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In Vitro Activities of Nine Antifungal Drugs against 81 Phialophora and Cyphellophora Isolates

Peiying Feng,^{a,b} M. Javad Najafzadeh,^{b,c,d} Jiufeng Sun,^{b,e} Sarah Ahmed,^b Liyan Xi,^f G. Sybren de Hoog,^{b,d,f,g} Wei Lai,^a Chun Lu,^a Corné H. Klaassen,ⁱ and Jacques F. Meis^{h,i}

Third Affiliated Hospital, Sun Yat-sen University, Guangzhou, China^a, CBS-KNAW Fungal Biodiversity Centre, Utrecht, The Netherlands^b; Department of Parasitology and Mycology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran^c; Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, Amsterdam, The Netherlands^d; Key Laboratory of Tropical Disease Control, Department of Parasitology, Zhongshan School of Medicine, Sun Yat-sen University, Guangzhou, China^e; Sun Yat-sen Memorial Hospital, Sun Yat-sen University, Guangzhou, China^e; Peking University Health Science Center, Research Center for Medical Mycology, Beijing, China^e; Department of Medical Microbiology, Radboud University Nijmegen Medical Center, Nijmegen, The Netherlands^h; and Department of Medical Microbiology and Infectious Diseases, Canisius Wilhelmina Hospital, Nijmegen, The Netherlandsⁱ

Cyphellophora guyanensis (n = 15), other *Cyphellophora* species (n = 11), *Phialophora europaea* (n = 43), and other *Phialophora* species (n = 12) were tested *in vitro* against nine antifungal drugs. The MIC₉₀s across all of the strains (n = 81) were, in increasing order, as follows: posaconazole, 0.063 µg/ml; itraconazole, 0.5 µg/ml; voriconazole, 1 µg/ml; micafungin, 1 µg/ml; terbinafine, 2 µg/ml; isavuconazole, 4 µg/ml; caspofungin, 4 µg/ml; fluconazole, 8 µg/ml; amphotericin B, 16 µg/ml.

lthough dermatophytes and yeasts account for the majority of superficial and cutaneous fungal infections, members of the black yeast-like fungi are reported regularly (2, 18). Humid indoor environments such as bathrooms and swimming pools (14, 15) are potential reservoirs for black yeast infections. Cyphellophora and Phialophora species are black yeast-like fungi that form a phylogenetic "europaea clade" within the order Chaetothyriales (9) and are recovered from human skin and nails (8, 18). Phialophora europaea was found in 27% of positive cultures with black yeasts (18) which are found simultaneously with dermatophytes on the skin of diabetic patients (11). The taxonomy of the genus Cyphellophora has recently been revised, and 11 independent species were recognized in the "europaea clade," i.e., Cyphellophora laciniata, C. vermispora, C. pluriseptata, C. suttonii, C. fusarioides, C. pauciseptata, C. guyanensis, Phialophora ambigua, P. oxyspora, P. reptans, and P. europaea (9, 12). Cyphellophora laciniata, C. pluriseptata (5, 10), C. pauciseptata, and P. ambigua thus far have been isolated exclusively from superficial lesions in human infections (7, 9, 12), while C. fusarioides originated from bronchoalveolar lavage fluid from a patient after heart bypass surgery (19). Cyphellophora suttonii was originally isolated from a phaeohyphomycotic lesion in a dog's ear (1) but also from ulcerating skin lesions in a patient with sarcoidosis (17). At present, there is no information available on the antifungal susceptibility profiles of Cyphellophora and its relatives. This study aimed to determine the in vitro susceptibilities of a large collection of clinical and environmental isolates of Cyphellophora and Phialophora species belonging to the black yeast "europaea clade" to eight antifungal drugs and isavuconazole (13, 20, 21), a new triazole that currently is undergoing phase III clinical trials.

A total of 81 strains were obtained from the Centraalbureau voor Schimmelcultures (CBS; Utrecht, The Netherlands) Fungal Biodiversity Centre, which included *Cyphellophora guyanensis* (n = 15), *C. laciniata* (n = 3), *C. pauciseptata* (n = 1), *C. pluriseptata* (n = 3), *Cyphellophora suttonii* (n = 1), *C. vermispora* (n = 3), *Phialophora ambigua* (n = 1), *P. europaea* (n = 43), *P. oxyspora* (n = 3), and *P. reptans* (n = 8) (see Table S1 in the supplemental material). The strains were obtained from human (n = 55) and animal (n = 1) clinical samples, environmental samples (n = 12),

plant materials (n = 11), and unknown sources (n = 2). The set included all available ex-type strains of the species described and was supplemented with newly isolated strains. In addition to genotyping, species identity was confirmed by molecular characterization of the internal transcribed spacer region, the partial DNA-dependent RNA polymerase II largest-subunit gene, the beta-tubulin gene, and the nuclear large-subunit rRNA gene (9, 12). Antifungal susceptibility testing was performed as described in CLSI document M38-A2, with some modifications (6). Briefly, isolates were cultured on potato dextrose agar in the dark (25°C) for up to 7 days to induce sporulation. Inocula were prepared by scraping the surface of the fungal colonies with a cotton swab moistened with sterile physiological saline containing 0.05% Tween 40. Large particles were allowed to settle for 5 min, and then a suspension of spores was adjusted with a spectrophotometer (Spectronic 20D; Milton Roy, Rochester, NY) to 68 to 71% transmission (at 530 nm) and diluted 10-fold to yield a final inoculum of 1.5×10^4 to 5×10^4 CFU/ml. The test concentrations of amphotericin B (Bristol-Myers Squibb, Woerden, The Netherlands), itraconazole (Janssen Research Foundation, Beerse, Belgium), voriconazole (Pfizer Central Research, Sandwich, United Kingdom), posaconazole (Schering-Plough, Kenilworth, NJ), isavuconazole (Basilea Pharmaceutica International AG, Basel, Switzerland), and terbinafine (Novartis Pharma, Basel, Switzerland) ranged from 0.016 to 16 µg/ml; those of fluconazole (Pfizer) ranged from 0.063 to 64 µg/ml; and those of caspofungin (MSD, Haarlem, The Netherlands) and micafungin (Astellas Pharma, Ibaraki, Japan) ranged from 0.008 to 8 µg/ml. After 72 h of incubation at 25°C, MICs and minimum effective concentrations

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Address correspondence to Jacques F. Meis, j.meis@cwz.nl.

Supplemental material for this article may be found at http://aac.asm.org/. Copyright © 2012, American Society for Microbiology. All Rights Reserved. doi:10.1128/AAC.01112-12 TABLE 1 Geometric mean MICs, MIC ranges, $MIC_{50}s$, and $MIC_{90}s$ obtained by susceptibility testing of antimycotic agents against *Cyphellophora* and relatives

Organism (no. of strains/genotype ^{<i>a</i>}) and drug	MIC/MEC (µg/ml)				
	Range	50%	90%	Geometric mean	
Total $(n = 81)$					
Amphotericin B	0.125-16	4	16	4	
Fluconazole	2-16	8	8	27.655	
Itraconazole	≤0.016-1	0.125	0.5	0.097	
Voriconazole	0.063-2	0.25	1	0.323	
Posaconazole	<0.016-0.125	<0.016	0.063	0.027	
Isavuconazole	0.25-4	1	4	1 928	
Caspofungin	0.25_8	1	1	1.116	
Micofungin	< 0.008 4	0.125	1	0.07	
Tarbinatina	$\leq 0.006 - 4$	0.125	1	0.07	
Terbinanne	≥0.010-4	0.5	2	0.541	
Cyphellophora guvanensis ($n = 15$ /group C4)					
Amphotericin B	1–16	16	16	5.879	
Fluconazole	4-16	8	16	23 516	
Itraconazole	0.063-1	0.25	0.5	0.25	
Voriconazola	0.25 1	0.25	1	0.25	
Desesser	0.23 - 1	0.062	1 0.125	0.5	
Posaconazole	≤0.016-0.125	0.065	0.125	0.05	
Isavuconazole	0.25–4	2	4	3.703	
Caspotungin	1-4	1	2	1.361	
Micatungin	$\leq 0.008 - 1$	0.031	0.25	0.037	
Terbinafine	0.125-1	0.25	0.5	1.167	
Cyphellophora laciniata ($n = 3/\text{group A1}$)					
Amphotericin B	1-4	NC^b	NC	1.587	
Fluconazole	4-16	NC	NC	25 398	
Itraconazole	0.063-0.125	NC	NC	0.079	
Voriconazole	0.25 1	NC	NC	0.63	
Desegementele	0.23 - 1	NC	NC	0.03	
Posaconazole	≤0.010-0.031	NC	NC	0.02	
Isavuconazole	1-2	NC	NC	5.1/5	
Caspotungin	1-4	NC	NC	1.58/	
Micatungin	≤0.008-≤0.008	NC	NC	0.008	
Terbinafine	0.125-0.125	NC	NC	0.5	
Cyphellophora pauciseptata ($n = 1$ /group A6)					
Amphotericin B	0.5	NC	NC	NC	
Fluconazole	32	NC	NC	NC	
Itraconazole	0.125	NC	NC	NC	
Voriconazole	0.5	NC	NC	NC	
Posaconazola	0.031	NC	NC	NC	
Iouruconazola	2	NC	NC	NC	
Savuconazole	2	NC	NC	NC	
Casporungin	0.5	NC	NC	NC	
Micatungin	0.031	NC	NC	NC	
Terbinafine	0.063	NC	NC	NC	
<i>Cyphellophora pluriseptata</i> ($n = 3$ /groups C1, C2)					
Amphotericin B	4-8	NC	NC	6.35	
Fluconazole	4-16	NC	NC	40.317	
Itraconazole	0.125-0.25	NC	NC	0.157	
Voriconazole	0.25-0.5	NC	NC	0.315	
Posaconazole	0.031-0.063	NC	NC	0.039	
Isavuconazole	1_2	NC	NC	2.52	
Compfungin	2.9	NC	NC	1	
Missfurgin	2-0	NC	NC	4	
	≤0.008-0.016	NC	NC	0.01	
Terbinafine	≤0.016-0.25	NC	NC	0.316	
<i>Cyphellophora suttonii</i> ($n = 1$ /group A4)					
Amphotericin B	2	NC	NC	NC	
Fluconazole	32	NC	NC	NC	
Itraconazole	0.25	NC	NC	NC	
Voriconazole	0.5	NC	NC	NC	

(Continued on following page)

TABLE 1 (Continued)

Organism (no. of strains/genotype ^{<i>a</i>}) and drug	MIC/MEC (µg/ml)	MIC/MEC (µg/ml)				
	Range	50%	90%	Geometric mean		
Posaconazole	0.063	NC	NC	NC		
Isavuconazole	8	NC	NC	NC		
Caspofungin	1	NC	NC	NC		
Micafungin	0.25	NC	NC	NC		
Terbinafine	2	NC	NC	NC		
<i>Cyphellophora vermispora</i> ($n = 3$ /group A2)						
Amphotericin B	0.5-1	NC	NC	0.707		
Fluconazole	8-8	NC	NC	32		
Itraconazole	0.063-0.5	NC	NC	0.177		
Voriconazole	0.5–2	NC	NC	1		
Posaconazole	0.031-0.063	NC	NC	0.044		
Isavuconazole	2-4	NC	NC	5.657		
Caspofungin	0.5-4	NC	NC	0.707		
Micafungin	0.063-1	NC	NC	0.251		
Terbinafine	0.063-0.063	NC	NC	0.25		
<i>Phialophora ambigua</i> ($n = 1$ /group C3)						
Amphotericin B	8	NC	NC	NC		
Flucon0azole	32	NC	NC	NC		
Itraconazole	0.5	NC	NC	NC		
Voriconazole	0.25	NC	NC	NC		
Posaconazole	0.125	NC	NC	NC		
Isavuconazole	1	NC	NC	NC		
Caspofungin	4	NC	NC	NC		
Micafungin	0.25	NC	NC	NC		
Terbinafine	1	NC	NC	NC		
<i>Phialophora europaea</i> ($n = 43$ /group E)						
Amphotericin B	0.125-16	2	16	2.341		
Fluconazole	2–16	4	8	21.246		
Itraconazole	≤0.016-0.5	0.063	0.25	0.071		
Voriconazole	0.125-1	0.25	0.5	0.25		
Posaconazole	≤0.016-0.125	≤0.016	0.031	0.021		
Isavuconazole	0.25–2	1	1	1.506		
Caspofungin	0.25–4	1	2	1.065		
Micafungin	$\leq 0.008 - 4$	0.25	1	0.167		
Terbinafine	0.25–2	0.5	1	0.533		
<i>Phialophora oxyspora</i> ($n = 3$ /group D)						
Amphotericin B	4–16	NC	NC	11.314		
Fluconazole	4-8	NC	NC	32		
Itraconazole	0.25-0.25	NC	NC	0.25		
Voriconazole	0.5-1	NC	NC	0.707		
Posaconazole	≤0.016-0.125	NC	NC	0.089		
Isavuconazole	2-4	NC	NC	8		
Caspofungin	1-4	NC	NC	4		
Micafungin	0.5–2	NC	NC	2		
Terbinafine	0.5–2	NC	NC	1.414		
<i>Phialophora reptans</i> ($n = 8$ /group B)						
Amphotericin B	1–16	NC	NC	4		
Fluconazole	4-8	NC	NC	22.627		
Itraconazole	≤0.016-0.125	NC	NC	0.044		
Voriconazole	0.063-0.5	NC	NC	0.177		
Posaconazole	≤0.016-0.063	NC	NC	0.019		
Isavuconazole	0.25–2	NC	NC	1.414		
Caspofungin	0.25–2	NC	NC	0.917		
Micafungin	≤0.008-0.5	NC	NC	0.045		
Terbinafine	≤0.008-0.063	NC	NC	0.037		

^{*a*} Molecular groups are those of Feng et al. (12).

 b NC, no comparison because ${<}10$ strains per species were available for testing.

Two species with sufficient numbers of isolates $(n \ge 10)$ were included to calculate the MIC₅₀ and MIC₉₀ values, viz., Phialophora europaea and Cyphellophora guyanensis (Table 1; see Table S2 in the supplemental material). All of the strains of *Cyphello*phora and Phialophora tested had low MICs of itraconazole, voriconazole, posaconazole, and micafungin, while most of the strains had high MICs of fluconazole and amphotericin B. The highest geometric mean MICs were 27.6 µg/ml of fluconazole, followed by 4 µg/ml of amphotericin B, and much lower geometric mean MICs of the triazoles, echinocandins, and allylamine (isavuconazole, 1.9 µg/ml; caspofungin, 1.1 µg/ml; voriconazole, 0.3 µg/ml; terbinafine, 0.3 µg/ml; itraconazole, 0.1 µg/ml; micafungin, 0.07 μ g/ml; posaconazole, 0.03 μ g/ml). Posaconazole was the drug with the best overall activity. The eight species for which few or single isolates were available (P. ambigua CBS 235.93; C. pauciseptata CBS 284.85; C. suttonii CBS 449.91; C. laciniata CBS 174.79, CBS 190.61, and CBS 239.91; C. pluriseptata CBS 285.85, CBS 286.85, and CBS 109633; C. vermispora CBS 277.86, CBS 228.86, and CBS 122852; P. oxyspora CBS 416.89, CBS 698.73, and CBS 124686; and P. reptans CBS 113.85, CBS 152.90, CBS 458.92, CBS 101467, CBS 110814, CBS 120903, CBS 120913, and CBS 123271) yielded the lowest MICs of posaconazole, micafungin, itraconazole, and voriconazole (Table 1). In general, the MIC₉₀s of itraconazole, voriconazole, and isavuconazole were at least 2 log, dilution steps higher than those of posaconazole. The environmental isolates of C. guyanensis had higher MICs of terbinafine and isavuconazole than the clinical strains of P. europaea. The two echinocandins showed marked differences in their MECs; in most cases, micafungin had potent activity, with a geometric mean MEC and MEC₉₀ showing at least 2 log₂ dilution steps higher activity than caspofungin. These data are in agreement with previously reported findings on the black yeasts Cladophialophora (3), Fonsecaea (16), and Rhinocladiella (4). There was a significant difference (P < 0.001) between the MICs of the eight antifungals for *P. europaea* (group E), *P. reptans* (group D), and *C.* guyanenesis (group B) (Table 1). Phialophora europaea isolates, when divided into skin and nail subgroups, showed no difference in susceptibility (see Table S2 in the supplemental material). The present paper provides the first antifungal susceptibility data on clinical species of Cyphellophora and its relatives in the genus Phialophora, which compose a phylogenetic "europaea clade" within the order Chaetothyriales. The results suggest that the species in this clade of black yeast-like organisms are susceptible in vitro to the newer azoles and to micafungin; however, these in vitro data still need to be complemented by clinical confirmation in vivo.

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