Do physiological limits of human palpation prevent clinicians from appreciating differences in spinal stiffness?

Gregory Neil Kawchuk[1]*, Stephen Miazga[1], Isabelle Pagé[1,2], Michael Swain[3,4], Diana De Carvalho[5], Martha Funabashi[1], Alexander Breen[6], Arnold YL Wong[7]

[1] Department of Physical Therapy, University of Alberta, Edmonton, AB, Canada

[2] Department of Anatomy, Université du Québec à Trois-Rivières, Trois-Rivières, QC, Canada

[3] Musculoskeletal Health Sydney, School of Public Health, Sydney Medical School, University of Sydney, Australia.

[4] Department of Chiropractic, Faculty of Science and Engineering, Macquarie University, Australia

[5] Faculty of Medicine, Memorial University of Newfoundland, St. John's, NL, Canada

[6] Institute for Musculoskeletal Research and Clinical Implementation, Anglo-European College of Chiropractic. Bournemouth, UK

[7] Department of Rehabilitation Sciences, The Hong Kong Polytechnic University, Hong Kong SAR, China

*Corresponding author

E-mail: greg.kawchuk@ualberta.ca (GNK)

Abstract

Objective: To determine if the physiological limits of human palpation influence traditional examination procedures.

Methods: In this study, the threshold at which a change in spinal stiffness was detected was quantified in 12 experienced clinicians by changing the differential stiffness in two inflatable targets until the clinician could no longer identify which was stiffer. In the second part of the study, clinicians were then asked to palpate pre-identified pairs of vertebrae in an asymptomatic volunteer and to identify the stiffer of the pair (T7&L3, T7&L4, L3&L4) while the biomechanical stiffness of each vertebral pair was quantified objectively by a validated instrument.

Results: The mean stiffness detection threshold for the clinicians was 8%. Objective measurement of the stiffness differential between vertebral pairs was 30% for T7* & L3 and 20% for T7* & L4 and 10% for L3* & L4 (*denotes the stiffer of the pair). Ten of 12 clinicians correctly identified T7 as more stiff when compared to L3 and T7 as more stiff than L4. Alternatively, when the differential vertebral pair stiffness was similar to the stiffness detection threshold (~8%), clinicians were less successful in identifying the stiffer vertebra of the pair; 4/12 clinicians correctly identified L3 as being more stiff compared to L4.

Conclusion: These results suggest that the physiological limits of human palpation may limit the ability of clinicians to identify small alterations in spine stiffness.

Introduction

Low back pain (LBP) is a very common condition with an estimated lifetime prevalence as high as 84% worldwide.[1,2] Characterized by recurrent or intermittent episodes with a third of cases becoming chronic,[3,4] LBP is the leading cause of global disability.[5] Presently, the cause of LBP is notoriously difficult to identify in any given person, often resulting in inefficient approaches toward prevention, diagnosis and treatment. Consequently, LBP is responsible for substantial societal burdens in terms of morbidity, disability and cost.[6–8]

Despite these challenges, a contemporary theoretical framework exists with which to contextualize back pain with the aim of reducing these burdens. This framework, the biopsychosocial model, comprises three domains (biological, psychological and social).[9,10] While there are many well-developed tools that measure self-reported psychological and social domains of LBP, such as the Start Back Screening Tool,[10] the Tampa Scale for Kinesiophobia[11] and the Fear-Avoidance Beliefs Questionnaire[12], the same cannot be said for tools that measure physical aspects of the biological construct. In the same way that the absence of objective tests of mechanical heart function would negatively impact cardiac care, the absence of objective physical measures that quantify clinically meaningful mechanical function could limit, or possibly bias, our current understanding of LBP within the biopsychosocial model.

Toward this, our group has recently demonstrated that in people with LBP, those who report clinical improvement following conservative care, also experience immediate and significant changes in three objective biomechanical measures: spinal stiffness, dynamic paraspinal muscle thickness and disc perfusion.[13] Those who report no improvement with the same

intervention do not experience these biomechanical changes. These findings suggest that there may be specific phenotypes of LBP.

This observation raises the question of whether clinicians would have the ability to detect stiffness changes of this magnitude using traditional examination procedures such as manual palpation. Although palpation remains a widely taught skill to evaluate spinal stiffness and guide conservative treatment plans[14,15] previous research suggests that palpation has poor intra- and inter-rater reliability.[16–18] In addition, palpation is a sense that has not yet benefited from technology in the same way that the stethoscope has improved auscultation or optics have improved vision. Therefore, there is a possibility that technological advances that improve the human sense of palpation may outperform traditional techniques of manual palpation.

Given this small, but significant magnitude of change in spinal stiffness quantified in our previous research,[13] we asked the following question: "Do physiologic limitations in the ability of human touch prevent clinicians from detecting clinically important changes in tissue stiffness?" The objectives of the current study were to quantify the threshold at which clinicians can detect a difference in spinal stiffness via palpation, and then determine if this detection threshold would impact a clinician's ability to identify changes in spinal stiffness as measured by an objective instrument. As such, our hypothesis was that a clinician's ability to identify the stiffer of two vertebrae through manual palpation would decrease as the relative stiffness between the vertebrae approached the clinician's physiological limit of stiffness detection.

Materials and methods

Participants

Clinicians from the   were recruited for this controlled

laboratory study. Specifically, physical therapists from the   region were recruited

directly through email using a list provided by Physiotherapy Alberta College + Association, and

chiropractors were recruited by email through the   Chiropractic Society. No eligible

clinicians were refused from participating. This study received ethics approval from the

University of Alberta Human Research Ethics Review Board (Protocol number Pro00041451)

and all participants signed an informed consent prior to testing.

Detection of stiffness threshold

To determine the detection threshold of palpated stiffness, two inflatable palpation targets were

used. Stiffness of these targets could be controlled remotely by two independent hand pumps

with pressure relief valves (Fig 1). The same investigator operated this equipment throughout

the trial to achieve the desired pressure in each target by monitoring digital pressure readouts.

At all times, the investigator's actions and the digital readouts were blocked from the

participants view (Fig 1). The differential stiffness of these targets was altered using the

staircase methodology. A detailed description of the staircase method can be found

elsewhere.[19] Briefly, this method is an adaptive psychometric technique that can be used to

determine sensation thresholds perceived by participants by presenting a series of stimuli in a

pattern of varying intensities ranging from large (obvious) to small.[19] Digital pressure gauges

were used to record the pressure in each bladder before the clinician began testing any specific

combination of pressures. The pressure differential was then reduced in a step-like manner until

it was too small for the participant to judge without making a mistake. At this point, the pattern

was reversed and intensities increased again to the last identifiable level and then decreased

again from that point. This pattern was repeated until the minimally detectable threshold was

identified. All clinicians were given standardized instruction at the start of the study. Specifically,

they were instructed to apply posteroanterior forces vertically to the inflatable targets and the spine during stiffness assessments. The clinicians wore opaque goggles to block their vision during all palpation tasks and they were instructed to evaluate the spinal stiffness only (Fig 1). Clinicians were instructed to palpate the targets using the pisiform area of their palm placed overtop of a rigid metal disc placed in the centre of the inflatable target. The disc was of the same diameter of the instrument used to measure spinal stiffness (below).

Fig 1: Participant conducting the staircase protocol with an examiner recording the pressure from two inflatable bags connected to digital pressure meters

Vertebral pair testing

Clinicians were then asked to palpate pre-identified pairs of vertebrae in prone asymptomatic volunteers and identify the stiffer of the pair (T7&L3, T7&L4, L3&L4). These locations were marked in advance by an expert clinician using ultrasonography[20] (Fig 2) and did not change over the course of the study. Briefly, this assessment involves the application of a manual posteroanterior force to a specific body landmark (e.g. spinous process) or to a spinal region in general (e.g. lumbar) by a clinician and then the clinician perceives the corresponding spinal movement or stiffness.[21] As with testing on the inflatable targets, palpation consisted of the clinician using the pisiform area of their palm to apply direct pressure to the spinous process of pre-identified vertebra within a time limit of five minutes or less. This was done while the human volunteer was asked to fully exhale without exertion. While there are many spinal motion palpation techniques, spinal stiffness assessment performed in this way is a widely used skill among manual therapy professionals.[22] Although we did not time the clinician's palpation of the subject, each took approximately less than 10 seconds in making their assessment.

Fig 2: Skin markings to identify vertebral pair targets at the T7, L3 and L4 levels. Marks are to identify the vertebral segment without identifying the spinous process itself.

Following palpation by the clinician, the biomechanical stiffness of each vertebral pair in the asymptomatic volunteer was quantified with a validated instrument (Fig 3). This device has demonstrated excellent within-day reliability and has been used in multiple studies to quantify spinal stiffness in people with and without LBP.[13,23–25] In brief, the device consists of a computer-controlled indentation probe that can be moved vertically at a predetermined speed of 2 mm/s. The probe is attached in-series with a compression-tension load cell transducer (Entran, Fairfield, NJ, USA) and a rotary encoder (Dual Motion Motor, Haydon Kerk Motion Solutions Inc., Waterbury, CT, USA) to measure indentation forces and the corresponding displacements, respectively. These measures are obtained from placing the probe on the skin surface above the spinous process of the lumbar vertebrae of interest while the subject is asked to exhale completely as in the manual palpation procedure. A customized programming language (Labview, National Instruments Inc., Austin, TX, USA) was used to control data collection from the above sensors at 200 Hz. The indentation process could be stopped at any time via hardware or software emergency switches.

The determination of a correct palpation (yes/no) was made when the clinician's estimate of which vertebra is stiffer agreed (or disagreed) with the results obtained from the indentation instrument.

Fig 3: Indentation equipment used to obtain spinal stiffness (photograph taken without skin markings).

Data processing and analysis

The stiffness detection threshold was calculated as the average minimal difference between stiffness values obtained by palpating the inflatable objects that were correctly detected by the participants and then expressed as a percentage of the maximal detectable difference (i.e. Weber fraction).[26] For the vertebral pair testing, stiffness (N/mm) was calculated as the maximally applied force (60 N) divided by the resulting displacement of the blunt indentation probe (mm). The percentage difference was calculated by considering the highest stiffness of the pair as 100%.

Results

Although 15 clinicians were recruited, three did not attend the testing session. Therefore, data from 12 clinicians were included in the analysis.

Table 1 presents data from the staircase stiffness threshold detection as well as from the vertebral pair testing. The staircase test revealed that the mean stiffness detection threshold for all participants was 8% (± 3%). Objective measurement of the stiffness differential between vertebral pairs was 30% for the T7*- L3 pair, 20% for the T7*- L4 pair and 10% for the L3* - L4 pair (*demotes the stiffer of the pair). While T7 was the stiffer vertebra when compared to L3 and L4, L3 was stiffer compared to L4 vertebra.

Table 1: Threshold detection results from manual palpation of vertebral pairings. The determination of correct (yes/no) is made when the clinician's estimate of which vertebra is stiffer agrees (or disagrees) with the results obtained from the indentation instrument.

| | Staircase Results | Vertebral Stiffness Testing | | | | | |
Subject	Threshold (%)	T7/L3 difference (%)	Correct?	T7/L4 difference (%)	Correct?	L3/L4 difference (%)	Correct?
1	5.2	24.3	Yes	15.1	No	9.3	Yes
2	7.5	24.3	Yes	15.1	Yes	9.3	Yes
3	5.0	31.2	Yes	20.6	Yes	10.8	No
4	5.5	31.2	Yes	20.6	No	10.8	No
5	6.1	31.2	Yes	20.6	Yes	10.8	No
6	6.8	31.2	No	20.6	Yes	10.8	No
7	7.6	31.2	No	20.6	Yes	10.8	Yes
8	8.0	31.2	Yes	20.6	Yes	10.8	No
9	9.1	31.2	Yes	20.6	Yes	10.8	No
10	9.5	31.2	Yes	20.6	Yes	10.8	No
11	11.2	31.2	Yes	20.6	Yes	10.8	Yes
12	13.1	31.2	Yes	20.6	Yes	10.8	No

Mean (SD)	7.9 ± 2.5	30.1 ± 2.7	N/A	19.7 ± 2.1	N/A	10.6 ± 0.6	N/A
% correct			83.3		83.3		33.3

For the vertebral pair testing, when the stiffness difference within each vertebral pair exceeded the detection threshold of the clinician (~8%), clinicians had greater success in identifying the stiffer vertebra of the pair. Specifically, 10/12 clinicians correctly identified T7 as more stiff when compared to L3. The same number of clinicians (10/12) also correctly identified T7 as more stiff than L4. Alternatively, when the differential vertebral pair stiffness was similar to the stiffness detection threshold, clinicians had less success in identifying the stiffer vertebra of the pair. In this circumstance, only 4/12 clinicians correctly identified L3 as being more stiff compared to L4. Of note, the stiffness difference between L3 and L4 (~10%) observed in the current study was similar to the value observed in prior studies whose subjects reported improvement following spinal manipulation (~10%).[13]

Discussion

This study found that clinicians are able to accurately detect large stiffness differences by using manual palpation, but were less able to accurately detect small differences in stiffness. Although previous studies have investigated both palpation and spinal stiffness separately, this is the first study to combine both in terms of a known threshold of detectable stiffness change.

Interestingly, when clinicians palpated the vertebral pair with the smallest stiffness differential as measured by instrumentation (L3 and L4), their responses were not distributed randomly

between the two (i.e. 50% selecting L3 and 50% selecting L4). Further, because stiffness detection thresholds were determined individually, there were clinicians whose threshold were above and below the average value of 8%. These data suggest that when clinicians reach the limits of their ability to provide a confident answer regarding differential stiffness, other factors not measured here may come into play (e.g. guessing, ineffective incorporation of other sense on other senses).

Related studies

Multiple studies and reviews have accentuated poor intra- and inter-rater reliability of spinal motion palpation when assessing spinal stiffness manually.[16,27–30] This low reliability may be attributed to the fact that many factors can affect spinal stiffness perceived by clinicians[21] including vision[31], technique[31], intent[32] and training.[33] Although manual PA spinal stiffness assessment has face validity in assessing spinal biomechanical changes following manual therapy [34], the sensitivity of clinicians in detecting differences/changes in spinal stiffness through manual palpation techniques has been unclear. While prior estimates suggest that clinicians can discriminate differences in stiffness as low as 11%,[31,35] this threshold does nothing to suggest the magnitude of stiffness changes in pathological tissues which in theory, may occur below or above the detection threshold of clinicians.

Previously, Koppenhaver et al.[36] demonstrated that estimates of spinal stiffness obtained by manual palpation were not correlated to measures obtained from an instrument. Specifically, their study compared manual stiffness assessments from the spinous processes of L1-L5 to a single measurement of stiffness at L3. One strength of the current study is that manual and instrumented assessments of stiffness were always taken from the same locations. In addition, the current study created a range of stiffness gradients which demonstrated that clinicians can

indeed discriminate changes in stiffness, but this discrimination is modulated by the magnitude of the change.

Clinical advances as a result of assistive technologies

Significant advances in our understanding of health conditions and their treatment occur when new technology augments a basic sense. From a rolled up tube of paper creating the first stethoscope, to modern day microphones, improving a clinician's sense of hearing has revealed new pathologies, improved diagnosis and created more timely treatment.[37] Similar advances have also occurred by augmenting vision with loupes and microscopes.[38]

In contrast to hearing and sight, the sense of touch remains relatively unassisted by technology. This is likely the result of being unable to increase the sense of touch directly as would be the case for amplifying auditory signals in the ear or magnifying images provided to the eye. Therefore, sensory augmentation must be achieved in a different way. Technologies to evaluate aspects of touch that cannot be enhanced directly within the human experience do exist. An example of this would be thermometers which work not by improving the ability of humans to perceive temperature, but by providing a surrogate technology with the capability of visually displaying changes in temperature that are not perceptible with human touch.

In addition to temperature, humans can also feel changes in texture such as a nano-size wrinkles on a smooth surface.[39] This is an extension of detecting changes in an object's stiffness, a regular human experience. Clinically, changes in stiffness can be linked to pathological alterations in tissue as in the case in glaucoma.[40] However, where once clinicians evaluated the eye by gently pressing their fingers into the closed eyelid of their patients, the advent of tonometers, sophisticated technology capable of measuring ocular pressure, has

again led to important advances in understanding eye disease and its treatment.[41,42] Similarly, technologies such as elastography are helping clinicians visualize changes in prostate stiffness related to cancer.[43,44]

While these technologies have created important advancements in understanding diseases of relatively homogeneous tissues (eye, prostate), heterogeneous tissues pose a significant challenge in determining stiffness related to pathology, disease or injury. The spine is such an example where changes in its stiffness may reflect specific conditions.[45,46] Unfortunately, there are few, if any, technologies that can quantify these changes to better aid clinicians. In addition, we do not know if the threshold of human stiffness detection is sufficient for detection of clinically important changes in spinal stiffness.

Relevance of the findings to clinical practice

This study revealed that while a majority of the clinicians could identify large differences in vertebral pair stiffness, the study sample had difficulty in successfully detecting differences in vertebral pairs when the stiffness gradient approximated the stiffness detected threshold. Since this threshold falls in the range of stiffness that has been observed to occur following clinical interventions such as spinal manipulation,[13] it appears that practitioners may have difficulty in detecting these post-intervention changes in stiffness of magnitudes similar to the detection threshold.

If there is an inability to detect a small magnitude change in stiffness that is clinically significant, it is possible to imagine a scenario where a practitioner may inappropriately change, or maintain a course of therapy based on faulty sensory information. In this case, the development of technologies to make this information available in the decision-making processes, may improve outcomes for musculoskeletal conditions.

Limitations

As can be the case in the Weber faction, we report our results as percentages so that data can be compared between circumstances such as differences in the surfaces from which stiffness data was collected (air bladder on a rigid surface, human subject lying prone on a padded plinth) and the surface area of palpation (the area of the clinician/subject interface and the area of the indenter). As a result, comparison of absolute stiffness values is not possible. Although the staircase method is very efficient in identifying sensory thresholds, this method relies on the reporting of the participant.19 The possibility that a participant has manipulated the results purposefully or through unintentionally guessing cannot be ruled out. Although our sample was small and the generalizability of these results is limited, physiological thresholds of human senses tends to be similar.47 In addition, we designed the study to be representative of what clinicians do in practice which meant giving them a sufficient amount of time to make their palpation determination rather than giving them multiple attempts. Therefore, we cannot know from this work if multiple palpation attempts would have improved clinician performance but speculate it would not as the physiological limitations of the clinician would not have changed with additional attempts. Last, although we did not measure spinal stiffness in the subject following palpation, prior reliability testing of the device shows that in symptomatic subjects, this measure is stable within the testing period.[24]

Conclusions

While clinicians can detect large stiffness changes between spinal regions, they are less able to detect smaller changes in stiffness that are similar in magnitude to previously published change changes in stiffness following successful interventions. These results allow us to speculate that the physiological limits of human touch may act to restrict the ability of clinicians to appreciate

changes in spinal stiffness as they approach the physiological limits of palpation which may influence clinical decisions although we do not yet know the clinical meaning of these small changes or if they are common. Given that instrumentation is used commonly to assist clinical hearing (stethoscope) and sight (loupes, ultraviolet light), assistive technology to aid a clinician's sense of touch (stiffness testing) may be helpful in better understanding how biological/biomechanical changes may be related to back pain within the biopsychosocial model.

Acknowledgements

References

1. Balagué F, Mannion AF, Pellisé F, Cedraschi C. Non-specific low back pain. *Lancet.* 2012;379(9814):482-491.

2. Rubin DI. Epidemiology and risk factors for spine pain. *Neurol Clin.* 2007;25(2):353-371.

3. Von Korff M, Saunders K. The course of back pain in primary care. *Spine* . 1996;21(24):2833-2837; discussion 2838-2839.

4. Pengel LHM, Herbert RD, Maher CG, Refshauge KM. Acute low back pain: systematic review of its prognosis. *BMJ.* 2003;327(7410):323.

5. Hoy D, March L, Brooks P, et al. The global burden of low back pain: estimates from the Global Burden of Disease 2010 study. *Ann Rheum Dis.* 2014;73(6):968-974.

6. Gore M, Sadosky A, Stacey BR, Tai K-S, Leslie D. The burden of chronic low back pain: clinical comorbidities, treatment patterns, and health care costs in usual care settings.

Spine . 2012;37(11):E668-E677.

7. Katz JN. Lumbar Disc Disorders and Low-Back Pain: Socioeconomic Factors and Consequences. *J Bone Joint Surg.* 2006;88(suppl_2):21.

8. Maniadakis N, Gray A. The economic burden of back pain in the UK. *Pain.* 2000;84(1):95-103.

9. Waddell G. *The Back Pain Revolution, 2nd Edition.* Churchill Livingstone; 2004.

10. Hill JC, Whitehurst DGT, Lewis M, et al. Comparison of stratified primary care management for low back pain with current best practice (STarT Back): a randomised controlled trial. *Lancet.* 2011;378(9802):1560-1571.

11. Korri SH, Miller RP, Todd DD. Kinesiophobia: a new view of chronic pain behaviour. *Pain Manag.* 1990;3:35-43.

12. Waddell G, Newton M, Henderson I, Somerville D, Main CJ. A Fear-Avoidance Beliefs Questionnaire (FABQ) and the role of fear-avoidance beliefs in chronic low back pain and disability. *Pain.* 1993;52(2):157-168.

13. Wong AYL, Parent EC, Dhillon SS, Prasad N, Kawchuk GN. Do Participants With Low Back Pain Who Respond to Spinal Manipulative Therapy Differ Biomechanically From Nonresponders, Untreated Controls or Asymptomatic Controls? *Spine* . 2015;40(17):1329-1337.

14. Delitto A, Erhard RE, Bowling RW. A treatment-based classification approach to low back syndrome: identifying and staging patients for conservative treatment. *Phys Ther.* 1995;75(6):470-485; discussion 485-489.

15. Flynn T, Fritz J, Whitman J, et al. A clinical prediction rule for classifying patients with low back pain who demonstrate short-term improvement with spinal manipulation. *Spine* . 2002;27(24):2835-2843.

16. Haneline MT, Cooperstein R, Young M, Birkeland K. Spinal motion palpation: a comparison of studies that assessed intersegmental end feel vs excursion. *J Manipulative Physiol Ther.* 2008;31(8):616-626.

17. Seffinger MA, Najm WI, Mishra SI, et al. Reliability of spinal palpation for diagnosis of back and neck pain: a systematic review of the literature. *Spine* . 2004;29(19):E413-E425.

18. Hestœk L, Leboeuf-Yde C. Are chiropractic tests for the lumbo-pelvic spine reliable and valid? A systematic critical literature review. *J Manipulative Physiol Ther.* 2000;23(4):258-275.

19. Cornsweet TN. The Staircase-Method in Psychophysics. *Am J Psychol.* 1962;75(3):485-491.

20. Mieritz RM, Kawchuk GN. The Accuracy of Locating Lumbar Vertebrae When Using Palpation Versus Ultrasonography. *J Manipulative Physiol Ther.* 2016;39(6):387-392.

21. Wong AYL, Kawchuk GN. The Clinical Value of Assessing Lumbar Posteroanterior Segmental Stiffness: A Narrative Review of Manual and Instrumented Methods. *PM R.*

December 2016. doi:10.1016/j.pmrj.2016.12.001

22. Tuttle N. Is it reasonable to use an individual patient's progress after treatment as a guide to ongoing clinical reasoning? *J Manipulative Physiol Ther*. 2009;32(5):396-403.

23. Fritz JM, Koppenhaver SL, Kawchuk GN, Teyhen DS, Hebert JJ, Childs JD. Preliminary investigation of the mechanisms underlying the effects of manipulation: exploration of a multivariate model including spinal stiffness, multifidus recruitment, and clinical findings. *Spine* . 2011;36(21):1772-1781.

24. Wong AYL, Kawchuk G, Parent E, Prasad N. Within- and between-day reliability of spinal stiffness measurements obtained using a computer controlled mechanical indenter in individuals with and without low back pain. *Man Ther*. 2013;18(5):395-402.

25. Wong AYL, Parent EC, Prasad N, Huang C, Chan KM, Kawchuk GN. Does experimental low back pain change posteroanterior lumbar spinal stiffness and trunk muscle activity? A randomized crossover study. *Clin Biomech* . 2016;34:45-52.

26. Karadogan E, Williams RL 2nd, Howell JN, Conatser RR Jr. A stiffness discrimination experiment including analysis of palpation forces and velocities. *Simul Healthc*. 2010;5(5):279-288.

27. Hicks GE, Fritz JM, Delitto A, Mishock J. Interrater reliability of clinical examination measures for identification of lumbar segmental instability. *Arch Phys Med Rehabil*. 2003;84(12):1858-1864.

28. Stochkendahl MJ, Christensen HW, Hartvigsen J, et al. Manual examination of the spine: a systematic critical literature review of reproducibility. *J Manipulative Physiol Ther*. 2006;29(6):475-485, 485.e1-e10.

29. Dishman RW. Static and dynamic components of the chiropractic subluxation complex: a literature review. *J Manipulative Physiol Ther*. 1988;11(2):98-107.

30. Huijbregts PA. Spinal Motion Palpation: A Review of Reliability Studies. *J Man Manip Ther*. 2002;10(1):24-39.

31. Maher C, Adams R. A psychophysical evaluation of manual stiffness discrimination. *Aust J Physiother*. 1995;41(3):161-167.

32. Haneline MT, Young M. A review of intraexaminer and interexaminer reliability of static spinal palpation: a literature synthesis. *J Manipulative Physiol Ther*. 2009;32(5):379-386.

33. Anders H-L, Corrie M, Jan H, et al. Standardized simulated palpation training--development of a palpation trainer and assessment of palpatory skills in experienced and inexperienced clinicians. *Man Ther*. 2010;15(3):254-260.

34. Bialosky JE, Bishop MD, Price DD, Robinson ME, George SZ. The mechanisms of manual therapy in the treatment of musculoskeletal pain: a comprehensive model. *Man Ther*. 2009;14(5):531-538.

35. Nicholson L, Adams R, Maher C. Reliability of a discrimination measure for judgements of non-biological stiffness. *Man Ther*. 1997;2(3):150-156.

36. Koppenhaver SL, Hebert JJ, Kawchuk GN, et al. Criterion validity of manual assessment of spinal stiffness. *Man Ther*. 2014;19(6):589-594.

37. Harbison J. "The old guessing tube": 200 years of the stethoscope. *QJM*. 2017;110(1):9-10.

38. Hajdu SI. The First Use of the Microscope in Medicine. *Annals of Clinical & Laboratory Science*. 2002;32(3):309-310.

39. Skedung L, Arvidsson M, Chung JY, Stafford CM, Berglund B, Rutland MW. Feeling small: exploring the tactile perception limits. *Sci Rep*. 2013;3:2617.

40. Liu B, McNally S, Kilpatrick JI, Jarvis SP, O'Brien CJ. Aging and ocular tissue stiffness in glaucoma. *Surv Ophthalmol*. June 2017. doi:10.1016/j.survophthal.2017.06.007

41. Zareei A, Razeghinejad MR, Nowroozzadeh MH, Mehrabi Y, Aghazadeh-Amiri M. Intraocular pressure measurement by three different tonometers in primary congenital glaucoma. *J Ophthalmic Vis Res*. 2015;10(1):43-48.

42. Okafor KC, Brandt JD. Measuring intraocular pressure. *Curr Opin Ophthalmol*. 2015;26(2):103-109.

43. Rouvière O, Melodelima C, Hoang Dinh A, et al. Stiffness of benign and malignant prostate tissue measured by shear-wave elastography: a preliminary study. *Eur Radiol*. 2017;27(5):1858-1866.

44. Wang Y, Yao B, Li H, et al. Assessment of Tumor Stiffness With Shear Wave Elastography in a Human Prostate Cancer Xenograft Implantation Model. *J Ultrasound Med*. 2017;36(5):955-963.

45. Li C, Sun H-L, Lu H-Z. Comparison of the effect of posterior lumbar interbody fusion with pedicle screw fixation and interspinous fixation on the stiffness of adjacent segments. *Chin Med J* . 2013;126(9):1732-1737.

46. Krenn MH, Ambrosetti-Giudici S, Pfenniger A, Burger J, Piotrowski WP. Minimally invasive intraoperative stiffness measurement of lumbar spinal motion segments. *Neurosurgery*. 2008;63(4 Suppl 2):309-313; discussion 313-314.

47. Nicholson LL, Adams RD, Maher CG. Manual discrimination capability when only viscosity is varied in viscoelastic stiffness stimuli. *J Manipulative Physiol Ther*. 2003;26(6):365-373.