

Sezgin Kara¹, Sergii Afonin¹, Véronique Doan², Andrea Bordessa², Stephan L. Grage¹, Gregory Chaume², Norio Takeshita³, Thierry Brigaud², Anne S. Ulrich^{1,4}

¹ Institute of Biological Interfaces (IBG-2), Karlsruhe Institute of Technology; ² Laboratory SESCO - Université de Cergy-Pontoise; Institute of Applied Biosciences (IAB), Karlsruhe Institute of Technology; ⁴ Institute of Organic Chemistry (IOC), Karlsruhe Institute of Technology

contact: sezgin.kara@kit.edu

Introduction

Peptaibols

- Natural membrane-active peptides isolated from fungi
- Abundant in Aib (U) (α -aminoisobutyric acid), possess a C-terminal alcohol and N-terminal acetylation/alkylation
- Display wide range of antimicrobial activities
- Able to lyse lipid membranes by pore formation

Harzianin HK-VI (HZ wt) is an ultra-short peptaibol (11-mer) isolated from *T. pseudokoningii* [1] with the sequence:

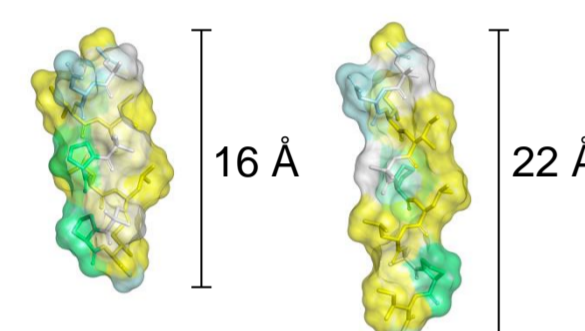
Ac-U-N-I-I-U-P-L-L-U-P-L-ol

Research aim

To solve the structure of membrane-bound HZ and get insights into its interactions with lipid bilayers.

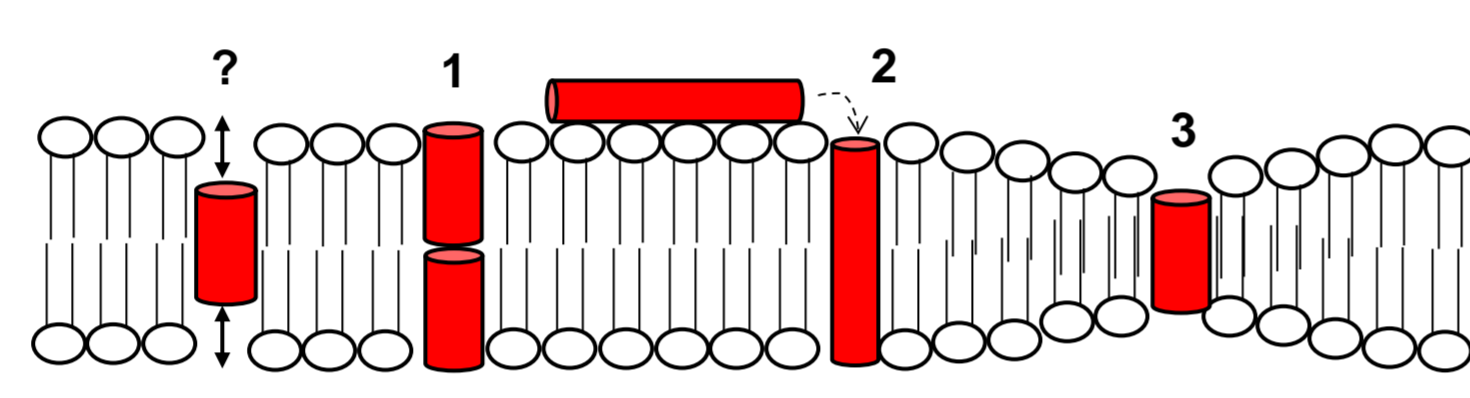
How can the short HZ span the bilayer with the expected α -helical structure?

HZ as α -helix and 3_{10} -helix



Inconsistency between peptide length and bilayer thickness.
D_b: bilayer thickness; 2D_c: thickness of hydrocarbon region
*T=30°C [2]

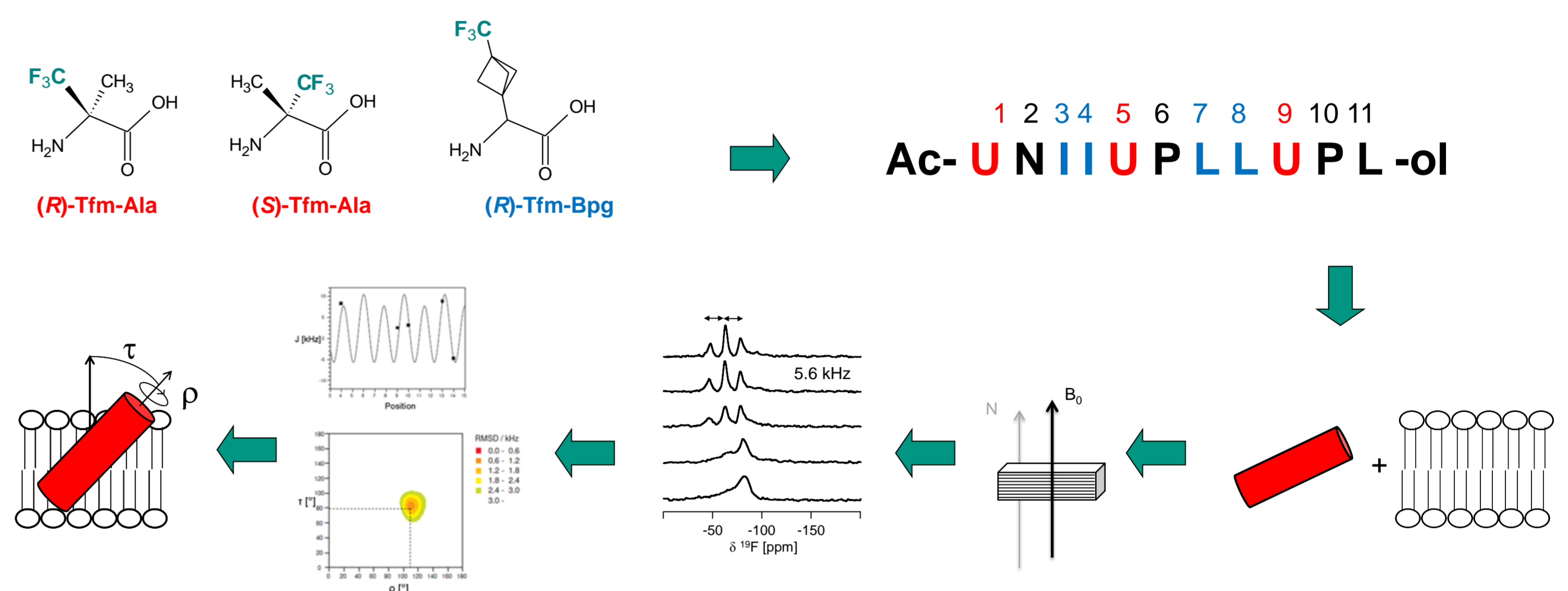
Lipid	D _b [Å]	2D _c [Å]
DLPC	32.6	21.7
DMPC	36.7	25.7
POPC	39.1	28.8
DPhPC	35.4	27.2



Suggested mechanisms of peptide/bilayer interactions for short pore-forming peptides: arranging as double-layered channels (1); conformational change to extended conformations; (2) membrane thinning (3).

Methods

- Solid-state ¹⁹F-NMR and synchrotron circular dichroism (SRCD) spectroscopy to determine the structure and alignment of HZ wt in lipid bilayers
- ¹⁹F-NMR is enabled by one-at-a-time incorporation of synthetic α -trifluoromethylated amino acids: (*R*)-, (*S*)-Tfm-Ala [3,4] and (*R*)-Tfm-Bpg [5]
- Synthetic peptides are reconstituted in mechanically aligned (oriented) lipid bilayers
- From the ¹⁹F-NMR dipolar couplings, the **structure, orientation** and **dynamics** is determined [6]



- Oriented SRCD from non-labelled peptide (HZ wt) with the synchrotron UV/VIS light source ANKA complements the results for the overall alignment of peptides

Results

Antimicrobial tests

Bacteria	MIC [μ g/ml]	
	HZ wt	HZ Tfm analogues
<i>E. coli</i> K12	> 256	> 256
<i>S. aureus</i> DSM 1104	128	> 256
<i>S. xylophilus</i> DSM 20267	> 256	> 256
<i>E. faecalis</i> DSM 2570	> 256	> 256
<i>B. subtilis</i> ATCC 6633	> 256	> 256
Fungi		
<i>A. nidulans</i> (GR5)	> 256	> 256
<i>C. tropicalis</i>	> 256	> 256
<i>M. oryzae</i> (Guy 11)	32	32
<i>T. harzianum</i>	> 256	> 256

Antimicrobial activity (MIC) of HZ wt and ¹⁹F-labeled analogues.

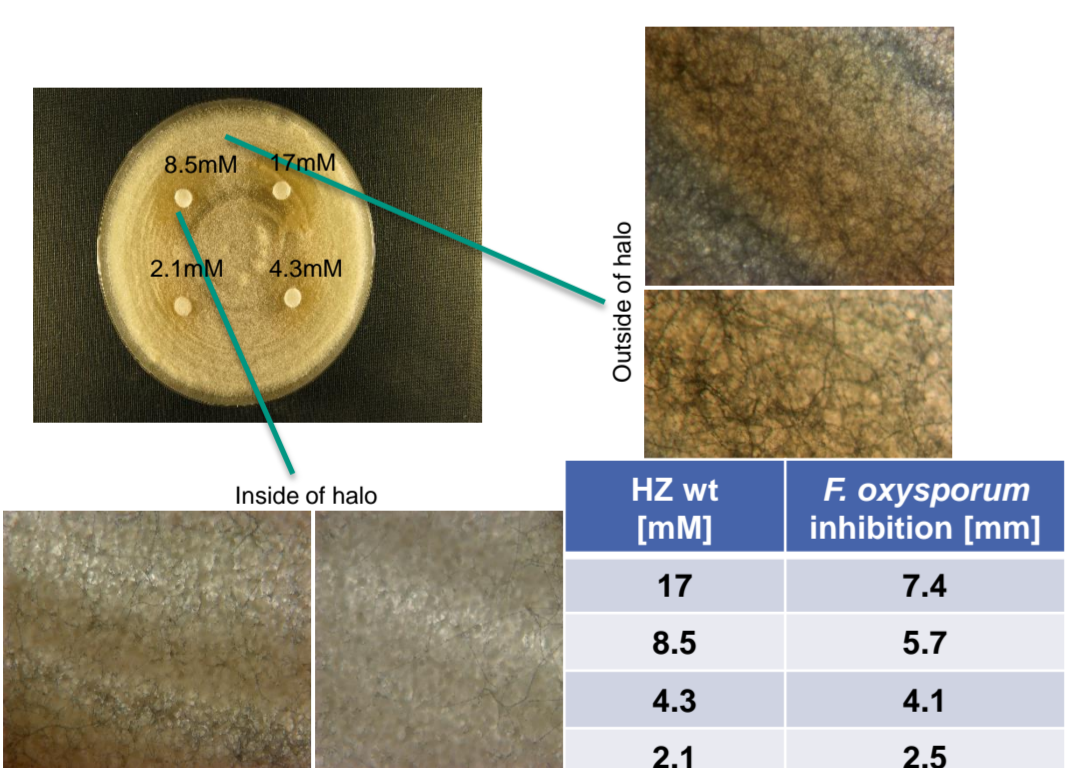
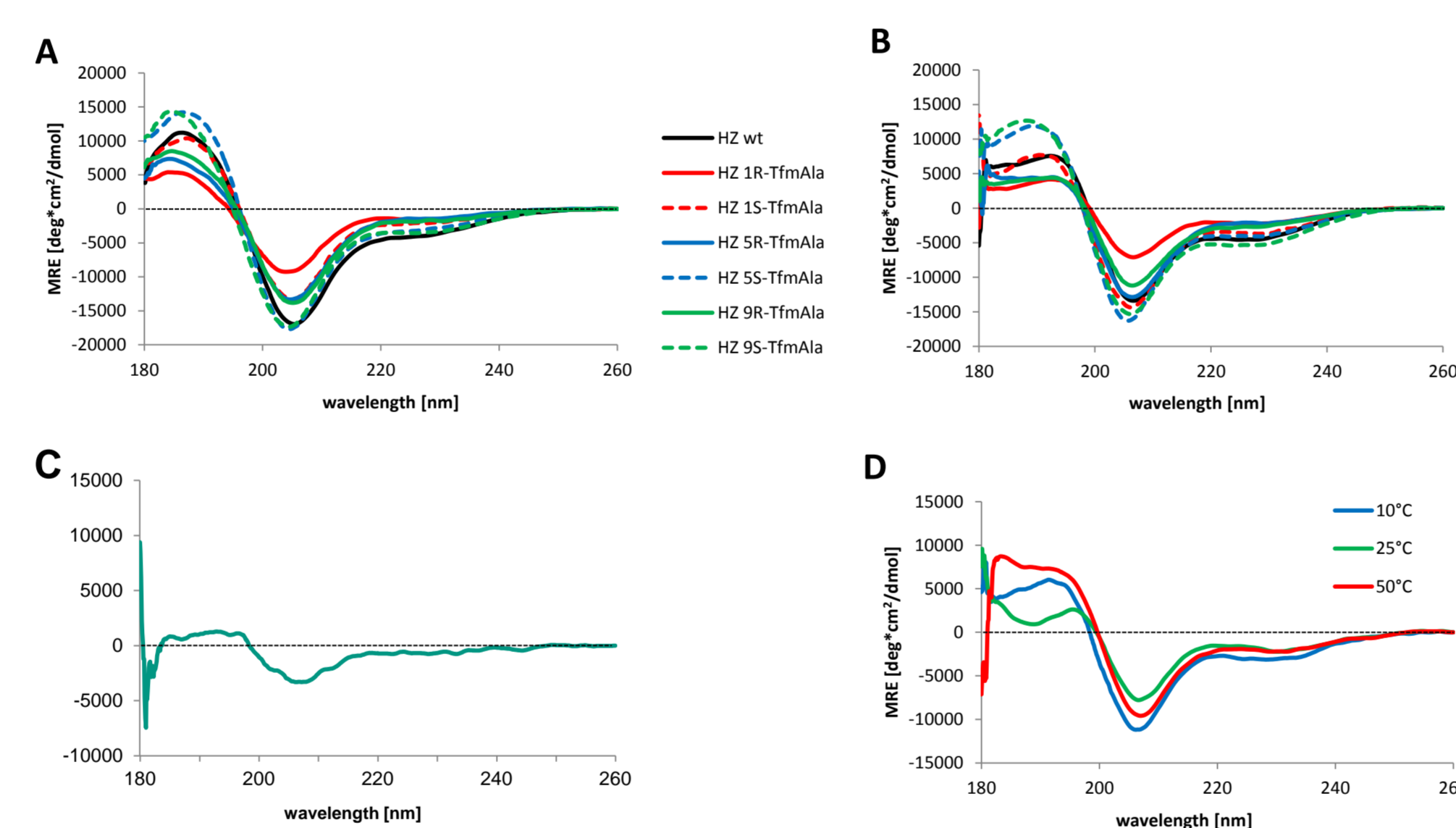


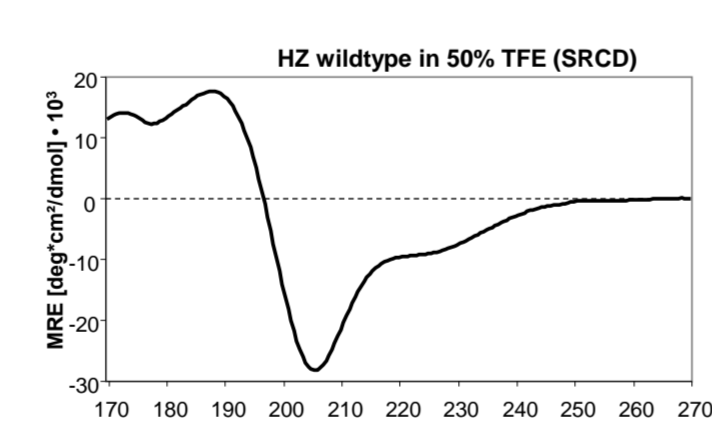
Plate diffusion assay testing HZ wt against *F. oxysporum*.

No pronounced antibacterial activity
Low antifungal effect against selected plant pathogenic fungi

Structure determination by CD



CD spectra of HZ peptides in organic solvent (TFE) (A); in detergent micelles (DPC, P/D 1/200) (B); in phospholipid liposomes (DPhPC) (C) and DMPC (D), P/L 1/20.

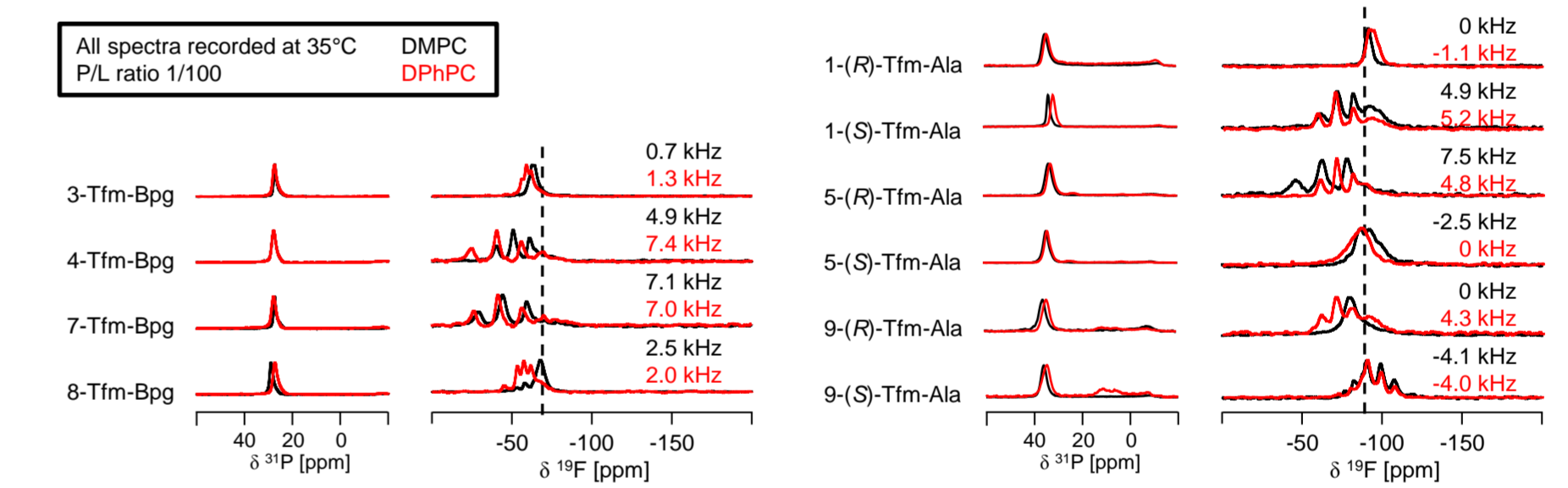


Structural analysis of the SRCD spectrum of HZ wt in 50% TFE.

Algorithm	α	3_{10}	β	turns	PPII	NRMSD
CONTIN	31.3	10.4	0	14	9.8	0.253
SELCON3	29.8	10.2	-2.3	13.5	8.8	0.132
CDSSTR	39	10	5	11	9	0.005

3_{10} -helical structure in various membrane models, whereas deconvolution suggests a predominant α -helical conformation

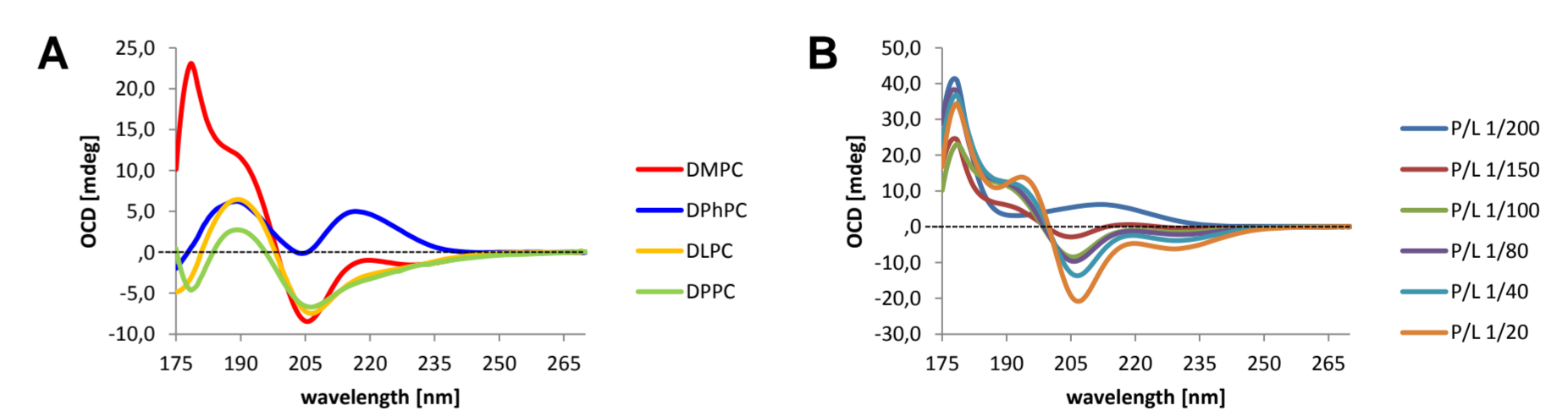
Peptide orientation by ssNMR and OCD



Solid-state ³¹P- and ¹⁹F-NMR spectra and observed ¹⁹F-NMR dipolar splittings of ¹⁹F-labeled HZ analogues in oriented DMPC and DPhPC bilayers; dotted line: isotropic position.

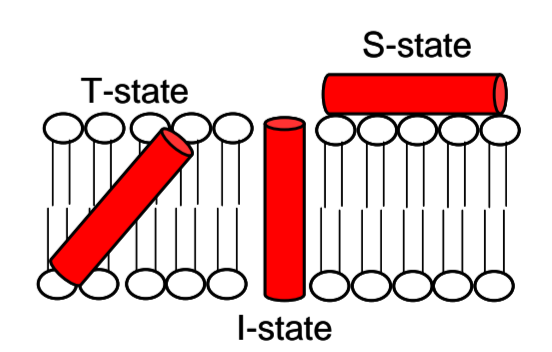
Structure model	Lipid	τ [°]	ρ [°]	S _{mol}	RMSD
3_{10} -helix (ideal)	DMPC	126	172	0.4	2.05
	DPhPC	78	10	0.5	2.0
α -helix (ideal)	DMPC	92	116	0.5	2.23
	DPhPC	96	102	0.5	0.59
β -bend ribbon spiral [7]	DMPC	102	118	0.6	1.38
	DPhPC	124	106	0.4	0.87

Putative alignment (τ and ρ angles) and dynamics (S_{mol}) of HZ in DMPC and DPhPC lipid bilayers assuming different models for secondary structure.



SR-OCD spectra of HZ wt in oriented phosphatidylcholine membranes of different composition (P/L 1/100) (A); HZ wt in oriented DMPC bilayers at varying PL (B).

DMPC: β -bend ribbon and S-state
DPhPC: α -helix and S-state
OCD suggests two different states



Outlook

- Structure determination of HZ wt by NMR in solution
- Synthesis of ¹⁵N-labeled HZ peptides to get more information on peptide alignment in lipid bilayers
- Analysis of membrane thinning (²H-NMR, MD simulations)
- Channel conductance measurements

Acknowledgements

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