

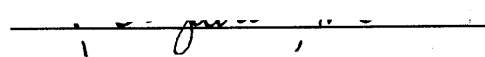
# Identification and Characterization of *fullback*, a Novel Posteriorly-Expressed *Xenopus* Gene

Master's thesis submitted by M. Elizabeth Hick


Massachusetts Institute of Technology

May 19, 1999

The author hereby grants to MIT permission to reproduce and to distribute publicly paper and electronic copies of this thesis document in whole or in part

 M. Elizabeth Hick, Student

 Hazel Sive, Advisor

 Terry Orr-Weaver, Co-Chair of Graduate Committee,  
Department of Biology

Science

## Abstract:

We have identified *fullback*, a *Xenopus laevis* gene which shows highest similarity to the low affinity nerve growth factor receptor *p75*. Extracellular portions of the protein are conserved between *fullback* and *p75*, although regions of *p75* which have been implicated in its role in apoptosis are not especially conserved in *fullback*. *fullback* is expressed in the posterior of the embryo from gastrulation onwards and is found dorsally during tailbud stages. This pattern of expression apparently does not coincide with the incidence of apoptosis in the early *Xenopus* embryo and therefore indicates that *fullback* may have a different function during embryogenesis.

## Results:

The low affinity nerve growth factor receptor *p75* is a member of the "death receptor" family of transmembrane proteins, most of which have some role in apoptosis (Liepinsh, et al, 1997). This receptor sends a pro-apoptotic signal leading to neuronal death unless it binds a neurotrophic ligand such as NGF which causes it to send an anti-apoptotic signal instead (reviewed in Bredesen and Rabizadeh, 1997). Using a yeast invertase screen to find secreted and transmembrane proteins which may play a role in patterning the early embryo, we have identified *fullback*, a *Xenopus* gene with highest similarity to *p75* (Jacobs et al, 1997). Here we report a sequence comparison of *fullback* to *p75* in other vertebrates and a spatial and temporal analysis of *fullback* expression in *Xenopus*.

*fullback* encodes a 427 amino acid protein with a signal sequence peptide at the amino terminus (von Heijne, 1986) and a single putative transmembrane domain. While the *p75* genes from human, rat and chicken share between 75-85%

identity with each other, *fullback* is only approximately 40% similar to any of them and is therefore unclear whether *fullback* is a *Xenopus* homolog of p75 (Fig 1). The death receptor family of proteins is characterized by their cysteine-rich extracellular domains (Liepinsh et al, 1997). A protein sequence alignment of *fullback* with the three p75 proteins illustrates that all 25 of the extracellular cysteines in p75 are also conserved in this region of *fullback* (Fig 1, boxed). This implies that *fullback* may have extracellular structural similarity to p75. However, the "death domain", a six-helix portion of the intracellular domain present in most death receptor family members which is thought to be essential for apoptotic signaling (underlined, Fig 1) (Tartaglia, et al, 1993) is not well conserved in *fullback*, suggesting that this protein may have an entirely different, non-apoptotic function in the embryo.

Northern analysis of *fullback* expression (Fig 2), shows a full length transcript of approximately 3kb. Weak maternal expression can be detected by RT-PCR (data not shown), but is not apparent by Northern blot. Zygotic expression of *fullback* begins at midgastrula stages, peaks during neurulation and then declines by swimming tadpole stages.

In situ hybridization analysis was performed in order to determine the location of *fullback* transcripts within the embryo (Fig 3). During gastrula and neurula stages, *fullback* expression is posteriorly localized. At early gastrula (Fig 3A, panel a) weak expression is found in a wide rim around the blastopore. At midgastrula (panel b) *fullback* expression remains around the blastopore and increases in intensity. A cross section through a halved embryo of this stage (panel f) shows that *fullback* is expressed in the superficial tissue of the blastopore lip but not in the involuted mesoderm. At neurulation (panel c) *fullback* reaches its highest levels of expression and is found in a ring around the closed blastopore in the

posterior of the embryo, both in the ectoderm and in the underlying mesoderm (panel g). At this stage, dorsal expression begins to extend out from the posterior of the embryo in a small streak along the midline (arrow, panel c). Cross sections of neurula embryos suggest that this staining does not coincide with the notochord but may correspond to migrating neural crest. At late neurula (panel d) expression persists in the posterior of the embryo and continues to extend dorsally into regions near the developing somites. A comparative *in situ* done on halved embryos with *fullback* and *m-actin* probes clearly shows non-overlapping regions of expression, implying that this dorsal *fullback* expression is not myotomal (panels h and j). At tailbud stages *fullback* is maintained in the posterior and the dorsal expression expands ventrally (panel e). A transverse cut through these embryos reveals that the dorsal staining overlaps with *muscle actin* staining in the somite, implying that at this stage *fullback* RNA is present in the myotome (panels i and k).

In order to assess whether *fullback* plays a role in modulating apoptosis, we compared its expression with TUNEL assays which profile the temporal and spatial occurrence of apoptosis within the *Xenopus* embryo (Hensey and Gautier, 1998). Using this assay, these authors found that apoptosis begins at midgastrula stages in a variable pattern including every region of the embryo except the cells immediately surrounding the blastopore. Later, TUNEL staining coincides with regions of neurogenesis in the ectoderm during neurula and tailbud stages. These regions of the embryo do not correspond with those expressing *fullback*, implying that *fullback* is not essential for apoptotic signaling. These data, together with the significant intracellular sequence divergence between p75, suggest that the function of *fullback* is different from that of p75.

## Methods:

The full length clone of *fullback* was isolated from a random-primed cDNA library made from ventral mesoderm and ectoderm from mid-gastrula *Xenopus* embryos (stage 11.5). Embryos were staged according to Nieuwkoop and Faber, 1967. Two oligos were used to form an *EcoR* I adapter: BIS3, 5'-AATTCCATAGCAACAAACAGTA-3', unphosphorylated and BIS4, 5'-TACTGTTTGTGCTATGGG-3', phosphorylated.

The random-primed cDNA library was subjected to a yeast selection as described previously (Jacobs et al., 1997). The Genbank accession number of *fullback* is AF131890.

*in situ* hybridization was performed as described in Kuo, et al., 1998. Most *in situ* panels show albino embryos, although some show wild type embryos which have been bleached in 2% hydrogen peroxide for 45 minutes.

Northern blot analysis was performed as described in Sagerström et al., 1997.

fullback - - - - -MDKRGPIVTLCLLLLISKISAEDVCEESGLYTN 32  
 hp75 -MGAGATGRAMDGPRLLLLLLLLGVS LGGAKEACP TGLYTH 39  
 rp75 MRRAGAACSAMDRLRLLLLL ILGVSSGGAKETCS TGLYTH 40  
 cp75 - - - - -MAGFVPLLLLLLLPAGPTWG-SKEKCLTKMYTT 31

fullback SGKCCSLCPAGFGVVVPCGDSDTKCEPC IENSTFSDVRS A 72  
 hp75 SGECCKACNLGEGVAQPCGANQTVCEPCLD SVTFSDVVSA 79  
 rp75 SGECCKACNLGEGVAQPCGANQTVCEPCLDN VTFSDVVSA 80  
 cp75 SGECCKACNLGEGVVQPCGVNQTVCEPCLD SVTYSDTVSA 71

fullback KAKCQPCFTCCQSPSLTLESNCTREQD TVCRCPERQYLD S - 111  
 hp75 TEPCKPCTE CVG - LQSMSAPC VEADDAVCRCAYGYYQDET 118  
 rp75 TEPCKPCTE CLG - LQSMSAPC VEADDAVCRCAYGYYQDEE 119  
 cp75 TEPCKPCTQ CVG - LHSMSAPC VESDDAVCRCA YGYFQDEL 110

fullback NGICLPCQLCSKGHGVVSQC THNKNTVCCQLCSSGFYSEVK 151  
 hp75 TGRCEACRVCEAGSGLVFS CQDKQNTVCEECPDGTYSDEA 158  
 rp75 TGHCEACSVCEVGSGLVFS CQDKQNTVCEECP EGTYSDEA 159  
 cp75 SGSCKECSICEVGFGLMFP CRDSQD TVCEECP EGTFSDEA 150

fullback SSESPCLPCTRTECKETE VQIGDCVPQHDILCMDKDVPILK 191  
 hp75 NHVDPCLPCTVCEDETERQLRECTRWADAEC EEPGRWIT 197  
 rp75 NHVDPCLPCTVCEDETERQLRECTPWADAEC EEPGRWIP 198  
 cp75 NFVDPCLPCTICEENEVMVKECTATSDAEC RDLHPRWTT 189

fullback RTE--GGEN-----G-----TSAG 203  
 hp75 RSTPPEGSDS TAPSTQEPEAPEQDL IASTVAGV VTTVMG 237  
 rp75 RSTPPEGSDS TAPSTQEPEVPPEQDLVPSTVADM VTTVMG 238  
 cp75 HTPSLAGSDSPEPITRDP--FNTEGMATTLAD IVTTVMG 226

fullback SPH-FIPQDNSKNIPVYCS ILAAVVVGLIAYVAFKCYTT 242  
 hp75 SSQPVVTRGTTDNLIPVYCS ILAAVVVGLVAYIAFKRWNS 277  
 rp75 SSQPVVTRGTTDNLIPVYCS ILAAVVVGLVAYIAFKRWNS 278  
 cp75 SSQPVVSRGTADNLIPVYCS ILAAVVVGLVAYIAFKRWNS 266

fullback CKQKKQLAKARAGELATSTEGEK LHNDSGVFLDTHSLQE - 281  
 hp75 CKQNKQGANSRPVNQTPPEGEK LHSDSG ISVDSQS LHDQ 317  
 rp75 CKQNKQGANSRPVNQTPPEGEK LHSDSG ISVDSQS LHDQ 318  
 cp75 CKQNKQGANNRPVNQTPSPEGEK LHSDSG ISVDSQS LHDQ 306

fullback -PNHLSKAKIEPK---LYINLPPHKQSEVERLLADTSLG 316  
 hp75 QPHTQTASGQALKGDGGLYSSLP PAKREEVEKLLNG-SAG 356  
 rp75 QHTQTASGQALKGDGNLYSSLP LTKREEVEKLLN---G 354  
 cp75 QPPNQSTQGPA PKGDGSLYASLP PPSKQEEVEKLLSS-SAE 345

fullback KDWQR LASLLGYEEETIDTFGRGEDPVHTLLTDWSSK ESS 356  
 hp75 D TWRHLAGELGYQPEHIDSFTHEACPVRALLASWATQDSA 396  
 rp75 D TWRHLAGELGYQPEHIDSFTHEACPVRALLASWGAQDSA 394  
 cp75 E TWRQLAGELGYKEDLIDCF TREESPARALLADWSAKE TA 385

fullback TLEVLC AALVNMERADVVENLNS TNDASSV V  
 hp75 TLDALLAALRR IQRADLVESLCSESTATSPV 427  
 rp75 TLDALLAALRR IQRADIVESLCSESTATSPV 425  
 cp75 TLDALLVALRK IQRGDIAESLYSESTATSPV 416  
 387

Figure 1: Protein Sequence Alignment of fullback with p75 from Other Vertebrates

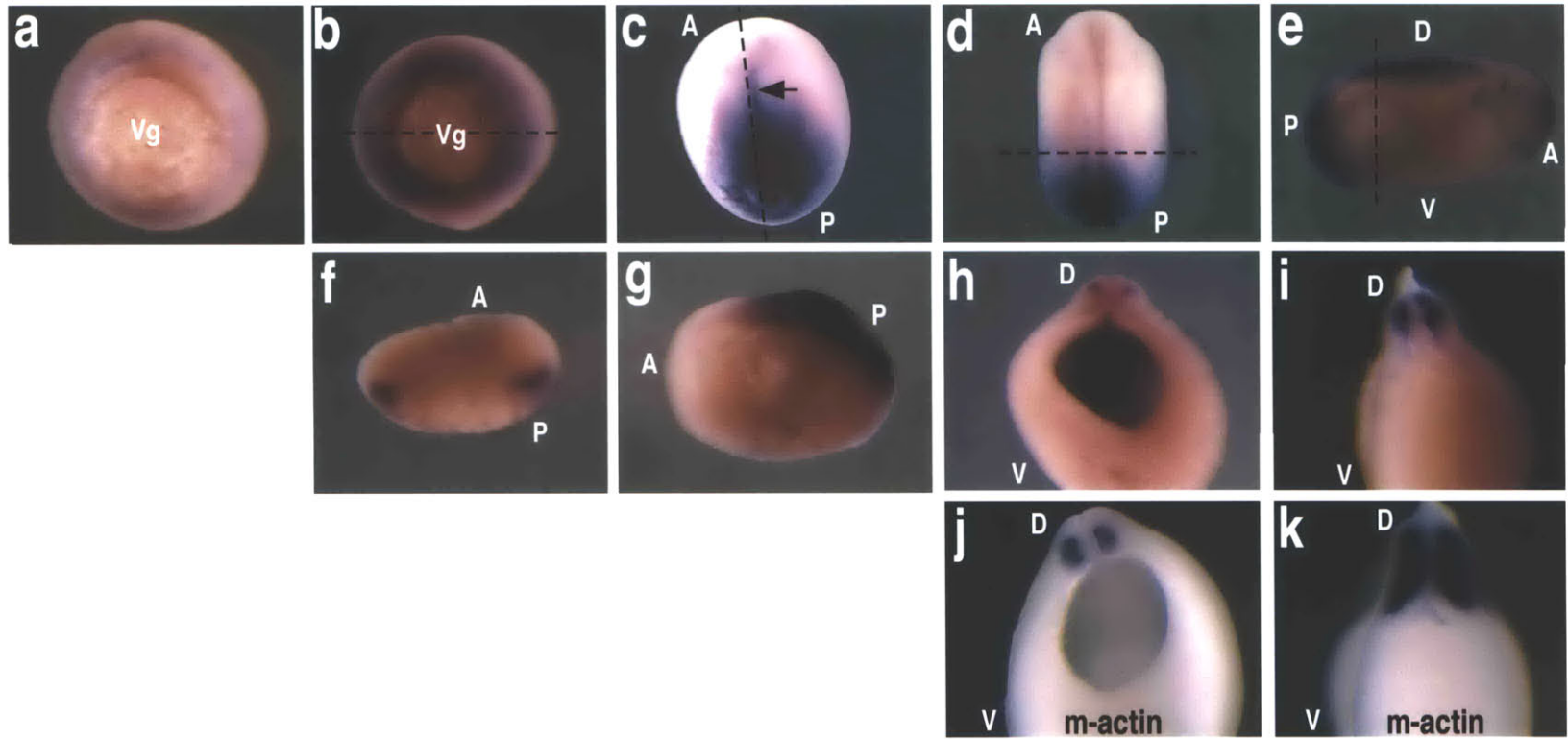
The p75 proteins shown are from human (hp75), rat (rp75) and chicken (cp75). Identical amino acids are shaded. Numbers to the right of the sequences indicate the numerical positions of the amino acids. Boxes denote cysteines conserved among all four proteins. The six-helix death domain region is underlined and overlined.





Figure 2: Northern Analysis of *fullback* mRNA

*fullback* RNA (upper panel). Lane 1 contains unfertilized egg RNA. Lane 2 contains RNA from early gastrula, stage 10. Lane 3 contains RNA from midgastrula, stage 11. Lane 4 contains RNA from early neurula, stage 13. Lane 5 contains RNA from midneurula, stage 15. Lane 6 contains RNA from early tailbud, stage 22. Lane 7 contains RNA from swimming tadpole, stage 35. 18s Ribosomal RNA is shown in the lower panel as a loading control.



### Figure 3: *In Situ* Hybridization Analysis of *fullback* RNA Expression

In situ hybridization was performed on whole (panels a-e) or halved (panels f-k) *Xenopus* embryos. Dashed lines in panels b-e denote the plane in which each embryo was cut in half.

a. Vegetal view of an early gastrula (stage 10.5). b. Posterior view of an embryo at mid-gastrula (stage 11.5). c. Dorsal view of an early neurula (stage 13). Arrow indicates the weakly perceptible stripe of staining at the dorsal midline. d. Dorsal view of a late neurula (stage 18). e. Lateral view of a tailbud (stage 25). f. Interior view of a halved embryo at mid-gastrula (stage 11.5). g. Interior view of an early neurula (stage 13) which has been parasagittally cut in half. h. Interior view of a late neurula halved transversely. The dorsal expression of *fullback* is shown. i. Interior view of a late tailbud (stage 25) embryo cut transversely in half.

Panels j and k: Comparative *in situ* hybridizations were performed using a probe for *muscle actin*, which is expressed in the myotome of the *Xenopus* somite. Panel j shows a late neurula embryo (stage 18) cut in half transversely. Panel k shows a tailbud stage embryo (stage 25).

In all panels, Vg= vegetal, A= anterior, P= posterior, D= dorsal, V= ventral.

## References:

Bredesen, D. E., and Rabizadeh, S. (1997). p75NTR and apoptosis: Trk-dependent and Trk-independent effects. *Trends Neurosci* 20, 287-90.

Hensey, C., and Gautier, J. (1998). Programmed cell death during *Xenopus* development: a spatio-temporal analysis. *Dev Biol* 203, 36-48.

Jacobs, K. A., Collins-Racie, L. A., Colbert, M., Duckett, M., Golden-Fleet, M., Kelleher, K., Kriz, R., La Vallie, E. R., Merberg, D., Spaulding, V., Stover, J., Williamson, M. J., and McCoy, J. M. (1997). A genetic selection for isolating cDNAs encoding secreted proteins. *Gene* 198, 289-96.

Johnson, D., Lanahan, A., Buck, C. R., Sehgal, A., Morgan, C., Mercer, E., Bothwell, M., and Chao, M. (1986). Expression and structure of the human NGF receptor. *Cell* 47, 545-54.

Kaplan, D. R., and Miller, F. D. (1997). Signal transduction by the neurotrophin receptors. *Curr Opin Cell Biol* 9, 213-21.

Kuo, J. S., Patel, M., Gamse, J., Merzdorf, C., Liu, X., Apekin, V., and Sive, H.L. (1998) opl: A Zinc Finger Protein that Regulates Neural Determination and Patterning in *Xenopus*. *Development* 125, 2867-2882.

Large, T. H., Weskamp, G., Helder, J. C., Radeke, M. J., Misko, T. P., Shooter, E. M., and Reichardt, L. F. (1989). Structure and developmental expression of the nerve growth factor receptor in the chicken central nervous system. *Neuron* 2, 1123-34.

Liepinsh, E., Ilag, L. L., Otting, G., and Ibanez, C. F. (1997). NMR structure of the death domain of the p75 neurotrophin receptor. *Embo J* 16, 4999-5005.

Nieuwkoop, P., and Faber, J. (1967). Normal table of *Xenopus laevis* (Daudin), 2nd Edition (Amsterdam: North-Holland Publishing Co.).

Radeke, M. J., Misko, T. P., Hsu, C., Herzenberg, L. A., and Shooter, E. M. (1987). Gene transfer and molecular cloning of the rat nerve growth factor receptor. *Nature* 325, 593-7.

Sagerström, C. and Sive, H. L. (1997). Northern hybridization analysis. *RNA Methods*. Krieg, P., ed. Academic Press, pp 83-104.

Tartaglia, L. A., Ayres, T. M., Wong, G. H., and Goeddel, D. V. (1993). A novel domain within the 55 kd TNF receptor signals cell death. *Cell* 74, 845-53.

von Heijne, G. (1986). A new method for predicting signal sequence cleavage sites. *Nucleic Acids Res* 14, 4683-90.