



Institutional Repository - Research Portal

Dépôt Institutionnel - Portail de la Recherche

researchportal.unamur.be

RESEARCH OUTPUTS / RÉSULTATS DE RECHERCHE

Draft Genome Sequences of Three Capnocytophaga canimorsus Strains Isolated from Septic Patients

Manfredi, Pablo; Renzi, Francesco; Cornelis, Guy

Published in:
Genome Announcements

DOI:
[10.1128/genomeA.00193-15](https://doi.org/10.1128/genomeA.00193-15)

Publication date:
2015

Document Version
Publisher's PDF, also known as Version of record

[Link to publication](#)

Citation for pulished version (HARVARD):
Manfredi, P, Renzi, F & Cornelis, G 2015, 'Draft Genome Sequences of Three Capnocytophaga canimorsus Strains Isolated from Septic Patients' Genome Announcements. <https://doi.org/10.1128/genomeA.00193-15>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Draft Genome Sequences of Three *Capnocytophaga canimorsus* Strains Isolated from Septic Patients

Pablo Manfredi,^a Francesco Renzi,^{a,b} Guy R. Cornelis^{a,b}

Biozentrum der Universität Basel, Basel, Switzerland^a; University of Namur, Namur, Belgium^b

***Capnocytophaga canimorsus* is a bacterium from the normal oral flora of dogs and cats that causes rare generalized infections in humans. In an attempt to determine whether infections could be caused by a subset of strains and to identify pathogenicity factors, we sequenced the genomes of three strains isolated from human infections.**

Received 12 February 2015 Accepted 28 April 2015 Published 28 May 2015

Citation Manfredi P, Renzi F, Cornelis GR. 2015. Draft genome sequences of three *Capnocytophaga canimorsus* strains isolated from septic patients. *Genome Announc* 3(3): e00193-15. doi:10.1128/genomeA.00193-15.

Copyright © 2015 Manfredi et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Guy R. Cornelis, guy.cornelis@unamur.be.

Capnocytophaga canimorsus is a Gram-negative commensal bacterium from the normal canine oral flora that causes life-threatening septicemia in patients who have been in contact with dogs or cats (1). Infections present with fulminant sepsis (2, 3), including cases of meningitis, endocarditis, or myocarditis. Fastidious growth of the pathogen and lack of symptoms during the initial stages of infection often lead to an unattended wound (4), resulting in a mortality rate of as high as 30% (1). Predisposing factors, like splenectomy (33%) or alcohol abuse (24%), have indeed been reported, but in 41% of the cases, patients do not show any obvious risk factors (1). Recent molecular studies on the pathogenicity of the strain Cc5 (5) led to the identification of different pathogenic factors of *C. canimorsus* (6–12). In particular, the data presented here allowed the identification of a new type of iron import system required for growth of the pathogen in human serum (13).

The three clinical strains of *C. canimorsus*, Cc2, Cc11, and Cc12 (i.e., ATCC 35979), were isolated from human patients (blood samples) who developed septicemia (7). Genomic DNA was extracted using the Genomic-tip 500/G DNA extraction kit (catalog no. 10262; Qiagen), according to the manufacturer's instructions, followed by an additional phenol-chloroform purification step. A total of 1.3 million (Cc11), 2.3 million (Cc12), and 2.4 million (Cc2) paired-end microreads (36 bp; fragment length, 205 ± 50 bp) were generated at Fasteris SA, Geneva (Switzerland), from a single run of Solexa/Illumina GAII EAS269 on 100 tiles. Additional sequencing data were generated with the Roche genome sequencing FLX system DNA pyrosequencing at Microsynth, Balgach, Switzerland, corresponding to 75,000 to 80,000 reads per strain of approximately 315-bp read length. The assemblies also included primer walking Sanger sequencing data generated on cherry-picked regions. The final hybrid assembly was performed with MIRA (14) and included pseudoreads corresponding to contigs mapped onto the reference chromosome of *C. canimorsus* strain 5 (Cc5) (5) using MAQ (15) and to contigs from the *de novo* assembly generated with Velvet with optimized parameters (16). Genome annotation and preliminary analyses were performed by LABGeM, France Génomique (17). The genomic metrics of the

three draft assemblies (206 [Cc2], 249 [Cc11], and 81 [Cc12] contigs) were similar to those of Cc5 (5), with draft assembly sizes ranging from 2.39 to 2.52 Mb (2.57 for Cc5), G+C content between 36.08% and 36.23% (36.11% for Cc5), and a total of 2,702 to 2,874 coding sequences (CDS) detected (2,519 for Cc5). The *C. canimorsus* core genome based on the four strains mentioned above included 1,292 genes, which corresponded to only 45 to 51% of the total genes in each strain, and therefore indicates high genomic plasticity within the taxon. With respect to pathogenicity factor candidates, 177 clusters of orthologs were found conserved in the four clinical isolates but not in six additional *C. canimorsus* strains isolated from a dog's mouth (18). Also, 118 clusters with unknown function, 18 genes involved in oxidative respiration, 10 in ion and peptide transport, 10 in mobile element transposition, 4 in transcriptional regulation, and 3 in cell adhesion formed the predominant functional classes of the list.

Nucleotide sequence accession numbers. These whole-genome shotgun projects have been deposited in ENA under the accession numbers [CDOJ00000000](https://ena.ebi.ac.uk/ena/record/CDOJ00000000) (Cc2), [CDOK00000000](https://ena.ebi.ac.uk/ena/record/CDOK00000000) (Cc11), and [CDOE00000000](https://ena.ebi.ac.uk/ena/record/CDOE00000000) (Cc12). The versions described in this paper are the initial versions.

ACKNOWLEDGMENTS

This work was supported by grants 3100A0-128659 from the Swiss National Science Foundation and ERC 2011-ADG 20110310 from the European Research Council.

F.R. is a postdoctoral research fellow (chargé de recherche) of the Belgian Fonds National de la Recherche Scientifique (FNRS).

REFERENCES

1. Lion C, Escande F, Burdin JC. 1996. *Capnocytophaga canimorsus* infections in human: review of the literature and cases report. *Eur J Epidemiol* 12:521–533. <http://dx.doi.org/10.1007/BF00144007>.
2. De Boer MG, Lambregts PC, van Dam AP, van 't Wout JW. 2007. Meningitis caused by *Capnocytophaga canimorsus*: when to expect the unexpected. *Clin Neurol Neurosurg* 109:393–398. <http://dx.doi.org/10.1016/j.clineuro.2007.02.010>.
3. Pers C, Gahrn-Hansen B, Frederiksen W. 1996. *Capnocytophaga canimorsus* septicemia in Denmark, 1982–1995: review of 39 cases. *Clin Infect Dis* 23:71–75. <http://dx.doi.org/10.1093/clinids/23.1.71>.

4. Tierney DM, Strauss LP, Sanchez JL. 2006. *Capnocytophaga canimorsus* mycotic abdominal aortic aneurysm: why the mailman is afraid of dogs. *J Clin Microbiol* 44:649–651. <http://dx.doi.org/10.1128/JCM.44.2.649-651.2006>.
5. Manfredi P, Pagni M, Cornelis GR. 2011. Complete genome sequence of the dog commensal and human pathogen *Capnocytophaga canimorsus* strain 5. *J Bacteriol* 193:5558–5559. <http://dx.doi.org/10.1128/JB.05853-11>.
6. Shin H, Mally M, Meyer S, Fiechter C, Paroz C, Zaehring U, Cornelis GR. 2009. Resistance of *Capnocytophaga canimorsus* to killing by human complement and polymorphonuclear leukocytes. *Infect Immun* 77:2262–2271. <http://dx.doi.org/10.1128/IAI.01324-08>.
7. Shin H, Mally M, Kuhn M, Paroz C, Cornelis GR. 2007. Escape from immune surveillance by *Capnocytophaga canimorsus*. *J Infect Dis* 195:375–386. <http://dx.doi.org/10.1086/510243>.
8. Renzi F, Manfredi P, Mally M, Moes S, Cornelis GR, Jenö P, Cornelis GR. 2011. The *N*-glycan glycoprotein deglycosylation complex (Gpd) from *Capnocytophaga canimorsus* deglycosylates human IgG. *PLoS Pathog* 7:e1002118. <http://dx.doi.org/10.1371/journal.ppat.1002118>.
9. Meyer S, Shin H, Cornelis GR. 2008. *Capnocytophaga canimorsus* resists phagocytosis by macrophages and blocks the ability of macrophages to kill other bacteria. *Immunobiology* 213:805–814. <http://dx.doi.org/10.1016/j.imbio.2008.07.019>.
10. Manfredi P, Renzi F, Mally M, Sauteur L, Schmalzer M, Moes S, Cornelis GR, Jenö P, Cornelis GR. 2011. The genome and surface proteome of *Capnocytophaga canimorsus* reveal a key role of glycan foraging systems in host glycoproteins deglycosylation. *Mol Microbiol* 81:1050–1060. <http://dx.doi.org/10.1111/j.1365-2958.2011.07750.x>.
11. Mally M, Shin H, Paroz C, Landmann R, Cornelis GR. 2008. *Capnocytophaga canimorsus*: a human pathogen feeding at the surface of epithelial cells and phagocytes. *PLoS Pathog* 4:e1000164. <http://dx.doi.org/10.1371/journal.ppat.1000164>.
12. Ittig S, Lindner B, Stenta M, Manfredi P, Zdorovenko E, Knirel YA, dal Peraro M, Cornelis GR, Zähringer U. 2012. The lipopolysaccharide from *Capnocytophaga canimorsus* reveals an unexpected role of the core-oligosaccharide in MD-2 binding. *PLoS Pathog* 8:e1002667. <http://dx.doi.org/10.1371/journal.ppat.1002667>.
13. Manfredi P, Lauber F, Renzi F, Hack K, Hess E, Cornelis GR. 2015. New iron acquisition system in *Bacteroidetes*. *Infect Immun* 83:300–310. <http://dx.doi.org/10.1128/IAI.02042-14>.
14. Chevreux B, Pfisterer T, Drescher B, Driesel AJ, Müller WE, Wetter T, Suhai S. 2004. Using the miraEST assembler for reliable and automated mRNA transcript assembly and SNP detection in sequenced ESTs. *Genome Res* 14:1147–1159. <http://dx.doi.org/10.1101/gr.1917404>.
15. Li H, Ruan J, Durbin R. 2008. Mapping short DNA sequencing reads and calling variants using mapping quality scores. *Genome Res* 18:1851–1858. <http://dx.doi.org/10.1101/gr.078212.108>.
16. Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
17. Vallenet D, Belda E, Calteau A, Cruveiller S, Engelen S, Lajus A, Le Fèvre F, Longin C, Mornico D, Roche D, Rouy Z, Salvignol G, Scarpelli C, Thil Smith AA, Weiman M, Médigue C. 2013. MicroScope—an integrated microbial resource for the curation and comparative analysis of genomic and metabolic data. *Nucleic Acids Res* 41:D636–D647. <http://dx.doi.org/10.1093/nar/gks1194>.
18. Manfredi P, Renzi F, Cornelis GR. 2015. Draft genome sequences of three *Capnocytophaga canimorsus* strains isolated from healthy canine oral cavities. *Genome Announc* 3(3):e00199-15. <http://dx.doi.org/10.1128/genomeA.00199-15>.