



Institutional Repository - Research Portal

Dépôt Institutionnel - Portail de la Recherche

researchportal.unamur.be

RESEARCH OUTPUTS / RÉSULTATS DE RECHERCHE

Reprogramming of tumor-associated macrophages with anticancer therapies

Genard, Géraldine; Lucas, Stéphane; Michiels, Carine

Published in:
Frontiers in Immunology

DOI:
[10.3389/fimmu.2017.00828](https://doi.org/10.3389/fimmu.2017.00828)

Publication date:
2017

Document Version
Publisher's PDF, also known as Version of record

[Link to publication](#)

Citation for pulished version (HARVARD):
Genard, G, Lucas, S & Michiels, C 2017, 'Reprogramming of tumor-associated macrophages with anticancer therapies: Radiotherapy versus chemo- and immunotherapies' *Frontiers in Immunology*, vol. 8, no. JUL, 828. <https://doi.org/10.3389/fimmu.2017.00828>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



Reprogramming of Tumor-Associated Macrophages with Anticancer Therapies: Radiotherapy versus Chemo- and Immunotherapies

Géraldine Genard^{1,2}, Stéphane Lucas² and Carine Michiels^{1*}

¹URBC – NARILIS, University of Namur, Namur, Belgium, ²Laboratory of Analysis by Nuclear Reaction (LARN/PMR) – NARILIS, University of Namur, Namur, Belgium

OPEN ACCESS

Edited by:

Anahid Jewett,
University of California,
Los Angeles, United States

Reviewed by:

Nejat K. Egilmez,
University of Louisville,
United States
Benjamin Bonavida,
University of California,
Los Angeles, United States

*Correspondence:

Carine Michiels
carine.michiels@unamur.be

Specialty section:

This article was submitted to Cancer
Immunity and Immunotherapy,
a section of the journal
Frontiers in Immunology

Received: 25 April 2017

Accepted: 30 June 2017

Published: 14 July 2017

Citation:

Genard G, Lucas S and Michiels C
(2017) Reprogramming of
Tumor-Associated Macrophages with
Anticancer Therapies: Radiotherapy
versus Chemo- and
Immunotherapies.
Front. Immunol. 8:828.
doi: 10.3389/fimmu.2017.00828

Tumor-associated macrophages (TAMs) play a central role in tumor progression, metastasis, and recurrence after treatment. Macrophage plasticity and diversity allow their classification along a M1–M2 polarization axis. Tumor-associated macrophages usually display a M2-like phenotype, associated with pro-tumoral features whereas M1 macrophages exert antitumor functions. Targeting the reprogramming of TAMs toward M1-like macrophages would thus be an efficient way to promote tumor regression. This can be achieved through therapies including chemotherapy, immunotherapy, and radiotherapy (RT). In this review, we first describe how chemo- and immunotherapies can target TAMs and, second, we detail how RT modifies macrophage phenotype and present the molecular pathways that may be involved. The identification of irradiation dose inducing macrophage reprogramming and of the underlying mechanisms could lead to the design of novel therapeutic strategies and improve synergy in combined treatments.

Keywords: tumor-associated macrophages, reprogramming, polarization immunotherapy, chemotherapy, radiotherapy, nuclear factor kappa B, reactive oxygen species

Abbreviations: AMPK, AMP-activated protein kinase; AP-1, activator protein 1; Arg 1, arginase 1; ATM, ataxia telangiectasia mutated; C/EBP β , CCAAT-enhancer-binding proteins; CCL, chemokine ligand; CCR2, C–C chemokine receptor type 2; CD206, cluster of differentiation 206 or mannose receptor; CL, clodrolip; COX, cyclooxygenase; CpG-ODN, unmethylated cytosine-guanine (CpG) oligodeoxynucleotides; CSF-1, colony-stimulating factor 1; CSF-1R, colony-stimulating factor 1 receptor; CTLA4, cytotoxic T lymphocyte-associated protein 4; CXCL12, C–X–C motif chemokine 12; ECM, extracellular matrix; ERK, extracellular signal-regulated kinase; FDA, food and drug administration; FR β , folate receptor β ; HDI, high doses of irradiation; HIF, hypoxia-inducible factor; HLA-DR, human leukocyte antigen-cell surface receptor; HO1, heme oxygenase 1; HRG, histidine-rich glycoprotein; IFN γ , interferon γ ; IL, interleukin; iNOS, inducible nitric oxide synthase; IR, ionizing radiation; IRF, interferon-regulatory factor; JNK, c-Jun N-terminal kinase; KLF4, Kruppel-like factor 4; LDI, low doses of irradiation; LET, linear energy transfer; LLC, Lewis lung cancer; LPS, lipopolysaccharide; mAb, monoclonal antibody; MAPK, mitogen-activated protein kinase; MARCO, pattern recognition scavenger receptor; MCP-1, monocyte chemoattractant protein 1; M-CSF, macrophage colony-stimulating factor; MDI, moderate doses of irradiation; MHC, major histocompatibility complex; MMP, matrix metalloproteinase; MnSOD, manganese superoxide dismutase; MPK1, MAPK phosphatase 1; MRC1, mannose receptor-C 1; NAC, N-acetyl cysteine; NADPH, nicotinamide adenine dinucleotide; NEMO, NF κ B essential modulator; NF κ B, nuclear factor kappa B; NK, natural killer; NO, nitric oxide; NOX, NADPH oxidase; Nrf2, nuclear erythroid derived 2-related factor; p38, MAPK; PAMPs, pathogen-associated molecular patterns; PD-1, programmed cell death 1; PD-L1, programmed death ligand 1; PDA, pancreatic ductal adenocarcinoma; PEC, peritoneal exudate cells; PI3K γ , phosphoinositide 3-kinase gamma; PlGF, placental growth factor; PPAR γ , peroxisome proliferator-activated receptor gamma; RANTES, regulated on activation, T cell expressed and secreted; RELM α , resistin-like molecule alpha; RNS, reactive nitrogen species; ROS, reactive oxygen species; RT, radiotherapy; STAT, signal transducer and activator of transcription; SUMO, small ubiquitin-like modifier; TAMs, tumor-associated macrophages; TGF β , transforming growth factor β ; TLR, toll-like receptor; TNF α , tumor necrosis factor α ; TRAIL-R2, TNF α -related apoptosis-inducing ligand receptor; VCAN, versican, extracellular matrix proteoglycan; VEGF, vascular endothelial growth factor; ZA, zoledronic acid.

INTRODUCTION

In 1863, Rudolf Virchow was the first to highlight the infiltration of leukocytes in tumor, thereby proposing a link between inflammation and tumorigenesis. Two centuries later, an inflammatory microenvironment within the tumor is part of the hallmarks of cancer (1). In fact, in emerging cancer disease, inflammation is a two-edge sword. On the one hand, the immune system can recognize tumor cells and kill them. On the other hand, chronic inflammation promotes cancer invasion, angiogenesis, and immunosuppression (2, 3). In this context, the immune system was assigned tumor immunoediting functions. The emergence of neoplastic cells induces an inflammatory environment, contributing to the rejection of the tumor (the *elimination phase*). However, the immune system establishes a selective pressure on cancer cells hence selecting resistant cells (the *equilibrium phase*). Finally, tumor evades from the immune system and moves into the *escape phase* (4). This last phase can occur through different mechanisms: reduced immune recognition, increased cancer cell resistance or survival, and immunosuppressive tumor microenvironment (5). In this process, macrophages act as an orchestrator of inflammation and are the main players of immunosurveillance. In the elimination phase, transformed cells are recognized and their antigens are presented to the effectors of the immune system by macrophages, promoting antitumor immunity (6). However, in the escape phase, macrophages play an important role in tumor progression by stimulating angiogenesis, metastasis, tumor growth, and immunosuppression notably through the secretion of polyamines, M-CSF, vascular endothelial growth factor (VEGF), IL-10, and transforming growth factor β (TGF β) (5).

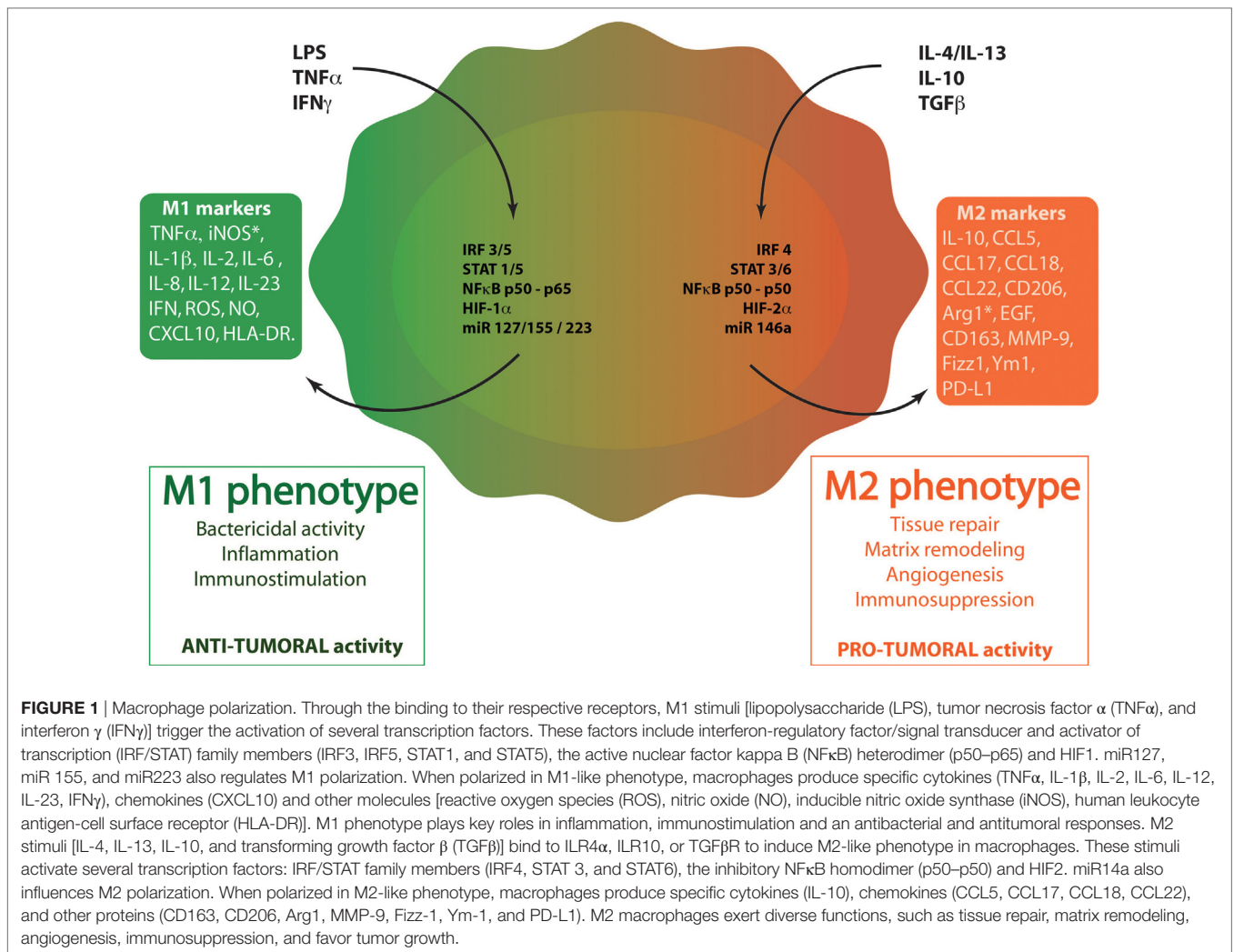
COMPLICITY OF TUMOR-ASSOCIATED MACROPHAGES (TAMs) IN TUMOR PROGRESSION

More than 50% of tumor-infiltrating cells are macrophages, named TAMs (7, 8). The recruitment and accumulation of TAMs into tumors are initiated by macrophage chemoattractants [e.g., CCL2/monocyte chemoattractant protein 1 (MCP-1), colony-stimulating factor 1 (CSF-1)] and it is well established that TAMs drive tumor progression (9, 10). In fact, cancer prognosis is closely linked to the number of TAMs with an inverse correlation: increased number of TAMs is associated with a reduced cancer patient survival (11). In healthy tissues, macrophages offer a remarkable plasticity to efficiently respond to environmental cues (12). In tumors, an identical plasticity is described: TAMs are educated by the tumor microenvironment, providing multiple phenotypes with a range of functions (13). TAM phenotypes can be featured as a linear scale where M1 and M2 phenotypes represent the two extremes, similarly to the T_H1–T_H2 classification (Figure 1). M1 macrophages are recognized as classically activated macrophages and show enhanced ability to phagocytose pathogens. More importantly, these cells have antitumoral properties. Macrophages can also be polarized into the M2 phenotype, the alternative activated state of macrophages. M2 macrophages are requested in infection-free healing circumstances and have pro-tumoral functions. The polarization of macrophages can be

driven by different microenvironmental molecules and leads to the production by the macrophages of different cytokines and chemokines. M1 macrophages are activated during acute inflammation by toll-like receptor (TLR) ligands [lipopolysaccharide (LPS)] or T_H1 cytokines [interferon γ (IFN γ)–tumor necrosis factor α (TNF α)]. M1 macrophages display an enhanced production of pro-inflammatory cytokines (TNF α and interleukins: IL-1 β , IL-2, IL-6, IL-12, IL-23), reactive oxygen species (ROS), nitric oxide (NO) and present antigens *via* major histocompatibility complex class II molecules (14). Stimulation of macrophages with IL-4/IL-13, IL-10, TGF β , or glucocorticoids leads to the M2 phenotype and the subsequent production of anti-inflammatory cytokines (IL-10, TGF β) that have an inhibitory effect on cytotoxic CD8⁺ T cells. Macrophages with M2 phenotype also express cell surface scavenger receptor (CD206), hemoglobin receptor (CD163) and produce extracellular matrix (ECM) components (14–16). M2 macrophages facilitate the resolution of inflammation and promote tissue repair by T_H2 response, tissue remodeling, and immune tolerance. They also favor tumor growth (17, 18). This phenotype can be subclassified into M2a, M2b, or M2c according to the roles these macrophages exert (19). In cancer disease, TAMs usually exhibit a M2 phenotype and participate to tumor angiogenesis, tumor invasion and metastasis, immunosuppression and cell activation. All these features led Qian and Pollard to classify TAMs in six functional subtypes: angiogenic, immunosuppressive, invasive, metastasis associated, perivascular, and activated macrophages (20).

MOLECULAR PATHWAYS FOR M1–M2 POLARIZATION

The polarization of macrophages is influenced by several mechanisms driven by transcription factors and miRNAs (Figure 1). The activation of macrophages into M1 or M2 phenotype is mainly induced by interferon-regulatory factor/signal transducer and activator of transcription (IRF/STAT) signaling pathways. IRF3, IRF5, STAT1, and STAT5 are responsible for driving M1 polarization while IRF4, STAT3, and STAT6 provide M2 activation signals (14, 21). Besides IRF/STAT transcription factors, hypoxia also influences macrophage polarization. The lack of oxygen can activate hypoxia-inducible factors (HIF) differently in M1 versus M2 macrophages. Indeed, T_H1 cytokines are able to induce HIF1 α stabilization, triggering M1 response whereas HIF2 α is activated by T_H2 cytokines in M2 macrophages. These differences rely on the ability of HIF1 and HIF2 to, respectively, activate or suppress NO synthesis (22). Furthermore, nuclear factor kappa B (NF κ B) fulfills a central role in the pro-inflammatory macrophages (M1). Indeed, the active heterodimer NF κ B (p50–p65) promotes the transcription of inflammatory genes while the inhibitory heterodimer NF κ B (p50–p50) prevents the transcription of these genes in anti-inflammatory macrophages (M2). Other transcription factors including AP-1, Kruppel-like factor 4 and PPAR γ can modulate the activated state of macrophages. Finally, miRNA also interfere with the polarization of macrophages. Among miRNAs of interest, miR127, miR155, and miR223 are key regulators of M1 polarization (23, 24). In contrast, miR146-a promotes M2 polarization while decreasing the expression of M1 markers (25).



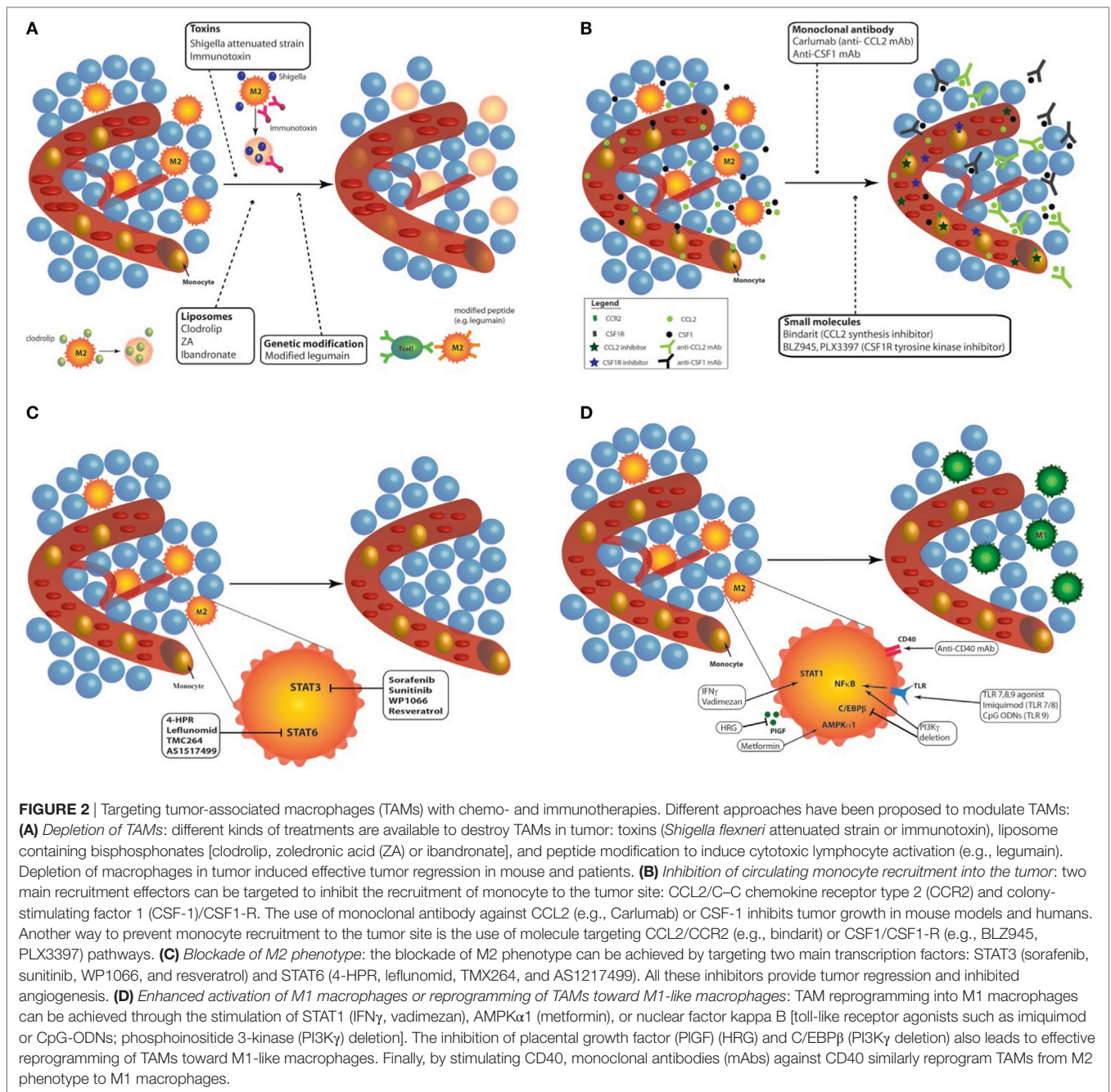
TARGETING TAMs WITH CHEMOTHERAPY OR IMMUNOTHERAPY

In response to microbial signals, tissue damage, cytokines and metabolic products, monocytes, and macrophages from healthy tissues are able to undergo reprogramming (12, 21, 26, 27). The pro-tumoral functions of TAMs and their ability to be reprogrammed, from M2-like macrophages toward M1 phenotype, make them an attractive target for anticancer therapies. Currently, different approaches have been proposed to modulate TAMs: (1) depletion of TAMs; (2) inhibition of circulating monocyte recruitment into the tumor; (3) blockade of M2 phenotype; and (4) enhanced activation of M1 macrophages or reprogramming of TAMs toward M1-like macrophages (27, 28). Each of these approaches will be detailed here under (Figure 2).

Depletion of TAMs

Three alternatives are available to destroy TAMs in tumor (Figure 2A). The first way is the use of chemical compounds, especially bisphosphonates, such as clodronate encapsulated

in liposomes [clodrolip (CL)]. These liposomes containing clodronate are phagocytosed by macrophages and are disrupted by lysosomal processes, leading to the release of clodronate in the cells. Clodronate is metabolized to an analog of ATP, cytotoxic for macrophages (29). CL is currently used in immune research and clinical trials to eliminate macrophages and phagocytes in multiple malignancies. For example, TAM depletion with CL impaired tumor engraftment, reduced tumor growth, and favored mouse survival in a chronic lymphocytic leukemia model (30). However, despite its effective tumor regression effects, it also induced severe side effects such as sensitivity toward infections and weight loss in mice and patients (31–34). A similar approach was developed using zoledronic acid (ZA), which inhibited tumor progression, angiogenesis, and metastasis in association with sorafenib in two human hepatocellular carcinoma mouse models (34). ZA depleted matrix metalloproteinase 9 (MMP-9) expressing TAMs (M2-like macrophages) but also impaired the differentiation of myeloid cells into TAMs (35). However, ZA, as ibandronate (another bisphosphonate), has generated conflicting results.



The targeting of MMP-9 positive macrophages was also obtained with dasatinib, a tyrosine kinase inhibitor well tolerated by patients and approved by food and drug administration (FDA) for chronic myeloid leukemia (36). However, dasatinib has been reported to inhibit other immune cells as natural killer (NK) cells and T cells, then aborting tumor immune response. Finally, trabectedin (ET-743), another drug approved by FDA, exhibited a cytotoxic effect against TAMs and showed antitumor activity. This drug was shown to not damage lymphocyte subgroups. This is due to the differential expression of TNF α -related apoptosis-inducing ligand receptor (TRAIL) (TNF α -related apoptosis-inducing ligand) receptor

by the different types of leukocytes: trabectedin targets TAMs while human NK cells and CD8⁺ lymphocytes were resistant to TRAIL-mediated toxicity. In fact, blood monocytes and TAMs expressed higher level of TRAIL-R2 compared to NK cells and CD8⁺ lymphocytes (37, 38).

A second alternative is the use of toxin-conjugated monoclonal antibodies (mAbs) or attenuated bacteria that kill macrophages. An anti-scavenger receptor A (CD204) antibody conjugated to the saporin toxin reduced the number of vascular leukocytes and inhibited tumor progression in a murine ovarian cancer model (39). An immunotoxin composed of portions of anti-folate receptor beta antibody conjugated

to *Pseudomonas* exotoxin A was also used to deplete TAMs: reduced tumor growth was observed in nude mice bearing C6 glioma xenografts (40). In the same line, *Shigella flexneri* is a bacterial pathogen inducing the apoptosis of macrophages. Galmbacher and his colleagues developed an attenuated strain of *S. flexneri* to infect breast cancer-bearing mice. The treatment led to TAM depletion and to a complete tumor regression in tumor-bearing mice (41).

The last alternative is the activation of cytotoxic T lymphocytes against macrophages. Legumain is an asparaginyl endopeptidase that contributes to the degradation of the ECM and to angiogenesis. TAMs have been found to express abundant amounts of legumain (42). This discovery led Smahel and his team to modify the gene sequence of legumain to enhance the efficacy of immunization against legumain. These modifications induced reduced legumain maturation and impaired cellular localization, and resulted in T helper (CD4⁺) and cytotoxic (CD8⁺) lymphocyte-dependent elimination of TAMs (43).

Depletion of macrophages has thus met successful results in the tumor regression in mouse models and patients. However, a systemic depletion of macrophages obviously renders the organism more sensitive to infections and other aggressions. Therefore, these types of treatments need to be localized to the tumor site.

Inhibition of Circulating Monocyte Recruitment into Tumor

The inhibition of CCL2/C-C chemokine receptor type 2 or CSF1/CSF1-R pathways by several methods was shown to efficiently induce tumor regression (**Figure 2B**). CCL2 is a chemokine produced by cancer cells that is responsible for the recruitment of monocytes at the tumor site. CCL2 overexpression in tumor is correlated with macrophage infiltration and poor prognosis in human cancers (44–46). Furthermore, CCL2 is a key actor of metastasis since it enhances the retention of metastasis-associated macrophages in breast cancer metastasis. CCL2 blockade was reported to block the mobilization of monocytes from the bone marrow to the blood in a murine breast cancer model (47). The inhibition of CCL2 by an anti-CCL2 monoclonal antibody (e.g., carlumab) or through the inhibition of its synthesis (e.g., bindarit, trabectedin) prevented the recruitment of macrophages into the tumor site. Most of these treatments are now tested in clinical trials. Carlumab, a human monoclonal antibody against CCL2, was safely used in metastatic castration-resistant prostate cancer (48) and in a phase Ib study in association with other chemotherapy agents (docetaxel, gemcitabine, paclitaxel or carboplatin and pegylated liposomal doxorubicin) (49). This monoclonal antibody led to an efficient depletion of macrophages into the tumor and can significantly delay tumor regrowth following chemotherapy (49). Besides mAb targeting CCL2, other compounds (e.g., bindarit, trabectedin) were found to inhibit the synthesis of CCL2/MCP-1. Bindarit reduced TAM and myeloid-derived suppressor cell infiltration in a breast cancer model and resulted in impaired metastatic disease in a prostate cancer model (50). This treatment also targeted angiogenesis and tumor growth in

human melanoma xenografts (51). In addition to deplete TAMs, trabectedin is used to inhibit monocyte recruitment. This drug targets CCL2 synthesis by interacting with the transcription machinery (52) but also modulates the DNA repair machinery (53). In clinic, it is used to treat ovarian and breast cancer as well as soft tissue sarcomas (53, 54).

Colony-stimulating factor 1/CSF1-R signaling drives the recruitment and the differentiation of TAMs toward a M2 phenotype in tumor. CSF1-deficient mice showed a 50% decrease in macrophage infiltration while neutrophil infiltration was increased during tumor progression in a mouse model of pancreatic islet cancer (55). mAbs and small molecules targeting CSF1 (mAb anti-CSF1) or CSF1-R (BLZ945, emactuzumab, PLX3397) were subjected to numerous studies and were shown to deplete macrophages in a tissue-specific manner (56, 57). Overall, CSF1-R inhibitors deplete TAMs and abolish tumor growth, angiogenesis, and metastasis. BLZ945, a highly selective small molecule inhibitor of CSF1-R tyrosine kinase, attenuated the turnover rate of TAMs while increasing the number of CD8⁺ T cells in murine cervical and breast carcinoma models (56). In addition, blockade of CSF-1/CSF1-R by mAb anti-CSF1 or with PLX2297 (a small molecule targeting the tyrosine kinase domain of CSF1-R) was reported to reprogram remaining TAMs at the tumor site to support antitumor immunity in pancreatic cancer mouse models (58). PLX3397 also delayed the recurrence of glioblastoma after radiation by modifying the recruitment and polarization of myeloid cells in an intracranial xenograft model (59). However, orally administered PLX3397 showed no efficacy for human glioblastoma in a phase II study (60). Other chemoattractants for macrophages such as VEGF, C-X-C motif chemokine 12, CCL4 are also under investigation and may be considered as potential targets to inhibit macrophage recruitment and hence tumor progression.

Blockade of M2 Phenotype

Two main transcription factors have been largely reported to block M2 polarization: STAT 3 and STAT6 (**Figure 2C**). Tyrosine kinase inhibitors (sunitinib and sorafenib) inhibit STAT3 in macrophages. Sorafenib was shown to restore the secretion of IL-12 while suppressing IL-10 expression in prostaglandin E2-conditioned macrophages, indicating a reverse effect on the immunosuppressive cytokine profile in TAMs (61). Inhibition of STAT3 with WP1066 reversed immune tolerance in patients with glioblastoma multiforme. This treatment stimulated the secretion of pro-inflammatory cytokines and activated T cells (62). In the same line, resveratrol has also been used to suppress M2-like polarization of TAMs with parallel inhibition of tumor growth in a mouse lung cancer xenograft model. The blockade of M2-like polarization of TAMs by resveratrol was linked to the decreased activity of STAT3 (63).

Similarly, fenretinide (4-HPR) inhibited the phosphorylation of STAT6 and skew M2 polarization. In a colorectal mouse model, the effects were accompanied by a reduction in the number of M2-like macrophages in tumor and by an inhibition of angiogenesis (64). Other inhibitors of STAT6 activation (TMC264, AS1517499) were developed but no one underwent clinical studies (65–67).

Enhanced Activation of M1 Phenotype or Reprogramming of TAMs toward M1-Like Macrophages

Several options are currently used to select M1 phenotype from TAMs or to reprogram TAMs from M2 to M1 phenotype: TLR agonists, mAb targeting inhibitory proteins of M1 phenotype as well as other compounds (Figure 2D). TLR agonists represent a promising antitumor therapy. However, different agonists have been shown to promote different immune responses. Imiquimod is a ligand of TLR7 and acts mainly by increasing the number of infiltrating CD43⁺ lymphocytes. This TLR agonist was found to induce the nuclear translocation of NF κ B in J774A macrophages, leading to the production of pro-inflammatory proteins, such as TNF α , IL-6, IL-12, and CCL2 (68). The combination of topical imiquimod with low doses of cyclophosphamid and radiotherapy (RT) revealed a synergic antitumor effect and an increased survival response. This allowed the prevention of recurrence, with tumor rejection over 2 months after the end of treatment in a cutaneous breast cancer model (69). Multiple phase I and phase II clinical trials for imiquimod resulted in the use of topical application of imiquimod in clinic for skin metastasis and carcinomas. Synthetic unmethylated cytosine-guanine (CpG) oligodeoxynucleotides (CpG-ODNs) also offered high immunostimulatory activity. These molecules act by enhancing the production of pro-inflammatory cytokines, such as TNF α , IL-6 and IL-12, in macrophages and by upregulating NF κ B activity in these cells. Indeed, these nucleotides are frequently found in viral and bacterial genomes and are recognized as pathogen-associated molecular patterns by TLR9 (70–72). Multiple studies aimed to improve the delivery of these CpG-ODNs by coupling them to gold nanoparticles for example (73). Combination of CpG-ODNs with other therapies, such as anti-CD40 exhibited promising results associated with the repolarization of TAMs (74, 75), even in poorly immunogenic cancer models such as a preclinical glioma model (76).

Another alternative to favor cytotoxic functions of TAMs is the stimulation of CD40 with mAbs. CD40 mAbs have demonstrated antitumor T-cell responses in mouse models of cancer and in clinical trials (77). Macrophages also express CD40 in their plasma membrane. Anti-CD40 mAbs were shown to promote macrophage tumoricidal activity, especially through enhanced secretion of NO and TNF α . Hence, it could induce CD8⁺ T cell-dependent inhibition of tumor growth and metastasis (78–80). Clinical studies revealed objective tumor response in solid tumors (81). Since anti-CD40 mAbs can induce programmed death ligand 1 (PD-L1) upregulation in TAM and tumor-infiltrating monocyte plasma membrane, the blockade of PD-L1 axis combined with anti-CD40 and anti-CTLA-4 (anti-cytotoxic T-Lymphocyte-associated protein 4) mAbs showed extensive survival in colon and breast cancer models (77). Finally, MARCO is a pattern recognition scavenger receptor and is expressed by immunosuppressive TAMs. The targeting of this receptor is a new promising way to treat mammary carcinoma, colon carcinoma, and melanoma through the reprogramming of immunosuppressive TAMs toward a pro-inflammatory phenotype and by increasing tumor immune response (82).

A third alternative to reprogram TAMs is the use of different chemical compounds. The most famous one is IFN γ (83), approved by the FDA. In the same line, a small flavonoid-like compound, vadimezan (DMXAA), was found to repolarize macrophages in M1 phenotype. Reprogrammed macrophages then released cytokines and chemokines, including high local levels of TNF α , to induce a subsequent CD8⁺ T cell activation (84). Vadimezan has been the subject of numerous preclinical studies and clinical trials. Alternatively, macrophage phosphoinositide 3-kinase γ (PI3K γ) was shown to control a critical switch between immune suppression and activation. Selective deletion of PI3K γ simultaneously activates NF κ B and inhibits C/EBP β in macrophages. In combination with anti-PD-L1, PI3K γ depletion promoted tumor regression and prolonged survival in head and neck, lung, and breast cancer murine models (85). The plasma protein histidine-rich glycoprotein (HRG) is also able to block TAMs into a M1 phenotype through the downregulation of the expression of the placental growth factor (PlGF), a member of the VEGF family. In mice, HRG promoted antitumor immune responses and normalization of the vessel network (86). Finally, metformin was shown to skew TAMs polarization into a pro-inflammatory phenotype, partially through AMPK α 1 activation. This effect was concomitant with a decrease in the number of metastases in Lewis lung cancer intravenous model (87).

While depletion of macrophages induced toxicities in mouse models and humans, the inhibition of macrophage/monocyte recruitment is one of the most used therapies in clinical studies. This type of therapy is usually combined to chemotherapy or RT and generated encouraging results in patients. On the other hand, the reprogramming of TAMs by chemotherapies or immunotherapies seems another very attractive way to target macrophages in tumors. However, there is a huge need for clinical studies to confirm these preclinical data in humans. Furthermore, it has to be noted that these treatments need to be localized to avoid an activation of M1 macrophages outside of the tumor and a systemic inflammatory response.

TARGETING TAMs WITH RT

In addition to target TAMs using chemo- and immunotherapies, it is also possible to influence the macrophage polarization with RT. Herein, we explore how low linear energy transfer (LET) radiotherapies (X-rays and γ -rays) can repolarize TAMs *in vitro* and *in vivo*. In a first part, the doses which have to be applied to the tumor in order to induce TAM reprogramming will be described and in a second part, the mechanisms involved in RT-induced reprogramming will be detailed.

Macrophage Radioresistance and Recruitment after RT

Macrophages are one of the most radioresistant cells in humans (7). This radioresistance is brought by a huge production of anti-oxidative molecules, such as manganese superoxide dismutase (MnSOD), a scavenger of superoxide (O₂⁻) ions. The high expression of MnSOD confers cellular resistance against damaging effects. These damaging effects are mainly produced

by radiation-induced radicals, such as ROS or reactive nitrogen species (RNS). Indeed, the expression of MnSOD was increased after irradiation in THP-1, HL60, and KG-1 myelocytic cell lines. The mechanism of radioresistance conferred to macrophages was shown to be dependent on TNF α signaling. Indeed, endogenous production of TNF α is required for MnSOD expression following ionizing radiation (IR) (88–90). This TNF α -induced MnSOD expression is mediated by PKC-dependent activation of cAMP-responsive element-binding protein-1/ATF-1-like factors and would be dependent on NF κ B activation (90–92). Currently, the radioresistance provided by endogenous MnSOD to macrophages is mimicked in healthy cells using MnSOD plasmid/liposome gene therapy to confer radioresistance to healthy cells (93–95).

Radiotherapy is used for more than 50% of cancer patients and showed tumor regression in most of the cases (33, 96). However, in addition to the intrinsic radioresistance of macrophages, IR also elicits a high recruitment of myeloid cells at the tumor site, possibly leading to tumor recurrence and tumor regrowth (59). Depletion of macrophages by liposomal clodronate before IR promoted the antitumor effects of RT and highlighted the role of recruited TAMs in tumor regrowth (90). Macrophage recruitment after IR is mediated by CCL2 and CSF1. Indeed, RT stimulated CSF1 production in prostate cancer and was shown to be responsible for TAM accumulation (97). Similarly, the inhibition of CSF1 receptor with PLX3397 prevented the myeloid cells recruitment after IR, increased survival and postponed the recurrence of glioblastoma in intracranial xenograft models (59). RT also promotes macrophage infiltration, from the peritumoral environment to the tumor site, in a CCL2 dependent way (98, 99). However, to the best of our knowledge, there is no study combining RT to CCL2 inhibition described yet.

Dose-Dependent Effects of Irradiation on Macrophage Reprogramming

Although IR slightly affects the viability of macrophages, radiotherapy modifies the macrophage phenotype. To analyze the effects of radiotherapy on macrophage reprogramming, we classified these effects according to the dose. According to the UNSCEAR (United Nations Scientific Committee on the Effects of Atomic Radiation), low dose of X-ray radiation are doses under 0.1 Gy, but clinically applied low doses are doses under 1 Gy (100). In this review, the irradiation doses were classified as followed: low doses as doses lower than 1 Gy, moderate doses (MDI) as doses ranging from 1 to 10 Gy, and high doses as doses higher than 10 Gy.

High Doses of Irradiation (HDI)

In vitro, M1 Raw264.7 macrophages were reprogrammed toward M2-like macrophages after HDI (20 Gy). Sustained M2 phenotype after irradiation resulted from NF κ B p50 activation, leading to high IL-10 production and reduced TNF α secretion. These results were confirmed *in vivo* since HDI (3 Gy \times 20 Gy over 3 days) promoted M2-like macrophages at the tumor site, in mice with Panc02 cell xenografts (101). In another study, high doses of X-rays (25 Gy in one shot or 60 Gy fractioned over 3 weeks) increased the number of M2-like TAMs in a murine model of

prostate cancer (TRAMP-C1 cell line). This was evidenced by a low inducible nitric oxide synthase (iNOS) level in macrophages and by an increase in Arg1 and COX-2 mRNA expression. Such high doses induced angiogenesis and tumor growth in this cancer model (7). An increased number of M2-like macrophages after high doses of radiation (12 Gy) was also observed in an oral cancer model. The recolonization of tumor by M2-like macrophages elicited the secretion of pro-angiogenic factors that contribute to neoangiogenesis, favoring tumor growth (102). In addition, Seifert et al. similarly described accelerated tumor growth after irradiation (12 Gy) of murine pancreatic tumor models. This was driven by an early infiltration of M2-polarized TAMs into the tumor after radiotherapy and resulted in a subsequent T-cell suppressive response. This T cell response was linked to the upregulation of PD-L1 expression in tumor. The authors showed that RT accelerated the progression of pancreatic dysplasia to invasive carcinoma but also promoted tumor growth in invasive pancreatic ductal adenocarcinoma (PDA). Indeed, a decrease in iNOS, IRF5 and H2eb1 mRNA expression and a higher expression of Arg1, CD206, and PD-L1 were observed in TAMs from pancreatic dysplasia and PDA irradiated with hypofractionated (12 Gy) or fractionated (3 Gy \times 6 Gy) doses (103). When TAMs were collected from tumor-bearing mice after RT and transferred to other tumor-bearing mice, tumor growth was accelerated to a higher extent than with unirradiated-macrophage transfer (7). Altogether, these results showed that HDI skew TAMs in a M2-like phenotype (Table 1). This is why the blockade of M-CSF, in order to prevent macrophage recruitment, in combination with HDI is attractive. Indeed, it revealed interesting results: M-CSF inhibition combined to RT promotes tumor regression. This reinforces the idea that HDI-irradiated TAMs take part to the tumor growth and tumor radioresistance. Furthermore, high levels of TAM-derived IL-10 elicited T cell anergy in irradiated tumor (103). This is why an adoptive CD4⁺ T cell transfer has not met any success, except with the use of anti-M-CSF therapy combined to RT. This observation highlights the necessity to reprogram macrophages toward a M1-like phenotype to overcome the immunosuppression and the tumor regrowth after RT.

Moderate Doses of Irradiation (MDI)

In vitro, human unpolarized monocyte-derived macrophages shifted toward M1-like macrophages after MDI (2 Gy \times 5). This is highlighted by the upregulation of pro-inflammatory markers (M1 phenotype) such as human leukocyte antigen-cell surface receptor (HLA-DR) and CD86, but also by the downregulation of anti-inflammatory markers (M2 phenotype) such as decreased mRNA expression of CD163, MRC1 (C-type mannose receptor 1, CD206) and versican (ECM proteoglycan) and reduced secretion of IL-10. In these macrophages, phagocytosis, associated with the M1-like phenotype, was increased after MDI while irradiation did not influence the ability of cocultured macrophages to promote the invasion of cancer cells and angiogenesis, features of M2-like macrophages (104). In the same line, unpolarized Raw 264.7 macrophages exhibited a higher expression of M1 markers (iNOS, TNF α , IL-12p70, IFN γ RANTES, and MCP-1) and a higher abundance of the phosphorylated p65 subunit from NF κ B after γ -irradiation (2 Gy) (105). The γ -irradiation (2 Gy) of a human

TABLE 1 | Macrophage reprogramming after HDI, MDI, and LDI.

	Reference	Dose	Radiation type	Effect on polarization	Model
HDI	(7)	4 Gy × 5 × 3 week = 60 Gy	X-ray	M2 polarization	Mouse TRAMP-C1 (prostate)
	(101)	20 Gy × 3/day	X-ray	M2 polarization	Mouse Panc02 (pancreas)
	(102)	12 Gy	X-ray	M2 polarization	Human oral cancer (OSC-19 cells)—xenograft
	(103)	12 Gy or 3 × 6 Gy	X-ray	M2 polarization	Orthotopic pancreatic ductal adenocarcinoma model pancreas
MDI	(108)	2 Gy × 5 (10 Gy)	X-ray	Mixed M1/M2 phenotype	Human monocyte-derived macrophages (hMDM) with RKO or SW1463 cells
	(105)	5–10 Gy	X-ray	Increased inducible nitric oxide synthase (iNOS)/nitric oxide (NO) production	Raw 264.7 stimulated with lipopolysaccharide (LPS)/interferon γ (IFN γ)
	(104)	2 Gy × 5	X-ray	Reduced M2 markers	hMDMs
	(107)	2–4 Gy	γ -ray	M1 polarization	Raw 264.7, THP-1, hMDM
	(33)	2 Gy	γ -ray	M1 polarization	Raw 264.7 and peritoneal macrophages from RT5 mice
	(96)	2 Gy	γ -ray	M1 polarization	Tumor-associated macrophages from RT5 mice + CD8 T cell transfer (pancreas)
	(118)	1–2 Gy	X-ray	M2 polarization	Peritoneal macrophages from BALB/c mice + LPS/Raw 264.7
	LDI	(105)	0.3–0.6–1.25 Gy	X-ray	Decreased iNOS/NO production
(109)		0.5–1 Gy	X-ray	M2 polarization	Raw 264.7
(110)		0.5–0.7 Gy	X-ray	M2 polarization	THP-1 monocytes + LPS and monosodium urate crystals
(111)		0.5–0.7 Gy	X-ray	Reduced M1 markers	THP-1 monocytes + LPS and monosodium urate crystals
(112)		0.01–0.7 Gy	X-ray	M2 polarization	Peritoneal macrophages/Raw 264.7

Red shading/font means M2 polarization, green shading/font means M1 polarization.

HDI, high doses of irradiation; MDI, moderate doses of irradiation; LDI, low doses of irradiation.

macrophage cell line (U937) also provoked a higher expression of TNF α and IFN γ compared to unirradiated macrophages (106). In another study, CD11b⁺/Gr-1 peritoneal macrophages from RT5 mice were γ -irradiated (2 Gy) *ex vivo* and showed an increased iNOS expression, related to M1 phenotype (33). Finally, PMA-differentiated macrophages (THP-1), murine macrophages (Raw 264.7) and human monocyte-derived macrophages (hMDM) revealed a pro-inflammatory profile after moderate doses of γ -rays (2 and 4 Gy). This was evidenced by increased mRNA levels for TNF α , IFN γ , IL-6, and IL-23 and higher protein levels for IL-1 β and IL-8. While most of the previous studies showed the activation of NF κ B p65 for macrophage reprogramming, this last study revealed that IR-induced M1-like phenotype was promoted by the transcriptional expression of IRF5 and was ataxia telangiectasia mutated (ATM)-dependent (107). Altogether, the results from *in vitro* experiment showed that unpolarized macrophages tend to acquire a M1 phenotype after MDI.

In addition to program unpolarized macrophages to M1 phenotype, MDI also potentiated the already acquired M1 phenotype. Indeed, MDI (2 Gy × 5) promoted a pro-inflammatory profile in M1 macrophages stimulated with LPS/IFN γ , as indicated by an increased expression of HLA-DR (104). Another *in vitro* experiment revealed that doses under 1 Gy prevented the polarization of macrophages toward M1 phenotype whereas doses of 5 and 10 Gy promoted the M1 phenotype in murine Raw 264.7 macrophages when stimulated with LPS/IFN γ . This is emphasized by an increased expression of iNOS and a higher NO production in irradiated macrophages (105). In contrast, in hMDMs stimulated with M-CSF and IL-10 (M2 macrophages), MDI did not influence the expression of pro- and anti-inflammatory markers (104). In other words, the effect of MDI on polarized macrophages showed an enhanced M1 phenotype in M1

macrophages but no change in M2 macrophages, meaning that MDI could not reprogram TAMs *in vitro*.

Cocultured experiments were performed between radiosensitive (RKO cells) or radioresistant (SW1463 cells) colorectal cancer cell lines and human unpolarized monocyte-derived macrophages. After irradiation, the mRNA expression of some pro-inflammatory (CCR7, IL1 β , CXCL8) markers was decreased whereas the mRNA expression of anti-inflammatory markers was unchanged when macrophages were cocultured with RKO cells. MDI also promoted cancer cell invasion and migration of cocultured RKO cells. However, when macrophages were cocultured with SW1463 cells, there was an upregulation of the expression of pro-inflammatory (CCR7, CD80) and anti-inflammatory markers (IL-10 and CCL18) but no change of cancer cell migration and invasion. It means that following MDI, unpolarized macrophages adopt a different phenotype according to the type of cancer cells with which they interact (108).

Interestingly, *in vivo* experiments revealed promising results. A group of researchers suggested that moderated single dose (2 Gy) of γ -ray was able to completely reprogram TAMs in tumor. Klug and Prakash analyzed M1 (iNOS protein level, NO production) and M2 (Ym-1, Fizz-1 and Arg 1 protein level) markers in CD11b⁺ peritoneal macrophages from RT5 mice after whole-body irradiation (WBI). While untreated mice exhibited elevated expression of M2 markers in peritoneal macrophages, whole-body irradiated mice revealed increased M1 marker expression and decreased M2 marker expression. However, local irradiation was not able to modify the expression of M1 and M2 markers when measured in tumor tissue lysate. As local MDI (1, 2, and 6 Gy) induced a decline of T cell infiltration, the authors used CD8⁺ T cell transfer in addition to local irradiation (2 Gy) to induce a shift in the polarization of TAMs toward a M1 phenotype

and to promote T cell infiltration in tumor. Local irradiation in combination with CD8⁺ T cell transfer induced an enhanced expression of IL-12p40 and IFN γ (M1 markers) and a reduced expression of IL-10 (M2 marker) in tumor tissue lysate. Indeed, the infiltration of tumor by T cells was exclusively dependent on the reprogrammed TAMs. When TAMs was depleted with CL treatment before local irradiation and CD8⁺ T cell transfer, there was an inhibition of T cell recruitment into irradiated tumors. The infiltration of CD8⁺ T cell into tumor is made feasible with the normalization of the vasculature. Indeed, pro-inflammatory macrophages elicited the normalization of the vasculature and allowed an efficient T cell transfer, leading to the eradication of the tumor and an increased survival in irradiated RT5 mice. However, it remains elusive how CD8⁺ T cell transfer impacts TAM reprogramming. Interestingly, local irradiation alone is not able to reprogram TAMs toward a M1-like phenotype. Then, it seems clear that CD8⁺ T cell transfer plays a role in macrophage reprogramming. The underlying mechanism needs to be further investigated (96) **Table 1** summarized the effects of MDI on macrophage polarization described here above.

Low Doses of Irradiation (LDI)

Low doses of irradiation represents a good alternative in order to bypass the toxicities observed during intensive radiotherapy (HDI) (**Table 1**). However, LDI rather favor the M2 phenotype of TAMs. Indeed, LDI decreased the iNOS level and the NO production in Raw 264.7 macrophages polarized in M1 phenotype, resulting in repolarization of M1 macrophages toward M2 phenotype (105). Other results in favor of a M2-like phenotype were described after low doses of γ -radiation in murine macrophages. Indeed, irradiation of murine macrophages (RAW 264.7) with a dose of 0.5 Gy inhibited MPK1 [mitogen-activated protein kinase (MAPK) phosphatase]. MPK1 dephosphorylates the MAPKs p38, c-Jun and ERK1/2 and the dephosphorylation of p38 was associated with a decrease in TNF α and IL1 β levels. Irradiation (0.5 Gy) of RAW264.7 macrophages stimulated with LPS also led to a decrease in p38 phosphorylation and TNF α production (109). In addition, Frey and Lodermann showed that low doses of X-rays (0.5–0.7 Gy) induced an anti-inflammatory phenotype in LPS-activated human THP-1 cells as evidenced by reduced amounts of secreted IL-1 β in the medium. This was linked to a lower p65 NF κ B nuclear translocation and a weak p38 phosphorylation (110, 111). Closely related to these results, it was also demonstrated that doses under 2 Gy favored the repolarization

of M1 macrophages into M2-like macrophages, as evidenced by an increased level of TGF β in the culture medium. This was associated to a reduced nuclear translocation of p65 NF κ B (112).

In conclusion, HDI and LDI show no effect on M2 to M1 macrophage polarization while MDI clearly evoke TAM reprogramming. However, most of studies performed in this field are *in vitro* experiments, i.e., macrophages alone with no contact with cancer cells. Further *in vivo* experiments should reinforce the results obtained by Klug et al. with MDI. Furthermore, to our knowledge, no clinical study has been designed to analyze the effect of MDI on TAM reprogramming. It is thus difficult to transpose these data to the human tumors.

Whole-Body Irradiation

In vivo studies showed that exposure to low doses of WBI in healthy mice activated the innate immune cells including macrophages (113–115) (**Table 2**). In healthy mice, a single dose of WBI resulted in a T_H1 cytokine expression profile whereas fractionated dose irradiation drove a T_H2 shift in spleen and blood of mice (116). In the same line, single low doses (0.075 and 2 Gy) of total body radiation in healthy mice polarized peritoneal macrophages toward M1 phenotype as indicated by increased IL-12 and IL-18 secretion. This polarization is strongly linked to the activation of p65 NF κ B and MyD88 a few hours after WBI (117). Only a few studies were published on WBI of tumor-bearing mice, as this treatment is irrelevant for human. However, as WBI would reprogram TAMs, studying the underlying mechanisms could be very helpful. A first study indicated that low doses (0.01–0.1 Gy) of WBI induced opposite effects on macrophages and NK cells in BALB/c radiosensitive mice and C57BL/6 radioresistant mice. Peritoneal macrophages exhibited a M1 phenotype in BALB/c mice, evidenced by an increased production of IL1 β , IL-12 and TNF α . However, M2 macrophages were observed in tumor from C57BL/6 mice after WBI, partly elucidating the radioresistance of this mouse strain (118). Klug et al. also showed that peritoneal macrophages from RT5 mice, irradiated with a systemic dose of 2 Gy, revealed a higher iNOS level and NO production whereas the expression of M2 markers was decreased in these macrophages, indicating macrophage reprogramming. Furthermore, macrophages from total body irradiated mice, when transferred to tumor-bearing mice in parallel with CD8⁺ T cell adoptive transfer, induced tumor regression, similar to local irradiation in combination with CD8⁺ T cell adoptive transfer (96). In parallel, Prakash et al. described a switch of TAM polarization toward

TABLE 2 | Macrophage reprogramming after WBI.

Reference	Dose	Radiation type	Effect	Model
(113)	0.2 and 2 Gy	γ -ray	M1 polarization	WBI (without tumor)
(117)	0.075 and 2 Gy	X-ray	M1 polarization	WBI (without tumor)
(114)	0.5–6 Gy	X-ray	M1 polarization	WBI (without tumor)
(118)	0.01–0.02–0.1 Gy \times 10/day \times 2 weeks (LDI)	X-ray	Increased cytotoxic activity and nitric oxide	Macrophages from BALB (L1)/c and C57BL/6 (Lewis lung cancer) mice (WBI)
(96)	0.5–2 Gy \times 1 (MDI)	X-ray	M1 polarization	Mouse RT5 insulinoma/Human MeWo melanoma xenograft/Human pancreas
(33)	2 Gy \times 2 (MDI)	γ -ray	M1 polarization	Rip1 Tag5 tumor-bearing mouse (WBI)

Green shading means M1 polarization.

WBI, whole-body irradiation.

M1 phenotype when RT5 insulinoma bearing mice were treated by systemic irradiation (2 Gy/week on 2 weeks). Indeed, tumor lysates presented a higher expression of M1 markers [iNOS, pSTAT3, TNF α , IL-12 (p70)] and a lower expression of M2 markers (CD206, Fizz-1, Ym-1 and Arg1). Peritumoral macrophages showed an elevated iNOS expression and NO production after WBI, modifying the tumor microenvironment with the normalization of the tumor vasculature.

To explain the discordance between local irradiation and WBI on the reprogramming of TAMs, the authors suggested that WBI allowed the mobilization of fresh reprogrammed macrophages from various lymphoid organs to infiltrate the tumor site and the surrounding microenvironment. Infiltration of tumor by reprogrammed macrophages can result in the activation of antitumor T-cell responses and can thereby act as an “endogenous vaccine” (33). On the other hand, local irradiation is not able to invite macrophages from lymphoid organs to the tumor site. It could be hypothesized that the transfer of CD8⁺ T cells would induce macrophage reprogramming in lymphoid organs. The combination of T cell transfer to irradiation would promote the normalization of the tumor vasculature, allowing tumor perfusion by CD8⁺ T cells and reprogrammed macrophages.

Molecular Pathways Responsible for the Repolarization of TAMs after RT

There are several mechanisms proposed to explain the influence of the irradiation dose on the polarization of TAMs. These mechanisms are partly different from those involved in the reprogramming mediated by chemotherapy and immunotherapy, and they include ROS, DNA damage, p50–p65 NF κ B activation, and MAPK phosphorylation. The differences between the different doses could be due to the activation of different pathways according to the dose, i.e., a switch in the NF κ B subunit balance for moderate doses while high doses would induce apoptosis.

NF κ B Balance

Nuclear factor kappa B plays an important role in the polarization of macrophages. It was shown that M2 macrophages acquired their phenotype under the effect of p50–p50 NF κ B homodimer while M1 macrophages are answerable to p50–p65 NF κ B heterodimer (24). Once p65–p50 NF κ B is translocated into the nucleus, it allows the expression of pro-inflammatory mediators [TNF α , IL-1 β , IL-6, IL-12(p40), IFN γ , CXCL10 and NOS2 (14)] whereas p50–p50 NF κ B inhibits the expression of these genes. Indeed, several experiments revealed the need for p65 translocation to activate M1 polarization (24, 119). Crittenden and his colleagues evidenced that HDI (60 Gy) produced M2 phenotype through p50–p50 dimer activation, leading to a subsequent IL-10 production (101). In the same line, low X-ray radiation dose (<1 Gy) reduced the translocation of p50–p65 NF κ B in M1 activated macrophages (112). LDI on human macrophages and LPS-activated THP-1 macrophages reduced the nuclear amount of p65 NF κ B, a phenomenon also correlated to a decreased secretion of IL-1 β (M1 cytokine) (110). Interestingly, Teresa Pinto et al. revealed an enhanced phosphorylation of RelA (p65) in macrophages following MDI (2, 6, and 10 Gy), correlated to M1 repolarization (104). Hildebrandt et al. also evidenced the role of NF κ B in TAM

reprogramming in Raw264.7 macrophages after 2 Gy irradiation (105). In an *in vivo* study, whole-body γ -irradiation (2 Gy)-induced the phosphorylation of NF κ B p65 in tumor lysate. The activation of NF κ B p65 is closely related to a reprogramming of TAMs toward M1-like macrophages (33) (**Figure 3**).

In sepsis, a sustained activation of macrophages by LPS leads to a tolerance and a reprogramming of macrophages toward M2 phenotype. This reprogramming is driven by p21, involved in the shifting of the balance between p65–p50 and p50–p50 NF κ B dimers. Indeed, p21 deficiency is linked to a reduced DNA-binding affinity of the p50–p50 homodimer and a prevalence for the p65–p50 heterodimer in macrophages stimulated with LPS (120, 121). In some cases, ubiquitination is controlled by the phosphorylation of a protein substrate (122). Indeed, some effectors of the DNA repair machinery are described to be involved in the ubiquitin proteasome pathway and regulate p21 protein level in ML-1 cells (myelocytic leukemia cell line) (123). Therefore, a high ubiquitination of p21 would lead to a lower level of p21 protein. This could be correlated with an increased p65–p50 DNA-binding affinity and could potentially favor a M1 phenotype after IR. Further investigations are, however, required to understand the possible role of p21 in regulating the NF κ B balance after IR.

ATM Kinase

Radiotherapy induces major effects on cells by creating DNA damage. DNA damage and ROS produced by γ -radiation (2–20 Gy) induced the phosphorylation of ATM in U937 human macrophage cell line (106). When DNA damage appeared, ATM plays a central role in the detection and the activation of the DNA repair machinery [for review see Ref. (124)]. The activation of ATM notably leads to the ubiquitination of NEMO (NF κ B Essential Modulator), a subunit of the IKK complex, a few hours

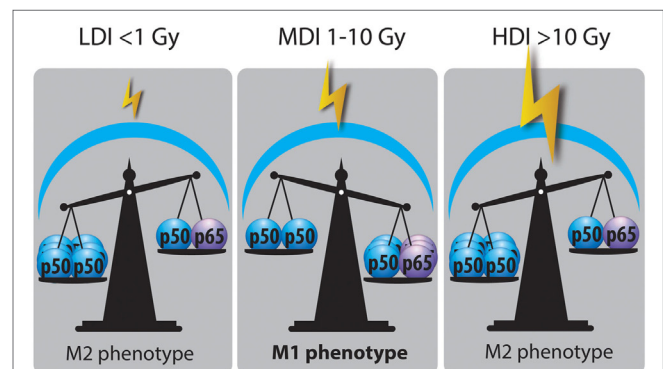


FIGURE 3 | Nuclear factor kappa B (NF κ B) balance state after LDI, MDI, and HDI. Irradiation dose showed opposite effects on NF κ B balance in macrophages: LDI or doses lower than 1 Gy did not modify the abundance of p50–p50 NF κ B in macrophage nucleus after radiation. MDI or doses from 1 to 10 Gy induced a switch of NF κ B balance from the inactive homodimer (p50–p50) to the active heterodimer (p50–p65), correlated with a reprogramming of tumor-associated macrophages (TAMs). HDI or doses higher than 10 Gy were not able to change the NF κ B balance, skewing TAMs in a M2 phenotype (HDI, high doses of irradiation; MDI, moderated doses of irradiation; LDI, low doses of irradiation).

after macrophage irradiation. The ubiquitination of NEMO is ATM-dependent as indicated by the reduced ubiquitination of NEMO when an ATM inhibitor (Ku5593) was used in combination with γ -rays (125). Ubiquitinated NEMO then drives the activation of IKK complex in the cytoplasm. This activation goes through ubiquitination of IK β and the subsequent degradation of this inhibitor by the proteasome. p65–p50 NF κ B is, therefore, released in the cytoplasm and free to move into the nucleus where the heterodimer acts as a transcriptional factor [for review see Ref. (126)].

After γ -radiation (2 Gy), the phosphorylation of NEMO was followed by the activation of NF κ B (p65). A subsequent upregulation of M1 markers (CD86, CD40, HLA-DR, TNF α , IFN γ) and a reduced secretion of IL-4 (M2 marker) followed the activation of NF κ B. These changes are partially generated by elevated ROS content in cells, as indicated by the increase or decrease in the phosphorylation of NEMO when U937 monocytic cells were irradiated and treated with L-buthionine sulphoximine or N-acetyl cysteine (NAC), respectively. Indeed, L-buthionine sulphoximine irreversibly inhibits γ -glutamylcysteine synthase, hence depleting GSH level and increasing ROS content. NAC is a potent antioxidant, known to decrease ROS level by increasing GSH levels (106). However, another team showed that γ -rays (0, 3, 6, and 10 Gy) induced NF κ B activation in Raw 264.7 macrophages, as highlighted by the degradation of the IK β protein. The activation of NF κ B was related to a higher NO production in macrophages. This phenomenon is dependent on DNA damage more than ROS level, since NAC did not affect NO production and IK β degradation in irradiated cells (127).

Moreover, modulators of DNA repair, such as Olaparib (PARP inhibitor), increased ATM activation and upregulated IRF5 transcription, resulting in macrophage activation toward a pro-inflammatory phenotype (107). In conclusion, IR is able to induce DNA damage by direct or ROS-dependent interactions with DNA. DNA damage leads to the recruitment of the ATM kinase and DNA repair machinery. Consequently, ATM contributes to the reprogramming of macrophages after irradiation.

Reactive Oxygen Species

Radiation therapy promotes the formation of radicals, such as ROS or RNS in cells. ROS quantities depend on the dose applied to the cells. In macrophages, different ROS concentrations can differently influence the cellular responses and the polarization of macrophages. Indeed, the role of ROS in the polarization of macrophages is not so clear but the most accepted idea is that ROS play key roles in M1 responses (e.g., defense against invading microbes) (128) and regulates M2 polarization (129, 130).

M1 Polarization

The polarization of macrophages toward M1 or M2 phenotype goes through several pathways, including NF κ B. There is a balance between the inhibitory heterodimer NF κ B (p50–p50) and the active heterodimer NF κ B (p50–p65) (24). When this balance leans on one side or the other, macrophages undergo M2 or M1 polarization, respectively. It is also well known that ROS generation mediated the production of pro-inflammatory cytokines (131). Indeed, this ROS production activates MAPK and NF κ B pathways

(132). It was shown that H₂O₂ enhanced p65 NF κ B DNA-binding and promoted p65 NF κ B phosphorylation. This NF κ B activation is followed by its translocation into the nucleus in macrophage and monocytic cell lines, and then induces M1 polarization (133). In parallel, it was shown that H₂O₂ altered protein–protein interaction of p50–p50 NF κ B, therefore reducing p50–p50 NF κ B activation, supporting M1 activation (134). As previously said, ROS production is observed after irradiation. This is due to the direct ionization of molecules by incident photons. Another way for γ -irradiation to induce ROS generation is the activation of NADPH oxidase (NOX). The NADPH oxidase (NOX 1 and NOX 2) catalyzes the oxidation of nicotinamide adenine dinucleotide in the NADP⁺ + H⁺ while reducing O₂ in O₂⁻, leading to the formation of H₂O₂. γ -irradiation (2 Gy) promoted ROS production in Raw264.7. This ROS production was partly inhibited by the use of NAC and by diphenylene iodonium, a NOX inhibitor. In parallel, the protein expression of NOX2 was increased after γ -irradiation in PMA-differentiated THP-1 cell and in Raw264.7 macrophages. Altogether, these results indicated that moderate doses of γ -irradiation generated NOX2-dependent ROS production. Elevated ROS content yielded ATM activation, as suggested by the absence of ATM phosphorylation in presence of NAC or siNOX2 when Raw264.7 macrophages were irradiated. This NOX2-dependent ATM activation controlled the polarization of Raw 264.7, THP-1 and hMDM in M1-like macrophages, through the regulation of IRF5 transcription. Moreover, in patients, the perturbation of ATM-dependent NOX2 signaling pathway was associated with a decreased iNOS macrophage number and poor tumor responses to radiotherapy (107).

M2 Polarization

Not only, NADPH oxidase activity drives M1 polarization, but its inhibition conversely triggers M2 polarization. A previous study showed that the inhibition of NOX promoted anti-inflammatory microglial activation (M2 phenotype) during neuroinflammation (135). Furthermore, another group revealed that the deletion of NOX1/2 reduced ROS production during macrophage differentiation. This double knockout is not critical for M1-like polarization of mouse bone marrow-derived macrophages, but prevented M2 polarization, as evidenced by the reduced expression of mCCL17, mCCL24 and RELM α . Indeed, NOX1/2 is crucial for the activation of c-Jun N-terminal kinase (JNK) and extracellular signal-regulated kinase (ERK) MAPKs during monocyte to macrophage differentiation and then affects M2 polarization (136). High concentrations of ROS play a decisive role in the differentiation and the polarization of macrophages into M2 phenotype and antioxidants treatment affected M2 but not M1 macrophage polarization. The use of butylated hydroxytoluene and other ROS scavengers repressed tumorigenesis by blocking the occurrence of TAMs (129). H₂O₂ stimulates the production of TNF α and activates the transcription factor STAT6, responsible for the expression of M2 markers such as Fizz-1 in macrophages (137, 138).

All these data showed that high concentrations of ROS trigger M2 polarization while smaller concentrations are responsible for M1 polarization. On the one hand, the production of ROS driven by HDI prevents the reprogramming of TAMs toward M1-like

macrophages. On the other hand, LDI induce small ROS production, probably not enough to reach the range of concentrations able to reprogram macrophages. However, MDI allows to reach the right range of ROS concentrations in macrophages to activate p65 NFκB, IRF5, and MAPK notably through ATM activation, hence favoring M1 phenotype.

Further investigations are needed to better understand the effect of IR on the ROS generation in TAMs and the molecular pathways involved in the polarization of macrophages *via* ROS production.

IRF5

Interferon-regulatory factor/signal transducer and activator of transcription (IRF/STAT) signaling is a central pathway in macrophage polarization. IRF can interact with two adaptors, MyD88 and TRIF, downstream of cytokine receptors or TLR. IRF5 and IRF4 are in competition to the binding of MyD88 and the subsequent activation of pro-inflammatory transcription factors, including NFκB. The activation of IRF5 through MyD88 then promotes M1 phenotype in macrophages. As a competitor for MyD88, IRF4 acts as an antagonist and can suppress M1 macrophage polarization. The balance between IRF4 and IRF5 in cells is critical for M2 and M1 polarization, respectively (24, 139).

Only one *in vitro* study established the role of IRF5 in macrophage reprogramming after irradiation. In this study, the authors showed that macrophages acquired a M1-like phenotype after exposition to γ -irradiation (2 Gy). This polarization is made feasible by the activation of NOX2 and ATM, hence driving the transcription of IRF5 (107). While these results are very interesting, the contribution of IRF/STAT pathway to macrophage polarization mediated by irradiation needs to be further investigated *in vivo* and in patients. Furthermore, IRF5 modulation is also known to drive M1 polarization through the activation of NFκB. The link between IRF and NFκB in macrophage reprogramming after radiotherapy should also be considered.

iNOS Level and NO Production

Nitric oxide mediator is a double-edge sword key regulator in tumor progression. On the one hand, NO has antitumoral effects as it promotes DNA damage, elicits cell death, and enhances anticancer therapeutic efficacy. Indeed, patients with lower iNOS expression levels, hence low NO production, showed recurrence of tumors and metastasis after RT. On the other hand, NO mediator is also known for its pro-tumoral effects since it induces antiapoptotic effects and promotes cell cycle, cancer progression, metastasis, angiogenesis, and chemoprotection. High level of iNOS is associated with poor prognosis and tumor-promoting effects (140). It thus seems that critical NO concentrations and iNOS levels finely modulate the fate of the tumor.

High doses of irradiation induced NO production and high iNOS expression level in macrophages. For instance, high doses of γ -irradiation (20 Gy) led to iNOS activation and NO production, favoring tumor growth and recurrence after RT (141). Another *in vivo* study using high doses (25 Gy or 3 Gy \times 20 Gy) also generated high level of iNOS in murine macrophages after IR, contributing to the tumor growth (7). NO production contributes to DNA repair by inducing p53, poly(ADP ribose) polymerase

and DNA-dependent protein kinase activation, leading to tissue regeneration and tumor regrowth. In addition, NO mediator is related to tumor angiogenesis and promotes reoxygenation. Therefore, targeting iNOS in murine macrophages enhanced postradiotherapeutic efficacy after HDI (141).

On the other hand, WBI (2 Gy \times 2 Gy) on RT5 mice induced an increased NO production and iNOS level, enough to induce tumor regression. In addition, *ex vivo* radiation with similar doses on peritoneal macrophages from RT5 mice also stimulated an increased in iNOS level and NO production (33). Indeed, NO can be considered as a free radical and as a source of reactive oxygen and nitrogen species, inducing DNA damage (142). Another team hypothesized that iNOS and NO production induced by MDI (2 Gy) remodeled vasculature allowing cytotoxic lymphocyte recruitment, and subsequent tumor eradication (15). NO leads to higher tumor oxygenation by promoting vasodilatation and hence radiosensitization (142). It can diffuse toward bystander cancer cells leading to their radiosensitization (143).

High doses of irradiation would induce high level of NO in macrophages, leading to cancer progression. Contrarily, MDI drive a slight increase in NO production in macrophages, enough to participate to the reprogramming of TAMs and favoring tumor regression.

MAPK (p38–ERK–JNK)

Mitogen-activated protein kinases are serine threonine kinases activated by diverse stimuli including cytokines and ROS. MAPKs regulate diverse pathways such as cell proliferation, apoptosis, and differentiation. MAPKs include extracellular signal-regulated kinase 1 and 2 (ERK 1/2), JNK and p38. They mediate immune cell functional responses to diverse signals and play a role in macrophage polarization. MAPK kinases (MAPKKs) activate MAPKs through dual phosphorylation on tyrosine and threonine residues (Thr–X–Tyr motif) located in the activation loop of the kinase domain (144). The phosphorylation of MAPK activates pro-inflammatory gene transcription and is correlated with M1-like macrophages. At the opposite, MAPKs are inactivated through dephosphorylation of threonine and/or tyrosine residues within the activation loop by MAPK phosphatase (MKP-1) (145). MKP-1-deficient macrophages exhibit skewed activation profiles: macrophages showed an enhanced pro-inflammatory phenotype in response to IFN γ and TNF α while this deficiency severely suppressed “M2-like” phenotype after IL-4 stimulation (146). Compared to wild-type mice, MKP-1-deficient mice produced greater amounts of TNF α , IL-1 β , CCL2, granulocyte/macrophage colony-stimulating factor (GM-CSF), IL-6, IL-10, and IL-12p70 (147, 148).

Mitogen-activated protein kinases are also regulated by ROS and DNA damage (132). Within a few minutes, low doses (0.5 and 1 Gy) of γ -radiation induced a dephosphorylation of both ERK 1/2 and p38 MAPKs. This dephosphorylation depended on MKP-1 and induced the suppression of TNF α production in Raw 264.7 cells. These results correlated with the inability of LDI to reprogram M2 macrophages (109). Another study showed enhanced phosphorylation of ERK 1/2 and p38 following moderated doses (2 Gy) of γ -radiation in resident peritoneal exudate cells. This activation was related to a decreased

dephosphorylation exerted by MKP-1 and was correlated with $\text{TNF}\alpha$ production. The explanation relies on the oxidation of the catalytic cysteine of MKP that inactivates the phosphatase, then triggering the activation of MAPK cascade including JNK and p38. These results correlated with the reprogramming of macrophages induced by MDI (149). Conflicting results revealed the inhibition of p38 phosphorylation in macrophages after WBI of tumor-bearing mice. The inhibition of p38 phosphorylation in macrophages allowed the reprogramming of TAMs toward M1 phenotype in tumor. This inhibition could be linked to the upregulation of MKP-1 (33).

Nuclear Erythroid Derived 2-Related Factor (Nrf2)

The Nrf2 is involved in the management of intracellular oxidative stress. It is also known to repress inflammation by inhibiting pro-inflammatory cytokine expression. Indeed Nrf2 is able to bind the close proximity of IL-6, IL-1 β , and IL-1 α genes in LPS-stimulated macrophages. This binding hence inhibits the recruitment of RNA polymerase II to the transcription start site without affecting the p65 NF κ B recruitment. The Nrf2 binding to these genes is independent on ROS level since the use of antioxidant (NAC)

did not affect the inhibition of pro-inflammatory gene transcription mediated by Nrf2 (150). The transcription factor Nrf2 was translocated into the nucleus even after 0.1 Gy γ -radiation, leading to the upregulation of antioxidant proteins such as heme oxygenase-1 in Raw 264.7. Nrf2 translocation increased with the dose (0.1–2.5 Gy). The regulation of Nrf2 is mediated by ERK 1/2 pathway after the irradiation of macrophages (121). It is also known that in oxidative stress conditions, p21 interacts with Nrf2 domain of Nrf2, upregulating Nrf2-mediated antioxidant response (151). This could link the Nrf2 pathway to NF κ B in the reprogramming of TAMs after IR: the absence of p21 prevents the anti-inflammatory response mediated by Nrf2 and favors the p50–p65 NF κ B DNA-binding. The fate of irradiated macrophages would then depend on the connection between ROS level/DNA damage and the downstream-activated pathways.

Overall View Regarding TAM Reprogramming upon Radiotherapy

Local radiotherapy can finely tune the balance between immunosuppression and immune antitumor properties. The shift toward

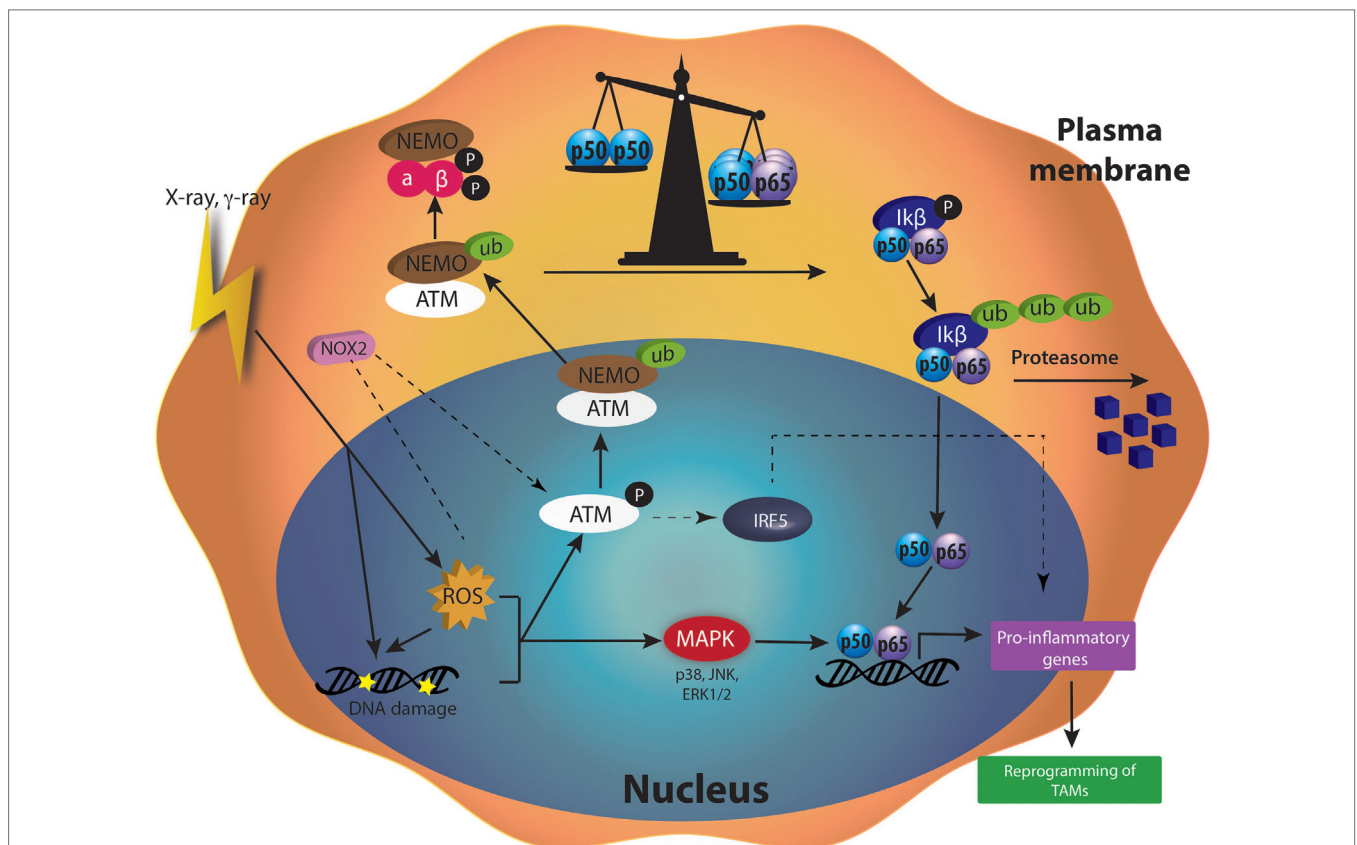


FIGURE 4 | Molecular mechanisms activated after moderated doses of irradiation in tumor-associated macrophages (TAMs). Ionizing radiation (X-rays or γ -rays) induces DNA damage and elevated reactive oxygen species (ROS) content in cells. DNA repair machinery [such as ataxia telangiectasia mutated (ATM)] is activated by DNA damage and initiates the ubiquitination of NF κ B essential modulator (NEMO), a subunit of the IKK complex. Therefore, ubiquitinated NEMO can drive the activation of IKK complex in the cytoplasm. The degradation of I κ B protein by the proteasome allows the release of p50–p65 nuclear factor kappa B (NF κ B) in the cytoplasm. p50–p65 NF κ B is then translocated into the nucleus and induces the transcription of pro-inflammatory genes, leading to the reprogramming of TAMs. ROS are also able to stimulate mitogen-activated protein kinase (MAPKs). Once phosphorylated, MAPKs also participate to the activation of NF κ B and hence, to the transcription of pro-inflammatory genes.

one or the other immune state depends on the dose per fraction, the total dose and the cancer type. Whereas local IR finds difficulties to completely reprogram TAMs, WBI can be helpful in the understanding of TAMs reprogramming since it activates the immune system outside the tumor site, leading to the complete eradication of the tumor. It, thus, seems that local IR needs something more than its sole effects on macrophages to induce a total reprogramming of TAMs and to restore a full immune antitumor response.

The mechanisms underlying macrophage reprogramming after radiation therapy are summarized in **Figure 4**. This figure emphasizes the central role of NF κ B in macrophage reprogramming after MDI. Although most of these studies described in this review are promising, there is still a huge need for further investigations and *in vivo* confirmations. Indeed, cancer cell irradiation generates damage-associated molecule patterns (DAMPs), such as high mobility group box 1 (HMGB1). DAMPs bind on their corresponding pattern-recognition receptor, such as TLR4 for HMGB1 in macrophages, triggering a pro-inflammatory phenotype (152). The irradiation then promotes antitumoral responses. However, it is possible that the contact between macrophages and cancer cells could offer the response of macrophages to radiotherapy. Indeed, cancer cells secrete different cytokines and chemokines that could influence the faith of macrophage polarization. More precisely, the type of cancer cells in the tumor can modulate macrophage reprogramming after irradiation, as evidenced by the opposite results acquired with different cancer cell lines in cocultured with macrophages (108).

In parallel, IR can also alter other types of cells in tumor, such as lymphocytes and dendritic cells, as well as the tumor vascularization (152). For example, ablative radiation therapy promotes CD8⁺ T cell accumulation into the tumor (153, 154). Indeed, single moderate doses (10 Gy) of irradiation mobilized CD8⁺ T cell through dendritic cells activation hence reducing or eradicating tumor. Therefore, MDI fulfills mobilization of effectors from the immune system and is able to reprogram TAMs in an antitumor phenotype. However, the issue about the fractionation should also be addressed, as in patients the treatments are usually given in several doses. It was shown that hyperfractionation (5 Gy \times 4 Gy) could completely abrogate the effect on CD8⁺ T cell mobilization observed with single high dose (20 Gy) (154). The effects of fractionation on macrophage reprogramming remain elusive and need to be further investigated.

CONCLUSION AND FUTURE DIRECTIONS

Chemo- and immunotherapies, as well as moderate dose of irradiation are able to reprogram TAMs into M1-like macrophages. TAM reprogramming is induced through various mechanisms.

REFERENCES

1. Mantovani A. Cancer: inflaming metastasis. *Nature* (2009) 457(7225):36–7. doi:10.1038/457036b
2. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med* (2013) 19(11):1423–37. doi:10.1038/nm.3394

These pathways mainly include NF κ B, NO production, and STAT1 that can act together. MDI-induced reprogramming is made possible through DNA damage-dependent ATM activation, the production of a specific range of ROS and NO amounts, a shift of the NF κ B balance to the active p50–p65 heterodimer, the transcription of IRF5 and the activation of the MAPK pathway. Amongst them, NF κ B pathway seems to be a central target for macrophage reprogramming.

Local IR is able to kill cancer cells and to activate the immune system by the release of cancer cell antigens and of immune-associated factors by stromal cells and vascular endothelial cells, but it also depletes T cells and antigen-presenting cells (155). Combined radiotherapy and immunotherapy have shown promising results in reprogramming macrophages and may thus be useful for tumor elimination. However, the choice of T cell adoptive transfer can determine the fate of the tumor and macrophage reprogramming, as CD8⁺ T cell adoptive transfer is more effective than CD4⁺ T cell adoptive transfer (96, 103). Indeed, the use of a anti CD4⁺ T cell therapy combined to γ -irradiation (5 Gy) in a mammary mouse model improved the efficacy of radiotherapy (156). The use of MDI combined to adoptive CD8⁺ T cell transfer or stimulating CD8⁺ T cell therapy (such as anti PD-L1/programmed cell death 1) may give promising results as effective anticancer therapies. The transfer of irradiated macrophages was also the subject of numerous studies in murine models. However, human clinical studies using transferred macrophages have also been performed but showed no or few improvements regarding cancer regression (157).

Another avenue to explore would be the effect of high LET particles, for example protons, since protontherapy is more and more developed in clinic. As the mechanism of action is different from that of conventional radiotherapy (X-rays), the effects on macrophage reprogramming could be potentiated. Indeed, charged particles can drive huge NF κ B activation compared to X-rays or γ -rays, as shown in HEK/293 cells (158).

Targeting the immune system in cancer diseases demonstrated successful improvement on cancer elimination and has become a highly promising therapy. Therefore, MDI combined to chemo- or immunotherapies targeting macrophage reprogramming could also synergize their effects on tumor regression.

AUTHOR CONTRIBUTIONS

GG wrote the review and designed the figures and the tables. SL and CM supervised the whole work, contributed to writing, and critically revised the paper.

FUNDING

GG is recipient of a Télévie grant (FNRS, Belgium).

3. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* (2011) 144(5):646–74. doi:10.1016/j.cell.2011.02.013
4. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoeediting: integrating immunity's roles in cancer suppression and promotion. *Science* (2011) 331(6024):1565–70. doi:10.1126/science.1203486
5. Mittal D, Gubin MM, Schreiber RD, Smyth MJ. New insights into cancer immunoeediting and its three component phases – elimination,

- equilibrium and escape. *Curr Opin Immunol* (2014) 27:16–25. doi:10.1016/j.coi.2014.01.004
6. Gordon S, Martinez FO. Alternative activation of macrophages: mechanism and functions. *Immunity* (2010) 32(5):593–604. doi:10.1016/j.immuni.2010.05.007
 7. Tsai CS, Chen FH, Wang CC, Huang HL, Jung SM, Wu CJ, et al. Macrophages from irradiated tumors express higher levels of iNOS, arginase-I and COX-2, and promote tumor growth. *Int J Radiat Oncol Biol Phys* (2007) 68(2):499–507. doi:10.1016/j.ijrobp.2007.01.041
 8. van Ravenswaay Claassen HH, Kluijn PM, Fleuren GJ. Tumor infiltrating cells in human cancer. On the possible role of CD16+ macrophages in antitumor cytotoxicity. *Lab Invest* (1992) 67(2):166–74.
 9. Allavena P, Sica A, Garlanda C, Mantovani A. The Yin-Yang of tumor-associated macrophages in neoplastic progression and immune surveillance. *Immunol Rev* (2008) 222:155–61. doi:10.1111/j.1600-065X.2008.00607.x
 10. Noy R, Pollard JW. Tumor-associated macrophages: from mechanisms to therapy. *Immunity* (2014) 41(1):49–61. doi:10.1016/j.immuni.2014.06.010
 11. Lewis CE, Pollard JW. Distinct role of macrophages in different tumor microenvironments. *Cancer Res* (2006) 66(2):605–12. doi:10.1158/0008-5472.CAN-05-4005
 12. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. *Nat Rev Immunol* (2008) 8(12):958–69. doi:10.1038/nri2448
 13. Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer* (2004) 4(1):71–8. doi:10.1038/nrc1256
 14. Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nat Immunol* (2010) 11(10):889–96. doi:10.1038/ni.1937
 15. De Palma M, Coukos G, Hanahan D. A new twist on radiation oncology: low-dose irradiation elicits immunostimulatory macrophages that unlock barriers to tumor immunotherapy. *Cancer Cell* (2013) 24(5):559–61. doi:10.1016/j.ccr.2013.10.019
 16. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* (2010) 141(1):39–51. doi:10.1016/j.cell.2010.03.014
 17. Gordon S, Taylor PR. Monocyte and macrophage heterogeneity. *Nat Rev Immunol* (2005) 5(12):953–64. doi:10.1038/nri1733
 18. Mantovani A, Germano G, Marchesi F, Locatelli M, Biswas SK. Cancer-promoting tumor-associated macrophages: new vistas and open questions. *Eur J Immunol* (2011) 41(9):2522–5. doi:10.1002/eji.201141894
 19. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* (2004) 25(12):677–86. doi:10.1016/j.it.2004.09.015
 20. Qian BZ, Pollard JW. New tricks for metastasis-associated macrophages. *Breast Cancer Res* (2012) 14(4):316. doi:10.1186/bcr3143
 21. Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* (2012) 122(3):787–95. doi:10.1172/JCI59643
 22. Takeda N, O’Dea EL, Doedens A, Kim JW, Weidemann A, Stockmann C, et al. Differential activation and antagonistic function of HIF- α isoforms in macrophages are essential for NO homeostasis. *Genes Dev* (2010) 24(5):491–501. doi:10.1101/gad.1881410
 23. Ying H, Kang Y, Zhang H, Zhao D, Xia J, Lu Z, et al. MiR-127 modulates macrophage polarization and promotes lung inflammation and injury by activating the JNK pathway. *J Immunol* (2015) 194(3):1239–51. doi:10.4049/jimmunol.1402088
 24. Wang N, Liang H, Zen K. Molecular mechanisms that influence the macrophage m1-m2 polarization balance. *Front Immunol* (2014) 5:614. doi:10.3389/fimmu.2014.00614
 25. Huang C, Liu XJ, QunZhou, Xie J, Ma TT, Meng XM, et al. MiR-146a modulates macrophage polarization by inhibiting Notch1 pathway in RAW264.7 macrophages. *Int Immunopharmacol* (2016) 32:46–54. doi:10.1016/j.intimp.2016.01.009
 26. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdts S, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity* (2014) 41(1):14–20. doi:10.1016/j.immuni.2014.06.008
 27. Mantovani A, Allavena P. The interaction of anticancer therapies with tumor-associated macrophages. *J Exp Med* (2015) 212(4):435–45. doi:10.1084/jem.20150295
 28. Komohara Y, Fujiwara Y, Ohnishi K, Takeya M. Tumor-associated macrophages: potential therapeutic targets for anti-cancer therapy. *Adv Drug Deliv Rev* (2016) 99(Pt B):180–5. doi:10.1016/j.addr.2015.11.009
 29. Lehenkari PP, Kellinsalmi M, Napankangas JP, Ylitalo KV, Monkkonen J, Rogers MJ, et al. Further insight into mechanism of action of clodronate: inhibition of mitochondrial ADP/ATP translocase by a nonhydrolyzable, adenine-containing metabolite. *Mol Pharmacol* (2002) 61(5):1255–62. doi:10.1124/mol.61.5.1255
 30. Galletti G, Scielzo C, Barbaglio F, Rodriguez TV, Riba M, Lazarevic D, et al. Targeting macrophages sensitizes chronic lymphocytic leukemia to apoptosis and inhibits disease progression. *Cell Rep* (2016) 14(7):1748–60. doi:10.1016/j.celrep.2016.01.042
 31. Zeisberger SM, Odermatt B, Marty C, Zehnder-Fjallman AH, Ballmer-Hofer K, Schwendener RA. Clodronate-liposome-mediated depletion of tumour-associated macrophages: a new and highly effective antiangiogenic therapy approach. *Br J Cancer* (2006) 95(3):272–81. doi:10.1038/sj.bjc.6603240
 32. Fritz JM, Tennis MA, Orlicky DJ, Lin H, Ju C, Redente EF, et al. Depletion of tumor-associated macrophages slows the growth of chemically induced mouse lung adenocarcinomas. *Front Immunol* (2014) 5:587. doi:10.3389/fimmu.2014.00587
 33. Prakash H, Klug F, Nadella V, Mazumdar V, Schmitz-Winnenthal H, Umansky L. Low doses of gamma irradiation potentially modifies immunosuppressive tumor microenvironment by retuning tumor-associated macrophages: lesson from insulinoma. *Carcinogenesis* (2016) 37(3):301–13. doi:10.1093/carcin/bgw007
 34. Zhang W, Zhu XD, Sun HC, Xiong YQ, Zhuang PY, Xu HX, et al. Depletion of tumor-associated macrophages enhances the effect of sorafenib in metastatic liver cancer models by antimetastatic and antiangiogenic effects. *Clin Cancer Res* (2010) 16(13):3420–30. doi:10.1158/1078-0432.CCR-09-2904
 35. Melani C, Sangaletti S, Barazzetta FM, Werb Z, Colombo MP. Amino-biphosphonate-mediated MMP-9 inhibition breaks the tumor-bone marrow axis responsible for myeloid-derived suppressor cell expansion and macrophage infiltration in tumor stroma. *Cancer Res* (2007) 67(23):11438–46. doi:10.1158/0008-5472.CAN-07-1882
 36. Liang W, Kujawski M, Wu J, Lu J, Herrmann A, Loera S, et al. Antitumor activity of targeting SRC kinases in endothelial and myeloid cell compartments of the tumor microenvironment. *Clin Cancer Res* (2010) 16(3):924–35. doi:10.1158/1078-0432.CCR-09-1486
 37. Germano G, Frapolli R, Belgiovine C, Anselmo A, Pesce S, Liguori M, et al. Role of macrophage targeting in the antitumor activity of trabectedin. *Cancer Cell* (2013) 23(2):249–62. doi:10.1016/j.ccr.2013.01.008
 38. Mirandola P, Ponti C, Gobbi G, Sponzilli I, Vaccarezza M, Cocco L, et al. Activated human NK and CD8+ T cells express both TNF-related apoptosis-inducing ligand (TRAIL) and TRAIL receptors but are resistant to TRAIL-mediated cytotoxicity. *Blood* (2004) 104(8):2418–24. doi:10.1182/blood-2004-04-1294
 39. Bak SP, Walters JJ, Takeya M, Conejo-Garcia JR, Berwin BL. Scavenger receptor-A-targeted leukocyte depletion inhibits peritoneal ovarian tumor progression. *Cancer Res* (2007) 67(10):4783–9. doi:10.1158/0008-5472.CAN-06-4410
 40. Nagai T, Tanaka M, Tsuneyoshi Y, Xu B, Michie SA, Hasui K, et al. Targeting tumor-associated macrophages in an experimental glioma model with a recombinant immunotoxin to folate receptor beta. *Cancer Immunol Immunother* (2009) 58(10):1577–86. doi:10.1007/s00262-009-0667-x
 41. Galmbacher K, Heisig M, Hotz C, Wischhusen J, Galmiche A, Bergmann B, et al. Shigella mediated depletion of macrophages in a murine breast cancer model is associated with tumor regression. *PLoS One* (2010) 5(3):e9572. doi:10.1371/journal.pone.0009572
 42. Shen L, Li H, Shi Y, Wang D, Gong J, Xun J, et al. M2 tumour-associated macrophages contribute to tumour progression via legumain remodelling the extracellular matrix in diffuse large B cell lymphoma. *Sci Rep* (2016) 6:30347. doi:10.1038/srep30347
 43. Smahel M, Duskova M, Polakova I, Musil J. Enhancement of DNA vaccine potency against legumain. *J Immunother* (2014) 37(5):293–303. doi:10.1097/CJI.0000000000000040
 44. Roca H, Varsos ZS, Sud S, Craig MJ, Ying C, Pienta KJ. CCL2 and interleukin-6 promote survival of human CD11b+ peripheral blood mononuclear cells and induce M2-type macrophage polarization. *J Biol Chem* (2009) 284(49):34342–54. doi:10.1074/jbc.M109.042671
 45. Soria G, Ben-Baruch A. The inflammatory chemokines CCL2 and CCL5 in breast cancer. *Cancer Lett* (2008) 267(2):271–85. doi:10.1016/j.canlet.2008.03.018

46. Zhang J, Patel L, Pienta KJ. CC chemokine ligand 2 (CCL2) promotes prostate cancer tumorigenesis and metastasis. *Cytokine Growth Factor Rev* (2010) 21(1):41–8. doi:10.1016/j.cytogfr.2009.11.009
47. Kitamura T, Qian BZ, Soong D, Cassetta L, Noy R, Sugano G, et al. CCL2-induced chemokine cascade promotes breast cancer metastasis by enhancing retention of metastasis-associated macrophages. *J Exp Med* (2015) 212(7):1043–59. doi:10.1084/jem.20141836
48. Pienta KJ, Machiels JP, Schrijvers D, Alekseev B, Shkolnik M, Crabb SJ, et al. Phase 2 study of carlumab (CNTO 888), a human monoclonal antibody against CC-chemokine ligand 2 (CCL2), in metastatic castration-resistant prostate cancer. *Invest New Drugs* (2013) 31(3):760–8. doi:10.1007/s10637-012-9869-8
49. Brana I, Calles A, LoRusso PM, Yee LK, Puchalski TA, Seetharam S, et al. Carlumab, an anti-C-C chemokine ligand 2 monoclonal antibody, in combination with four chemotherapy regimens for the treatment of patients with solid tumors: an open-label, multicenter phase 1b study. *Target Oncol* (2015) 10(1):111–23. doi:10.1007/s11523-014-0320-2
50. Zollo M, Di Dato V, Spano D, De Martino D, Liguori L, Marino N, et al. Targeting monocyte chemotactic protein-1 synthesis with bindarit induces tumor regression in prostate and breast cancer animal models. *Clin Exp Metastasis* (2012) 29(6):585–601. doi:10.1007/s10585-012-9473-5
51. Gazzaniga S, Bravo AI, Guglielmotti A, van Rooijen N, Maschi F, Vecchi A, et al. Targeting tumor-associated macrophages and inhibition of MCP-1 reduce angiogenesis and tumor growth in a human melanoma xenograft. *J Invest Dermatol* (2007) 127(8):2031–41. doi:10.1038/sj.jid.5700827
52. D'Incalci M, Badri N, Galmarini CM, Allavena P. Trabectedin, a drug acting on both cancer cells and the tumour microenvironment. *Br J Cancer* (2014) 111(4):646–50. doi:10.1038/bjc.2014.149
53. D'Incalci M, Zambelli A. Trabectedin for the treatment of breast cancer. *Expert Opin Investig Drugs* (2016) 25(1):105–15. doi:10.1517/13543784.2016.1124086
54. Larsen AK, Galmarini CM, D'Incalci M. Unique features of trabectedin mechanism of action. *Cancer Chemother Pharmacol* (2016) 77(4):663–71. doi:10.1007/s00280-015-2918-1
55. Pyonteck SM, Gadea BB, Wang HW, Gocheva V, Hunter KE, Tang LH, et al. Deficiency of the macrophage growth factor CSF-1 disrupts pancreatic neuroendocrine tumor development. *Oncogene* (2012) 31(11):1459–67. doi:10.1038/onc.2011.337
56. Strachan DC, Ruffell B, Oei Y, Bissell MJ, Coussens LM, Pryer N, et al. CSF1R inhibition delays cervical and mammary tumor growth in murine models by attenuating the turnover of tumor-associated macrophages and enhancing infiltration by CD8+ T cells. *Oncimmunology* (2013) 2(12):e26968. doi:10.4161/onci.26968
57. Pradel LP, Ooi CH, Romagnoli S, Cannarile MA, Sade H, Ruttinger D, et al. Macrophage susceptibility to emactuzumab (RG7155) treatment. *Mol Cancer Ther* (2016) 15(12):3077–86. doi:10.1158/1535-7163.MCT-16-0157
58. Zhu Y, Knolhoff BL, Meyer MA, Nywening TM, West BL, Luo J, et al. CSF1/CSF1R blockade reprograms tumor-infiltrating macrophages and improves response to T-cell checkpoint immunotherapy in pancreatic cancer models. *Cancer Res* (2014) 74(18):5057–69. doi:10.1158/0008-5472.CAN-13-3723
59. Stafford JH, Hirai T, Deng L, Chernikova SB, Urata K, West BL, et al. Colony stimulating factor 1 receptor inhibition delays recurrence of glioblastoma after radiation by altering myeloid cell recruitment and polarization. *Neuro Oncol* (2016) 18(6):797–806. doi:10.1093/neuonc/nov272
60. Butowski N, Colman H, De Groot JF, Omuro AM, Nayak L, Wen PY, et al. Orally administered colony stimulating factor 1 receptor inhibitor PLX3397 in recurrent glioblastoma: an Ivy Foundation Early Phase Clinical Trials Consortium phase II study. *Neuro Oncol* (2016) 18(4):557–64. doi:10.1093/neuonc/nov245
61. Edwards JP, Emens LA. The multikinase inhibitor sorafenib reverses the suppression of IL-12 and enhancement of IL-10 by PGE(2) in murine macrophages. *Int Immunopharmacol* (2010) 10(10):1220–8. doi:10.1016/j.intimp.2010.07.002
62. Hussain SF, Kong LY, Jordan J, Conrad C, Madden T, Fokt I, et al. A novel small molecule inhibitor of signal transducers and activators of transcription 3 reverses immune tolerance in malignant glioma patients. *Cancer Res* (2007) 67(20):9630–6. doi:10.1158/0008-5472.CAN-07-1243
63. Sun L, Chen B, Jiang R, Li J, Wang B. Resveratrol inhibits lung cancer growth by suppressing M2-like polarization of tumor associated macrophages. *Cell Immunol* (2017) 311:86–93. doi:10.1016/j.cellimm.2016.11.002
64. Dong R, Gong Y, Meng W, Yuan M, Zhu H, Ying M, et al. The involvement of M2 macrophage polarization inhibition in fenretinide-mediated chemopreventive effects on colon cancer. *Cancer Lett* (2017) 388:43–53. doi:10.1016/j.canlet.2016.11.029
65. Sakurai M, Nishio M, Yamamoto K, Okuda T, Kawano K, Ohnuki T. TMC-264, a novel inhibitor of STAT6 activation produced by Phoma sp. TC 1674. *J Antibiot (Tokyo)* (2003) 56(6):513–9. doi:10.7164/antibiotics.56.513
66. Miljkovic D, Samardzic T, Mostarica Stojkovic M, Stosic-Grujicic S, Popadic D, Trajkovic V. Leflunomide inhibits activation of inducible nitric oxide synthase in rat astrocytes. *Brain Res* (2001) 889(1–2):331–8. doi:10.1016/S0006-8993(00)03181-4
67. Chiba Y, Todoroki M, Nishida Y, Tanabe M, Misawa M. A novel STAT6 inhibitor AS1517499 ameliorates antigen-induced bronchial hypercontractility in mice. *Am J Respir Cell Mol Biol* (2009) 41(5):516–24. doi:10.1165/rcmb.2008-0163OC
68. De Meyer I, Martinet W, Schrijvers DM, Timmermans JP, Bult H, De Meyer GR. Toll-like receptor 7 stimulation by imiquimod induces macrophage autophagy and inflammation in atherosclerotic plaques. *Basic Res Cardiol* (2012) 107(3):269. doi:10.1007/s00395-012-0269-1
69. Dewan MZ, Vanpouille-Box C, Kawashima N, DiNapoli S, Babb JS, Formenti SC, et al. Synergy of topical toll-like receptor 7 agonist with radiation and low-dose cyclophosphamide in a mouse model of cutaneous breast cancer. *Clin Cancer Res* (2012) 18(24):6668–78. doi:10.1158/1078-0432.CCR-12-0984
70. Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update on toll-like receptors. *Nat Immunol* (2010) 11(5):373–84. doi:10.1038/ni.1863
71. Kumagai Y, Takeuchi O, Akira S. TLR9 as a key receptor for the recognition of DNA. *Adv Drug Deliv Rev* (2008) 60(7):795–804. doi:10.1016/j.addr.2007.12.004
72. Hemmi H, Takeuchi O, Kawai T, Kaisho T, Sato S, Sanjo H, et al. A toll-like receptor recognizes bacterial DNA. *Nature* (2000) 408(6813):740–5. doi:10.1038/35047123
73. Chen N, Wei M, Sun Y, Li F, Pei H, Li X, et al. Self-assembly of polyadenine-tailed CpG oligonucleotide-gold nanoparticle nanoconjugates with immunostimulatory activity. *Small* (2014) 10(2):368–75. doi:10.1002/smll.201300903
74. Buhtoiarov IN, Sondel PM, Wigginton JM, Buhtoiarova TN, Yanke EM, Mahvi DA, et al. Anti-tumour synergy of cytotoxic chemotherapy and anti-CD40 plus CpG-ODN immunotherapy through repolarization of tumour-associated macrophages. *Immunology* (2011) 132(2):226–39. doi:10.1111/j.1365-2567.2010.03357.x
75. Shi Y, Felder MA, Sondel PM, Rakhmilevich AL. Synergy of anti-CD40, CpG and MPL in activation of mouse macrophages. *Mol Immunol* (2015) 66(2):208–15. doi:10.1016/j.molimm.2015.03.008
76. Jordan M, Waxman DJ. CpG-1826 immunotherapy potentiates chemotherapeutic and anti-tumor immune responses to metronomic cyclophosphamide in a preclinical glioma model. *Cancer Lett* (2016) 373(1):88–96. doi:10.1016/j.canlet.2015.11.029
77. Zippelius A, Schreiner J, Herzig P, Muller P. Induced PD-L1 expression mediates acquired resistance to agonistic anti-CD40 treatment. *Cancer Immunol Res* (2015) 3(3):236–44. doi:10.1158/2326-6066.CIR-14-0226
78. Lum HD, Buhtoiarov IN, Schmidt BE, Berke G, Paulnock DM, Sondel PM, et al. Tumoristic effects of anti-CD40 mAb-activated macrophages involve nitric oxide and tumour necrosis factor-alpha. *Immunology* (2006) 118(2):261–70. doi:10.1111/j.1365-2567.2006.02366.x
79. Johnson EE, Buhtoiarov IN, Baldeshwiler MJ, Felder MA, Van Rooijen N, Sondel PM, et al. Enhanced T-cell-independent antitumor effect of cyclophosphamide combined with anti-CD40 mAb and CpG in mice. *J Immunother* (2011) 34(1):76–84. doi:10.1097/CJI.0b013e318200b28a
80. Weiss JM, Ridnour LA, Back T, Hussain SP, He P, Maciag AE, et al. Macrophage-dependent nitric oxide expression regulates tumor cell detachment and metastasis after IL-2/anti-CD40 immunotherapy. *J Exp Med* (2010) 207(11):2455–67. doi:10.1084/jem.20100670
81. Vonderheide RH, Flaherty KT, Khalil M, Stumacher MS, Bajor DL, Hutnick NA, et al. Clinical activity and immune modulation in cancer

- patients treated with CP-870,893, a novel CD40 agonist monoclonal antibody. *J Clin Oncol* (2007) 25(7):876–83. doi:10.1200/JCO.2006.08.3311
82. Georgoudaki AM, Prokopec KE, Boura VF, Hellqvist E, Sohn S, Ostling J, et al. Reprogramming tumor-associated macrophages by antibody targeting inhibits cancer progression and metastasis. *Cell Rep* (2016) 15(9):2000–11. doi:10.1016/j.celrep.2016.04.084
 83. Dunn GP, Koebel CM, Schreiber RD. Interferons, immunity and cancer immunoediting. *Nat Rev Immunol* (2006) 6(11):836–48. doi:10.1038/nri1961
 84. Jassar AS, Suzuki E, Kapoor V, Sun J, Silverberg MB, Cheung L, et al. Activation of tumor-associated macrophages by the vascular disrupting agent 5,6-dimethylxanthone-4-acetic acid induces an effective CD8+ T-cell-mediated antitumor immune response in murine models of lung cancer and mesothelioma. *Cancer Res* (2005) 65(24):11752–61. doi:10.1158/0008-5472.CAN-05-1658
 85. Kaneda MM, Messer KS, Ralainirina N, Li H, Leem CJ, Gorjestani S, et al. PI3Kgamma is a molecular switch that controls immune suppression. *Nature* (2016) 539(7629):437–42. doi:10.1038/nature19834
 86. Rolny C, Mazzone M, Tugues S, Laoui D, Johansson I, Coulon C, et al. HRG inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization through downregulation of PlGF. *Cancer Cell* (2011) 19(1):31–44. doi:10.1016/j.ccr.2010.11.009
 87. Ding L, Liang G, Yao Z, Zhang J, Liu R, Chen H, et al. Metformin prevents cancer metastasis by inhibiting M2-like polarization of tumor associated macrophages. *Oncotarget* (2015) 6(34):36441–55. doi:10.18632/oncotarget.5541
 88. Hachiya M, Shimizu S, Osawa Y, Akashi M. Endogenous production of tumour necrosis factor is required for manganese superoxide dismutase expression by irradiation in the human monocytic cell line THP-1. *Biochem J* (1997) 328(Pt 2):615–23. doi:10.1042/bj3280615
 89. Wong GH. Protective roles of cytokines against radiation: induction of mitochondrial MnSOD. *Biochim Biophys Acta* (1995) 1271(1):205–9. doi:10.1016/0925-4439(95)00029-4
 90. Meng Y, Beckett MA, Liang H, Mauzeri HJ, van Rooijen N, Cohen KS, et al. Blockade of tumor necrosis factor alpha signaling in tumor-associated macrophages as a radiosensitizing strategy. *Cancer Res* (2010) 70(4):1534–43. doi:10.1158/0008-5472.CAN-09-2995
 91. Sugino N, Karube-Harada A, Sakata A, Takiguchi S, Kato H. Nuclear factor-kappa B is required for tumor necrosis factor-alpha-induced manganese superoxide dismutase expression in human endometrial stromal cells. *J Clin Endocrinol Metab* (2002) 87(8):3845–50. doi:10.1210/jcem.87.8.8771
 92. Chung YW, Kim HK, Kim IY, Yim MB, Chock PB. Dual function of protein kinase C (PKC) in 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced manganese superoxide dismutase (MnSOD) expression: activation of CREB and FOXO3a by PKC-alpha phosphorylation and by PKC-mediated inactivation of Akt, respectively. *J Biol Chem* (2011) 286(34):29681–90. doi:10.1074/jbc.M111.264945
 93. Niu Y, Shen H, Epperly M, Zhang X, Nie S, Cao S, et al. Protection of esophageal multi-lineage progenitors of squamous epithelium (stem cells) from ionizing irradiation by manganese superoxide dismutase-plasmid/liposome (MnSOD-PL) gene therapy. *In Vivo* (2005) 19(6):965–74.
 94. Epperly MW, Sikora CA, DeFilippi SJ, Gretton JE, Bar-Sagi D, Archer H, et al. Pulmonary irradiation-induced expression of VCAM-1 and ICAM-1 is decreased by manganese superoxide dismutase-plasmid/liposome (MnSOD-PL) gene therapy. *Biol Blood Marrow Transplant* (2002) 8(4):175–87. doi:10.1053/bbmt.2002.v8.pm12014807
 95. Epperly MW, Guo HL, Jefferson M, Nie S, Gretton J, Bernarding M, et al. Cell phenotype specific kinetics of expression of intratracheally injected manganese superoxide dismutase-plasmid/liposomes (MnSOD-PL) during lung radioprotective gene therapy. *Gene Ther* (2003) 10(2):163–71. doi:10.1038/sj.gt.3301852
 96. Klug F, Prakash H, Huber PE, Seibel T, Bender N, Halama N, et al. Low-dose irradiation programs macrophage differentiation to an iNOS(+)/M1 phenotype that orchestrates effective T cell immunotherapy. *Cancer Cell* (2013) 24(5):589–602. doi:10.1016/j.ccr.2013.09.014
 97. Xu J, Escamilla J, Mok S, David J, Priceman S, West B, et al. CSF1R signaling blockade stanches tumor-infiltrating myeloid cells and improves the efficacy of radiotherapy in prostate cancer. *Cancer Res* (2013) 73(9):2782–94. doi:10.1158/0008-5472.CAN-12-3981
 98. Morganti JM, Jopson TD, Liu S, Gupta N, Rosi S. Cranial irradiation alters the brain's microenvironment and permits CCR2+ macrophage infiltration. *PLoS One* (2014) 9(4):e93650. doi:10.1371/journal.pone.0093650
 99. Kalbasi A, Komar C, Tooker GM, Liu M, Lee JW, Gladney WL, et al. Tumor-derived CCL2 mediates resistance to radiotherapy in pancreatic ductal adenocarcinoma. *Clin Cancer Res* (2017) 23(1):137–48. doi:10.1158/1078-0432.CCR-16-0870
 100. Rodel F, Frey B, Gaipf U, Keilholz L, Fournier C, Manda K, et al. Modulation of inflammatory immune reactions by low-dose ionizing radiation: molecular mechanisms and clinical application. *Curr Med Chem* (2012) 19(12):1741–50. doi:10.2174/092986712800099866
 101. Crittenden MR, Cottam B, Savage T, Nguyen C, Newell P, Gough MJ. Expression of NF-kappaB p50 in tumor stroma limits the control of tumors by radiation therapy. *PLoS One* (2012) 7(6):e39295. doi:10.1371/journal.pone.0039295
 102. Okubo M, Kioi M, Nakashima H, Sugiura K, Mitsudo K, Aoki I, et al. M2-polarized macrophages contribute to neovasculogenesis, leading to relapse of oral cancer following radiation. *Sci Rep* (2016) 6:27548. doi:10.1038/srep27548
 103. Seifert L, Werba G, Tiwari S, Gao LY, Nguyen S, Alothman S, et al. Radiation therapy induces macrophages to suppress T-cell responses against pancreatic tumors in mice. *Gastroenterology* (2016) 150(7):1659–72.e5. doi:10.1053/j.gastro.2016.02.070
 104. Teresa Pinto A, Laranjeiro Pinto M, Patricia Cardoso A, Monteiro C, Teixeira Pinto M, Filipe Maia A, et al. Ionizing radiation modulates human macrophages towards a pro-inflammatory phenotype preserving their pro-invasive and pro-angiogenic capacities. *Sci Rep* (2016) 6:18765. doi:10.1038/srep18765
 105. Hildebrandt G, Radlingmayr A, Rosenthal S, Rothe R, Jahns J, Hindemith M, et al. Low-dose radiotherapy (LD-RT) and the modulation of iNOS expression in adjuvant-induced arthritis in rats. *Int J Radiat Biol* (2003) 79(12):993–1001. doi:10.1080/09553000310001636639
 106. Parker JJ, Jones JC, Strober S, Knox SJ. Characterization of direct radiation-induced immune function and molecular signaling changes in an antigen presenting cell line. *Clin Immunol* (2013) 148(1):44–55. doi:10.1016/j.clim.2013.03.008
 107. Wu Q, Allouch A, Paoletti A, Leteur C, Mirjolet C, Martins I, et al. NOX2-dependent ATM kinase activation dictates pro-inflammatory macrophage phenotype and improves effectiveness to radiation therapy. *Front Immunol* (2017) 26(8):613. doi:10.3389/fimmu.2017.00613
 108. Pinto AT, Pinto ML, Velho S, Pinto MT, Cardoso AP, Figueira R, et al. Intricate macrophage-colorectal cancer cell communication in response to radiation. *PLoS One* (2016) 11(8):e0160891. doi:10.1371/journal.pone.0160891
 109. Tsukimoto M, Homma T, Mutou Y, Kojima S. 0.5 Gy gamma radiation suppresses production of TNF-alpha through up-regulation of MKP-1 in mouse macrophage RAW264.7 cells. *Radiat Res* (2009) 171(2):219–24. doi:10.1667/RR1351.1
 110. Lodermann B, Wunderlich R, Frey S, Schorn C, Stangl S, Rodel F, et al. Low dose ionising radiation leads to a NF-kappaB dependent decreased secretion of active IL-1beta by activated macrophages with a discontinuous dose-dependency. *Int J Radiat Biol* (2012) 88(10):727–34. doi:10.3109/09553002.2012.689464
 111. Frey B, Hehlhans S, Rodel F, Gaipf US. Modulation of inflammation by low and high doses of ionizing radiation: implications for benign and malignant diseases. *Cancer Lett* (2015) 368(2):230–7. doi:10.1016/j.canlet.2015.04.010
 112. Wunderlich R, Ernst A, Rodel F, Fietkau R, Ott O, Lauber K, et al. Low and moderate doses of ionizing radiation up to 2 Gy modulate transmigration and chemotaxis of activated macrophages, provoke an anti-inflammatory cytokine milieu, but do not impact upon viability and phagocytic function. *Clin Exp Immunol* (2015) 179(1):50–61. doi:10.1111/cei.12344
 113. Shin SC, Lee KM, Kang YM, Kim K, Kim CS, Yang KH, et al. Alteration of cytokine profiles in mice exposed to chronic low-dose ionizing radiation. *Biochem Biophys Res Commun* (2010) 397(4):644–9. doi:10.1016/j.bbrc.2010.05.121
 114. Ren H, Shen J, Tomiyama-Miyaji C, Watanabe M, Kainuma E, Inoue M, et al. Augmentation of innate immunity by low-dose irradiation. *Cell Immunol* (2006) 244(1):50–6. doi:10.1016/j.cellimm.2007.02.009
 115. Ibuki Y, Goto R. Contribution of inflammatory cytokine release to activation of resident peritoneal macrophages after in vivo low-dose gamma-irradiation. *J Radiat Res* (1999) 40(3):253–62. doi:10.1269/jrr.40.253

116. Song KH, Kim MH, Kang SM, Jung SY, Ahn J, Woo HJ, et al. Analysis of immune cell populations and cytokine profiles in murine splenocytes exposed to whole-body low-dose irradiation. *Int J Radiat Biol* (2015) 91(10):795–803. doi:10.3109/09553002.2015.1068461
117. Shan YX, Jin SZ, Liu XD, Liu Y, Liu SZ. Ionizing radiation stimulates secretion of pro-inflammatory cytokines: dose-response relationship, mechanisms and implications. *Radiat Environ Biophys* (2007) 46(1):21–9. doi:10.1007/s00411-006-0076-x
118. Nowosielska EM, Cheda A, Wrembel-Wargocka J, Janiak MK. Effect of low doses of low-let radiation on the innate anti-tumor reactions in radioresistant and radiosensitive mice. *Dose Response* (2012) 10(4):500–15. doi:10.2203/dose-response.12-018.Nowosielska
119. Hoesel B, Schmid JA. The complexity of NF-kappaB signaling in inflammation and cancer. *Mol Cancer* (2013) 12:86. doi:10.1186/1476-4598-12-86
120. Rackov G, Hernandez-Jimenez E, Shokri R, Carmona-Rodriguez L, Manes S, Alvarez-Mon M, et al. p21 mediates macrophage reprogramming through regulation of p50-p50 NF-kappaB and IFN-beta. *J Clin Invest* (2016) 126(8):3089–103. doi:10.1172/JCI83404
121. Tsukimoto M, Tamaishi N, Homma T, Kojima S. Low-dose gamma-ray irradiation induces translocation of Nrf2 into nucleus in mouse macrophage RAW264.7 cells. *J Radiat Res* (2010) 51(3):349–53. doi:10.1269/jrr.10002
122. Fukuchi K, Hagiwara T, Nakamura K, Ichimura S, Tatsumi K, Gomi K. Identification of the regulatory region required for ubiquitination of the cyclin kinase inhibitor, p21. *Biochem Biophys Res Commun* (2002) 293(1):120–5. doi:10.1016/S0006-291X(02)00198-5
123. Fukuchi K, Tomoyasu S, Nakamaki T, Tsuruoka N, Gomi K. DNA damage induces p21 protein expression by inhibiting ubiquitination in ML-1 cells. *Biochim Biophys Acta* (1998) 1404(3):405–11. doi:10.1016/S0167-4889(98)00089-5
124. Yang J, Xu ZP, Huang Y, Hamrick HE, Duerksen-Hughes PJ, Yu YN. ATM and ATR: sensing DNA damage. *World J Gastroenterol* (2004) 10(2):155–60. doi:10.3748/wjg.v10.i2.155
125. Huang TT, Wuerzberger-Davis SM, Wu ZH, Miyamoto S. Sequential modification of NEMO/IKKgamma by SUMO-1 and ubiquitin mediates NF-kappaB activation by genotoxic stress. *Cell* (2003) 115(5):565–76. doi:10.1016/S0092-8674(03)00895-X
126. Hayden MS, Ghosh S. Shared principles in NF-kappaB signaling. *Cell* (2008) 132(3):344–62. doi:10.1016/j.cell.2008.01.020
127. Ibuki Y, Mizuno S, Goto R. gamma-Irradiation-induced DNA damage enhances NO production via NF-kappaB activation in RAW264.7 cells. *Biochim Biophys Acta* (2003) 1593(2–3):159–67. doi:10.1016/S0167-4889(02)00385-3
128. Brune B, Dehne N, Grossmann N, Jung M, Namgaladze D, Schmid T, et al. Redox control of inflammation in macrophages. *Antioxid Redox Signal* (2013) 19(6):595–637. doi:10.1089/ars.2012.4785
129. Zhang Y, Choksi S, Chen K, Pobezinskaya Y, Linnoila I, Liu ZG. ROS play a critical role in the differentiation of alternatively activated macrophages and the occurrence of tumor-associated macrophages. *Cell Res* (2013) 23(7):898–914. doi:10.1038/cr.2013.75
130. Covarrubias A, Byles V, Horng T. ROS sets the stage for macrophage differentiation. *Cell Res* (2013) 23(8):984–5. doi:10.1038/cr.2013.88
131. Qin L, Liu Y, Wang T, Wei SJ, Block ML, Wilson B, et al. NADPH oxidase mediates lipopolysaccharide-induced neurotoxicity and proinflammatory gene expression in activated microglia. *J Biol Chem* (2004) 279(2):1415–21. doi:10.1074/jbc.M307657200
132. Kohchi C, Inagawa H, Nishizawa T, Soma G. ROS and innate immunity. *Anticancer Res* (2009) 29(3):817–21.
133. Kang JL, Go YH, Hur KC, Castranova V. Silica-induced nuclear factor-kappaB activation: involvement of reactive oxygen species and protein tyrosine kinase activation. *J Toxicol Environ Health A* (2000) 60(1):27–46. doi:10.1080/009841000156574
134. Taetzsch T, Levesque S, McGraw C, Brookins S, Luqa R, Bonini MG, et al. Redox regulation of NF-kappaB p50 and M1 polarization in microglia. *Glia* (2015) 63(3):423–40. doi:10.1002/glia.22762
135. Choi SH, Aid S, Kim HW, Jackson SH, Bosetti F. Inhibition of NADPH oxidase promotes alternative and anti-inflammatory microglial activation during neuroinflammation. *J Neurochem* (2012) 120(2):292–301. doi:10.1111/j.1471-4159.2011.07572.x
136. Xu Q, Choksi S, Qu J, Jang J, Choe M, Banfi B, et al. NADPH oxidases are essential for macrophage differentiation. *J Biol Chem* (2016) 291(38):20030–41. doi:10.1074/jbc.M116.731216
137. He C, Carter AB. The metabolic prospective and redox regulation of macrophage polarization. *J Clin Cell Immunol* (2015) 6(6):371–7. doi:10.4172/2155-9899.1000371
138. He C, Ryan AJ, Murthy S, Carter AB. Accelerated development of pulmonary fibrosis via Cu,Zn-superoxide dismutase-induced alternative activation of macrophages. *J Biol Chem* (2013) 288(28):20745–57. doi:10.1074/jbc.M112.410720
139. Gunthner R, Anders HJ. Interferon-regulatory factors determine macrophage phenotype polarization. *Mediators Inflamm* (2013) 2013:731023. doi:10.1155/2013/731023
140. Singh S, Gupta AK. Nitric oxide: role in tumour biology and iNOS/NO-based anticancer therapies. *Cancer Chemother Pharmacol* (2011) 67(6):1211–24. doi:10.1007/s00280-011-1654-4
141. Ryu YK, Lee MH, Lee J, Lee JW, Jang SJ, Kang JH, et al. gamma-Irradiated cancer cells promote tumor growth by activation of toll-like receptor 1-mediated inducible nitric oxide synthase in macrophages. *J Leukoc Biol* (2015) 97(4):711–21. doi:10.1189/jlb.3A0114-055R
142. De Ridder M, Verellen D, Verovski V, Storme G. Hypoxic tumor cell radiosensitization through nitric oxide. *Nitric Oxide* (2008) 19(2):164–9. doi:10.1016/j.niox.2008.04.015
143. Ghosh S, Maurya DK, Krishna M. Role of iNOS in bystander signaling between macrophages and lymphoma cells. *Int J Radiat Oncol Biol Phys* (2008) 72(5):1567–74. doi:10.1016/j.ijrobp.2008.08.006
144. Cargnello M, Roux PP. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol Mol Biol Rev* (2011) 75(1):50–83. doi:10.1128/MMBR.00031-10
145. Kim HS, Asmis R. Mitogen-activated protein kinase phosphatase 1 (MKP-1) in macrophage biology and cardiovascular disease. A redox-regulated master controller of monocyte function and macrophage phenotype. *Free Radic Biol Med* (2017) 109:75–83. doi:10.1016/j.freeradbiomed.2017.03.020
146. Kim HS, Tavakoli S, Piefer LA, Nguyen HN, Asmis R. Monocytic MKP-1 is a sensor of the metabolic environment and regulates function and phenotypic fate of monocyte-derived macrophages in atherosclerosis. *Sci Rep* (2016) 6:34223. doi:10.1038/srep34223
147. Salojin KV, Owusu IB, Millerchip KA, Potter M, Platt KA, Oravec T. Essential role of MAPK phosphatase-1 in the negative control of innate immune responses. *J Immunol* (2006) 176(3):1899–907. doi:10.4049/jimmunol.176.3.1899
148. Chi H, Barry SP, Roth RJ, Wu JJ, Jones EA, Bennett AM, et al. Dynamic regulation of pro- and anti-inflammatory cytokines by MAPK phosphatase 1 (MKP-1) in innate immune responses. *Proc Natl Acad Sci U S A* (2006) 103(7):2274–9. doi:10.1073/pnas.0510965103
149. Narang H, Krishna M. Effect of nitric oxide donor and gamma irradiation on MAPK signaling in murine peritoneal macrophages. *J Cell Biochem* (2008) 103(2):576–87. doi:10.1002/jcb.21429
150. Kobayashi EH, Suzuki T, Funayama R, Nagashima T, Hayashi M, Sekine H, et al. Nrf2 suppresses macrophage inflammatory response by blocking proinflammatory cytokine transcription. *Nat Commun* (2016) 7:11624. doi:10.1038/ncomms11624
151. Chen W, Sun Z, Wang XJ, Jiang T, Huang Z, Fang D, et al. Direct interaction between Nrf2 and p21(Cip1/WAF1) upregulates the Nrf2-mediated antioxidant response. *Mol Cell* (2009) 34(6):663–73. doi:10.1016/j.molcel.2009.04.029
152. Kalbasi A, June CH, Haas N, Papiwala N. Radiation and immunotherapy: a synergistic combination. *J Clin Invest* (2013) 123(7):2756–63. doi:10.1172/JCI69219
153. Gupta A, Probst HC, Vuong V, Landshammer A, Muth S, Yagita H, et al. Radiotherapy promotes tumor-specific effector CD8+ T cells via dendritic cell activation. *J Immunol* (2012) 189(2):558–66. doi:10.4049/jimmunol.1200563
154. Lee Y, Auh SL, Wang Y, Burnette B, Wang Y, Meng Y, et al. Therapeutic effects of ablative radiation on local tumor require CD8+ T cells: changing strategies for cancer treatment. *Blood* (2009) 114(3):589–95. doi:10.1182/blood-2009-02-206870

155. Hodge JW, Guha C, Neefjes J, Gulley JL. Synergizing radiation therapy and immunotherapy for curing incurable cancers. Opportunities and challenges. *Oncology (Williston Park)* (2008) 22(9):1064–84.
156. Shiao SL, Ruffell B, DeNardo DG, Faddegon BA, Park CC, Coussens LM. TH2-polarized CD4(+) T cells and macrophages limit efficacy of radiotherapy. *Cancer Immunol Res* (2015) 3(5):518–25. doi:10.1158/2326-6066.CIR-14-0232
157. Lee S, Kivimae S, Dolor A, Szoka FC. Macrophage-based cell therapies: the long and winding road. *J Control Release* (2016) 240:527–40. doi:10.1016/j.jconrel.2016.07.018
158. Hellweg CE, Baumstark-Khan C, Schmitz C, Lau P, Meier MM, Testard I, et al. Carbon-ion-induced activation of the NF-kappaB pathway. *Radiat Res* (2011) 175(4):424–31. doi:10.1667/RR2423.1

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer, BB, and handling editor declared their shared affiliation, and the handling editor states that the process nevertheless met the standards of a fair and objective review.

Copyright © 2017 Genard, Lucas and Michiels. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.