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Metabotropic and ionotropic glutamate receptors mediate the modulation of acetylcholine release at the frog neuromuscular junction

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Abstract

© 2016 Wiley Periodicals, Inc. There is some evidence that glutamate (Glu) acts as a signaling molecule at vertebrate neuromuscular junctions where acetylcholine (ACh) serves as a neurotransmitter. In this study, performed on the cutaneous pectoris muscle of the frog Rana ridibunda, Glu receptor mechanisms that modulate ACh release processes were analyzed. Electrophysiological experiments showed that Glu reduces both spontaneous and evoked guantal secretion of ACh and synchronizes its release in response to electrical stimulation. Quisqualate, an agonist of ionotropic α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic receptors and metabotropic Group I mGlu receptors, also exerted Glu-like inhibitory effects on the secretion of ACh but had no effect on the kinetics of quantal release. Quisqualate's inhibitory effect did not occur when a blocker of Group I mGlu receptors (LY 367385) or an inhibitor of phospholipase C (U73122) was present. An increase in the degree of synchrony of ACh quantal release, such as that produced by Glu, was obtained after application of N-methy--D-aspartic acid (NMDA). The presence of Group I mGlu and NMDA receptors in the neuromuscula r synapse was confirmed by immunocytochemistry. Thus, the data suggest that both metabotropic Group I mGlu receptors and ionotropic NMDA receptors are present at the neuromuscular synapse of amphibians, and that the activation of these receptors initiates different mechanisms for the regulation of ACh release from motor nerve terminals. © 2016 Wiley Periodicals, Inc.

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Keywords

cholinergic synapse, metabotropic Glu receptors, neuromodulation, neurotransmitter release, NMDA receptor

References

- Adámek S, Shakirzyanova AV, Malomouzh AI, Naumenko NV, Vyskočil F. 2010. Interaction of glutamate- and adenosine-induced decrease of acetylcholine quantal release at frog neuromuscular junction. Physiol Res 59:803–810.
- [2] Anderson MJ, Cohen MW. 1974. Fluorescent staining of acetylcholine receptors in vertebrate skeletal muscle. J Physiol 237:385–400.

- [3] Barthelemy-Requin M, Portalier P, Chamoin MC, Ternaux JP. 2000. Acetylcholine secretion enhanced by glutamate in rat embryonic spinal motoneurons: respective involvement of NMDA and AMPA receptors. Neurochem Res 25:377–384.
- [4] Berger UV, Carter RE, Coyle JT. 1995. The immunocytochemical localization of N-acetylaspartyl glutamate, its hydrolysing enzyme NAALADase, and the NMDAR-1 receptor at a vertebrate neuromuscular junction. Neuroscience 64:847–850.
- [5] Borodinsky LN, Spitzer NC. 2007. Activity-dependent neurotransmitter-receptor matching at the neuromuscular junction. Proc Natl Acad Sci U S A 104:335–340.
- [6] Boulland JL, Qureshi T, Seal RP, Rafiki A, Gundersen V, Bergersen LH, Fremeau RT Jr, Edwards RH, Storm-Mathisen J, Chaudhry FA. 2004. Expression of the vesicular glutamate transporters during development indicates the widespread corelease of multiple neurotransmitters. J Comp Neurol 480:264–280.
- [7] Bray JJ, Forrest JW, Hubbard JI. 1982. Evidence for the role of non-quantal acetylcholine in the maintenance of the membrane potential of rat skeletal muscle. J Physiol 326:285–296.
- [8] Bukcharaeva E, Kim K, Moravec J, Nikolsky E, Vyskocil F. 1999. Noradrenaline synchronizes evoked quantal release at frog neuromuscular junctions. J Physiol 517:879-888.
- [9] Burnstock G. 2004. Cotransmission. Curr Opin Pharmacol 4:47–52.
- [10] Burnstock G. 2009. Autonomic neurotransmission: 60 years since Sir Henry Dale. Annu Rev Pharmacol Toxicol 49:1–30.
- [11] Del Castillo J, Katz B. 1954. Quantal components of the end-plate potential. J Physiol 124:560-573.
- [12] Fu WM, Liou JC, Lee YH, Liou HC. 1995. Potentiation of neurotransmitter release by activation of presynaptic glutamate receptors at developing neuromuscular synapses of Xenopus. J Physiol 489:813–823.
- [13] Fu WM, Liou HC, Chen YH, Wang SM. 1998. Coexistence of glutamate and acetylcholine in the developing motoneurons. Chin J Physiol 41:127–132.
- [14] Giniatullin RA, Khazipov RN, Oranska TI, Nikolsky EE, Voronin VA, Vyskocil F. 1993. The effect of non-quantal acetylcholine release on quantal miniature currents at mouse diaphragm. J Physiol 466:105-114.
- [15] Gutierrez R. 2009. Co-existence and co-release of classical neurotransmitters. New York (NY): Springer.
- [16] Herzog E, Landry M, Buhler E, Bouali-Benazzouz R, Legay C, Henderson CE, Nagy F, Dreyfus P, Giros B, El Mestikawy S. 2004. Expression of vesicular glutamate transporters, VGLUT1 and VGLUT2, in cholinergic spinal motoneurons. Eur J Neurosci 20:1752–1760.
- [17] Hnasko TS, Edwards RH. 2012. Neurotransmitter corelease: mechanism and physiological role. Annu Rev Physiol. 74:225–243.
- [18] Israël M, Lesbats B, Bruner J. 1993. Glutamate and acetylcholine release from cholinergic nerve terminals, a calcium control of the specificity of the release mechanism. Neurochem Int 22:53–58.
- [19] Katz B, Miledi R. 1965. The measurement of synaptic delay, and the time course of acetylcholine release at the neuromuscular junction. Proc R Soc Lond B Biol Sci 161:483–495.
- [20] Kerkut GA, Shapira A, Walker RJ. 1967. The transport of 14C-labelled material from CNS to and from muscle along a nerve trunk. Comp Biochem Physiol 23:729-748.
- [21] Krause M, Wernig A. 1985. The distribution of acetylcholine receptors in the normal and denervated neuromuscular junction of the frog. J Neurocytol 14:765–780.
- [22] Kupfermann I. 1991. Functional studies of cotransmission. Physiol Rev 71:683-732.
- [23] Lin JW, Faber DS. 2002. Modulation of synaptic delay during synaptic plasticity. Trends Neurosci 25:449-455.
- [24] Liou HC, Yang RS, Fu WM. 1996. Potentiation of spontaneous acetylcholine release from motor nerve terminals by glutamate in Xenopus tadpoles. Neuroscience 75:325–331.
- [25] Malomouzh Al. 2012. Non-cholinergic signaling pathways at vertebrate neuromuscular junctions. In: Cseri J, editor. Skeletal muscle—from myogenesis to clinical relations. Rijeka (Croatia): InTech. p 143–164.
- [26] Malomouzh AI, Mukhtarov MR, Nikolsky EE, Vyskocil F, Lieberman EM, Urazaev AK. 2003. Glutamate regulation of non-quantal release of acetylcholine in the rat neuromuscular junction. J Neurochem 85:206–213.
- [27] Malomouzh AI, Nikolsky EE, Lieberman EM, Sherman JA, Lubischer JL, Grossfeld RM, Urazaev AKh. 2005. Effect of N-acetylaspartylglutamate (NAAG) on non-quantal and spontaneous quantal release of acetylcholine at the neuromuscular synapse of rat. J Neurochem. 94:257–267.
- [28] Malomouzh AI, Nurullin LF, Arkhipova SS, Nikolsky EE. 2011. NMDA receptors at the endplate of rat skeletal muscles: precise postsynaptic localization. Muscle Nerve 44:987-989.
- [29] Mays TA, Sanford JL, Hanada T, Chishti AH, Rafael-Fortney JA. 2009. Glutamate receptors localize postsynaptically at neuromuscular junctions in mice. Muscle Nerve 39:343-349.
- [30] Neale JH, Bzdega T, Wroblewska B. 2000. N-acetylaspartylglutamate: the most abundant peptide neurotransmitter in the mammalian central nervous system. J Neurochem 75:443–452.
- [31] Nikolsky EE, Zemkova H, Voronin VA, Vyskocil F. 1994. Role of non-quantal acetylcholine release in surplus polarization of mouse diaphragm fibres at the endplate zone. J Physiol 477:497–502.

- [32] Nikolsky EE, Oranska TI, Vyskocil F. 1996. Non-quantal acetylcholine release in the mouse diaphragm after phrenic nerve crush and during recovery. Exp Physiol 81:341–348.
- [33] Nikolsky E, Vyskocil F, Bukharaeva E, Samigullin D, Magazanik L. 2004. Cholinergic regulation of the quantal release at frog neuromuscular junction. J Physiol 560:77–88.
- [34] Nishimaru H, Restrepo CE, Ryge J, Yanagawa Y, Kiehn O. 2005. Mammalian motor neurons corelease glutamate and acetylcholine at central synapses. Proc Natl Acad Sci U S A 102:5245-5249.
- [35] Niswender CM, Conn PJ. 2010. Metabotropic glutamate receptors: physiology, pharmacology, and disease. Annu Rev Pharmacol Toxicol 50:295–322.
- [36] Petrov KA, Malomouzh AI, Kovyazina IV, Krejci E, Nikitashina AD, Proskurina SE, Zobov VV, Nikolsky EE. 2013. Regulation of acetylcholinesterase activity by nitric oxide in rat neuromuscular junction via N-methyl-D-aspartate receptor activation. Eur J Neurosci 37:181–189.
- [37] Pinard A, Robitaille R. 2008. Nitric oxide dependence of glutamate-mediated modulation at a vertebrate neuromuscular junction. Eur J Neurosci 28:577–587.
- [38] Pinard A, Lévesque S, Vallée J, Robitaille R. 2003. Glutamatergic modulation of synaptic plasticity at a PNS vertebrate cholinergic synapse. Eur J Neurosci 18:3241–3250.
- [39] Rinholm JE, Slettaløkken G, Marcaggi P, Skare Ø, Storm-Mathisen J, Bergersen LH. 2007. Subcellular localization of the glutamate transporters GLAST and GLT at the neuromuscular junction in rodents. Neuroscience 145:579–591.
- [40] Sun Y-a, Poo M-m. 1985. Non-quantal release of acetylcholine at a developing neuromuscular synapse in culture. J Neurosci 5:634-642.
- [41] Thesleff S. 1990. Functional aspects of quantal and non-quantal release of acetylcholine at the neuromuscular junction. Prog Brain Res 84:93–99.
- [42] Tsentsevitsky A, Nikolsky E, Giniatullin R, Bukharaeva E. 2011. Opposite modulation of time course of quantal release in two parts of the same synapse by reactive oxygen species. Neuroscience 189:93-99.
- [43] Urazaev A, Naumenko N, Malomough A, Nikolsky E, Vyskocil F. 2000. Carbachol and acetylcholine delay the early postdenervation depolarization of muscle fibres through M1-cholinergic receptors. Neurosci Res 37:255–263.
- [44] Vyas S, Bradford HF. 1987. Co-release of acetylcholine, glutamate and taurine from synaptosomes of Torpedo electric organ. Neurosci Lett 82:58–64.
- [45] Vyskocil F, Vrbova G. 1993. Non-quantal release of acetylcholine affects polyneuronal innervation on developing rat muscle fibres. Eur J Neurosci 5:1677–1683.
- [46] Waerhaug O, Ottersen OP. 1993. Demonstration of glutamate-like immunoreactivity at rat neuromuscular junctions by quantitative electron microscopic immunocytochemistry. Anat Embryol (Berl) 188:501–513.
- [47] Walder KK, Ryan SB, Bzdega T, Olszewski RT, Neale JH, Lindgren CA. 2013. Immunohistological and electrophysiological evidence that N-acetylaspartylglutamate is a co-transmitter at the vertebrate neuromuscular junction. Eur J Neurosci 37:118-129.
- [48] Willard SS, Koochekpour S. 2013. Glutamate, glutamate receptors, and downstream signaling pathways. Int J Biol Sci 9:948–959.