Biologicheskie Membrany 2017 vol.34 N2, pages 142-152

Analysis of Exo- and endocytosis in the mouse nerve ending in experimental diabetes mellitus

Yakovleva O., Zaharov A., Zefirov A., Sitdikova G. Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

Abstract

Diabetes mellitus (DM) is a systemic disease characterized by changes in many organs and tissues, including the motor system. The aim of this work was a study of the processes of exoand endocytosis in the motor nerve ending of mouse diaphragm during high-frequency activity in experimental alloxan model of DM. Endplate potentials (EPPs) was recorded using intracellular microelectrodes during single and high-frequency (50 Hz, 1 min) stimulation. In mice with experimental DM the amplitude-temporary parameters of EPPs did not differ from the control; however, an increase in the EPPs depression and a slower recovery was observed during high frequency stimulation. Using an endocytosis dye FM 1-43, it was shown that in animals with experimental DM the intensity of fluorescence of nerve terminals loaded by high-frequency stimulation was higher than in control. This effect was prevented by an inhibitor of slow dynamin-mediated endocytosis - 1-azakenpaullone (2 μ M). In addition, the bleaching of preloaded nerve terminals during high-frequency stimulation was slower in animals with DM. The obtained results suggest that in experimental DM the recycling of synaptic vesicles via the long path becomes more pronounced and the mechanisms of the vesicular transport are impaired. This hypothesis was confirmed by methods of mathematical modeling.

http://dx.doi.org/10.7868/S0233475517020098

Keywords

Diabetes mellitus, Endocytosis, Exocytosis, Mathematical modeling, Neuromuscular junction, Synaptic vesicles

References

- [1] Vincent A.M., Callaghan B.C., Smith A.L., Feldman E.L. 2011. Diabetic neuropathy: cellular mechanisms as therapeutic targets. Nat. Rev. Neurol. 7 (10), 573-583.
- [2] Said G. 2007. Focal and multifocal diabetic neuropathies. Arq. Neuropsiquiatr. 65 (4B), 1272-1278.
- [3] Constantini S., Schiller Y., Cohen A.M., Rahamimoff R. 1987. Pathophysiology of the neuromuscular junction in diabetic rats. Isr. J. Med. Sci. 23 (1-2), 101-106.
- [4] Kimura I., Okazaki M., Kimura M. 1993. Streptozocin-diabetes modifies acetylcholine release from mouse phrenic nerve terminal and presynaptic sensitivity to succinylcholine. Jpn. J. Pharmacol. 62 (1), 35-41.
- [5] Magarinos A.M., McEwen B.S. 2000. Experimental diabetes in rats causes hippocampal dendritic and synaptic reorganization and increased glucocorticoid reactivity to stress. Proc. Natl. Acad. Sci. USA. 91 (20), 11056-11061.
- [6] Fahim, A.M., Al Shuaib W., Davidson N. 1999. Depolarization affects neuromuscular junction of streptozotocindiabetic mice. Cell Mol. Biol. (Noisy-le-grand). 45 (2), 259-263.

- [7] Fahim M.A., Hasan M.Y., Alshuaib W.B. 2000. Early morphological remodeling of neuromuscular junction in a murine model of diabetes. J. Appl. Physiol. 89 (6),: 2235-2240.
- [8] Marques M.J., Santo Neto H. 2002. Acetylcholine receptors and nerve terminal distribution at the neuromuscular junction of non-obese diabetic mice. Anat. Rec. 267(2), 112-119.
- [9] Souayah N., Potian J.G., Garcia C.C., Krivitskaya N., Boone C., Routh V.H., Mc Ardle J.J. 2009. Motor unit number estimate as a predictor of motor dysfunction in an animal model of type 1 diabetes. Am. J. Physiol. Endocrinol. Metab. 297 (3), E602-608.
- [10] Garcia C.C., Potian J.G., Hognason K., Thyagarajan B., Sultatos L.G., Souayah N., Routh V.H., McArdle J.J. 2012. Acetylcholinesterase deficiency contributes to neuromuscular junction dysfunction in type 1 diabetic neuropathy. Am. J. Physiol. Endocrinol Metab. 303 (4), E551-561.
- [11] Rizzoli S.O., Betz W.J. 2005. Synaptic vesicle pools. Nat. Rev. Neurosci. 6(1), 57-69.
- [12] Richards D.A., Guatimosim C, Betz W.J. 2000. Two endocytic recycling routes selectively fill two vesicle pools in frog motor nerve terminals. Neuron. 27 (3),. 551-559.
- [13] Deák F., Schoch S., Liu X., Südhof T.C., Kavalali E.T. 2004. Synaptobrevin is essential for fast synaptic-vesicle endocytosis. Nat. Cell. Biol. 6 (11), 1102-1108.
- [14] Clayton E.L., Anggono V., Smillie K.J., Chau N., Robinson Ph. J., Cousin M.A. 2009. The phospho-dependent dynamin-syndapin interaction triggers activity-dependent bulk endocytosis of synaptic vesicles. J. Neurosci. 29, 7706-7717.
- [15] Clayton E.L., Sue N., Smillie K.J., O'Leary T., Bache N., Cheung G., Cole A.R., Wyllie D.J., Sutherland C, Robinson P.J., Cousin M.A. 2010. Dynamin I phosphorylation by GSK3 controls activity-dependent bulk endocytosis of synaptic vesicles. Nat. Neurosci. 13 (7), 845-851.
- [16] Saheki Y., DeCamilli P. 2012. Synaptic vesicle endocytosis. Cold Spring Harb. Perspect Biol. 4 (9), a005645.
- [17] Wu L-G., Hamid E., Shin W., Chiang H-Ch. 2014. Exocytosis and endocytosis: modes, functions, and coupling mechanisms. Annu. Rev. Physiol. 76, 22.1-22.31.
- [18] Lenzen S. 2008. The mechanisms of alloxan- and streptozocin-induced diabetes, Diabetologia. 51, 216-226.
- [19] Sitdikova G.F., Islamov R.R., Mukhameyarov, Permyakova V.V., Zefirov A.L., Palotas A. 2007. Modulation of neurotransmitter release by carbon monoxide at the frog neuro-muscular junction. Curr. Drug Metab. 8 (2), 177-184.
- [20] Yakovleva O.V., Shafigullin M.U., Sitdikova G.F. 2013. The role of nitric oxide in the regulation of neurotransmitter release and processes of exo and endocytosis of synaptic vesicles in mouse motor nerve endings. Neurochem. J. 7(2), 103-110.
- [21] Zakharov A.V, Petrov A.M., Kotov N.Y, Zefirov A.L. 2012. Experimental and modeling investigation of the mechanism of synaptic vesicles recycling. Biophysics. 57 (4), 508-518.
- [22] Betz W.J., Angelson J.K. 1998. The synaptic vesicle cycle. Annu. Rev. Physiol. 60, 347-363.
- [23] Forde J.E., Dale T.C. 2007. Glycogen synthase kinase 3: a key regulator of cellular fate. Cell Mol. Life Sci. 64 (15), 1930-1944.
- [24] Schneggenburger R., Meyer A.C., Neher E. 1999. Released fraction and total size of a pool of immediately available transmitter quanta at a calyx synapse. Neuron. 23 (2), 399-409.
- [25] Sakaba T., Neher E. 2001. Quantitative relationship between transmitter release and calcium current at the calyx of held synapse. J. Neurosci. 21 (2), 462-476.
- [26] Pirart J., Lauvaux J.P., Rey W. 1978. Degenerative diabetic complications. Is persistent hyperglycemia more dangerous than wide glycemie fluctuations? Nouv Presse Med. 7 (44), 4031-4035.
- [27] Brown M.J., Asbury A.K. 1984. Diabetic neuropathy. Ann. Neurol. 15(1), 2-12.
- [28] Guy R.J., Dawson J.L., Garrett J.R., Laws J.W., Thomas P.K., Sharma A.K., Watkins P.J. 1984. Diabetic gastroparesis from autonomic neuropathy: surgical considerations and changes in vagus nerve morphology. J. Neurol. Neurosurg. Psychiatry. 47 (7), 686-691.
- [29] Biessels G.J., Gispen W.H. 2005. The impact of diabetes on cognition: what can be learned from rodent models? Neurobiol. Aging. 1, 36-41.
- [30] Szkudelski T., Kandulska K., Okulicz M. 1998. Alloxan in vivo does not only exert deleterious effects on pancreatic B cells. Physiol. Res. 47 (5), 343-346.
- [31] Nimnual A.S., Chang W, Chang N.S., Ross A.F., Gelman M.S., Prives J.M. 1998. Identification of phosphorylation sites on AChR delta-subunit associated with dispersal of AChR clusters on the surface of muscle cells. Biochemistry. 37 (42), 14823-14832.
- [32] Kiss G., Somogui Y, Ver I. 2001. Streptozocin-induced diabetes alters the oligomerization pattern of acetylcholinesterase in rat skeletal muscle. Diabetologya. 44, 220-223.
- [33] Rosenmund C, Stevens CF 1996. Definition of the readily releasable pool of vesicles at hippocampal synapse. Neuron. 16(6), 1197-1207.

[34] Zhu Y., Xu J., Heinemann S.F. 2009. Two pathways of synaptic vesicle retrieval revealed by single-vesicle imaging. Neuron. 61 (3), 397-411.