian rendaman dinamik direka khusus untuk mensimulasikan persekitaran tulang kanselus manusia. Terdapat dua jenis ujian yang telah dijalankan dalam kajian ini: (1) aliran cecair dengan kadar aliran yang berbeza (0.025, 0.4 dan 0.8 ml/min) dan (2) aliran cecair dipadukan dengan kitaran beban (kitaran beban yang berbeza (1000, 2000 dan 3500  $\mu\epsilon$ ) di bawah kadar aliran malar 0.025 ml/min). Ujian rendaman dinamik telah dijalankan untuk 24, 48 dan 72 jam. Keputusan menunjukkan bahawa spesimen dengan peratusan yang lebih tinggi keliangan serta kawasan permukaan yang terdedah lebih cepat degradasi berbanding dengan yang lain. Pembentukan produk degradasi dan fenomena liang tersumbat dipengaruhi oleh tahap kadar aliran. Kesan kadar aliran yang berbeza terhadap keutuhan mekanikal magnesium berliang telah menunjukkan penurunan yang besar sebanyak 95% dari sifat-sifat mekanikal asalnya dalam tempoh 3 hari, yang telah merosot dalam kedua-dua fungsi; keliangan dan masa degradasi. Variasi kadar aliran yang digunakan menunjukkan bahawa degradasi bahan adalah tujuh kali lebih tinggi berbanding dengan persekitaran ujian rendaman statik. Tambahan pula, dipengaruhi oleh paduan aliran bendalir dan kitaran beban telah meningkat penurunan berat relatif dan kadar degradasi setinggi 61.56% dan 93,67%, masing-masing. Selain itu, sifat-sifat mekanikal telah bertambah baik dan meningkat daripada 53% kepada 87% berbanding dengan ujian rendaman dinamik menggunakan rangsangan mekanikal aliran bendalir sahaja. Oleh itu, ujian rendaman dinamik dipadukan dengan kitaran bebanan adalah lebih dipercayai dengan persekitaran realistik untuk penilaian degradasi berbanding dengan ujian rendaman statik untuk applikasi penggantian tulang kerana kajian ini menggunakan sempadan persekitaran tulang kanselus manusia.

## TABLE OF CONTENTS

<b>CHAPTER</b>	<b>TITLE</b>	<b>PAGE</b>
	<b>DECLARATION</b>	ii
	<b>DEDICATION</b>	iii
	<b>ACKNOWLEDGEMENT</b>	iv
	<b>ABSTRACT</b>	v
	<b>ABSTRAK</b>	vi
	<b>TABLE OF CONTENTS</b>	vii
	<b>LIST OF TABLES</b>	xi
	<b>LIST OF FIGURES</b>	xii
	<b>LIST OF SYMBOLS</b>	xvi
	<b>LIST OF ABBREVIATIONS</b>	xvii
	<b>LIST OF APPENDICES</b>	xviii
<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
	1.1 Background of the Study	1
	1.2 Problem Statements	4
	1.3 Objectives	5
	1.4 Scopes	6
	1.5 Significance of the Study	7
	1.6 Thesis Structure and Organization	7

<b>2</b>	<b>LITERATURE REVIEW</b>	<b>9</b>
2.1	Human Skeletal and Bone Structure	9
2.1.1	Bone Remodelling	10
2.1.2	Mechanical Stimuli as Regulator in Bone Remodelling	14
2.2	Biodegradable Materials as Bone Scaffolds	16
2.2.1	Biomimetic of Porous Scaffolds	18
2.2.2	Porous Magnesium for Bone Scaffolds Application	21
2.3	Degradation Mechanism of Magnesium in Body Fluid	23
2.3.1	Analytical Methods of the Degradation Rate	24
2.3.2	Degradation Rate of Magnesium in Body Fluid	25
2.4	Porous Magnesium: Concepts and Perspectives	29
2.5	Summary	31
<b>3</b>	<b>METHODOLOGY</b>	<b>32</b>
3.1	Research Flow Diagram	32
3.2	Specimens Preparation	33
3.3	Preparation of Body Fluid Solutions	36
3.4	In-Vitro Dynamic Immersion Tests	37
3.4.1	Body Fluid Passing Through Porous Magnesium	39
3.4.2	Porous Magnesium under Fluid Flow Integrate Cyclic Loading	42
3.5	Degradation Rate Determination	45
3.6	Material Characterization	45
3.7	Scanning Specimens Procedure	47
3.8	Compression Test Procedure	48
3.9	Summary	48



<b>4</b>	<b>DEGRADATION OF POROUS MAGNESIUM UNDER DYNAMIC IMMERSION TEST USING CONSTANT FLOW RATE</b>	<b>50</b>
4.1	Overview	50
4.2	Results	51
4.2.1	Morphology of Specimen and Corrosion Product Identification	51
4.2.2	Degradation Rate	54
4.2.3	Changes in pH and Suction Pressure	56
4.2.4	Mechanical Properties Degradation of Porous Magnesium	57
4.3	Discussion	60
<b>5</b>	<b>INFLUENCE OF DIFFERENT FLOW RATES ON DEGRADATION BEHAVIOUR OF POROUS MAGNESIUM UNDER DYNAMIC IMMERSION TEST</b>	<b>66</b>
5.1	Overview	66
5.2	Results	67
5.2.1	Morphologies of Specimens Under Different Flow Rates	67
5.2.2	Degradation Products Under Different Flow Rates	68
5.2.3	Different Flow Rates Induced Extensive Degradation	71
5.2.4	Effect of Different Flow Rates Towards pH and Suction Pressure	72
5.2.4	Mechanical Properties Under Different Flow Rates	75
5.3	Discussion	77

<b>6</b>	<b>EFFECTS OF CYCLIC LOADING ON DEGRADATION OF POROUS MAGNESIUM UNDER DYNAMIC IMMERSION TEST USING CONSTANT FLOW RATE</b>	<b>83</b>
6.1	Overview	83
6.2	Results	84
6.2.1	Morphology Specimens Under Dynamic Immersion Integrated Different Loading	84
6.2.2	Degradation Products Formation	85
6.2.3	Degradation Behaviour Under Dynamic Immersion with Different Cyclic Loading	87
6.2.4	Effects of Cyclic Loading in Dynamic Immersion Test Towards pH and Suction Pressure	90
6.2.5	Effects of Cyclic Loading on Mechanical Properties of Porous Mg Under Dynamic Immersion Test	91
6.2.6	Effects of Different Cyclic Loading on Secants Modulus Degradation of Porous Mg	93
6.3	Discussion	95
<b>7</b>	<b>CONCLUSION AND FUTURE RECOMENDATIONS</b>	<b>101</b>
7.1	Conclusion	101
7.2	Limitation and Future Recommendation	103
	<b>REFERENCES</b>	<b>104</b>
	Appendices A - F	120 - 143

## LIST OF TABLES

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Mechanical properties of bone.	13
2.2	Flow rates of different sites and medium.	15
2.3	Mechanical properties of porous scaffolds.	20
2.4	The average degradation rate (mm/yr) of Mg and its alloys in in-vitro and in-vivo conditions.	28
2.5	Bone healing periods at different fracture sites.	30
3.1	The chemical composition (in wt. %) of the commercially pure magnesium 99.9% purity.	34
3.2	The morphologic details of the porous pure magnesium specimens.	36
3.3	Composition for preparing 1 litre SBF solutions.	37
3.4	Experimental planning for fluid flow	41
3.5	Experimental planning for fluid flow integrate cyclic loading	44
4.1	Comparison between the degradation rate of porous magnesium.	62
4.2	The mechanical properties of porous magnesium compared to human bone.	65
5.1	Comparison of dynamic and static immersion degradation rate of pure Mg.	79
6.1	Comparison degradation rate of porous Mg under dynamic immersion test with different condition.	97

## LIST OF FIGURES

<b>FIGURE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Human skeleton and bone structure.	10
2.2	Bone remodelling process	12
2.3	Porous Mg Scaffolds: (a) AZ91D using NaCl spacer, (b) honeycomb structure using laser perforation techniques, (c) porous Mg using Ti wires, (d) porous Mg using fibre deposition hot pressing (FDHP) and (e) lotus type using metal/gas eutectic unidirectional solidification.	22
2.4	Degradations trends (a) percentage of weight loss, (b) hydrogen volume and (c) degradation rate of Mg and its alloys	27
2.5	The schematic diagram of concept bone healing and degradation behaviour of bone scaffolds.	29
3.1	Research flow diagram.	33
3.2	Specimens Preparations.	34
3.3	Photograph of three types of porous Mg specimens.	35
3.4	Schematic diagram of in vitro dynamic immersion test rig.	38
3.5	The dynamic immersion test rig of fluid passing through the porous Mg.	40

3.6	Relationship between suction pressure and flow rate under dynamic immersion system. (Note: selected flow rate was highlighted using dotted line circle)	40
3.7	The dynamic immersion test rig of fluid flow integrates cyclic loading.	43
3.8	A typical hysteresis loop stress-strain curve of compressive cyclic loading using strain controlled using cyclic loading of 1000 $\mu\epsilon$ for specimen A.	43
3.9	A vacuum pump and chamber equipment.	46
3.10	A Field Emission Scanning Electron Microscope (FESEM) equipment.	46
3.11	X-ray diffractometer (XRD).	47
3.12	Micro-computed tomography ( $\mu$ CT) scans equipment.	48
3.13	A universal testing machine equipment.	49
4.1	Morphology comparison of the specimens after the dynamic immersion tests before and after cleaning.	51
4.2	SEM images and EDS spectra of: (a) needle-like formations and (b) rod-like formations found on the surface, and (c) a uniform corrosion product that has cracked on drying with spherical deposits found on the pore.	53
4.3	The XRD patterns of degraded porous magnesium specimen C in dynamic immersion at 24, 48, and 72 hours.	54
4.4	Degradation rate of the porous magnesium specimens after immersion for 24, 48 and 72 hours were calculated based on: (a) relative weight loss and (b) degradation rate.	55
4.5	Changes in pH of the SBF solution after dynamic immersion over time.	56
4.6	Suction pressure profile of the solution during dynamic immersion test over 72 hours for specimen B (41%).	57

4.7	Typical compressive stress-strain curves of the porous magnesium specimens in comparison to solid pure magnesium before and after 72 hours of dynamic immersion tests.	58
4.8	The mechanical properties of the porous magnesium specimens: (a) yield strength, (b) compressive strength, and (c) Young's modulus.	59
4.9	Schematic illustration of the degradation process of, and SBF flow interaction in porous magnesium specimens during the dynamic immersion test.	63
5.1	Morphology of the specimens after 72 hours of dynamic immersion test.	68
5.2	SEM and EDS spectra of: (a) needle-like formation on the surface, (b) network-like crack with ball-like deposit found on the pore, and (c) rod-like formation, (d) clod rod-like formation, (e) flower-like formation, and (f) rod-plate-like formation on the surface.	69
5.3	XRD patterns of degraded porous Mg in dynamic immersion at 0.025, 0.4 and 0.8 ml/min.	71
5.4	Degradation rate of the porous Mg specimens at different flowrates and time immersion. (Note: X, Y and Z were referred to flowrate of 0.025, 0.4 and 0.8 ml/min respectively)	73
5.5	Changes of pH of the SBF solution after dynamic immersion over time at different flowrate.	74
5.6	Suction pressure profile of the solution during the dynamic immersion test over 72 hours at varying flowrate.	75
5.7	Mechanical properties of porous Mg specimens under different flowrate: (a) yield strength, (b) compressive strength, and (c) Young's modulus.	76
6.1	Morphology of the specimens with different cyclic loading after 72 hours of dynamic immersion test.	85

6.2	SEM and EDS spectra of: (a) needle-like formation, (b) crags-like formation, and (c) flake-like formation.	86
6.3	XRD patterns of degraded porous Mg in dynamic immersion test integrated different cyclic loading.	87
6.4	Degradation rate of the porous Mg specimens under dynamic immersion test integrating different cyclic loading for 24, 48 and 72 hours. (Note: L1, L2 and L3 were referred to flowrate of 1000, 2000 and 3500 $\mu\text{e}$ respectively)	89
6.5	Changes of pH of the SBF solution under dynamic immersion test integrated different cyclic loading.	90
6.6	Mechanical properties of porous Mg specimens under dynamic immersion integrated different cyclic loading: (a) yield strength, (b) compressive strength, and (c) Young's modulus.	91
6.7	Degradation of secant modulus of (a) the specimen B under 3500 $\mu\text{e}$ cyclic compressive for dry (control) and dynamic immersion, (b) the specimen B under different cyclic compressive loading in dynamic immersion test and (c) different specimen under 2000 $\mu\text{e}$ cyclic compressive in dynamic immersion test.	94

**LIST OF SYMBOLS**

$P_m$	-	Degradation Rate in Units of Penetration
$\Delta W_m$	-	Degradation Rate in Units of Weight Change
$\rho$	-	Density
$W_f$	-	Final Weight
$W_o$	-	Initial Weight
$d_h$	-	Inner diameter
$E_N$	-	Secants Modulus
$\varepsilon$	-	Strain
$\Delta\varepsilon$	-	Strain Range
$\sigma$	-	Stress
$\Delta\sigma$	-	Stress Range
$v$	-	Velocity
$\mu$	-	Viscosity
$E$	-	Young's Modulus



## LIST OF ABBREVIATIONS

ASTM	-	American Society for Testing and Materials
CO <sub>2</sub>	-	Bicarbonate
CP-Mg	-	Commercially pure magnesium
CAGR	-	compound annual growth rate
CAD	-	computer aided design
CNC	-	Computer Numeric Control
DAQ	-	Data Acquisition
DR	-	Degradation rate
DMEM	-	Dulbecco's Modified Eagle Medium
EDS	-	Energy Dispersive Spectrometer
FDHP	-	fibre deposition hot pressing
HP-Mg	-	high purity magnesium
HA	-	hydroxyapatite
IOF	-	International Osteoporosis Foundation
Mg	-	Magnesium
μ-CT	-	micro computed tomography
μm	-	micro-meter
με	-	micro-strain
NDE	-	negative difference effect
RANKL	-	receptor activator of nuclear factor kappa B ligand
Re	-	Reynolds number
SEM	-	Scanning Electron Microscope
SBF	-	Simulated Body Fluid
NaCl	-	Sodium Chloride (Salt)
TSWH	-	titanium wire space holder
XRD	-	X-ray diffractometer

**LIST OF APPENDICES**

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
A	Other SEM images of corrosion product observed under dynamic immersion test at constant flow rate	121
B	Other SEM images of corrosion product observed under dynamic immersion test integrated cyclic loading	125
C	Sample of pressure data for suction and discharge	129
D	Sample of compression test data	132
E	Sample of cyclic loading test data	137
F	List of publications	143

## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of the Study**

Bone grafting is a medical procedure for bone replacement or repairs. It is the second most highly performed surgical procedures on tissue transplantation after blood transfusion [1]. The usage frequency of bone graft has been paramount every year. The transparency market research has reported that due to the rising demand for bone graft substitutes, its market is expected to expand at a 4.5% compound annual growth rate (CAGR) between 2015 and 2023, and it is estimated to be worth USD 3.48 billion by 2023 [2]. Over two million orthopaedics procedures on bone-graft substitutes have been annually performed worldwide, with over 400,000 and 600,000 procedures recorded in Europe and the United States, respectively [3–6]. Asian Audit 2009 has reported from International Osteoporosis Foundation (IOF) [7] that in China, almost 69.4 million of people over the age of 50 years old suffering from osteoporosis. This huge number includes 0.687 and 1.8 million of hip and vertebral fractures, respectively stirring each year. Hong Kong and Singapore have demonstrated in the past four decades that hip fractures have remarkably increased in number by 300% and 500%, correspondingly. In Japan, 12 million people suffering from osteoporosis and the incidence rate of hip fractures are increasing radically among both men and women of age 75 years old and above.

The amplifying prevalence of bone and joint disorder such as deformities, trauma, tumor, degenerative and aging population have been accompanied by the increase in number of orthopaedics reconstructions. This has driven the high demand on bone graft substitutes. Bone-graft substitutes have been established with different types that are available in the worldwide market. The common types of bone-graft substitutes used are autografts, allografts and xenografts; which the existence depends on their sources. Autografts can be referred to as the gold standard in medical procedure to repair damaged bone or for bone replacement. It provides the best osteo-conductive, osteo-inductive and osteo-genic properties since their sources are from the parts of the patient's body [8]. As for allografts and xenografts, their tissues sources come from different members of the same species and different species respectively. However, those bone-graft substitutes have exhibited a vital concern of donor site morbidity (autografts), and limited supply and possibility of pathogen transmission and immune-rejection (allografts and xenografts) [9,10]. Therefore, researchers have developed the new generation of synthetic bone-grafts substitutes to eliminate susceptibility of the aforementioned drawbacks [11]. Hence, with the advancement of technology in biomaterials engineering, the biodegradable materials have attracted researchers to investigate a lot in favour of obtaining the ideal synthetic bone-grafts substitutes or known as bone scaffolds [12–14].

Biodegradable materials have been acknowledged as an ideal model in biomaterials that has inspired researchers to focus on. These materials serve as a device to provide temporary support for tissue regeneration while bone heals and gradually degrades after fulfilling its function [15]. Among biodegradable materials, polymer can be classified as an excellent material due to its good biodegradability, biocompatibility and easy to fabricate [16,17]. However, the advantages of polymer are retarded due to its low mechanical properties for load bearing applications [18,19]. This has led to the use of biodegradable metal which possesses good mechanical properties [14]. In comparison to iron-based and newly introduced zinc alloys, magnesium and its alloys are the most investigated biodegradable metals for their potential application as biomedical implants [12,14]. They have shown an excellent performance to human bone in terms of mechanical integrity and their mechanical properties (41-45 GPa of Young's Modulus [14,20]) is close to cortical bone (3–23 GPa of Young's Modulus [14,21]) while cancellous bone

(0.01–3.0 GPa of Young's Modulus [22]) and bioactivity of Mg stimulatory effects have induced the growth of new formation of bone-apatite like of hydroxyapatite (HA) crystallization [14,23,24] and are favourable to bone strength [25]. These two factors have favoured the idea of using biodegradable metals to be used as the materials for bone tissue engineering [26–28].

Recent advancements in bone tissues engineering is to develop the multifunctional capabilities of the scaffold to be well-integrated with the biological environment and physiological functions of natural bone [12,29]. Bone scaffolds are typically required to have porous structure to allow nutrient to be transported from the surrounding tissues and releases waste disposal from the regenerated tissues [30,31]. Ideally, this porous structure will have 25-90% porosity and a 10-1000  $\mu\text{m}$  pore size to provide an ideal condition for infiltration of essential nutrients, oxygen, and progenitor cells for cell survivability [32,33]. The porosity of porous structure can be controlled and regulated to a desired form. Though the employment of porous structure reduces the mechanical properties, it is still an advantage in obtaining the scaffold that has the mechanical properties which are well-matched with natural bone. Researchers have investigated the strength of porous scaffolds using polymers, ceramics, composites and metals. It is suggested to use metals due to their mechanical properties that are close to the mechanical properties of bone [34].

Biomechanically, cancellous bone adapts to the mechanical loading from the perpetual motion of physiological activities through the mechanobiological signalling of osteocytes [35,36]. Cancellous bone adapts the compressive strain level of 1000 – 3000  $\mu\epsilon$  that is generated from various activities and beyond 3500  $\mu\epsilon$  leads to bone fracture [37,38]. Due to the cyclic motion of compressive strain, it causes the bone marrow, which is the home for progenitor cells of osteoblasts and osteoclasts, moves as a fluid medium with a flowrate range of 0.0072-1.67 ml/min [36,39–42]. The interaction between the bone marrow movement and the cancellous bone structure induces mechanical stresses that stimulate the mechanobiological response to the bone quality and bone healing process [36]. The movement of bone marrow through the porous structure of cancellous bone due to pressure differences is generated by continuous cycles of mechanical loading

from physiological activities [43]. This cyclic motion of compressive strain and bone marrow movement in the cancellous bone must be considered as an actual boundary for testing the biodegradable materials of bone scaffolds.

## 1.2 Problem Statements

Bone scaffold were developed by a wide range of the biomaterials. Instead of metallic biomaterials, others than that have been produced is unsuitable for load bearing purposes [44]. By using load bearing bone scaffold, patients will able to speed up in performing their daily lives activities which could also contribute to a better healing process [35]. The metallic biomaterials that already approved and commonly used as biomedical implant are stainless steels, titanium and cobalt chromium based alloys [14]. However, the suitability to be acted as an ideal scaffold for bone was degraded by the possible release of toxic metallic ions and poor stimulation of new bone growth due to elastic moduli mismatch [45,46]. The interest in metallic biomaterials have expanded to biodegradable metal which exhibit the most promising properties and can be used temporarily during bone healing process [13]. In order to be well integrated with host tissue, bone scaffold is required to have characteristics such as porous, mechanical properties, and biocompatibility which are also very important for tissues regenerations [34].

The comprehensive degradation assessment systems of biodegradable metals must be carefully selected towards specific applications [47]. Biodegradable metals have an assorted degradation behaviour and mechanism contingent depending on the environments and types of measurements used [12]. Witte *et al.* have reported that the current ASTM degradation test methods for *in-vitro* test cannot be used to predict the *in-vivo* degradation rates [48]. They reported that the degradation rates of specimen with cylindrical rods in the *in-vitro* test have shown a four magnitude higher compared to the *in-vivo* test. It is crucial for the *in-vitro* test to be precisely mimicked the *in-vivo*

conditions, in order to provide more accurate information and to obtain promising implants [49]. In fact, the acceptable degradation rate of the bone scaffold should be 0.02 mm/y [19,50]. However, based on the literature findings, there was none of the studies have obtained the required degradation rate. Thus far, all studies conducted on porous biodegradable metals for potential bone scaffold applications have been done under static immersion tests only [26,51–53]. The static degradation assessment does not represent the actual boundary in human cancellous bone environment. The bone scaffolds made of biodegradable metals will be in contacted with cancellous bone and exposed to the surrounding environment once implanted [35,36]. Therefore, to address the existing gap, in this study, we had integrated a biomechanical condition of cancellous bone for testing porous magnesium specimens under a dynamic immersion condition.

### **1.3 Objectives**

The aim of this study is to analyse the degradation behaviour of porous magnesium under dynamic degradation test for bone scaffold applications. The specific objectives are:

- (i) To analyse the influences of different flow rates fluid passing through porous magnesium structure on dynamic immersion test.
- (ii) To analyse the effects of different cyclic loading of porous magnesium under a constant flow rate on dynamic immersion test.

## 1.4 Scopes

A commercially pure magnesium (Mg) rod with a diameter of 25.4 mm and 99.9% purity (Goodfellow Inc., Cambridge, UK) was used for developing porous specimens with three different percentage of porosity (30%, 41% and 55%). The porous structure was fabricated using CNC machine. The specific specimen chamber was developed for both experimental setups of fluid flow and different cyclic loading test. The specimens were cleaned internally and externally using interdental brush to remove any excess materials and chemicals and ground using abrasive paper, respectively. The dynamic immersion test rig has been built, equipped with data acquisition (record the pressure value of the fluid before and after the specimen chamber), water bath (heating the fluids medium to human body temperature) and peristaltic pump (pulsatile flow). The simulated body fluid (SBF) was used as fluid medium in this study. The dynamic immersion test was conducted for 24, 48 and 72 hours. The variation of flow rates and cyclic loading were used as the boundaries in the dynamic immersion test as there were to mimic the condition of fluid pass through cancellous bone structure and compressive strain levels of physiological activities. The universal testing machine (The FastTrack 8874, Instron, Norwood, USA) was used to perform the cyclic loading and to determine the mechanical properties of the specimen. The tested specimen was characterised using X-ray diffractometer (XRD), Scanning Electron Microscope (SEM) and Energy Dispersive Spectrometer (EDS). The weight loss measurement was used as the method to assess the degradation rates of the porous magnesium. Limitations of this study was not included the hydrogen evolution measurement as it required the hydrogen gas trapping system.



## **1.5 Significance of the Study**

This study has assessed the potential of the porous magnesium as bone scaffold using dynamic degradation under simulated environment of human cancellous bone. The implementation boundary of human cancellous bone environment in dynamic immersion integrated cyclic loading had demonstrated significant degradation behaviour and mechanical property changes of the porous magnesium compare with using static immersion test only. Hereby, through this study, the degradation assessment of biodegradable material especially metal is required to use the dynamic immersion test integrated cyclic loading. This will be very beneficial to the community because once bone scaffold implanted, the patients can perform daily routine as usual. Because the use of bone scaffold that has taken into account for load bearing purpose. Thus the more activities are carried out, it could improve the bone healing process and also the health of the patient himself. Not just that, when the process of bone healing occurs in a very good condition, then the failure of the bone scaffold that has happened in the past can be avoided so that patients no longer need a second surgery. This can reduce the costs to be incurred by the patient and the use of bone scaffold causing the patient can continue to perform the desired activity, thus reducing the time the patient gets treatment and contribute to a better living environment.

## **1.6 Thesis Structure and Organization**

Chapter 1 presents an introduction of this research which provides an overview and the needs of bone scaffolds. Background is provided on both mechanobiological of bone and degradation techniques used for biodegradable metal evaluation. Then, research aims, scopes and significance of this study are highlights. Chapter 2 is the literature review which contains reviews on bone remodelling process, the usage of biodegradable metal for bone scaffolds and concept of biodegradation. Chapter 3 explains how the dynamic degradation of porous magnesium were produced, prepared tested and analysed. The results and discussion of the study was presented in three

subsequent chapter. Chapter 4 reports the results and discussion of the effects using dynamic immersion test on degradation behaviour of porous magnesium under constant flowrates. Chapter 5 presents the results and discussion of the influences of variation flow rates towards degradation behaviour of porous magnesium in dynamic immersion test. Chapter 6 contains the results and discussion of the influenced integrating the cyclic loading on the dynamic degradation behaviour of porous magnesium under dynamic immersion test using constant flow rate. Finally, chapter 7 concludes the findings attained in this study. The limitations and recommendations also are highlight for future works.

## REFERENCES

1. V. Campana, G. Milano, E. Pagano, M. Barba, C. Cicione, G. Salonna, W. Lattanzi, G. Logroscino, Bone substitutes in orthopaedic surgery: from basic science to clinical practice, *J. Mater. Sci. Mater. Med.* (2014) 25 :2445–2461.
2. Transparency Market Research. (2016). Bone Grafts and Substitutes Market expected to reach USD 3.48 Billion Globally in 2023, website : <http://www.transparencymarketresearch.com/bone-grafts-substitutes-market.html>.
3. O. Faour, R. Dimitriou, C.A. Cousins, P. V. Giannoudis, The use of bone graft substitutes in large cancellous voids: Any specific needs?, *Injury.* (2011) 42 :S87–S90.
4. K.A. Hing, Bone repair in the twenty-first century: biology, chemistry or engineering?, *Philos. Trans. A. Math. Phys. Eng. Sci.* (2004) 362 :2821–50.
5. C.G. Finkemeier, Bone-grafting and bone-graft substitutes., *J. Bone Joint Surg. Am.* (2002) 84–A :454–64.
6. A.S. Greenwald, S.D. Boden, V.M. Goldberg, Y. Khan, C.T. Laurencin, R.N. Rosier, Bone-graft substitutes: facts, fictions, and applications., *J. Bone Joint Surg. Am.* (2001) 83–A Suppl :98–103.
7. A. Audit, The Asian Audit: Epidemiology, costs and burden of osteoporosis in Asia, *Int. Osteoporos. Found.* (2009).
8. S.K. Nandi, S. Roy, P. Mukherjee, B. Kundu, D.. De, D. Basu, Orthopaedic Applications of Bone Graft, *Indian J. Med. Res.* (2010) 132 :15–30.
9. P.X. Ma, R. Zhang, G. Xiao, R. Franceschi, Engineering new bone tissue in vitro on highly porous poly(alpha-hydroxyl acids)/hydroxyapatite composite scaffolds., *J. Biomed. Mater. Res.* (2001) 54 :284–93.

10. R. Langer, J.P. Vacanti, Tissue engineering, *Science* (80-. ). (1993) 260 :920–926.
11. V.T. Athanasiou, D.J. Papachristou, A. Panagopoulos, A. Saridis, C.D. Scopa, P. Megas, Histological comparison of autograft, allograft-DBM, xenograft, and synthetic grafts in a trabecular bone defect: an experimental study in rabbits., *Med. Sci. Monit.* (2010) 16 :BR24-R31.
12. Y.F. Zheng, X.N. Gu, F. Witte, Biodegradable metals, *Mater. Sci. Eng. R Reports.* (2014) 77 :1–34.
13. F. Witte, The history of biodegradable magnesium implants: a review., *Acta Biomater.* (2010) 6 :1680–92.
14. M.P. Staiger, A.M. Pietak, J. Huadmai, G. Dias, Magnesium and its alloys as orthopedic biomaterials: a review., *Biomaterials.* (2006) 27 :1728–34.
15. D.W. Hutmacher, Scaffolds in tissue engineering bone and cartilage., *Biomaterials.* (2000) 21 :2529–43.
16. L.S. Nair, C.T. Laurencin, Biodegradable polymers as biomaterials, *Prog. Polym. Sci.* (2007) 32 :762–798.
17. I. Vroman, L. Tighzert, Biodegradable polymers, *Materials (Basel).* (2009) 2 :307–344.
18. P.K.D. V Yarlagadda, M. Chandrasekharan, J.Y.M. Shyan, Recent advances and current developments in tissue scaffolding., *Biomed. Mater. Eng.* (2005) 15 :159–77.
19. G. Song, Control of biodegradation of biocompatible magnesium alloys, *Corros. Sci.* (2007) 49 :1696–1701.
20. X. Gu, Y. Zheng, Y. Cheng, S. Zhong, T. Xi, In vitro corrosion and biocompatibility of binary magnesium alloys, *Biomaterials.* (2009) 30 :484–498.
21. F. Witte, N. Hort, C. Vogt, S. Cohen, K.U. Kainer, R. Willumeit, F. Feyerabend, Degradable biomaterials based on magnesium corrosion, *Curr. Opin. Solid State Mater. Sci.* (2008) 12 :63–72.
22. M. Geetha, a. K. Singh, R. Asokamani, a. K. Gogia, Ti based biomaterials, the ultimate choice for orthopaedic implants - A review, *Prog. Mater. Sci.* (2009) 54 :397–425.
23. A. Bigi, G. Falini, E. Foresti, M. Gazzano, A. Ripamonti, N. Roveri, Magnesium influence on hydroxyapatite crystallization, *J. Inorg. Biochem.*

- (1993) 49 :69–78.
24. X. Zhang, X.-W. Li, J.-G. Li, X.-D. Sun, Preparation and mechanical property of a novel 3D porous magnesium scaffold for bone tissue engineering., *Mater. Sci. Eng. C. Mater. Biol. Appl.* (2014) 42 :362–7.
  25. H. Zreiqat, C.R. Howlett, A. Zannettino, P. Evans, G. Schulze-Tanzil, C. Knabe, M. Shakibaei, Mechanisms of magnesium-stimulated adhesion of osteoblastic cells to commonly used orthopaedic implants, *J. Biomed. Mater. Res.* (2002) 62 :175–184.
  26. A.H.M. Yusop, N.M. Daud, H. Nur, M.R.A. Kadir, H. Hermawan, Controlling the degradation kinetics of porous iron by poly(lactic-co-glycolic acid) infiltration for use as temporary medical implants, *Sci. Rep.* (2015) 5 :11194.
  27. A.H. Yusop, A.A. Bakir, N.A. Shaharom, M.R. Abdul Kadir, H. Hermawan, Porous biodegradable metals for hard tissue scaffolds: A review, *Int. J. Biomater.* (2012) 2012.
  28. A. Brown, S. Zaky, H. Ray, C. Sfeir, Porous magnesium/PLGA composite scaffolds for enhanced bone regeneration following tooth extraction, *Acta Biomater.* (2015) 11 :543–553.
  29. Y. Khan, Tissue Engineering of Bone: Material and Matrix Considerations, *J. Bone Jt. Surg.* (2008) 90 :36.
  30. M. Gravel, T. Gross, R. Vago, M. Tabrizian, Responses of mesenchymal stem cell to chitosan-coraline composites microstructured using coraline as gas forming agent, *Biomaterials.* (2006) 27 :1899–1906.
  31. B.J. Lawrence, S. V Madihally, Cell colonization in degradable 3D porous matrices ND ES SC CE ND ES SC RIB, (2008) :9–16.
  32. J. Rouwkema, N.C. Rivron, C.A. van Blitterswijk, Vascularization in tissue engineering, *Trends Biotechnol.* (2008) 26 :434–441.
  33. L. Polo-Corrales, M. Latorre-Esteves, J.E. Ramirez-Vick, Scaffold Design for Bone Regeneration, *J. Nanosci. Nanotechnol.* (2014) 14 :15–56.
  34. S. Bose, M. Roy, A. Bandyopadhyay, Recent advances in bone tissue engineering scaffolds, *Trends Biotechnol.* (2012) 30 :546–554.
  35. M.B. Schaffler, W.Y. Cheung, R. Majeska, O. Kennedy, Osteocytes: Master orchestrators of bone, *Calcif. Tissue Int.* (2014) 94 :5–24.
  36. T.A. Metzger, T.C. Kreipke, T.J. Vaughan, L.M. McNamara, G.L. Niebur, The In Situ Mechanics of Trabecular Bone Marrow: The Potential for

- Mechanobiological Response., *J. Biomech. Eng.* (2015) 137 :1–7.
37. E. Birmingham, G.L. Niebur, L.M. McNamara, P.E. McHugh, An Experimental and Computational Investigation of Bone Formation in Mechanically Loaded Trabecular Bone Explants, *Ann. Biomed. Eng.* (2015).
  38. L.M. McNamara, P.J. Prendergast, Bone remodelling algorithms incorporating both strain and microdamage stimuli., *J. Biomech.* (2007) 40 :1381–91.
  39. J. Grimm, J.L. Williams, *TECHNICAL Ok-*, (1997) 30 :743–745.
  40. F. Zhao, T.J. Vaughan, L.M. Mcnamara, Multiscale fluid-structure interaction modelling to determine the mechanical stimulation of bone cells in a tissue engineered scaffold., *Biomech. Model. Mechanobiol.* (2014).
  41. M.E. Gomes, V.I. Sikavitsas, E. Behraves, R.L. Reis, A.G. Mikos, Effect of flow perfusion on the osteogenic differentiation of bone marrow stromal cells cultured on starch-based three-dimensional scaffolds., *J. Biomed. Mater. Res. A.* (2003) 67 :87–95.
  42. E. Birmingham, T.C. Kreipke, E.B. Dolan, T.R. Coughlin, P. Owens, L.M. McNamara, G.L. Niebur, P.E. McHugh, Mechanical Stimulation of Bone Marrow In Situ Induces Bone Formation in Trabecular Explants., *Ann. Biomed. Eng.* (2014).
  43. S.P. Samuel, C.S.U.D. of C. and B. Engineering, Fluid/solid Interactions in Cancellous Bone, Cleveland State University, 2005.
  44. S. Wu, X. Liu, K.W.K. Yeung, C. Liu, X. Yang, Biomimetic porous scaffolds for bone tissue engineering, *Mater. Sci. Eng. R Reports.* (2014) 80 :1–36.
  45. J. Nagels, M. Stokdijk, P.M. Rozing, Stress shielding and bone resorption in shoulder arthroplasty, *J. Shoulder Elb. Surg.* (2003) 12 :35–39.
  46. J.J. Jacobs, N.J. Hallab, A.K. Skipor, R.M. Urban, Metal degradation products: a cause for concern in metal-metal bearings?, *Clin. Orthop. Relat. Res.* (2003) :139–47.
  47. J. Wang, C.E. Smith, J. Sankar, Y. Yun, N. Huang, Absorbable magnesium-based stent: physiological factors to consider for in vitro degradation assessments., *Regen. Biomater.* (2015) 2 :59–69.
  48. F. Witte, J. Fischer, J. Nellesen, H.-A. Crostack, V. Kaese, A. Pisch, F. Beckmann, H. Windhagen, In vitro and in vivo corrosion measurements of magnesium alloys, *Biomaterials.* (2006) 27 :1013–1018.
  49. N.T. Kirkland, J. Lespagnol, N. Birbilis, M.P. Staiger, A survey of bio-

- corrosion rates of magnesium alloys, *Corros. Sci.* (2010) 52 :287–291.
50. A. Atrens, M. Liu, N.I. Zainal Abidin, Corrosion mechanism applicable to biodegradable magnesium implants, *Mater. Sci. Eng. B.* (2011) 176 :1609–1636.
  51. X.N. Gu, W.R. Zhou, Y.F. Zheng, Y. Liu, Y.X. Li, Degradation and cytotoxicity of lotus-type porous pure magnesium as potential tissue engineering scaffold material, *Mater. Lett.* (2010) 64 :1871–1874.
  52. E. Aghion, Y. Perez, Effects of porosity on corrosion resistance of Mg alloy foam produced by powder metallurgy technology, *Mater. Charact.* (2014) 96 :78–83.
  53. M.H. Kang, T.S. Jang, S.W. Kim, H.S. Park, J. Song, H.E. Kim, K.H. Jung, H. Do Jung, MgF<sub>2</sub>-coated porous magnesium/alumina scaffolds with improved strength, corrosion resistance, and biological performance for biomedical applications, *Mater. Sci. Eng. C.* (2016) 62 :634–642.
  54. R.S. Taichman, W. Dc, R.S. Taichman, hematopoietic stem-cell niche Review article Blood and bone : two tissues whose fates are intertwined to create the hematopoietic stem-cell niche, (2011) 105 :2631–2639.
  55. M.L. Brandi, Microarchitecture, the key to bone quality, *Rheumatol. (United Kingdom)*. (2009) 48.
  56. B. Clarke, Normal Bone Anatomy and Physiology, *Clin. J. Am. Soc. Nephrol.* (2008) 3 :S131–S139.
  57. M.F. Holick, J.W. Nieves, Nutrition and bone health, *Nutr. Bone Heal.* (2015) :1–718.
  58. I. Fogelman, H. Van Der Wall, G. Gnanasegaran, Radionuclide and hybrid bone imaging, *Radionucl. Hybrid Bone Imaging.* (2012) 9783642024 :1–1046.
  59. M.J. Seibel, Biochemical markers of bone turnover: Part I: biochemistry and variability., *Clin. Biochem. Rev.* (2005) 26 :97–122.
  60. K. Irie, S. Ejiri, Y. Sakakura, T. Shibui, T. Yajima, Matrix mineralization as a trigger for osteocyte maturation., *J. Histochem. Cytochem.* (2008) 56 :561–7.
  61. T.A. Burgers, B.O. Williams, Regulation of Wnt/ $\beta$ -catenin signaling within and from osteocytes, *Bone.* (2013) 54 :244–249.
  62. M. Morita, a. Ebihara, M. Itoman, T. Sasada, Progression of osteoporosis in cancellous bone depending on trabecular structure, *Ann. Biomed. Eng.* (1994) 22 :532–539.

63. R. Eastell, *Osteoporosis, Medicine (Baltimore)*. (2013) 41 :586–591.
64. World Health Organization (WHO), *Assessment of fracture risk and its application to screening for postmenopausal osteoporosis: Report of a WHO study group*. WHO Technical Report Series, Report No. 843., SWITZERLAND, 1994.
65. B.M. Density, *Osteoporosis Health Center Bone Mineral Density*, (2011) :1–6.
66. P. Lips, N.M. van Schoor, *Quality of life in patients with osteoporosis.*, *Osteoporos. Int.* (2005) 16 :447–455.
67. C. Dowson, R. Lewis, *Rheumatology, Rheumatology*. (2010) :289–305.
68. E. Siris, P.D. Delmas, *Assessment of 10-year absolute fracture risk: A new paradigm with worldwide application*, *Osteoporos. Int.* (2008) 19 :383–384.
69. W.D. Leslie, *Absolute fracture risk reporting in clinical practice: A physician-centered survey*, *Osteoporos. Int.* (2008) 19 :459–463.
70. X.S. Liu, G. Bevill, T.M. Keaveny, P. Sajda, X.E. Guo, *Micromechanical analyses of vertebral trabecular bone based on individual trabeculae segmentation of plates and rods*, *J. Biomech.* (2009) 42 :249–256.
71. B. Helgason, E. Perilli, E. Schileo, F. Taddei, S. Brynjólfsson, M. Viceconti, *Mathematical relationships between bone density and mechanical properties: A literature review*, *Clin. Biomech.* (2008) 23 :135–146.
72. M.A. Velasco, C.A. Narváez-tovar, D.A. Garzón-alvarado, *Design , Materials , and Mechanobiology of Biodegradable Scaffolds for Bone Tissue Engineering*, (2015) 2015.
73. L.J. Gibson, *The mechanical behaviour of cancellous bone*, *J. Biomech.* (1985) 18 :317–328.
74. N.D. Sessions, B.P. Halloran, D.D. Bikle, T.J. Wronski, C.M. Cone, E. Morey-Holton, *Bone response to normal weight bearing after a period of skeletal unloading*, *Am. J. Physiol. - Endocrinol. Metab.* (1989) 257 :E606 LP-E610.
75. E.R. Morey, D.J. Baylink, *Inhibition of bone formation during space flight*, *Science (80-. )*. (1978) 201 :1138 LP-1141.
76. Z. Zhong, O. Akkus, *Effects of age and shear rate on the rheological properties of human yellow bone marrow*, *Biorheology*. (2011) 48 :89–97.
77. P.J. Prendergast, D. Taylor, *Prediction of bone adaptation using damage accumulation*, *J. Biomech.* (1994) 27 :1067–1076.
78. H.M. Frost, *Bone mass and the mechanostat: A proposal*, *Anat. Rec.* (1987) 219



- :1–9.
79. S.P. Fritton, K.J. McLeod, C.T. Rubin, Quantifying the strain history of bone: spatial uniformity and self-similarity of low-magnitude strains., *J. Biomech.* (2000) 33 :317–25.
  80. D.B. Burr, C. Milgrom, D. Fyhrie, M. Forwood, M. Nyska, A. Finestone, S. Hoshaw, E. Saiag, A. Simkin, In vivo measurement of human tibial strains during vigorous activity., *Bone.* (1996) 18 :405–10.
  81. J.R. Mosley, Osteoporosis and bone functional adaptation: mechanobiological regulation of bone architecture in growing and adult bone, a review., *J. Rehabil. Res. Dev.* (n.d.) 37 :189–99.
  82. D.R. Carter, D.P. Fyhrie, R.T. Whalen, Trabecular bone density and loading history: regulation of connective tissue biology by mechanical energy., *J. Biomech.* (1987) 20 :785–94.
  83. E. Birmingham, J.A. Grogan, G.L. Niebur, L.M. McNamara, P.E. McHugh, Computational modelling of the mechanics of trabecular bone and marrow using fluid structure interaction techniques, *Ann. Biomed. Eng.* (2013) 41 :814–826.
  84. T.R. Coughlin, G.L. Niebur, Fluid shear stress in trabecular bone marrow due to low-magnitude high-frequency vibration, *J. Biomech.* (2012) 45 :2222–2229.
  85. T.J. Vaughan, M. Voisin, G.L. Niebur, L.M. Mcnamara, Multiscale Modelling of Trabecular Bone Marrow: Understanding the Micromechanical Environment of Mesenchymal Stem Cells during Osteoporosis., *J. Biomech. Eng.* (2014) 137 :11003.
  86. D.J. Downey, P.A. Simkin, R. Taggart, The effect of compressive loading on intraosseous pressure in the femoral head in vitro., *J. Bone Joint Surg. Am.* (1988) 70 :871–7.
  87. N. Case, B. Sen, J.A. Thomas, M. Styner, Z. Xie, C.R. Jacobs, J. Rubin, Steady and oscillatory fluid flows produce a similar osteogenic phenotype., *Calcif. Tissue Int.* (2011) 88 :189–97.
  88. E.J. Arnsdorf, P. Tummala, R.Y. Kwon, C.R. Jacobs, Mechanically induced osteogenic differentiation--the role of RhoA, ROCKII and cytoskeletal dynamics., *J. Cell Sci.* (2009) 122 :546–53.
  89. M.J. Jaasma, N.A. Plunkett, F.J. O'Brien, Design and validation of a dynamic

- flow perfusion bioreactor for use with compliant tissue engineering scaffolds, *J. Biotechnol.* (2008) 133 :490–496.
90. A.S. Goldstein, T.M. Juarez, C.D. Helmke, M.C. Gustin, A.G. Mikos, Effect of convection on osteoblastic cell growth and function in biodegradable polymer foam scaffolds, *Biomaterials.* (2001) 22 :1279–1288.
  91. D. Li, T. Tang, J. Lu, K. Dai, Effects of Flow Shear Stress and Mass Transport on the Construction of a Large-Scale Tissue-Engineered Bone in a Perfusion Bioreactor, *Tissue Eng. Part A.* (2009) 15 :2773–2783.
  92. J. You, C.E. Yellowley, H.J. Donahue, Y. Zhang, Q. Chen, C.R. Jacobs, Substrate Deformation Levels Associated With Routine Physical Activity Are Less Stimulatory to Bone Cells Relative to Loading-Induced Oscillatory Fluid Flow, *J. Biomech. Eng.* (2000) 122 :387–393.
  93. L. Tan, X. Yu, P. Wan, K. Yang, Biodegradable Materials for Bone Repairs: A Review, *J. Mater. Sci. Technol.* (2013) 29 :503–513.
  94. M. Navarro, A. Michiardi, O. Castaño, J.A. Planell, Biomaterials in orthopaedics., *J. R. Soc. Interface.* (2008) 5 :1137–58.
  95. X. Wen, P.A. Tresco, Fabrication and characterization of permeable degradable poly(dl-lactide-co-glycolide) (PLGA) hollow fiber phase inversion membranes for use as nerve tract guidance channels, *Biomaterials.* (2006) 27 :3800–3809.
  96. R.J. Narayan, The next generation of biomaterial development, *Philos Trans A Math Phys Eng Sci.* (2010) 368 :1831–1837.
  97. X. Li, L. Wang, Y. Fan, Q. Feng, F.-Z. Cui, F. Watari, Nanostructured scaffolds for bone tissue engineering, *J. Biomed. Mater. Res. Part A.* (2013) 101A :2424–2435.
  98. T. Gong, J. Xie, J. Liao, T. Zhang, S. Lin, Y. Lin, Nanomaterials and bone regeneration., *Bone Res.* (2015) 3 :15029.
  99. M. Cheng, T. Wahafu, G. Jiang, W. Liu, Y. Qiao, X. Peng, T. Cheng, X. Zhang, G. He, X. Liu, A novel open-porous magnesium scaffold with controllable microstructures and properties for bone regeneration, *Sci. Rep.* (2016) 6 :24134.
  100. G. Lewis, Properties of open-cell porous metals and alloys for orthopaedic applications, *J. Mater. Sci. Mater. Med.* (2013) 24 :2293–2325.
  101. K. Bobe, E. Willbold, I. Morgenthal, O. Andersen, T. Studnitzky, J. Nellesen, W. Tillmann, C. Vogt, K. Vano, F. Witte, In vitro and in vivo evaluation of biodegradable, open-porous scaffolds made of sintered magnesium W4 short

- fibres., *Acta Biomater.* (2013) 9 :8611–23.
102. A. Syahrom, M.R. Abdul Kadir, J. Abdullah, A. Öchsner, Permeability studies of artificial and natural cancellous bone structures., *Med. Eng. Phys.* (2013) 35 :792–9.
  103. D.A. Shimko, V.F. Shimko, E.A. Sander, K.F. Dickson, E.A. Nauman, Effect of porosity on the fluid flow characteristics and mechanical properties of tantalum scaffolds, *J. Biomed. Mater. Res. Part B Appl. Biomater.* (2005) 73B :315–324.
  104. G.A.P. Renders, L. Mulder, L.J. van Ruijven, T.M.G.J. van Eijden, Porosity of human mandibular condylar bone, *J. Anat.* (2007) 210 :239–248.
  105. B.D. Snyder, S. Piazza, W.T. Edwards, W.C. Hayes, Role of trabecular morphology in the etiology of age-related vertebral fractures, *Calcif. Tissue Int.* (1993) 53 :14–22.
  106. E.F. Morgan, T.M. Keaveny, Dependence of yield strain of human trabecular bone on anatomic site, *J. Biomech.* (2001) 34 :569–577.
  107. T.M. Keaveny, E.F. Morgan, O.C. Yeh, *Bone Mechanics*, *Stand. Handb. Biomed. Eng. Des.* (2004) :8.1-8.23.
  108. C.E. Wen, Y. Yamada, K. Shimojima, Y. Chino, T. Asahina, M. Mabuchi, Processing and mechanical properties of autogenous titanium implant materials, *J. Mater. Sci. Mater. Med.* (2002) 13 :397–401.
  109. S. Tarafder, V.K. Balla, N.M. Davies, A. Bandyopadhyay, S. Bose, Microwave-sintered 3D printed tricalcium phosphate scaffolds for bone tissue engineering, *J. Tissue Eng. Regen. Med.* (2013) 7 :631–641.
  110. W. Xue, A. Bandyopadhyay, S. Bose, Polycaprolactone coated porous tricalcium phosphate scaffolds for controlled release of protein for tissue engineering, *J. Biomed. Mater. Res. Part B Appl. Biomater.* (2009) 91B :831–838.
  111. K. Zhang, Y. Wang, M.A. Hillmyer, L.F. Francis, Processing and properties of porous poly(l-lactide)/bioactive glass composites, *Biomaterials.* (2004) 25 :2489–2500.
  112. H.R. Ramay, M. Zhang, Preparation of porous hydroxyapatite scaffolds by combination of the gel-casting and polymer sponge methods, *Biomaterials.* (2003) 24 :3293–3302.
  113. Z.S. Seyedraoufi, S. Mirdamadi, Synthesis, microstructure and mechanical

- properties of porous Mg--Zn scaffolds., *J. Mech. Behav. Biomed. Mater.* (2013) 21 :1–8.
114. C.E. Wen, Y. Yamada, K. Shimojima, Y. Chino, H. Hosokawa, M. Mabuchi, Compressibility of porous magnesium foam: dependency on porosity and pore size, *Mater. Lett.* (2004) 58 :357–360.
  115. F. Witte, H. Ulrich, M. Rudert, E. Willbold, Biodegradable magnesium scaffolds: Part 1: appropriate inflammatory response., *J. Biomed. Mater. Res. A.* (2007) 81 :748–56.
  116. F. Witte, H. Ulrich, C. Palm, E. Willbold, Biodegradable magnesium scaffolds: Part II: peri-implant bone remodeling., *J. Biomed. Mater. Res. A.* (2007) 81 :757–65.
  117. F. Geng, L. Tan, B. Zhang, C. Wu, Y. He, J. Yang, K. Yang, Study on beta-TCP Coated Porous Mg as a Bone Tissue Engineering Scaffold Material, *J. Mater. Sci. Technol.* (2009) 25 :123–129.
  118. G. Jiang, G. He, A new approach to the fabrication of porous magnesium with well-controlled 3D pore structure for orthopedic applications, *Mater. Sci. Eng. C.* (2014) 43 :317–320.
  119. B. Guang, L. Song, A. Atrens, Corrosion Mechanisms of Magnesium Alloys \*\*, (1999) 2648 :10–33.
  120. J. Vormann, Magnesium: nutrition and metabolism., *Mol. Aspects Med.* (2003) 24 :27–37.
  121. R. Zeng, W. Dietzel, F. Witte, N. Hort, C. Blawert, Progress and Challenge for Magnesium Alloys as Biomaterials, *Adv. Eng. Mater.* (2008) 10 :B3–B14.
  122. G.C. Clark, D.F. Williams, The effects of proteins on metallic corrosion., *J. Biomed. Mater. Res.* (1982) 16 :125–34.
  123. G. Song, a. Atrens, Understanding Magnesium Corrosion—A Framework for Improved Alloy Performance, *Adv. Eng. Mater.* (2003) 5 :837–858.
  124. Z. Shi, G. Song, A. Atrens, Corrosion resistance of anodised single-phase Mg alloys, *Surf. Coatings Technol.* (2006) 201 :492–503.
  125. M. Liu, P. Schmutz, P.J. Uggowitzer, G. Song, A. Atrens, The influence of yttrium (Y) on the corrosion of Mg-Y binary alloys, *Corros. Sci.* (2010) 52 :3687–3701.
  126. Z. Shi, A. Atrens, An innovative specimen configuration for the study of Mg corrosion, *Corros. Sci.* (2011) 53 :226–246.

127. F. Cao, Z. Shi, J. Hofstetter, P.J. Uggowitzer, G. Song, M. Liu, A. Atrens, Corrosion of ultra-high-purity Mg in 3.5% NaCl solution saturated with Mg(OH)<sub>2</sub>, *Corros. Sci.* (2013) 75 :78–99.
128. A. Atrens, G.-L. Song, M. Liu, Z. Shi, F. Cao, M.S. Dargusch, Review of Recent Developments in the Field of Magnesium Corrosion, *Adv. Eng. Mater.* (2015) 17 :400–453.
129. N.T. Kirkland, N. Birbilis, M.P. Staiger, Assessing the corrosion of biodegradable magnesium implants: A critical review of current methodologies and their limitations, *Acta Biomater.* (2012) 8 :925–936.
130. M.C. Zhao, P. Schmutz, S. Brunner, M. Liu, G. ling Song, A. Atrens, An exploratory study of the corrosion of Mg alloys during interrupted salt spray testing, *Corros. Sci.* (2009) 51 :1277–1292.
131. M. Liu, P.J. Uggowitzer, A. V. Nagasekhar, P. Schmutz, M. Easton, G.L. Song, A. Atrens, Calculated phase diagrams and the corrosion of die-cast Mg-Al alloys, *Corros. Sci.* (2009) 51 :602–619.
132. A.H. Martinez Sanchez, B.J.C. Luthringer, F. Feyerabend, R. Willumeit, Mg and Mg alloys: how comparable are in vitro and in vivo corrosion rates? A review., *Acta Biomater.* (2015) 13 :16–31.
133. Y. Wang, M. Wei, J. Gao, J. Hu, Y. Zhang, Corrosion process of pure magnesium in simulated body fluid, *Mater. Lett.* (2008) 62 :2181–2184.
134. N.I. Zainal Abidin, B. Rolfe, H. Owen, J. Malisano, D. Martin, J. Hofstetter, P.J. Uggowitzer, A. Atrens, The in vivo and in vitro corrosion of high-purity magnesium and magnesium alloys WZ21 and AZ91, *Corros. Sci.* (2013) 75 :354–366.
135. S. Johnston, Z. Shi, A. Atrens, The influence of pH on the corrosion rate of high-purity Mg, AZ91 and ZE41 in bicarbonate buffered Hanks' solution, *Corros. Sci.* (2015) 101 :182–192.
136. K.-W. Lee, S. Wang, L. Lu, E. Jabbari, B.L. Currier, M.J. Yaszemski, Fabrication and characterization of poly(propylene fumarate) scaffolds with controlled pore structures using 3-dimensional printing and injection molding., *Tissue Eng.* (2006) 12 :2801–11.
137. T. Kokubo, H. Takadama, How useful is SBF in predicting in vivo bone bioactivity?, *Biomaterials.* (2006) 27 :2907–2915.
138. J. Lévesque, H. Hermawan, D. Dubé, D. Mantovani, Design of a pseudo-

- physiological test bench specific to the development of biodegradable metallic biomaterials, *Acta Biomater.* (2008) 4 :284–295.
139. M.D.M. Innocentini, R.K. Faleiros, R. Pisani, I. Thijs, J. Luyten, S. Mullens, Permeability of porous gelcast scaffolds for bone tissue engineering, *J. Porous Mater.* (2010) 17 :615–627.
  140. R.S. Ochia, R.P. Ching, Hydraulic Resistance and Permeability in Human Lumbar Vertebral Bodies, *J. Biomech. Eng.* (2002) 124 :533.
  141. M.E. Lynch, C. Fischbach, Biomechanical forces in the skeleton and their relevance to bone metastasis: Biology and engineering considerations, *Adv. Drug Deliv. Rev.* (2014) 79 :119–134.
  142. R.W. Revie, *Corrosion and Corrosion Control*, John Wiley & Sons, 2008.
  143. H. Wang, Z. Shi, In vitro biodegradation behavior of magnesium and magnesium alloy, *J. Biomed. Mater. Res. - Part B Appl. Biomater.* (2011) 98 B :203–209.
  144. Z. Qiao, Z. Shi, N. Hort, N.I. Zainal Abidin, A. Atrens, Corrosion behaviour of a nominally high purity Mg ingot produced by permanent mould direct chill casting, *Corros. Sci.* (2012) 61 :185–207.
  145. N.I. Zainal Abidin, A.D. Atrens, D. Martin, A. Atrens, Corrosion of high purity Mg, Mg<sub>2</sub>Zn<sub>0.2</sub>Mn, ZE41 and AZ91 in Hank's solution at 37°C, *Corros. Sci.* (2011) 53 :3542–3556.
  146. D. Tie, F. Feyerabend, N. Hort, R. Willumeit, D. Hoeche, XPS studies of magnesium surfaces after exposure to Dulbecco's modified eagle medium, Hank's buffered salt solution, and simulated body fluid, *Adv. Eng. Mater.* (2010) 12 :699–704.
  147. C. Liu, Y. Xin, G. Tang, P.K. Chu, Influence of heat treatment on degradation behavior of bio-degradable die-cast AZ63 magnesium alloy in simulated body fluid, *Mater. Sci. Eng. A.* (2007) 456 :350–357.
  148. R. Willumeit, J. Fischer, F. Feyerabend, N. Hort, U. Bismayer, S. Heidrich, B. Mihailova, Chemical surface alteration of biodegradable magnesium exposed to corrosion media, *Acta Biomater.* (2011) 7 :2704–2715.
  149. D. Ahmadkhaniha, M. Fedel, M. Heydarzadeh Sohi, A. Zarei Hanzaki, F. Deflorian, Corrosion behavior of magnesium and magnesium-hydroxyapatite composite fabricated by friction stir processing in Dulbecco's phosphate buffered saline, *Corros. Sci.* (2016) 104 :319–329.

150. F. Cao, Z. Shi, G.L. Song, M. Liu, A. Atrens, Corrosion behaviour in salt spray and in 3.5% NaCl solution saturated with Mg(OH)<sub>2</sub> of as-cast and solution heat-treated binary Mg-X alloys: X=Mn, Sn, Ca, Zn, Al, Zr, Si, Sr, *Corros. Sci.* (2013) 76 :60–97.
151. T. Kobayashi, S. Ono, S. Hirakura, Y. Oaki, H. Imai, Morphological variation of hydroxyapatite grown in aqueous solution based on simulated body fluid, *CrystEngComm.* (2012) 14 :1143.
152. W. Ma, Y. Liu, W. Wang, Y. Zhang, Effects of electrolyte component in simulated body fluid on the corrosion behavior and mechanical integrity of magnesium, *Corros. Sci.* (2015) 98 :201–210.
153. F. Ren, Y. Ding, X. Ge, X. Lu, K. Wang, Y. Leng, Growth of one-dimensional single-crystalline hydroxyapatite nanorods, *J. Cryst. Growth.* (2012) 349 :75–82.
154. Z. Li, G.-L. Song, S. Song, Effect of bicarbonate on biodegradation behaviour of pure magnesium in a simulated body fluid, *Electrochim. Acta.* (2014) 115 :56–65.
155. Y. Xin, T. Hu, P.K. Chu, Degradation behaviour of pure magnesium in simulated body fluids with different concentrations of HCO<sub>3</sub><sup>-</sup>, *Corros. Sci.* (2011) 53 :1522–1528.
156. X.B. Chen, N. Birbilis, T.B. Abbott, Effect of [Ca<sup>2+</sup>] and [] levels on the formation of calcium phosphate conversion coatings on die-cast magnesium alloy AZ91D, *Corros. Sci.* (2012) 55 :226–232.
157. M. Tomozawa, S. Hiromoto, Growth mechanism of hydroxyapatite-coatings formed on pure magnesium and corrosion behavior of the coated magnesium, *Appl. Surf. Sci.* (2011) 257 :8253–8257.
158. Y. Lu, P. Wan, B. Zhang, L. Tan, K. Yang, J. Lin, Research on the corrosion resistance and formation of double-layer calcium phosphate coating on AZ31 obtained at varied temperatures., *Mater. Sci. Eng. C. Mater. Biol. Appl.* (2014) 43 :264–71.
159. X.B. Chen, N. Birbilis, T.B. Abbott, A simple route towards a hydroxyapatite–Mg(OH)<sub>2</sub> conversion coating for magnesium, *Corros. Sci.* (2011) 53 :2263–2268.
160. S. Hiromoto, Self-healing property of hydroxyapatite and octacalcium phosphate coatings on pure magnesium and magnesium alloy, *Corros. Sci.*

- (2015) 100 :284–294.
161. S. V. Dorozhkin, Calcium orthophosphates: occurrence, properties, biomineralization, pathological calcification and biomimetic applications., (2011) :121–164.
  162. F. Witte, V. Kaese, H. Haferkamp, E. Switzer, a. Meyer-Lindenberg, C.J. Wirth, H. Windhagen, In vivo corrosion of four magnesium alloys and the associated bone response, *Biomaterials*. (2005) 26 :3557–3563.
  163. X.L. Ma, L.H. Dong, X. Wang, Microstructure, mechanical property and corrosion behavior of co-continuous  $\beta$ -TCP/MgCa composite manufactured by suction casting, *Mater. Des.* (2014) 56 :305–312.
  164. X.N. Gu, W.R. Zhou, Y.F. Zheng, Y. Cheng, S.C. Wei, S.P. Zhong, T.F. Xi, L.J. Chen, Corrosion fatigue behaviors of two biomedical Mg alloys - AZ91D and WE43 - In simulated body fluid., *Acta Biomater.* (2010) 6 :4605–13.
  165. J.M. Seitz, R. Eifler, J. Stahl, M. Kietzmann, F.W. Bach, Characterization of MgNd<sub>2</sub> alloy for potential applications in bioresorbable implantable devices, *Acta Biomater.* (2012) 8 :3852–3864.
  166. X. Liu, J. Sun, Y. Yang, F. Zhou, Z. Pu, L. Li, Y. Zheng, Microstructure, mechanical properties, in vitro degradation behavior and hemocompatibility of novel Zn-Mg-Sr alloys as biodegradable metals, *Mater. Lett.* (2016) 162 :242–245.
  167. M. Thomann, C. Krause, D. Bormann, N. Von Der Höh, H. Windhagen, A. Meyer-Lindenberg, Comparison of the resorbable magnesium alloys LAE442 und MgCa0.8 concerning their mechanical properties, their progress of degradation and the bone-implant-contact after 12 months implantation duration in a rabbit model, *Materwiss. Werksttech.* (2009) 40 :82–87.
  168. W.D. Mueller, M. Lucia Nascimento, M.F. Lorenzo De Mele, Critical discussion of the results from different corrosion studies of Mg and Mg alloys for biomaterial applications, *Acta Biomater.* (2010) 6 :1749–1755.
  169. S. V. Dorozhkin, Calcium orthophosphate coatings on magnesium and its biodegradable alloys, *Acta Biomater.* (2014) 10 :2919–2934.
  170. S. Shadanbaz, G.J. Dias, Calcium phosphate coatings on magnesium alloys for biomedical applications: A review, *Acta Biomater.* (2012) 8 :20–30.
  171. P. Tian, X. Liu, Surface modification of biodegradable magnesium and its alloys for biomedical applications, *Regen. Biomater.* (2015) 2 :135–151.



172. G. Bergmann, F. Graichen, A. Rohlmann, Hip joint loading during walking and running, measured in two patients, *J. Biomech.* (1993) 26 :969–990.
173. W.R. Taylor, M.O. Heller, G. Bergmann, G.N. Duda, Tibio-femoral loading during human gait and stair climbing, *J. Orthop. Res.* (2004) 22 :625–632.
174. S. Weinbaum, S.C. Cowin, Y. Zeng, A model for the excitation of osteocytes by mechanical loading-induced bone fluid shear stresses, *J. Biomech.* (1994) 27 :339–360.
175. A.P. Md. Saad, N. Jasmawati, M.N. Harun, M.R. Abdul Kadir, H. Nur, H. Hermawan, A. Syahrom, Dynamic degradation of porous magnesium under a simulated environment of human cancellous bone, *Corros. Sci.* (2016) 112 :1–12.
176. H.J. Donahue, Y. Zhang, Q. Chen, C.R. Jacobs, Associated With Routine Physical Activity Are Less Stimulatory to Bone Cells Relative to, (2013).
177. J. Wang, Y. Jang, G. Wan, V. Giridharan, G.L. Song, Z. Xu, Y. Koo, P. Qi, J. Sankar, N. Huang, Y. Yun, Flow-induced corrosion of absorbable magnesium alloy: In-situ and real-time electrochemical study, *Corros. Sci.* (2016) 104 :277–289.
178. S. Hiromoto, A. Yamamoto, N. Maruyama, H. Somekawa, T. Mukai, Influence of pH and flow on the polarisation behaviour of pure magnesium in borate buffer solutions, *Corros. Sci.* (2008) 50 :3561–3568.
179. M.C. Zhao, M. Liu, G.L. Song, A. Atrens, Influence of pH and chloride ion concentration on the corrosion of Mg alloy ZE41, *Corros. Sci.* (2008) 50 :3168–3178.
180. R.W. Revie, H.H. Uhlig, Thermodynamics: Pourbaix Diagrams, in: *Corros. Corros. Control*, John Wiley & Sons, Inc., Hoboken, NJ, USA, 2008: pp. 43–51.
181. J. Wang, V. Giridharan, V. Shanov, Z. Xu, B. Collins, L. White, Y. Jang, J. Sankar, N. Huang, Y. Yun, Flow-induced corrosion behavior of absorbable magnesium-based stents, *Acta Biomater.* (2014) 10 :5213–5223.
182. X. Chen, G. Cao, A. Han, V.K. Punyamurtula, L. Liu, P.J. Culligan, T. Kim, Y. Qiao, Nanoscale Fluid Transport: Size and Rate Effects, *Nano Lett.* (2008) 8 :2988–2992.
183. J.N. Israelachvili, Special Interactions: Hydrogen-Bonding and Hydrophobic and Hydrophilic Interactions, *Intermol. Surf. Forces.* (2011) 4 :151–167.

184. X. Zhang, X. Li, J. Li, X. Sun, Processing, microstructure and mechanical properties of biomedical magnesium with a specific two-layer structure, *Prog. Nat. Sci. Mater. Int.* (2013) 23 :183–189.
185. C. Chen, Z. Huang, W. Yuan, J. Li, X. Cheng, R. Chi, Pressure effecting on morphology of hydroxyapatite crystals in homogeneous system, *CrystEngComm.* (2011) 13 :1632.
186. L.F. Bonewald, Osteocytes as dynamic multifunctional cells, *Ann. N. Y. Acad. Sci.* (2007) 1116 :281–290.
187. T. Adachi, Y. Aonuma, S. ichi Ito, M. Tanaka, M. Hojo, T. Takano-Yamamoto, H. Kamioka, Osteocyte calcium signaling response to bone matrix deformation, *J. Biomech.* (2009) 42 :2507–2512.
188. T.A. Metzger, S.A. Schwaner, A.J. LaNeve, T.C. Kreipke, G.L. Niebur, Pressure and shear stress in trabecular bone marrow during whole bone loading, *J. Biomech.* (2015) 48 :3035–3043.
189. S.W. Verbruggen, T.J. Vaughan, L.M. McNamara, Fluid flow in the osteocyte mechanical environment: A fluid-structure interaction approach, *Biomech. Model. Mechanobiol.* (2014) 13 :85–97.
190. Y. Xiong, Q. Yu, Y. Jiang, An experimental study of cyclic plastic deformation of extruded ZK60 magnesium alloy under uniaxial loading at room temperature, *Int. J. Plast.* (2014) 53 :107–124.
191. M. Jamesh, S. Kumar, T.S.N. Sankara Narayanan, Corrosion behavior of commercially pure Mg and ZM21 Mg alloy in Ringer's solution – Long term evaluation by EIS, *Corros. Sci.* (2011) 53 :645–654.
192. F. Yoshida, T. Uemori, K. Fujiwara, Elastic-plastic behavior of steel sheets under in-plane cyclic tension-compression at large strain, *Int. J. Plast.* (2002) 18 :633–659.
193. S.M. Haddock, O.C. Yeh, P. V. Mummaneni, W.S. Rosenberg, T.M. Keaveny, Similarity in the fatigue behavior of trabecular bone across site and species, *J. Biomech.* (2004) 37 :181–187.
194. S.M. Bowman, X.E. Guo, D.W. Cheng, T.M. Keaveny, L.J. Gibson, W.C. Hayes, T. a McMahon, Creep contributes to the fatigue behavior of bovine trabecular bone., *J. Biomech. Eng.* (1998) 120 :647–654.
195. D. Taylor, Fatigue of bone and bones: An analysis based on stressed volume, *J. Orthop. Res.* (1998) 16 :163–169.