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Structure and Function of Hip, an Attenuator of the Hsp70 Chaperone Cycle

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1 Summary

Protein must fold into unique three-dimensional structure in order to become functionally active. In the crowded environment of the cell, misfolded proteins easily clump together and form aggregates. Such protein aggregation contributes to neurodegenerative diseases, such as Alzheimer's, Parkinson's and Huntington's disease. Protein aggregation is prevented by specialized proteins called molecular chaperones, which recognize misfolded proteins and promote their productive folding.

The Hsp70 chaperone system is central to the cellular defense against toxic protein aggregation and for maintenance of protein homeostasis. Cytosolic Hsp70 mediates the folding of newly-synthesized and stress-denatured proteins in an ATP-dependent reaction cycle, but also helps in the transfer of misfolded protein species to the degradation machinery. To accomplish protein folding, Hsp70 is regulated by co-chaperones of the J-protein family and by nucleotide exchange factors (NEFs). These stimulate ATP hydrolysis and ADP dissociation, respectively. A further Hsp70 regulator, the protein Hip/ST13 (Hsp70 interacting protein), has been suggested to delay substrate release by slowing ADP dissociation from Hsp70.

Hip was first described in 1995, but its significance for protein homeostasis has only recently been appreciated. Hip, a dimer of ~ 40 kDa subunits, stabilizes Hsp70-substrate complexes in the ADP-bound state. Hip is conserved in metazoans and functions in stabilizing complexes of Hsp70 with aggregation-prone substrate proteins, including α -synuclein in Parkinson's and Tau in Alzheimer's disease.

This study presents an in-depth structural and mechanistic analysis of the Hsp70 regulator Hip. In isothermal titration calorimetry (ITC) experiments Hip bound preferentially to the ADP-bound form of Hsp70, however with relatively modest affinity. Binding of Hip slowed the dissociation of a fluorescent ADP analog from Hsp70. Circular dichroism spectrometry and small angle X-ray scattering (SAXS) showed that the structured units in the multi-domain protein Hip are flexibly linked by long tethers, resulting in an extended dimer structure. We determined the crystal structures of the N-terminal dimerization domain and the tetratricopeptide repeat (TPR) middle domain of Hip by experimental phasing at 1.1 Å and 2.6 Å resolution, respectively. For

the C-terminal DP domain, we constructed a homology model. The TPR middle domain is sufficient for Hsp70 binding. The C-terminal domain presumably mediates contacts to substrate proteins.

Furthermore, we solved the crystal structure of the core complex of Hip with the nucleotide binding domain (NBD) of Hsp70 at 2.7 Å resolution. The complex structure showed that Hip forms a bracket over the ATPase domain of Hsp70, reducing its otherwise dynamic nature and locking the ADP-bound state. As a consequence, dissociation of ADP is retarded and Hsp70-client protein complexes are stabilized. Structure-guided mutational analysis targeting the interaction interface fully validated the observed interactions between Hip and Hsp70.

Comparison of the structures of the Hip and NEF complexes suggested that Hip and NEF binding to Hsp70 are mutually exclusive. This was confirmed using a Hip-Hsp70 fusion protein, which forms a functional intramolecular interface. A yeast reporter assay employing the Hsp90 client protein glucocorticoid receptor (GR) showed that Hip must work as a dimer and interact also with the Hsp70-bound substrate protein to compete effectively with NEF binding to Hsp70. Removal of either the dimerization or the DP domain was functionally equivalent to mutations that block interactions with the NBD.

The presented data are in line with a model where association of Hip with specific Hsp70-substrate complexes attenuates their active cycling. This mechanism explains why Hip enhances aggregation prevention by Hsp70 and facilitates the transfer of specific proteins to downstream chaperones such as Hsp90 or the proteasome. Pharmacological activation of Hip may prove useful in the removal of potentially toxic protein species. Our findings may thus inform new strategies for the treatment of protein deposition diseases.

2 Introduction

Proteins constitute the largest fraction of the cell interior and fulfill numerous cellular functions, as metabolic enzymes, cellular regulators and in signal transductions. The different types of protein are synthesized as polymers of the common 20 amino acids. The functional properties of a protein are determined by its three-dimensional structure, which is encoded by the amino acid sequence. Therefore, determination of protein structures provides important clues about how proteins perform their function.

2.1 Protein folding and protein aggregation

To become biologically functional, newly synthesized proteins must correctly fold to their native state with a unique three-dimensional structure. The information for the structure of a protein is encoded in its primary sequence. This was demonstrated for the first time by Christian Anfinsen, who in 1972 received the Nobel Prize in Chemistry for his pioneering discovery of protein folding *in vitro* (Anfinsen, 1973; Anfinsen and Haber, 1961; Anfinsen et al., 1954).

Protein folding is not a random process. It follows a direct pathway from unfolded forms to the energetically stable native state. The main driving force of protein folding is the gain of entropy in the overall system through the hydrophobic effect, when hydrophobic regions are buried to form the hydrophobic core and the solvation water is released into the surrounding space. Additional stabilization comes from hydrogen bonding, salt bridges and van der Waals interactions.

The free enthalpy of the native state is around 10-15 kcal/mol compared to the unfolded state. Along the folding trajectory, metastable non-native intermediates can be formed. The folding properties of a protein can be illustrated by a funnel shaped energy landscape, representing the free-energy potential during protein folding (Fig. 1).

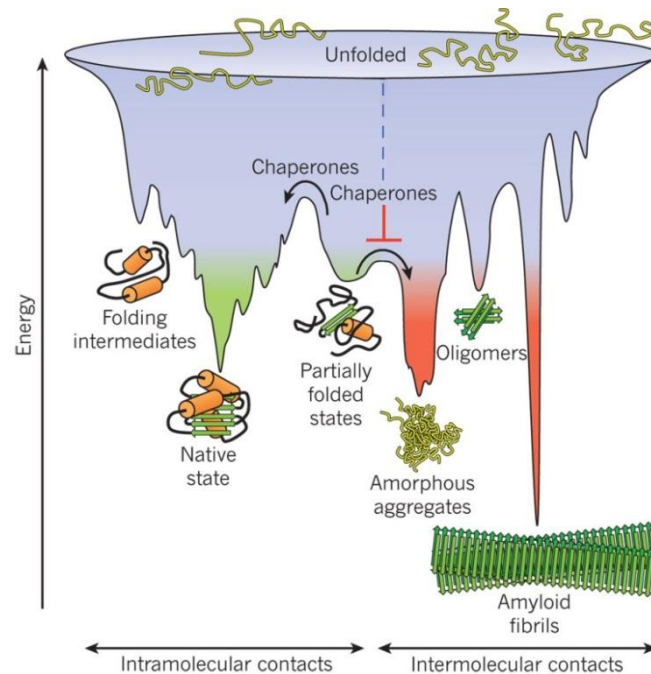


Figure 1: Energy landscape scheme of protein folding and aggregation. Local minima correspond to kinetically trapped intermediate states.

Left panel: Formation of intermediates towards native conformation *via* intramolecular interactions. Right panel: Misfolding through intermolecular interactions. The red color indicates the intermediates leading towards amorphous aggregates, toxic oligomers or amyloid fibrils *via* intermolecular contacts. Adapted from the review: (Hartl et al., 2011).

In the crowded environment of the cell, additionally intermolecular interactions contribute substantially to the folding energy landscape (Fig. 1). Partially folded or misfolded proteins tend to aggregate in a concentration-dependent manner, as they transiently expose hydrophobic residues to the solvent. Such aggregates are generally amorphous and their formation prevents productive folding. Alternatively, aggregation can lead to highly ordered, fibrillar aggregates named amyloid fibrils, in which β -strands run perpendicular to the long fibril axis (right panel in Fig. 1). Such forms appear to be energetically highly stable, perhaps more stable than the native state. These fibrillar amyloid aggregates are highly resistant to the cellular degradation machinery. The occurrence of such aggregates is common to neurodegenerative diseases such as Alzheimer's (AD), Parkinson's (PD), Huntington's (HD) disease as well as Amyotrophic lateral sclerosis (ALS) (Chiti and Dobson, 2006; Olzscha et al., 2011). A large load of misfolded species and aggregated protein are toxic to cells.

Aggregation of nonnative proteins is a by-product of *de novo* protein synthesis *in vivo*, whereby aggregation competes with protein folding. In addition, the aggregation of partially folded proteins and chain compaction is predicted to be enhanced by the crowded macromolecular environment, equivalent to a ~ 300-400 mg/ml protein solution, which increases the affinities between interacting proteins (Ellis, 2001). Crowding also provides a nonspecific force for macromolecular compaction and association (Minton, 2000), including the collapse of protein chains during folding and the interaction of nonnative proteins with molecular chaperones (Martin and Hartl, 1997).

2.2 Molecular chaperones

While in the test tube proteins may fold into their native state spontaneously, under cellular condition the folding efficiency is often limited by the side reactions of misfolding and aggregation. To overcome the problem of protein misfolding and aggregation, the cells employ a group of otherwise unrelated proteins called “molecular chaperones”.

Chaperones are highly conserved in all three branches of the tree of life, archaea, bacteria and eukaryotes. They only assist the protein folding and assembly process, but are not present in the final structures. Molecular chaperones interact with the nonnative protein during *de novo* folding and under stress conditions to increase the efficiency of protein folding or re-folding. They are distinguished from folding catalysts such as peptidyl-prolyl isomerases and protein disulphide isomerases (Schmid, 1993; Tuite and Freedman, 1994), while accelerate specific folding steps. Chaperones are constitutively expressed, but under stress conditions, their expression can be greatly increased, hence the term “heat-shock protein” or “stress protein”. They are also key players in the regulation of the heat shock response pathway, which is mediated by the transcription factor heat shock factor 1 (Hsf1). Chaperones are mainly divided into five major classes according to molecular weight: small Hsps, Hsp60, Hsp70, Hsp90 and Hsp100.

The molecular chaperone mechanism was originally proposed in 1987 (Ellis, 1987). The universal feature of molecular chaperones is their ability to recognize and bind non-native proteins. Therefore, they are essential for the correct and efficient folding of proteins. They perform mainly three activities: (1) recognizing and interacting with non-native or denatured polypep-

tide to form a stable chaperone protein complex; (2) preventing aggregation of unfolded and misfolded polypeptides and aiding their refolding; (3) helping the cell to maintain protein homeostasis. For the latter function, chaperones may transfer terminally misfolded proteins to proteolytic degradation by the ubiquitin proteasome system (UPS) or autophagy (Hartl et al., 2011; Rubinsztein, 2006). The cellular folding pathways mediated by molecular chaperones are well established (Fig. 2).

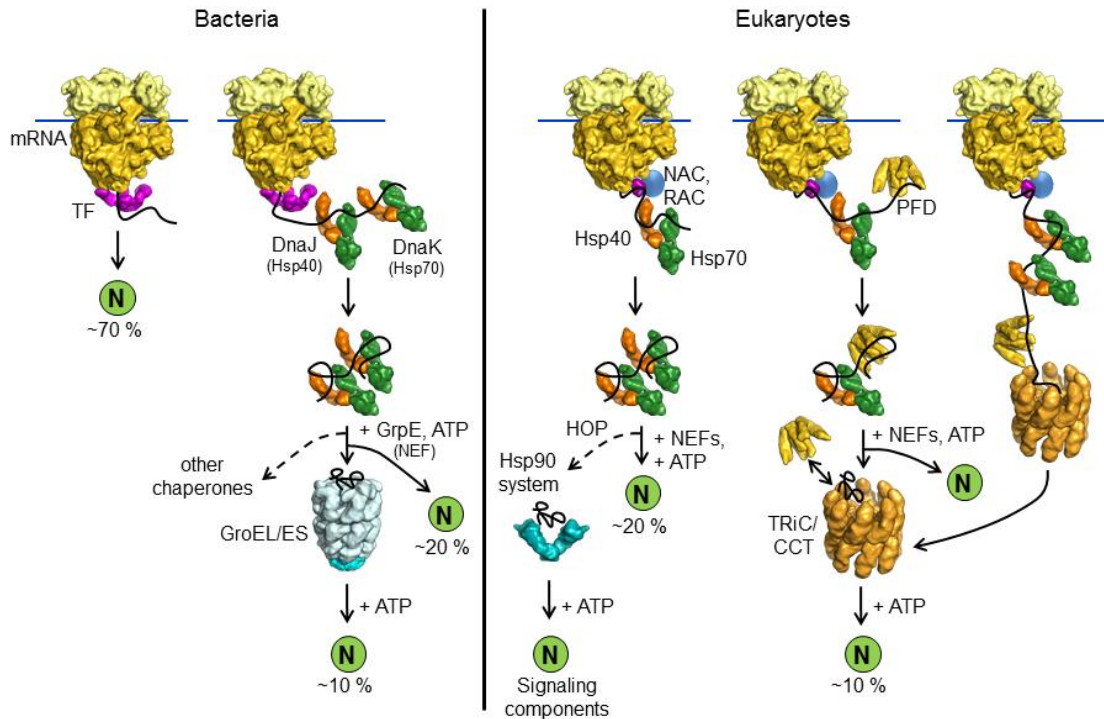


Figure 2: Model of the chaperone-mediated protein folding in the cytosol.

Left panel: In bacteria, newly synthesized polypeptide chains interact with trigger factor (TF) and around 70% of total proteins may fold directly to native state. Besides that, 20% of total require the DnaK system, bacterial homolog of Hsp70. The remaining 10% are subsequently transferred to the downstream chaperonin GroEL/GroES system to fold to the native state. Right panel: In eukaryotes, nascent chain-associated complex (NAC) interacts with nascent chain, similar to trigger factor. The Hsp70 family functions as second-tier player for longer nascent chains and mediates the co- or post-translational folding. Around 20% polypeptide can reach their native state through the assistance of ribosome-associated chaperones (RAC), Hsp70 and Hsp40. Hsp70 system also contributes around 10% protein to downstream chaperones to complete the folding, such as Hsp90 or chaperonin systems. Adapted from the review: (Hartl et al., 2011).

During synthesis of the polypeptide by the ribosome, part of the newly synthesized chain is shielded by the ribosomal exit tunnel. Beyond that, the hydrophobic side chains of partially synthesized polypeptide are exposed to the crowded environment of the cytosol. Thus the ex-

tended polypeptide needs to be protected from aberrant interactions and misfolding during translation. Therefore, molecular chaperones begin to act cotranslationally. A particular class of chaperones specifically recognizes the newly synthesized nascent-chain emerging from the ribosome. These are ubiquitously present in all domains of life. In the bacterial system, it consists of trigger factor (TF) and the Hsp70 system. In the eukaryotic system, the ribosome-associated chaperones (RAC) and nascent chain-associated complex (NAC) are involved besides Hsp70. The bacterial trigger factor is the best-characterized ribosome-associated chaperone. After capture by Hsp70, the polypeptide may be handed over to downstream chaperones such as the Hsp60 or Hsp90 systems for further folding and regulation. Therefore, we can summarize that protein folding starts cotranslationally and is completed posttranslationally.

The general understanding of the roles of chaperones in protein folding comes from the study of the chaperonins (Hsp60) and the Hsp70 system, which will be discussed below. The Hsp90 system will also be briefly described as an example for the complex mechanism of molecular chaperones.

2.2.1 Chaperonins

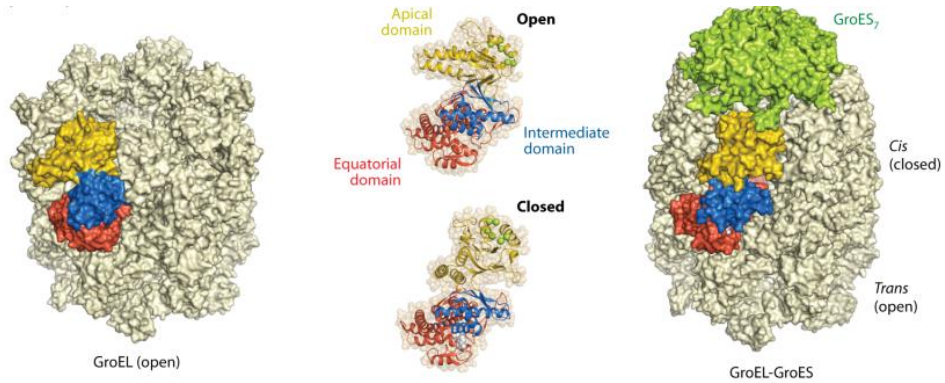
Chaperonins are a highly conserved class of macromolecular complexes. They form ~ 800 kDa large cylindrical folding compartments with a central cavity. Chaperonins are the downstream molecular machinery of Hsp70. The chaperonins have been classified into two families, groups I and II, with GroEL and TRiC as paradigms (Horwich et al., 2007; Tang et al., 2007).

Members of group I are found in bacteria (GroEL) and compartments derived from bacteria in eukaryotes, such as chloroplast (Cpn60) and mitochondria (mt-Hsp60). The chaperonin complexes of group I are composed of 14 subunits, forming two seven-membered rings. The double rings are stacked back-to-back and they cooperate with a co-chaperone (GroES, Cpn10 or mt-Hsp10), which acts as a lid and covers the top of the chaperonin central cavity (Fig. 3A).

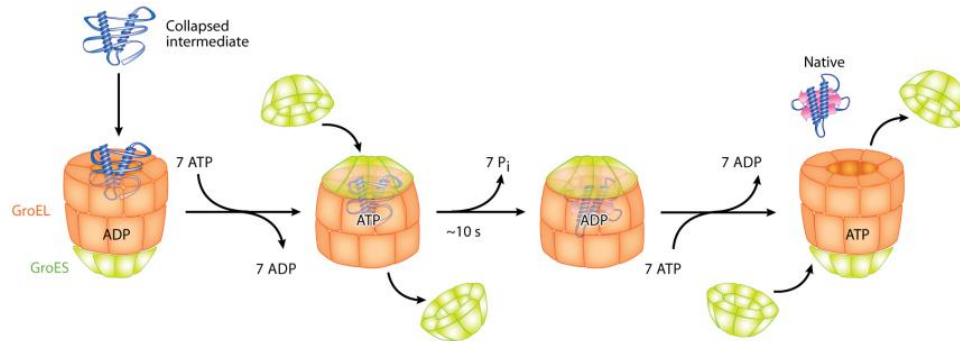
The best-studied and characterized bacterial system is GroEL-GroES, which function as a protein-folding cage (Hartl, 1996; Hartl and Hayer-Hartl, 2002; Horwich et al., 2007). The folding ability of GroEL depends on its molecular architecture (Fig. 3A). GroEL is a double-ring consisting of 14 identical subunits of ~ 57 kDa. Each subunit contains three structural parts: an

equatorial ATPase domain, an intermediate hinge domain and an apical substrate binding domain. Figure 3B shows the mechanism of GroEL-GroES in detail (Kim et al., 2013).

A)



B)



C)

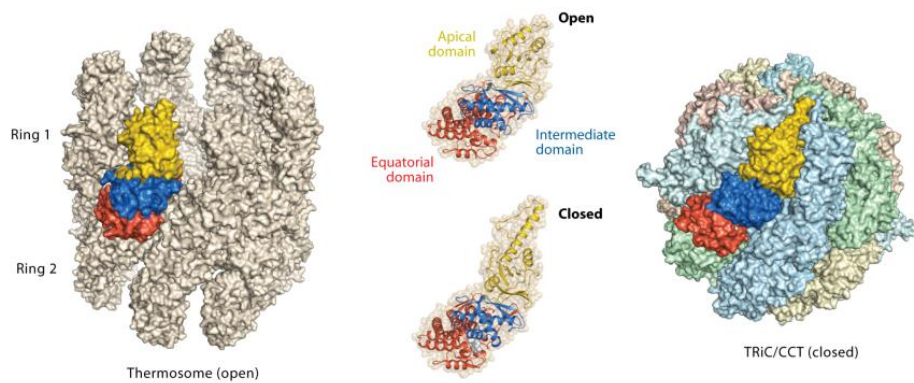


Figure 3: Structures and mechanism of the chaperonin system.

- A) Crystal structure of the group I chaperonin complex, GroEL-GroES. Left panel: The crystal structure of GroEL (PDB accession code 1SS8). Middle panel: Two conformational states of each subunit. The apical, intermediate and equatorial domains are colored in yellow, dark blue and red, respectively. Right panel: The asymmetrical GroEL-GroES complex (PDB accession code 1PF9).
- B) GroEL-GroES reaction mechanism. Substrate is bound by the open GroEL cis-ring of the asymmetrical GroEL-GroES complex. Binding of ATP to each of the seven GroEL subunits causes a conformational change in the apical domains. GroES binding traps the substrate in the cage. The encapsulated substrate is free to fold in the GroEL during the time needed to hydrolyze the seven ATP molecules bound to cis-ring (~ 10 s). GroES dissociates from the ring and the substrate is released.
- C) Crystal structures of group II chaperonins. Left panel: Open form of the homo-oligomeric thermosome (PDB accession code 3KFK). Middle panel: Subunit CCT3 of TRiC in two states, which is colored as follows to indicate its domain structures (equatorial nucleotide-binding domain in red; intermediated hinge domain in green and the apical substrate-binding domain in yellow). Right panel: Closed form of TRiC/CCT (PDB accession code 4D8Q).

The substrate is delivered from the upstream DnaK (Hsp70) system and is trapped on the GroEL ring. It may assume an extended or compact conformation. ATP-dependent apical domain movements result in sequential stretching and re-modeling the partially folded region. The binding of GroES traps the substrate in the cage. GroES is a dome-shaped homoheptamer of ~ 10 kDa subunits and regulates the ATPase activity of GroEL (Horwich et al., 2007; Xu et al., 1997). The substrate folds in the chaperonin cavity. After ATP hydrolysis, the substrate is released either in native state or as an incompletely folded intermediate state. The incompletely folded substrate is re-bound to GroEL and will start a new cycle. The whole folding process is regulated by ATP and GroES. Binding of GroES leads to allosteric conformational change of GroEL and results in an expansion of the central cavity (Hartl and Hayer-Hartl, 2009). Unlike Hsp70, GroEL recognizes a molten globule, locally expanded form of substrate, and promotes folding through cycles of encapsulation. Due to the volume of the cavity, the size of the trapped substrate is limited, ranging from 20-50 kDa. They have been identified by a proteomics analysis of GroEL/ES binding partners (Kerner et al., 2005).

Group II chaperonins such as thermosome and TRiC (TCP-1 ring Complex) exist in archaea and the eukaryotic cytosol (Fig. 3C). TRiC is also called CCT (Chaperonin Containing TCP-1). In contrast to group I chaperonins and thermosomes, TRiC is not stress inducible (Horwich et al., 2007). Though all chaperonins share a common cylindrical structure, substantial differences exist between groups I and II. TRiC contains eight different subunits per ring and each subunit is ~ 60 kDa. In addition, group II TRiC subunits contain α -helical protrusions that extend from the apical domain of the ring, which has been described as a built-in lid. Therefore, TRiC work independent from an additional GroES-like co-chaperone. Notably, TRiC subunits share se-

quence homology with GroEL at the ATP binding site, but not the substrate binding site. Therefore, TRiC was assumed to be specialized for a small subset of substrate proteins (Gomez-Puertas et al., 2004). A recent study suggested however that ~ 10 % newly synthesized proteins are TRiC substrates, ranging from the obligate substrate such as actin and tubulin to tumor suppressors (Yam et al., 2008). The broad range of substrate suggested that TRiC may work by a mechanism different from group I GroEL-GroES system.

2.2.2 The Hsp90 system

Similar to Hsp70, Hsp90 is a highly conserved and essential molecular chaperone in the mammalian cytosol and nucleus. Hsp90 functions in the downstream of Hsp70 and associates with non-native protein substrates. It mediates the conformational regulation of a wide range of client proteins involved in cell cycle control, apoptosis and especially signal transduction (Burrows et al., 2004; Pearl and Prodromou, 2000). The glucocorticoid receptor (GR) was identified as the first client protein of Hsp90 (Sanchez et al., 1985). Signaling components such as steroid-hormone receptors including progesterone receptor (PR), androgen receptor (AR) and kinases are the best-understood clients of Hsp90 (McClellan et al., 2007; Picard, 2006). Hsp90 is also essential for the cell's response to stress and is a key player in maintaining cellular homeostasis (Taipale et al., 2010).

Though a comprehensive understanding of the cellular functions and underlying mechanism of Hsp90 is still elusive, the crystal structures of full-length Hsp90 in complex with its co-chaperones provided deep insights into the structural dynamics and the regulation of Hsp90 (Ali et al., 2006; Pearl and Prodromou, 2000; Shiao et al., 2006).

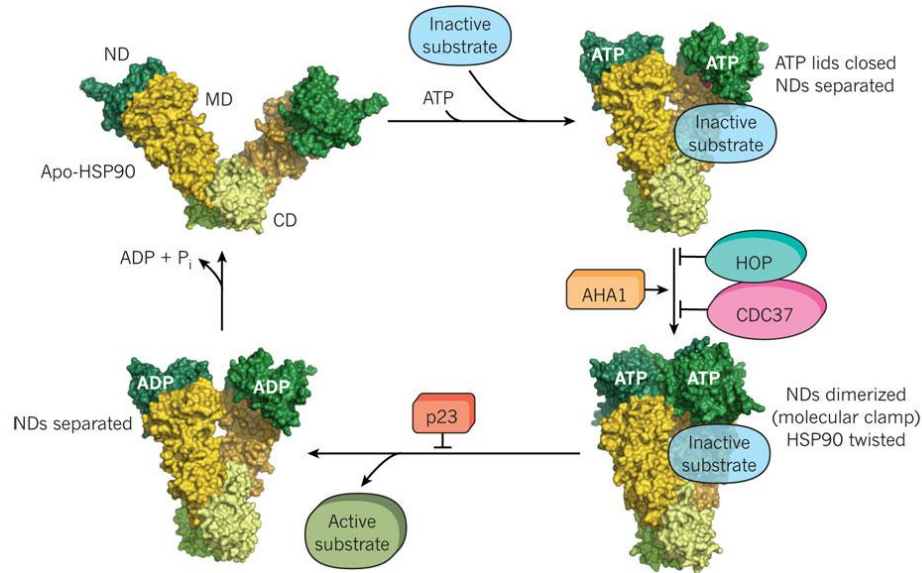


Figure 4: ATPase cycle of Hsp90.

Upon ATP binding, the N-terminal ATPase domain (ND) undergoes a conformational rearrangement, leading to the dimerization of ND. The dimerization forms a “molecular clamp” and results in a compact Hsp90 dimer. The ATP-bound state is metastable and committed to ATP hydrolysis. Transition to the ATP-hydrolysis competent state is regulated by co-chaperones. After ATP hydrolysis, the NDs dissociate and the protomers in the Hsp90 dimer remain connected by the C-terminal domain (CD). It has been suggested that the inactive substrate interacts mostly with the middle domain (MD) and is conformationally activated, when Hsp90 proceeds through the ATPase cycle. Adapted from the review: (Hartl et al., 2011).

Hsp90 exists as a flexible dimer, connected by its C-terminal domains. Each monomer consists of three domains: an N-terminal nucleotide binding domain (ND), followed with a middle domain (MD) and C-terminal dimerization domain (CD) (Figure 4). The ND possesses a deep nucleotide binding pocket, where ATP is bound in an unusual kinked manner. Similar to other chaperones, Hsp90 mediates ATP hydrolysis-dependent folding of substrates. The ATPase activity is essential for the function of Hsp90. Figure 4 shows the ATPase cycle of Hsp90 with its highly dynamic conformational changes. Besides the C-terminal dimerization domain, ATP binding promotes the dimerization of ND, resulting in a compact dimeric conformation of Hsp90. In this compact dimer, two monomers twist around each other and the NDs and MDs are also tightly associated. ATP hydrolysis leads to the dissociation of the NDs, resulting in the open conformational state of Hsp90, which enables the recognition of various substrates. Because of the broad extent of substrate specificities and the different structures of the known

substrates, Hsp90 is assumed to have multiple binding sites to substrates and the identification of binding sites on Hsp90 is still ongoing (Wandinger et al., 2008).

In addition, the CD domain contains an EEVD peptide motif at the end of C-terminus, which serves as a specific binding site for several co-chaperones, such as Hop and Chip, which bind to Hsp90 *via* TPR domains. Hop facilitates the cooperation between Hsp90 and Hsp70. Chip facilitates the cooperation of Hsp90 and proteasome. Hop inhibits N-terminal dimerization of Hsp90. It works as an adaptor for Hsp70 and Hsp90 and participates in steroid hormone receptor maturation by targeting clients to Hsp90 and forming the Hsp90-client complex. Moreover, Hsp90 is also targeted by several other co-chaperones without TPR domains, such as Cdc37, Aha1 and p23, which interact with Hsp90 in a sequential manner to regulate the chaperone machinery (Pratt and Toft, 2003; Smith, 1993). Cdc37 delivers specific kinase substrates to Hsp90 and acts as an ATPase inhibitor. Thereby, Cdc37 is thought to keep Hsp90 in an open state for transfer of the kinase. Conversely Aha1 stimulates ATP hydrolysis. In contrast to Aha1, p23 stabilizes the compact dimeric form of Hsp90 before ATP hydrolysis. In summary, the co-chaperones may control the conformational changes of Hsp90 and thereby modulate specific substrate processing by Hsp90.

It is widely recognized that the Hsp90 system is multifunctional and multicomponent chaperone machinery in the protein folding apparatus of the cell. In addition to client protein folding, Hsp90 has been shown to cooperate with the degradation machinery of the cell. This lead to a particular focus on Hsp90 with regard to pharmacological modulation (Schneider et al., 1996). Since it is responsible for the maturation of approximately 200 client proteins, among which several are oncoproteins (Workman et al., 2007), Hsp90 has gained attention as a critical target in cancer therapy, especially through the use of small compounds like geldanamycin that act as specific inhibitors of Hsp90. These drugs bind to the ATP binding pocket of Hsp90 blocking ATP binding. Thereby they specifically inhibit the ATPase function and arrest the Hsp90 functional cycle. The growing number of Hsp90 inhibitors has been evaluated in preclinical and clinical trials, by which a proof-of-concept has been established (Kim et al., 2009; Sharp and Workman, 2006; Workman et al., 2007).

2.2.3 The Hsp70 system

2.2.3.1 Diversity of Hsp70 isoforms

Hsp70s are the most ubiquitous chaperones and occur in all three domains of life: archaea, bacteria and eukaryotes (Hartl and Hayer-Hartl, 2002). The archaeal and bacterial Hsp70 is named DnaK. The eubacterium *Escherichia coli* has three Hsp70 isoforms (DnaK, HscA and HscC). The unicellular eukaryote *S. cerevisiae* contains four non-ribosome associated Hsp70s (Ssa1-4) and three ribosome-associated Hsp70s (Ssb1, Ssb2, Ssz1) in the cytosol, and organelle-specific isoforms in the endoplasmic reticulum (ER) and mitochondria (Lindquist and Craig, 1988; Nelson et al., 1992). In mammals, the constitutively expressed and stress-inducible isoforms are known as heat shock cognate 70 (Hsc70) and Hsp70, respectively, but more isoforms exist, 13 in total (Systematic names start with HSPA, followed by a number.). Hsc70 and Hsp70 share about 80 % sequence identity. The Hsp70s present in mitochondria and ER are known as mtHsp70/Grp75 and Bip/Grp78/Kar2, respectively. mtHsp70 is involved in the import of nuclear-encoded proteins into mitochondria. In the ER lumen, Bip is responsible for protein folding and quality control of proteins of the secretory pathway. Additional versatility is achieved by the employment of a multitude of Hsp40 family proteins (J proteins) and nucleotide exchange factors (NEFs) present in the eukaryotic cell, which act as co-chaperones. They control ATP hydrolysis of Hsp70 and recruit Hsp70 to specific substrate proteins.

2.2.3.2 Cellular functions of Hsp70

The Hsp70s have essential roles in protein metabolism under normal and stress conditions. They are central to many cellular processes, such as *de novo* protein folding, membrane translocation and misfolded protein degradation (Hartl and Hayer-Hartl, 2002; Neupert and Brunner, 2002; Ryan and Pfanner, 2001; Wang et al., 2013). One of the important roles of Hsp70 is to assist protein folding and turnover. Many neurodegenerative diseases involve aberrant accumulation of misfolded protein, and previous studies have linked Hsp70 to this process. *In vitro* studies showed that Hsp70 together with its co-chaperone Hsp40 can partially suppress the aggregation of huntingtin (htt) in the early prefibrillar state (Muchowski et al., 2000). Consistently, Hsp70 was found to co-localize with polyQ aggregates implicated in polyQ expansion diseases, and overexpression of Hsp70 and Hsp40 reduces polyQ aggregation and cytotoxicity

(Jana et al., 2000). Moreover, overexpression of Ssa1 (yeast homolog of Hsp70) decreases aggregation of htt and increased its SDS solubility (Krobitsch and Lindquist, 2000). Furthermore, Hsp70 has been reported to be sufficient to inhibit amyloid fibril formation by α -Synuclein, even in the absence of co-factors (Dedmon et al., 2005). Intriguingly, Hsp70 is able to recruit the co-chaperone ubiquitin ligase Chip for the ubiquitination of Tau, in contrast to Hsc70. One recent genetic study reported that Hsp70 directly stabilizes lysosomes and plays a key role in Niemann-Pick disease (Kirkegaard et al., 2010). All of the versatility of Hsp70 builds from its basic function. It binds the hydrophobic regions of unfolded polypeptides in an ATP-dependent manner.

In addition, Hsp70 appears to be a key component of cellular protein quality control. Hsp70 is abundantly expressed in many human tumors and this often correlates with metastasis and poor outcome in cancer patients (Brodsky and Chiosis, 2006; Patury et al., 2009). Up-regulated Hsp70 function is critical for the growth and survival of different human tumor cell lines (Brodsky and Chiosis, 2006; Patury et al., 2009; Powers et al., 2008). In light of the ample structural data available on this molecular chaperone, Hsp70 appears as an attractive putative drug target for the treatment of cancer.

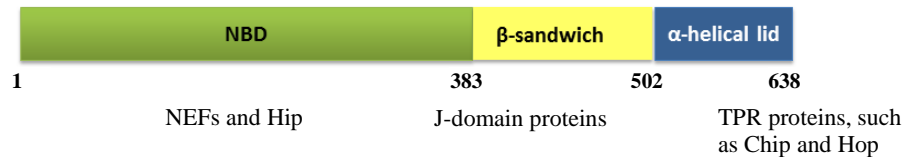
2.2.3.3 Structure and working cycle of Hsp70

All the Hsp70 forms are comprised of two domains, a ~ 45 kDa N-terminal nucleotide binding domain (NBD) and a ~ 25 kDa C-terminal substrate binding domain (SBD), which contains a ~10 kDa α -helical “lid” domain and a ~ 15 kDa β -sandwich (Fig. 5A). The NBD and SBD are connected *via* a highly conserved hydrophobic linker with 10-12 residues (Vogel et al., 2006). Structural studies have revealed that the NBD of Hsp70 is composed of two lobes, I and II. Each lobe consists of two subdomains named IA, IB and IIA, IIB, respectively. The two lobes form a heart-shaped structure with a large cleft at the bottom, where the nucleotide-binding site is located. The two subdomains look like a clamp that locks the nucleotide within the cleft. The nucleotide binding is coordinated by residues derived from all four subdomains. Subdomain IIB plays an essential role in the dissociation of nucleotide. It was well-known that NBD and SBD of Hsp70 in ADP-bound state are structurally and functionally almost independent (Chang et al., 2008; Swain et al., 2007). Conversely, in the ATP state, many biochemical and NMR

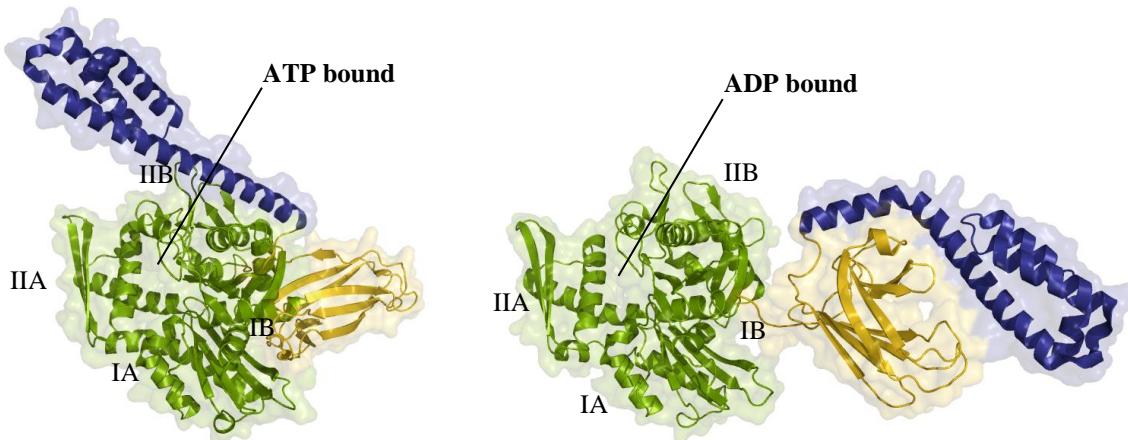
studies have suggested that NBD and SBD contact with each other (Buchberger et al., 1995; Mayer, 2010; Swain et al., 2007). However, the structure of ATP-bound full length Hsp70 had remained elusive until the crystal structure of an intact Hsp70 became recently (Kityk et al., 2012; Qi et al., 2013). In the ATP-bound state, the NBD and SBD form extensive contacts, which cause a dramatic conformational change in the SBD. The α -helical lid is separated from the β -sandwich domain and packs against the NBD, leading to an accessible peptide binding site. In the structure of an isolated SBD bound to a hydrophobic peptide, the β -sandwich harbors the peptide binding site, and the tightly associated α -helical domain forms a lid over the bound peptide (Zhu et al., 1996).

As a molecular machine, Hsp70 performs its function through crosstalk between ATPase activity in the NBD and substrate binding in the SBD. Hsp70 binds the hydrophobic polypeptide with its SBD, hydrolyzes ATP and then directs its substrate into a variety of distinct fates (Evans et al., 2010). Substrates of Hsp70 are thought to include newly synthesized linear polypeptides and partially folded proteins.

A)



B)



C)

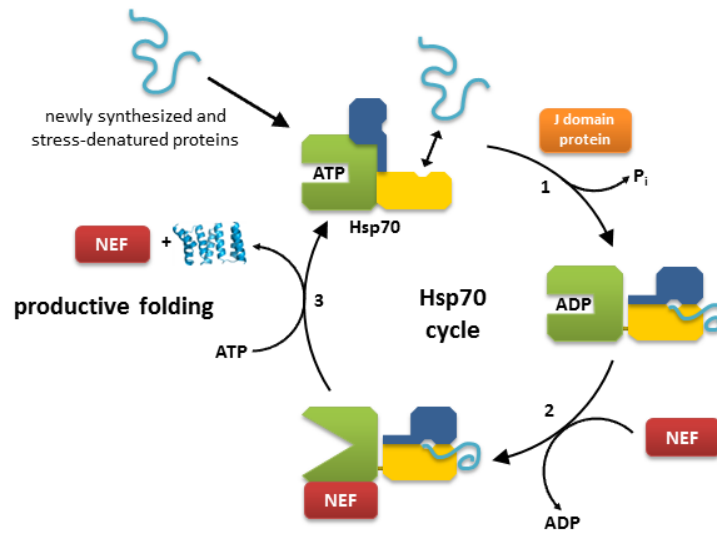


Figure 5: Structure and working cycle of Hsp70.

- A) Schematic representation of domain architecture of Hsp70 in DnaK *E.coli*. Hsp70 consists of two domains, the nucleotide-binding domain (NBD) in green color and the substrate-binding domain (SBD), which is composed of a ~ 15 kDa β -sandwich domain and 10 kDa α -helical “lid” domain, shown in gold and dark blue, respectively. Co-chaperone interaction sites with Hsp70. Only eukaryotic Hsp70 in cytosol contain an EEVD sequence at their C-terminus, which mediates the interaction with tetratricopeptide repeat TPR proteins, such as C-terminus of Hsc70 interacting protein (Chip) and Hsp70-Hsp90 organizing protein (Hop).
- B) Left panel: Crystal structure of bacterial Hsp70, DnaK in the ATP-bound open state. Right panel: Closed state in ADP-bound of *E. coli* DnaK (PDB: 2KHO), which was solved by NMR spectroscopy using the crystal structures of the individual domains.
- C) The ATP-dependent reaction cycle of Hsp70, regulated by J-domain proteins and nucleotide-exchange factors (NEFs).

Hsp70 is an allosteric protein. It has two conformational states with distinct substrate binding properties: the open state with low affinity to substrate, and closed states with high affinity (Fig. 5B). Substrate binding stimulates ATP hydrolysis, while ATP binding enhances substrate release. Figure 5C shows the model of Hsp70 working cycle, which is well-understood for the bacterial Hsp70 homolog DnaK. To accomplish the ATPase cycle, distinct classes of co-chaperones are required. The J-domain protein DnaJ is required for stimulation of ATP hydrolysis and the nucleotide exchange factor (NEF) GrpE for ADP dissociation from Hsp70. The collaboration with these important co-chaperones drives the transition of Hsp70 between the ATP and ADP states. ATP binding to the Hsp70 NBD increases the rate of substrate binding to SBD. Conversely, interactions between SBD and substrate increase the rate of ATP hydrolysis

through communication between the two functional domains (Mayer et al., 2000). In the absence of J-domain proteins, Hsp70 is predominantly in the ATP-bound state, since the intrinsic ATPase activity of Hsp70 is low (Schmid et al., 1994).

The elucidation of the DnaK mechanism in detail has provided the paradigm for the function of the eukaryotic Hsp70 chaperone system. There are additional co-chaperones to control substrate binding by independently binding to specific substrates, leading to a kind of substrate selection for Hsp70 (Kota et al., 2009; Vos et al., 2008). Recently, a novel protein Hikeshi was discovered that recognizes the ATP-bound state of Hsp70 and mediates its transport into the nucleus under heat-shock stress condition (Kose et al., 2012). Hsp70 distribution between cytosol and nucleus might be important for the decision between re-folding and degradation.

2.2.4 Hsp40 co-chaperones

The working cycle of Hsp70 starts with the action of J-domain proteins. They are also referred to as Hsp40, which induce tight binding of substrate to Hsp70. Hsp40 proteins have been well established to recognize and stabilize unfolded substrates for delivery to the hydrophobic pocket of Hsp70 (Gamer et al., 1996). They were proposed to interact with the NBD and the highly conserved hydrophobic linker of Hsp70 through its J-domain and to coupling substrate with its C-terminal substrate binding domain (Karzai and McMacken, 1996). Furthermore, substrate and J-domain protein binding synergistically stimulated ATP hydrolysis by Hsp70 (Laufen et al., 1999).

J-proteins are a heterogeneous group of multi-domain proteins. Though they are thought to act as co-chaperones of Hsp70, their number of homologs far exceeds that of Hsp70. There are six J-domain proteins in *E. coli*, 20 in *S. cerevisiae*, 33 in *C. elegans* and presumably 44 in human cells (Qiu et al., 2006). Based on their domain composition, they are subdivided into three classes as depicted in Figure 6: Type I: DnaJA (DnaJ *E.coli*, Ydj1 yeast, HDJ-2 human), Type II: DnaJB (Sis1 yeast, HDJ-1 human) and Type III: DnaJC (pyJ polio virus) (Cheetham and Caplan, 1998). Type I has full domain conservation with *E. coli* DnaJ, which contains four domains: N-terminal J-domain, followed by a glycine-phenylalanine rich region, a Zn-binding domain and a C-terminal domain. The α -helical J-domain is composed of ~ 70 amino acids and

responsible for the interaction with Hsp70 (Wall et al., 1994). Type II is highly similar to Type I apart from missing the Zn-binding domain. Type III is more divergent as it only shares similarity to the J-domain segment of types I and II. Since the C-terminal domains are highly divergent, this class is less explored so far.

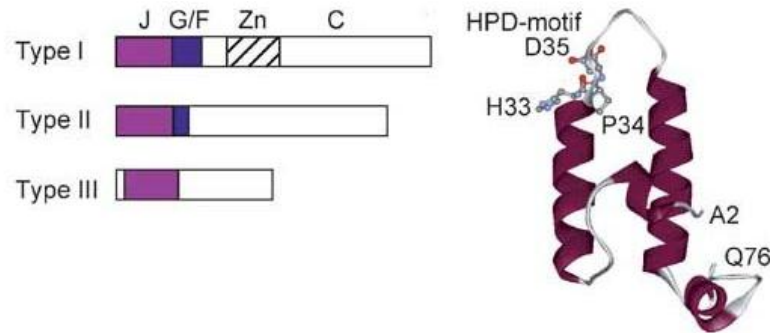


Figure 6: J-domain protein family.

Left panel: Schematic representation of domain architecture of three subfamilies of J-proteins. J: N-terminal J-domain (magenta color). G/F: Gly-Phe rich region. Zn: Zn^{2+} binding domain. C: C-terminal domain of homology. Right panel: Ribbon representation of the NMR structure of J-domain of DnaJ in *E. coli*. Adapted from review: (Mayer and Bukau, 2005)

The structure of the J-domain composed of four α -helices is common to all J-domain proteins (Fig. 6). The central two long anti-parallel α -helices are connected by an exposed loop with a conserved signature sequence, histidine-proline-aspartate (HPD). NMR and mutagenesis studies revealed that both of these two anti-parallel α -helices are necessary for the binding to Hsp70 and modulating ATP hydrolysis (Landry, 2003). The interactions between J-domain and Hsp70 trigger an allosteric conformation change, which accelerate ATP turnover by approximately seven-fold (Pierpaoli et al., 1997). In addition, a crystallographic study suggested that J-domain stimulated ATPase activity by directing the Hsp70 linker between NBD and SBD to displace their interactions (Jiang et al., 2007). Furthermore, binding of J-domain to Hsp70 displaced the SBD from NBD, which may activate Hsp70 for substrate binding through the SBD flexibility to capture diverse substrates (Jiang et al., 2007). Despite of its function in substrate targeting to Hsp70, the J-domains has been reported that it can stimulate the binding of Hsp70 to polypeptides (Misselwitz et al., 1998). The solid binding assay demonstrated that J-domain binding to Bip and activated Bip to bind more substrates, suggesting J-domain activation broadened the substrate binding range of Bip (Misselwitz et al., 1998). Therefore, it could be proposed that

substrate specificity of Hsp70 could be largely influenced by its partner, J-proteins. Moreover, the cooperation of J-domain proteins with Hsp70 can prevent the aggregation of unfolded protein.

2.2.5 Nucleotide exchange factors (NEFs)

A distinct class of co-chaperones involved in the Hsp70 ATPase cycle is the nucleotide exchange factors (NEFs), which catalyze ADP dissociation. Nucleotide dissociation is a crucial step and therefore it is regulated by a variety of NEFs in eukaryotes. Four major NEF families in the cellular cytosol are classified as: GrpE-like, BAG proteins, HspBP1 and Hsp110 (Harrison, 2003; Kabani et al., 2002; Kabbage and Dickman, 2008; Shaner and Morano, 2007). In the bacterial system, ADP is firmly trapped in DnaK. In the absence of GrpE, the bound nucleotide cannot be released effectively. This will prolong substrate binding by DnaK to an extent that cell function is disrupted. GrpE is therefore essential in *E. coli* (Brehmer et al., 2004). However, unlike the bacterial DnaK, the eukaryotic Hsp70 has a considerable off-rate for ADP. In principle, it could perform its work in cells in a NEF-independent way. Thereby, nucleotide exchange is apparently not the rate-limiting step in eukaryotes. This is in sharp contrast to the essential role of GrpE for DnaK. However, eukaryotic Hsp70 can interact with nucleotide exchange factors to further accelerate the rate of nucleotide dissociation (Höfelfeld et al., 1995).

A number of crystal structures of Hsp70-NEFs complexes has been determined (Harrison et al., 1997; Polier et al., 2008). All the NEFs appeared to trigger an outward rotation of subdomain IIB in the Hsp70 NBD, except for HspBP1. Consequently, the bound nucleotide can be released more easily. However, each family probably prefers a slight different mechanism.

Figure 7 shows an overview of the different forms of NEFs defined from different family.

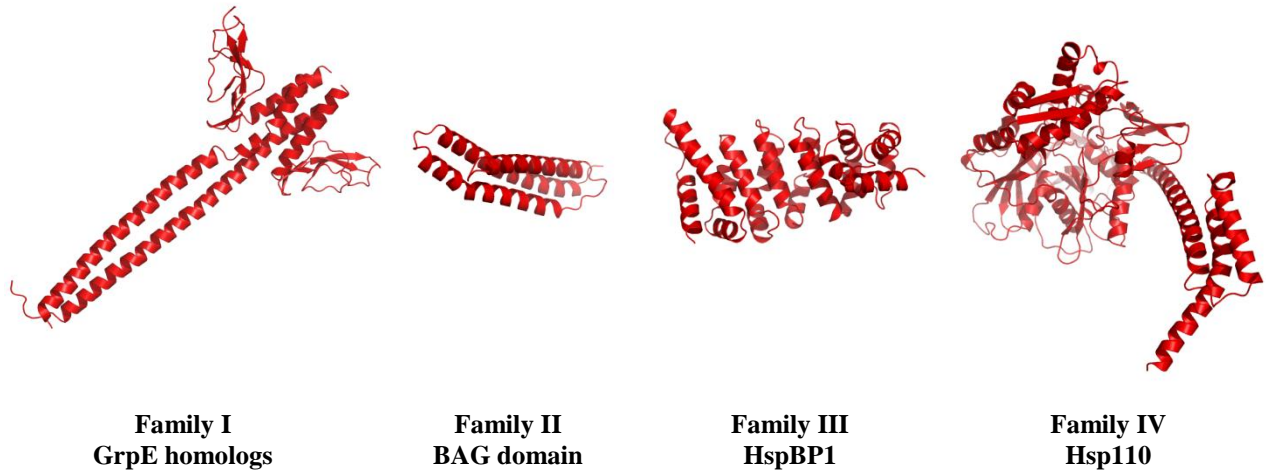


Figure 7: Structures of the different NEFs shown in ribbon representation.

GrpE is the NEF of the *E. coli* homolog of Hsp70, DnaK. It forms a homodimer, which is mediated by the α -helical domain. Form II, the BAG domain, is composed of three helices which are present in most BAG-domain proteins, except for BAG2. Family III shown here is the core domain of human HspBP1. It is an all- α -helical protein. Sse1 of family IV is the yeast homolog of Hsp110, which is structurally highly similar as Hsp70, containing NBD, β -sandwich and three α -helices domains. All structures are shown in ribbon representation and presented on the same scale. PDB accession numbers are as follows: 1DKG (Harrison et al., 1997), 1HX1 (Sondermann et al., 2001), 1XQS (Shomura et al., 2005) and 3D2F (Polier et al., 2008).

2.2.5.1 Family I

GrpE (GroP-like gene E) functions as a nucleotide exchange factor for bacterial Hsp70, DnaK. It is heat-inducible and essential in *E. coli*. GrpE eukaryotic homologs are found in mitochondria and chloroplasts cell organelles derived from bacteria. The yeast mitochondrial GrpE is named as Mge1, which is not heat-inducible in contrast to GrpE (Ikeda et al., 1994). Both share low sequence homology with only 34 % identity and 57 % similarity (Laloraya et al., 1994). Dimers of GrpE show an intriguing architecture with two almost parallel long α -helices contributing to a four-helix bundle and two proximal β -sheet domains, which look like two wings (Fig. 7). In the co-crystal complex structure with DnaK, one monomer is bent towards DnaK, which leads to an asymmetric dimer appearance (Harrison et al., 1997).

2.2.5.2 Family II

There are six homologs in the family of BAG proteins in humans, named BAG1 to BAG6. All the homologs contain one BAG domain, except for BAG5, which is comprised of five putative

BAG domains. In contrast to the α - β -dimer structure of GrpE, the BAG domain of BAG1 (Bcl2-associated athanogene) forms a three-helix bundle (Fig. 7). BAG-1 was initially thought as a cellular partner to bind the anti-apoptotic protein Bcl-2 and to promote cell survival (Takayama et al., 1995). The six BAG proteins are modular multi-domain proteins and seem to recruit Hsp70 for specific functions, though they employ a rather similar way to induce conformational change on Hsp70. Besides the BAG domain, BAG1 isoforms also share an ubiquitin-like Ub1 domain which was implicated in proteasomal degradation, similar to the co-chaperone Chip. BAG3 is involved in autophagy.

2.2.5.3 Family III

HspBP1 (Hsp70 binding protein 1) has another structural framework that was recruited for NEF function, a succession of armadillo repeats found in diverse structural contexts. HspBP1 is relatively abundant and it was initially described as an Hsp70-interacting protein that inhibited Hsp70-mediated protein refolding (Raynes and Guerriero, 1998). HspBP1 homologs represent a distinct class of conserved NEFs and relative abundant in cytosol, comparing to BAG domain proteins. The HspBP1 homolog in *S. cerevisiae* cytosol was named Fes1p. The structure of the core fragment of HspBP1 has an all- α -helical structure containing four armadillo repeats (Fig. 7) (Shomura et al., 2005). In contrast to the other NEFs, HspBP1 binding to the NBD causes much more drastic structural changes. It was suggested that HspBP1 distorts the lobe I of Hsp70NBD and separates it from lobe II.

2.2.5.4 Family IV

The principal class of eukaryotic NEF as judged from its abundance appears to be Hsp110, which is predominantly located in cytosol and its homolog Grp170 in the ER-lumen. After Hsp70 and Hsp90, it is the third most abundant heat shock protein in most mammalian cell lines and tissues. There are three human Hsp110 homologs, Hsp105, Apg1 and Apg2, in mammalian cytosol. Two homologs, Sse1 and Sse2, are found in the cytosol of *S. cerevisiae* (Easton et al., 2000). They are highly homologous with 76 % sequence identity. Both of them are up-regulated upon stress. Sse1 is abundantly expressed under normal cellular conditions. Disrup-

tion of Sse1 causes a growth defect and deletion of Sse2 causes no distinct phenotype, however double deletion results in synthetic lethality (Mukai et al., 1993).

Interestingly, Hsp110 is a remote homolog of Hsp70 and shares the general domain composition with Hsp70. It consists of an NBD, a β -sandwich domain and a three-helix bundle domain (Fig. 7). The main difference with Hsp70 is the presence of an acidic insertion into the β -sandwich domain and an extended C-terminus. It was previously hypothesized that the Sse1-ATP structure represents an evolutionary vestige of Hsp70-ATP (Liu and Hendrickson, 2007). Intriguingly, the recent available structure of DnaK-ATP showed a similar structure, confirming that Hsp70s and Hsp110 are homologs (Qi et al., 2013). The most remarkable difference between the DnaK-ATP and Sse1-ATP structures was the altered conformation of the β -sandwich and three-helix bundle domains.

2.2.5.5 Cellular functions of NEFs

Through their crucial role as nucleotide exchange factors, Hsp70 NEFs perform a wide variety of cellular functions, ranging from transcriptional regulation to the control of cell migration. In addition, some may participate in the targeting of substrates to the proteasome (Lüders et al., 2000). For the BAG-1 protein a role in coordinating chaperone and degradation pathways was suggested. The ER homolog of HspBP1 in humans, Sil1, has been revealed as an essential component of the endoplasmic-reticulum-associated protein degradation (ERAD) machinery and the unfolded protein response. Moreover, disruption of Sil1 leads to accumulation of misfolded protein in ER and nucleus, which causes a neurodegenerative disease, Marinesco-Sjögren syndrome (Anttonen et al., 2005; Senderek et al., 2005). Hsp110 is a distant Hsp70 homolog, possessing holdase activity, which can stabilize unfolded proteins and prevent aggregation. It was initially characterized as “holdase”, as they can keep denatured protein in solution (Oh et al., 1997; Oh et al., 1999). In addition, it can bind to a misfolded protein, but alone lacks substrate refolding activity (Dragovic et al., 2006). Passive substrate binding has been attributed to Hsp110 which might be related to Hsp70-mediated folding (Polier et al., 2010). Furthermore, Hsp110s were found associated with amyloidogenic proteins and disease-related protein aggregates such as from mutant superoxide dismutase 1 (SOD1) or tau (Eroglu et al., 2010; Olzscha et al., 2011; Yamashita et al., 2007). More recently, Hsp110 has been implicated

to empower Hsp70-Hsp40 to efficiently resolubilize and reactivate substrate protein *in vitro* (Rampelt et al., 2012). Moreover, down-regulation of Hsp110 triggered formation of heat shock-induced luciferase aggregates in the model system *C.elegans*. This finding demonstrated that *C. elegans* unable to resolubilize luciferase aggregate in the absence of Hsp110, suggesting Hsp110 is an essential component of protein disaggregation machinery (Rampelt et al., 2012). In line with this finding, deletion of Hsp105 causes accumulation of hyper-phosphorylated tau and neurofibrillary tangles in mice (Eroglu et al., 2010). Taken together, the multiple functions of NEFs indicate the functional significance of NEFs in the Hsp70 molecular machinery.

2.3 Hsp70 protein-protein network

2.3.1 Eukaryotic Hsp70 protein-protein network

Hsp70 as a major chaperone in the mammalian cytosol and nucleus has been widely recognized not to act alone, but to cooperate with multiple cofactors and other chaperone systems to maintain a functional proteome (Buchner, 1999; Caplan, 1999; Frydman and Höfelfeld, 1997). These cofactors do not only regulate the ATPase activity and substrate affinity of Hsp70, but also recruit Hsp70 to other chaperones and to components of the protein degradation machinery, thereby distributing functional proteins to functional complexes and organelles and damaged and misfolded proteins to proteolytic compartments. Figure 8 provides an overview of the main interactions of Hsp70 summarized by online database STRING (<http://string-db.org/>).

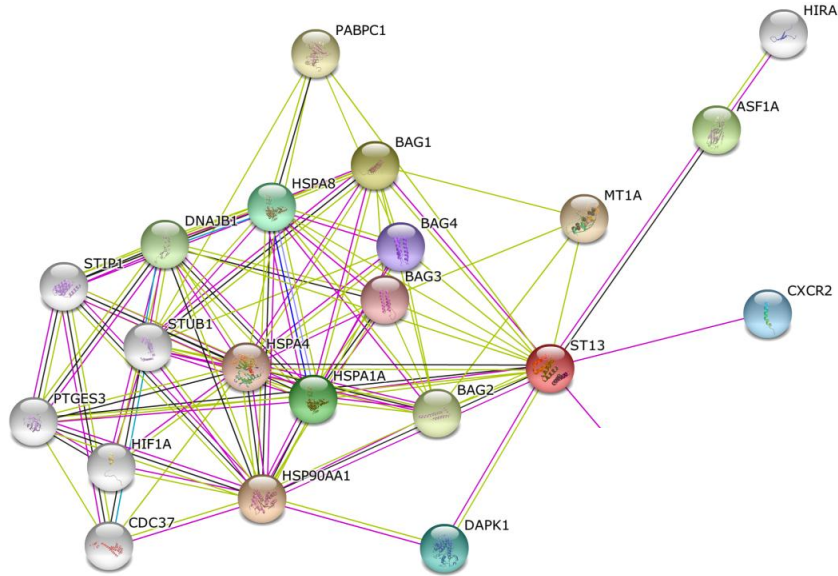


Figure 8: Overview of protein-protein interactions in the human Hs

The network was generated by database STRING. The interactions include direct and indirect associations.

For the cooperation of Hsp70 with the downstream Hsp90 system and the ubiquitin proteasome system (UPS), mainly three cofactors have been implicated in Figure 8: Hip (ST13), Hop (STIP1) and Chip (STUB1). In the following chapters, each of them is discussed in greater detail, but with special emphasis on Hip.

2.3.2 Hsp70 interacting protein (Hip)

Animals, plants and protozoa contain a conserved additional regulator of the Hsp70 reaction cycle, the cytosolic Hsp70 interacting protein (Hip). Alternative names for Hip are suppressor of tumorigenicity (ST13) and human p48. Human p48 shares over 91 % identity with rat Hip. It is a highly conserved protein found in most eukaryotic lines, but absent in *S. cerevisiae* and other fungi (Appendix 8.2). The Hip concentration in reticulocyte lysate is $\sim 1 \mu\text{M}$, similar to Hsc70. Hip is a dimeric protein of protomers with $\sim 43 \text{ kDa}$ molecular weight (human Hip). Hip was originally identified as a 368 residue protein binding specifically to the NBD of Hsp70 in the presence of ADP and participating in the regulation of eukaryotic Hsp70 protein (Höfelfeld et al., 1995). Independently, Hip was identified as a component (p48) of a complex of a progesterone receptor folding intermediate, which also contained Hsp70 and Hsp90 (Smith et

al., 1995). Hip was also found as interaction partner of the chemokine receptors C-X-C-motif chemokine receptors type 2 and 4 (CXCR2 and CXCR4), required for receptor internalization (Fan et al., 2002).

The existence of Hip suggests a regulatory mechanism in the eukaryotic cytosol distinct from bacterial DnaK. Hip was proposed to stimulate the chaperone activity of Hsp70 by preventing premature substrate release (Mayer and Bukau, 2005). Besides affecting the Hsp70 chaperone activities *in vitro* and *in vivo*, Hip possesses intrinsic molecular chaperone activity, as it binds specifically to unfolded proteins and prevents their aggregation (Höhfeld et al., 1995; Velten et al., 2002; Velten et al., 2000). Hip however cannot fold substrate proteins independently from Hsp70 and has no intrinsic ATPase activity (Bruce and Churchich, 1997; Höhfeld et al., 1995; Velten et al., 2000). Hip may be specifically recruited to Hsp70 by certain substrates. In addition, Hip was also proposed to compete with BAG1 for binding to the NBD of Hsp70, thereby preventing BAG1 stimulated ADP release (Höhfeld and Jentsch, 1997).

2.3.2.1 Predicted domain structure of Hip

Hip is thought to be composed of five domains, a dimerization domain at its N-terminus, followed by a tetratricopeptide repeats (TPR) domain flanked by highly negatively charged regions and a conserved region at its C-terminus containing several GGMP peptide repeats and a DP domain. In the absence of structural data, most of these domains were proposed based on sequence alignment criteria, structural modelling and mutagenesis studies (Irmer and Höhfeld, 1997; Prapapanich et al., 1996b; Velten et al., 2000). The schematic representation of the domain structure of Hip is shown in Figure 9.



Figure 9: Overview of the domain structure of Hip.

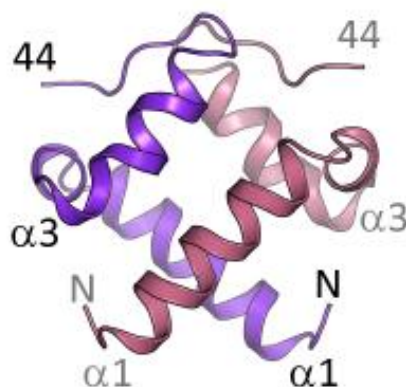
The residue numbers refer to Hip from *Rattus norvegicus*. The dimerization domain (HipN), Hsp70-binding TPR domain (HipM) and the DP domain are shown in purple, blue, and orange, respectively. White, acidic linker regions; Yellow, GGMP repeat segment.

2.3.2.2 The N-terminal dimerization domain of Hip

Initially the number of Hip subunits in the Hip homo-oligomer was unclear (Höfheld et al., 1995; Ziegelhoffer et al., 1996). Later Hip was unambiguously shown by a combination of size-exclusion chromatography and analytical ultracentrifugation to form elongated dimers (Velten et al., 2000). Deletion of 14 residues from the N-terminus of Hip resulted in the apparent loss of Hip homo-oligomerization. This indicates that dimerization was required for Hip-assisted nuclear receptor maturation *in vivo* (Irmer and Höfheld, 1997; Nelson et al., 2004; Nollen et al., 2001).

In the course of my master project, I cloned the conserved 44 residues at the N-terminus of rat Hip, named Hip(1-44), and determined its crystal structure at 1.1 Å resolution (Zhuo Li, Master thesis). An exchange experiment using Hip(1-44) and full length Hip revealed slow exchange between the dimers (Figure 10C), suggesting stable dimerization on a molecular time scale (Zhuo Li, Master thesis).

A)



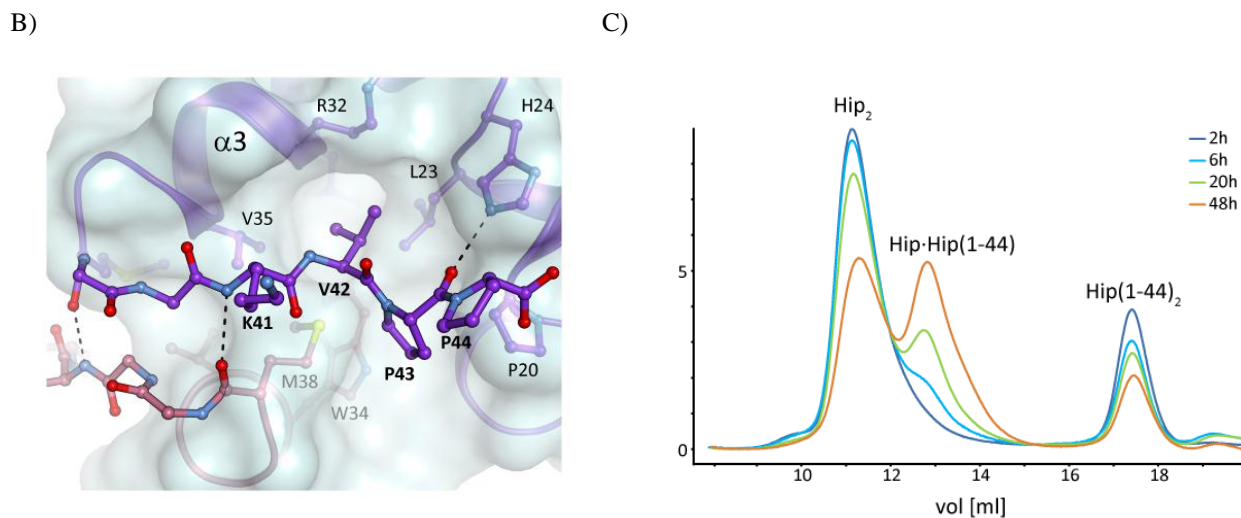


Figure 10: The dimerization domain of Hip.

- A) Ribbon representation of the crystal structure of the dimerization domain of Hip. Protomers are shown in purple and salmon. Chain termini are indicated.
- B) The dimer structure is stabilized by the contacts at the C-terminal end of the dimerization domain. The hydrogen bonds involved are indicated by dashed lines.
- C) Slow subunit exchange between Hip dimers (Hip_2 and $\text{Hip}(1-44)_2$) analyzed by size-exclusion chromatography. A stoichiometric mixture of Hip and $\text{Hip}(1-44)$ was incubated at 20 °C and analyzed at the times indicated.

The structure showed a compact, α -helical dimer with each subunit contributing three helices (Fig. 10A). Interestingly, the helices are oriented at nearly 90° angles. This leads to a box-like parallel dimer structure, which is stabilized by van-der-Waals interactions of the side chains in the hydrophobic core. The C-terminal loops are stabilized by hydrogen bonds between the backbones at Lys41 and Met38 (Fig. 10B). No structural homologs were found in the Protein Data Bank with the DALI server. Similarly, no sequence homologs were found apart from sequences that were likely Hip homologs. This domain type might thus be unique to the Hip protein family.

To characterize the stability of the Hip dimer, the monomer exchange between Hip_2 and $\text{Hip}(1-44)_2$ at 20 °C was investigated over time by size-exclusion chromatography. During hours, the Hip-Hip(1-44) heterodimer complex band gradually increased, suggesting slow exchange (Fig. 10C). The estimated half-time of exchange is ~ 6 h. This indicates that Hip forms a constitutive dimer on a physiological timescale.

2.3.2.3 The TPR domain of Hip

Tetratricopeptide repeat (TPR) domain is composed of TPR motif, which is composed of 34 amino acids (Lamb et al., 1995). Typically three successive TPR motifs and a C-terminal capping helix form a TPR domain. Specialized TPR domains recognizing the C-terminal EEVD motifs in Hsp70 and Hsp90 are present in their co-factors Hop, Chip, phosphoprotein phosphatase 5 (PP5), aryl hydrocarbon receptor interacting protein (AIP) and Hsp90-binding immunophilins Cyp-40, FK506-binding protein FKBP51 and FKBP52 (Ballinger et al., 1999; Buchner, 1999; Dolinski et al., 1998; Pratt and Toft, 1997; Scheufler et al., 2000).

Hip contains a predicted TPR domain between residues 113-214 (Höhfeld et al., 1995; Irmer and Höhfeld, 1997; Prapapanich et al., 1996b; Velten et al., 2002). Analysis of deletion constructs suggested that both the predicted TPR domain and the following positively charged region are required for efficient binding to the ATPase domain of Hsp70 (Prapapanich et al., 1996b). The TPR domain alone was not sufficient for binding to Hsp70 in the yeast two-hybrid system (Irmer and Höhfeld, 1997). The shortest construct displaying interaction with the ATPase domain of Hsp70 comprised residues 15-257 of Hip (Irmer and Höhfeld, 1997).

Truncation of the TPR domain from Hip unable it bind to Hsp70 (Prapapanich et al., 1996a), but its binding to CXCR2 was maintained and CXCR2-mediated mitogen-activated protein kinase activation can be preserved. However, overexpression of the TPR depletion mutant of Hip significantly attenuated the internalization of the chemokine receptors CXCR2 and CXCR4 (Fan et al., 2002).

2.3.2.4 The GGMP repeat region

Hip comprises seven imperfect Gly-Gly-(Met/Phe)-Pro (GGMP) tetrapeptide repeats between residues 278-311, and this motif is located close to the C-terminus of Hip together with an additional Sti1-related region (Höhfeld et al., 1995). It is highly conserved and is expanded to over 20 tandem repeats in a Plasmodium homolog of Hip (Uparanukraw et al., 1993). Similar repeats were also located in other chaperones, for example close to the peptide binding site of the C-terminus of Hsc70 as part of a regulatory motif (Höhfeld et al., 1995). Notably, inducible Hsp70 lacks this motif, as previously noted (Prapapanich et al., 1996a). It is interesting that Hip

combines structural elements found in Hsc70, Hsc70 and Hsp90-associated co-chaperones. Because of its proline-rich nature, it was predicted that neither α -helix nor β -sheet structure could be present in this region (Prapapanich et al., 1996a). The *in vitro* and *in vivo* studies suggested that this motif was not involved in regulating Hsp70 chaperone activity (Irmer and Höfled, 1997; Nollen et al., 2001). The exact function of the GGMP repeats in the interaction with Hsp70 or other cellular components is not yet understood.

2.3.2.5 The DP domain of Hip

The C-terminal region of Hip (residues 312-368) comprises two Asp-Pro (DP) repeat motifs with similarity to segments of the Hsp70-Hsp90 organizing protein (Hop) and Rad23. Little is known about how the DP region participates in co-chaperon function (Nelson et al., 2003).

The C-terminal domain of Hip was implicated in binding to the C-X-C-motif chemokine receptors type 2 and 4 (CXCR2 and CXCR4). Truncation of the C-terminal 66 amino acids completely abolished the interaction of Hip with CXCR2 (Fan et al., 2002), suggesting substrate binding ability of C-terminus of Hip. In addition, it was previously found that mutants that destabilized the DP domain can stabilize interaction with Hsp70, while inhibiting PR assembly (Prapapanich et al., 1998). Moreover, a recent study identified Hip as a novel substrate of G protein-coupled receptor kinase 5 (GRK5), which recognizes Hip by a motif within the DP domain, residues 303-319, and phosphorylates Hip at Ser346 *in vitro* and *in vivo* (Barker and Benovic, 2011). GRK5 had previously not been implicated in signaling and trafficking of CXCR4. Phosphorylation of Hip by GRK5 was shown to play an essential role in modulating CXCR4 internalization.

2.3.3 C-terminus of Hsc70 interacting protein (Chip)

Aberrant proteins or protein aggregates destined for degradation need to be labeled with a multi-ubiquitin chain. The ubiquitination process is mediated by the cooperation of three enzymes: activator E1, ubiquitin conjugating E2 and ubiquitin ligase E3 enzyme. C-terminus of Hsc70 interacting protein, Chip, a ~ 35 kDa protein, was the first known ubiquitin ligase that directly associated with a chaperone. Chip was initially identified in a screen for human TPR-

containing protein (Ballinger et al., 1999). Chip is a dimeric protein with subunits containing TPR and U-box domains at N- and C-terminus, respectively. The TPR domain in Chip enables the binding to the C-terminal EEVD motifs of Hsp70 and Hsp90, and the U-box domain participates in the ubiquitin conjugation. Chip was shown to associate with Hsp70, block the ATPase cycle of Hsp70 and inhibit its ability to refold non-native protein (Ballinger et al., 1999). Similarly, Chip may abolish the cooperation of Hsp90 with other co-chaperones needed for productive chaperone function. However, based on the modular structure of Chip, it may also enable a direct link between Hsp70 and Hsp90, ubiquitinate the clients and target them to proteasome for degradation.

Chip was shown by isothermal titration calorimetry (ITC) to have a six-fold higher affinity for Hsp90 than Hsp70 *in vitro* (Stankiewicz et al., 2010). The same study also suggested that Chip preferentially ubiquitinated Hsp70-bound substrates, such as luciferase (Stankiewicz et al., 2010). Chip was however also shown to promote the degradation of glucocorticoid hormone receptor (GR) by eliciting release of the regulatory cofactor p23 from the ternary complex (Connell et al., 2001). In line with this finding, Chip targeted the degradation of the ubiquitinated Hsp90-associated immature cystic-fibrosis transmembrane-conductance regulator (CFTR) (Meacham et al., 2001). Importantly, overexpression of the U-box truncation fragment in Chip blocked CFTR ubiquitination, indicating the essential function of U-box domain for targeting substrate degradation (Meacham et al., 2001). In addition to CFTR, Chip has been shown to be involved in degradation of mutant superoxide dismutase 1 (SOD1), which is implicated in protein misfolded disease (Urushitani et al., 2004). In addition, Chip was implicated in mammalian longevity through defective protein control. A Chip-knockout mouse model exhibited a markedly reduced life span, along with accelerated age-related pathophysiological phenotypes (Min et al., 2008). Taken together, Chip appears to play a crucial role in maintaining protein quality control in cells by modulating the balance of folding and degradation of chaperone substrates.

2.3.4 The Hsp70-Hsp90 organizing protein (Hop)

The yeast homolog to Hop, Sti1, was initially identified in a genetic screen for proteins that mediate the heat-shock response in *S. cerevisiae* (Nicolet and Craig, 1989). Subsequently, in

studies on the maturation of progesterone receptor (PR), mammalian Hop has been identified together with Hsp40, Hsp70 and Hip as an essential component for the recruitment of this steroid receptor to Hsp90 (Smith et al., 1993). Hop showed effective enhancement of the maturation of PR during *in vitro* reconstitution experiments. It is however not required for GR maturation in the same system (Morishima et al., 2000).

Hop is an abundant and highly conserved protein in eukaryotes. The homolog in *S. cerevisiae* is however not essential (Chang and Lindquist, 1994). Hop is a multi-domain adaptor protein that can bind Hsp70 and Hsp90 simultaneously. So far Hop has not been shown to act as a chaperone independently of Hsp70 and Hsp90 (Bose et al., 1996).

The various homologs of Hop are structurally characterized by the presence of three TPR domains, TPR1, TPR2A and TPR2B (residues 4-118, 223-352 and 353-477 in human Hop). The N-terminal TPR1 was shown to preferentially bind to the highly conserved EEVD motif at the C-terminus of Hsp70 (Scheufler et al., 2000). Interestingly, both TPR1 and TPR2B were identified to contribute to the Hsp70 interaction and they have redundant or overlapping functions *in vivo* (Flom et al., 2006). Conversely, TPR2A and TPR2B domains interact preferentially with the EEVD residues at the C-terminus of Hsp90 (Scheufler et al., 2000; Young et al., 1998). Notably, Hop contains two DP domains following TPR1 and TPR2B, called DP1 and DP2, respectively. Their sequences are 27 % identical and they form the arrangement in TPR1-DP1-TPR2A-TPR2B-DP2. Both of the two DP domains contribute to Hop functions (Carrigan et al., 2004; Chen et al., 1998), though one study has shown that DP2 domain is dispensable for the Hsp70 and Hsp90 interactions (Flom et al., 2007). Since Hop simultaneously interacts with Hsp70 and Hsp90 *via* separate TPR domains, Hop was postulated to be essential for integrating Hsp70-Hsp90 interactions. This was demonstrated by using purified Hsp70 and Hsp90s *in vitro*, which associate with each other only when Hop was present (Chen et al., 1996). Thereby, it has been proposed that Hop plays a crucial role in the transfer of client proteins from Hsp70 to Hsp90. Surprisingly, Hop was also reported as an ADP/ATP exchange factor in the recycling of Hsp70 system (Gross and Hessefort, 1996). In line with this study, the yeast homolog Sti1 was found to stimulate ATPase activity of yeast Hsp70 Ssa1 by a factor of about 200, indicating that Sti1 is a potent novel effector for the Hsp70 ATPase activity (Wegele et al., 2003). Furthermore, Sti1 binds to the N- and C-terminal part of Hsp90 and prevents the N-terminal dimerization reaction that is required for efficient ATP hydrolysis by Hsp90. Therefore, we can

surmise that Hop might work as a functional homolog to GrpE in the eukaryotic cytosol, promoting the dissociation of the substrate from Hsp70 for the efficient transfer to downstream Hsp90 chaperone system.

2.3.4.1 Similarities between Hip and Hop

Both Hip and Hop contain TPR domains, which mediate binding to chaperones. In addition, they also share DP motifs at their C-termini (Höhfeld et al., 1995; Nelson et al., 2003). Figure 11 shows a comparison of the domain structure. The first DP unit between Hip and Hop has the same sequence DPEV, but the second, DPAM, is slightly different in Hop. Interestingly, the combination of TPR and DP domains can only be found in Hip and Hop homologs. Besides sharing TPR and DP domains, both contain highly charged regions. Both the sequences at the end of the DP domain of Hop and between the TPR and DP domains of Hip show a preference for the residues such as D, E, K, Q, S and T.

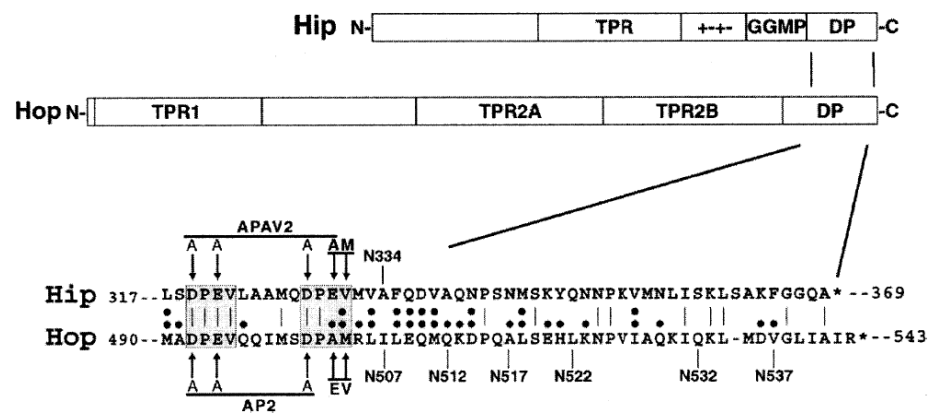


Figure 11: Diagram representation of Hip and Hop domains and shared C-terminal sequences.

Though the respective DP regions of Hip and Hop share sequence similarity, a functional defect in Hop was observed upon introduction of the double-point mutation DPAM to DPEV in the second DP motif of Hop, whereas the corresponding mutation in Hip maintains its function (Fig. 11) (Nelson et al., 2003). Therefore, they are not functionally equivalent. Moreover, both of them bind to Hsp70 but through distinct binding sites. For Hip binding, the N-terminal ATPase domain of Hsp70 is sufficient. Conversely, Hop recognizes the C-terminus of Hsc70

(Demand et al., 1998). Notably, both of Hip and Hop are involved in the steroid receptor complex maturation. In Hop, truncation of the entire DP region or DP point mutants blocked Hop binding to Hsp70 and assembly into the progesterone receptor (PR) complex (Chen and Smith, 1998). In Hip, DP mutants bind to Hsp70 more stably than wild type Hip, but block recruitment of Hop and Hsp90 to the receptor complex (Prapapanich et al., 1998).

2.4 Role of Hip in cellular protein quality control

Hip has been shown to be important for stabilizing Hsp70 in the ADP-bound state (Höhfeld et al., 1995). It was implicated in progesterone receptor (PR) pathway and presumably mediates substrate binding to Hsp70 in early receptor complexes (Prapapanich et al., 1996a). In collaboration with Hsp70, Hip prevents the formation of large cytotoxic aggregates of α -Synuclein and a polyQ-expanded mutant of androgen receptor (AR), hallmarks of Parkinson's disease and spinobulbar muscular atrophy (SBMA) (Kennedy's disease), respectively (Howarth et al., 2009; Roodveldt et al., 2009). Recently, Hip was also implicated in the ubiquitylation and proteasomal degradation of neuronal nitric oxide synthase and polyQ androgen receptor (Wang et al., 2013). Notably, a similar effect in promoting the degradation of polyQ androgen receptor and of hyper-phosphorylated Tau was achieved by the Hsp70-binding compounds MKT-077 and YM-1. These chemically related compounds stabilize Hsp70-substrate complexes and have been described as mimics of Hip function (Rousaki et al., 2011; Wang et al., 2013).

2.4.1 Regulation of progesterone receptor maturation

Steroid receptors are rapidly turned over in cells. In the absence of hormone, the receptor remains inactive. Upon hormone binding to the receptor, the complex disassembles, leading to release of the active receptor. The activation is mediated by intricate chaperone machinery in the cytosol. Hip was found to associate with Hsp70 in the ADP-bound state. It was suggested that Hip might contribute to the interaction of Hsc70 with various target proteins (Höhfeld et al., 1995). In addition, Hip was noted as a transient component during the cell-free assembly of progesterone receptor complex (Smith et al., 1995). Thereby, it was proposed to be an additional accessory factor paralleling Hsp70 in the assembly process. The identification of Hip as a

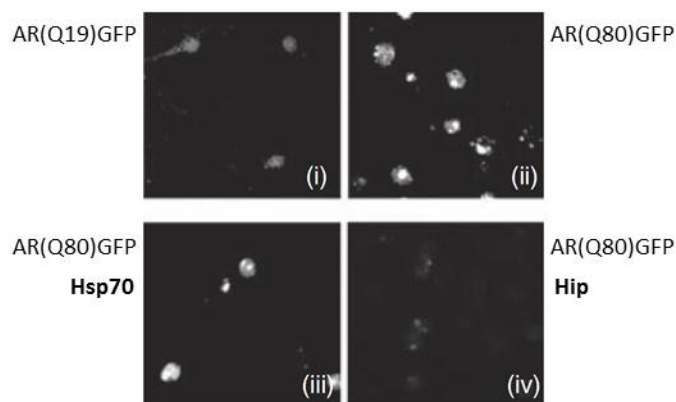
component of the hormone receptor complex provides the possibility to understand the interactions between Hip, Hsp70 and Hsp90 *in vivo*, since all of them are components of the same multi-protein complex (Smith, 1993; Smith et al., 1995). Hsp90 and its co-factors, which are essential for hormone binding and the activation of receptor, were suggested to stabilize the receptor in the proper hormone binding conformation.

Therefore, the presence of Hip in the progesterone receptor complex provides us with an example for the intracellular cooperation between the molecular chaperones. Especially, Hip might work as a signaling protein to link Hsp70 with downstream Hsp90.

2.4.2 Aggregation prevention in models of neurodegenerative disease

Protein misfolding and aberrant self-assembly may cause cell toxicity, resulting in protein conformational disorders and neurodegeneration. Many neurodegenerative diseases such as Alzheimer's (AD), Parkinson's (PD) and Huntington's diseases (HD) are characterized by the deposition of insoluble protein inclusions (Chiti and Dobson, 2006). The role of small molecular co-chaperones in facilitating the removal of protein inclusions has been studied (Jana and Nukina, 2005).

A)



B)

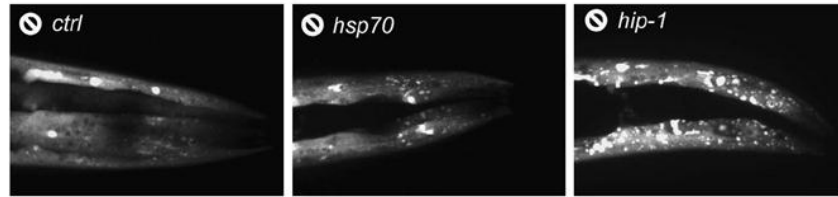


Figure 12: Suppression of protein deposit formation by Hip *in vivo*.

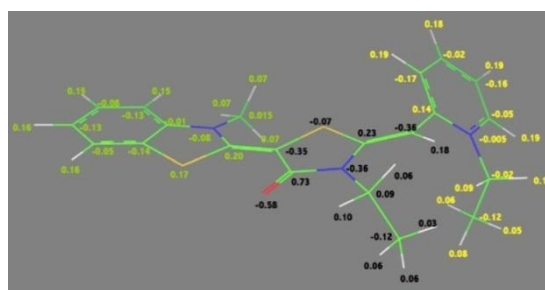
- A) Cortical neurons were transduced with vectors expressing AR(Q19)GFP and AR(Q80)GFP fusion proteins with or without presences of additional chaperones. (i) vectors expressing AR(Q19)GFP; (ii) vectors expressing AR(Q80)GFP; (iii) AR(Q80)GFP in presence of Hsp70; (iv) AR(Q80)GFP in presence of Hip (Howarth et al., 2009).
- B) α -Syn inclusion formation in the head region of transgenic *C. elegans* worms. α -Syn-YFP transgenic *C. elegans* were fed with bacteria expressing either empty vector (L4440), RNAi for knockdown of Hsp70 and knockdown of Hip. Quantification of the inclusion formation was performed in age-synchronized young adult *C. elegans*, after feeding RNAi bacteria for two generations (Roodveldt et al., 2009).

The effect of overexpressing Hip was initially investigated in a cellular polyglutamine (polyQ) model of spinal and bulbar muscular atrophy (SBMA) and Huntington's diseases (Howarth et al., 2009). Overexpression of Hip alone significantly reduced inclusion body formation (Fig. 12A). In addition, Parkinson's disease (PD) patients had consistently low Hip expression levels in their blood relative to healthy controls, even in the early stages of the disease (Scherzer et al., 2007). Intra-neuronal inclusions in the brains of PD patients known as Lewy bodies contain a large proportion of aggregated α -Synuclein. The effect of Hip on α -Synuclein aggregation was investigated *in vitro* and the *C. elegans* model system (Roodveldt et al., 2009). In the study, addition of Hip to Hsp70 in the presence of ATP efficiently suppressed α -Synuclein fibrils' formation *in vitro* and no fibrils can be observed under transmission electron microscopy (TEM). Importantly, this anti-amyloidogenic activity of Hip is mediated by Hsp70, as overexpression of Hip alone cannot decrease the formation of α -Synuclein fibrils. Depletion of Hsp70 by RNA interference caused only a non-significant increase of α -Synuclein-YFP inclusions in transgenic worms, whereas knockdown of Hip significantly caused increased inclusion formation, suggesting an essential function of Hip in suppressing α -Syn aggregation (Fig. 12B). It can be therefore postulated that depletion of Hip might impair the ability of proteins to resist aggregation and that Hip is required to maintain protein homeostasis.

2.4.3 Hip promotes degradation of misfolded protein

Hip preferentially binds to Hsp70 in the ADP-bound state. Interestingly, a small molecule functional homolog of Hip named MKT-077 was identified, which also binds to the ADP-bound state of Hsp70, but not ATP state (Rousaki et al., 2011).

A)



B)

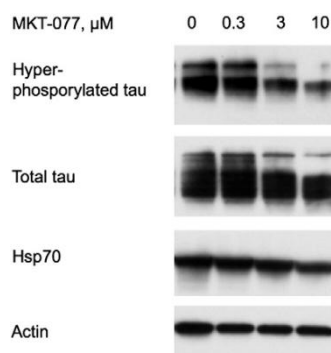


Figure 13: MKT-077, a compound mimicking Hip.

- A) The molecular structure of MKT-077, based on quantum mechanical calculations.
 B) MKT-077 promotes the clearance of hyper-phosphorylated tau in HeLa C3 cell line in a dose-dependent manner. SDS-PAGE analysis was performed after 24h incubation (Pitsis and Visouli, 2011).

MKT-077 is a rhodacyanine dye analog with the systematic name 1-ethyl-2-[[[3-ethyl-5-(3-methylbenzothiazolin-2-yliden)]-4-oxothiazolidin-2-ylidenemethyl] pyridinium chloride (Fig. 13A). MKT-077 has been identified initially as potent anticancer compound that interacted with mtHsp70 in pull-down studies (Deocaris et al., 2007; Wadhwa et al., 2000). Later MKT-077 was proposed to act as an “allosteric drug”, which blocks the cycle of cytosolic Hsp70 through stabilization of the ADP state (Rousaki et al., 2011). The K_D value for the complex of MKT-077 and the ADP-bound NBD of Hsp70 was estimated to be in the range of 1 – 10 μM by NMR titration. Selectivity towards the presumed binding site, a negatively charged pocket

between subdomains IA and IIA of Hsp70 NBD, might be driven by the cationic character of MKT-077 (Rousaki et al., 2011). This location is close to the nucleotide binding site, but not identical to it.

In preclinical studies, MKT-077 was found to inhibit proliferation of multiple human cancer cell lines, including colon, breast carcinoma cells and showed no toxicity against normal kidney cells (Koya et al., 1996). However, because of renal toxicity in the clinical Phase I trial against solid tumors, testing of MKT-077 was stopped.

One of the hallmarks of Alzheimer's disease is the accumulation of hyper-phosphorylated Tau in neurons besides extracellular amyloid- β deposits. Interestingly, MKT-077 promoted the clearance of hyper-phosphorylated Tau in a model of Alzheimer's disease, suggesting a potential link from Hip to the cellular degradation machinery (Fig. 13B) (Rousaki et al., 2011). Indeed, Hip overexpression facilitated ubiquitination and promoted proteasomal degradation of neuronal nitric oxide synthase (nNOS) and polyQ androgen receptor (AR) in cultured cells (Wang et al., 2013). A similar effect of increased ubiquitination and degradation of client proteins was observed with the compound YM-1, a stable and soluble derivative of MKT-077. As predicted, this compound also promoted binding of Hsp70 to denatured luciferase.

2.5 Aim of study

It had been suggested that Hsp70 interacting protein, Hip delays substrate release from Hsp70 by slowing the dissociation of Hsp70-bound ADP. However, the physiological role of Hip and its interplay with other Hsp70 co-factors were far from clear. Despite considerable achievements in understanding the Hip structure and its interaction with Hsp70, the exact three-dimensional structure of Hip remained a mystery. The exact and detailed structural mechanism by which Hip regulates Hsp70 was unknown. Such data could provide the basis for rational drug design and novel therapeutic interventions.

Based on these considerations, this work focuses on a detailed structural and functional analysis of Hip. Our aim was to determine the crystal structures of Hip and the Hip:Hsp70 complex. To achieve our goal, we first delineated the probable domain boundaries in rat Hip from sequence alignment, which guided the design of various Hip constructs. Functional analyses by isother-

mal titration calorimetry (ITC) and MABA-ADP dissociation assay identified the essential Hsp70 binding domain of Hip. With the information provided by the functional assays, the crystal structures of the middle domain of Hip (HipM) and the Hip:Hsp70 protein core complex were successfully determined. Subsequently, a detailed characterization of Hip:Hsp70 complex based on structure-guided site-directed mutagenesis of Hip was undertaken. These experiments characterized the Hip interface with Hsp70 and the structural requirement for the stabilization and regulation of Hsp70. Comparison of the Hip-Hsp70 and NEF-Hsp70 complex structures suggested Hip and NEF binding to Hsp70 are mutually exclusive. Thus, the association of Hip with Hsp70-substrate complexes attenuates their active cycling. Finally, the functional interaction between Hip and Hsp70 was probed *in vivo*, using the maturation of glucocorticoid receptor as a read-out.

In summary, the structural and biochemical analyses performed in this study were designed to provide a mechanistic model for the cooperation of Hip with Hsp70 in protein folding and aggregation prevention.

3 Materials and Methods

3.1 Chemicals

Unless otherwise stated, chemicals were of pro analysis quality and purchased from Sigma-Aldrich (Steinheim, Germany) or Merck (Darmstadt, Germany).

| Suppliers | Chemicals |
|--------------------------------------|---|
| Applied Biosystems (Foster, USA) | Gal-Screen chemiluminescence reagent |
| Biomol (Hamburg, Germany) | HEPES |
| Biozym (Hessisch Oldendorf, Germany) | Biozym LE Agarose |
| Clontech (Heidelberg, Germany) | Herring testis carrier DNA |
| Difco (Heidelberg, Germany) | Bacto Agar |
| | Bacto Tryptone |
| | Bacto peptone |
| | Bacto Yeast Extract |
| | Bacto Yeast Nitrogen Base |
| Fermentas (St. Leon-Rot, Germany) | GeneRuler 100 b und 1 kb DNA Ladder Page Ruler Protein Ladder |
| | PageRuler Prestained Protein Ladder |
| | Restriction enzyme <i>EheI</i> (Enzyme) |
| Fluka (Deisenhofen, Germany) | Polyethylene glycols of different molecular weights |
| | Sodium cacodylate |
| GE Healthcare (München, Germany) | Chloramphenicol |
| | MES |
| Hampton Research (Aliso Viejo, USA) | Index Screen |
| Interchim (Montluçon, France) | 8-((4-AMINO)BUTYL)-AMINO-ATP-MANT (MABA-ATP) |

| | |
|--|---|
| Invitrogen (Karlsruhe, Germany) | pProEx-HtA, pProEx-HtB protein marker for SDS-PAGE SYBR Safe DNA gel stain |
| J.M. Gabler Saliter GmbH & Co.KG (Obergingzburg, Germany) | Skimmed milk powder |
| Metabion (Martinsried, Germany) | dNTP Oligonucleotides (Primers) |
| New England Biolabs (Frankfurt am Main, Germany) | BSA Restriction endonucleases (Enzyme) T4 DNA ligase (Enzyme) |
| Promega (Mannheim, Germany) | <i>Pfu</i> polymerase (Enzyme) Wizard Plus SV Miniprep DNA Purification System Wizard SV Gel and PCR Clean-Up System |
| Qiagen (Hilden, Germany) | Qiagen Plasmid Midi Kit Qiagen Plasmid Maxi Kit |
| Roche (Basel, Switzerland) | DTT EDTA-free Complete protease inhibitor cocktail Hexokinase (Enzyme) Proteinase K (Enzyme) Shrimp alkaline phosphatase (Enzyme) |
| Roth (Karlsruhe, Germany) | Ampicillin IPTG KCl |
| Serva (Heidelberg, Germany) | Acrylamide-Bis Coomassie Blue R250 PMSF SDS |

Table 1: Chemicals

3.2 Enzyme and antibodies

| Suppliers | Chemicals |
|---|---|
| Fermentas (St. Leon-Rot, Germany) | Restriction enzyme <i>EheI</i> |
| New England Biolabs (Frankfurt am Main, Germany) | Restriction enzymes T4 ligase |
| Novagen /ToyoBo (Darmstadt, Germany) | KOD Hot Start DNA polymerasae |
| Promega (Mannheim, Germany) | <i>Pfu</i> polymerase T4 ligase |
| Department of Cellular Biochemistry, MPI (Martinsried, Germany) | Rat polyclonal Anti-Hip, second bleeding, dilution 1:1000 |
| Invitrogen (Karlsruhe, Germany) | Mouse polyclonal anti-PGK, dilution 1:5000 |
| Invitrogen (Karlsruhe, Germany) | Mouse monoclonal anti-Hsc70, dilution 1:2500 |
| Roche (Mannheim, Germany) | Mouse monoclonal anti-GFP, dilution 1:1000 |
| Sigma-Aldrich (Steinheim, Germany) | HRP-coupled secondary antibodies |

Table 2: Enzyme and antibodies

3.3 Strains

| Suppliers | Strains |
|---|--|
| Novagen (Darmstadt, Germany) | <i>E. coli</i> DH5 α |
| Stratagene (Heidelberg, Germany) | <i>E. coli</i> BL21 (DE3) |
| Stratagene (Heidelberg, Germany) | <i>E. coli</i> BL21 (DE3) codon plus RIL |
| M.B. Cox (University of Texas, El Paso, Texas, USA) | <i>S. cerevisiae</i> DSY-1100 |

Table 3: Strains

3.4 Instruments

| Suppliers | Reagents |
|---|---|
| Agilent Technologies (Santa Clara, USA) | Variable wavelength detector Agilent 1100 series |
| Applied Photophysics (Surrey, UK) | SX. 18V stopped-flow instrument |
| Bachofer (Reutlingen, Germany) | Hybridization oven |
| Beckman Coulter GmbH (Krefeld, Germany) | Centrifuges (GS-6R, Allegra-6R, Avanti J-25 with rotors JKA 10.500 and JA 25.50, J6-MI with rotor JS 4.2) DU 640 UV/VIS Spectrophotometer DU 800 UV/VIS Spectrophotometer Optima LE 80K ultracentrifuge with rotor 45 Ti |
| Bibby Scientific (Stone, UK) | Stuart rotating wheel |
| Biometra (Göttingen, Germany) | PCR thermocycler |
| Bio-Rad (München, Germany) | Chemidox XRS Horizontal agarose gel electrophoresis Mini Protean II electrophoresis cell Power Pac 300 (Wide) Mini-SUB CELL GT |
| Biospec (Bartlesville, USA) | Bead Beater |
| Eppendorf (Hamburg, Germany) | Centrifuges (5415D and 5417R) Pipettes (2, 10, 20, 100, 200, 1000 µl) Thermomixer comfort |
| Fischer Scientific (Schwerte, Germany) | Accumet Basic pH meter |
| Fuji/Raytest (Straubenhardt, Germany) | Fuji-LAS3000 luminescence and densitometry system with Image Reader LAS-3000 Gel imaging software: AIDA v.3.5.0 |
| GE-Healthcare (München, Germany) | Äkta FPLC Äkta Purifier/Explorer Electrophoresis Power Supply – EPS600 Ettan LC |

| | |
|--|--|
| | Pre-packed chromatography columns as following: HiTrap Chelating HP, HiTrap Blue HP, HiPrep 26/10 Desalting, HiLoad 16/60 Superdex 200 prep grade, MonoQ HR 10/16, Mono S HR 10/10, NAP-5 Sephadex G25 desalting, NAP-10 Sephadex G25 desalting, NAP-25 Sephadex G25 desalting |
| Hampton Research (Aliso Viejo, USA) | Mounted cryo loops Siliconized glass square cover slides (22 mm) VDX Plate with sealant |
| Hofer Scientific Instruments (San Francisco, USA) | SemiPhor blotting systems |
| Mettler Toledo (Gießen, Germany) | Balances (AG285, PB602) |
| MicroCal (Northampton, USA) | MicroCal VP-ITC MicroCalorimeter |
| Millipore (Schwalbach, Germany) | Centriprep concentrators (3000, 10.000, 30.000 and 50.000 Da MWCO) Steritop Filtration System (pore size 0.22 µm) |
| Misonix (Farmingdale, USA) | Sonicator 3000 |
| New Brunswick Scientific | Innova 44 incubator / shaker |
| PEQLAB (Erlangen, Germany) | Nanodrop 1000 |
| Roth (Karlsruhe, Germany) | ZelluTrans dialysis membrane |
| Scientific Industries, Inc. (Bohemia/NY, USA) | Vortex-Genie 2 |
| Whatman GmbH (Dassel, Germany) | Whatman Protran nitrocellulose membrane |
| Wissenschaftlich Technische Werkstätten (Weilheim, Germany) | pH meter |

Table 4: Instruments

3.5 Media and buffers

Media were prepared with deionized water and subsequently autoclaved. M9 minimal media was prepared with autoclaved water or sterile filtered ingredients.

| Media | Reagents |
|------------|---|
| LB medium: | 10 g/l Bacto peptone 5 g/l Bacto Yeast Extract 10 g/l NaCl Adjust pH 7.0 with 5 M NaOH (~ 0.7 ml 5 M NaOH/liter) (15 g/l agar for solid medium) |
| SC medium | 6.7 g/l Bacto Yeast Nitrogen Base 2 g/l drop-out mix Adjust pH 5.6 with 5 M KOH, autoclave 40 ml 50 % (w/v) glucose (sterile-filtered) If required, the following amino acids were added in, which were prepared in solution. 55.3 mg/l adenine (autoclaved) 62.9 mg/l L-histidine (autoclaved) 22.4 mg/l uracil (autoclaved) 81.7 mg/l L-tryptophan (sterile-filtered) 182.6 mg/l L-lysine (autoclaved) 219 mg/ml L-leucine (autoclaved) |
| SC plates: | 20 g/l Agar solution and SC medium (Bacto Yeast Nitrogen Base/drop out mix) were autoclaved separately, as the low pH of SC medium will cause acid hydrolysis of the agar. After autoclaving, the agar solution was mixed with SC medium. |

Table 5: Media

| Buffers | Reagents |
|-------------------------------|---|
| 6 x DNA-loading buffer | 0.25 % (w/v) bromophenol blue 0.25 % (w/v) xylene cyanol FF |
| Coomassie staining solution | 0.1 (w/v) Serva Coomassie Blue R250 40 % (v/v) Ethanol 10 % (v/v) Acetic acid |
| Coomassie destaining solution | 10 % (v/v) Ethanol 10 % (v/v) Acetic acid |
| 2 x SDS-loading buffer | 100 mM Tris-HCl pH 6.8 4 % (w/v) SDS 200 mM DTT 20 % (v/v) glycerol 0.2 % (w/v) bromphenolblue |
| 10 x SDS-running buffer | 3.02 % (w/v) Tris 14.4 % (w/v) glycine 1 % (w/v) SDS |
| Ponceau S stain | 0.2 % (w/v) Ponceau S 3 % (v/v) trichloro-acetic acid |
| 10 x PBS | 92 mM Na ₂ HPO ₄ •2H ₂ O 16 mM NaH ₂ PO ₄ •2H ₂ O 1.5 M NaCl Adjust pH 7.2 with NaOH |
| 1x PBS-T | 1 x PBS 0.1 % (v/v) Tween 20 |

| | |
|---------------------------|--|
| 2 x SDS-loading buffer | 100 mM Tris-HCl pH 6.8 200 mM DTT 4 % (w/v) SDS 0.2 % (w/v) Bromophenol blue 20 % glycerol |
| Stripping membrane buffer | 62.5 mM Tris-HCl pH 6.8 100 mM β -mercaptoethanol 2 % (w/v) SDS |
| 50 x TAE | 2 M Tris-acetate 50 mM EDTA pH 8.0 |
| 10 x TBS | 500 mM Tris-HCl pH 8.0 1.37 M NaCl 27 mM KCl |
| 1 x TBS-T | 50 mM Tris-HCl pH 8.0 137 mM NaCl 2.7 mM KCl 0.1 % (v/v) Tween 20 |
| Tfb I buffer (200 ml) | 30 mM K-Acetate 100 mM RbCl 10 mM CaCl ₂ 15 % (v/v) glycerol Adjust pH 5.8 with 0.1 M acetic acid Adjust final volume of 180 ml with deionized H ₂ O After autoclaving, 20 ml of sterile-filtered 500 mM MnCl ₂ •2H ₂ O was added |

| | |
|-----------------------------|---|
| Tfb II buffer | 10 mM MOPS 75 mM CaCl ₂ 10 mM RbCl 15 % glycerol Adjust pH 6.5 with 0.5 M NaOH, fresh sterile filtration |
| Western Blot buffer | 50 mM Tris 20 % (v/v) Methanol 1.441 % (w/v) glycine |
| Yeast alkaline lysis buffer | 0.2 M NaOH 1 % (w/v) SDS |
| Buffer A | 20 mM HEPES-KOH, pH 7.4 200 mM KCl 1 mM MgCl ₂ |
| Buffer B | 20 mM HEPES pH 7.4 200 mM KCl 1 mM MgCl ₂ 1 M imidazole |
| Buffer C | 25 mM HEPES-KOH pH 7.4 300 mM KCl 4 mM MgCl ₂ 5 mM β-mercaptoethanol |
| Buffer D | 25 mM HEPES pH 7.4 300 mM KCl 4 mM MgCl ₂ 1 M imidazole |

| | |
|-------------|--|
| Buffer E | 40 mM HEPES-KOH pH 7.4 500 mM KCl 20 mM MgCl ₂ 5 % (w/v) glycerol 2 mM β-mercaptoethanol 5 mM ATP |
| Buffer F | 40 mM HEPES pH 7.4 500 mM KCl 20 mM MgCl ₂ 1 M imidazole 5 % (w/v) glycerol 2 mM β-mercaptoethanol 5 mM ATP |
| Buffer BA | 20 mM KH ₂ PO ₄ -KOH pH 8.0 5 mM β-mercaptoethanol |
| Buffer BB | 20 mM KH ₂ PO ₄ -KOH pH 8.0 1 M NaCl 5 mM β-mercaptoethanol |
| Buffer CD | 10 mM NaKHPO ₄ -buffer pH 7.5 0.5 mM MgCl ₂ |
| Buffer HKM2 | 10 mM HEPES-KOH pH7.4 150 mM KCl 1 mM MgCl ₂ |
| Buffer GF | 20 mM HEPES-KOH pH7.4 100 mM KCl |

| | |
|-------------|--|
| Buffer GFM | 20 mM HEPES-KOH pH7.4 100 mM KCl 1 mM MgCl ₂ |
| Buffer PK | 20 mM Tris-HCl pH 8.0 100 mM NaCl 1 mM DTT |
| Buffer QA | 10 mM HEPES-KOH pH7.4 1 mM MgCl ₂ 10 mM KCl |
| Buffer QB | 10 mM HEPES-KOH pH7.4 1 mM MgCl ₂ 1 M KCl |
| Buffer SAXS | 20 mM HEPES-KOH pH7.4 100 mM KCl 1 mM DTT fresh addition |

Table 6: Buffers

3.6 Molecular biology methods

3.6.1 DNA analytical methods

DNA concentrations were determined by UV spectroscopy at a wavelength of 260 nm with a Nanodrop 1000 spectrophotometer (PEQLAB), where one absorption unit corresponds to 50 ng/ μ l double stranded DNA. DNA-nuclease free water served as a reference. The absorbance ratio 260/280 nm for pure DNA should be over 1.85. Deviations from this absorbance ration ratio indicate contaminations such as RNA or protein.

DNA electrophoresis was performed with 0.8 % (w/v) agarose gels submerged in 1 x TAE buffer. To prepare the gels, the agarose was dissolved in TAE buffer containing SYBR-SAFE

(1:5000). Prior to loading DNA samples on the gel, these were mixed with 6 x DNA loading buffer. Agarose gel electrophoresis was carried out at a constant voltage of 80 V for 20 minutes.

The authenticity of cloned plasmid constructs was confirmed by restriction digestion with suitable restriction enzymes before DNA sequencing by the Core Facility at the MPI of Biochemistry in Martinsried, Germany. Sequencing chromatograms were visualized by the program Chromas (C. McCarthy, Griffith University, Australia). Comparative sequence analysis was performed by using the MultAlin (<http://multalin.toulouse.inra.fr/multalin>) analysis online program (Corpet, 1988).

3.6.2 Polymerase chain reaction (PCR)

To amplify DNA target sequences from plasmid or annealed PCR oligonucleotides was used. Two kinds of Polymerases, KOD and *Pfu*, were used for the amplification of Hip constructs. The following standard protocols and cycling conditions were applied, respectively (Table 7 & Table 8).

| Component | Volume |
|-----------------------|--------------------|
| Template [100 ng] | 0.5 μ l |
| Primers [0.5 μ M] | 1.5 μ l (each) |
| KOD buffer 10 x | 5 μ l |
| MgSO ₄ | 3 μ l |
| dNTPs [2 mM] | 5 μ l |
| ddH ₂ O | 32.5 μ l |
| KOD Polymerase | 1 μ l |
| Total volume | 50 μ l |

Table 7: Standard PCR amplification protocol using KOD polymerase. For *Pfu* Polymerase, see Table 9.

| | Process | Temperature | Duration |
|--------|----------------------|--------------------|--------------------|
| Step 1 | Initial denaturation | 95 °C | 2 min |
| Step 2 | Cycle denaturation | 94 °C | 20 s |
| Step 3 | Primer annealing | 53 °C | 10 s |
| Step 4 | Primer extension | 70 °C | 1 kb/min |
| | | | Steps 2 – 4 (x 35) |
| Step 5 | Final extension | 70 °C | 5 min |
| Step 6 | Cooling | 4 °C | ∞ |

Table 8: Thermocycling conditions for PCR amplification.

3.6.3 Site-directed mutagenesis

For site-directed mutagenesis in existing plasmids, a whole-plasmid PCR was performed. A complete list of primer pairs used for site-directed mutagenesis introduced in this study can be found in the Appendix (Table 17). Table 9 and Table 10 list the standard protocol and thermocycling conditions. Modifications to the procedure were made when necessary. The success of the PCR amplification was checked by agarose gel electrophoresis. Afterwards, the product band was excised from the gel, dissolved and the DNA purified. 1 μ l *Dpn*I was added to 50 μ l PCR Clean-Up sample and incubated at 37 °C for 1 h. The *Dpn*I specifically digests methylated DNA, i.e. the template DNA produced in *E.coli*. Therefore, the parent DNA strands were selectively removed from the sample (Weiner et al., 1994). Finally, 5 μ l of the sample was directly transformed into *E. coli* DH5 α .

| Component | Volume |
|------------------------|--------------------|
| Template [100 ng] | 0.5 μ l |
| Primers [0.5 μ M] | 1.5 μ l (each) |
| <i>Pfu</i> buffer 10 x | 5 μ l |
| dNTPs [2 mM] | 5 μ l |
| ddH ₂ O | 28.5 μ l |
| <i>Pfu</i> Polymerase | 1 μ l |
| Total volume | 50 μ l |

Table 9: Standard PCR amplification protocol using *Pfu* polymerase.

| | Process | Temperature | Duration |
|--------|----------------------|--------------------|--------------------|
| Step 1 | Initial denaturation | 95 °C | 5 min |
| Step 2 | Cycle denaturation | 94 °C | 1 min |
| Step 3 | Primer annealing | 53 °C | 1 min |
| Step 4 | Primer extension | 70 °C | 1 kb/min |
| | | | Steps 2 – 4 (x 18) |
| Step 5 | Final extension | 70 °C | 5 min |
| Step 6 | Cooling | 4 °C | ∞ |

Table 10: Thermal cycling conditions for site-directed mutagenesis.

3.6.4 DNA restriction and ligation

The amplified PCR products or plasmids of interest were digested with the appropriate restriction enzymes at the respective restriction conditions as follows: 19.5 µl water, 5 µl plasmid (~ 0.5 µg), 3 µl of the appropriate buffer ± 0.5 µl BSA, 1 µl of each restriction enzyme in 30 µl total reaction volume; 50 µl PCR product, 5.6 µl of appropriate buffer ± 0.5 µl BSA, 1 µl of each restriction enzyme. Samples were incubated at 37 °C for 1 h. The mixtures were separated by agarose gel electrophoresis, and the target bands directly extracted with the PCR Clean-up System (Promega).

For ligation, approximately 50 ng digested plasmid DNA and a three-fold molar excess of insert DNA were mixed with 1 µl T4 DNA ligase and 1 µl T4 ligase buffer in a total volume of 10 µl. As a control reaction for re-ligation of incompletely restricted plasmid DNA, the same reaction without addition of insert DNA was carried out. The ligations were either performed for 1 h at room temperature or for 3 h at 18 °C. Afterwards, 5 µl of the ligation product was directly transformed into *E. coli* DH5a cells. The cells were subsequently spread on selective agar medium (AMP for pProEx) and incubated overnight at 37 °C (~ 15 h). Successful ligation was confirmed by restriction analysis and subsequent DNA sequencing.

3.6.5 Plasmid DNA and DNA fragment purification

E. coli DH5 α was used for the amplification of plasmid DNA. The transformed strains were grown in LB medium in presence of the appropriate antibiotics as selective markers for 8-16 h at 37 °C (Sambrook, 1989). Subsequently, plasmids were isolated by anion exchange chromatography using the Wizard Plus SV Miniprep DNA Purification System, according to the manufacturer's instructions, apart from reducing the washing step to 500 μ l Membrane Wash Solution in our protocol. For DNA Midiprep and Maxiprep, the Qiagen Plasmid Midi Kit and Qiagen Plasmid Maxi Kit were used, respectively.

To purify DNA fragments after agarose gel electrophoresis or enzymatic reactions (see section 3.6.1 and 3.6.4), the Wizard SV Gel and PCR Clean-Up system were used. Purification and isolation were performed according to the manufacturer's instructions.

3.6.6 Cloning strategies

For the present study, a complete list of plasmids constructed and primer pairs used can be found in the Appendix section 8.1, respectively. The expression vectors pProEx-HtA and pProEx-HtB were used for the heterologous expression of protein constructs in *E. coli*. Proteins were expressed with as an N-terminal His₆-tag fusion, which enables easy protein purification by immobilized metal-ion affinity chromatography (IMAC). The N-terminal His₆-tag was subsequently cleaved with TEV protease.

Cloning of rat Hip constructs: Apart from Hip(1-44) and Hip(312-368), which were cloned into the pETM30 plasmid with restriction sites *Nco*I and *Hind*III (Zhuo Li, Master thesis), the Hip constructs were generated by PCR using pProEx-HtB-Hip(1-368) as a template (from Dr. Andreas Bracher), *via* primers flanked with *Ehe*I/*Hind*III or *Ehe*I/*Kpn*I restriction sites. The trimmed amplicates were cloned into the expression plasmid pProEx-HtB (Invitrogen), which was cut with the same respective restriction sites. For generation of Hip mutants, whole plasmid site-directed mutagenesis was used to mutate the residues of interest in the respective plasmid *via* a PCR based strategy (see section 3.6.3) (Weiner et al., 1994). Primers are listed in Appendix 8.1 (Table 16 and 17).

Cloning of Hip fusion proteins: To construct Hip(78-234)-linker-Hsp70N and HipM-linker-Hsp70N fusion proteins, Hip(78-234) and Hip(78-247) (HipM) were amplified by PCR using primers introducing *EheI* and *EcoRI* restriction sites and cloned into plasmid pProEx-Hsp70N (plasmid stock, #796, Department of Cellular Biochemistry of MPI, Martinsried) between the restriction sites for *EheI* and *EcoRI*. The coding sequence for the artificial linker sequences GGS, GGSGGS and GAGGGSGGS between Hip and Hsp70 was constructed by nested PCR method. For construction of the reverse fusion protein Hsp70N-HipM, no artificial linker was introduced, since the N-terminal of Hip encodes an unstructured region, which might work as a flexible tether. *XhoI* restriction sites were introduced by PCR at the 3' end of Hsp70N and at the beginning of HipM. The *EcoRI/XhoI* and *XhoI/HindIII* digested PCR products were subsequently inserted into pProEx-HtA digested with *EcoRI* and *HindIII*. For generation of mutants of Hip fusion protein, site-directed mutagenesis with the appropriate primer pairs was employed *via* PCR (see section 3.6.3) (Weiner et al., 1994). Primers were listed in the Appendix 8.1 (Table 18).

Cloning of Hip constructs into yeast expression plasmids: For cloning the Hip constructs into the yeast expression plasmids p423GPD and p423ADH, *SpeI* and *EcoRI* restriction sites were introduced into the Hip constructs by PCR. Before cloning these Hip variants into the yeast plasmids, they were subcloned into a modified pProEx without the original *SpeI* site. For the Hip mutants, site-directed mutagenesis of this pProEx plasmid with the appropriate primer pairs was performed *via* PCR (Weiner et al., 1994). Primers are listed in the Appendix 8.1 (Table 19). All plasmid inserts and mutants were verified by DNA sequencing. Subsequently, the inserts were excised from the pProEx-HtB expression plasmid by digestion with *SpeI* and *EcoRI* and cloned into the yeast expression plasmids p423GPD and p423ADH (Mumberg et al., 1995), which were complementarily cleaved with *SpeI* and *EcoRI*. Amplification of the constructed yeast expression plasmids was performed in *E. coli* DH5 α .

3.6.7 Competent *E. coli* cell preparation and transformation

3.6.7.1 Chemocompetent *E. coli* cells and chemical transformation

For preparation of chemically competent *E. coli* cells, the RbCl method was utilized (Hanahan, 1983). A single colony of the respective *E. coli* strain was grown in 5 ml LB over-night culture.

200 ml of fresh Psi broth were inoculated with 2 ml of the overnight culture and grown to an OD₆₀₀ of 0.5 at 37 °C. Cells were then chilled on ice for 15 min and mixed frequently to speed up the cooling process. Afterwards, the cell suspensions were transferred into 50 ml-Falcon and centrifuged at 4 °C for 20 min at 3700 rpm. The supernatant was removed, and the cell pellets resuspended by gentle shaking in 40 ml freshly prepared, ice-chilled TfbI buffer. After incubation on ice for 15 min, the mixture was centrifuged again for 10 min at 3700 rpm at 4 °C. Following decantation of the supernatant, the harvested cells were placed on ice and resuspended in 8 ml ice-chilled TfbII buffer by gentle shaking during 15 min on ice. Subsequently, 50 µl aliquots of the cell suspension were pipetted into pre-cooled tubes (-20 °C), and immediately frozen in liquid nitrogen. Afterwards, the tubes were stored at -80 °C.

For transformation, 50 µl chemically competent *E. coli* cells were mixed with 50-100 ng plasmid DNA or 5 µl ligation mixture, and incubated on ice for 30 min. Subsequently, the cells were heat-shocked at 37 °C for 5 min and afterwards immediately cooled on ice for 1 min. 950 µl of LB were added to the cells and incubated for 1 h at 37 °C under gentle shaking (450 rpm). The cell suspension was then plated on plates with selective agar LB medium and incubated overnight at 37 °C (~ 15 h). Single colonies of transformants were picked and inoculated in liquid medium for further analysis.

3.6.7.2 Electrocompetent *E. coli* cells and electroporation

Electrocompetent *E. coli* cells were made by inoculating 500 ml of LB medium with 1 ml overnight culture and grown at 37 °C to OD₆₀₀ of 0.5. Subsequently, cells were chilled on ice for 15 min and centrifuged for 15 min at 4200 rpm (Beckmann centrifuge Avanti J-25 with rotor JS 4.2) at 4 °C. Followed supernatant decantation, cell pellets were resuspended in 100 ml cold sterile water followed by centrifugation as described above. The washing step was repeated and cells were resuspended in 50 ml pre-cooled sterile 10 % (v/v) glycerol. Centrifugation was carried out for 10 min at 3700 rpm (Beckmann centrifuge Avanti J-25 with rotor JA 25.50) at 4 °C and the cell pellet was resuspended in 1 ml of pre-cooled sterile 10 % (v/v) glycerol. Each 50 µl were aliquoted into pre-cooled tubes (-20 °C), flash-frozen in liquid nitrogen and stored at -80 °C.

Before the start of the electroporation, electroporation cuvettes were pre-cooled and the competent cells were thawed on ice. 5 μ l of the ligation mix or \sim 100 ng of plasmid DNA were transferred to the cells, and this mixture was subsequently transferred into the electroporation cuvette and pulsed (1250 V, 25 μ F, 200 Ω). 800 μ l LB-medium were immediately added into the cuvette after the signal tone. The mixture was subsequently transferred into an Eppendorf tube and incubated for 1 h at 37 $^{\circ}$ C with 450 rpm. Afterwards, the cells were pelleted for 1 min at 7000 rpm (Eppendorf 5415D). Cells were resuspended in \sim 100 μ l medium, plated on selective LB agar medium and incubated overnight at 37 $^{\circ}$ C (\sim 15 h).

3.6.8 Lithium acetate transformation of *S. cerevisiae* cells

All yeast transformations in this study were performed following to the lithium acetate (LiAc) method (Gietz et al., 1995; Schiestl and Gietz, 1989). A single colony of the *S. cerevisiae* strain DSY-1100 (Genotype MATa leu2-112, ura3-1, trp1-1, his3-11, 15 ade2-1, can1-100 GAL SUC2 carrying plasmids pDS-125 (GR/TRP) and pDS-362 (GRE-lacZ/URA)) was picked from a freshly streaked SC-Ade-Leu plate and inoculated into 5 ml SC-Ade-Leu medium as a start overnight culture, which was incubated at 30 $^{\circ}$ C with vigorous shaking 200 rpm. This culture was diluted with SC-Ade-Leu medium in a 200 ml flask to an OD₆₀₀ of \sim 0.15 in a total volume of 50 ml. Cells were harvested at OD₆₀₀ of 0.5 by centrifugation for 5 min with a speed of 3700 rpm at 4 $^{\circ}$ C. The yeast pellet was suspended in 25 ml autoclaved water and centrifuged again. The resulting cell pellet was washed with 1 ml 100 mM LiAc and transferred into a 1.5 ml Eppendorf tube, which was centrifuged for 15 s with top speed. The cell pellet was resuspended in 500 μ l 100 mM LiAc and chilled on ice. For each transformation, 50 μ l cells were needed. Cells were centrifuged at 2000 rpm for 1 minute. After removal of the LiAc solution, the following components were added sequentially: 240 μ l sterile-filtered 50 % PEG3350, 36 μ l 1 M LiAc, 5 μ l 10 mg/ml herring testes carrier DNA, 60 μ l autoclaved water and 5 μ l plasmid DNA (p423GPD-Hip or p423ADH-Hip) at a concentration of \sim 100 ng/ μ l. Each tube was vigorously vortexed at room temperature until all the components were mixed well, then incubated in a shaker at 600 rpm and 30 $^{\circ}$ C for 30 min. Cells were then heat-shocked at 42 $^{\circ}$ C for 20 min and centrifuged at 8000 rpm for 30 s. Afterwards, the cell pellet was resuspended in 1 ml SC medium and incubated at 30 $^{\circ}$ C and 600 rpm. After 2 h, the cells were harvested by centrifugation at

8000 rpm for 1 min at room temperature and resuspended in 100 μ l of appropriate selective medium for plating on SC-Ade-Leu-His medium. These plates were then incubated for maximal two days at 30 $^{\circ}$ C.

3.7 Protein biochemical and biophysical methods

3.7.1 Protein expression and purification

For protein overexpression in a BL21 codon plus strain, 1 L of LB medium containing suitable antibiotics was directly inoculated with \sim 1 ml of the transformation mixture grown overnight at 33 $^{\circ}$ C with 180 rpm of gentle shaking until next morning (\sim 15 h), following incubation, the cells were further grown at 37 $^{\circ}$ C with 200 rpm to OD₆₀₀ of 0.7.

Protein purifications were performed at 4 $^{\circ}$ C unless stated. All the steps of purification were monitored by SDS-PAGE, and protein concentrations were measured by Bradford assay and A₂₈₀.

3.7.1.1 Hip constructs

Firstly, constructs Hip(1-368), Hip(1-267), Hip(78-267) and Hip(78-368) were cloned by Dr. Andreas Bracher. Genes encoding Hip(1-44) and Hip(312-368) were cloned into bacterial expression vector pETM30 (Zhuo Li, Master thesis) and the other constructs Hip(45-267), Hip(45-368), Hip(78-234), Hip(78-247) and Hip(107-267) were cloned into pProEx-HtB containing an N-terminal 6x His tag. The proteins were expressed in the *E.coli* strain BL21 (DE3) codon Plus RIL (Novagen) at 37 $^{\circ}$ C in LB medium containing 0.1 mg/ml AMP (ampicillin for pProEx) or 0.025 mg/ml Kan (kanamycin for pETM30) and 0.034 mg/ml CM (chloramphenicol). The cultures were grown at 37 $^{\circ}$ C until OD₆₀₀ of 0.7 and expression was induced with 0.5 mM IPTG at 37 $^{\circ}$ C for 3 h (Hip(1-44) and Hip(312-368)) or at 18 $^{\circ}$ C for 16 h (all other constructs). The harvested cells were removed by centrifugation at 4000 rpm (Beckman Coulter J6-Micentrifuge with JS 4.2 rotor) and 4 $^{\circ}$ C for 25 min. Each 1 L cell pellet was resuspended in 20 ml cell lysis buffer A and supplemented with 1 mM PMSF. Subsequently the cells were mechanically disrupted by ultrasonication with a Misonix Sonicator 3000 cell disruptor. The cells were placed in an ice bath and treated with 12 cycles of 25 s pulsing with force 6.0 and 95 s

chilling. Cell debris was clarified by centrifugation at a speed 20000 rpm (Beckmann centrifuge Avanti J-25 with JA 25.50 rotor) and 4 °C for 40 min. Supernatant was loaded onto a Ni-Chelating Sepharose (GE Healthcare), which was equilibrated with 1 % buffer B (dilution with buffer A) (Table 6). The bound protein was eluted with an imidazole gradient of increasing buffer B content, and fractions containing protein were pooled. For cleavage of the eluted His-tagged protein, the protein was incubated with TEV protease at a mass ratio of 1:100 in the presence of 1 mM DTT and 0.5 mM EDTA at 4 °C overnight. The uncleaved His-tagged protein and TEV protease were removed on Ni-Chelating Sepharose (GE Healthcare). Before loading on Ni-Chelating Sepharose, the imidazole-containing buffer was removed by buffer exchange against buffer HKM2 (Table 6) using a HiPrep 26/10 Desalting column (GE Healthcare). If necessary, TEV-cleaved protein in flow-through fractions was further purified by additional anion-exchange chromatography (MonoQ, GE Healthcare) using a linear gradient of 0.01 to 1 M KCl mixed from buffers QA and QB (Table 6). Finally the proteins were purified by size exclusion chromatography on the Superdex 200 (GE Healthcare) using the buffer GF (Table 6) as equilibration and running buffer. The purity and molecular mass were verified by SDS-PAGE.

3.7.1.2 Hip fusion constructs

Hip-Hsp70 fusion proteins were expressed and purified similar to Hip variants. The modifications are described in the following. Overnight expression (~ 16 h) of the fusion proteins at 18 °C was induced with 0.8 mM IPTG. Harvested cell pellets from 2 L were resuspended in 40 ml 1 % buffer D (diluted with buffer C) (Table 6), supplemented with 1 mM PMSF and 1 x Complete protease inhibitor cocktail. The His₆-tagged fusion protein was eluted using an imidazole gradient of increasing buffer D. In the size exclusion chromatography on the Superdex 200 (GE Healthcare), the running buffer was GF (Table 6) containing 1 mM DTT. The protein was directly pooled and concentrated to 16-20 mg/ml. Aliquots of this material were flash-frozen in liquid nitrogen and stored at - 80 °C.

3.7.1.3 Hsp70 homologs

All the full-length human Hsp70 constructs (wild type, Hsp70(D199A) and Hsp70(T204A)) and Hsp70 NBD were expressed as N-terminally His₆-tagged proteins in *E. coli* BL21(DE3)-Codon Plus RIL cells. All the strains were grown at 37 °C in LB medium containing 0.1 mg/ml ampicillin and 0.034 mg/ml chloramphenicol. At an OD₆₀₀ of 0.7, full-length proteins and Hsp70 NBD were induced with 0.2 mM IPTG and 0.8 mM IPTG, respectively. Cells were subsequently grown at 18 °C for 16 h. Cells were harvested by centrifugation at 4 °C and 4200 rpm for 25 min. The pellets were suspended in 1 % buffer F (diluted with buffer E) for full-length Hsp70 variants or 1 % buffer D (diluted with buffer C) for Hsp70 NBD containing 1 mM PMSF and Complete protease inhibitor cocktail and lysed by ultrasonication as described above in 3.7.1.1. After 30 minutes centrifugation at 4 °C with 20000 rpm, the supernatant will be loaded onto Ni-HiTrap Chelating Sepharose (GE Healthcare), which was pre-equilibrated with 50 ml 1 % buffer F (full-length Hsp70) or 1 % buffer D (Hsp70 NBD) containing 10 mM imidazole. The bound protein was eluted with a linear gradient of imidazole. The fractions containing highly concentrated protein were pooled together. For cleavage of His₆-tag, the pool of protein was incubated with His₆-TEV protease at a molar ratio of 1:100 in the presence of 1 mM DTT and 0.5 mM EDTA at 4 °C overnight. To remove the residual cleaved His₆-tagged protein and TEV protease, the proteins were first transferred into HKM2 buffer (Table 6) and then passed onto Ni-HiTrap Chelating Sepharose again, which was equilibrated with 2 % buffer F (full-length Hsp70) or 2 % buffer D (Hsp70 NBD) containing 20 mM imidazole. The flow-through containing the protein without His₆-tag was pooled. After the Ni-affinity chromatography, anion exchange MonoQ column chromatography was performed. Before loading onto MonoQ (GE Healthcare), proteins were changed into buffer QA containing low salt by using HiPrep 26/10 Desalting column. When necessary, size exclusion chromatography on Superdex 200 using buffer GF (Table 6) followed as the final purification step. Purified proteins were flash-frozen in liquid nitrogen and stored at - 80 °C.

To remove the bound nucleotides from the full-length Hsp70, after 2nd Ni-affinity chromatography, the protein was incubated with 50 mM EDTA on a rotation plate at room temperature for 1 h. Subsequently the nucleotide was partially removed by the size exclusion chromatography on Superdex 200 (GE Healthcare) with running buffer GF. After Superdex 200, Hsp70 was incubated with Calf Intestinal Alkaline Phosphatase (CIP, New England Biolabs) at a ratio

of 3 U/mg Hsp70 at room temperature for 3 h and then incubated on ice overnight. CIP phosphatase and reaction products were removed by size exclusion chromatography on Superdex 200.

Another effective method to remove nucleotide of Hsp70 NBD is using the HiTrap Blue Sepharose column (GE Healthcare), which was described in a previous study (Arakawa et al., 2011). After Hsp70N was purified with anion exchange chromatography, protein buffer was exchanged to buffer BA (Table 6) using HiPrep 26/10 Desalting column (GE Healthcare). Protein was loaded onto a 5 ml the HiTrap Blue Sepharose which was equilibrated with buffer BA, and eluted with buffer BB containing 1 M NaCl salt gradient in 0.5 ml/min flow-rate (Table 6).

3.7.2 Protein analytical methods

3.7.2.1 Protein quantification

Protein concentrations were determined either colorimetrically using the Bradford assay or spectrophotometrically by measuring the absorption at 280 nm. Samples of cell lysate or protein mixtures were quantified by the Bradford assay according to the manufacturer's instructions (Bradford 1976). 1-5 μ l protein were added to 1 ml of a 1:4 dilution Bio-Rad protein assay reagent and the absorbance at 595 nm measured with a DU640 photometer (Beckman Coulter) against the reagent background. Concentrations of pure proteins were always determined by A_{280} using a Nanodrop 1000 (PEQLAB). As a reference, protein-free buffer was used. Concentration was determined with the theoretical molar extinction coefficient ($M^{-1} cm^{-1}$), which was calculated based on the protein sequence with the online program ExPASy ProtParam (<http://web.expasy.org/protparam>).

3.7.2.2 SDS-PAGE

To separate and analyze protein mixtures based on molecular weight, sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) was used (Laemmli, 1970). Protein analysis was performed under denaturing and reducing conditions. Gels were prepared as described in Table 11 and water was layered over the resolving gel while the gel matrix polymerized. Protein samples were mixed with 2 x SDS-sample buffer. Prior to loading, samples were heated for

5 min at 95 °C and spun down shortly at maximum speed in a table centrifuge (Eppendorf 5415D). Gel electrophoresis was completed in Bio-Rad Mini-Protean 2 electrophoresis chambers filled with SDS-PAGE running buffer employing a constant voltage of 200 V for 45 min.

| Solutions (for 4 gels) | Resolving gel | | | Stacking gel |
|---------------------------|---------------|--------|---------|--------------|
| | 10 % | 12 % | 15 % | |
| deionized water | 7.9 ml | 6.6 ml | 4.6 ml | 2.7 ml |
| 1.5 M Tris-HCl pH 8.8 | 5 ml | 5 ml | 5 ml | — |
| 1.0 M Tris-HCl pH 6.8 | — | — | — | 0.5 ml |
| 30 % Acrylamide | 6.7 ml | 8.0 ml | 10.0 ml | 0.67 ml |
| 10 % (w/v) SDS | 200 µl | 200 µl | 200 µl | 40 µl |
| 10 % (w/v) APS | 200 µl | 200 µl | 200 µl | 40 µl |
| TEMED | 8 µl | 8 µl | 8 µl | 4 µl |

Table 11: Composition of resolving and stacking gels for SDS-PAGE.

3.7.2.3 Coomassie blue staining of SDS-PAGE

Coomassie blue staining was used for the visualization of protein bands in SDS-PAGE. Coomassie blue dye is a triphenylmethane coloring agent that binds to proteins. The dye integrates with cationic, nonpolar and hydrophobic side chains of a polypeptide. Protein bands were fixed and stained in the gel matrix by immersion into freshly prepared Coomassie blue staining solution, followed by gentle shaking for 20 min. The background destaining was performed by incubation with Coomassie destaining solution for at least 1 h at RT under gentle shaking.

3.7.2.4 Western blotting and immunodetection

Proteins were separated on SDS-PAGE and subsequently proteins were transferred from the gel onto nitrocellulose membranes using a semi-dry western blot system (SemiPhor). Transfer was performed in Western blot buffer and accomplished by employing a constant current of 80 mA for 75 min for one or two gels. Transfer efficiency was determined by staining with Ponceau S solution for 2-3 min, followed by water destaining to remove background. Membranes were afterwards immediately subjected to immunodetection.

Nitrocellulose membranes were blocked in 3-5 % milk powder in PBST/TBST solution for 1 h at RT. The primary antibody was diluted in blocking solution and incubated with the membrane for 2 h at room temperature or at 4 °C overnight. The membranes were washed for 3-4 times with TBST solution, each time 10 min long. After washing, the membrane was incubated with HRP-conjugated (horseradish peroxidase) secondary antibody at a dilution of 1:5000 in TBST for 1 h at RT. The membranes were washed again as above and the bound HRP-coupled antibodies were detected by enhanced chemiluminescence (ECL) staining using a 1:1 ratio of Rodeo ECL detection reagents 1 and 2. The detection reagent should completely cover the membranes. Chemiluminescence was detected and documented with the Fuji-LAS3000 luminescence and densitometry system. When necessary, membranes were stripped with stripping buffer at 70 °C for 45 min.

3.7.2.5 Circular dichroism spectroscopy (CD)

The secondary structure of purified Hip variants was determined by CD spectroscopy. Proteins were exchanged into buffer CD by using Superdex 200 (Table 6). The CD spectra were measured with a Jasco-715 spectrometer equipped with a Peltier-thermostat. 200 µl of each protein sample with concentration of ~ 0.1 mg/ml or 2.5 µM was analyzed in 0.1 cm glass cuvettes at 4 °C. Wavelength scans ranging from 250-195 nm were recorded four times and averaged, followed by correction with the analysis buffer baseline. All the spectra were fitted with the programs CDPro analysis and CONTIN to estimate the secondary structure content.

3.7.2.6 Small angle X-ray scattering (SAXS)

Samples of Hip constructs for SAXS measurement were applied to an analytical Superdex 200 column operated at 18 °C in buffer SAXS (see Table 6). Three different concentrations: ~ 2, 5 and 10 mg/ml were prepared. After concentrations were accurately measured with the Nanodrop photometer (2 µl loading), 1 mM fresh DTT was added to each sample. The SAXS experiments were performed with a monochromatic X-ray beam of 0.931 Å wavelength at ESRF beamline ID14-3. As a reference, a BSA sample was measured. For each Hip sample, ten measurements of 10 s exposures were recorded and averaged. The buffer background scattering curve was subtracted. The program Primus was used to process the protein scattering data

(Konarev et al., 2003). Radii of gyration (R_g) were determined using the Guinier approximation. Scattering curves were fitted with GNOM (Svergun, 1992).

3.7.2.7 Multi-angle light scattering (MALS)

For determination of molecular mass and hydrodynamic radius of Hip fusion proteins, size exclusion chromatography/multi-angle light scattering (MALS) was performed by Dr. Manajit Hayer-Hartl. 80 μ l of each Hsp70N-HipM and HipM-Hsp70N were respectively injected on a gel filtration column using a 100 μ l injection loop. The column was developed at a flow rate 0.2 ml/min. The GFM buffer used as running buffer contained 1 mM fresh DTT. The system was connected to DAWN EOS MALS detector (960 nm laser), variable wavelength detector (absorbance at 280 nm, Agilent 1100 series), and Optilab DSP refractive index detector (690 nm). Masses and molecular diameters were calculated using the ASTRA software with the dn/dc value set to 0.185 mg/g.

3.7.3 Protein crystallization and structure determination

3.7.3.1 Crystallization

Before setting up the crystallization screens, particles and precipitated protein were first removed from the protein solution by centrifugation with 12000 rpm at 4 $^{\circ}$ C for 40 min. Initial crystal screens were conducted at the Crystallization Facility of the MPI of Biochemistry, Martinsried. In the initial screening, the sitting drop vapor diffusion method was used mixing 100 nl protein solution and 100 nl reservoir solution.

The precipitant kits Hampton Research Index, Qiagen Classics, Qiagen pH Clear 1 and Qiagen pH Clear 2 were used for initial screenings at 18 $^{\circ}$ C for Hip(78-247), named HipM in the following. Initial crystals of HipM were obtained from the Hampton Research Index and Qiagen pH Clear 1 screens with 1.4 M tri-sodium citrate, 0.1 M HEPES pH 7.5 and 2.4 M Na-malonate pH 7.0. Crystallization conditions were optimized by systematically varying the precipitant concentration. To get larger crystals, 1 μ l HipM at the same concentration, 18 mg/ml, was mixed with 1 μ l of reservoir solution and equilibrated against 500 μ l of reservoir solution at 18 $^{\circ}$ C using the hanging drop vapor diffusion method. Crystal grew in clusters of thin plates.

Optimization of the tri-sodium citrate condition was not successful. With the Na-malonate pH 7.0 condition, single crystals can hardly be achieved. Thin plates were only observed after several rounds of micro-seeding. For this crystal form, experimental phasing using heavy atom soaks failed. Growing similar crystals of SeMet-substituted HipM also failed. This protein formed spheroids, and single crystal could not be obtained by seeding. However, a second crystal form was found at pH 6.0 with sodium malonate as precipitant. Single, cube-shaped tetragonal crystals were obtained after three weeks in presence of 2.3 M Na-malonate pH 6.0 at 18 °C. For cryo-protection of the crystals, they were transferred in two steps into a cryo-solution. First they were mixed with a 1:1 mixture of mother liquor and cryo-solution for 15 min. The mother liquor contained 2.5 M Na-malonate pH 6.0 and the cryo-solution with additionally 15 % glycerol. After equilibration, they were transferred to 100 % cryo-solution and incubated for 15 min, followed by flash-cooling in liquid nitrogen.

To crystallize the Hsp70N-HipM fusion protein, the precipitant kits Qiagen PEGs, Hampton Research Index and Qiagen Classics Lite were used for initial screening at 4 °C and 18 °C. Initial crystals were obtained from Qiagen PEGs screening kit with 0.2 M NH₄I and 20 % PEG 3350 at 4 °C and 18 °C. Improved crystals were achieved by hanging drop vapor diffusion method after micro-seeding in presence of 0.2 M NH₄I and 17 % PEG 3350 at 18 °C. For cryo-protection, the original drop was first mixed with a 1:1 mixture of mother liquor and cryo-solution for 15 min. The mother liquor was 0.2 M NH₄I, 20 % PEG3350 and cryo-solution with additional 15 % glycerol. Finally, they were transferred to 100 % cryo-solution and incubated for 15 min, followed by flash-cooling in liquid nitrogen.

3.7.3.2 Data collection, structure solution and refinement

Diffraction data were collected at beamlines ID14-4, ID23-1, ID23-2 and ID29 of ESRF Grenoble, France. The data collection strategy was devised with iMosflm from two wedges of reflection images 90 ° apart (Battye et al., 2011). The images were integrated with XDS (Kabsch, 2010) and converted to CCP4 format with Pointless (Evans, 2006). SCALA (Evans, 1997) was utilized for scaling as implemented in the CCP4i interface (Collaborative Computational Project, 1994).

HipM crystal form I (P4₃2₁2): The native dataset was from a crystal soaked with hydroxyl-mercuribenzoic acid. The mercury ligand was however not bound or disordered in the structure. The form I structure of HipM was initially determined by single isomorphous replacement with anomalous scattering (SIRAS) from this native dataset and derivative dataset from a K₂[OsO₄]-soaked crystal by direct methods using Shelx-CDE as implemented in HKL2MAP at 5.6 Å resolution (Schneider and Sheldrick, 2002). SHARP was used to refine the three heavy atom sites and for calculating initial phases (Vonrhein et al., 2007). Phase extension and density modification was performed with Resolve, resulting in an interpretable electron density map (Terwilliger, 2000). The unit cell contained two copies of HipM, which were symmetrically associated *via* C-terminal interaction with the opposing neighbor molecule. Density for the residues 78-93 and 213-247 was not discernible. The respective portions of the protein were either disordered or cleaved off in both copies. The final model was refined against native dataset of 2.6 Å resolution.

HipM crystal form II (P2₁): The diffraction data was initially integrated in the orthorhombic space group C222₁ with lattice parameters of 68.9, 92.4 and 126.6 Å. This structure was solved by molecular replacement with the HipM fragment coordinates from the Hsp70N-HipM fusion protein using Molrep (Vagin and Isupov, 2001). However, refinement of the model failed to lower the R-factors. The actual space group turned out to be P2₁ with quite similar lattice parameters a and c with 57.29 Å and 57.31 Å, respectively. The crystals were twinned and each unit cell contained four copies of HipM. Residues 82-241 were ordered in all chains.

Hsp70N-HipM fusion protein: This fusion protein was solved by molecular replacement using Molrep with the Hsp70(1-393)-ADP complex and the HipM monomer from crystal form I as search templates (Vagin and Isupov, 2001). Residues 1-382 of human Hsp70 and residues 78-243 of rat Hip were defined in the density as well as ADP-Mg²⁺ and 17 iodine ions. The positions of 17 resolved iodine ions from the precipitant solution were confirmed from the anomalous signal in the dataset collected from another crystal at 2.0 Å wavelength.

All of the models were built interactively using Coot (Emsley and Cowtan, 2004). Refinement was performed with REFMAC5 (Murshudov et al., 1997). Non-glycine residues without detectable side chain density facing solvent channels were modeled as alanines. Together with the

previous solved dimerization domain of Hip, all of the atomic coordinates and structure factors of HipM and Hsp70N-HipM fusion protein have been deposited in the Protein Data Bank under the accession code of 4J8C (HipN), 4J8E (HipM, crystal form I), 4J8D (HipM, crystal form II) and 4J8F (Hsp70N-HipM). Structural figures were generated with PyMOL (Schrödinger, 2010).

3.7.4 Structure analysis

Coordinates were aligned with Lsqman (Kleywegt and Jones, 1994). The sequence alignment was prepared with ClustalW (Larkin et al., 2007; Thompson et al., 1994) and ESPript (Gouet et al., 1999). The interaction surface in the Hip:Hsp70 complex was analyzed with Areaimol (Lee and Richards, 1971), Contact (Skarzynski, 1988), Naccess (Hubbard and Thornton, 1993). Surface shape complementarity between Hip and Hsp70 complex was estimated using SC (Lawrence and Colman, 1993).

3.7.5 Functional characterization in *vitro*

3.7.5.1 Limited proteolysis with Proteinase K

In order to determine stably folded protein fragments, protein samples were subjected to Proteinase K treatment. Aliquots of 6 μ M purified protein were incubated with a series of Proteinase K concentrations from 0.4 to 2.56 μ g/ml on ice for 1 h. The reaction was then quenched by addition of 100 mM PMSF to a final concentration of 4 mM PMSF, followed by 5 minutes incubation on ice. To remove insoluble PMSF, the samples were centrifuged for 1 min. 10 μ l supernatant were mixed with 10 μ l 2 x SDS loading buffer and heated to 95 $^{\circ}$ C for 5 min. The samples were analyzed by SDS-PAGE on 15 % acrylamide gels after Coomassie blue staining.

3.7.5.2 Isothermal titration calorimetry (ITC)

To measure the binding affinity of Hip to Hsp70 under various conditions, ITC experiments were performed with a MicroCal VP-ITC at 22 $^{\circ}$ C. All of the reactant solutions were dialyzed against 20 mM HEPES-KOH pH 7.4 and 100 mM KCl buffer at 4 $^{\circ}$ C overnight. The samples and some dialysis buffer were centrifuged at 12,000 rpm and 20 $^{\circ}$ C for degassing and removal

of particles. The sample cell chamber was rinsed with degassed buffer, then around 1.8 ml Hsp70 or Hsp70N (164 – 214 μM) was carefully injected into the chamber using a syringe with a long needle until liquid became visible in the aperture. Around 320 μl Hip variant concentrate (1.38 – 1.76 mM) sample were loaded into the injection syringe so that no air bubbles were trapped inside the syringe. During each injection, an 8 μl portion of Hip protein from the syringe was added into the calorimeter chamber containing Hsp70. Injections were applied at 180 s intervals. The data were evaluated with MicroCal Origin software (v7.0), assuming independent binding sites.

3.7.5.3 Nucleotide release assay

To determine the effect of Hip on nucleotide release from Hsp70 and its competition with NEFs, we employed the fluorescent nucleotide analog MABA-ADP, which was enzymatically prepared from MABA-ATP (8-[(4-(N-methyl-anthraniloyl)-amino)butyl]-amino-adenosine-5'-triphosphate, sodium salt, Interchim). The assay was performed with a SX.18V stopped-flow instrument (Applied Photophysics, Surrey, UK) as previously described (Gässler et al., 2001). All components were in GFM buffer containing additional 1 mM DTT. MABA-ADP complexes were formed by mixing 2.5 μM Hsp70N, Hsp70 or an Hsp70N fusion protein with an equimolar concentration of MABA-ADP, followed by incubation at 30 $^{\circ}\text{C}$ for 30 min. The nucleotide exchange solutions contained 250 μM ATP and protein factors as indicated. To determine the nucleotide release rates, equal volumes of the solutions were mixed at 30 $^{\circ}\text{C}$ in the stopped-flow apparatus. The release of nucleotide MABA-ADP fluorescence was monitored via the time-dependent decrease in fluorescence (excitation at 360 nm wavelength, emission cut-off filter 400 nm) for 1-500 s at 30 $^{\circ}\text{C}$. The curves were fitted and analyzed assuming first order kinetics and a constant drift from bleaching. Rate constants were determined by averaging the kinetic constants from each three repeats in three independent experiments.

3.7.6 Functional characterization *in vivo*

3.7.6.1 β -Galactosidase assay

This β -Galactosidase assay was performed according to an established protocol (Nelson et al., 2004). The reporter strain *S. cerevisiae* DSY-1100 (Genotype MATa leu2-112, ura3-1, trp1-1, his3-11, 15 ade2-1, can1-100 GAL SUC2 carrying plasmids pDS-125 (GR/TRP) and pDS-362 (GRE-lacZ/URA)) expresses mammalian Glucocorticoid Receptor (GR) and the lacZ gene under control of a Glucocorticoid Responsive Element (GRE) promoter. This strain was transformed with plasmids p423GPD and p423ADH into which the different Hip constructs had been cloned to test the influence on β -Galactosidase (lacZ is the gene for β -Galactosidase) activity. The empty plasmids p423GPD and p423ADH were also transformed as background controls. In the assay, the resultant strains were grown in 2 ml SC medium lacking uracil and tryptophan at 30 °C overnight, diluted to OD₆₀₀ of 0.05 – 0.10 in a final volume of 1 ml and grown at 25 °C and 600 rpm for 90 min. Cell growth was monitored spectrophotometrically to ensure that the cells were in the exponential growth phase. The glucocorticoid hormone Deoxycorticosterone (DOC) was then added to a final concentration of 50 nM. Hormone binding is needed for GR activation. 90 minutes after hormone addition, 50 μ l samples were withdrawn at 15-min intervals, the OD₆₀₀ determined and an equal volume Gal-Screen chemiluminescence reagent (Applied Biosystems) added. After 2 h incubation in a dark box, the reactions were analyzed with a luminometer (Berthold LB 9507). The emitted light at the different time points was plotted against the OD₆₀₀, and the slope was determined by linear regression as a measure of functional β -galactosidase expression. Cells transformed with p423GPD and p423ADH served as background controls. All the experiments were repeated at least three times.

3.7.6.2 Protein expression analysis by Western blotting

To analyze the expression levels of the Hip truncation constructs and Hip mutants in the p423ADH yeast strains, Western blotting of whole cell lysates was performed. The strain carrying the empty vector p423ADH was used as a background control. The overnight cultures were diluted to OD₆₀₀ 0.1 to 0.5 and grown to mid-log phase OD₆₀₀ 0.5 – 1.0 at 30 °C in selective medium lacking uracil and tryptophan. Equal amounts corresponding to yeast cell OD₆₀₀ 2 in 1.5 ml were harvested by centrifugation at 8000 rpm for 3 minutes at room temperature (Eppendorf 5417). Cell pellets were subjected to alkaline lysis buffer (Kushnirov, 2000). 10 μ l of the final samples were loaded on a 12 % SDS-PAGE and further analyzed by Western blotting.

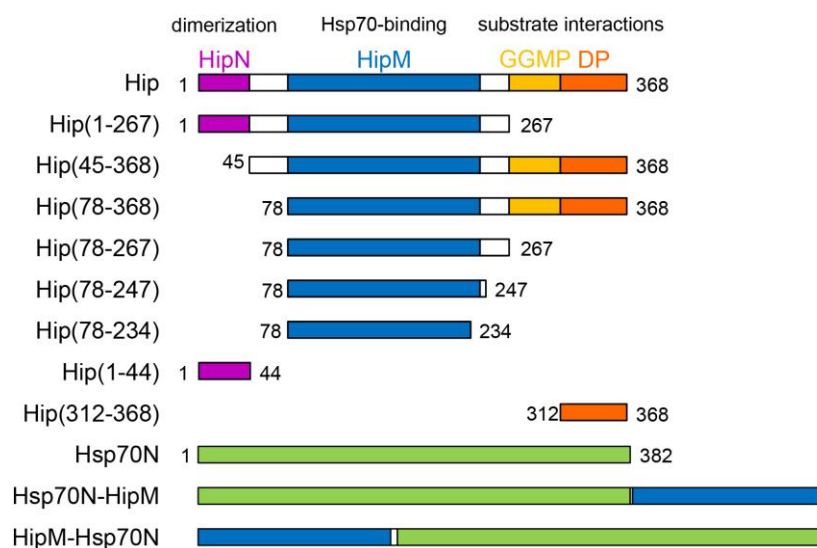
4 Results

In order to dissect Hip into functional modules and to determine their crystal structures, a series of truncation mutants and fusion proteins was generated. The constructs are schematically shown in Figure 14A. The construct boundaries were chosen based on sequence conservation between distantly related Hip homologs (for the respective sequence alignment, see Appendix 8.2).

4.1 Purification of Hip

All the proteins used in this project were overexpressed in the *E. coli* BL21 codon plus strain as N-terminal His-tag fusion proteins (Fig. 14A). Immobilized metal ion affinity chromatography (IMAC) of the cleared lysate on Ni-Chelating Sepharose (GE) served as first purification step. Proteins were eluted with a step gradient of increasing imidazole concentration. The purification step was monitored by SDS-PAGE analysis. Samples of whole cells (C), insoluble pellet (P), supernatant (lysate) (SN), flow-through (FT), wash steps (W) and elution fractions (E) were analyzed on SDS-PAGE gels. A representative example of such a Coomassie-stained gel from the purification of HipM, Hip(78-247), is shown in Figure 14B.

A)



B)

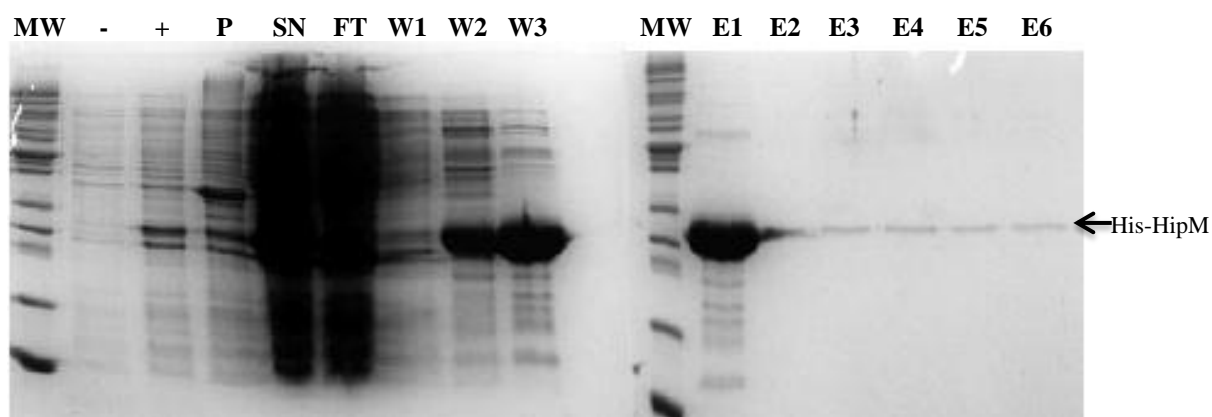


Figure 14: Partial purification of HipM on Ni-Chelating Sepharose.

- A) Schematic representation of the Hip constructs analyzed. The dimerization domain (HipN), Hsp70-binding (HipM) and DP domains of rat Hip are shown in purple, blue and orange, respectively. The GGMP repeat segment is indicated in yellow; the acidic linker regions in white. The NBD of human Hsp70 used to construct the Hsp70N-HipM and HipM-Hsp70N fusion proteins is shown in green.
- B) 15 % SDS-PAGE gel with samples from a crude purification of His-tagged HipM by nickel-affinity chromatography under native conditions. MW: Molecular weight marker (lanes 1); Whole cell lysate before induction (lane 2); Sample after induction with 0.5 mM IPTG at 20 °C for 16 hours (lane 3); P: Insoluble pellet (lane 4); SN: Supernatant (lane 5); FT: Flow-through (lane 6); Wash fractions (W1-W3); Elution fractions (E1-E6).

Fractions W3 and E1 were pooled and subjected to TEV protease digestion performed at 4 °C overnight to cleave the His-tag segment. To remove imidazole, the protein was subjected to buffer exchange by application to a GE desalting column on the next morning, which was equilibrated with buffer HKM2. The pool of the protein-containing fractions from the desalting column was subsequently applied a second time to the Ni-Chelating Sepharose column to remove the His-tagged TEV protease, the cleaved His-tag and the remaining His-HipM.

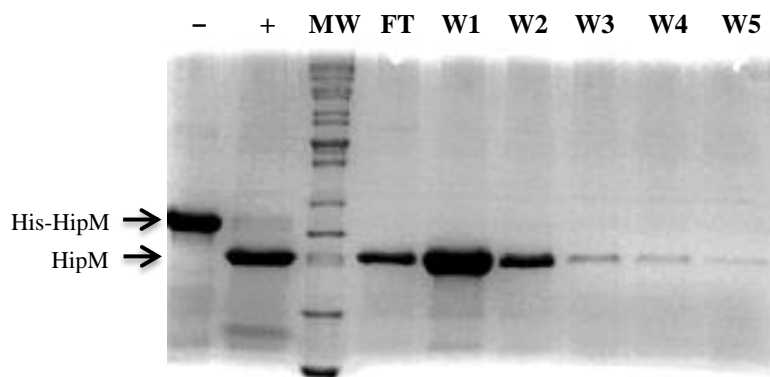
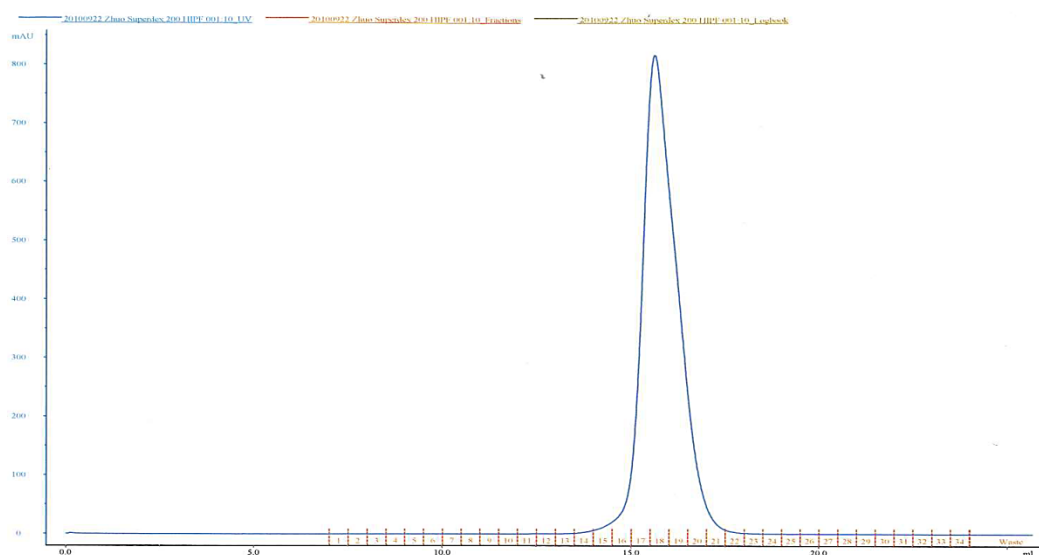


Figure 15: Nickel-affinity chromatography step after TEV protease cleavage in a representative purification of HipM construct.

15% SDS-PAGE analysis after TEV cleavage and further purification of HipM with Nickel-affinity chromatography. Sample before TEV cleavage (lane 1); Sample after cleavage over night at 4 °C (lane 2); Molecular marker (lanes 3); FT: Flow-through (lane 4); W1-W5: wash fractions

After the second run through Ni-Chelating Sepharose, the flow-through and wash fractions were loaded on 15 % SDS-PAGE for checking the cleavage quality and elution efficiency (Fig. 15). It can be seen that almost complete cleavage took place. The His-tagged proteins were removed and HipM protein was eluted in the fractions flow-through, W1 and W2, which were pooled and concentrated to a total volume of ~ 500 µl. The concentrate was subjected to size exclusion chromatography on a Superdex-200 column (GE) yielding mono-disperse and pure protein (Fig. 16A). HipM was eluted at an apparent molecular weight of 20 kDa, as analyzed by SDS-PAGE (Fig. 16B).

A)



B)

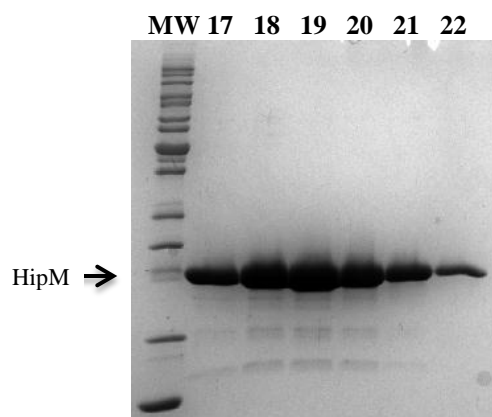


Figure 16: Size exclusion chromatography step of a representative purification.

- A) Size exclusion chromatography by using Superdex 200 GL10/300 (GE).
 B) 15 % SDS-PAGE gel with fractions from the purification of HipM with size exclusion chromatography. The location of the HipM band is indicated. MW: Molecular weight marker (lane 1); Elution fractions: numbers are indicated above the gel.

After size exclusion chromatography, samples were pooled and concentrated to ~ 18-22 mg/ml for protein crystallization. The average yield was ~ 20-30 mg of pure protein from 1L *E. coli* BL21 codon plus grown in a shaking culture of LB medium. All the purification steps were performed within 4 days. The purified protein was subsequently stored at - 80 °C.

4.2 Characterization of the Hip-Hsp70 interaction by isothermal titration calorimetry

4.2.1 Selectivity of Hip for the ADP-bound state of Hsp70

To gain first insight into the mechanism by which Hip modulates the Hsp70 reaction cycle, we used isothermal titration calorimetry (ITC) to quantify Hip binding to Hsp70 and to determine the affinity constant. These experiments were performed with full-length rat Hip and human Hsp70, which shares 97.4 % sequence identity with rat Hsp70. First, we analyzed the interaction of Hip with different conformational states of full-length Hsp70 in the absence of nucleotide (apo) and presence of ADP and ATP.

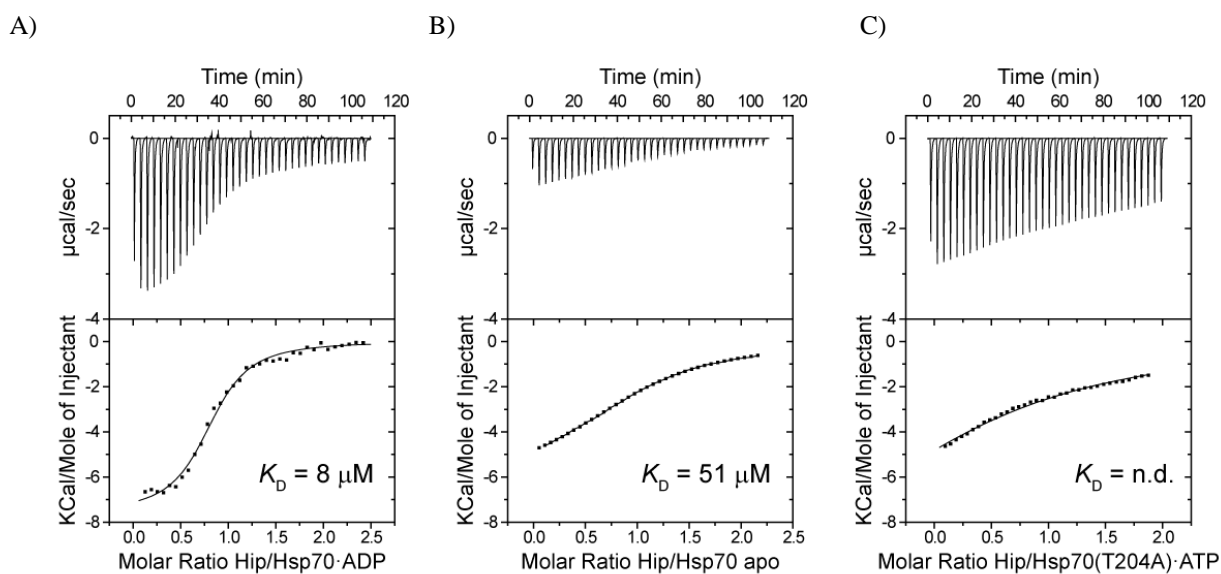


Figure 17: Affinity measurements by isothermal titration calorimetry (ITC) of Hip with Hsp70 in the apo, ADP and ATP-bound conformational states. Upper panel: Energy flow as a result of injection of Hip plotted against time. Lower panel: Heat release per injection plotted against the molar ratio of Hip:Hsp70.

- A) Interaction of Hip with Hsp70 in the ADP-bound state. The experiments were performed at 22 °C using a buffer of 20 mM HEPES-KOH pH 7.4 containing 100 mM KCl. 5 mM Mg-ATP was added when indicated. All data were analyzed assuming that Hip protomers act as independent binding partners.
- B) Interaction of Hip with nucleotide-free Hsp70 (apo).
- C) Interaction of Hip with Hsp70 in the ATP-induced conformational state. The hydrolysis-defective mutant Hsp70(T204A) was used (Barthel et al., 2001; Kityk et al., 2012). The curve-fitting of the binding parameters for Hsp70(T204A)·ATP data was unstable.

| Construct | ΔH [calmol ⁻¹] | ΔS [calmol ⁻¹ K ⁻¹] | N [molar ratio] | K_D [μ M] |
|------------------|------------------------------------|--|-----------------|------------------|
| Hsp70•ADP | -7573 | -2.21 | 0.82 | 8 |
| Hsp70•Apo | -6232 | -1.49 | 1.01 | 51 |
| Hsp70•ATP | -6192 | -0.02 | 1.05 | 26 |
| Hsp70(T204A)•ATP | -11650 | -23.20 | 1 | n.d. |
| Hsp70(D199A)•ATP | -7332 | -2.84 | 1.04 | 16 |

Table 12: The binding parameters as determined by curve-fitting the ITC experimental data of Hip with Hsp70.

Our data showed that Hip had a strong preference for the ADP-bound form of Hsp70 ($K_D = 8 \mu\text{M}$), relative to the nucleotide-free apo state ($K_D = 51 \mu\text{M}$) (Fig. 17A & 17B, Table 12). We also studied the affinity of Hip for Hsp70·ATP. In an ITC experiment with wild type Hsp70 in the presence of a large excess of ATP, we determined an affinity of 26 μM , close to the value of ADP-bound Hsp70 (Table 12, data not shown). Because we suspected that Hip might preferentially recognize and stabilize transiently formed ADP-bound Hsp70, we decided to also test ATP-hydrolysis-defective mutants of Hsp70. The Hsp70 mutation D199A is ATPase-inactive and fails to undergo an ATP-induced conformational transition to a more compact state (Buchberger et al., 1995; McCarty and Walker, 1991). Using this mutant we found an affinity for Hip of 16 μM , suggesting that ATP binding to Hsp70 has only a minor influence on Hip binding (Table 12, data not shown). In DnaK, T199A is a well-characterized mutation, which effectively abolishes ATPase activity, but maintains allosteric coupling. In the crystal structure, this mutant adopts a compact conformation, in which NBD and SBD are closely associated (Kityk et al., 2012; Qi et al., 2013). The co-existence of the “open” and “closed” conformations in the Hsp70 ensemble due to ATP hydrolysis appears to be largely suppressed in this mutant. Based on sequence alignment, the homologous mutation in human Hsp70 is T204A. In the

presence of ATP, Hip bound with strikingly low affinity to the ATPase-inactive Hsp70 mutant T204A. The titration curve was too shallow for proper quantification (Fig. 17C).

Despite the strong preference of Hip for the ADP-bound state of Hsp70, it should be noted that the affinity to Hsp70·ADP is rather moderate ($K_D = 8 \mu\text{M}$) in comparison to Hsp70 nucleotide exchange factors (NEFs), which all display substantially higher affinity for Hsp70 ($K_D \approx 0.1 \mu\text{M}$). This suggests that the interaction with Hip must be enhanced by additional contacts for allowing Hip to effectively compete with the binding of nucleotide exchange factors (NEFs). For example, interactions of Hip with Hsp70-bound substrate protein may contribute to the strength of association.

Besides the binding affinity, ITC curves may also reveal the binding stoichiometry and the existence of distinct binding sites in the receptor component. The ITC binding curves indicated that one Hip dimer bound two molecules of Hsp70·ADP (Table 12). We found no evidence for any negative interference between the Hsp70 binding sites in the Hip dimer. Thus avidity effects might enhance complex formation of Hip with entities containing multiple Hsp70 molecules.

4.2.2 Identification of binding site of Hip on Hsp70

We have demonstrated that Hip selectively binds to full length Hsp70 in the ADP-bound state, which is in line with previous studies (Höfheld et al., 1995). But Hip was originally identified as binding partner of the NBD of Hsp70 (Höfheld et al., 1995). We attempted to confirm this by ITC analysis.

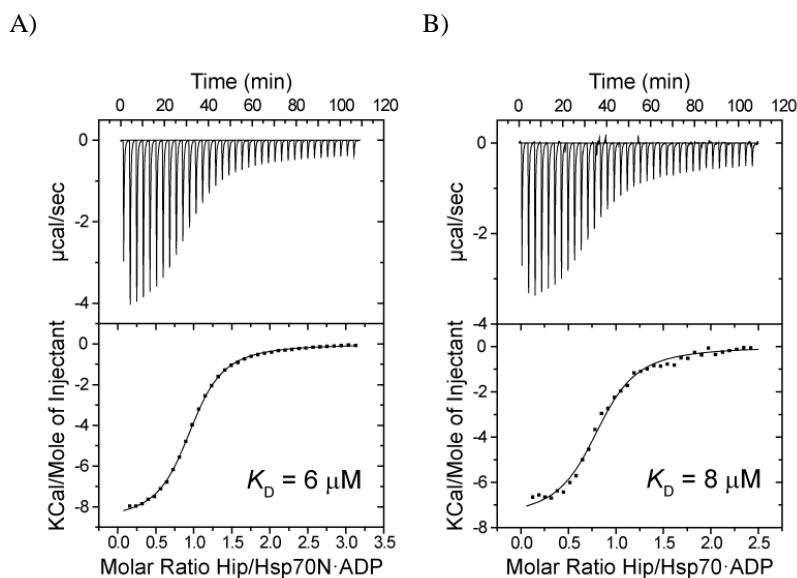


Figure 18: Hip affinity towards the ADP-bound states of Hsp70NBD and full length Hsp70, respectively.

- A) ITC titration of full-length Hip with the NBD of Hsp70 (Hsp70N) in the ADP-bound conformational state. The data were analyzed based on the Hip protomer concentration. Prior to dialysis against GF buffer, 1 mM ADP was added to Hsp70N.
- B) ITC titration of full-length Hip with full-length Hsp70 in the ADP-bound conformational state.

Based on ITC, the affinity of full-length Hip for the ADP-bound Hsp70N ($K_D = 6 \mu\text{M}$) is essentially the same as that for full-length Hsp70 ($K_D = 8 \mu\text{M}$) (Fig. 18). Thus, the SBD of Hsp70 does not detectably contribute to the interaction with Hip *in vitro*. It might however influence Hip binding in presence of substrate *in vivo*.

4.2.3 Identification of the Hip minimal Hsp70-binding domain

For the exact identification of the region in Hip responsible for the binding to Hsp70N, truncation constructs were analyzed by ITC (Fig. 19). Höhfeld and coworkers had already found that both the TPR domain and the C-terminally adjacent charged region in Hip are needed for Hsp70 binding (Irmer and Höhfeld, 1997). Residues 107-241 were predicted to form a TPR domain. In a sequence alignment including distantly related Hip homologs (Appendix 8.2), this region is flanked by conserved segments, starting at residue 78 and ending at 267. For these regions, however, no secondary structure was predicted by J-Pred (data not shown), suggesting intrinsically poor ordering, which might be a potential obstacle for crystallization. To our sur-

prise, the construct encompassing residues 107-267 was not expressed as a soluble protein in *E. coli* (data not shown).

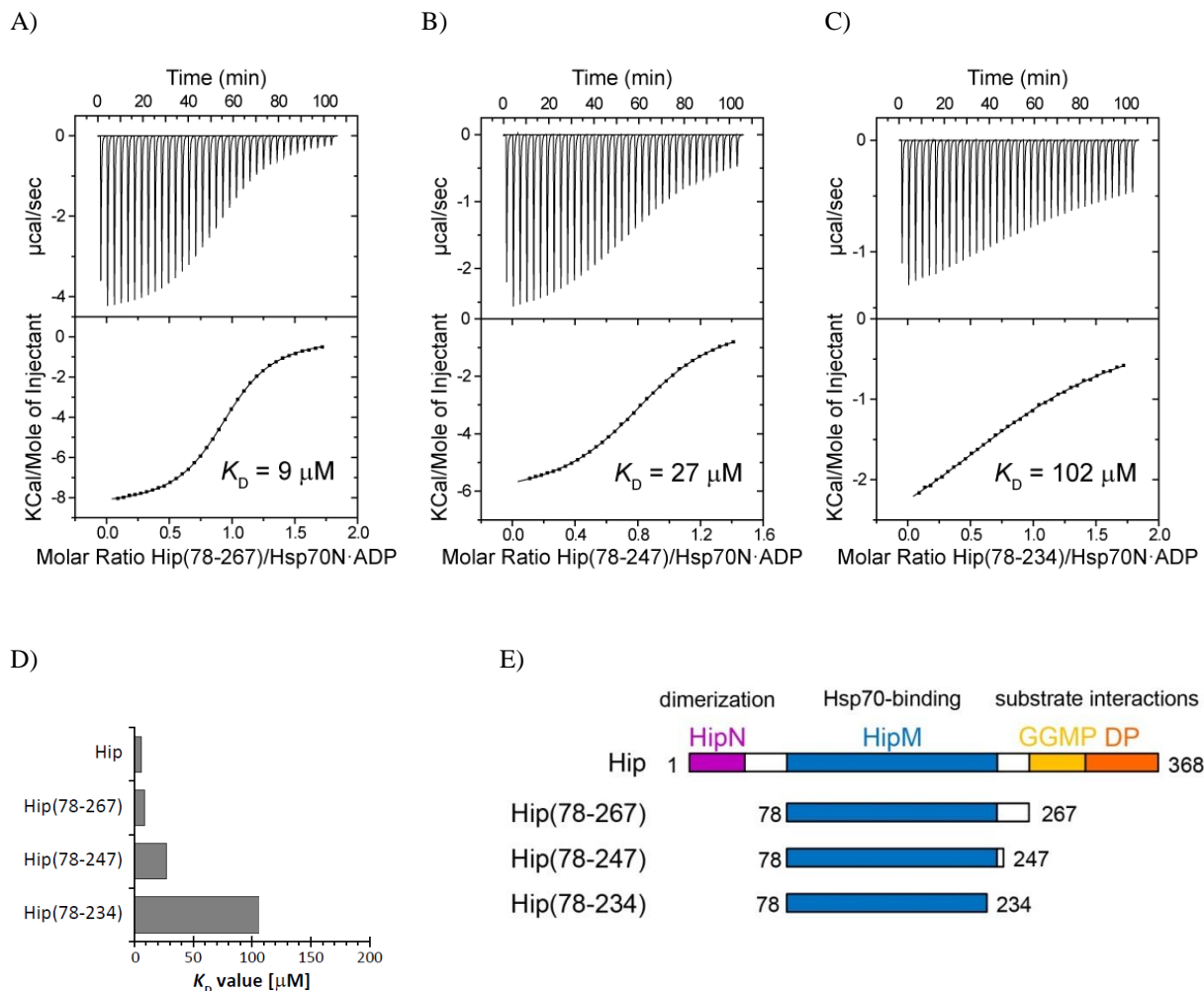


Figure 19: Affinity of Hip truncation constructs towards Hsp70N•ADP.

(A-C) ITC curves for binding of truncation constructs Hip(78-267), Hip(78-247) and Hip(78-234) to Hsp70N•ADP. Prior to dialysis against ITC buffer, 1 mM ADP was added to Hsp70N.

D) Direct comparison of the binding affinities of the truncation constructs towards Hsp70N•ADP.

E) Schematic representation of the Hip truncation constructs analyzed.

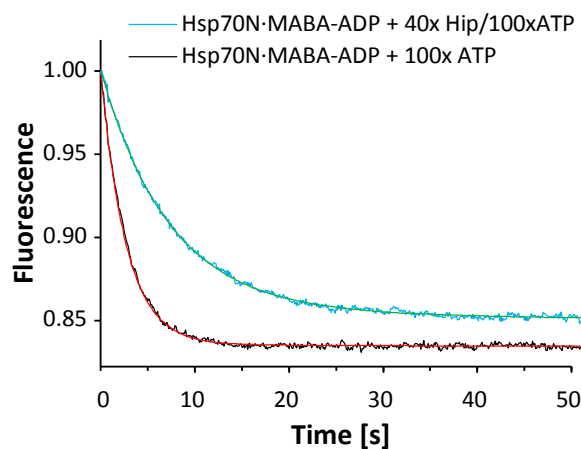
Hip(78-267) contains the whole conserved segment and thus the complete Hsp70-binding region as predicted by Höhfeld and coworkers. Using ITC, we found virtually the same affinity to Hsp70N•ADP ($K_D = 9 \mu\text{M}$) as for full-length Hip ($K_D = 8 \mu\text{M}$), suggesting that this segment indeed contains the complete Hsp70-binding region in Hip. C-terminal truncation to Hip(78-247) lowered the affinity slightly to $K_D = 27 \mu\text{M}$ (Fig. 19B & 19D). A sharp decrease in affinity

was found for Hip(78-234) with $K_D = 102 \mu\text{M}$, suggesting that residues 235-247 contribute substantially to Hsp70 binding (Fig. 19C, 19D & 19E).

4.3 Hip binding decelerates nucleotide release from Hsp70

Our ITC analysis demonstrated that Hip preferentially binds to Hsp70 in the ADP-bound state. Hip might thereby also stabilize Hsp70 binding to ADP (Höhfeld et al., 1995). In presence of high ATP concentrations such as in the cytosol, this would only take effect however if ADP dissociation was also slowed down. In order to monitor nucleotide dissociation from Hsp70 in presence of Hip, we used the fluorescent ADP analog MABA-ADP as a reporter. Previous studies have shown that the Hsp70-binding and dissociation properties of MABA-ADP are closely similar to ADP (Brehmer et al., 2004). For the assay, Hsp70 or Hsp70N were incubated with a stoichiometric amount of MABA-ADP to prepare the Hsp70-MABA-ADP complex. Mixing with a great excess of ATP in a stopped-flow apparatus prevents re-binding of MABA-ADP to Hsp70, resulting in decreased fluorescence (Fig. 20A).

A)



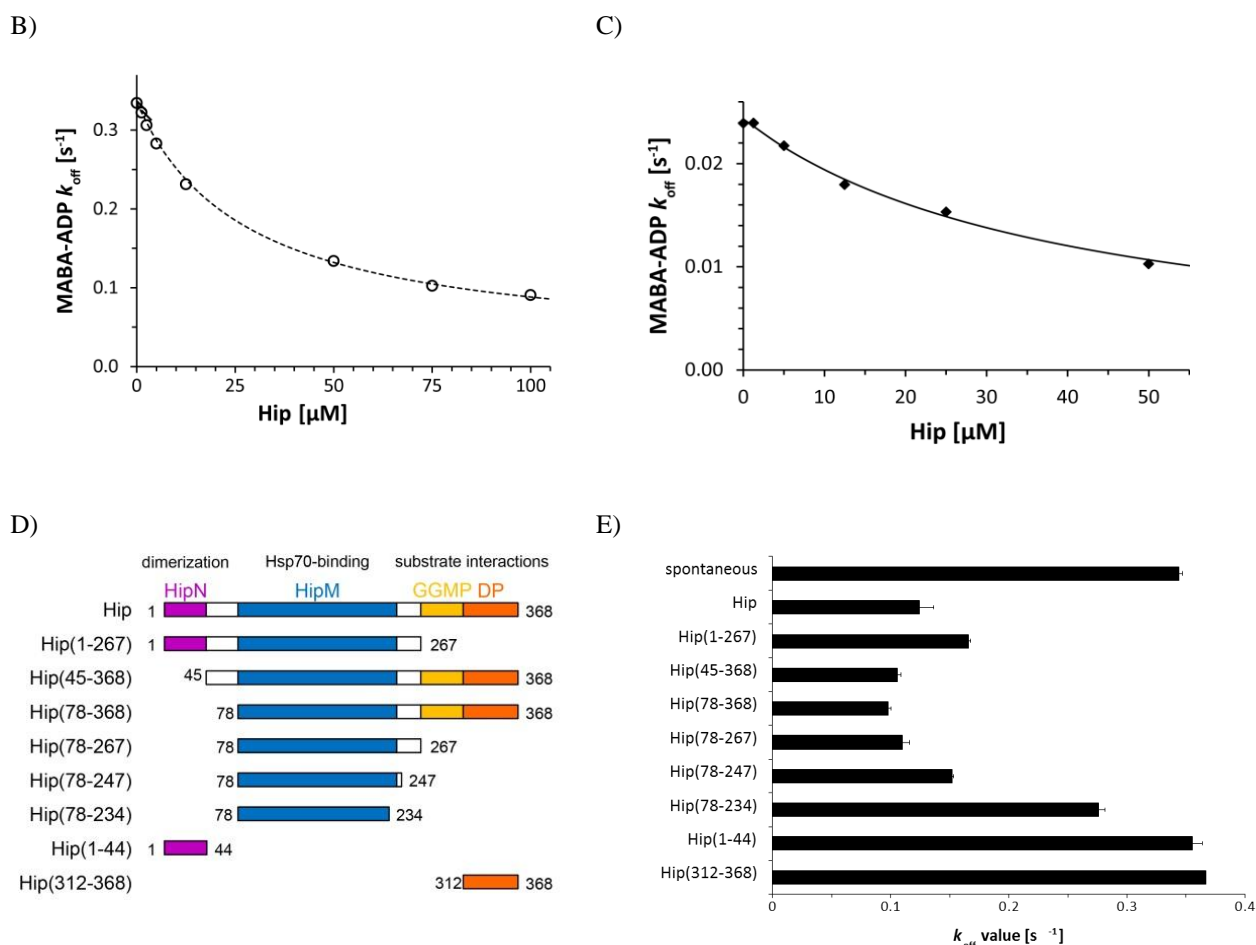


Figure 20: Deceleration of MABA-ADP release from Hsp70N by Hip.

- A) Fluorescence traces reflecting the time-dependent dissociation of Hsp70N-bound MABA-ADP. The fluorescent MABA-ADP was incubated for 30 min with 2.5 μ M Hsp70N at 30 $^{\circ}$ C, and then mixed in a stopped-flow apparatus with buffer containing 250 μ M ATP either with or without 100 μ M of Hip protein (i.e., 40-fold excess over Hsp70N). The fluorescence traces were curve-fitted using a model assuming single-exponential decay and a linear drift to account for bleaching. The red line is the spontaneous release of MABA-ADP from Hsp70N. The blue line is in the presence of 100 μ M of Hip.
- B) Hip-mediated attenuation of MABA-ADP dissociation from Hsp70N in buffer without phosphate. For determining the concentration dependence, the Hip concentration in the ATP solution was varied. Otherwise the solution was as described above. Apparent k_{off} rates were determined from curve-fitting the exponential decay of MABA-ADP fluorescence and plotted against the final Hip concentration.
- C) Hip-mediated attenuation of MABA-ADP dissociation from Hsp70N in buffer containing 10 mM phosphate.
- D) Schematic representation of the Hip constructs analyzed. The dimerization domain (HipN), Hsp70-binding (HipM) and DP domains of rat Hip are shown in purple, blue and orange, respectively. The GGMP repeat segment is indicated in yellow; the acidic linker regions in white.
- E) MABA-ADP dissociation as a proxy for estimating the affinity of different Hip truncation constructs towards Hsp70. Nucleotide dissociation rates from Hsp70N were measured in presence of each of the Hip truncation constructs. The Hsp70N:MABA-ADP complex was mixed with buffer containing 250 μ M ATP and 100 μ M of the respective Hip construct. Apparent k_{off} rates were determined from curve-fitting the exponential decay

of MABA-ADP fluorescence. The averages and standard deviations from three independent experiments are shown.

We first investigated the dependence of the MABA-ADP dissociation rate on the Hip concentration under condition of the release assay. In the absence of Hip, the spontaneous MABA-ADP off-rate was $\sim 0.34 \text{ s}^{-1}$ (red line, Fig. 20A), similar to the ADP dissociation rate for the bacterial homolog of Hsp70, DnaK (Gässler et al., 2001). At an equimolar concentration relative to Hsp70N, Hip lowered the ADP-dissociation rate only marginally, in line with its modest affinity to Hsp70N (Fig. 20B). However, a substantial decrease of k_{off} ($\sim 0.13 \text{ s}^{-1}$) was observed upon the addition of full length Hip in 40-fold excess over Hsp70N (blue line, Fig. 20A), indicating that Hip can stabilize Hsp70 in the ADP bound state.

The ADP release rate from eukaryotic Hsp70 was reported to be greatly lowered in the presence of inorganic phosphate, a component of the eukaryotic cytosol (Brehmer et al., 2001). We suspected that Hip and phosphate might have a multiplicative effect on the dissociation rate. In presence of 10 mM inorganic phosphate, a possible physiological concentration, the spontaneous MABA-ADP off-rate was lowered by a factor of ~ 15 . However, in relative terms the dependence on Hip concentration did not change substantially in presence of phosphate ($\sim 60\%$ decrease at 50 μM Hip and 1.25 μM Hsp70N), arguing against a synergistic effect of Hip and phosphate (Fig. 20C).

Next, we used the MABA-ADP dissociation from Hsp70N as a functional assay for our Hip truncation constructs. To get a robust signal, a 40-fold excess of the respective construct over Hsp70N•MABA-ADP was used (Fig. 20D & 20E).

The N-terminal dimerization domain Hip(1-44) and C-terminal DP repeat region Hip(312-368) alone did not change the MABA-ADP dissociation rate from Hsp70N, consistent with Hsp70 exclusively binding to the middle segment of Hip. Moreover, the dissociation rates in the presence of full-length Hip(1-368) and the C-terminally truncated construct Hip(1-267) were closely similar, suggesting that the C-terminal GGMP motifs and the DP repeat regions do not contribute to Hsp70-binding (Fig. 20D & 20E). When we compared the off-rates in presence of dimeric full-length Hip(1-368) with monomeric Hip(45-368) and Hip(78-368), we found a

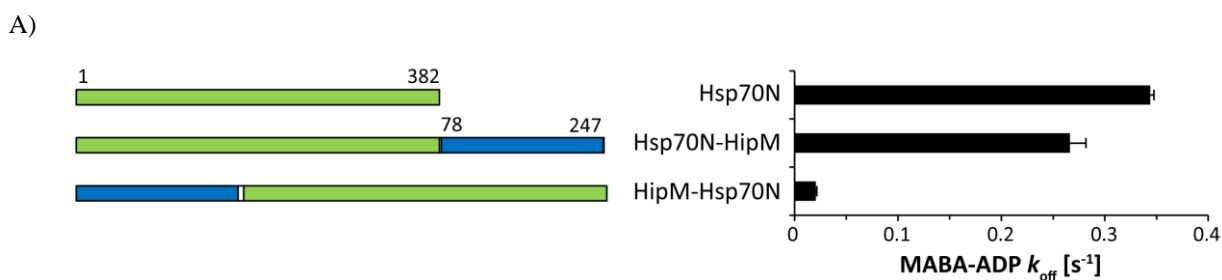
slightly increased stabilizing effect with the N-terminal truncation mutants, indicating that dimerization might disturb the interaction with Hsp70 to some degree.

Hip(78-247) and Hip(78-267) slowed MABA-ADP dissociation from Hsp70N to similar extents, consistent with our ITC affinity data. Compared to Hip(78-247) and Hip(78-267), the shorter construct Hip(78-234) had a substantially lower activity, confirming that the functional MABA-ADP dissociation assay can be used as a proxy for affinity.

Taken together, the ITC experiments demonstrated that Hip has a clear preference for the Hsp70•ADP complex. In addition, the NBD of Hsp70 is sufficient for Hip binding. Consistent with the loose domain tethering in the structural model of the ADP complex of Hsp70, the SBD of Hsp70 is neutral towards Hip binding. Moreover, the ability of Hip truncation mutants to attenuate MABA-ADP release from Hsp70N correlated with their binding affinity to Hsp70N-ADP. The investigation of Hip truncation mutants revealed that both conserved flanking regions to the TPR domain are required for the interaction with Hsp70.

4.4 Nucleotide release is virtually abolished in a Hip-Hsp70 fusion protein

Even at a high 40-fold excess of Hip only a partial reduction of the MABA-ADP dissociation rate from Hsp70 was observed *in vitro*. To estimate the maximal effect of Hip on nucleotide dissociation under saturation conditions, we generated two fusion proteins of HipM and Hsp70N differing in the sequence of the domain segments, HipM-Hsp70N and Hsp70N-HipM, respectively (Fig. 21A).



B)

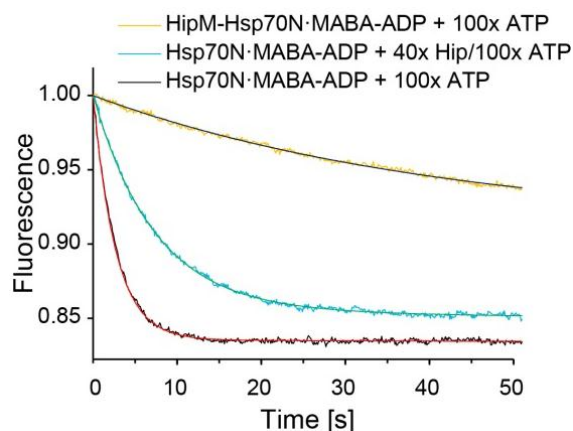


Figure 21: Analysis of the HipM-Hsp70N fusion proteins with the MABA-ADP dissociation assay.

- A) Left panel: Schematic representation of the fusion protein constructs. The NBD of human Hsp70 is shown in green; HipM in blue. Right panel: MABA-ADP dissociation rates from the fusion proteins as determined by curve-fitting.
- B) Fluorescence traces reflecting the time-dependent dissociation of MABA-ADP. The fluorescent MABA-ADP was incubated with $2.5 \mu\text{M}$ of the respective protein at 30°C for 30 min, and then mixed in a stopped-flow apparatus with buffer containing $250 \mu\text{M}$ ATP. The fluorescence traces were curve-fitted using a model assuming single-exponential decay and a linear drift to account for bleaching. The yellow line was recorded with the HipM-Hsp70N fusion protein. For comparison, the spontaneous dissociation from Hsp70N and the triggered dissociation in presence of a 40-fold excess of Hip are shown in red and blue (same as in Figure 20A).

In the HipM-Hsp70N fusion construct, the spontaneous MABA-ADP dissociation rate was strongly reduced to a k_{off} value of 0.02 s^{-1} , indicating a functional intramolecular interface formed between the domains (Fig. 21A, and yellow line in Fig. 21B). This demonstrates that there is ample room for enhancement of the functional effect of Hip by additional interactions stabilizing the Hip:Hsp70 complex. Conversely, the reversed Hsp70N-HipM showed a k_{off} value close to the spontaneous off-rate, indicating that this fusion protein failed to form a functional intramolecular interface in this orientation (Fig. 21A).

4.5 Structural analysis of Hip

4.5.1 Hip domain structure

We delineated the probable domain boundaries in rat Hip from the alignment with remote homologs from protists, which became available only after most of the previous functional

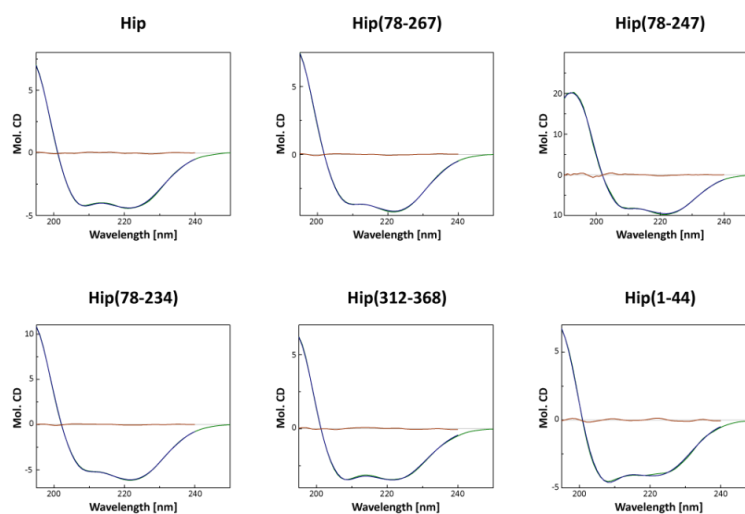
work of Hip had been completed, shown in Appendix 8.2. Including these additional sequences revealed the functional, structured regions as boxes of increased conservation.

Limited proteolysis of full-length Hip gave unclear results (data not shown). The first block of high conservation encompassing the dimerization domain terminates at Pro44 in rat Hip. This is followed by an acidic linker of ~ 30 amino acids, which is likely to be unstructured. The net negative charge might help to force a Hip dimer into an extended conformation by electrostatic repulsion. The following block of high sequence conservation, residues 78-242 in rat Hip, encompasses additional regions flanking the predicted core TPR domain (residues 107-232) and likely forms a structured middle domain. This is followed by another acidic stretch of ~ 20 residues and a variable segment of imperfect repeats of the motif Gly-Gly-(Met/Phe)-Pro (GGMP). At the C-terminus a conserved DP domain is located (residues 312-368 in rat Hip).

4.5.2 Circular dichroism spectroscopy of Hip domains

Hip has been proposed to be an almost all α -protein (Prapapanich et al., 1998). The secondary structure of a polypeptide chain (α -helix, β -sheet and random coils) shows a characteristic circular dichroism (CD) spectra. Therefore, we analyzed our Hip truncation constructs using CD spectroscopy to test whether the conserved regions are indeed independently folded domains. The secondary structure content was estimated by deconvolution of the CD spectra with the program CONTIN (Fig. 22).

A)



B)

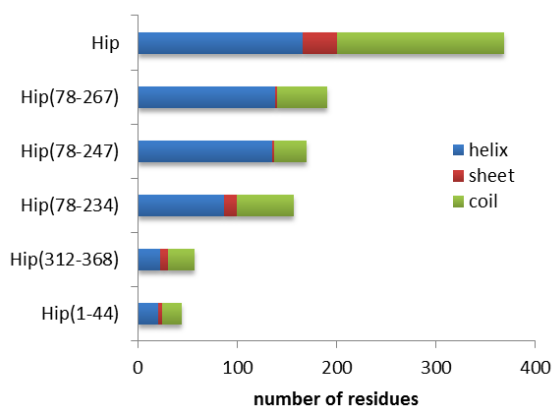


Figure 22: CD Spectroscopic analysis of Hip truncation constructs.

- A) CD spectra of Hip constructs. A strong maximum at 195 nm and two strong minima at 208 and 222 nm are indicative of α -helical structures. A minimum at 218 nm and maximum at 195 nm suggest β -sheet structures. The mean molar ellipticities per residue for indicated constructs are shown in green. Calculated spectra are shown in blue and residual differences in red.
- B) Comparison of the putative secondary structure contents, represented as bar graphs. The α -helical, β -sheet and random-coil structure is indicated in blue, red and green, respectively.

The CD spectra indicate that full-length Hip and the analyzed truncation constructs indeed contain a large amount of α -helices (Fig. 22A). To compare the truncation constructs with each other, the secondary structure distribution for each construct is shown in Figure 22B. The secondary structure content of full-length Hip is almost identical to the sum of N-terminal dimerization domain Hip(1-44), middle domain Hip(78-247) and the DP domain Hip(312-368), suggesting that these regions indeed represent the structured domains in Hip. The linker regions and the GGMP repeats do not seem to form regular secondary structure.

Furthermore, the analysis of the secondary structure content of the middle domain constructs Hip(78-267), Hip(78-247) and Hip(78-234) revealed that Hip(78-267) and Hip(78-247) had approximately the same amount of α -helical structure, suggesting that the C-terminal charged flanking region is unstructured in isolation. Notably, Hip(78-234) had much less helical content, suggesting that residues 235-247 are required for full helix formation in the domain. These findings explain why among the three Hip constructs only Hip(78-247) was successfully crystallized (see section 4.5.4).

In contrast to Hip(1-44), which was crystallized successfully, we were unable to obtain crystals of the C-terminal Hip(312-368) (data not shown). This suggests that the DP domain of Hip is not stably structured in isolation, and may only assume a defined structure in presence of a bound substrate.

4.5.3 The DP domain of Hip

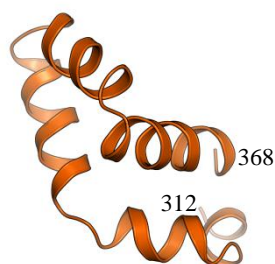


Figure 23: Structure model of the DP domain (312-368) in Hip.

The C-terminal DP domain beginning at residue 312 is quite well-conserved. Analysis of the CD spectra of the domain constructs versus full-length Hip suggested that DP domain of Hip is partially structured (Fig. 22). We further analyzed the domain constructs Hip(1-44), Hip(78-234) and Hip(312-368) by one-dimensional $^1\text{H-NMR}$ spectroscopy (Rehm et al., 2002) (Zhuo Li, Master thesis). The dispersion of amide resonances indicated that the N-terminal domain was stably structured, whereas this was less so in the spectrum of the C-terminal domain, suggesting that this region was structurally more dynamic (data not shown). Fortunately, the NMR structures of the DP domains in Stt1 (yeast Hop) were solved independently in the meantime (Schmid et al., 2012). Thereby we were able to create a homology model for the respective segment in Hip based on the DP1 domain of Stt1, which is largely consistent with our CD data (Fig. 22).

4.5.4 Crystallization of the Hsp70-interacting domain of Hip

Since our initial attempts to crystallize full-length Hip failed, we chose to determine the crystal structures of individual Hip segments (Zhuo Li, Master thesis). At the beginning of the doctoral

study, we had already solved the structure of the N-terminal dimerization domain of Hip (Zhuo Li, Master thesis). In NMR spectra, the C-terminal DP domain appeared only partially structured. Consistently, crystallization of this segment did not succeed. Therefore, we concentrated on the structure of the Hsp70-binding domain of Hip, which has been predicted to contain a tetratricopeptide repeat (TPR) domain. All three truncation constructs Hip(78-267), Hip(78-247) and Hip(78-234) encompassing this putative TPR domain were tested in crystallization trials based on commercial and in-house factorial screens.

Only the domain construct Hip(78-247) was however successfully crystallized. In the following, this construct is designated HipM – for Hip middle domain. Initial crystals were obtained with two different precipitant compositions: 2.4 M sodium malonate pH 7.0 and 1.4 M tri-sodium citrate, 0.1 M HEPES pH 7.5. These crystals grew in clusters of thin plates. To refine the crystal morphology, we varied the pH value from 7.0 to 6.0 in the sodium malonate condition. Single, cube-shaped tetragonal crystals appeared after three weeks (Fig. 24A). Conversely, single crystals could only be obtained from Na-malonate pH 7.0 conditions through several rounds of micro-seeding. Unfortunately, the optimization of tri-sodium citrate condition was not successful, even when employing micro-seeding and streak-seeding.

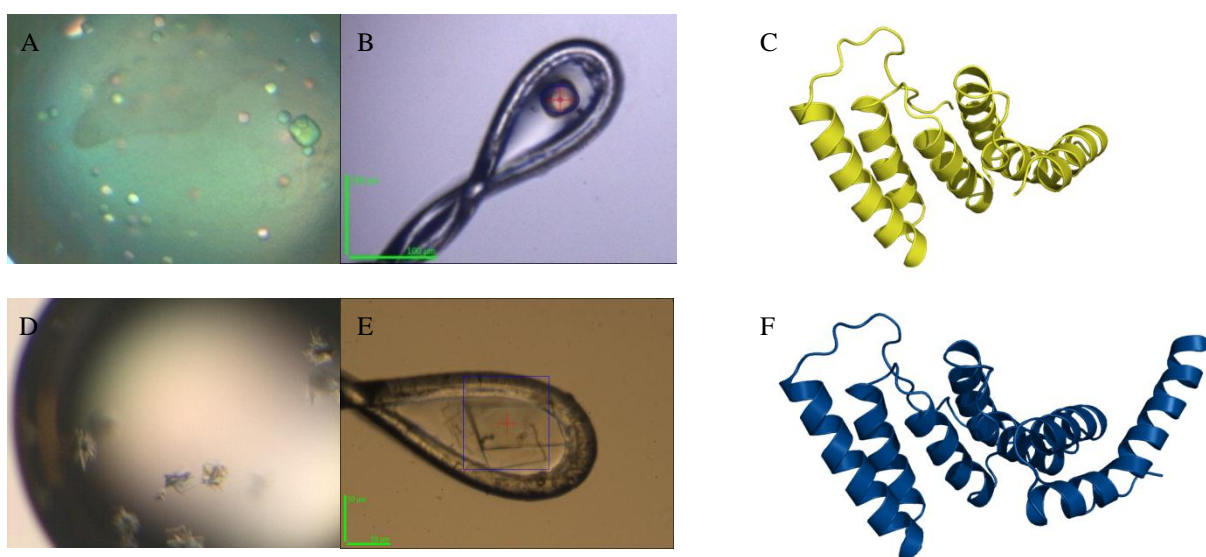


Figure 24: Representative images of Hip(78-247) crystal forms and structures.

-
- A) Crystals obtained in a hanging-drop setup with 2.1M Na-malonate pH 6.0 as precipitant at 18 °C.
 - B) A tetragonal crystal of Hip(78-247) from the Na-malonate condition at pH 6.0 mounted in a cryo loop. The green scalebar corresponds to 100 μm .
 - C) Structure of the tetragonal crystal form. Crystal form I was solved by Os-SIRAS at a resolution of 2.6 Å. The R and R_{free} values are 18.4 % and 24.8 %, respectively.
 - D) Crystals obtained in a hanging-drop setup with 2.15 M Na-malonate pH 7.0 at 18 °C.
 - E) Monoclinic plate-shaped crystals of Hip(78-247) from the Na-malonate condition at pH 7.0 mounted in a cryo loop. The green scalebar corresponds to 50 μm .
 - F) Structure of the monoclinic crystal form. Crystal form II was solved by molecular replacement at a resolution of 2.8 Å. The R and R_{free} values are 24.6 % and 29.9 %, respectively.

4.5.5 Structure determination and refinement of HipM

From the construct HipM we obtained two crystal forms, I and II (Fig. 24B & 24E), with native diffraction to 2.6 Å and 2.8 Å, respectively. Table 13 provides an overview of data collection and refinement statistics. The structure of form I was crystallized in space group $P4_32_12$ with unit cell dimension of $a \cdot b \cdot c = 79.86 \text{ \AA} \cdot 79.86 \text{ \AA} \cdot 97.31 \text{ \AA}$. The structure was initially solved by Os-SIRAS at a resolution of 4 Å, containing two protein chains per asymmetric unit. The Os substructure was found using the Shelx program package (ShelxC, ShelxD) and contained three Os-atom positions. The heavy atom positions were refined and initial phases were calculated in SHARP (Vonrhein et al., 2007). After further density modifications by Resolve (Terwilliger, 2000), an interpretable map was obtained, which was sufficient to manually build initial models with Coot (Emsley and Cowtan, 2004). The final model building was carried out against a high-resolution native dataset of 2.6 Å resolution. Subsequently, several rounds of refinement were performed with Coot and REFMAC (Murshudov et al., 1997), respectively. The refinement statistics was greatly improved by the application of TLS parameterization in REFMAC. This could indicate that the Hip TPR domains undergo rigid body movements with respect to each other in the crystal lattice. The final structure of form I (Fig. 24C) was refined to a crystallographic R-factor of 18.4% (R_{free} 24.8%).

The structure of the second crystal form II (Fig. 24F) was solved by molecular replacement only after the Hsp70N-HipM crystal structure had been determined (see below), using the HipM segment as a search template. The crystals are twinned, belonging to space group $P2_1$ with lattice parameters of $a \cdot b \cdot c = 57.29 \text{ \AA} \cdot 127.00 \text{ \AA} \cdot 57.31 \text{ \AA}$, and harbor four copies of HipM per asymmetric unit. The best native diffraction data at 2.8 Å resolution was collected at the European Synchrotron Radiation Facility (ESRF).

Compared to form I, in which residues 95-212 are ordered, almost the complete HipM construct (residues 81-241 in all chains) was visible in crystal form II. Therefore this crystal form will be discussed first.

| Construct | Hip(1-44) | | Hip(78-247) | | | Hsp70N-Hip |
|---|-------------------------------|-----------------------------|--|--|-----------------------------|-------------------------------|
| | EtHgPO ₄ | native | K ₂ [OsO ₄]-I | native-I | native-II | native |
| Space group | <i>P</i> 2 ₁ | <i>P</i> 2 ₁ | <i>P</i> 4 ₃ 2 ₁ 2 | <i>P</i> 4 ₃ 2 ₁ 2 | <i>P</i> 2 ₁ | <i>P</i> 4 ₁ |
| Cell dimensions | | | | | | |
| a, b, c (Å) | 28.80, 43.89, 32.24 | 28.78, 43.98, 32.21 | 79.05, 79.05, 96.82 | 79.86, 79.86, 97.31 | 57.29, 127.00, 57.31 | 65.60, 65.60, 141.47 |
| α, β, γ (°) | 90, 93.40, 90 | 90, 93.44, 90 | 90, 90, 90 | 90, 90, 90 | 90, 105.93, 90 | 90, 90, 90 |
| Wavelength (Å) | 1.005 | 0.9000 | 1.140 | 1.005 | 0.9189 | 0.8726 |
| Resolution (Å)* | 43.90 - 1.57 (1.67 - 1.57) | 43.98 - 1.1 (1.16 - 1.1) | 48.41 - 4.0 (4.22 - 4.0) | 48.84 - 2.6 (2.74 - 2.6) | 45.74 - 2.8 (2.95 - 2.8) | 48.10 - 2.65 (2.79 - 2.65) |
| <i>R</i> _{merge} * | 0.104 (0.375) | 0.057 (0.323) | 0.187 (0.432) | 0.054 (0.338) | 0.110 (0.427) | 0.068 (0.491) |
| <i>I</i> / <i>σ</i> <i>I</i> * | 16.9 (6.7) | 13.4 (4.1) | 13.4 (6.2) | 24.0 (5.5) | 8.3 (2.5) | 16.1 (2.9) |
| Completeness (%) * | 99.9 (99.7) | 98.6 (97.5) | 99.8 (99.0) | 99.5 (98.5) | 98.7 (96.0) | 99.9 (99.8) |
| Redundancy * | 7.3 (7.3) | 3.6 (3.5) | 13.1 (13.4) | 6.3 (6.6) | 3.1 (3.1) | 5.1 (5.1) |
| Refinement | | | | | | |
| Resolution (Å) | - | 20 - 1.1 | - | 20 - 2.6 | 20 - 2.8 | 20 - 2.7 |
| No reflections | - | 30476 | - | 9666 | 18235 | 15540 |
| <i>R</i> _{work} / <i>R</i> _{free} | - | 0.135 / 0.162 | - | 0.184 / 0.248 | 0.246 / 0.299 | 0.194 / 0.261 |
| Number of atoms | | | | | | |
| Protein | - | 819 | - | 1865 | 4884 | 4243 |
| Ligand/ion | - | 6 | - | 12 | - | 50 |
| Water | - | 147 | - | 11 | - | 45 |
| <i>B</i> -factors | | | | | | |
| Protein | - | 11.73 | - | 50.14 | 36.26 | 55.25 |
| Ligand/ion | - | 22.63 | - | 61.42 | - | 47.46 |
| Water | - | 32.10 | - | 30.41 | - | 41.76 |
| R.m.s. deviations | | | | | | |
| Bond length (Å) | - | 0.015 | - | 0.013 | 0.007 | 0.010 |
| Bond angles (°) | - | 1.722 | - | 1.363 | 1.039 | 1.243 |

* Values in parenthesis for outer shell.

Table 13: Summary of Hip data collection and refinement statistics.

4.5.6 Structure of HipM

HipM comprising residues 78-247 is composed of two parts, the N-terminal extension (residues 78-106) and TPR module, consisting of three TPR repeats (residues 107-211) and a capping helix $\alpha 10$ (residues 214-241) (Fig. 25A). Interpretable electron density was visible for residues 81-241 in all four, almost identical copies of HipM in the asymmetric unit of crystal form II (Fig. 25B).

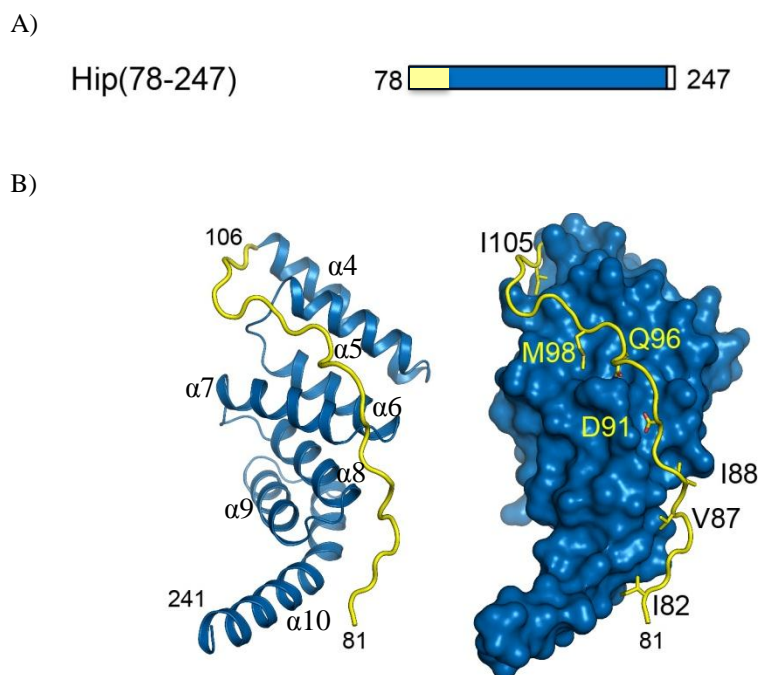


Figure 25: Crystal structure of the Hsp70-binding domain of Hip, HipM.

- A) Representation of the architecture of the HipM domain. N-terminal extension (residues 81-106) is highlighted in yellow. The core TPR domain is shown in dark blue.
- B) Left panel: Cartoon representation of the overall structure of HipM. Right panel: Detailed view of the interactions between the N-terminal extension and the core TPR domain. The N-terminal extension is shown against the molecular envelope of the TPR domain. Selected side chains in the N-terminal linker extension are labeled and highlighted in stick representation.

The N-terminal extension traverses the helix bundle in the groove between helices $\alpha 5$ and $\alpha 7$ of TPR core. Subsequently, it sneaks along the opposite edge to reach the C-terminal capping helix $\alpha 10$, which protrudes beyond the classical TPR domain structure, bringing N- and C-terminal segments into close proximity (Fig. 25B). As a result of this arrangement, the negatively charged residues 81-85 are placed in the vicinity of the positively charged C-terminus of

helix $\alpha 10$. The N-terminal segment is firmly attached to the TPR core by extensive hydrophobic (Ile82, Val87, Ile88 and Ile105) and polar contacts (Asp91, Gln96 and Met98) (Fig. 25B). The buried surface area is 1086 \AA^2 , suggesting a stable interaction between the segments in HipM. Consistently, HipM also appeared stable in solution, which was determined by CD spectrometry with a melting point of $56 \text{ }^\circ\text{C}$ (data not shown).

Notably, truncation of N-terminal residues 78-106 might destabilize the TPR domain by exposing large hydrophobic surface areas. This is consistent with the insolubility of the construct Hip(107-267). Remarkably, the C-terminal extended final helix $\alpha 10$ might not be stably structured. This is suggested by two lines of evidence. A previous study found that Hip was proteolytically truncated at residues 226 and 237 (Velten et al., 2002). Furthermore, the final helix $\alpha 10$ is not present in crystal form I (Fig. 26A), indicating that it was disordered or cleaved during crystallization. Perhaps prior proteolysis in the N-terminal extension is required to stabilize helix $\alpha 10$. Figure 26 provides an structure overview of the two crystal forms of HipM.

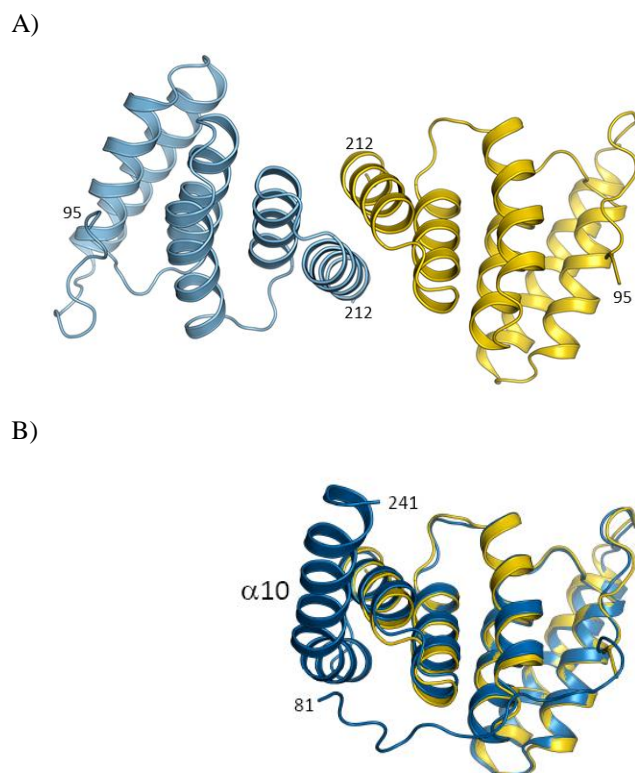


Figure 26: Comparison of the two crystal forms of HipM.

A) Structure of the HipM dimer in crystal form I. The two segments are shown in ribbon representations in light blue and gold, respectively. Residue numbers are indicated for the termini of the ordered segment.

-
- B) Superposition of the HipM units in crystal forms I and II. For clarity, only one representative chain from each crystal form is shown. The chain from crystal form II is shown in dark blue. The additional helix is indicated as $\alpha 10$.

Form I contained two protein chains per asymmetric unit, forming a symmetrical dimer (Fig. 26A). Only residues 95-212 were ordered in both copies, i.e. density for the capping helix $\alpha 10$ was completely absent. Otherwise, the structured part was closely similar to crystal form I as shown in Figure 26B. Subsequent SDS-PAGE analysis of dissolved crystals revealed an approximate 1:1 mixture of two chain lengths, one similar to full-length HipM (data not shown), suggesting partial proteolytic degradation. The exact composition of the crystals remains therefore unclear.

Size exclusion chromatography gave no indication for dimerization of HipM in solution (Fig. 16A). The buried surface area of $\sim 600 \text{ \AA}^2$ at the interface is probably too small for a stable interaction. Two opposed cysteines were found at the interface, however no disulfide bond formation was observed. Therefore, this dimer formation is probably a consequence of crystal packing.

4.5.7 Small angle X-ray scattering (SAXS)

Since full-length Hip could not be crystallized, the structural features of Hip as a whole remained unknown. Small angle X-ray scattering (SAXS) analysis offers complementary information about macromolecular folding, conformation and assembly state in solution with resolution range of about 50 \AA to 10 \AA . Together with the solved three-dimensional structures of individual domains of Hip, we attempted to create a model for a complete Hip dimer.

Mainly two parameters can be estimated from SAXS data, the particle radius of gyration R_g and the maximum linear dimension of the scattering particle D_{\max} . R_g can be obtained by curve-fitting with the programs Guinier and GNOM using Guinier's law and D_{\max} can be calculated with GNOM. When the atomic structures are available, the theoretical curves can be calculated from the pdb files by evaluating all the interatomic distances in the structure. In the direct comparison of such theoretical curves with SAXS data, the Chi value (χ^2) is used to estimate the

agreement between the atomic model with the experimental data from SAXS, which is similar to the R-factor in crystallography by indicating the goodness-of-fit.

Following this methodology, an R_g of 6.05 nm was identified for full-length Hip. The respective pair distance distribution function $p(r)$ is shown as insert, suggesting a large D_{\max} of 202 Å (Fig. 27A). These dimensions are consistent with previous observations (Dores-Silva et al., 2012; Velten et al., 2000), suggesting an elongated conformation for the complete ~ 80 kDa Hip dimer with a maximum extension of ~ 200 Å (Fig. 27A). The Kratky plot representation of the full-length Hip data indicated that a substantial fraction of Hip is unstructured (Fig. 27B), which is consistent with the secondary structure composition derived from CD spectroscopy.

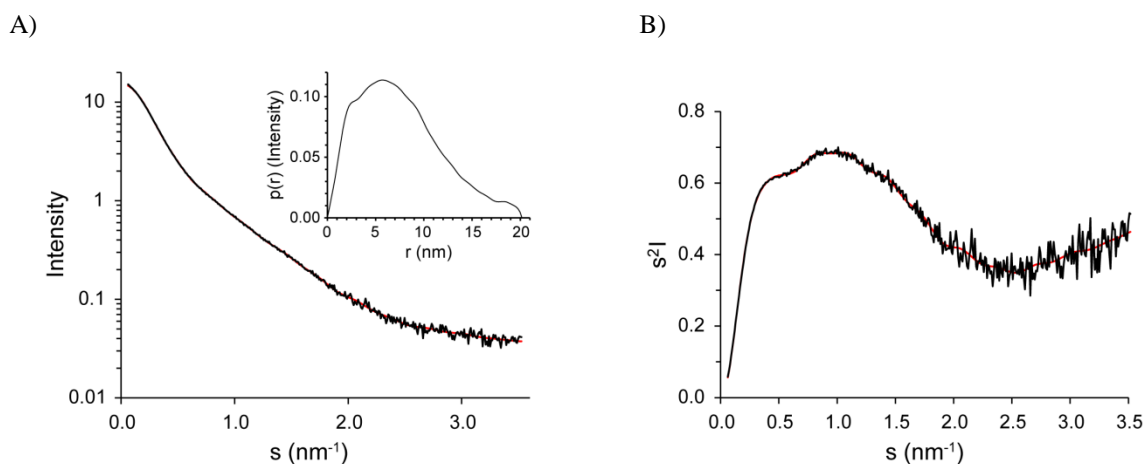


Figure 27: SAXS Structural analysis of the Hip dimer.

- A) X-ray scattering curve of Hip at 1.8 mg/ml in the presence of 20 mM HEPES, 100 mM KCl and 1 mM DTT, pH 7.4. The red line indicates the best fit obtained with the indirect Fourier transform method using the program GNOM (Svergun, 1992). The respective pair distance distribution $p(r)$ is shown as insert, suggesting a large D_{\max} of 202 Å for the 80 kDa particle. The radius of gyration (R_g), 6.05 nm, was determined using the Guinier approximation, as implemented in PRIMUS (Konarev et al., 2003).
- B) Kratky plot representation of the scattering data. The high signal of $s^2 \cdot I$ at $s > 2.5$ nm indicates that a substantial fraction of Hip is unstructured.

Besides full-length Hip, two Hip truncation mutants, Hsp70 and the HipM-Hsp70 fusion protein were also studied (data not shown). Table 14 summarizes the key results derived from SAXS data.

| Construct | SAXS Scattering curve | | Atomic model | | Chi fit (χ^2) |
|-------------|-----------------------|----------------|--------------|----------------|----------------------|
| | R_g [nm] | D_{max} [nm] | R_g [nm] | D_{max} [nm] | |
| Hip | 6.05 \pm 0.04 | 20.18 | – | – | – |
| Hip(1-267) | 5.08 \pm 0.01 | 17.77 | – | – | – |
| Hip(78-247) | 2.25 \pm 0.01 | 8.31 | 1.78 | 6.50 | 14.64 |
| Hip(78-234) | 1.98 \pm 0.004 | 6.33 | 1.68 | 6.02 | 10.78 |
| Hsp70N | 2.44 \pm 0.01 | 8.39 | 2.10 | 7.17 | 5.197 |
| HipM-Hsp70N | 2.73 \pm 0.01 | 9.56 | 2.54 | 8.51 | 4.567 |

Table 14: Comparison of structural data from SAXS and crystallographic analysis

Both Hip(78-247) and Hip(78-234) showed quite high Chi values. One explanation could be that these two constructs might show more open conformations in solution than suggested from the crystal structure of HipM, Hip(78-247). Especially the SAXS data for the functional fusion protein HipM-Hsp70N are in good agreement with the determined Hip:Hsp70 core complex crystal structure (see below). Furthermore, the D_{max} of the fusion protein is similar to Hsp70N, indicating a compact conformation of this protein in solution.

4.6 Structural analysis of the Hip:Hsp70 complex

4.6.1 Crystallization of the Hip:Hsp70 core complex

To gain further insight into the mechanism by which Hip stabilizes Hsp70, we determined the crystal structure of Hsp70 in complex with Hip.

Since the individual crystal structures of Hsp70N and the Hsp70-binding domain of Hip, HipM, were known, we first tried to co-crystallize Hsp70N and HipM. ITC experiments had shown that the binding ratio of these two proteins was 1:1 with a modest binding affinity ($K_D = 27 \mu\text{M}$). The co-crystallization trials were first performed at a molar ratio of 1:1. In order to increase complex formation, we modified the molar ratio of 1:1.2 and 1:1.5, assuming that HipM was less likely to crystallize by itself than Hsp70N, which crystallized readily. Unfortunately, the co-crystallization of Hip with Hsp70N failed. The crystals invariably contained only Hsp70N in the apo or ADP-bound form, depending on whether exogenous ADP was added or not (data not shown). To obtain the structure of a Hip:Hsp70 complex, we therefore changed

our strategy by employing the fusion proteins of HipM and Hsp70N. The proximity of the domains was assumed to enforce intramolecular interactions. As described above, we designed two different types of fusion proteins, the functional HipM-Hsp70N and the rather non-functional Hsp70N-HipM (Fig. 21A). While crystals could not be obtained from the HipM-Hsp70N fusion protein, the reverse construct, Hsp70N-HipM, produced crystals. These appeared as needles in the initial screen with 0.2 M NH_4I and 20 % PEG 3350 as a precipitant at both 4 °C and 18 °C (Fig. 28A & 28B). To improve crystal quality, the crystallization condition was optimized and micro-seeding was performed at 18 °C. A single rod crystal could be obtained by micro-seeding with 0.2 M NH_4I and 17 % PEG 3350 as precipitant using the hanging drop method (Fig. 28D).

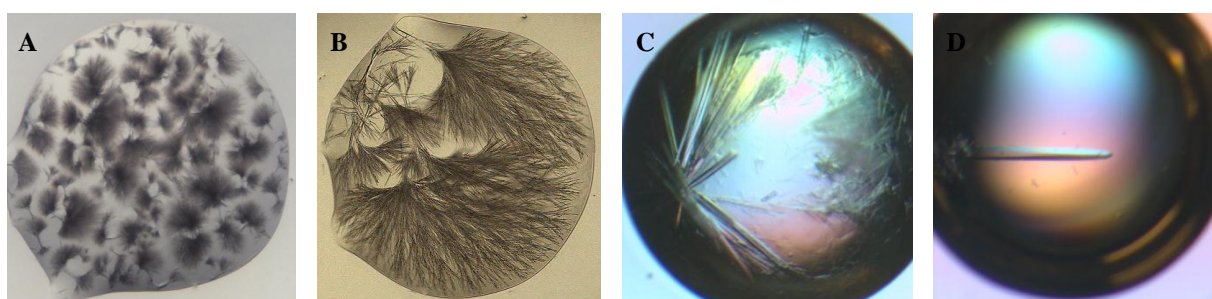


Figure 28: Representative images of the HipM-Hsp70N crystal form.

- A) Initial crystals obtained in the sitting-drop setup under the condition using 0.2 M NH_4I and 20 % PEG 3350 at 4 °C
- B) Initial crystals obtained in the sitting-drop setup under the same conditions at 18 °C
- C) Crystals obtained in the hanging-drop setup with of 0.2 M NH_4I and 17 % PEG 3350 at 18 °C
- D) A single crystal obtained after seeding in the hanging-drop setup using 0.2 M NH_4I and 17 % PEG 3350 as a precipitant.

4.6.2 Structure determination of the Hsp70-Hip fusion protein

X-ray diffraction data were collected from the single rod crystal to 2.7 Å resolution. The analysis showed that the space group is $P4_1$ with lattice parameters of $a \cdot b \cdot c = 65.5 \text{ \AA} \cdot 65.5 \text{ \AA} \cdot 141.47 \text{ \AA}$ (Table 13). The structure of the Hsp70N-HipM fusion protein was solved by molecular replacement using the program Molrep. Hereby, the Hsp70(1-393)-ADP complex and the HipM monomer form I were used as the searching templates. Hsp70N-HipM was almost fully defined in the density including the human Hsp70 (residues 1-382), a short linker and rat Hip (residues 78-243), as well as bound $\text{ADP} \cdot \text{P}_i \cdot \text{Mg}^{2+}$. 17 structurally defined iodine ions from the

precipitant solution were confirmed with the anomalous signal in a dataset collected from a different crystal at 2.0 Å wavelength. Several of these iodide ions are located at the domain-domain interface and at crystal contacts, presumably stabilizing crystal packing interactions (Fig. 29A & 29B). Therefore, the presence of 200 mM NH₄I in the crystallization condition presumably plays an important role in crystal stabilization.

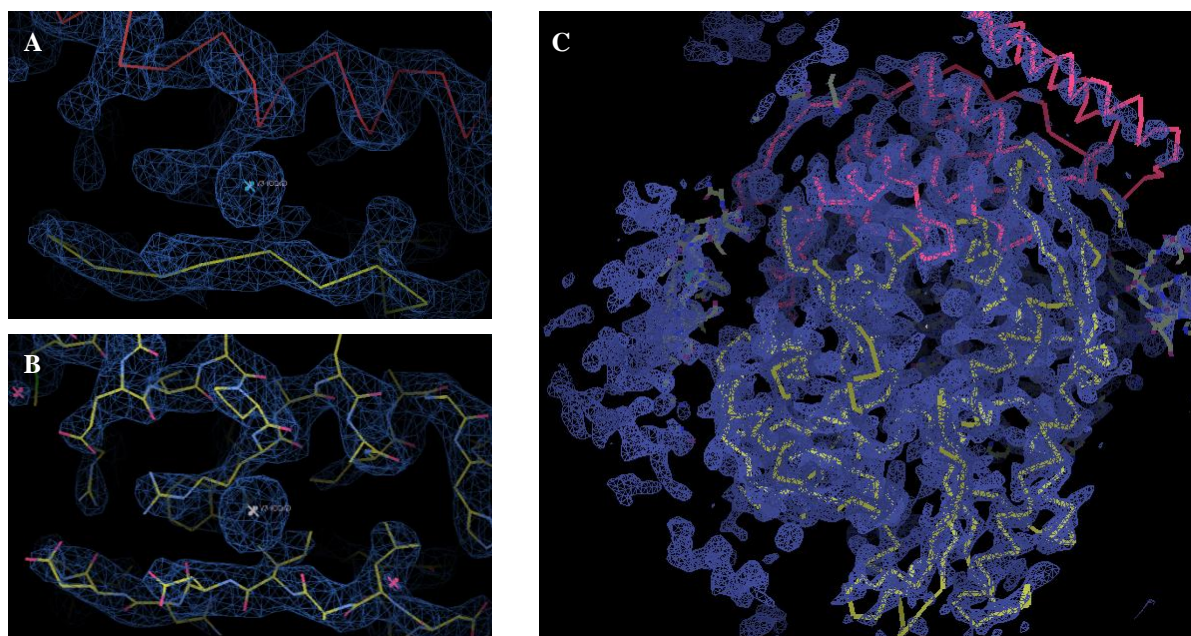


Figure 29: Electron density map of the fusion protein Hsp70N-HipM.

- A) Representative portion of an initial electron density map. The intermolecular interface of HipM (red), Hsp70N (yellow) and resolved iodine ion shown in cross.
- B) The final structural model is overlaid and shown as color coded stick representation.
- C) The representative intermolecular interface of the fusion protein in the initial electron density map.

The complex structure was refined to an R-factor of 19.4 % (R_{free} of 26.1 %). All models were built interactively using the program Coot. Refinement was performed with the program REFMAC. Residues (apart from glycine) facing solvent channels were modeled as alanine, when side chain density was not discernible. Interestingly, besides the intramolecular interface within the fusion protein Hsp70N-HipM, the likely intermolecular interface between Hsp70N and HipM could be observed in the electron density map (Fig. 29C).

4.6.3 Two interaction interfaces between Hip and Hsp70

In the electron density map, two plausible interfaces between Hsp70N and HipM domains were observed. Figure 30A and 30B show the two different interfaces between Hip and Hsp70 together with the surface conservation in Hip at the respective region.

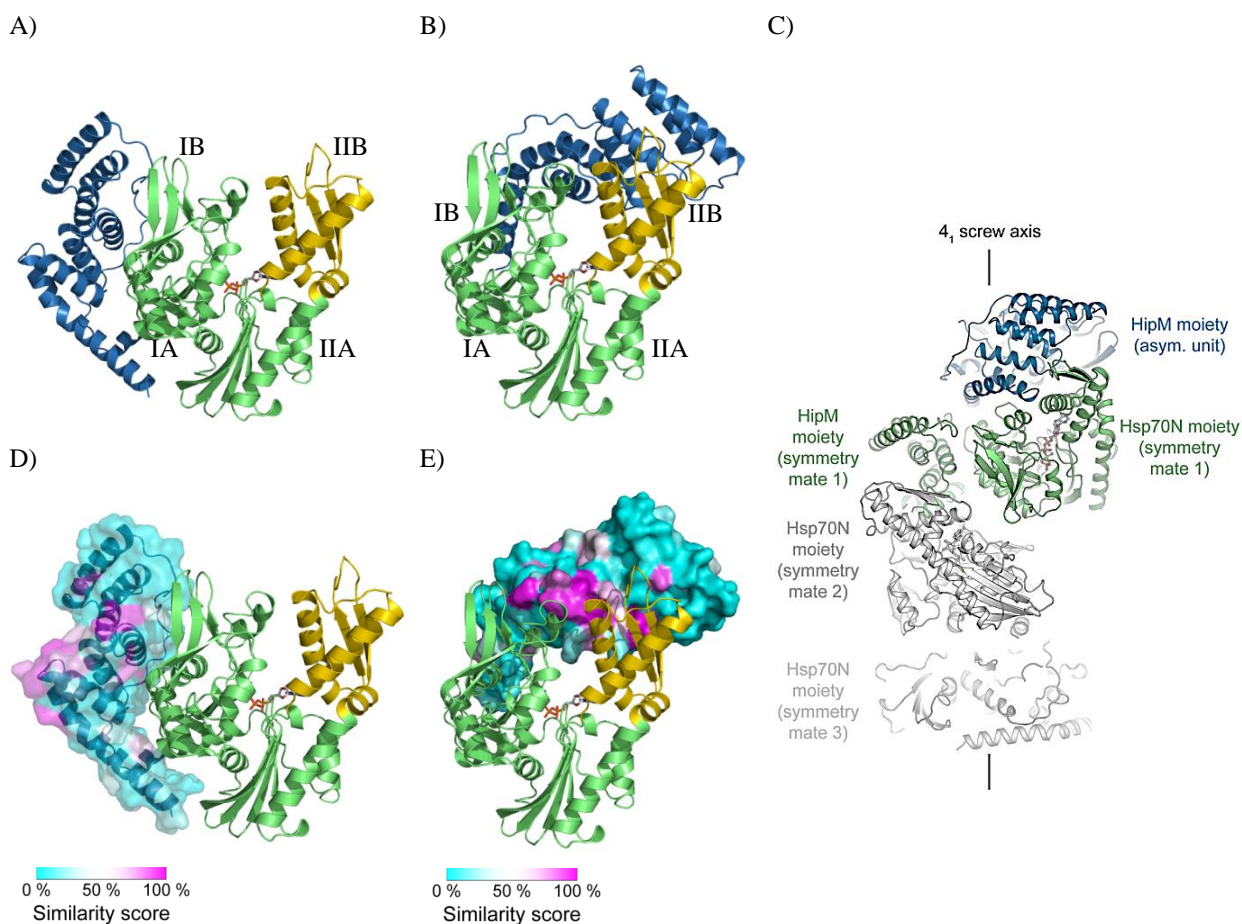


Figure 30: Intramolecular- and intermolecular interfaces of the Hsp70N-HipM fusion protein.

- A) Crystal structure of the Hsp70N-HipM fusion protein showing the intramolecular domain-domain interface. The protein structure is shown in ribbon representation. The Hsp70N and HipM segments are colored green and dark blue, respectively; subdomain IIB of Hsp70N is highlighted in yellow. The bound ADP is shown in stick representation.
- B) Intermolecular interface between HipM and Hsp70N with functional contacts.
- C) Packing of Hsp70N-HipM fusion protein units in the crystal lattice. The functional contact between Hsp70N and HipM moieties from symmetry mates in the crystal lattice is high-lighted in blue (HipM partner) and green (Hsp70N partner). The fusion proteins pack via these functional HipM-Hsp70N contacts into supra-helical fibers along the 4_1 vertical screw axis.
- D) Lack of surface conservation of HipM at the intramolecular interface of the non-functional Hsp70N-HipM fusion protein.
- E) Increased surface conservation of HipM at the intermolecular interface of functional Hsp70N-HipM fusion protein.

4.6.3.1 Intramolecular interface

The fused Hsp70N and HipM domains were tightly associated in the crystal structure (Fig. 30A). The observed intramolecular domain-domain interface buried 949 Å² of surface area, potentially sufficient for a stable interaction. Hip is located on the side of subdomain I of Hsp70 and makes no contacts to subdomain IIB of Hsp70N, which is a well-known to regulate access to the nucleotide binding pocket (Liu et al., 2010). It is hard to see how this binding mode of Hip could stabilize Hsp70 in the ADP-bound state. The interaction of Hip with Hsp70 is on the convex face of the TPR domain instead of the concave face (Fig. 30D). Moreover, this interface proved to be poorly conserved, which is in conflict with surface sequence conservation normally found at protein-protein interfaces because of co-evolution of the binding partners. Of note, the surface of the NBD of Hsp70 is strongly conserved all over. Together this raises strong doubts on the functional relevance of the intramolecular interface.

Moreover, this interaction is apparently forced by favorable contacts to the artificial linker between Hsp70 and HipM, residues Leu76 and Glu77, which were introduced during cloning, the respective sequence serving as a restriction site between the Hsp70 and HipM coding sequences (Fig. 31).

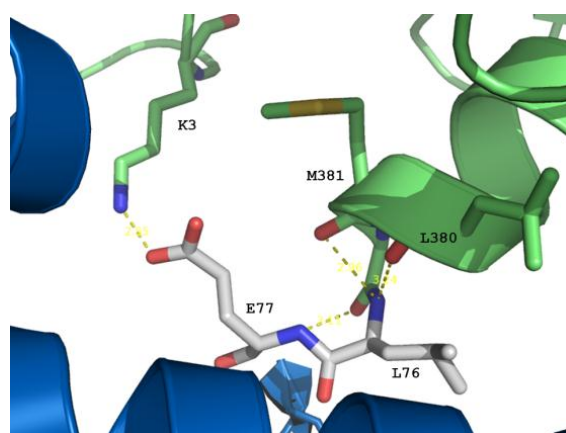


Figure 31: Detailed interactions between the artificial linker, Hsp70 and HipM.

The linker residues undergo multiple polar interactions. Back bone hydrogen bonds are formed between Leu76/Glu77 and Leu380/Met381 at the C-terminus of Hsp70N (color green), respectively. In addition, a likely salt bridge between Glu77 and side chain of Lys3 of Hsp70 was observed.

Remarkably, the linker was structured, forming an extension to the C-terminal final helix of Hsp70N. To prove that the intramolecular interface is without function, the double mutation H196S/E199A targeting interruption of the intramolecular interface was investigated in our functional assays (see below). We confirmed that this mutation is indeed neutral, suggesting that the contact is an experimental artifact.

4.6.3.2 Intermolecular interface

Surprisingly, an intermolecular interface of Hsp70N-HipM fusion proteins forming a contact between symmetry mates in the crystal lattice showed the characteristics of a functional interface (Fig. 30B & 30E). The interaction buries 1039 Å² surface area, similar to the intramolecular interface (949 Å²). Moreover, the surface of HipM shows the highest conservations score between residue 176-181 and 210-214 (Fig. 32), which contact subdomains IA, IB and IIB of Hsp70N in the intermolecular interface (Fig. 30B & Fig. 30E). This is in line with the regulatory role of subdomain IIB of Hsp70. The intermolecular contact therefore provides a promising candidate for the functionally relevant interface between Hip and Hsp70.

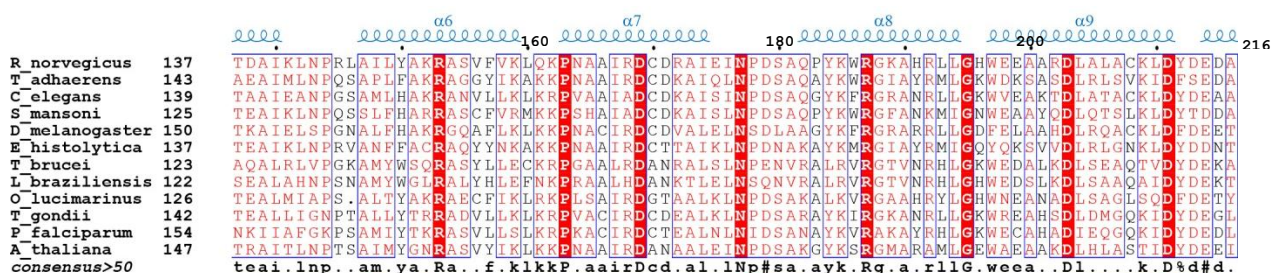


Figure 32: Evolutionary conservation of the central part of HipM, from residues 137 to 216.

In this section of the alignment of Hip sequences similar residues are shown in red and identical residues in white on red background, respectively.

4.6.4 Structure of the Hip:Hsp70 core complex

As mentioned above, the crystal lattice suggested a promising candidate for the interface between Hip and Hsp70. This interface and some key residue contacts are described in detail be-

low (Fig. 33). We later confirmed the functional significance of this interface by mutational analysis (see below).

The NBD of Hsp70 is thought to be conformationally dynamic, with subdomain IIB acting as a mobile regulator for the nucleotide (Liu et al., 2010). The crystal structure of HipM:Hsp70 core complex showed that Hip forms a bracket over the NBD, arresting the otherwise mobile subdomain IIB in a closed conformation and locking the nucleotide in the binding pocket (Fig. 33A).

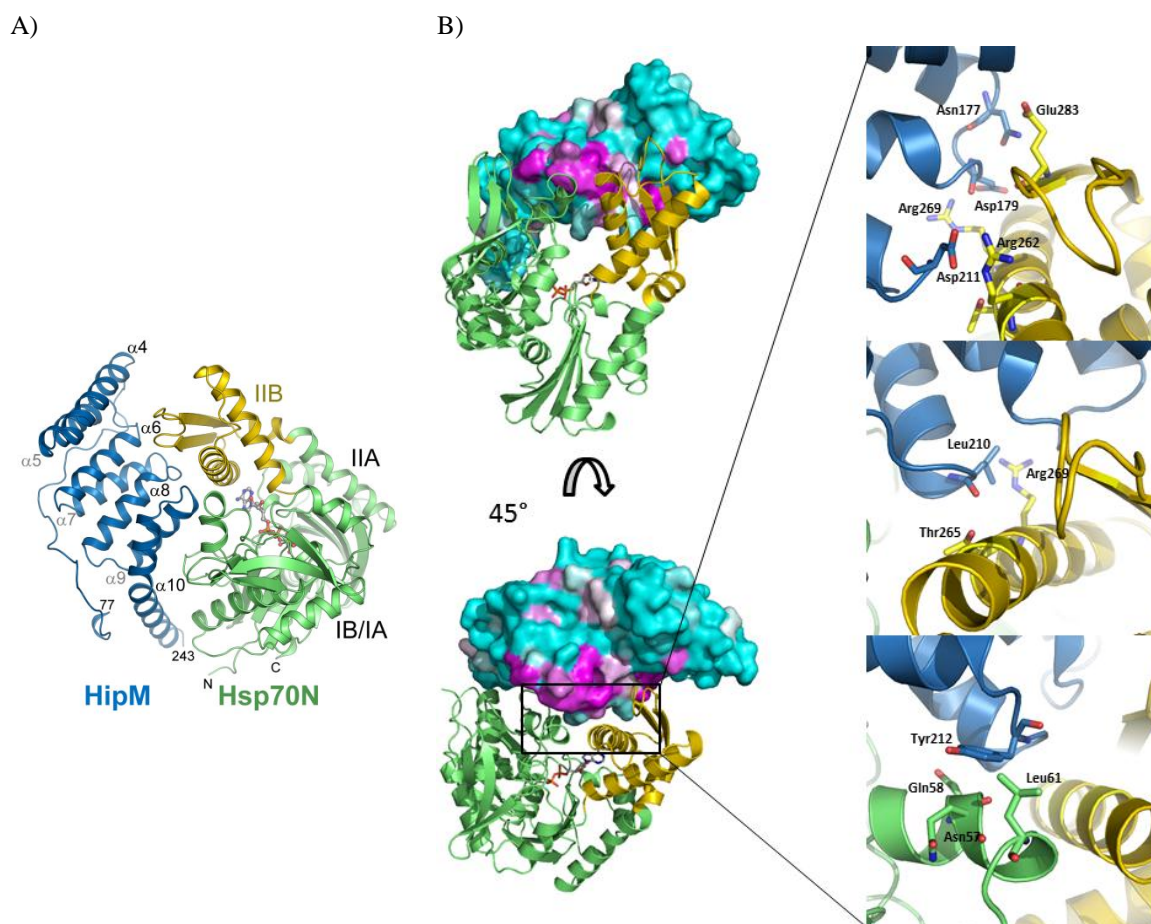


Figure 33: Structure of core complex of Hip and Hsp70.

- A) Ribbon representation of the HipM:Hsp70 complex. HipM and Hsp70N are colored blue and green, respectively. Subdomain IIB of Hsp70N is highlighted in yellow. The bound ADP-phosphate-Mg²⁺ complex are shown in ball-and stick representation. The subdomain structure of the Hsp70N is indicated.
- B) Left panel: Surface conservation of the interface between HipM and Hsp70N. The similarity score from thesequence alignment (Appendix 8.2) is mapped onto the molecular surface of HipM using a color gradient from magenta (high similarity) to cyan (low similarity). Right panel: Key contacts between HipM and Hsp70N are shown in detail.

The conformation of Hsp70N in complex with HipM is virtually identical to that of the ADP·Mg·P_i complex of the NBD of bovine Hsc70 (Wilbanks and McKay, 1995), with the nucleotide being deeply buried in the center of the structure (Fig. 34A). Conversely, Hsp70 NEFs accelerate ADP release by stabilizing Hsp70N in an open conformation.

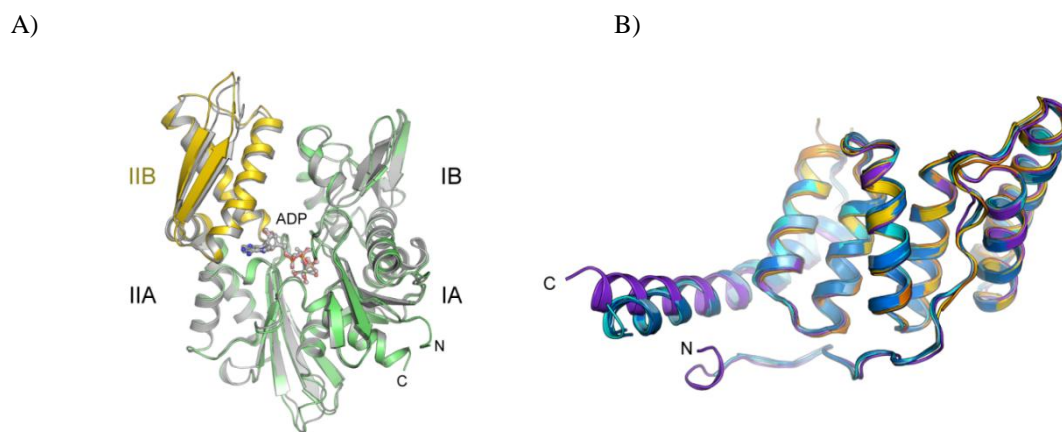


Figure 34: Analysis of Hip:Hsp70 interaction.

- A) Superposition of Hsp70N from the Hsp70N-HipM structure with the ADP·P_i·Mg²⁺ complex of the NBD from bovine Hsc70. The latter structure is shown in gray (PDB code 1HPM (Wilbanks and McKay, 1995)). The subdomain structure is indicated. The Hsp70N fragment has essentially the same structure with an r.m.s.d. of 0.674 Å, consistent with Hip recognizing and stabilizing the ADP-bound state.
- B) Superposition of HipM structures. The copies of HipM in the asymmetric units of crystal forms I (two copies) and II (four copies) were superposed on the respective segment in the structure of the fusion protein. Backbone traces from crystal forms I and II are indicated by yellow and blue, respectively. The backbone of the Hip segment in the fusion protein is shown in purple. Both of the N- and C-termini are indicated.

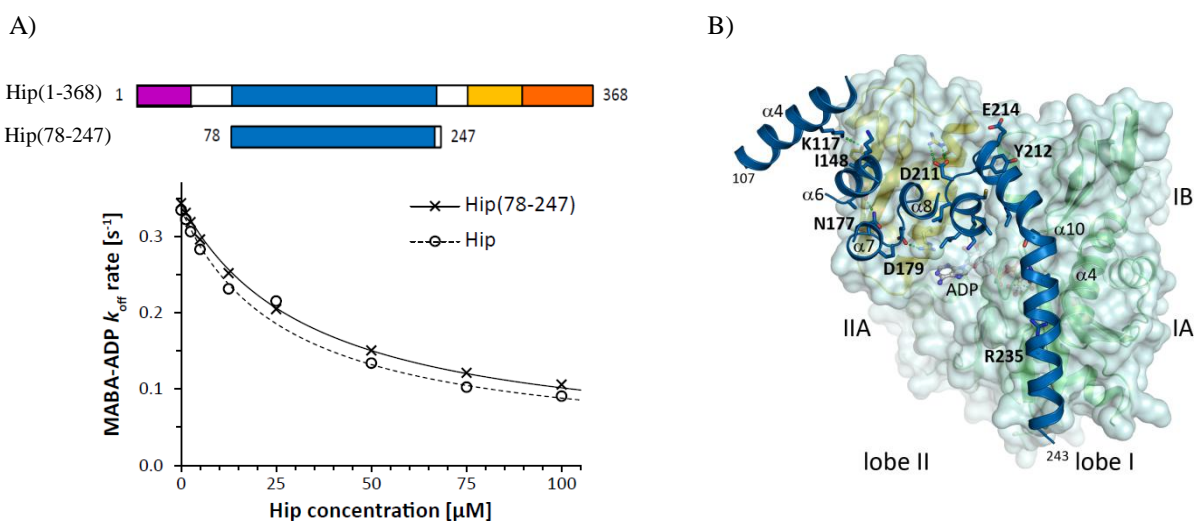
The structure of HipM in the Hip:Hsp70 complex is essentially identical to the structure of HipM alone in crystal form II (r.m.s.d.s 0.834 – 0.873 Å), indicating that the functionally active conformation is favored also in absence of Hsp70. Since the conformation of HipM in the complex with Hsp70N is virtually indistinguishable from that of the isolated domain, the surface of HipM for Hsp70 binding is probably pre-formed (Fig. 34B). The extended interface with Hsp70N at the upper edge of the Hip TPR domain is complementary in shape to the surroundings of the Hsp70 nucleotide binding cleft. This mode of interaction differs markedly from other TPR modules, most of which bind extended peptides in a groove at the concave face of the TPR solenoid (D'Andrea and Regan, 2003).

The intermolecular interface comprises a large number of polar and hydrophobic interactions, shown in right panel of Figure 33B: (i) Asn177 of HipM forms a hydrogen bond with Glu283 in Hsp70N; (ii) Asp179 and Asp211 engage in salt bridges to Arg269 and Arg262 in Hsp70N, respectively; (iii) Tyr212 of HipM is in van-der-Waals contact to residues Asn57, Gln58 and Leu61 in lobe IB of Hsp70N; (iv) Leu210 forms hydrophobic contacts to residues Arg269 and Thr265 of Hsp70N. The interaction of the TPR core domain with subdomains IA, IB and IIB of Hsp70N is reinforced by contacts of the elongated terminal helix $\alpha 10$ of Hip with subdomain IA. This rather shallow interface appears to be stabilized by electrostatic interactions between numerous basic residues (233-HRRKYERKR-241) at the C-terminus of helix $\alpha 10$ and the helix dipole of helix $\alpha 4$ in Hsp70N.

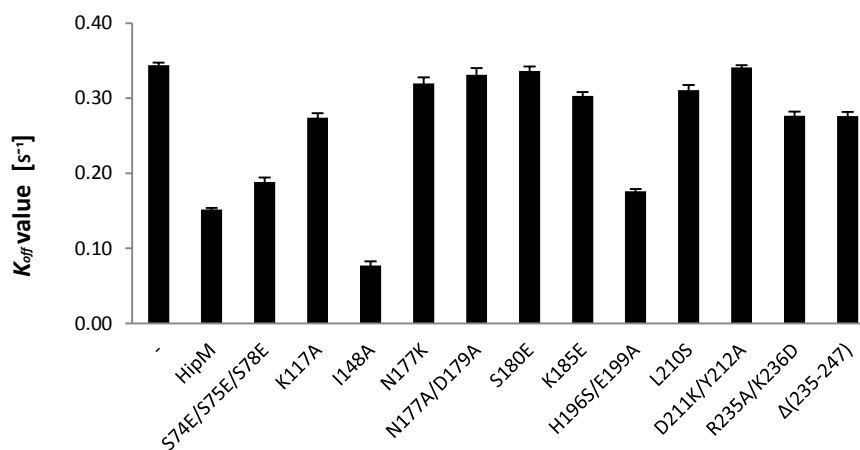
4.6.5 Mutational analysis of the binding interface between Hip and Hsp70

Since a large number of residues within the conserved interface were involved in the interaction between Hip and Hsp70, we next probed the functional significance of the interface by mutation of key residues in Hip, using the MABA-ADP dissociation assay as a read-out.

The mutations were introduced in HipM, Hip residues 78-247 (Fig. 35B), because this fragment exhibited a similar effect on MABA-ADP dissociation as full-length Hip (Fig. 35A). The assay is a suitable proxy for the Hsp70 binding ability.



C)



D)

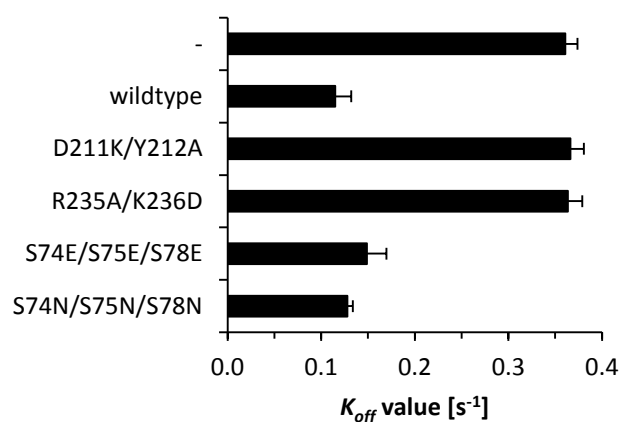


Figure 35: Mutational analysis of the interaction between Hip and Hsp70.

- A) Schematic representation of the full-length Hip(1-368) and Hip(78-247) (HipM) and the analysis of MABA-ADP release assay. Hip-mediated attenuation of MABA-ADP dissociation from Hsp70N. In a stopped-flow apparatus, 2.5 μ M Hsp70N with MABA-ADP complex was mixed with 250 μ M ATP solution containing increasing concentrations of full-length Hip or HipM at 30 $^{\circ}$ C. Apparent k_{off} rates were determined from the exponential decay of MABA-ADP fluorescence and plotted against the final Hip concentration.
- B) Key contacts in the functional HipM:Hsp70N interface. The contacting elements of HipM are shown above the molecular surface of Hsp70N. Interacting side chains are highlighted in stick representation. Key polar contacts are indicated by green dotted lines. The bulk of the HipM TPR domain in the foreground was removed for clarity.
- C) Probing the interface between Hip and Hsp70 by HipM mutation. Hsp70N•MABA-ADP was mixed with a 40-fold excess of the respective HipM mutant. K_{off} rates of MABA-ADP obtained from the resultant fluorescence traces are shown. Error bars represent standard deviations from at least three independent experiments.
- D) Probing the interface of full-length Hip by mutation. Hsp70N•MABA-ADP was mixed with a 40-fold excess of the full-length Hip mutants.

Mutations N177K, N177A/D179A, L210S and D211K/Y212A in the central region of the interface resulted in a virtual loss of the ability of Hip to decelerate the dissociation of MABA-ADP from Hsp70N (Fig. 35B & 35C). The mutation N177K resulted in very weak binding to Hsp70-ADP, as determined by ITC (Table 15). The double mutant D211K/Y212A resulted in virtually complete loss of binding in ITC. The inactivating mutation K185E disrupts a conserved intramolecular salt bridge with Asp213, and thus presumably destabilizes Hip. Mutations K117A, R235A/K236D and $\Delta(235-247)$ at the periphery of the interface exhibited a similar milder functional impairment. The double mutation R235A/K236D at the C-terminus of HipM was intended to prevent dissipation of the dipole moment in $\alpha 4$ helix in Hsp70N by Hip. It induced a similar K_{off} rate as the truncation construct of HipM, $\Delta(235-247)$, consistent with an important role of dissipation of the dipole moment for binding.

| HipM mutation | K_D value [μ M] |
|---------------|------------------------|
| N177K | 192 |
| K185E | 85 |
| D211K/Y212A | n.d. |
| R235A/K236D | 68 |

Table 15: The dissociation constants of selected HipM mutants determined by ITC.

As expected, the mutations showed lower binding affinity to Hsp70N than wild type HipM ($K_D = 27 \mu$ M), consistent with the MABA-ADP release assay.

Notably, mutation L210S (L211S in human Hip) was previously identified in a screen for loss-of-function mutants of human Hip (Place, 2011). In contrast to the above listed mutants, the double mutation H196S/E199A targeting the non-conserved intramolecular interface of the Hsp70N-HipM fusion protein had no effect on HipM activity (Fig. 35C), a result confirming that this interface is not functionally relevant.

Subsequently, two phosphomimetic mutations, S74E/S75E/S78E and S180E, were tested. Cell cycle-dependent phosphorylation of Hip at S74, S75 and S78 was previously detected in a large scale proteomics studies (Olsen et al., 2006). Phosphorylation at Hip residue S180 was independently found (Olsen et al., 2010). We found that the phosphor-mimetic mutant S180E located at the interface was inactive (Fig. 35C). However, the triple mutation S74E/S75E/S78E had activity similar to wild type, which is most likely because of its location far from the Hsp70

interface. Interestingly, the mutation I148A located within the functional Hsp70N-HipM interface was reported before to increase Hsp70 binding activity (Nelson et al., 2004). This surprising finding could be confirmed with our functional assay. The mutation might enhance polar contacts of Lys117 and Lys152 with Asp292, Tyr294 and Asp285 of Hsp70N, respectively. The finding that the I148A mutation has not been selected for during evolution suggests that *in vivo* the affinity of Hip for Hsp70 must be carefully balanced with that of other Hsp70 regulators.

To complete the analysis, we further tested selected mutations in the context of full-length Hip (Fig. 35D). The selected mutations all exhibited the same trend in the functional assay as the respective mutations in HipM. Only the double mutation R235A/K236D showed a more serious functional impairment in full-length Hip compared to the HipM mutant. No clear difference was observed between the phosphor-mimetic S74E/S75E/S78E mutation and the phosphorylation-resistant mutant S74N/S75N/S78N, showing that modification at this site can only indirectly affect Hip activity.

In order to test for subtle differences between mutations, which virtually abolish Hip function in the HipM construct, some of these mutations were introduced into the HipM-Hsp70N fusion protein with the functional intramolecular interface, which has been identified in section 4.4 (Fig. 21).

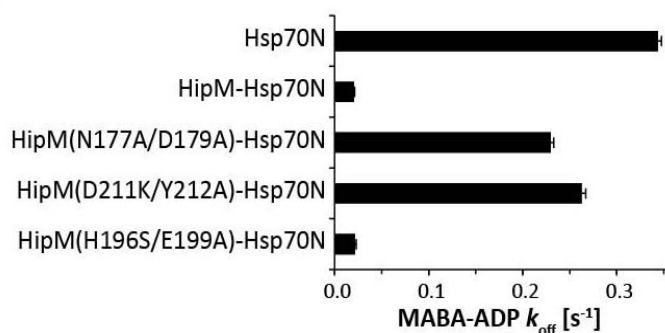


Figure 36: Point mutations in the intramolecular interface of the HipM-Hsp70N fusion protein. Mutants markedly accelerate MABA-ADP dissociation, whereas the mutation H196S/E199A in the HipM-Hsp70N interface is neutral.

The double mutation D211K/Y212A in HipM-Hsp70N almost restored the MABA-ADP off-rate to the level of free Hsp70N (Fig. 36). The double mutant N177A/D179A had a somewhat

milder effect. As expected, the double mutation H196S/E199A in the non-functional interface left the off-rate of HipM-Hsp70N unchanged (Fig. 36).

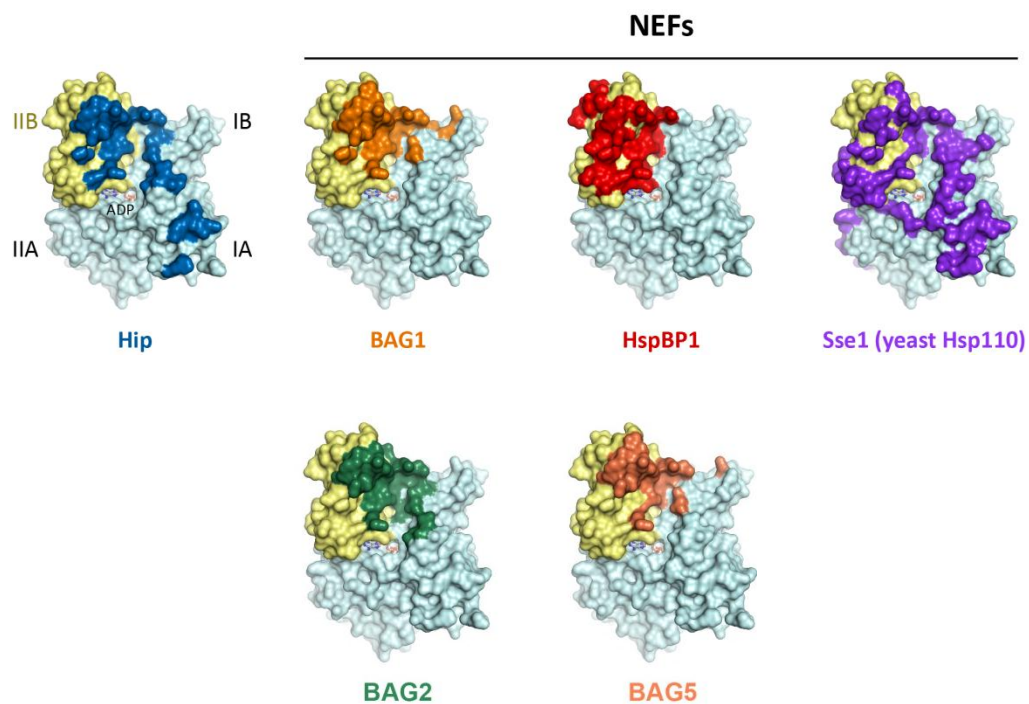
Taken together, the mutational analysis fully validated the intermolecular contacts in the Hsp70N-HipM crystal structure as the functional binding interface between Hip and Hsp70.

4.7 Interplay between Hip and Hsp70 NEFs

Two co-chaperones target the ADP state of Hsp70. Hip thereby stabilizes this state, whereas NEFs stimulate the release of ADP and the progression of the Hsp70 folding cycle. Therefore, we hypothesized that Hip and NEFs might act as direct competitors for Hsp70 binding.

To explore this idea, we mapped the binding areas of Hip and NEFs on Hsp70N based on the solved complex structures. Figure 37A shows that the binding surfaces on the Hsp70 NBD strongly overlap, suggesting mutually exclusive binding of Hip and NEFs to Hsp70 (Polier et al., 2008; Schuermann et al., 2008; Shomura et al., 2005; Sondermann et al., 2001; Xu et al., 2008).

A)



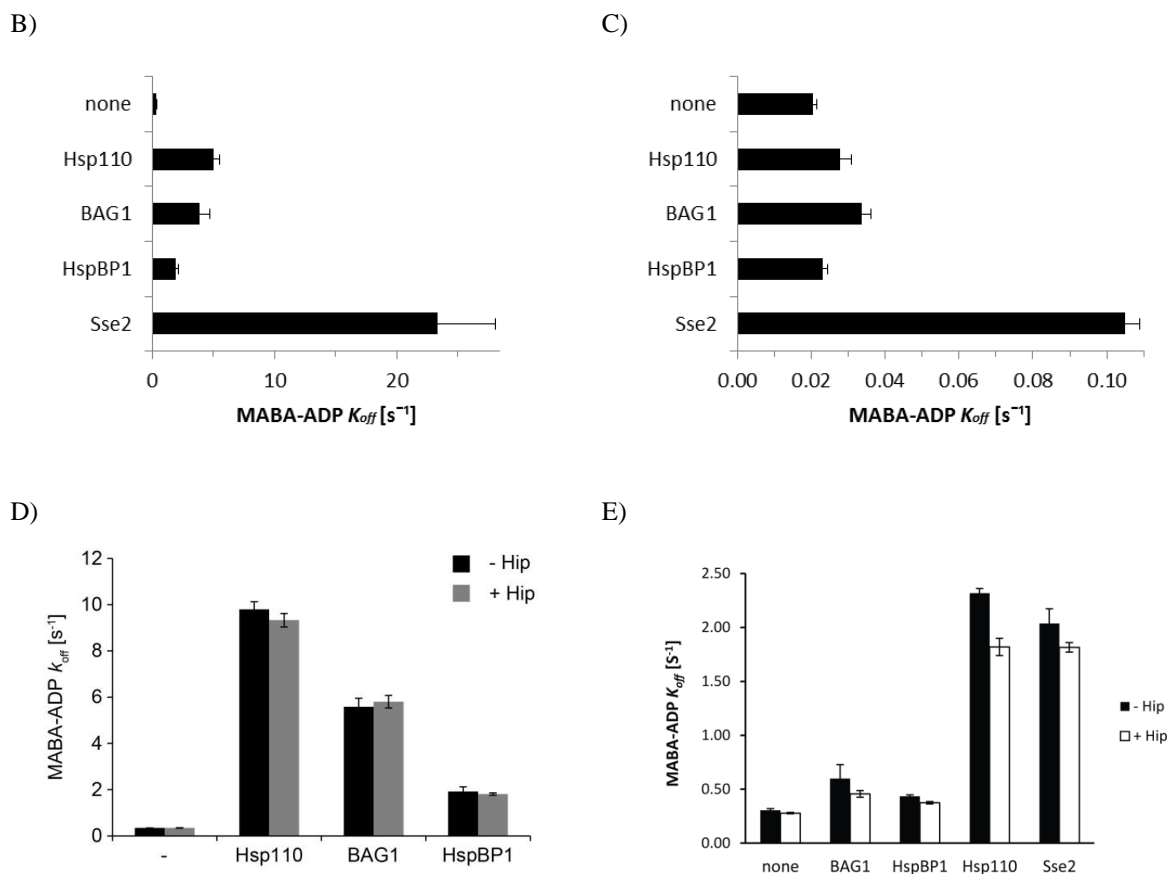


Figure 37: Interplay of Hip and Hsp70 NEFs.

- A) Comparison of Hip- and NEF-binding surfaces on Hsp70N. Residues of the NBD that form contacts with Hip, BAG1, BAG2, BAG5, HspBP1 and Sse1 in the respective crystal structures are indicated on the surface of the human Hsp70NBD in the ADP state. Subdomain IIB of the NBD of Hsp70 is highlighted in yellow. In the crystal structures of NEF-Hsp70 NBD complexes, subdomains IIB are rotated outwards. The nucleotide binding pocket is located in the center of the molecule.
- B) Effect of NEFs on MABA-ADP dissociation from the Hsp70N•MABA-ADP complex in the absence of Hip. Equimolar concentrations (2.5 μ M) of the Hsp70 and NEFs (Hsp110, BAG1, HspBP1 and Sse2) were used in the assay. Error bars represent standard deviations from three independent experiments.
- C) Rates of MABA-ADP dissociation from the functional HipM-Hsp70N fusion protein in the presence of NEFs (Hsp110, BAG1, HspBP1 and Sse2). Equimolar concentrations of the proteins (2.5 μ M) were used. Please note the 100-fold difference in scale compared to panel B
- D) Effect of Hip on MABA-ADP dissociation from Hsp70N in the presence of NEFs (Hsp110, BAG1, HspBP1). Equimolar amounts of the proteins were used (2.5 μ M). MABA-ADP K_{off} rates for Hsp70N (black) and an equimolar mixture of Hip and Hsp70N (gray) are shown. Error bars represent standard deviations from three independent experiments.
- E) Competition at likely cytosolic concentration ratios. The complex of MABA-ADP and Hsp70N (2.5 μ M) was mixed with buffer containing ATP (250 μ M) and NEFs (BAG1, HspBP1, Hsp110, Sse2) (0.5 μ M) either in presence or absence of Hip (2.5 μ M) at 30 °C in a stopped-flow apparatus. K_{off} rates for Hsp70N (black) and an equimolar mixture of Hip and Hsp70N (white) are shown. Error bars represent standard deviations from three independent experiments.

First we tested the stimulatory effect of NEFs on the MABA-ADP dissociation from the Hsp70N•MABA-ADP complex in the absence of Hip. Equimolar amount of Hsp70 and NEFs (Hsp110, BAG1, HspBP1 and Sse2) were studied (Fig. 37B). Compared to spontaneous ADP dissociation from Hsp70 ($k_{off} \sim 0.34 \text{ s}^{-1}$), all the NEFs strongly accelerated ADP release. Addition of equimolar amounts of Hip had almost no effect (Fig. 37D), in line with the roughly ~ 100 -fold higher affinity of NEFs for Hsp70 found in previous studies (Raviol et al., 2006; Shomura et al., 2005; Sonderrmann et al., 2001). However, the maximal stabilization effect of Hip can be observed by employment of the fusion protein HipM-Hsp70N (Fig. 37C). Interestingly, the yeast Hsp110 protein Sse2 competed somewhat more efficiently, perhaps because a Hip homolog does not exist in yeast (Fig. 37B).

Hip was reported to be present in the cytosol at concentrations similar to Hsc70 ($\sim 1 \mu\text{M}$) (Höhfeld et al., 1995), while most NEFs are present only at ~ 10 -fold lower levels. To mimic a more physiological ratio in the MABA-ADP release assay, equimolar amounts of Hip and Hsp70 in 5-fold molar excess to NEFs were used (Fig. 37E). This excess was however not sufficient to suppress NEF action on Hsp70. Therefore, Hip cannot efficiently interfere with the binding of NEFs to Hsp70N under the assay conditions.

Since the free Hip cannot efficiently compete with the binding of NEFs under the assay conditions, we employed the functional fusion protein HipM-Hsp70N as a model for maximum Hip binding (Fig. 38).

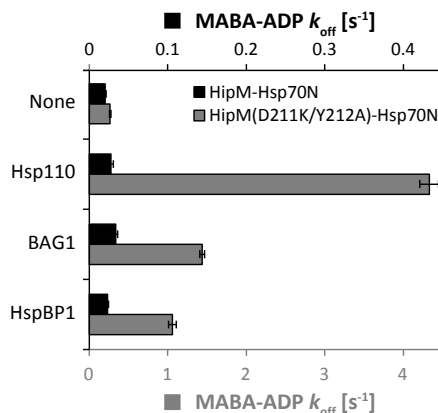


Figure 38: Effect of mutations in the HipM-Hsp70N interface on NEF-triggered MABA-ADP release.

Rates of MABA-ADP dissociation from HipM-Hsp70N fusion proteins in the presence of NEFs (Hsp110, BAG1, HspBP1). Equimolar concentrations of the proteins were used. K_{off} rates for the functional HipM-Hsp70N fusion protein (black) and the inactive mutant HipM(D211K/Y212A)-Hsp70 (gray) are shown with standard deviations from three independent measurements. Note the difference in scale of the x-axes.

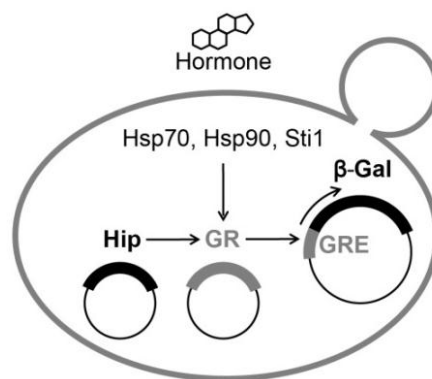
Virtually no acceleration of MABA-ADP release was observed, when stoichiometric amounts of the mammalian NEFs (Hsp110, BAG and HspBP1) were added to the MABA-ADP complex of the functional HipM-Hsp70N fusion protein (black bar in Fig. 38). The k_{off} rate of the MABA-ADP dissociation remained below $\sim 0.04 \text{ s}^{-1}$. The NEF effect was restored in the double mutation HipM(D211K/Y212A)-Hsp70N, which disrupts the functional interface between Hip and Hsp70, reflecting that presence of HipM attached to Hsp70N (gray bar in Fig. 38).

In the presence of saturating concentrations of Hip, the NEFs thus do not effectively bind to Hsp70. When Hip binding is reinforced by other means, Hip should therefore effectively compete with NEFs.

4.8 Hip function *in vivo*

To evaluate the functional relevance of the dimerization and of the putative substrate binding region of Hip *in vivo*, we employed a β -galactosidase reporter (GR). This established *in vivo* model system evaluates the effects of various molecular chaperones on the hormone-dependent maturation of glucocorticoid receptor (GR) in *S. cerevisiae*. (Nelson et al., 2004). In this model system lacking endogenous Hip, GR activity is measured *via* the expression of a steroid-dependent β -galactosidase reporter (Fig. 39A). Almost complete conservation of the Hip contact residues in the yeast Hsp70s Ssa1 and Ssa2 suggests that mammalian Hip interacts functionally with yeast Hsp70. At limiting hormone levels, Hip expression increased the reporter activity ~ 3 -fold, reflecting the ability of Hip to enhance the conformational maturation of GR (Fig. 39C)

A)



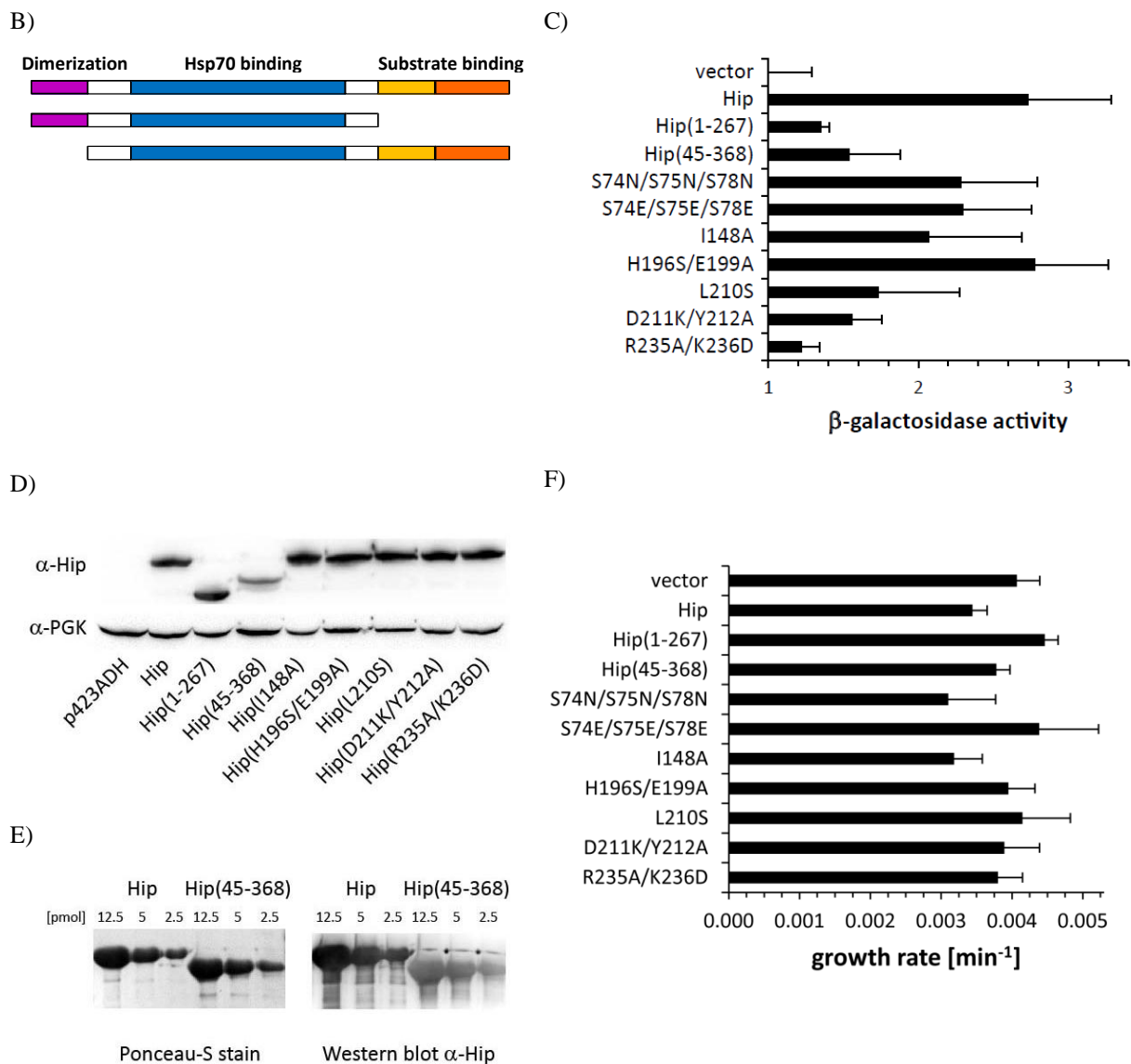


Figure 39: Hip functional study *in vivo*.

- A) The model system. Hip constructs and the glucocorticoid receptor (GR) protein from *R. norvegicus* were co-expressed in a *S. cerevisiae* reporter strain expressing β -galactosidase (β -Gal) under control of the hormone-dependent GR promoter; GRE (Riggs et al., 2003). Hip and GR functionally interact with the yeast Hsp70 and Hsp90 machinery (Hsp70, Hsp90, Sti1).
- B) Schematic representation of the Hip, Hip(1-267) and Hip(45-368) constructs.
- C) Reporter gene activity upon expression of Hip mutants. β -Gal levels were measured upon the expression of Hip constructs. The relative amounts of functional GR compared to control cells without expressing Hip are indicated. Averages and standard deviations (s.d.) are from five independent experiments.

-
- D) Expression levels of Hip variants in yeast cells. Western blot of yeast reporter strains. *S. cerevisiae* cultures were grown to OD 1.5-2.0 at 30 °C in selective medium. Equal amounts of cells were harvested and subjected to alkaline lysis. Cell extracts were separated on 12 % SDS-PAGE and probed by Western blotting against Hip using a polyclonal antiserum (Höhfeld et al., 1995). The strain transformed with p423ADH (empty vector) served as a control for background; phosphoglycerate kinase (PGK) was used as a loading control.
- E) Western blot signal of purified Hip and Hip(45-368). Left panel: Equimolar amounts of purified Hip and Hip(45-368) were analyzed by Ponceau-S protein stain; Right panel: Western blotting with anti-Hip antibody.
- F) The growth rate of the reporter strains. The growth rate for the different Hip variants in liquid shaking culture was analyzed by OD₆₀₀ measurements during the assay.

To dissect the impact of different features of Hip on its *in vivo* functionality, we also expressed selected Hip mutants in the model system. All Hip variants had similar expression levels (Fig. 39D). The growth rates of the reporter strains were closely similar (Fig. 39F). In accordance with previous findings (Nelson et al., 2004), the monomeric construct Hip(45-368) had significantly reduced activity (Fig. 39B & 39C), although the protein was expressed similar to wild-type Hip (Fig. 39D). This finding strongly suggests that avidity plays an important role in the interaction of Hip with Hsp70-substrate complexes. In addition, direct contacts of the substrate with the two arms of Hip might be important. This notion is supported by the poor *in vivo* activity of Hip(1-267), lacking the putative substrate binding region. However, in our *in vitro* MABA-ADP release assay, Hip(1-267) was as active as full-length Hip (Fig. 20E), suggesting that the direct interaction with Hsp70 is unperturbed in this mutant.

The mutants L210S, D211K/Y212A and R235A/K236D, which disrupted the Hip-Hsp70 interface similarly, diminished the ability of Hip to support GR activation. Thus, *in vivo* function of Hip depends on direct interactions with Hsc70 (Fig. 39C). Interestingly, expression of the hyper-active mutant I148A slightly reduced GR maturation relative to wild-type Hip. This is again evidence for careful control of Hip activity *in vivo*. Expression of Hip under a stronger promoter caused a temperature-sensitive phenotype in the reporter system (Nelson et al., 2004). The phosphomimetic S74E/S75E/S78E mutant and the S74N/S75N/S78N mutation used as negative control for *in vivo* phosphorylation were again more-or-less neutral (Fig. 39C), similar to the MABA-ADP release assay (Fig. 35D).

In summary, the *in vivo* studies of Hip suggested that function requires Hip dimerization and substrate binding capacity in addition to the capability to directly interact with Hsc70.

5 Discussion

Protein folding to a well-defined conformation is essential to perform biological function. Moreover, misfolded proteins cause neurodegenerative diseases such as Alzheimer's, Parkinson's and Huntington's disease. The Hsp70 chaperones are central to the cellular defense against toxic protein aggregation through an ATP-dependent reaction cycle, which is regulated by co-chaperones of the J-protein family and by nucleotide exchange factors (NEFs).

Hip, Hsp70 interacting protein, is so far the only known Hsp70 co-chaperone that stabilizes Hsp70-client protein complexes by retarding the dissociation of ADP from Hsp70. In this study, we present the first complete structural model of this novel regulator of metazoan Hsp70 including the crystal structures of the dimerization domain and the TPR domain. We performed an in-depth mechanistic analysis of the interaction of the Hip core domain with the nucleotide binding domain (NBD) of human Hsp70. The structure-guided mutational analysis showed that Hip forms a bracket over the ATPase domain, reducing its otherwise dynamic nature and locking the ADP-bound state. Moreover, Hip directly counteracts the binding of various NEFs, which catalyze ADP dissociation and facilitate substrate release upon ATP re-binding. To become fully functionally effective, Hip must interact not only with the NBD but also with the Hsp70-bound client protein. Our data explains how Hip enhances aggregation prevention by Hsp70 and suggests how Hip would potentially divert the client proteins to the downstream proteostasis machinery, such as Hsp90 or the proteasome.

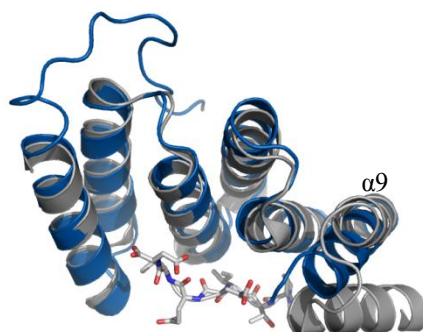
In the following sections, we contrast and compare the results of this study with previous studies and present an integrated model for the Hsp70 mechanism.

5.1 Comparison of the TPR domains in Hip and Hop

The basic function of TPR domains is to mediate protein-protein interactions, particularly facilitating interactions with heat shock proteins. The binding residues are normally presented on the concave face of the TPR solenoid and bind with high specificity to the target peptide (D'Andrea and Regan, 2003). Hop is the paradigm for TPR domain interaction with Hsp70 and Hsp90. The crystal structures of the TPR domain-peptide complexes revealed that Hop specifi-

cally recognizes the C-terminal pentapeptides of Hsp70 and Hsp90 through distinct TPR domain modules (Scheufler et al., 2000).

A)



B)

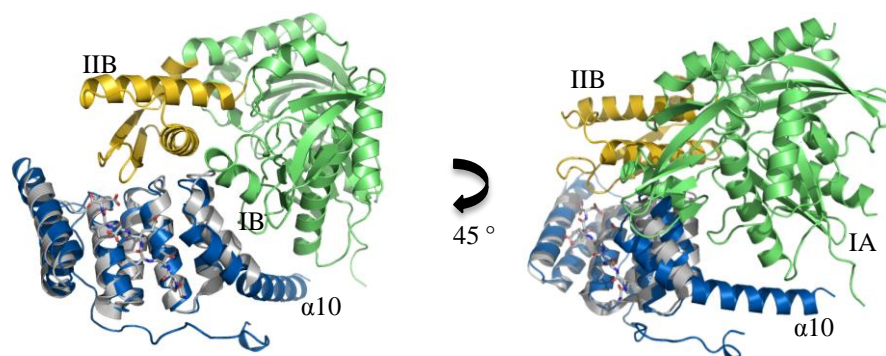


Figure 40: Comparison of the binding sites in the TPR domains of Hip and Hop.

A) Overlay of crystal form I of Hip(78-247) (dark blue) and the Hop-TPR1 peptide complex (gray). The proteins are shown in ribbon representation. The backbone of the Hop-bound peptide is represented as stick model. The last α -helix of Hop is absent in form I of Hip(78-247).

B) Ribbon representation of the overlay between the complex structures of the Hip-Hsp70 and Hop-peptide. Hsp70N is shown in green with subdomain IIB in gold. Hip(78-247) is shown in blue.

In comparison to the TPR1 domain of Hop, the solenoid formed by the TPR repeats in Hip has a more pronounced curvature. However, unlike Hop, Hip does not employ the concave face of the solenoid for binding of Hsp70. It instead employs the upper edge of the solenoid, which is shape-complementary to the Hsp70 surfaces around the closed nucleotide binding cleft in subdomains IB and IIB (left panel in Fig. 40B). In addition, the elongated C-terminal helix α 10 of Hip contacts lobe IA in the NBD of Hsp70 (right panel in Fig. 40B). This rather flat interface appears to be stabilized by electrostatic interactions with the negative end of one α -helix dipole in Hsp70. Substitution of the positively charged residues Arg235 and Lys236 had a surprisingly

strong effect on Hip function (see Table 15, Fig. 35C, Fig. 35D and Fig. 39C). The crystal structures of the middle domain of Hip show that the TPR domain is a versatile structural scaffold for protein-protein interactions that can support both interactions with extended peptide motifs as well as surface motifs in folded proteins.

5.2 Tentative model for the overall structure of Hip dimer

With structural models of the folded domains at hand, we can now construct a tentative model for the complete Hip dimer (Fig. 41).

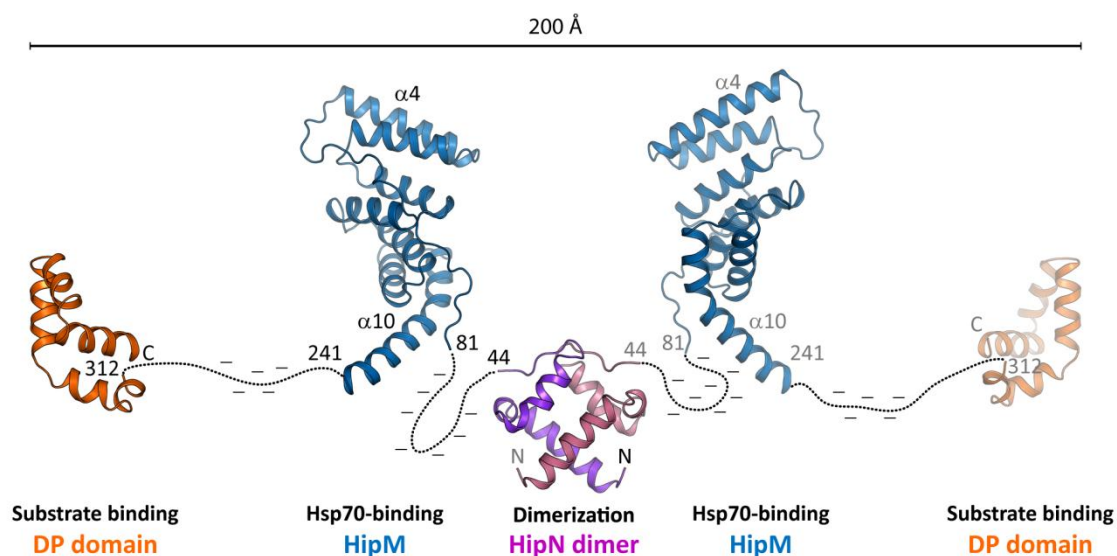


Figure 41: Tentative model of dimeric full-length Hip.

Consistent with previous observations (Dores-Silva et al., 2012; Velten et al., 2000), our small angle X-ray scattering (SAXS) analysis indicated an elongated conformation for the complete ~80 kDa Hip dimer with a maximum extension of ~ 200 Å. In the model, it is important to note that all the folded domain structures are connected by long, negatively-charged linkers, which could span much greater distances (assuming a contour length of 3.6 Å per residue). Considering the elongated shape of HipM, the DP domains at the ends of long tethers might not be resolved in the SAXS experiments at all, because of limited contrast against the solvent background. Electrostatic repulsion between the pronounced negative charges in the linkers might

force the Hip dimer into an open, extended conformation, allowing independent and simultaneous binding of two Hsp70 molecules at each arm of the dimer.

Analysis of the CD spectra and the Kratky plot suggests the absence of regular secondary structure and disorder for most of the linker parts in Hip. This disorder may provide the conformational flexibility for Hip to recognize a wide range of Hsp70-substrate protein complexes.

The helices of the dynamic DP domains form hydrophobic grooves, which could serve as binding sites for exposed hydrophobic segments in non-native substrate proteins. Since the DP domains are expendable for Hsp70 binding *in vitro*, but important for the enhancement of the conformational maturation of GR *in vivo*, such interactions appear to play an important role for Hip functionality. This appears to work however only in dimeric Hip, suggesting an avidity-enhanced binding mechanism requiring multiple recognition sites in the substrate complex.

5.3 Specific recognition of the Hsp70 ADP state by Hip

Consistent with the earlier proposal by Höhfeld et al. in 1995, our detailed ITC analysis demonstrated preferential binding of Hip to ADP-bound Hsp70 ($K_D = 8 \mu\text{M}$), compared to the apo- and ATP states of Hsp70 ($K_D = 51 \mu\text{M}$ and not determinable, respectively). Our analysis showed that the presence of ATP in the binding pocket of Hsp70 as such had only a minor influence.

Superposition of the Hip-Hsp70 core complex with the recently solved crystal structure of a mutant full-length bacterial Hsp70 (DnaK) in complex with ATP indicates how Hip might discern the ADP-bound from the ATP-bound form of the molecular chaperone (Kityk et al., 2012) (Fig. 42).

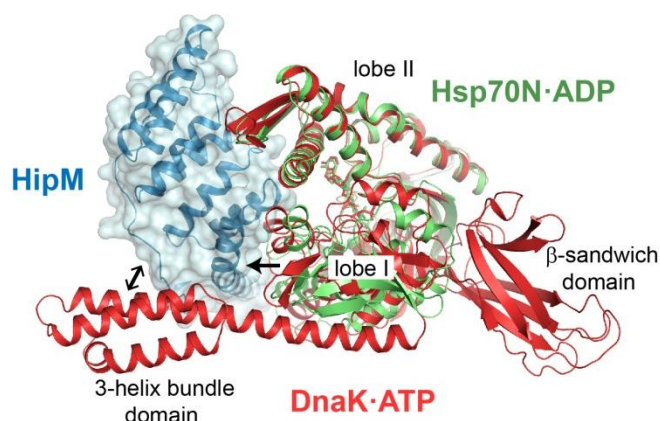


Figure 42: Comparison of the crystal structures of DnaK-ATP and the HipM-Hsp70N-ADP complex.

An overlay of the two structures is shown. For superposition of the structures, the lobe II parts (subdomains IIA and IIB) of the NBDs were aligned. DnaK-ATP is shown in red, Hsp70N-ADP in green and HipM in blue ribbons, respectively. HipM is enveloped by a molecular surface.

In the ATP bound state, DnaK is in a compact conformation and the 3-helix bundle domain of the Hsp70 SBD is associated with the flank of the NBD. ATP binding to Hsp70 furthermore induces a twist between lobes I and II of the Hsp70 NBD, causing a slight rotation of lobe I of DnaK-ATP around the vertical axis, resulting in a large (4 Å) relative shift of the top-left part of lobe I compared to ADP-bound Hsp70. This part contributes critically to the interface with Hip (indicated by an arrow). The ATP-induced shift would cause a severe clash with HipM. Therefore, Hip can no longer bind to both lobes of Hsp70 in the ATP-bound conformation simultaneously. Furthermore, the 3-helix bundle domain of the Hsp70 SBD would clash with the N-terminal region of HipM, which is indicated by a double arrow (Fig. 42). In the ADP-bound state, the 3-helix bundle domain of DnaK forms a lid over the peptide binding β -sandwich domain, which is loosely tethered *via* a flexible linker roughly opposite from the Hip interaction site. Thus it is unlikely to interfere with Hip binding.

The nucleotide-free state of Hsp70 (apo-state) is probably lowly populated in the nucleotide-rich environment of the cytosol. In principle, it should have protein substrate affinity similar to the ADP-bound state, and as such should be a target for Hsp70-substrate complex stabilization. The lower affinity of Hip to this state is probably caused by increased flexibility of the Hsp70 NBD, as suggested by molecular dynamic simulations (Zhuravleva and Gierasch, 2011). This would transiently disrupt the simultaneous contacts of Hip to subdomains IB and IIB in Hsp70.

5.4 Mechanism of Hip-mediated stabilization of Hsp70 in the ADP state

The crystal structure of the Hip:Hsp70 core complex revealed the molecular details of the functional interaction. Through a large number of polar and hydrophobic interactions, Hip stabilizes Hsp70N by forming a bracket over the nucleotide binding cleft, locking ADP in the nucleotide pocket. The binding interface involves the highly conserved edge of the TPR domain (residue 176-181 and 210-214) in Hip, which contacts subdomains IA, IB and IIB of Hsp70N.

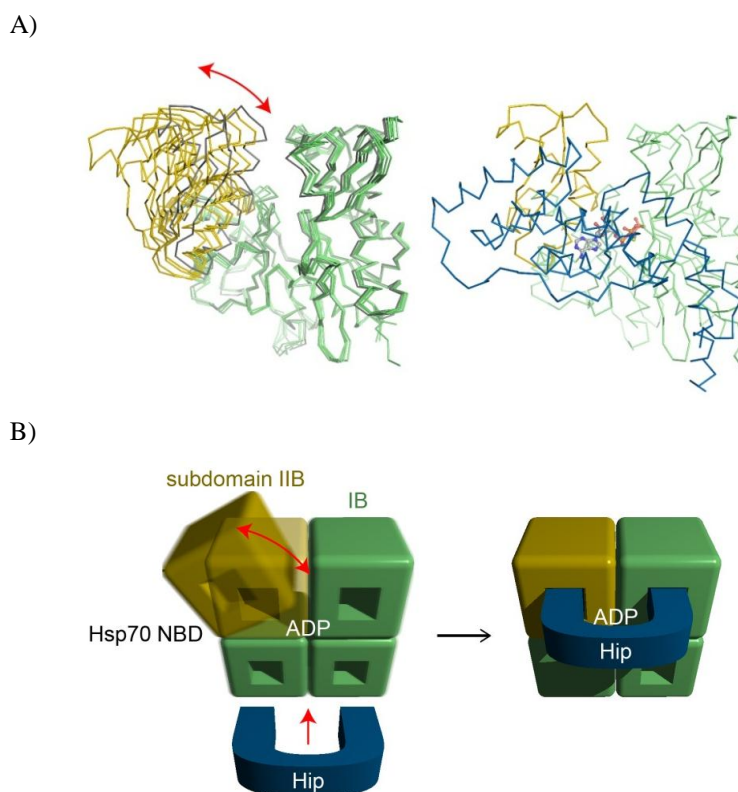


Figure 43: Interpretation of the Hip:Hsp70 structure.

- A) Left panel: Superposition of Hsp70 NBD structures. Subdomain IIB indicated in yellow is attached *via* a flexible hinge to the rest of the NBD (green). This becomes apparent by comparing the conformations of the NBD in crystal structures of NBD:NEF complexes and the free ADP-bound form (gray) (Arakawa et al., 2010; Polier et al., 2008; Schuermann et al., 2008; Sondermann et al., 2001; Wilbanks and McKay, 1995; Xu et al., 2008). Right panel: The structure of the Hip:Hsp70 core complex is shown. Peptide backbones are represented as α -carbon traces.
- B) Schematic representation of structural dynamics of the Hsp70 NBD and the mechanism for stabilization of the closed conformation by Hip. The outward rotation of subdomain IIB - indicated by a curved red arrow - breaks the respective contacts to ADP, thereby accelerating nucleotide dissociation (Liu et al., 2010). The contacts of the middle domain of Hip (dark blue) to subdomains IIB and IB/IA lock the domain in the closed conformation with ADP buried in the center. Absence of nucleotide greatly enhances the dynamics of the Hsp70 NBD, which would prevent stable Hip binding. In the ATP-bound conformation, the halves of the NBD are twisted with respect to each other, displacing the Hip-binding interface segments.

Figure 44 shows an overview of the structural dynamics of the NBD of Hsp70. It is interesting to note the motion of subdomain IIB to allow ADP release, which is otherwise firmly surrounded by Hsp70 and cannot easily escape. A simulation of the rotatory movement is shown in Figure 44A by comparing the NBD in crystal structures of NBD:NEF complexes and the ADP-bound form, which is closely similar to the structure of the NBD in the core complex with Hip. Interaction of Hip with both subdomain IIB and the opposing lobe I dampens this dynamic movement, thereby slowing dissociation of ADP.

5.5 Antagonism between Hip and NEFs

In principle, Hip and NEFs have opposing effects on the reaction cycle of Hsp70. Hip binding would rather slow or inhibit the cycle, while NEFs together with J-domain proteins would drive it forward. Comparison of the respective structures and our biochemical data show that Hip and NEF binding to Hsp70 are mutually exclusive.

Notably, Hip has much lower binding affinity to Hsp70 *in vitro* ($K_D \sim 10 \mu\text{M}$) than the NEFs, which have remarkably similar affinities ($K_D \sim 0.1 \mu\text{M}$) (Raviol et al., 2006; Shomura et al., 2005; Sondermann et al., 2001). The interaction between Hip and substrate-free Hsp70 appears to be largely, if not exclusively, mediated by the TPR domain of Hip (HipM) and the Hsp70 NBD, but with an affinity probably too low to be biologically relevant. In the cytosol, this may be partially compensated by a favorable concentration ratio between Hip and NEF (NEFs inhibit Hsp70 at 1:1 ratio). While the competition under physiological conditions remains to be experimentally investigated, our preliminary analysis of Hip-mediated attenuation of MABA-ADP dissociation from Hsp70N in the presence of 10 mM phosphate suggests that increased concentrations of this metabolite might be in favor of Hip binding (Fig. 20C). In presence of phosphate, ADP appears to bind more tightly to Hsp70 (Arakawa et al., 2011), suggesting that the dynamics of the NBD is decreased. The binding characteristics of Hip as judged from the concentration dependence in presence of 10 mM phosphate, appear unaltered (Fig. 20C). NEFs however appear to merely capture and stabilize the open conformation of the NBD, which will be less populated under such circumstances. Stabilization of ADP complexes of eukaryotic Hsp70 by phosphate might also explain why NEFs are required in the eukaryotic cytosol, alt-

hough the spontaneous off-rate is sufficient to drive the folding reaction *in vitro*. This is in contrast to the bacterial homolog DnaK, which also depends on the NEF GrpE *in vitro*. The finding that the hyper-activating mutation I148A of Hip has not been selected during evolution indicates that Hip can effectively compete with NEFs under physiological conditions.

Under non-stressed conditions, Hip likely needs direct or indirect interactions with Hsp70-bound substrate to effectively compete with NEFs. We were unable to demonstrate this effect *in vitro* with denatured firefly luciferase or carboxymethylated bovine α -lactalbumin using our MABA-ADP release assay (data not shown). As judged from GR maturation, the putative substrate binding capability of Hip is as important as its capacity to interact with Hsp70 *in vivo*. However, avidity could greatly enhance the binding of Hip dimers to both substrates and Hsp70, as long as they form complexes with multiple binding sites.

Our findings therefore suggest that in order to effectively compete with NEF binding to Hsp70, Hip dimers would have to interact simultaneously with two Hsp70 molecules bound to different regions of the same substrate protein or to Hsp70s associated with small protein aggregates. The binding of multiple Hsp70 molecules may reflect the requirement of a specific substrate protein for prolonged chaperone stabilization. Thus, conformational properties of the client protein would determine the extent to which Hip decelerates Hsp70 cycling. Future experiments will have to test this element of the model, and the 140-residue protein α -synuclein might be a promising substrate candidate, which was shown to undergo nucleotide-dependent interactions with Hsp70 (Roodveldt et al., 2009).

5.6 Possible role of Hip in protein quality control

Hip is so far the only known Hsp70 co-chaperone that stabilizes Hsp70-client protein complexes by retarding the dissociation of ADP from Hsp70. Previous model suggested the regulation of Hip as a tetramer to stabilize Hsp70 in the ADP-bound state (Ziegelhoffer et al., 1996).

Hsp70 is both involved in the folding of newly made and stress-denatured proteins as well as in the timely degradation of terminally misfolded proteins. Figure 44 shows an overview for the potential role of Hip in the cellular protein homeostasis machinery.

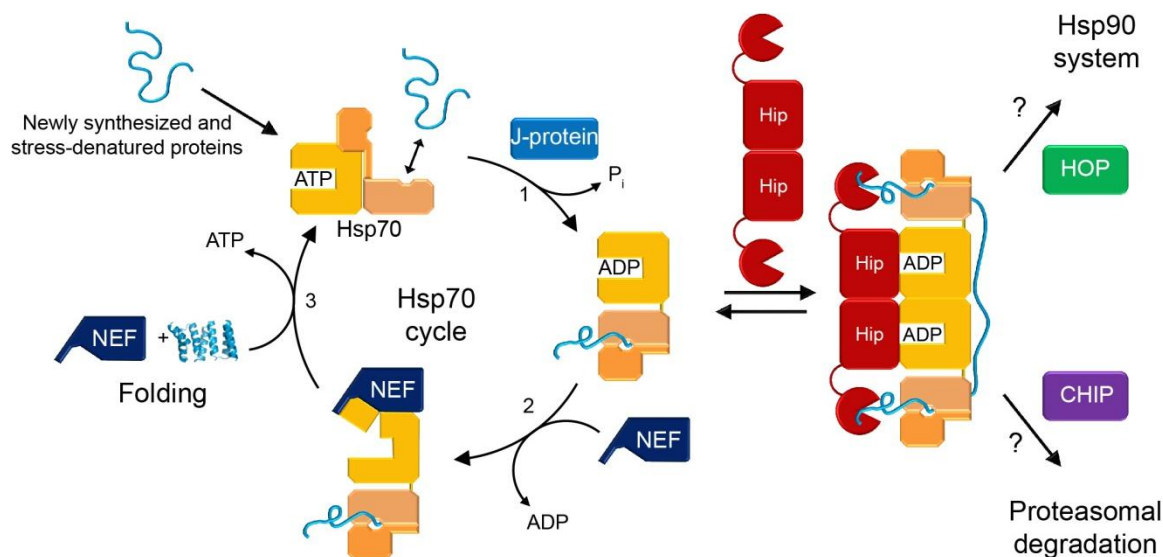


Figure 44: Model for the role of Hip in protein quality control.

A schematic model for the cooperation of Hsp70 and its cochaperones in protein folding is depicted. Hsp70 and Hip are indicated in yellow and red color, respectively. J-domain proteins and NEFs (blue color) drive the Hsp70 cycle of substrate binding and release. Hip attenuates Hsp70 cycling by stabilizing the ADP state, thereby increasing the substrate “holdase” activity of Hsp70 and diverting certain Hsp70-ADP-substrate complexes towards Hsp90 or proteasomal degradation.

J-domain proteins and NEFs drive the Hsp70 cycle of substrate binding and release. For both factors multiple isoforms co-exist in the cytosol, presumably adapting Hsp70 cycling rates to specific substrates.

Hip attenuates Hsp70 cycling by stabilizing its ADP state, thereby increasing the substrate “holdase” activity of Hsp70. This might occur by specific recognition of the bound substrate, enabling slow cycling and limiting unnecessary ATP expenditure on recalcitrant substrates. In addition, Hip preferentially targets and stabilizes Hsp70-ADP-substrate complexes containing multiple Hsp70 molecules. Binding of multiple Hsp70 molecules may reflect the requirement of a specific client protein for prolonged chaperone stabilization. Notably, the conformational properties of the client protein would determine the extent to which Hip decelerates Hsp70 cycling. Prolonged residence on Hsp70 mediated by Hip may prevent aggregation or facilitate substrate transfer to downstream Hsp90 or to the degradation machinery. The decision to fold

or degrade the client might depend on the competition between downstream co-chaperones, Hop/Sti1 and the ubiquitin ligase Chip. Both of them target the C-terminal EEVD motif of Hsp70 (Scheufler et al., 2000; Zhang et al., 2005). Previous studies showed that Hip cooperated with both factors. Hop and Hip synergize in the maturation of GR (Nelson et al., 2004) and Hip and Chip support increased proteasomal degradation of iNOS (Wang et al., 2013).

We have demonstrated that Hip binds to the NBD domain of Hsp70. It was known that Hop binds to the SBD of Hsp70. Thus a simultaneous interaction of Hop and Hip with Hsp70 seems possible. Consistently, a complex containing Hsp70, Hsp90, Hip and Hop has been identified as an intermediate during progesterone receptor activation (Smith et al., 1995). Besides Hop, Chip also interacts with Hsp70 and plays an essential role in balancing substrate protein folding and degradation. It was reported that Chip associated with Hsp70 and blocked the ATPase cycle of the chaperone (Ballinger et al., 1999). Apparently Hsp70 selects substrates for Chip-mediated ubiquitylation. Thereby Chip might associate with Hsp70 to promote the ubiquitylation of diverse substrates.

The balance between the diverse pathways, active protein folding by Hsp70, hand-over to Hsp90 or to the protein degradation machinery, apart from the cellular concentrations of the components and phosphate levels – might also be regulated by the post-translational modifications of Hip. Phosphorylation of Hip at serine residues 74, 75 and 78 adjacent to its Hsp70-binding domain and within the DP domain likely modulates the activity of Hip, providing additional regulation of substrate flux (Barker and Benovic, 2011; Olsen et al., 2006).

Both Hop and Chip could also be involved in the interaction of Hip with Hsp70 and substrate, but the underlying mechanism remains elusive. One possibility to study the simultaneous interactions of Hip-Hsp70 with Hop or Chip in detail would be to determine the crystal structures of complexes of the full-length proteins.

6 Implications and outlook

Determination of three-dimensional structures of Hip and the Hip-Hsp70 core complex was the main achievement in this study. We presented the crystal structures of the N-terminal dimerization domain of Hip, the TPR binding domain for Hsp70 and the core complex with the ATPase domain of Hsp70. Thus, this work provides novel insights into the function of Hip at the molecular level.

The biochemical and structural data presented in this work lead to a detailed model for the function of metazoan Hsp70 and provide a preliminary model of the regulatory role of Hip in protein quality control. Hip may function as a central scaffolding protein that coordinates the interplay between multiple chaperones such as Hsp40, Hsp70 and Hsp90 and the proteasomal degradation machinery. Since Hip associates with Hsp70-substrate complexes, an important and interesting future goal will be to identify the client proteins of Hip. If the substrates include signaling molecules known to interact with Hsp90, this would provide clear evidence that Hip contributes to the conformational regulation of these clients and cooperates with Hsp90 in their biogenesis.

Increasing the level of Hip has been shown to reduce pathologic protein aggregation in cellular models of Parkinson's and Huntington's disease (Howarth et al., 2009; Roodveldt et al., 2009). Enhancing Hip activity or mimicking its effect on Hsp70 pharmacologically (Wang et al., 2013) may prove useful in facilitating the proteolytic clearance of toxic, aggregation-prone proteins. Similarly, boosting Hip function might be advantageous in cancer therapy (Zhou et al., 2012) by accelerating the clearance of metastable, oncogenic mutant proteins.

Given that mammalian cells likely possess multiple isoforms of J-proteins and different types of NEFs in the same compartment, controlling the ATPase activity of Hsp70 appears to be one central determinant for regulation of chaperone activity. Hip competes with NEFs for the binding to the ADP state of Hsp70, which might have far-reaching effects. It will be interesting to see whether retarding ADP-release from Hsp70 is also utilized more extensively in the regulation of protein quality control. This might create new opportunities for drug design and therapeutic interventions.

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8 Appendix

8.1 List of primers

Oligonucleotides utilized in this study are listed here. Oligonucleotides were purchased from Metabion (Martinsried, Germany).

| Construct | Primer | Sequence 5' → 3' | Endonuclease |
|--------------|----------|--|-------------------|
| Hip(1-44) | LZ_HIP1f | CATGCCATGGACCCCCGCAAAGTGAGCGAGC | <i>NcoI</i> |
| | LZ_HIP2r | CCCAAGCTTAAGGTGGTACTTTACCCCCATGCTCTCC | <i>HindIII</i> |
| Hip(1-267) | AB99f | phos-GCCCGCGGTGGAATGGACCCCCGCAAAGTGAGCGAGC | <i>EheI-SacII</i> |
| | AB101r | CCCAAGCTTATTCTTCCCTTTGGGCTTTTTTCATGCTCTTCTCG | <i>HindIII</i> |
| Hip(1-368) | AB99f | phos-GCCCGCGGTGGAATGGACCCCCGCAAAGTGAGCGAGC | <i>EheI-SacII</i> |
| | AB102r | CCCAAGCTTATGAGTGACCTCCAAACTTGGCTGACAA TTTACTG | <i>HindIII</i> |
| Hip(45-267) | LZ_HIP5f | CCCCATGGCTACTCATAAAGCGAAGTCAGAAGAA A AC | <i>NcoI</i> |
| | LZ02r | CGGGGTACCTTATTCTTCCCTTTGGGCTTTTTTCATGCTCTTCTCG | <i>KpnI</i> |
| Hip(45-368) | LZ_HIP5f | CCCCATGGCTACTCATAAAGCGAAGTCAGAAGAAA AC | <i>NcoI</i> |
| | AB102r | CCCAAGCTTATGAGTGACCTCCAAACTTGGCTGACAA TTTACTG | <i>HindIII</i> |
| Hip(78-234) | AB100f | phos-GCCCGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <i>EheI-SacII</i> |
| | LZ_HIP4r | CCCAAGCTTACCGATGTTTACGCAATTTTTTGAGCCCG AGG | <i>HindIII</i> |
| Hip(78-247) | AB100f | phos-GCCCGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <i>EheI-SacII</i> |
| | LZ20r | CCCAAGCTTATTTTATCTCTCGCTTTCACGTTTTTCGCT CATA | <i>HindIII</i> |
| Hip(78-267) | AB100f | phos-GCCCGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <i>EheI-SacII</i> |
| | AB101r | CCCAAGCTTATTCTTCCCTTTGGGCTTTTTTCATGCTCTTCTCG | <i>HindIII</i> |
| Hip(78-368) | AB100f | phos-GCCCGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <i>EheI-SacII</i> |
| | AB102r | CCCAAGCTTATGAGTGACCTCCAAACTTGGCTGACAA TTTACTG | <i>HindIII</i> |
| Hip(107-267) | LZ23f | GCCGAGGCGATGATGGATGAAGCAAATGAAAAG | <i>EheI</i> |
| | AB101r | CCCAAGCTTATTCTTCCCTTTGGGCTTTTTTCATGCTCTTCTCG | <i>HindIII</i> |
| Hip(312-368) | LZ_HIP3f | CATGCCATGGGACTCAACGAAATCCTCAGTGACCCAG AG | <i>NcoI</i> |
| | AB102r | CCCAAGCTTATGAGTGACCTCCAAACTTGGCTGACAA TTTACTG | <i>HindIII</i> |

Table 16: Primer utilized for generation of Hip constructs in this study.

The names of constructs, primers with respective nucleotide sequences and the corresponding restriction endonuclease sites are listed in the respective columns. The underlined residues indicate restriction endonuclease sites introduced for cloning. The 5'-phosphorylation of forward primers is indicated by phos.

| Hip Mutant | Primer | Sequence 5' → 3' |
|----------------|--------------|---|
| K117A | LZ89f_K117A | ATGAAGCAAATGAAAAG CGGGGG GCTGCCATCGACGCTC |
| | LZ90r_K117A | GAGCGTCGATGGCAGCCCC GCCTTTT CATTTGCTTCAT |
| G118V | LZ34f_HIPFM1 | GAAGCAAATGAAAAGAAG GTGG GCTGCCATCGACGCTCTA |
| | LZ35r_HIPFM1 | TAGAGCGTCGATGGCAGCCAC CTTCTTTT CATTTGCTTC |
| I148A | LZ93f_I148A | CTAAACCCTCGCTTGGCC GCCT GTATGCCAAGAGA |
| | LZ94r_I148A | TCTCTTGGCATA CAGGGCGG CCAAGCGAGGGTTTAG |
| N177K | LZ70f_N177K | GACAGAGCTATTGAAATA AAAGC CTGATTCAGCTCAGCCA |
| | LZ71r_N177K | TGGCTGAGCTGAATCAG GCTTT TATTTCAATAGCTCTGTC |
| S180E | LZ96f2_S180E | ATTGAAATA AAACCCT GAT GAGG CTCAGCCATACAAATGGAGA |
| | LZ97r2_S180E | TCTCCATTTGTATGGCTGAG CCTCAT CAGGGTTTATTTCAAT |
| K185E | LZ72f_K185E | GATTCAGCTCAGCCATAC GAAT GGAGAGGGAAAGCGCAC |
| | LZ73r_K185E | GTGCGCTTTCCCTCTCCAT TC GTATGGCTGAGCTGAATC |
| L210S | LZ87f_L210S | TTGCCCTGGCCTGTAAAT TCCG ACTATGATGAGGACGC |
| | LZ88r_L210S | GCGTCCTCATCATAGT CGGATTT ACAGGCCAGGGCAA |
| N177A/D179A | LZ85f_ND | AGAGCTATTGAAATAG CCCCTGC CTCAGCTCAGCCATAC |
| | LZ86r_ND | GTATGGCTGAGCTGAG GCAGGG GCTATTTCAATAGCTCT |
| H196S/E199A | LZ91f_HE | ACAGACTCCTGGG TTCTT GGGAAG CCG CAGCTCGCGAT |
| | LZ92r_HE | ATCGCGAGCTG CGGCTT CCAAGA ACCC AGGAGTCTGT |
| D211K_Y212A | LZ74f_2M | CTGGCCTGTAAATTGA AGGCC GATGAGGACGCCAGTGCA |
| | LZ75r_2M | TGCACTGGCGT CCTCATCGG CCTTCAATTTACAGGCCAG |
| R235A/K236D | LZ83f_RK | AAATTGCTGAACATCG GGCCG ACTATGAGCGAAAACGTGAAGAGC |
| | LZ84r_RK | GCTCTTACGTTTT CGCTCATAGT CGGCCCGATGTT CAGCAATTT |
| S74E/S75E/S78E | LZ95f_3E | GCCGGAG AGGAGG AGGAGGAGGATCTAGAAATTGACAATGAAGGTGTAATTGAA |
| | LZ20r | CC CAAGCTT ATTTTTATCTCTCGCTCTT CAGTTTT CGCT CATA |
| S74E/S75E/S78E | LZ100f_3E | AAGACAGAGGAGCCAG AGGAGG AGGAGGAGGATCTAGAAATTT |
| | LZ101r_3E | AATTTCTAGAT CCTCCTCCTCCT CTGGCTCCTCTGTCTT |
| S74N/S75N/S78N | LZ98f_3N | AAGACAGAGGAGCCAA ACAAC GAGGAGA AC GATCTAGAAATTGAC |
| | LZ99r_3N | GTCAATTTCTAGAT CGTTCTCCTCGTTG TTTGGCTCCTCTGTCTT |

Table 17: Primers utilized for site-directed mutagenesis on Hip.

Point mutations were introduced *via* a PCR based mutagenesis strategy into Hip. The sites of point mutations are listed on the left column. The respective names of the primer and the corresponding oligonucleotide sequences are listed. The modified sites are highlighted by bold characters.

| Construct | Primer | Sequence 5' → 3' | Endonuclease |
|------------------------|--------------|---|-------------------|
| Hip(78-234)-3aa-Hsp70N | AB100f | phos-G <u>CCC</u> CGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <u>EheI-SacII</u> |
| | LZ43r | <u>CCCGGATCC</u> ACCCCGATGTTTCAGCAATTTTT TGAGCCCCGAGG | <u>BamHI</u> |
| | LZ51r | <u>CCCGAATTC</u> GGATCCACCCCGATGTTTCAGCA ATTT | <u>EcoRI</u> |
| Hip(78-234)-6aa-Hsp70N | AB100f | phos-G <u>CCC</u> CGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <u>EheI-SacII</u> |
| | LZ44r | <u>CCCGGATCC</u> ACCGGAGCCACCCCGATGTTCA GCAATTTTTTGAGCCCC | <u>BamHI</u> |
| | LZ52r | <u>CCGAATTC</u> GGATCCACCGGAGCCACCCCG | <u>EcoRI</u> |
| Hip(78-234)-9aa-Hsp70N | AB100f | phos-G <u>CCC</u> CGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <u>EheI-SacII</u> |
| | LZ45r | GGAGCCACCTCCAGCACCCCGATGTTTCAGCA ATTTTTTGAGCCCC | — |
| | LZ46r | <u>CCCGGATCC</u> ACCGGAGCCACCTCCAGCACCC CG | <u>BamHI</u> |
| | LZ53r | <u>CCCGAATTC</u> GGATCCACCGGAGCCACCTCC | <u>EcoRI</u> |
| HipM-3aa-Hsp70N | AB100f | phos-G <u>CCC</u> CGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <u>EheI-SacII</u> |
| | LZ76r_F3aa | <u>CCCGGATCC</u> ACCTTTTATCTCTCGCTCTTCAC- GTTTTCGCTCA | <u>BamHI</u> |
| HipM-6aa-Hsp70N | AB100f | phos-G <u>CCC</u> CGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <u>EheI-SacII</u> |
| | LZ77r_F6aa | <u>CCCGGATCC</u> ACCGGAGCCACCTTTTATCTCTC GCTCTTCACGTTTTTCGCTCA | <u>BamHI</u> |
| HipM-9aa-Hsp70N | AB100f | phos-G <u>CCC</u> CGCGGTGGAAGCGATCTAGAAAT TGACAATGAAGGTGTAATTGAAGC | <u>EheI-SacII</u> |
| | LZ78r_F9aa | GGAGCCACCTCCAGCACCTTTTATCTCTCGCT CTTCACGTTTTTCGCTCA | — |
| | LZ79r_F9aa | <u>CCCGGATCC</u> ACCGGAGCCACCTCCAGCACCTT TTAT | <u>BamHI</u> |
| Hsp70N-HipM | LZ80f_Hsp70N | <u>CCCGAATTC</u> ATGGCCAAAGCCGCGGCGATCG | <u>EcoRI</u> |
| | LZ81r_Hsp70N | <u>CCCTCGAG</u> CCCCATCAGGATGGCCGCCTG | <u>XhoI</u> |
| | LZ82f_HIPF | GGG <u>CTCGAG</u> AGCGATCTAGAAATTGACAATG AAGGTGTAATTGAA | <u>XhoI</u> |
| | LZ20r | <u>CCCAAGCTT</u> ATTTTATCTCTCGCTCTTCACGTT TTCGCTCATA | <u>HindIII</u> |

Table 18: Primers utilized for generation of Hip-Hsp70 fusion protein in this study.

The names of fusion proteins, primers with respective nucleotide sequences and the corresponding restriction endonuclease sites are listed in columns. The underlined residues indicate the restriction sites introduced for cloning. The phosphorylation of forward primers is indicated by phos.

| Construct | Primer | Sequence 5' → 3' | Endonuclease |
|--|----------------------------|--|--------------------------------------|
| Hip(1-267) | LZ102f | phos-GCC <u>ACTAGT</u> ATGGATCCCCGCAAAGTGAG CGAG | <i>EheI-SpeI</i> -ATG start codon |
| | LZ110r | CCGAATTCTTATTCTTCCCTTTGGGCTTTTTTCAT GCTCTTC TCG | <i>EcoRI</i> |
| Hip(1-368) | LZ102f | phos-GCC <u>ACTAGT</u> ATGGATCCCCGCAAAGTGAGC GAG | <i>EheI-SpeI</i> -ATG start codon |
| | LZ109r | CCGAATTCTTATGAGTGACCTCCAAACTTGGCTG ACAATTTACTG | <i>EcoRI</i> |
| Hip(45-368) | LZ103f | phos-GCC <u>ACTAGT</u> ATGGCTACTCATAAAGCGAAG TCAGAAGAAAACAC | <i>EheI-SpeI</i> -ATG start codon |
| | LZ109r | CCGAATTCTTATGAGTGACCTCCAAACTTGGCTG ACAATTTACTG | <i>EcoRI</i> |
| pProEx-HtB (<i>SpeI</i> site mutant) | LZ111f_ <i>SpeI</i> mut | CTACGTCGACGAGCTCT ATAGT CGCGGCCGCTT TCGAA | — |
| | LZ112r_ <i>SpeI</i> mut | TTCGAAAGCGGCCGCG ACTATAG AGCTCGTCGA CGTAG | — |

Table 19: Primers utilized for generation of Hip constructs for the *in vivo* studies in yeast.

The names of constructs, primers with respective nucleotides and the corresponding restriction endonucleases are listed in the respective column. The underlined residues indicate restriction endonuclease sites introduced for cloning purposes. The phosphorylation of forward primers is indicated by phos. Since the yeast vectors p423GPD or p423ADP contains no start codon, ATG start codons were introduced *via* PCR amplification. ATG start codon and mutated *SpeI* site in pProEx-HtB vector are also highlighted in bold. The Hip mutants for yeast study were generated using the respective primers in Table 17.

| Hsp70 Mutant | Primer | Sequence 5' → 3' |
|--------------|--------------|---|
| Hsp70_D199A | LZ120f_D199A | AACGTGCTCATCTTT GCC CTGGGCGGGGGCACCT |
| | LZ121r_D199A | AGGTGCCCCCGCCCAG GGC AAAGATGAGCACGTT |
| Hsp70_T204A | T204A_f | TTT GAC CTGGGCGGGGG CGC GTTCGACGTGTCCAT |
| | T204A_r | ATGGACACGT CGA AC CGC CCCCCGCCAGGTCAA |

Table 20: Primers utilized for site-directed mutagenesis on full length human Hsp70.

Point mutants of Hsp70 variants were investigated *via* a PCR based mutagenesis strategy. The sites of point mutants are listed on the left column. The names of primer and the corresponding nucleotides are listed, respectively. The mutated sites are highlighted in bold.

8.2 Hip alignment

α1 α2 α3 α4 α5 α6 α7 α8 α9 α10

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R_norvegicus 1 .....MDPRKVSELRAFVKMCQRDPSVLHTEEMRFLREWVES..MGGKVPPAT.....HKAKSE.....ENTKEEKRD
T_adhaerens 1 .....MATSIQHLRELEQFIDHCKNNPAMLRLPELKFPADFLVS..LGGKIPEAAE.....PEEKVE.....ESINDESES
C_elegans 1 .....MDHVALLKQFVGMCQANPAVLHAPFEGFFKDYLVS..LGATLPKPA.....DKPEGK.....CPMSEEPKKE
S_mansoni 1 .....MNPEKVALLKQFVELLREKPEVLNDPELSFFKDWLKS..LGANVPSVQD.....NRP.....TENSFSD
D_melanogaster 1 ..MAFTMQTDGLKKLKYFIDFALENPTFLNMPLQFVKDFVEK..FGGTVPGQF.....NGGSAGGKCPFGGVAGAKANE
E_histolytica 1 ..MESFGVTSQIQALKVFINLLQHPEFLNDDRLDFLRNYIIS..LGGKIPEKK.....EPKSE
T_brucei 1 ..MHTLSSDFEALNRVLDFLRHPSEIHRDEFNGLRAFLTS..LGATLPSTYEPK.....TTAGPEANE
L_braziliensis 1 ..MPLYSDADVSGLRRILEALSRQPGDIHLSELAELKAWTAA..AGASFPAAAVPT.....VPVEAATLE
O_lucimarinus 1 MTRERAPTATLEALLRHLVRAAKERDSAILDPALEFFRRLWLEEDLGATIPARTTTT.....TGTGTDALE
T_gondii 1 ..MAALSPQKVAELKAFIGMCDPSILHRPELSFFKELQS..LKAAIPAER.....QAGAGSPDRPTSPVPEASSD
P_falciiparum 1 .....MIDEKKVEDLKQFVTLCQENPSILLKPELGFPKFIES..FGGKVSKDKEFEKSASDESTEEKSDEEEVEEDEND
A_thaliana 1 .....MDSTKLSELKVFIDQCKSDPSLLTPSLSFPRYLES..LGAKIPTGV.....HEEDKDTKRSFFVESDDD
consensus>50 .....v..Lk.fvd..edp..l..pel.ff.df..s.lga.ip.....HEEDKDKTRSFVVESESDD

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contacts to Hsp70 post-transl. modif. mutation sites

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R_norvegicus 62 KTTEDN.....IKTEEPSSEESDLEIDNEGVIEADTDPAQEMGDENAEITEAMMDEANEKKGAAIDALNDGELKAIDLF
T_adhaerens 66 QVGSANYI..ILCPEKESEESDLDIEAEGVTEGDDPAQPMGDGSKEVTEEMLEEASEKRQNANIALADGNIDEGIKLF
C_elegans 62 TPAAEATP..EPEIPKEEIPPKIDNEVEPEEEAVALMGDSAKEATEDEIEKASEERGQEAFSNGDFTALTF
S_mansoni 58 DSGADETS..ES.....EIEF.....DDEVLPKEPVPDLAMGDDSIEVTDEMREKAEEKKCEAMAKMSDGDLTGAVDLF
D_melanogaster 73 PANAPEDS..EEEKSLSDPESDVELMDEGVIEADSDPAQPMGNYSKKATEEEVEQASTELRAQASAYGGQKFDEAIALY
E_histolytica 59 EKMEEK..KEEETAKEKPEEDIKIDDPVIPGDTIEPETIN.MDIEVTEEMVEQASTEKSEAMEAFNNGEVDKAINTI
T_brucei 63 EDTASEPD.....EELWKLEDVADGGIPAGSGDPSPEQEEKAMELKAAADCAADGRLDEAVDLF
L_braziliensis 62 QDDKSDS.....EEHWTLVDEDPEEIPAKYSEPSDADVDAAVSAKEAVELHDAGKLEAITKM
O_lucimarinus 67 IEDDEAMA.....AESDDLSATAMGAETAPETLGEAEEAKSAEKRLASEAFAREAWEEAIERY
T_gondii 71 DSSLES.....EVEEFDESSLKDSE..VIPPETSPLPLAPEGEKELTDDELDKLGKLEASAACEAGNSERALEKY
P_falciiparum 75 KDDVEEEEE..EDDVQEDDEEEDDERCKDFVEEPTICPPLAPIVEDLSDEVLEEISNLKIEAELVQNKFEALERY
A_thaliana 67 MDEVEVKPKVEEEEEDEIVSDVELGGVEPEDNDPPKMGDSSVEVTDNREAAQEAKKAMELSGNFDEAIELH
consensus>50 .....ee..e.....e.vie.d..pq..d..e.teee.e.a.e.k.ea.ea.dgn.aaeia.e.y

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contacts to Hsp70 post-transl. modif. mutation sites

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R_norvegicus 137 TDAIKLNPRLAILYAKRASVFVKLQKPNAIAIRDCDRAIEINPDSAQEYKWRGKAHRLLGHWEEARDLALACKDDYDEDA
T_adhaerens 143 AEAIMLNPQSAPLFAKRRAGYIKAKKPNAIAIRDCDKAIQLNPDSAQAYKWRGIAYRMLGKWDKSASDLRLSVKDFSEDA
C_elegans 139 TAAIEANPGSAMLHAKRANVLLKRVPAAIADCDKAISINPDSAQGYKWRGRANRLLGKWVEAKTDLATACKDDYDEA
S_mansoni 125 TEAIKLNPQSSLFHARRASCFVRMKPSHAIAADCDKAISINPDSAQEYKWRGFANKIGNWEAAYQDLQTSLKDDYTDDA
D_melanogaster 150 TKAIELSPGNALFHARRGOAFLLKKNACIRDCVALELSDLAAGYKFRGRARLLGDFELAAHDLROACKDDEET
E_histolytica 137 TEAIKLNPRVANFFACRAQYYNKAKKENAIRDCTAIKLNPDNAKYWRGIAYRMIQYQKSVVDLRLGNKLDDDET
T_brucei 123 AQALRLVPGKAMYWSQRASYLLECKRPAALRDANRALSLNPENVRALRVRGTVNRHLGWEDALKDLSEAQTVDYDEKA
L_braziliensis 122 SEALAHNFSNAMYWGLRALYHLFNKPAALDANKTLELSQNVRALRVRGTVNRHLGHWESLKDLSAQATDYDEKT
O_lucimarinus 126 TEALMIAPS..ALYAKRAECFIKLRKPLSAIRDGTAALKLNPDSAKALKVRGAAHRLGHWNEANADLSAGLSODFDETY
T_gondii 142 TEALIGNPTALLYTRADVLLKRPVACIRDCEALNLNPDSARAYKIRGKANRLLGKWEAKSDLMGQKDDYDEL
P_falciiparum 154 NKIAFGKPSAMIYTKRASVLSLKRPKACIRDCEALNLNIDSANAYKVRAKAYRHLGKWECAHADIEQGKTDYDEDL
A_thaliana 147 TRAITLNPTSAMYGNRASVLLKKPNAIAIRDANALEINPDSAKYKSGMARALGWEAEAKDLHLASTDYDEL
consensus>50 tea.i.lnp..am.ya.Ra..f.klkkp.aairDcd.al.lnp#sa.ayk.Rg.a.rllG.weea..Dl....k.D#d.d.

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contacts to Hsp70 post-transl. modif. mutation sites

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R_norvegicus 217 SAMLREVQPRAQKTAEHRRKYERKEEEIKER..IERVKK.....AREHEKAQREE..ARRO.....SGSQFG
T_adhaerens 223 QEVLNYVQPRAQRIQEHQRKIERRKAEKEIKER..RERVRL.....AQEAKEKARKEEERARKEESGTPGGFPGAGAG
C_elegans 219 NEWLKEVEPNAHKIQEYNRAVERQKADIELAER..RERVRR.....AQEANKKAAEEAKRQAEQOG.....D
S_mansoni 205 NEAIKEIEPKHKRIFEHNMKYERKRQEKLERER...RERIRK.....AQEE.....RERARQRTEK.....S
D_melanogaster 230 DEWLKEVTPNAKIEQHRLKQERRQAERKIKER...QRDQRR.....ARKEQ.....EKHNASSGSS.....S
E_histolytica 217 YELLQVVEKFVAIEKQNEKRHNQQEEKPQEEQHOCHCHG.....GEKPGCDGHGCHRQQGCCG.....GQGGEK
T_brucei 203 DALLRLVQEKANARRO...SRRRKEEERAKRQEALRRQRE.....MEMOREEEERRQNQROAPPSGGFPGGFPGMPG
L_braziliensis 202 NETLKFVQFRAMORHKRELARQHAEEAAVARQEELRRQQ.....REAEEEAATAEAQASGMPPGMPPGMPG
O_lucimarinus 205 GEMHKKVLSVVHELHVR...EGKARAAEAKEREELEKRR.....AAEAAARKEAAAKDAGGPFGQPG.....AGFPG
T_gondii 222 WDMQKLVDEKFKIEEHERKIVRKEAEEKKRREKEARKR.....RAAAQRAYEEQKR.....DAAGS.....GGFPG
P_falciiparum 234 WEMQKLIEEKYKIEEKRRFKNREEEKKRKERKLKRLARKAAEKAYKANNKRENDSDSSSYSEPDFSGDFPG
A_thaliana 227 SAVLKKVEPNAHKLEEHRRKYDRLREREDEKAERDLR.....RAEAQAYDKAKEE.....QSSSRSPSGGFPGMPG
consensus>50 .e.lk.e.a.i.e.....er.ee.e.kere.er..r.....aq.a.e.....g...g

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contacts to Hsp70 post-transl. modif. mutation sites

```

R_norvegicus 279 SFP...GGFP.....GGMPGN...FP...GGMPGMG...GAMPGM.....GAMPG...GA
T_adhaerens 296 GEPGAGGPPGAGGFPGAGAGGPPGAGGFPGAGAGGFPGA.....GMPGA.....GA
C_elegans 279 DFDEPEGGFP.....GGMPGGFPGA.....GMPGGFPGA.....GMPGPGPAPGGGAPG...
S_mansoni 259 DFD.....RRKEEERER.....GGMPGGFPGA.....IPDS.....GNIPG...
D_melanogaster 286 EFS.....RRKEEERER.....GGMPGGFPGA.....IPDS.....GNIPG...
E_histolytica 284 GMPNMGSMPN.....GMPGGFPGA.....GMPGGFPGA.....GNIPG...
T_brucei 275 GFP.....GMPGGFPGA.....GMPGGFPGA.....GMPGPGPAPGGGAPG...
L_braziliensis 277 GFP.....GMPGGFPGA.....GMPGGFPGA.....GMPGPGPAPGGGAPG...
O_lucimarinus 271 GAG.....DLPPEV.....GMPGGFPGA.....GMPGGFPGA.....GMPGPGPAPGGGAPG...
T_gondii 286 GFP...EGFPG..FGG...AGFPPGAGG...GYSGGMPGTFPSSCGGS...GMTG.TPGCCGCCGMGGGCG...GM
P_falciiparum 314 GMP...GMPGGMPGM...PGGFPGGFP...GMPGGMPGGFPGGMPGGMPGGMP...GMPGGMPGGFPGGMPGGMP...GM
A_thaliana 300 GEP...GMPGGFPGGMPGGFPGGMG...GMGMPGGFPGGMPGGMPGGMPGG...GMPGPGMPGGMPGGMPGGM
consensus>50 gfp...p...p...gmpg

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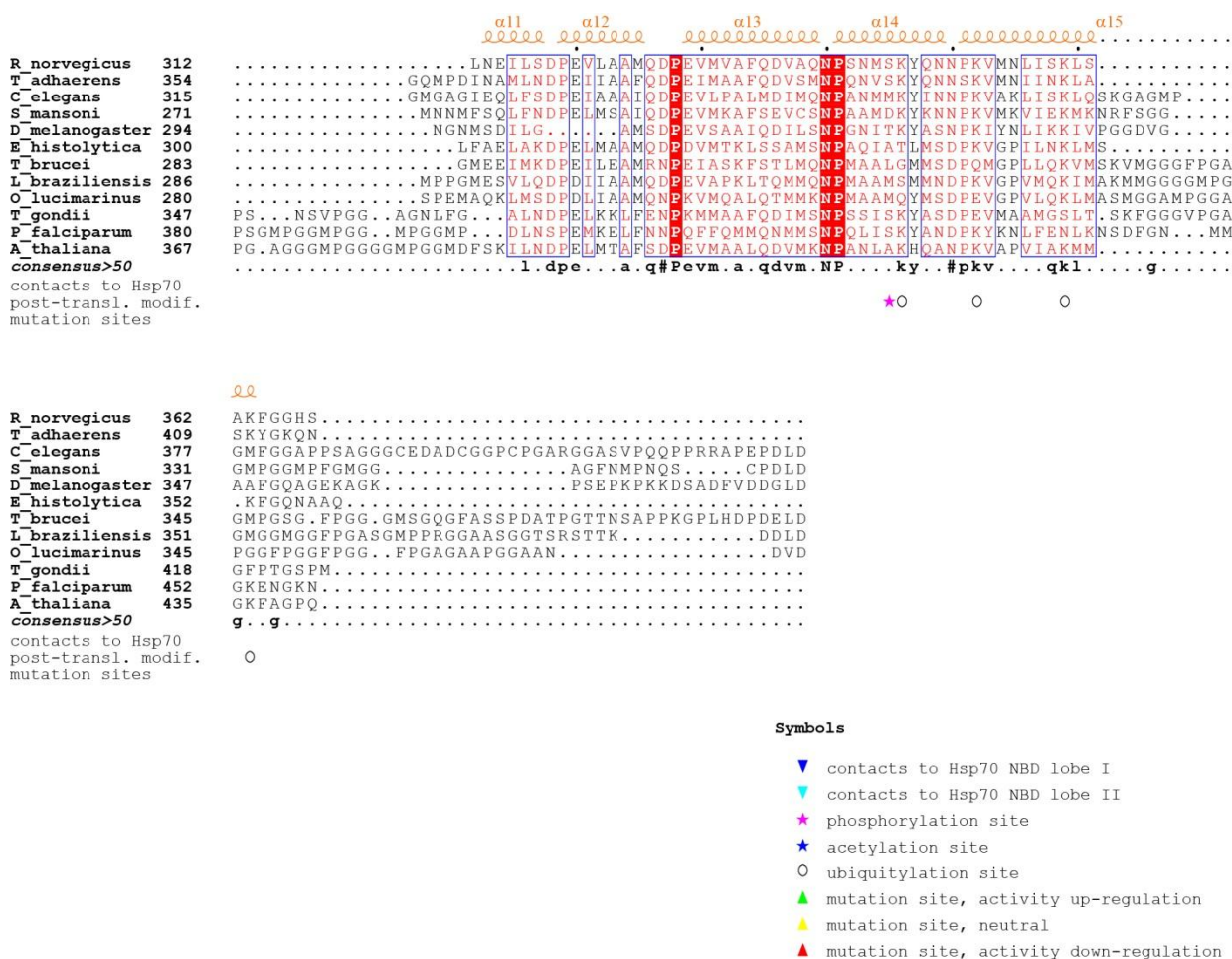


Figure 45: Alignment of Hip sequences.

Amino acid sequences of selected Hip homologs were aligned using the EBI Clustal-X server. Secondary structure elements for Hip from *Rattus norvegicus* are indicated above the sequences. The Hip domain structure is indicated by purple, blue and orange coloring of secondary structure elements in the dimerization, TPR and DP domains, respectively. Similar residues are shown in red and identical residues in white on a red background. Blue frames indicate homologous regions. The consensus sequence is shown at the bottom. Downward pointing arrowheads indicate interface residues facing the NBD of Hsp70. Blue and cyan colors indicate contacts to subdomains IA/IB and IIB of the Hsp70 NBD, respectively. Known acetylation and phosphorylation sites are indicated by asterisks in dark blue and purple, respectively. Reported ubiquitylation sites are shown as hollow circles. Mutation sites are indicated by upward pointing arrowheads, and colored according to their effect on Hsp70 binding. Green, yellow and red colors indicate decreased, neutral and increased dissociation rates of MABA-ADP from Hsp70N. The Uniprot/TREMBL accession codes for the sequences are: P50503, *Rattus norvegicus*; B3RY90, *Trichoplax adhaerens*; G5EE04, *Caenorhabditis elegans*; G4VJ1, *Schistosoma mansoni*; Q86DS1, *Drosophila melanogaster*; C4M2C4, *Entamoeba histolytica*; C9ZKP5, *Trypanosoma brucei gambiense* (strain MHOM/CI/86/DAL972); A4HH33, *Leishmania braziliensis*; A4S4D2, *Ostreococcus lucimarinus* (strain CCE9901); B9Q4N1, *Toxoplasma gondii*; Q8I3J0, *Plasmodium falciparum* (isolate 3D7); Q93YR3, *Arabidopsis thaliana*.

8.3 Abbreviations

| | |
|---------------|---|
| AD | Alzheimer's Disease |
| ADP | Adenosine 5'-diphosphate |
| AIP | Aryl hydrocarbon receptor interacting protein |
| AMP | Ampicillin |
| APS | Ammonium peroxydisulfate |
| AR | Androgen receptor |
| ATP | Adenosine 5'-triphosphate |
| BAG | Bcl2-associated athanogene |
| BLAST | Basic Local Alignment Search Tool |
| bp | base pairs |
| BSA | Bovine serum albumin |
| CD | Circular dichroism |
| CD | C-terminal domain |
| CFTR | Cystic-fibrosis transmembrane-conductance regulator |
| CHIP | C-terminus of Hsp70-interacting protein |
| CIP | Calf intestinal phosphatase |
| CV | Column volume |
| CXCR2 | C-X-C-motif chemokine receptors type 2 |
| CXCR4 | C-X-C-motif chemokine receptors type 4 |
| DNA | Deoxyribonucleic acid |
| DnaJ | Bacterial Hsp40 chaperone |
| DnaK | Bacterial Hsp70 chaperone |
| dNTPs | Deoxyribonucleoside triphosphates |
| DOC | Deoxycorticosterone |
| DTT | Dithiothreitol |
| ECL | enhanced chemiluminescence |
| <i>E.coli</i> | <i>Escherichia coli</i> |
| EDTA | Ethylenediaminetetraacetic acid |
| ER | Endoplasmic reticulum |

| | |
|------------------|--|
| ESRF | European Synchrotron Radiation Facility |
| FKBP | FK506-binding protein |
| GR | Glucocorticoid Receptor |
| GRE | Glucocorticoid Responsive Element |
| GRK5 | G protein-coupled receptor Kinase 5 |
| GroEL | Large growth E gene product |
| GroES | Small growth E gene product |
| GrpE | Growth P-like gene E |
| HEPES | N-(2-hydroxyethyl)piperazin-N'-2-ethanesulfonic acid |
| Hip | Hsp70-interacting protein |
| His ₆ | Hexa-histidine tagged |
| Hsf1 | Heat shock factor 1 |
| Hop | Hsp70-Hsp90 organizing protein |
| HPD | histidine-proline-aspartate |
| Htt | Huntingtin |
| Hsp | Heat shock protein |
| HspBP1 | Hsp70 binding protein 1 |
| IMAC | Immobilised metal ion affinity chromatography |
| IPTG | Isopropyl- β -D-1 thiogalactopyranoside |
| ITC | Isothermal titration calorimetry |
| Kan | Kanamycin |
| LB | Luria Bertani medium |
| MD | Middle domain |
| MOPS | 3-(N-morpholino)propanesulfonic acid |
| MW | Molecular weight |
| NAC | Nascent chain-associated complex |
| NBD | Nucleotide binding domain |
| ND | N-terminal domain |
| NEF | Nucleotide exchange factor |
| NMR | Nuclear magnetic resonance |
| OD | Optical density |

| | |
|---------------------|--|
| PCR | Polymerase chain reaction |
| PDB | Protein data bank |
| PEG | Polyethylene glycol |
| <i>Pfu</i> | <i>Pyrococcus furiosus</i> |
| PGK | Phosphoglycerate kinase |
| PMSF | Phenylmethylsulfonylfluoride |
| PP5 | Phosphoprotein phosphatase 5 |
| RAC | Ribosome-associated complex |
| rpm | revolutions per minute |
| RT | Room temperature |
| SBD | Substrate binding domain |
| SC | Synthetic complete |
| <i>S.cerevisiae</i> | <i>Saccharomyces cerevisiae</i> |
| SDS-PAGE | Sodiumdodecylsulfate polyacrylamide gelelectrophoresis |
| sHsp | Small heat shock protein |
| SIRAS | Single isomorphous replacement with anomalous scattering |
| SLS | Swiss Synchrotron Light Source |
| SOD | Superoxide dismutase 1 |
| TEM | Transmission Electron microscopy |
| TEMED | N, N, N', N'-tetramethylethylenediamine |
| TEV | Tobacco etch virus |
| TF | Trigger factor |
| TPR | Tetratricopeptide repeat |
| TRiC | TCP-1 ring complex |
| Tris | Tris(hydroxymethyl)aminomethane |
| UPS | Ubiquitin Proteasome System |
| UV | Ultra violet |
| V | Volts |
| v/v | Volume per volume |
| w/v | Weight per volume |

8.4 Curriculum Vitae

Name: Zhuo Li
Date of birth: 05.April.1980
Birth of place: Dalian, Liaoning, China
Citizenship: Chinese

EDUCATION

11/2009 – present

PhD Dissertation

Max Planck Institute of Biochemistry, Martinsried, Germany
Cellular Biochemistry Department of Prof. Dr. F. U. Hartl

Title: “Structure and Function of Hip, an Attenuator of the Hsp70 Chaperone Cycle”

05/2007 – 10/2009

Master of Science (M. Sc.)

Ludwig-Maximilians-University Munich (LMU), Germany
Master thesis in the Cellular Biochemistry Department of Prof. Dr. F. U. Hartl, Max Planck Institute of Biochemistry, Martinsried

Title: „Structural and Functional Characterization of the Hsp70 Co-chaperone Hip”

09/2003 – 04/2007

Bachelor of Science (B. Sc.)

Ludwig-Maximilians-University Munich (LMU), Germany
Bachelor thesis in the lab Prof. Dr. Paul Knochel at LMU

Title: „Towards a New Method for Palladium-catalyzed *N*-arylation“

09/1998 – 07/2002

Bachelor of Art (B. A.)

Liaoning University, Shenyang, China

PUBLICATIONS

Zhuo Li, F. Ulrich Hartl and Andreas Bracher
Structure and function of Hip, an attenuator of the Hsp70 chaperone
Nature Structural & Molecular Biology, 20, 929-935 (2013)

Hongjun Ren, **Zhuo Li** and Paul Knochel
Chemoselective C(sp³)-H-Bond Activation for the Preparation of
Condensed *N*-Heterocycles
Chem. Asian J. 2007, **2**, 416-433.