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**Author:** Urszula Guzik, Izabela Greń, Danuta Wojcieszńska, Sylwia Łabużek

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## ISOLATION AND CHARACTERIZATION OF A NOVEL STRAIN OF *STENOTROPHOMONAS MALTOPHILIA* POSSESSING VARIOUS DIOXYGENASES FOR MONOCYCLIC HYDROCARBON DEGRADATION

Guzik Urszula; Greń Izabela; Wojcieszńska Danuta; Łabużek Sylwia

Department of Biochemistry, Faculty of Biology and Environment Protection, University of Silesia, Katowice, Poland

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### ABSTRACT

A Gram-negative bacterium, designated as strain KB2, was isolated from activated sludge and was found to utilize different aromatic substrates as sole carbon and energy source. On the basis of morphological and physicochemical characteristics and 16S rRNA gene sequence analysis, the isolated strain KB2 was identified as *Stenotrophomonas maltophilia*. Strain KB2 is from among different *Stenotrophomonas maltophilia* strains the first one described as exhibiting the activities of three types of dioxygenases depending on the structure of the inducer. The cells grown on benzoate and catechol showed mainly catechol 1,2-dioxygenase activity. The activity of 2,3-dioxygenase was detected after phenol induction. Protocatechuate 3,4-dioxygenase was found in crude cell extracts of this strain after incubation with 4-hydroxybenzoic acid, protocatechuic acid and vanillic acid. Because of broad spectrum of dioxygenases' types that *Stenotrophomonas maltophilia* KB2 can exhibit, this strain appears to be very powerful and useful tool in the biotreatment of wastewaters and in soil decontamination.

**Key words:** biodegradation, *Stenotrophomonas*, aromatic compounds, dioxygenases

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### INTRODUCTION

Aromatic compounds are environmental pollutants due to their widespread use and potential toxicity to higher organisms. The catabolic versatility exhibited by microorganisms plays an important role in the carbon cycle and the bioremediation of polluted environments. Pathways for aerobic biodegradation of phenolic compounds are diverse, but they usually involve the formation of catechols as common intermediates. Catechol 1,2-dioxygenase [EC 1.13.11.1] catalyzes an intradiol cleavage reaction of catechol at 1,2-(*ortho*) position to form *cis,cis*-muconate. The catechol 2,3-dioxygenase [EC 1.13.11.2], an extradiol-ring fission enzyme, catalyzes the conversion of catechol to 2-hydroxymuconic semialdehyde by breaking the C-C bond at 2,3-(*meta*) position (6, 19, 26). Protocatechuate is one of the key intermediates for microbial catabolism of benzoic molecules. These compounds are degraded through one of two (*ortho* or *meta*) aromatic ring cleavage pathways, too.

Protocatechuate may be cleaved by protocatechuate 3,4-dioxygenase [EC 1.13.11.3], which catalyzes the intradiol addition of molecular oxygen and forms 3-carboxy-*cis,cis*-muconate or by protocatechuate 4,5-dioxygenase [1.13.11.8]. This last enzyme catalyzes extradiol addition of molecular oxygen, which results in the aromatic ring cleaving and 4-carboxy-2-hydroxy-*cis,cis*-muconic semialdehyde forming (6,20,26).

In our lab we isolated a bacterial strain, which was analyzed for the type of ring-cleavage dioxygenase physiologically induced during growth in the presence of six monocyclic aromatic compounds (phenol, catechol, benzoic acid, 4-hydroxybenzoic acid, protocatechuic acid and vanillic acid), which served as carbon and energy sources for strain KB2. Although *Stenotrophomonas* species capable of degrading aromatic compounds have been reported earlier (4,17) little has been investigated on the possibility of occurrence of the heterogeneous metabolic pathways in one strain of this genus (28). So far *Stenotrophomonas* species were mainly described

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\*Corresponding Author. Mailing address: Department of Biochemistry, Faculty of Biology and Environment Protection, University of Silesia, Jagiellonska 28, 40-032 Katowice, Poland. Phone: +48322009576. E-mail: urszula.guzik@us.edu.pl

as polycyclic hydrocarbon degraders or opportunistic human pathogens (2,3,4,8,10).

In this study, for the first time the isolation and characterization of a Gram-negative *Stenotrophomonas maltophilia* strain, designated as KB2, which exhibits activities of three types of dioxygenases while growing in the presence of monocyclic hydrocarbons, has been reported.

## MATERIALS AND METHODS

### Isolation of an aromatic-degrading bacterium

An aromatic compound degrading strain was isolated from the activated sludge of a sewage treatment plant in Bytom Miechowice in Poland using the classical enrichment technique with phenol as a selection factor. The mixed microbial population from the activated sludge was adapted to grow on phenol at a concentration of 10 mM. The adaptation process was carried out at 20°C in an aeration chamber. On the 10th day of adaptation, a 2 ml sample was taken from the culturing chamber and 100 µl of 10<sup>-1</sup> to 10<sup>-7</sup> were spread on agar plates containing mineral salts medium (Na<sub>2</sub>HPO<sub>4</sub> · 12H<sub>2</sub>O 3.78 g; KH<sub>2</sub>PO<sub>4</sub> 0.5 g; NH<sub>4</sub>Cl 5 g; MgSO<sub>4</sub> · 7H<sub>2</sub>O; per litre distilled water) with 3 mM phenol to obtain pure cultures. Agar plates were incubated at 30°C for 24h and single colonies were isolated and transferred to nutrient agar plates to test their purity. Strain was kept on nutrient agar slopes at 4°C and systematically transferred to the new ones every month.

### Morphological, physiological and biochemical characterization of the isolated strain

The isolated strain was phenotypically and biochemically characterized using standard techniques (Gram staining, colony shape, size and colour on nutrient agar plate, catalase and oxidase test, etc.), according to Bergey's Manual of Determinative Bacteriology (12). Additional biochemical and physiological characteristics were determined using the API 20NE and API 20E system (BioMerieux, Lyon, France).

Isolation of fatty acids was performed according to Sasser (23). Analysis of FAMES was performed using an HP 5890 gas chromatograph (Hewlett Packard, Rolling Meadows, IL, US) equipped with an HP 25 m x 0.2 mm cross-linked methyl-silicone capillary column. The initial oven temperature was 170°C, increased 5°C min<sup>-1</sup> to 260°C, the increased 40°C min<sup>-1</sup> and held constant at 320°C for 1.5 min (9). Helium was used as the carrier gas. FAMES were identified with Sherlock software (TSBA library, version 3.9, Microbial ID, Newark, NJ, USA), based on the actual calibration retention times run prior to sample analysis.

### Analytical methods

The aromatic substrates were determined by a Merck HITACHI HPLC with a LiChromospher® RP-18 column (4 × 250 mm) and a DAD detector (Merck HITACHI). The wavelength of

detection, composition of elution solvent and the flow rate were developed separately for each aromatic compound.

### Culture conditions

In order to verify, which aromatic substrate can serve as the sole source of carbon and energy, adaptations to the increasing concentrations of various aromatic substrates were carried out. Cells were proliferated in mineral medium with 3 mM of phenol (at 30°C on a rotary shaker at 130 rpm), harvested by centrifugation (5,000 × g at 4°C for 15 min) and washed with fresh sterile medium. Such prepared cells were used as inoculum for the experiments with adaptation. Cultures in 250-ml flask containing 100 ml of sterile mineral salts medium supplemented with 1 mM of the tested aromatic compound, were inoculated with previously prepared cells to the final optical density about 0.1 in absorbance scale at λ = 600 nm, and incubated shaking at 130 rpm at 30°C for 24 hours. Chromatographic analyses of the culture fluid and measurements of the cultures growth (spectrophotometrically at 600 nm) were carried out every 24 hours. If growth of the cultures and complete degradation of the aromatic substrate was observed, the successive dose (2 mM and higher) of the aromatic substrate was introduced and the cultures were left for incubation for the next 24 hours. Lack of aromatic substrate degradation and absence of growth for three following days finished the adaptation process.

The aromatic substrates, which could serve as the source of carbon and energy, were used as the inducers of enzymes activities. Induction experiments were carried out in 1-litre flasks, containing 500 ml of mineral salts medium and aromatic substrate in concentration of 3 mM. Inducer was added every 24 hours for three days in order to obtain final A<sub>600</sub> of 0.9-1.0.

These induction's were preceded by preparation of the cells suspensions as follows: colonies were picked from nutrient agar and transferred to a 50-ml conical flask containing 20 ml of mineral salt medium with 3 mM of the appropriate aromatic substrate. After 24 hours of incubation the suspension of the cells was transferred to a 250-ml flask containing 80 ml of mineral medium with 3 mM of inducer. The cells were allowed to grow the next 24 hours and this inoculum was used to prepare induction experiments. No-induced enzyme levels were determined in the cells coming from the cultures carried out in mineral salts medium supplemented with 1 g/L glucose.

### Preparation of cell extracts

Cells were harvested in the late exponential phase of growth by centrifugation at 4,500 g for 15 min at 4°C. The cells were washed with 50 mM phosphate buffer, pH 7.0, and resuspended in the same buffer. Cell-free extracts were prepared by sonication 6 times for 15 s and centrifugation at 9,000 g for 30 min at 4°C. The clear supernatant was used as crude extract for enzyme assays.

### Enzyme assays

Activity of catechol 1,2-dioxygenase [EC 1.13.11.1] was measured spectrophotometrically by formation of *cis,cis*-muconic acid at 260 nm ( $\epsilon_{260} = 16,800 \text{ M}^{-1} \text{ cm}^{-1}$ ). The reaction mixture contained 20  $\mu\text{l}$  of catechol (50 mM), 67  $\mu\text{l}$   $\text{Na}_2\text{EDTA}$  (20 mM), 893  $\mu\text{l}$  of phosphoric buffer pH 7.4 (50 mM) and 20  $\mu\text{l}$  of crude extracts in a total volume of 1 ml. When activity of catechol 2,3-dioxygenase was detected, crude cell extract was incubated with 5%  $\text{H}_2\text{O}_2$  prior to determination of catechol 1,2-dioxygenase. In order to determine catechol 2,3-dioxygenase [EC 1.13.11.2] activity, the formation of 2-hydroxymuconic semialdehyde was measured at 375 nm ( $\epsilon_{375} = 36,000 \text{ M}^{-1} \text{ cm}^{-1}$ ) in a reaction mixture containing 20  $\mu\text{l}$  of catechol (50 mM), 960  $\mu\text{l}$  of phosphoric buffer pH 7.4 (50 mM) and 20  $\mu\text{l}$  of crude extract in a total volume of 1 ml (11). Activity of protocatechuate 3,4-dioxygenase [EC 1.13.11.3] was measured by protocatechuate depletion at 290 nm in a reaction mixture containing 20  $\mu\text{l}$  of protocatechuate (50 mM), 960  $\mu\text{l}$  of Tris-HCl buffer pH 7.4 (50 mM) and 20  $\mu\text{l}$  of crude extract in a total volume of 1 ml. A molar extinction coefficient of  $2,300 \text{ M}^{-1} \text{ cm}^{-1}$  was used, which is the difference between  $\epsilon_{290}$  of protocatechuate ( $3,890 \text{ M}^{-1} \text{ cm}^{-1}$ ) and  $\epsilon_{290}$  of the product 3-carboxy-*cis cis*-muconate ( $1,590 \text{ M}^{-1} \text{ cm}^{-1}$ ). In order to determine protocatechuate 4,5-dioxygenase [EC 1.13.11.8] activity, the formation of 2-hydroxy-4-carboxymuconic semialdehyde was measured at 410 nm ( $\epsilon_{410} = 9,700 \text{ M}^{-1} \text{ cm}^{-1}$ ). The reaction mixture contained 20  $\mu\text{l}$  of protocatechuate (50 mM), 960  $\mu\text{l}$  of Tris-HCl buffer pH 7.4 (50 mM) and 20  $\mu\text{l}$  of crude extract in a total volume of 1 ml (25). One unit of enzyme activity was defined as the amount of enzyme required to generate 1  $\mu\text{mol}$  of product per minute. Protein concentrations of the crude extract from different inducer-cultured bacteria were determined by the Bradford method using lysozyme as a standard (5).

### Phylogenetic analysis

Bacterial DNA was isolated from pure culture using the DNA Mini Prep Kit (Qiagen). For 16S rRNA gene amplification the Bacteria-specific primers 8F 5' AGTTTGATCATCGCTCAG 3' and 1492R 5' GGTTACCTTGTTACGACTT3' were used (18). Amplification was carried out through a program consisting of initial denaturation at 94°C for 300 s, 3 cycles at 94°C for 45 s, 57°C for 30 s, 72°C for 120 s; 3 cycles at 94°C for 45 s, 56°C for 30 s, 72°C for 120 s; 3 cycles at 94°C for 45 s, 55°C for 30 s, 72°C for 120 s; 26 cycles at 94°C for 45 s, 53°C for 30 s, 72°C for 120 s; and a final elongation cycle at 72°C for 300 s. The nucleotide sequencing of the gene was done by using Big Dye<sup>®</sup> Terminator Cycle Sequencing Kit (Applied Biosystem) and AbiPrism<sup>®</sup>3100 Genetic Analyzer. The MegaBLAST program was used for homology searches with the standard program default. Multiple sequence alignments were performed and the neighbour-joining phylogenetic tree was constructed using CLC Free Workbench 4.5.1 program. The 16S rRNA gene sequence determined in

this study has been deposited in the GeneBank database of NCBI under the accession number **DQ230920**.

### Chemicals

Phenol, monochlorophenols, cresols, 2,3-, 2,4-, 2,6-dichlorophenols, catechol, hydroquinone, cinnamic and gallic acid were obtained from E. Merck AG, Darmstadt, Germany. 2,5- and 3,4-dichlorophenol, vanillic, syringic, 4-hydroxybenzoic and protocatechuic acid were obtained from FLUKA, Fluka AG, Buchs, Switzerland. Resorcinol, caffeic, sinapic and *p*-coumaric acid were obtained from Sigma Chemical Co., St. Louis, Mo. Ferulic acid and 2,5-dihydroxybenzoic acid were obtained from Aldrich Chemical Co., Inc., Milwaukee, Wis. Benzoic and salicylic acid, and other chemicals were of analytical quality and were obtained from local suppliers.

## RESULTS AND DISCUSSION

### Isolation and identification of strain degrading aromatic compounds

The present work was focused on isolation and characterization of bacterial strain capable of degrading high concentrations of aromatic compounds. Although phenol was supplied as the carbon and energy source during the enrichment process, this strain could also utilize different kind of aromatic compounds such as catechol, benzoic acid, 4-hydroxybenzoic acid (4-HB), protocatechuic acid (3,4-DHB), vanillic acid. All substrates that were tested for their ability to support growth of KB2 strain, are summarized in Table 1.

Morphological, physiological and biochemical characterisation and 16S rRNA gene sequencing were used to identify strain KB2. Cells of strain KB2 were Gram-negative, rod shaped, motile and nonspore forming. Colonies of strain KB2 were found to be circular, smooth, convex and with the entire margin during growing on agar plates for 48 hours. The biochemical and physiological characteristic of strain KB2 are summarized in Table 2. The pattern generated with API 20NE test was a good match with *Xanthomonas maltophilia*. Similar results were obtained by Radianingtyas and co-workers (22). These authors identified the particular microorganisms of consortium degrading 4-chloroaniline. Strain SB5 which was able to grow on 4-chlorocatechol was identified using API 20NE test as *Xanthomonas maltophilia* and by using 16S rRNA method as *Stenotrophomonas maltophilia* (22). In Table 3 fatty acids from strain KB2 are listed. The cellular fatty acid content gave little information what species the isolated strain KB2 could belong to. Glucksman and co-workers (9) during analysis of fatty acid content of *Stenotrophomonas maltophilia* ATCC 13637 observed the largest contribution of some typical for *Pseudomonas* genus fatty acids: 16:0 and 16:1  $\omega$ 7c (9). These cellular fatty acids were also observed in fatty acid profile of the strain KB2. Because there is no clear data what kinds of cellular fatty acids are the most typical for genus *Stenotrophomonas*, in

**Table 1** Utilization of aromatic compounds as the sole source of carbon by the strain KB2 (in brackets the highest concentration degraded by strain within 24 hours is given).

Aromatic compounds	Results	Aromatic compounds	Results
phenol	+(12 mM)	catechol	+(6 mM)
2-chlorophenol	-	resorcinol	-
3-chlorophenol	-	hydroquinone	+(4 mM)
4-chlorophenol	-	vanillic acid	+(6 mM)
<i>o</i> -cresol	+(6 mM)	syringic acid	-
<i>m</i> -cresol	+(6 mM)	protocatechuic acid	+(13 mM)
<i>p</i> -cresol	+(5 mM)	benzoic acid	+(10 mM)
2-nitrophenol	-	4-hydroxybenzoic acid	+(9 mM)
3-nitrophenol	-	gentisic acid	-
4-nitrophenol	-	cinnamic acid	-
2,3-dichlorophenol	-	salicylic acid	-
2,4-dichlorophenol	-	gallic acid	-
2,5-dichlorophenol	-	caffeic acid	-
2,6-dichlorophenol	-	ferulic acid	-
3,4-dichlorophenol	-	<i>p</i> -coumaric acid	-
3,5-dichlorophenol	-	sinapic acid	-

order to confirm phylogenetic relationship of strain KB2 with *Stenotrophomonas* genus, genomic DNA was isolated and the gene coding for 16S rRNA was amplified. The nearly complete 1445-bp-long 16S rRNA gene sequence of strain KB2 was found to be 99% identical to that of *Stenotrophomonas maltophilia* strain D1 ([EU340025.1](#)). A high percentage identity was also found with *Stenotrophomonas* sp. strain Sea-8, a phenol degrader (28) (Fig. 1). In accordance with these data, the isolate was included in the genus *Stenotrophomonas* and named as *Stenotrophomonas maltophilia* KB2.

#### Induction of the various types of dioxygenase in cells of strain KB2

To investigate which degradation pathway was used by the strain KB2 for growth on aromatic compounds, the presence of enzymes involved in the *ortho*- and *meta*- cleavage pathways for dissimilation of catechol or protocatechuate was determined using crude cell extracts after sonication. Depending on the structure of inductor three various dioxygenases were induced in *Stenotrophomonas maltophilia* KB2 strain cells (Table 4). These results suggest that strain KB2 has three main metabolic pathways to degrade aromatic compounds. The existence of two different metabolic routes for the aerobic breakdown of aromatic compounds is now well established in bacteria (26).

**Table 2** Differential phenotypic characteristics of strain KB2.

Characteristic	Results	Characteristic	Results
Growth in the absence of NaCl	+	Assimilation of:	
Growth in the presence of 1.5% (w/v) NaCl	+	Glucose	+
Growth in the presence of 3% (w/v) NaCl	+	Arabinose	+
Growth at 4°C	-	Mannose	+
Growth at 42°C	+	Mannitol	+
Oxidase	+	Inositol	+
Catalase	+	Sorbitol	+
Hydrolysis of esculin	+	Rhamnose	+
Hydrolysis of gelatin	+	Sucrose	+
Fermentation of glucose	-	Melbiose	-
Arginine dihydrolase	-	Maltose	+
Urease	-	Gluconate	-
H <sub>2</sub> S production	-	Caprate	+
Indol production	-	Adipate	-
Nitrate reduction	-	Malate	+
Acetoin production	-	Citrate	+
Tryptophane desaminase	-	Phenylacetate	-
Ornithine decarboxylase	-	N-acetyl-glucosamine	+
Beta-galactosidase	-	Amygdalin	+

+, positive reaction; -, negative reaction.

**Table 3.** Percentage of total fatty acid from *Stenotrophomonas maltophilia* KB2.

Fatty acids	% of total fatty acids	Fatty acids	% of total fatty acids
Saturated		18:0	0.94
10:0	0.64	19:0	0.22
12:0	2.39	19:0 <i>iso</i>	0.23
12:0 2OH	4.81	19:0 <i>cyclo</i> ω8c	1.29
14:0	0.45	Unsaturated	
14:0 3OH	0.08	16:1 ω7c	16.33
15:0	0.26	17:1 ω8c	0.18
15:0 <i>iso</i> 2OH	3.31	18:1 ω7c	23.45
16:0	32.09	18:1 ω9c	0.3
17:0	0.25	other	0.24
17:0 <i>cyclo</i>	12.46	Sat./unsat. ratio	1.48
17:0 <i>iso</i>	0.08		

ω – methyl end of fatty acid;

c – *cis* configuration of the double bound;

*cyclo*-cyclopropane fatty acid;

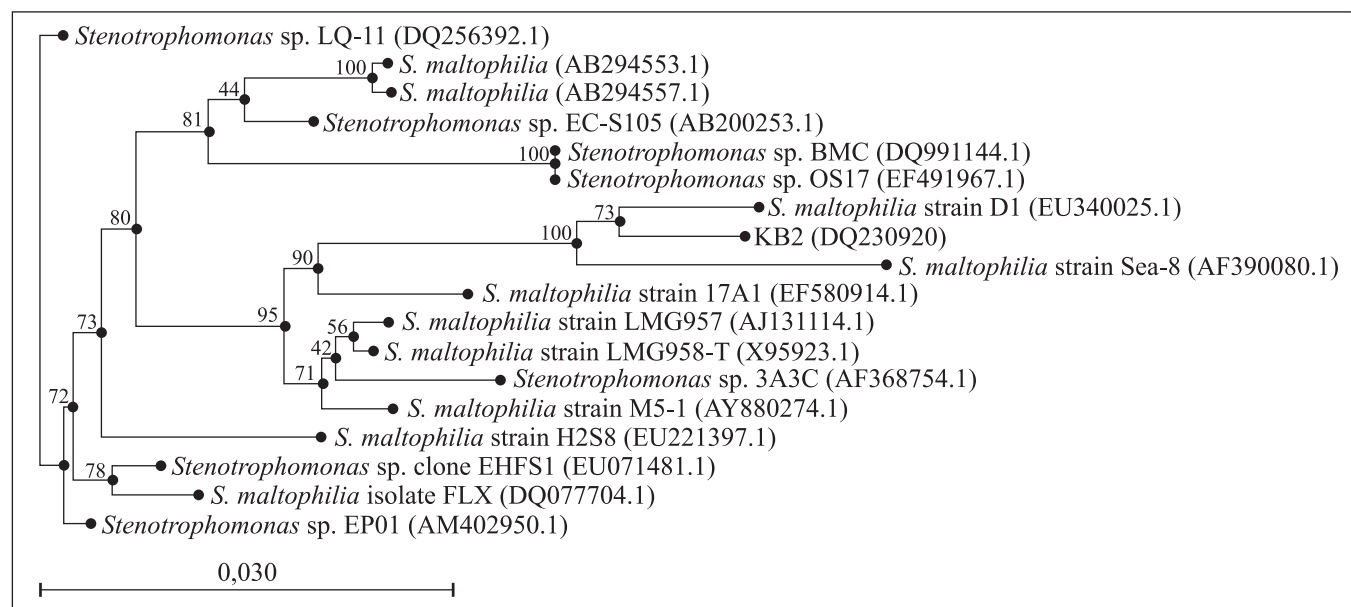
-OH indicates the position of hydroxyl group from the acid end;

*iso*-branched fatty acids.

**Table 4.** Dioxygenase activities in cell-free extracts from *Stenotrophomonas maltophilia* KB2 grown on 3 mM aromatic compounds. Catechol was used as substrate in the reaction mixture for detection of catechol dioxygenase activities and protocatechuate – for protocatechuate dioxygenases. The results are an average of at least three independent measurements.

Inductor	Specific activity $\mu\text{mol min}^{-1} \text{mg}^{-1}$ of protein			
	Catechol 1,2-dioxygenase	Catechol 2,3-dioxygenase	Protocatechuate 3,4-dioxygenase	Protocatechuate 4,5-dioxygenase
Phenol	0.06 ± 0.02	4.92 ± 0.34	ND <sup>a</sup>	ND
Catechol	1.03 ± 0.01	0.37 ± 0.00	ND	ND
4-HB	ND	ND	6.37 ± 0.72	ND
3,4-DHB	ND	0.01 ± 0.00	2.98 ± 0.72	ND
Vanillic acid	0.12 ± 0.03	0.04 ± 0.02	8.70 ± 1.36	ND
Benzoic acid	11.97 ± 3.04	0.02 ± 0.01	ND	ND
Glucose	ND	ND	ND	ND

<sup>a</sup>ND – not detected; 4-HB – 4-hydroxybenzoic acid; 3,4-DHB – 3,4-dihydroxybenzoic acid.



**Figure 1.** Neighbor-joining tree showing the phylogenetic position of the aromatic substances-degrading bacterium strain KB2 and related species of the genus *Stenotrophomonas* based on partial 16S rRNA gene sequences. The GenBank accession number for each microorganism used in the analysis is shown in parentheses after the species name. Bootstrap values (expressed as percentage of 100 replicons) are shown at the branch.

Growth on benzoate mainly induced either catechol 1,2- and 2,3- dioxygenase (24) or 1,2-benzoate dioxygenases and catechol 1,2-dioxygenase (1). Although activity of 1,2-benzoate dioxygenase activity in the crude cell extract of strain KB2 was not determined, but the absence of 3,4-protocatechuate dioxygenase activity after benzoic acid induction suggested

that *Stenotrophomonas maltophilia* KB2 degraded benzoate through the catechol pathway. Lack of monooxygenase activity after benzoate induction (data not shown) also confirmed that hypothesis. Among the *Stenotrophomonas maltophilia* species only one described as strain Sea-8 was found to express catechol dioxygenases activities after phenol induction, although the



activity of catechol 2,3-dioxygenase in that case was rather small (28). Contrary to Sea-8 strain, *Stenotrophomonas maltophilia* strain KB2 growing on 3 mM phenol showed trace activity of intradiol catechol dioxygenase (Table 3). After growth on hydroxybenzoic acids, additional cleaving dioxygenase is usually induced (26). 4-Hydroxybenzoic acid as well as vanillic acid were found to induce mainly protocatechuate 3,4-dioxygenase (7,13,14,16,21). Gentisate dioxygenase was also found in the cells grown on hydroxybenzoic acids (6,27). But to the authors' knowledge, so far only one strain *Pseudomonas* sp. K82 was described as to be able to induce three different kinds of dioxygenases including protocatechuate 4,5-dioxygenase for *p*-hydroxybenzoate degradation, catechol 1,2-dioxygenase for benzoate decomposition and catechol 2,3-dioxygenase for breakdown of aniline. By contrast in *Stenotrophomonas maltophilia* KB2 strain the protocatechuate 3,4-dioxygenase pathway was the only one induced by 4-hydroxybenzoate, protocatechuate or vanillic acid exposure (Table 3). Presence of metabolic pathways for both catechol and protocatechuic acid breakdown increases the biodegradation potential of *Stenotrophomonas maltophilia* strain KB2 and its ability to perform a role of the universal tool to fight the increasing pollution of the environment. Co-metabolic biodegradation of aromatics by strain KB2 and activity of dioxygenases in such conditions are under investigations.

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#### RESUMO

##### **Isolamento e caracterização de uma nova cepa de *Stenotrophomonas maltophilia* com várias dioxygenases para degradação de hidrocarbonetos monocíclicos**

Uma bactéria Gram-negativa, denominada KB2, foi isolada de lodo ativado, verificando-se ser capaz de utilizar substratos aromáticos com única fonte de carbono e energia. Com base nas características morfológicas e físico-químicas, e na análise da sequência do gene 16SrRNA, esta bactéria foi identificada como *Stenotrophomonas maltophilia*. Entre as diversas cepas de *S. maltophilia* já descritas, essa cepa é a primeira com atividade de três tipos de dioxygenases, dependendo da estrutura do indutor. As células cultivadas em benzoato e catecol apresentaram atividade de catecol 1,2-dioxygenase principalmente. A atividade de 2,3-dioxygenase foi detectada após indução com fenol. Após incubação com ácidos 4-hidrobencóico, ácido protocatecuico e vanílico, encontrou-se protocatecuato 3,4-dioxygenase no extrato celular. Devido ao amplo espectro de atividade das diferentes

dioxygenases de *S. maltophilia* KB2, esta cepa parece ser uma ferramenta poderosa e útil para o biotratamento de efluentes e descontaminação do solo.

**Palavras-chave:** biodegradação, *Stenotrophomonas*, compostos aromáticos, dioxygenases

#### REFERENCES

- Adebusoye, S.A.; Picardal, F.W.; Ilori, M.O.; Amund, O.O.; Fuqua, C.; Grindle, N. (2007). Aerobic degradation of di- and trichlorobenzenes by two bacteria isolated from polluted tropical soils. *Chemosphere*, 66, 1939-1946.
- Almeida, M.T.G.; Rubio, F.G.; Garcia, D.O.; Pavarino-Bertelli, E.C.; Rossit, A.R.B.; Bando, S.Y.; Silbert, S.; Goloni-Bertollo, E.M.; Soares, M.M.C.N.; Martinez, M.B. (2007). Genetic relatedness among clinical strains of *Stenotrophomonas maltophilia* in tertiary care hospital settings in Sao Paulo State, Brazil. *Braz. J. Microbiol.*, 38, 278-284.
- Assih, E.A.; Ouattara, A.S.; Thierry, S.; Cayol, J-L.; Labat, L.; Macarie, H. (2002). *Stenotrophomonas acidaminiphila* sp. nov., a strictly aerobic bacterium isolated from an upflow anaerobic sludge blanket (UASB) reactor. *Int. J. Syst. Evol. Microbiol.*, 52, 559-568.
- Boonchan, S.; Britz, M.L.; Stanley, G.A. (1998). Surfactant-enhanced biodegradation of high molecular weight polycyclic aromatic hydrocarbons by *Stenotrophomonas maltophilia*. *Biotechnol. Bioengin.*, 59, 482-494.
- Bradford, M.M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.*, 72, 248-258.
- Bugg, T.D.H. (2003). Dioxygenase enzymes: catalytic mechanisms and chemical models. *Tetrahedron*, 59, 7075-7101.
- Deveryshetty, J.; Suvekkala, V.; Varadamshetty, G.; Phale, P.S. (2007). Metabolism of 2-, 3- and 4-hydroxybenzoate by soil isolates *Alcaligenes* sp. strain PPH and *Pseudomonas* sp. strain PPD. *FEMS Microbiol. Lett.*, 268, 59-66.
- Finkmann, W.; Altendorf, K.; Stackebrandt, E.; Lipski, A. (2000). Characterization of N<sub>2</sub>O-producing *Xanthomonas*-like isolates from biofilters as *Stenotrophomonas nitritireducens* sp. nov., *Luteimonas mephitis* gen. nov., sp. nov. and *Pseudoxanthomonas broegbermensis* gen. nov., sp. nov. *Int. J. Syst. Evol. Microbiol.*, 50, 273-282.
- Glucksman, A.M.; Skipper, H.D.; Brigmon, R.L.; Santo Domingo, J.W. (2000). Use of the MIDI-FAME technique to characterize groundwater communities. *J. Appl. Microbiol.*, 88, 711-719.
- Hauben, L.; Vauterin, L.; Moore, E.R.B.; Hoste, B.; Swings, J. (1999). Genomic diversity of the genus *Stenotrophomonas*. *Int. J. Syst. Bacteriol.*, 49, 1749-1760.
- Hegeman, G.D. (1966). Synthesis of enzymes of the mandelate pathways by *Pseudomonas putida*. Synthesis of enzyme by the wild type. *J. Bacteriol.*, 91, 1140-1154.
- Holt, J.G.; Krieg, N.R.; Sneath, P.H.A.; Staley, J.T.; Williams, S.T. (1994). *Bergey's manual of determinative bacteriology*, Williams and Wilkins, USA
- Kahng, H-Y.; Cho, K.; Song, S-Y.; Kim, S-J.; Leem, S-H.; Kim, S.I. (2002). Enhanced detection and characterization of protocatechuate 3,4-dioxygenase in *Acinetobacter lwoffii* K24 by proteomics using a column separation. *Biochem. Biophys. Res. Commun.*, 295, 903-909.
- Kim, D.; Kim, S.W.; Choi, K.Y.; Lee, J.S.; Kim, E. (2008). Molecular cloning and functional characterization of the genes encoding benzoate and *p*-hydroxybenzoate degradation by the halophilic *Chromohalobacter* sp. strain HS-2. *FEMS Microbiol. Lett.*, 280, 235-241.

15. Kim, S.; Kim, J.Y.; Yun, S-H.; Kim, J-Y.; Leem, S-H.; Lee, C. (2004). Proteome analysis of *Pseudomonas* sp. K82 biodegradation pathways. *Proteomics*, 4, 3610-3621.
16. Kim, Y.H.; Cho, K.; Yun, S-H.; Kim, J.Y.; Kwom, K-H.; Yoo, J.S.; Kim, S.I. (2006). Analysis of aromatic catabolic pathways in *Pseudomonas putida* KT 2440 using a combined proteomic approach: 2-DE/MS and cleavable isotope-coded affinity tag analysis. *Proteomics*, 6, 13301-1318.
17. Liu, Z.; Yang, C.; Qiao, C.H. (2007). Biodegradation of *p*-nitrophenol and 4-chlorophenol by *Stenotrophomonas* sp. *FEMS Microbiol. Lett.*, 277, 150-156.
18. Lonergan, D.J.; Jenter, H.L.; Coates, J.D.; Phillips, E.J.; Schmidt, T.M.; Lovley, D.R. (1996). Phylogenetic analysis of dissimilatory Fe (III) – Reducing Bacteria. *J. Bacteriol.*, 178, 2402-2408.
19. Matthew, W.; Vetting, M.W.; Douglas, H.; Ohlendorf, D.H. (2008). The 1.8 Å crystal structure of catechol 1,2-dioxygenase reveals a novel hydrophobic helical zipper as a subunit linker. *Structure*, 8, 429-440.
20. Orville, A.M.; Lipscomb, J.D.; Ohlendorf, D.H. (1997). Crystal structures of substrate and substrate analog complexes of protocatechuate 3,4-dioxygenase: endogenous Fe<sup>3+</sup> ligand displacement in response to substrate binding. *Biochemistry*, 36, 10052-10066.
21. Park, S-H.; Kim, J-W.; Yun, S-H.; Lem, S.H.; Kahng, H-Y.; Kim, S.I. (2006). Characterization of β-ketoadipate pathway from multi-drug resistance bacterium, *Acinetobacter baumannii* DU202 by proteomic approach. *J. Microbiol.*, 44, 632-640.
22. Radianingtyas, H.; Robinson, G.K.; Bull, A.T. (2003). Characterization of a soil-derived bacterial consortium degrading 4-chloroaniline. *Microbiology*, 149, 3279-3287.
23. Sasser, M. (1990). *Technical note #*, 101-110.
24. Song, J.; Sung, J.; Kim, Y.M.; Zylstra, G.J.; Kim, E. (2000). Roles of the *meta*- and the *ortho*-cleavage pathways for the efficient utilization of aromatic hydrocarbons by *Sphingomonas yanoikuyae* B1. *J. Microbiol.*, 38, 245-249.
25. Stanier, R.Y.; Ingraham, J.L. (1954). Protocatechuic acid oxidase. *J. Biol. Chem.*, 210, 799-808.
26. Vaillancourt, F.H.; Bolin, J.T.; Eltis, L.D. (2006). The Ins and Outs of ring-cleaving dioxygenases. *Crit. Rev. Biochem. Mol. Biol.*, 41, 241-267.
27. Yoon, Y-H.; Yun, S-H.; Park, S-H.; Seol, S-Y.; Leem, S-H.; Kim, S.I. (2007). Characterization of a new catechol branch of the β-ketoadipate pathway induced for benzoate degradation in *Acinetobacter lwoffii* K24. *Biochem. Biophys. Res. Commun.*, 360, 513-519.
28. Zaki, S. (2006). Detection of *meta*- and *orto*-cleavage dioxygenases in bacterial phenol-degraders. *J. Appl. Sci. Environ. Manage.*, 10, 75-81.