Environmental Toxicology 2017 vol.32 N3, pages 989-1006

Species-specific differences in peroxisome proliferation, catalase, and SOD2 upregulation as well as toxicity in human, mouse, and rat hepatoma cells induced by the explosive and environmental pollutant 2,4,6trinitrotoluene

Naumenko E., Ahlemeyer B., Baumgart-Vogt E. Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

Abstract

© 2016 Wiley Periodicals, Inc. 2,4,6-Trinitrotoluene (TNT) has been widely used as an explosive substance and its toxicity is still of interest as it persisted in polluted areas. TNT is metabolized in hepatocytes which are prone to its toxicity. Since analysis of the human liver or hepatocytes is restricted due to ethical reasons, we investigated the effects of TNT on cell viability, reactive oxygen species (ROS) production, peroxisome proliferation, and antioxidative enzymes in human (HepG2), mouse (Hepa 1-6), and rat (H4IIEC3) hepatoma cell lines. Under control conditions, hepatoma cells of all three species were highly comparable exhibiting identical proliferation rates and distribution of their cell cycle phases. However, we found strong differences in TNT toxicity with the lowest IC 50 values (highest cell death rate) for rat cells, whereas human and mouse cells were three to sevenfold less sensitive. Moreover, a strong decrease in cellular dehydrogenase activity (MTT assay) and increased ROS levels were noted. TNT caused peroxisome proliferation with rat hepatoma cells being most responsive followed by those from mouse and human. Under control conditions, rat cells contained fivefold higher peroxisomal catalase and mitochondrial SOD2 activities and a twofold higher capacity to reduce MTT than human and mouse cells. TNT treatment caused an increase in catalase and SOD2 mRNA and protein levels in human and mouse, but not in rat cells. Similarly, human and mouse cells upregulated SOD2 activity, whereas rat cells failed therein. We conclude that TNT induced oxidative stress, peroxisome proliferation and mitochondrial damage which are highest in rat cells rendering them most susceptibl e toward TNT. © 2016 Wiley Periodicals, Inc. Environ Toxicol 32: 989-1006, 2017.

http://dx.doi.org/10.1002/tox.22299

Keywords

catalase, hepatoma cells, human, mouse, peroxisome proliferation, PEX14, rat, SOD2, superoxide radical anion, TNT

References

- [1] Ahlborg G Jr, Sorsa M. 1988. Mutagenic activity and metabolites in the urine of workers exposed to trinitrotoluene (TNT). Br J Ind Med 45:353–358.
- [2] Ahlemeyer B, Neubert I, Kovacs WJ, Baumgart-Vogt E. 2007. Differential expression of peroxisomal matrix and membrane proteins during postnatal development of mouse brain. J Comp Neurol 505:1–17.
- [3] Ahlemeyer B, Vogt JF, Michel V, Hahn-Kohlberger P, Baumgart-Vogt E. 2014. Microporation is an efficient method for siRNA-induced knockdown of PEX5 in HepG2 cells: Evaluation of the transfection efficiency, the PEX5 mRNA and protein levels and induction of peroxisomal deficiency. Histochem Cell Biol 142:577–591.
- [4] Ammerschläger M, Beigel J, Klein KU, Müller SO. 2004. Characterization of species specificity of peroxisome proliferation in rat and human hepatocytes. Toxicol Sci 78:229–240.
- [5] Arlt VM, Hewer A, Sorg BL, Schmeiser HH, Phillips DH, Stiborova M. 2004. 3-aminobenzanthrone, a human metabolite of the environmental pollutant 3-nitrobenzanthrone, forms DNA adducts after metabolic activation by human and rat liver microsomes: Evidence for activation by cytochrome P450 1A1 and P450 1A2. Chem Res Toxicol 17:1092–1101.
- [6] Bai J, Rodriguez AM, Melendez JA, Cederbaum AI. 1999. Overexpression of catalase in cytosolic or mitochondrial compartment protects HepG2 cells against oxidative injury. J Biol Chem 274:2617–2624.
- [7] Banerjee HN, Verma M, Hou LH, Ashraf M, Dutta SK. 1999. Cytotoxicity of TNT and its metabolites. J Biol Med 72:1–4.
- [8] Baumgart E. 1997. Application of in situ hybridization, cytochemical and immunocytochemical techniques for the investigation of peroxisomes. Histochem Cell Biol 108:185–210.
- [9] Berthe-Corti L, Jacobi H, Kleihauer S, Witte I. 1998. Cytotoxicity and mutagenicity of a 2,4,6-trinitrotoluene (TNT) and hexogen contaminated soil in S. typhimurium and mammalian cells. Chemosphere 37:209–218.
- [10] Berridge MV, Herst PM, Tan AS. 2005. Tetrazolium dyes as tools in cell biology: New insights into their cellular reduction. Biotechnol Annu Rev 11:127–152.
- [11] Bolt HM, Degen GH, Dorn SB, Plöttner S, Harth V. 2006. Genotoxicity and potential carcinogenicity of 2,4,6-TNT trinitrotoluene: Structural and toxicological considerations. Rev Environ Health 21:217–228.
- [12] Borbath I, Leclercq I, Moulin P, Sempoux C, Horsmans Y. 2007. The PPARgamma agonist pioglitazone inhibits early neoplastic occurrence in the rat liver. Eur J Cancer 43:1755–1763.
- [13] Bradford MM. 1976. A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein dye binding. Anal Biochem 72:248–254.
- [14] Brocard C, Es-Souni M, Ramirez LC, Latruffe N, Bournot P. 1993. Stimulation of peroxisomal palmitoyl-CoA oxidase activity by ciprofibrate in hepatic cell lines: Comparative studies in Fao, MH1C1 and HepG2 cells. Biol Cell 77:37-41.
- [15] Bueding E, Jolliffe N. 1946. Metabolism of trinitrotoluene (TNT) in vitro. J Pharmacol Exp Ther 88:300–312.
- [16] Butterworth BE, Earle LL, Strom S, Jirtle R, Michalopoulos G. 1983. Induction of DNA repair in human and rat hepatocytes by 1,6-dinitropyrene. Mutat Res 122:73–80.
- [17] Channon HJ, Mills GT, Williams RT. 1944. The metabolism of 2:4:6-trinitrotoluene (alpha-T.N.T.). Biochem J 38:70–85.
- [18] Chien CC, Kao CM, Chen DY, Chen SC, Chen CC. 2014. Biotransformation of trinitrotoluene (TNT) by Pseudomonas spp. isolated from a TNT-contaminated environment. Environ Toxicol Chem 33:1059–1063.
- [19] Cho YS, Lee BU, Kahng HY, Oh KH. 2009. Comparative analysis of 2,4,6-trinitrotoluene (TNT)-induced cellular responses and proteomes in Pseudomonas sp. HK-6 in two types of media. J Microbiol 4:220–224.
- [20] Colasante C, Chen J, Ahlemeyer B, Baumgart-Vogt E. 2015. Peroxisomes in cardiomyocytes and the peroxisome/peroxisome proliferator-activated receptor-loop. Thromb Haemost 113:425-436.
- [21] Cornu-Chagnon MC, Dupont H, Edgar A. 1995. Fenofibrate: Metabolism and species differences for peroxisome proliferation in cultured hepatocytes. Fundam Appl Toxicol 26:63–74.
- [22] Deng Y, Meyer SA, Guan X, Escalon BL, Ai J, Wilbanks MS, Welti R, Garcia-Reyero N, Perkins EJ. 2011. Analysis of common and specific mechanisms of liver function affected by nitrotoluene compounds. PLoS One 6:e14662.
- [23] Elcombe CR, Bell DR, Elias E, Hasmall SC, Plant NJ. 1986. Peroxisome proliferators: Species differences in response of primary hepatocyte cultures. Ann NY Acad Sci 804:628–635.
- [24] El-Naa NM, El-Refaei MF, Nasif WA, Abduljawad SH, El-Brairy AI, El-Readi MZ. 2015. In-vivo antioxidant and antiinflammatory activity of rosiglitazone, a peroxisome proliferator-activated receptor-gamma (PPAR-γ) agonists in animal model of bronchial asthma. J Pharm Pharmacol 67:1421-1430.
- [25] Esteve-Nunez A, Caballero A, Ramos JL. 2001. Biological degradation of 2,4,6-trinitrotoluene. Microbiol Mol Biol Rev 65:335-352.
- [26] Fahimi HD, Baumgart E, Völkl A. 1993. Ultrastructural aspects of the biogenesis of peroxisomes in rat liver. Biochimie 75:201–208.

- [27] Fahimi HD, Beier K, Lindauer M, Schad A, Zhan J, Pill J, Rebel W, Völkl A, Baumgart E. 1996. Zonal heterogeneity of peroxisome proliferation in rat liver. In: Reddy JK, Suga T, Mannaerts GP, Lazarow PB, Subramani S, editors. Peroxisomes. Biology and Role in Toxicology and Disease. New York, NY: New York Academy of Sciences. Vol. 804, pp 341–361.
- [28] Fransen M, Nordgren M, Wang B, Apanasets O. 2011. Role of peroxisomes in ROS/RNS metabolism: Implications for human disease. Biochim Biophys Acta 1822:1363-1373.
- [29] George SE, Kohan MJ, Warren SH. 1996. Hepatic DNA adducts and production of mutagenic urine in 2,6dinitrotoluene-treated B6C3F1 male mice. Cancer Lett 102:107-111.
- [30] Glass KY, Newsome CR, Tchounwou PB. 2005. Cytotoxicity and expression of c-fos, HSP70, and GADD45/153 proteins in human liver carcinoma (HepG2) cells exposed to dinitrotoluenes. Int J Environ Res Public Health 2:355–361.
- [31] Gonzalez FJ, Shah YM. 2008. PPARalpha: Mechanism of species differences and hepatocarcinogenesis of peroxisome proliferators. Toxicology 246:2–8.
- [32] Goodsell DS. 2004. Catalase. Molecule of the Month. RCSB Protein Data Bank.
- [33] Grant P, Ahlemeyer B, Karnati S, Berg T, Stelzig I, Nenicu A, Kuchelmeister K, Crane DI, Baumgart-Vogt E. 2013. The biogenesis protein PEX14 is an optimal marker for the identification and localization of peroxisomes in different cell types, tissues, and species in morphological studies. Histochem Cell Biol 140:423–442.
- [34] Homma-Takeda S, Hiraku Y, Ohkuma Y, Oikawa S, Murata M, Ogawa K, Iwamuro T, Li S, Sun GF, Kumagai Y, Shimojo N, Kawanishi N. 2002. 2,4,6-trinitrotoluene-induced reproductive toxicity via oxidative DNA damage by its metabolite. Free Radic Res 36:555–566.
- [35] Homma-Takeda S, Hiraku Y, Ohkuma Y, Oikawa S, Murata M, Ogawa K, Iwamuro T, Li S, Sun Joo HJ, Yim YH, Jeong PY, Jin YX, Lee JE, Kim H, Jeong SK, Chitwood DJ, Paik YK. 2009. Caenorhabditis elegans utilizes dauer pheromone biosynthesis to dispose of toxic peroxisomal fatty acids for cellular homoeostasis. Biochem J 422:61–71.
- [36] Immenschuh S, Baumgart-Vogt E. 2005. Peroxiredoxins, oxidative stress, and cell proliferation. Antioxid Redox Signal 7:768–777.
- [37] Islinger M, Li KW, Seitz J, Völkl A, Lüers GH. 2009. Hitchhiking of Cu/Zn superoxide dismutase to peroxisomes—Evidence for a natural piggyback import mechanism in mammals. Traffic 10:1711-1721.
- [38] Karnati S, Baumgart-Vogt E. 2008. Peroxisomes in mouse and human lung: Their involvement in pulmonary lipid metabolism. Histochem Cell Biol 130:719-740.
- [39] Kiria-Sakai M, Kumagai Y, Li S, Shimojo N. 1995. Generation of hydrogen peroxide during reduction of 2,4,6trinitrotoluene and its related compounds. Sangyo Eiseigaku Zasshi 41:11–12.
- [40] Kumagai Y, Kikushima M, Nakai Y, Shimojo N, Kunimoto M. 2004. Neuronal nitric oxide synthase (nNOS) catalyses one-electron reduction of 2,4,6-trinitoluene, resulting in decreased nitric oxide production and increased nNOS gene expression: Implication for oxidative stress. Free Rad Biol Med 37:350–357.
- [41] Lachance B, Robidoux PY, Hawari J, Ampleman G, Thiboutot S, Sunahara GI. 1999. Cytotoxic and genotoxic effects of energetic compounds on bacterial and mammalian cells in vitro. Mutat Res 444:25–39.
- [42] Lai DY. 2004. Rodent carcinogenicity of peroxisome proliferators and issues on human relevance. J Environ Sci Health C Environ Carcinog Ecotoxicol Rev 22:37-55.
- [43] Miliukiene V, Cenas N. 2008. Cytotoxicity of nitroaromatic explosives and their biodegradation products in mice splenocytes: Implications for their immunotoxicity. Z Naturforsch 63:519–525.
- [44] Mukherjee R, Jow L, Noonan D, McDonnell DP. 1994. Human and rat peroxisome proliferator activated receptors (PPARs) demonstrate similar tissue distribution but different responsiveness to PPAR activators. J Steroid Biochem Mol Biol 51:157–166.
- [45] Naumenko EA, Naumov AV, Suvorova ES, Gerlach R, Ziganshin AM, Lozhkin AP, Naumova RP. 2008. Participation of oxygen in the bacterial transformation of 2,4,6-trinitrotoluene. Biochemistry (Moscow) 73:463–469.
- [46] Naumenko EA, Sibgatullina GV, Mukhitov AR, Rodionov AA, Ilinskaya ON, Naumova RP. 2013. 2,4,6-Trinitrotoluene as a trigger for oxidative stress in Fagopyrum tatricum callus cells. Russ J Plant Physiol 60:404–410.
- [47] O'Brien ML, Spear BT, Glauert HP. 2005. Role of oxidative stress in peroxisome proliferator-mediated carcinogenesis. Crit Rev Toxicol 35:61–88.
- [48] Oswal DP, Balanarasimha M, Loyer JK, Bedi S, Soman FL, Rider SD Jr, Hostetler HA. 2013. Divergence between human and murine peroxisome proliferator-activated receptor alpha ligand specificities. J Lipid Res 54:2354–2365.
- [49] Reddy JK, Azarnoff DL, Hignite CE. 1980. Hypolipidaemic hepatic peroxisome proliferators form a novel class of chemical carcinogens. Nature 283:397-398.

- [50] Rodriguez C, Noe V, Cabrero A, Ciudad CJ, Laguna JC. 2000. Differences in the formation of PPARalpha-RXR/acoPPRE complexes between responsive and nonresponsive species upon fibrate administration. Mol Pharmacol 58:185–193.
- [51] Sarlauskas J, Nemeikaite-Ceniene A, Anusevicius Z, Miseviciene L, Julvez MM, Medina M, Gomez-Moreno C, Cenas N. 2004. Flavoenzyme-catalyzed redox cycling of hydroxylamino- and amino metabolites of 2,4,6trinitrotoluene: Implications for their cytotoxicity. Arch Biochem Biophys 425:184–192.
- [52] Schrader M, Wodopia R, Fahimi HD. 1999. Induction of tubular peroxisomes by UV irradiation and reactive oxygen species in HepG2 cells. Histochem Cytochem 47:1141–1148.
- [53] Schrader M, Fahimi HD. 2006. Peroxisomes and oxidative stress. Biochim Biophys Acta 1763:1755-1766.
- [54] Stangl H, Kovacs W, Böck P, Kremser K. 1995. Differential induction of peroxisomal enzymes by hypolipidaemics in human (HepG2) and rat (MH1C1) hepatoma cell lines. Eur J Clin Chem Clin Biochem 33:775–783.
- [55] Stenuit B, Lamblin G, Cornelis P, Agathos SN. 2012. Aerobic denitration of 2,4,6-trinitrotoluene in the presence of phenazine compounds and reduced pyridine nucleotides. Environ Sci Technol 46:10605–10613.
- [56] Stier H, Fahimi HD, Van Veldhoven P, Mannaerts GP, Völkl A, Baumgart E. 1998. Maturation of peroxisomes in differentiating human hepatoblastoma cells (HepG2): Possible involvement of the peroxisome proliferatoractivated receptor a (PPARa). Differentiation 64:55–66.
- [57] Taub J, Lau JF, Ma C, Hahn JH, Hoque R, Rothblatt J, Chalfie M. 1999. A cytosolic catalase is needed to extend adult lifespan in C. elegans daf-C and clk-1 mutants. Nature 399:162–166.
- [58] Tchounwou PB, Wilson BA, Ishaque AB, Schneider J. 2001. Transcriptional activation of stress genes and cytotoxicity in human liver carcinoma cells (HepG2) exposed to 2,4,6-trinitrotoluene, 2,4-dinitrotoluene, and 2,6-dinitrotoluene. Environ Toxicol 16:209–216.
- [59] Thiele S, Fernandes E, Bollag JM. 2002. Enzymatic transformation and binding of labeled 2,4,6-trinitrotoluene to humic substances during an anaerobic/aerobic incubation. J Environ Qual 31:437–444.
- [60] Tugwood JD, Aldridge TC, Lambe KG, Macdonald N, Woodyatt NJ. 1996. Peroxisome proliferator-activated receptors: Structures and function. Ann NY Acad Sci 804:252–265.
- [61] Woodyatt NJ, Lambe KG, Myers KA, Tugwood JD, Roberts RA. 1999. The peroxisome proliferator (PP) response element upstream of the human acyl CoA oxidase gene is inactive among a sample human population: Significance for species differences in response to PPs. Carcinogenesis 20:369–372.
- [62] Xiao Y, Karnati S, Qian G, Nenicu A, Fan W, Tchatalbachev S, Höland A, Hossain H, Guillou F, Lüers GH, Baumgart-Vogt E. 2012. Cre-mediated stress affects sirtuin expression levels, peroxisome biogenesis and metabolism, antioxidant and proinflammatory signaling pathways. PLoS One 7:e41097.
- [63] Yu J, Shen B, Chu ES, Teoh N, Cheung KF, Wu CW. 2010. Inhibitory role of peroxisome proliferator-activated receptor gamma in hepatocarcinogenesis in mice and in vitro. Hepatology 51:2008–2019.
- [64] Zaripov SA, Naumov AV, Suvorova ES, Garusov AV, Naumova RP. 2004. Initial stages of 2,4,6-trinitrotoluene transformation by microorganisms. Microbiology 73:398–403.
- [65] Zhang XC, Hu JP. 2008. FISSION1A and FISSION1B proteins mediate the fission of peroxisomes and mitochondria in Arabidopsis. Mol Plant 1:1036–1047.
- [66] Ziganshin AM, Gerlach R, Borch T, Naumov AV, Naumova RP. 2007. Production of eight different hydride complexes and nitrite release from 2,4,6-trinitrotoluene by Yarrowia lipolytica. Appl Environ Microbiol 73:7898–7905.
- [67] Ziganshin AM, Naumov AV, Suvorova ES, Naumenko EA, Naumova RP. 2010. Hydride-mediated reduction of 2,4,6-trinitrotoluene by yeasts as the way to its deep degradation. Mikrobiologica 76:766-773.
- [68] Ziganshin AM, Ziganshina EE, Byrne J, Gerlach R, Struve E, Biktagirov T, Rodionov A, Kappler A. 2015. Fe III mineral reduction followed by partial dissolution and reactive oxygen species generation during 2,4,6trinitrotoluene transformation by aerobic yeast Yarrowi lipolytica. AMB Express 5:8–20.