

Soil Respiration

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I. Background

Understanding the role of terrestrial ecosystems in governing the isotopic composition of atmospheric CO_2 requires knowledge of environmental controls of $\delta^{13}C$ and $\delta^{18}O$ of CO_2 in biosphere-atmosphere CO_2 exchange. Heterotrophic respiration contributes approximately half of the total respiratory backflow of CO_2 from terrestrial ecosystems to the atmosphere, but too little is known about environmental controls of C and O isotopic signatures of CO_2 released from soils to be able to reliably quantify the contribution of different CO_2 flux components (photosynthesis, above- and belowground autotrophic as well as heterotrophic respiration) to net ecosystem CO_2 exchange.

II. Experiment The dependencies of $\delta^{13}C$ and $\delta^{18}O$ of CO $_2$

temperature

spruce

incubated

from heterotrophic soil respiration on soil

investigated. Samples of the organic layer (O)

and humified A horizon (Ah) of a Norway

(Höglwald close to Augsburg, Germany) were

moisture levels in gas-tight incubation tubes

(exetainers), followed by isotope-ratio mass spectrometer (IRMS) analysis of the $\rm CO_2$ formed during heterotrophic soil respiration.

The increase of CO2 mixing ratios during the

incubations, monitored by IRMS at different

sampling times, allowed for calculation of the

 $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ of the respiratory source of the

CO₂ by the Keeling plot methodology

(L.)

different temperature

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soil

moisture

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forest

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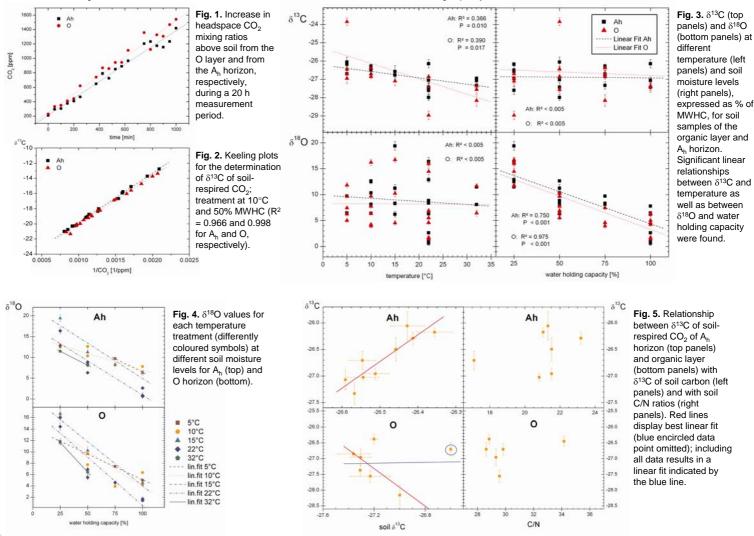
III. Method

Samples of the O layer below the needle litter and of the ${\rm A}_{\rm h}$ horizon were taken on the 10th of May 2007 and 30th of August 2007, taken back to the lab on the same day, and kept refrigerated at 5°C until incubation and analysis. Preparation of refrigerated soil samples started two days prior to analytical determination of the $\delta^{13} C$ and $\delta^{18} O$ of soil-respired CO_2. First, samples were sieved at 4 mm in order to homogenate the sample material and to exclude fine roots. Subsequently, the sieved samples of O and A_h horizon were air-dried for one night. Then, some g of soil material, depending on the temperature and humidity treatment, were placed into 12 mL exetainers (Labco Ltd., UK). For each temperature and moisture level and each horizon 18 replicates were analyzed. The specific amount of water required to achieve the target soil moisture level was added to each exetainer. To avoid drying of samples, the exetainers were sealed with parafilm. Samples were then incubated for one night at the respective temperature, which was controlled with a custommade sample tray connected to a water bath. The parafilm was removed and the samples were aerated for another 1-2 hours before sealing the exetainers with the appropriate septum screw caps prior to IRMS analysis with a gas bench coupled to a continuous-flow mass spectrometer (Delta^{Plus}XP, ThermoFisher, Bremen, Germany).



IV. Results

Values for δ^{13} C are given in ‰ vs. PDB, values for δ^{18} O in ‰ vs. SMOW; MWHC = maximum water holding capacity.



V. Summary

The δ^{13} C of the CO₂ respired from root-free organic layer and A_h horizon did not show any significant relation to soil water content. In contrast, δ^{13} C had a negative significant linear relationship with temperature. Data fluctuated around an average δ^{13} C of -25.6‰ for the A_h horizon and -26.4‰ for the organic layer, reflecting the fact that the organic layer was more depleted in ¹³C than the A_h horizon. Likewise, the CO₂ from the organic layer was more depleted in ¹³O than the CO₂ from the A_h horizon. In contrast to the δ^{13} C results, no significant relationship of δ^{18} O to temperature could be observed. However, a highly significant decrease in δ^{19} O of respiratory CO₂ from both soil layers with increasing soil moisture was found, decreasing from an average of +14‰ vs. SMOW at 25% MWHC to +4‰ at 100% MWHC.

KIT – a cooperation between Karlsruhe Research Center and University of Karlsruhe

