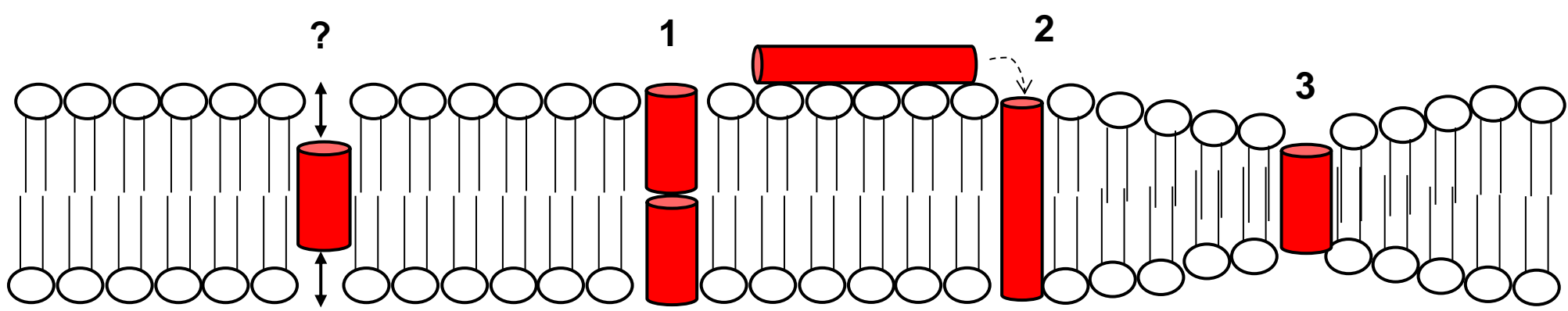


Introduction

Peptaibols are naturally occurring peptides isolated from fungal sources. They contain a high amount of **Aib (U)** (α -aminoisobutyric acid), possess a C-terminal alcohol and N-terminal acetylation/alkylation. They are able to permeabilize bacterial membranes by pore formation. **Harzianin HK-VI (HZ)** is a short peptaibol (11-mer) isolated from *Trichoderma pseudokoningii*. [1]

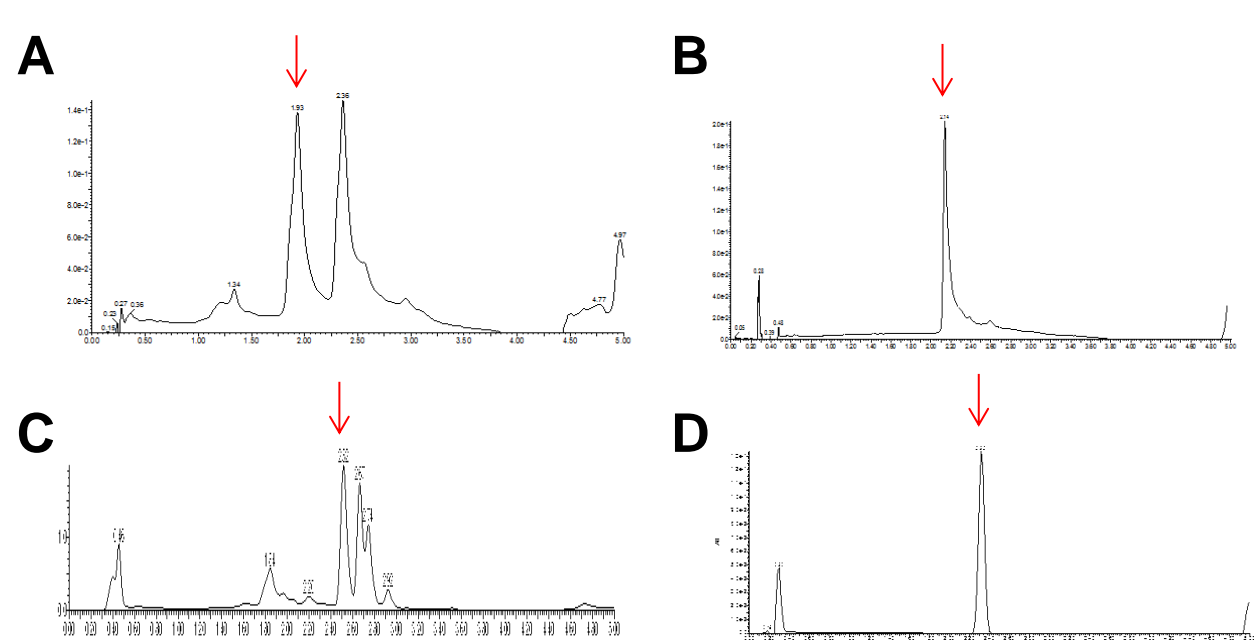
Aims

We aim to solve the structure of membrane-bound **HZ** and understand its interactions with lipid bilayers.

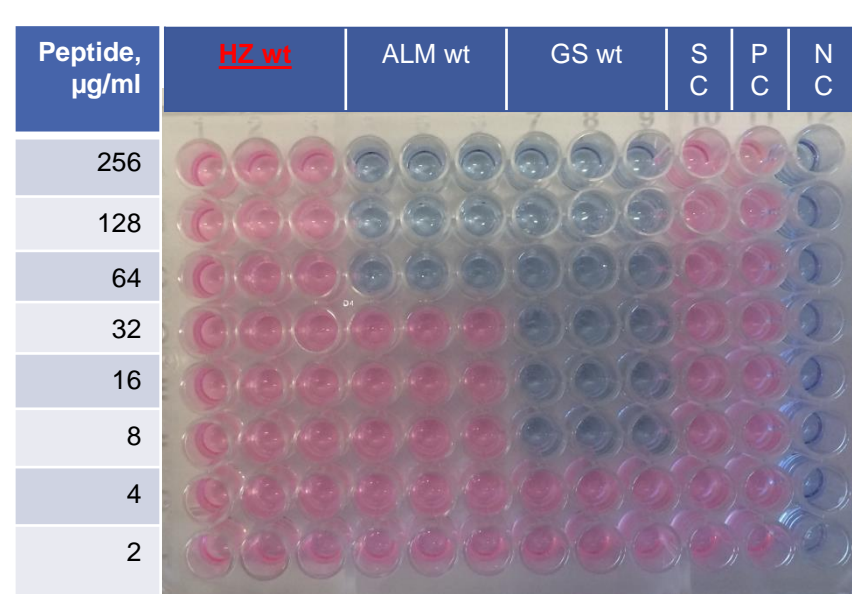


Hypothetical mechanisms how short pore-forming peptides can interact with lipid bilayers: by forming double-layered channels (1), by changing to an extended conformation (2), by causing membrane thinning (3).

Results



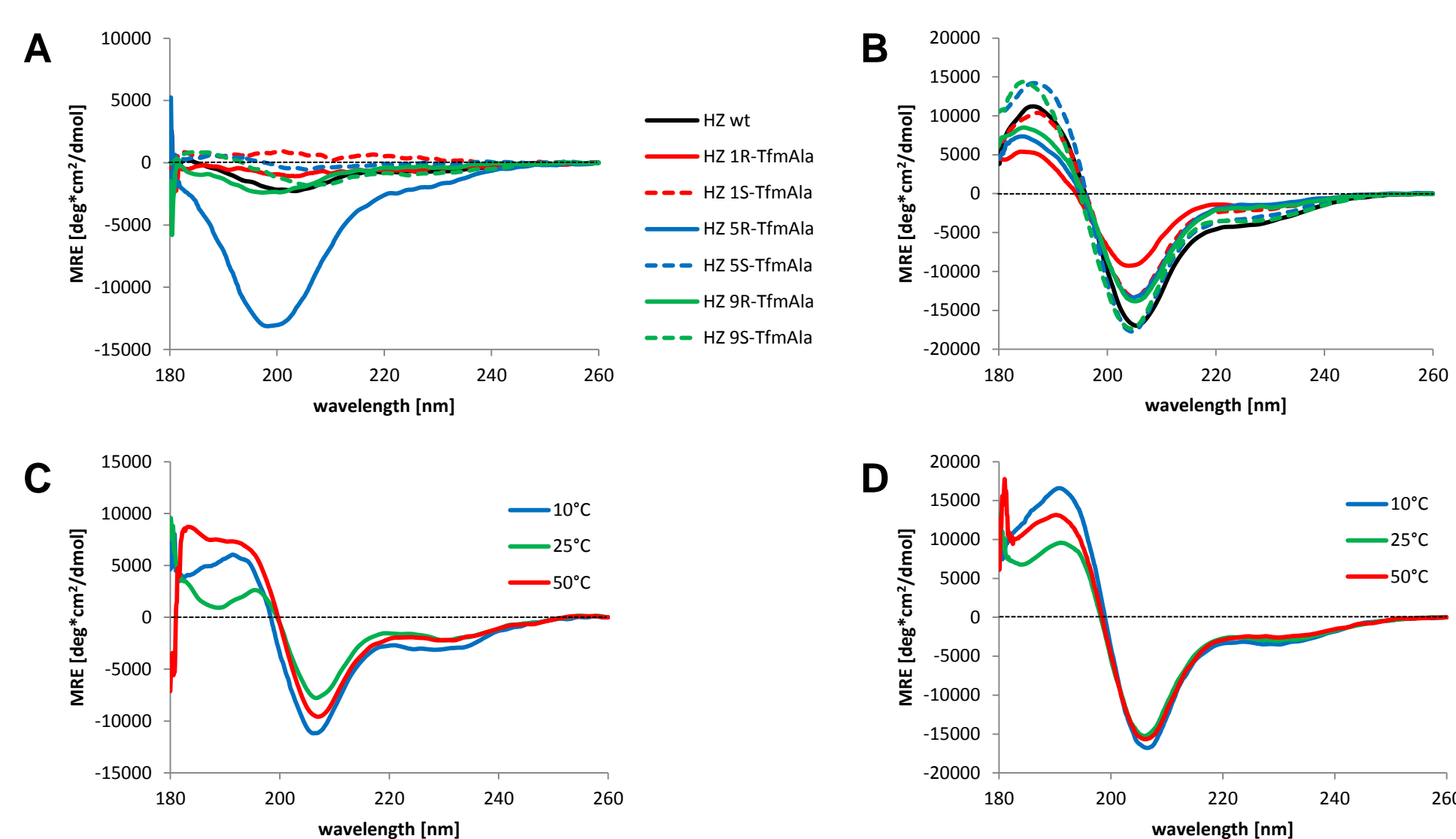
HPLC profiles of synthesized peptides: **HZ wt** crude (A) and pure (B), **HZ 1-(S)-Tfm-Ala** crude (C) and pure (D).



Representative MIC experiment. Strain: *B. subtilis* ATCC 6633. ALM: Alamehycin F30/3; GS: Gramicidin S; SC: solvent control; PC: positive control; NC: negative control.

Strain	MIC [μ g/ml]	
	HZ wt	HZ Tfm-Ala peptides
<i>E. coli</i> K12	> 256	> 256
<i>S. aureus</i> DSM 1104	128	> 256
<i>S. xyloso</i> us DSM 20267	> 256	> 256
<i>E. faecalis</i> DSM 2570	> 256	> 256
<i>B. subtilis</i> ATCC 6633	> 256	> 256

MIC values for HZ.

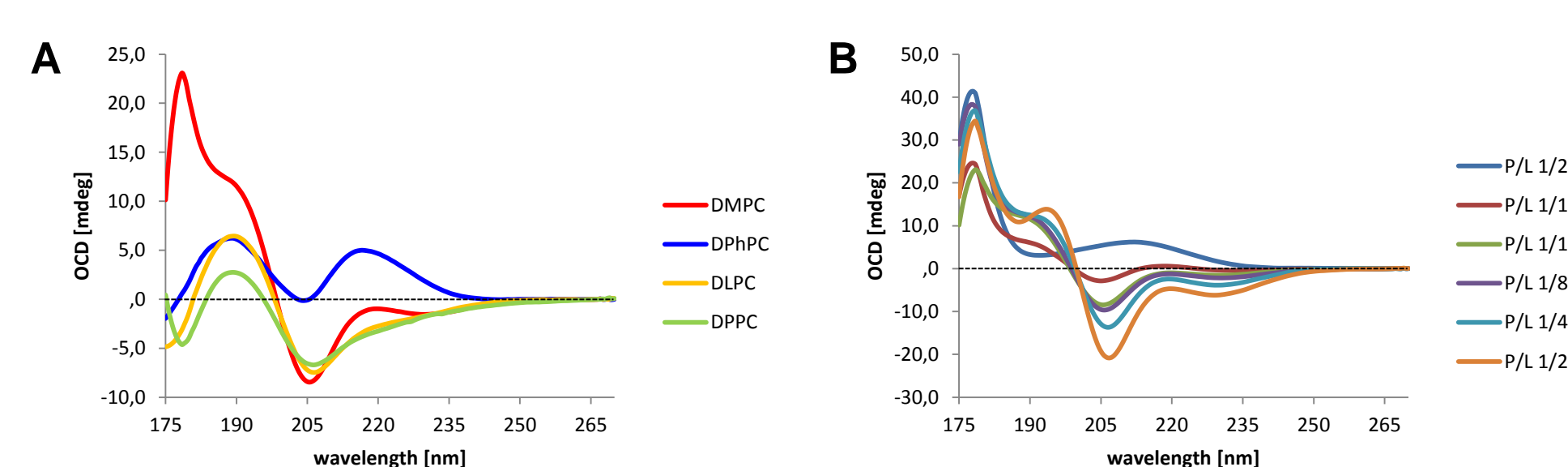


CD spectra of HZ peptides in H₂O (A) and TFE (B); HZ wt (C) and HZ 5-(S)-Tfm-Ala (D) in DMPC vesicles (P/L 1/20).

3₁₀-helical conformation

Algorithm	α	3_{10}	β	turns	PPII	NRMSD
CONTIN	31.3	10.4	0	14	9.8	0.253
SELCON3	29.8	10.2	-2.3	13.5	8.8	0.132
CDSSTR	39	10	5	11	9	0.005

Deconvolution of the SR-CD spectrum for HZ wt in 50% TFE.

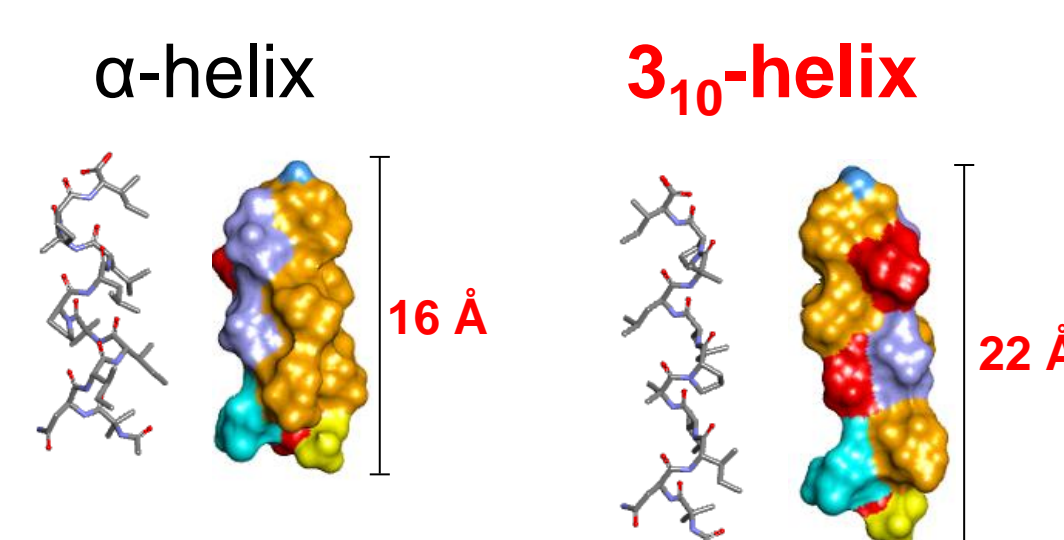


SR-OCD spectra: HZ wt in different lipid membranes at constant concentration (P/L 1/100) (A); HZ in DMPC at varying P/L (B).

Conclusions

HZ peptides

- possess no antimicrobial activity
- adopt a 3₁₀-helical structure when bound to the lipid membrane
- show re-orientation in lipid membranes, dependent on the lipid composition and the lipid phase state



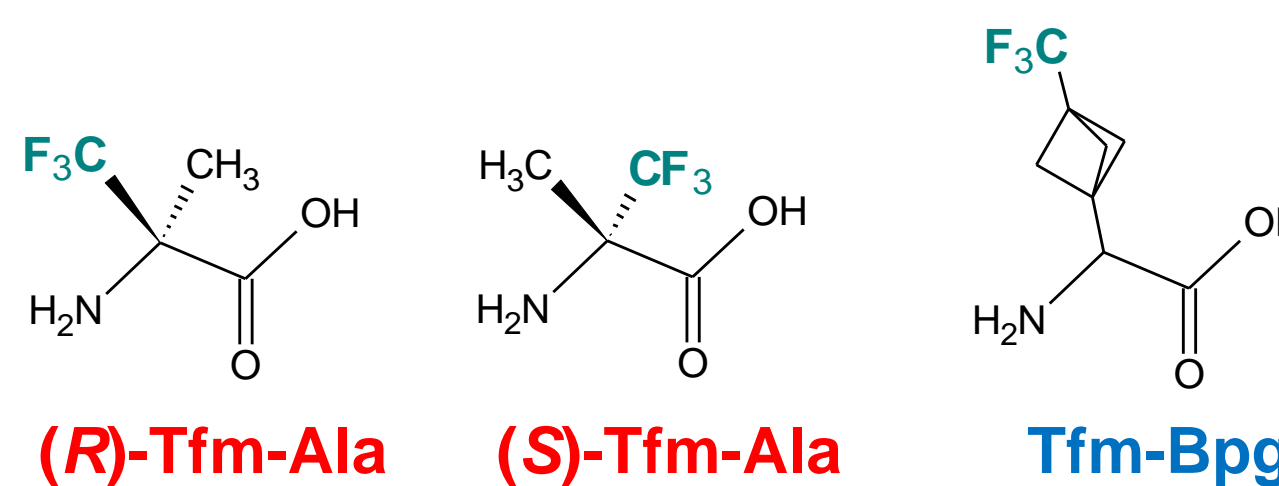
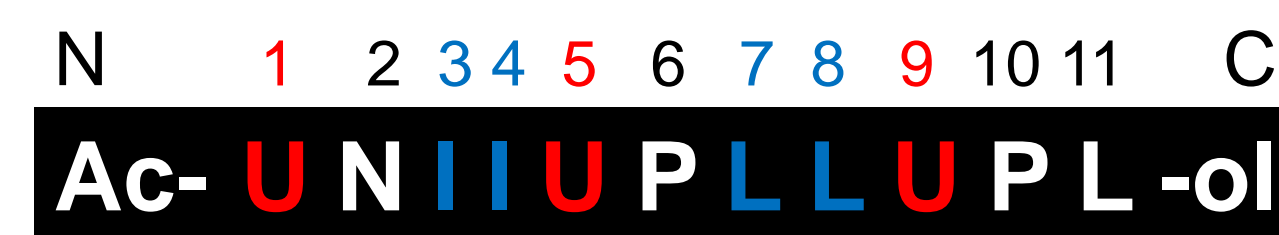
References:

[1] S. Rebuffat, S. Hlimi, Y. Prigent, C. Goulard, B. Bodo, J. Chem. Soc., Perkin Trans. 1 (1996) 2021; [2] F. Huguenot, T. Brigaud, J. Org. Chem. 71 (2006) 7075–7078; [3] D. Maisch, P. Wadhvani, S. Afonin, C. Böttcher, B. Koksich, A.S. Ulrich, J. Am. Chem. Soc. 131 (2009) 15596–15597; [4] P.K. Mikhailiuk, S. Afonin, A.N. Chernega, E.B. Rusanov, M. Platonov, G. Dubinina, M. Berditsch, A.S. Ulrich, I.V. Komarov, Angewandte Chem. 118 (2006), 5787–5789; [5] A.S. Ulrich, Progress in Nuclear Magnetic Resonance Spectroscopy 46 (2005) 1–21; [6] I. Segalas, Y. Prigent, D. Davoust, B. Bodo, S. Rebuffat, Biopolymers (1999) 71–85.

Methods

Solid-state ¹⁹F-NMR and circular dichroism (CD) spectroscopy are used to determine the structure and alignment of **HZ** in lipid bilayers. For ¹⁹F-NMR analysis, we use α -trifluoromethylated (Tfm) amino acids (**R**-, (**S**)-Tfm-Ala [2,3] and Tfm-Bpg [4] as substitutes of **Aib** and **Leu/Ile** residues, respectively.

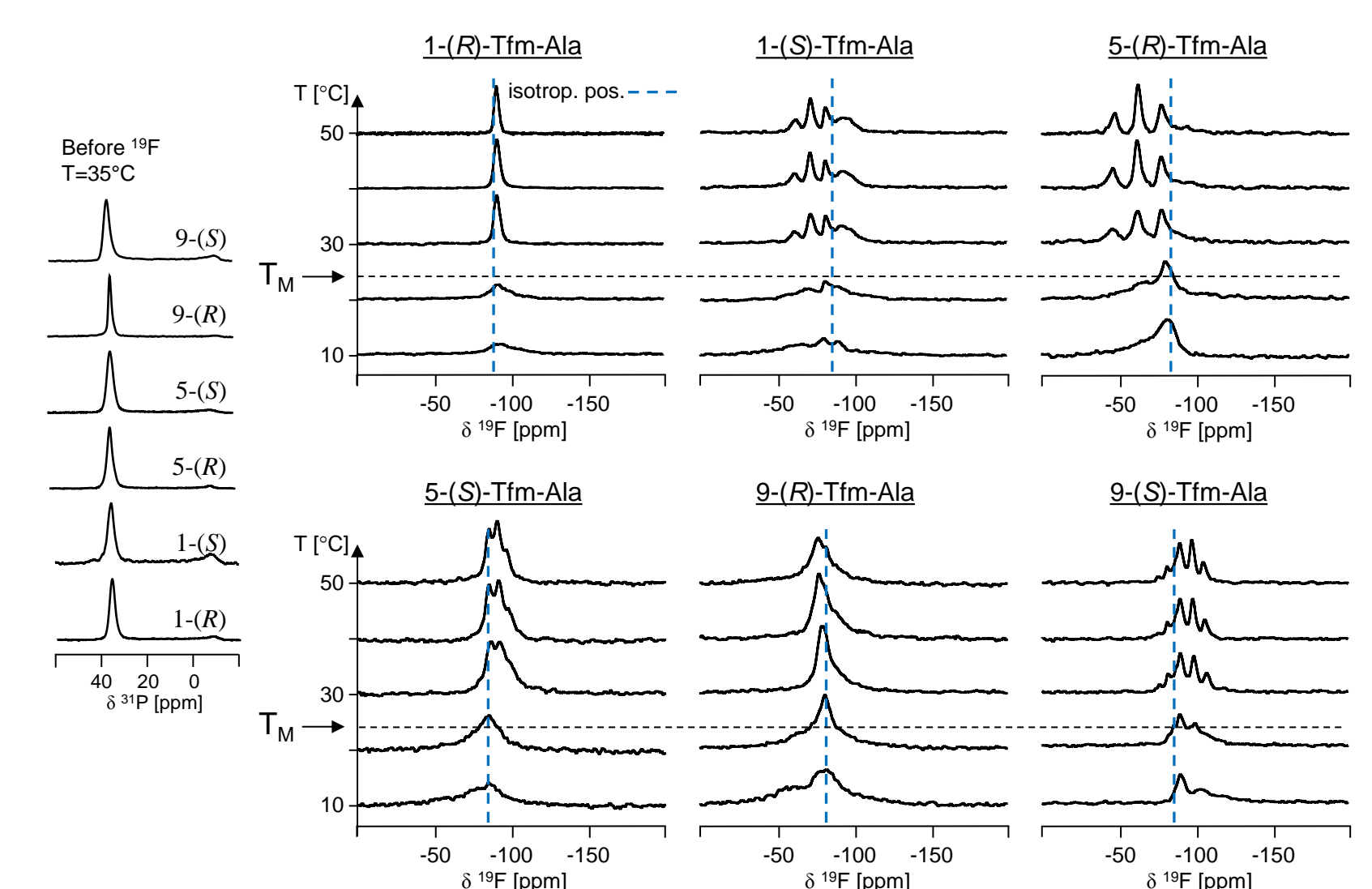
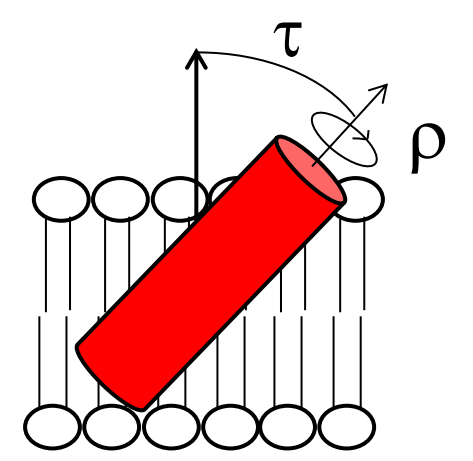
Sequence, labelling strategy and the labels:



Peptide	Sequence
HZ	Ac-U-N-I-I-U-P-L-L-U-P-L-ol
HZ 1-(R)-Tfm-Ala	Ac-(R)-Tfm-Ala-N-I-I-U-P-L-L-U-P-L-ol
HZ 1-(S)-Tfm-Ala	Ac-(S)-Tfm-Ala-N-I-I-U-P-L-L-U-P-L-ol
HZ 5-(R)-Tfm-Ala	Ac-U-N-I-I-(R)-Tfm-Ala-P-L-L-U-P-L-ol
HZ 5-(S)-Tfm-Ala	Ac-U-N-I-I-(S)-Tfm-Ala-P-L-L-U-P-L-ol
HZ 9-(R)-Tfm-Ala	Ac-U-N-I-I-U-P-L-L-(R)-Tfm-Ala-P-L-ol
HZ 9-(S)-Tfm-Ala	Ac-U-N-I-I-U-P-L-L-(S)-Tfm-Ala-P-L-ol
HZ 3-Tfm-Bpg	Ac-U-N-Tfm-Bpg-I-U-P-L-L-U-P-L-ol
HZ 4-Tfm-Bpg	Ac-U-N-I-Tfm-Bpg-U-P-L-L-U-P-L-ol
HZ 7-Tfm-Bpg	Ac-U-N-I-I-U-P-Tfm-Bpg-L-U-P-L-ol
HZ 8-Tfm-Bpg	Ac-U-N-I-I-U-P-L-Tfm-Bpg-U-P-L-ol

Synthetic peptides are reconstituted in oriented lipid bilayers. From the ¹⁹F-NMR dipolar couplings, the structure and membrane alignment of peptides is determined. [5]

Oriented CD with the synchrotron UV/VIS light source ANKA complements the results for the overall alignment of peptides.



³¹P- and ¹⁹F-NMR spectra of Tfm-Ala-labeled HZ analogues in oriented DMPC lipid bilayers (P/L 1/100) as a function of temperature.

Lipid	Dipolar splitting [kHz]					
	1-(R)	1-(S)	5-(R)	5-(S)	9-(R)	9-(S)
DMPC	0	4.9	7.4	-2.7	0	-4.0
DPhPC	-1.2	5.2	4.8	0	4.6	-4.0

CF₃ dipolar splitting values of Tfm-Ala labeled HZ peptides in DMPC and DPhPC lipid bilayers (P/L 1/100).

Structure model	Lipid	τ [°]	ρ [°]	S_{mol}	RMSD
3 ₁₀ -helix (ideal)	DMPC	126	172	0.4	2.05
	DPhPC	78	10	0.5	2.0
α -helix (ideal)	DMPC	92	116	0.5	2.23
	DPhPC	96	102	0.5	0.59
β -bend ribbon spiral [6]	DMPC	102	118	0.6	1.38
	DPhPC	124	106	0.4	0.87

Putative alignment (τ and ρ angles) and dynamics (S_{mol}) of HZ in DMPC and DPhPC lipid bilayers assuming different models for secondary structure.

Outlook

- Synthesis of ¹⁵N-labeled HZ peptides and purification of Tfm-Bpg-labeled HZ analogues to obtain more NMR constraints
- Investigation of the possibility of membrane thinning (²H-NMR, MD simulations)
- Structure analysis of free HZ by NMR in solution and/or X-ray crystallography

Acknowledgements

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