



Phase transition, ordering and lateral diffusion in phospholipid bilayers in the presence of poly(ethylene oxide)

Andrey Filippov,^{*a,b} Bulat Munavirov^b and Oleg N. Antzutkin^{a,c}^a Institute of Physics, Kazan (Volga Region) Federal University, 420008 Kazan, Russian Federation.Fax: +7 843 231 5189; e-mail: Andrey.Filippov@ksu.ru^b Chemistry of Interfaces, Luleå University of Technology, SE-97187 Luleå, Sweden^c Department of Physics, Warwick University, CV4 7AL Coventry, UK

DOI: 10.1016/j.mencom.2012.09.007

The thermal behaviour, molecular orientation and lateral diffusion in the bilayered systems of dimyristoylphosphatidylcholine (DMPC) in the presence of poly(ethylene oxide) (PEO) were studied by NMR and DSC techniques, and it was found that PEO decreases the melting temperature (of vesicles and flat multibilayers) and affects the degree of orientation of DMPC molecules relative to the bilayer normal, but it does not influence the lateral diffusion of DMPC molecules.

Poly(ethylene oxides) (PEOs) are biomembrane-active agents frequently used in pharmacy, medicine and molecular biology.^{1,2} PEOs are highly soluble in water, and they have low toxicity and low immunogenicity. Therefore, the mechanisms by which PEO molecules interact with phospholipids is of interest. In this work, we studied alterations in phospholipid bilayer properties, such as the phase state and melting behaviour of the lipid system, lipid molecule orientation and lateral diffusion in the membrane that were induced by the PEO–lipid interaction.

Commercial phospholipid dimyristoylphosphatidylcholine (DMPC) from Avanti Polar Lipids and PEO (MW 6000) from Sigma-Aldrich were used. For differential scanning calorimetry (DSC) measurements, DMPC was hydrated to achieve a concentration of 2 mM unilamellar vesicles, as described previously.³ DSC thermograms were generated by heating and cooling from 5–45 °C with a rate of 10 K h⁻¹. Four temperature scans were performed, where the first scan was discarded to obtain a common thermal history of all samples. The measurements were performed on a MicroCal VP-DSC Microcalorimeter (Täby, Sweden). The macroscopically oriented multibilayers of DMPC were prepared according to a previously described procedure.^{3,4} The PEO concentration in the water phase of the oriented bilayers was ~2 wt%. A Chemagnetic InfinityPlus NMR spectrometer (Agilent) operating at proton frequencies of 360 MHz was used. The ³¹P NMR spectra were recorded at 145.703 MHz using single pulse excitation. The ¹H NMR diffusion measurements were performed at 359.92 MHz using the stimulated echo pulsed field gradient procedure. A square sample tube containing macroscopically oriented lipid multibilayers was placed in a specifically designed goniometer probe, which enabled the bilayers to be oriented with the bilayer normal at the magic angle (54.7°) with respect to the constant magnetic field of the spectrometer. This causes the dipolar interactions to vanish, resulting in a significant reduction of the line width. The NMR PFG method for measuring lipid lateral diffusion on macroscopically oriented bilayers was described elsewhere.⁵ For all measurements, we applied the stimulated echo pulse sequence.⁶ The echo amplitude diffusion decay (DD) in the case of a one-component, non-associated liquid is the dependence of the echo amplitude (*A*) on the pulse sequence parameters (δ , *g* and t_d) and the self-diffusion coefficient of molecules (*D*):

$$I = I_0 \exp(-2\tau_2/T_2) \exp(-\tau_1/T_1) \exp[-\gamma^2 g^2 \delta^2 D(\Delta - \delta/3)], \quad (1)$$

where I_0 is the factor proportional to the proton content of the system; T_1 and T_2 are spin–lattice and spin–spin relaxation times, respectively; τ_2 and τ_1 are time intervals in the pulse sequence; γ is the gyromagnetic ratio for protons; *g* and δ are the amplitude and duration of the gradient pulse, respectively; $t_d = (\Delta - \delta/3)$ is the diffusion time; $\Delta = (\tau_2 + \tau_1)$ is the time interval between the two gradient pulses. In our experiments, $g = 1.15 \text{ T m}^{-1}$ was set constant and δ was varied in the range of 1.1–8.2 ms, $\tau_2 = 11 \text{ ms}$ and $\tau_1 = 100 \text{ ms}$. We Fourier transformed the experimental echoes into corresponding sets of spectra and calculated the diffusion coefficient from a non-linear fit of the obtained signal decay. For bilayers oriented at the magic angle, the lateral diffusion coefficient

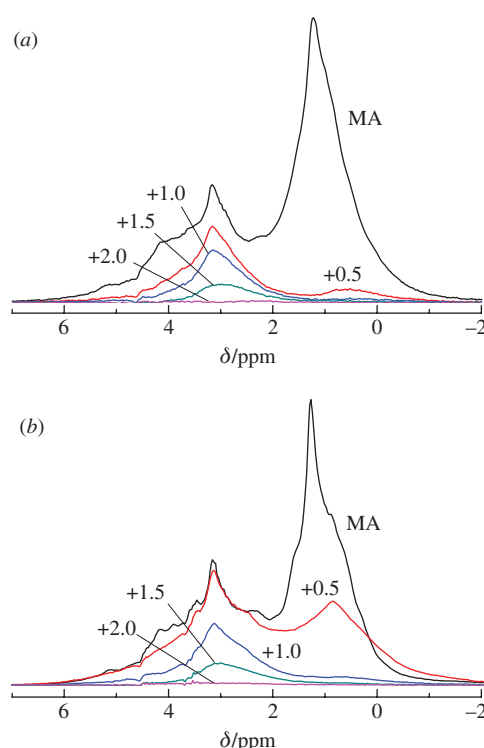


Figure 1 ¹H NMR spectra obtained after Fourier transformation of the descending half of stimulated echo of DMPC bilayers (a) without and (b) with PEO, oriented at the magic angle (black) and at angles subsequently deviating from magic angle by 0.5°. PEO/DMPC weight ratio, 0.02. Temperature, 30 °C.