Supplementary Material

Effect of osteocyte-ablation on inorganic phosphate metabolism: analysis of bone-kidney-gut axis

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SUPPLEMENTARY METHODS

Determination of bile acid content in mouse small intestine epithelial cells and feces

Tissue (30 mg) was homogenized with 1.0 ml of 70% ethanol and incubated at 55°C for 4 h. The ethanol extracts were evaporated to dryness and resuspended in 300 μ l of 0.5-M phosphate buffer (pH 7.0) (1). To determine fecal bile acid excretion, feces from individually housed mice were collected over a 72-h period, weighed, and dried. Then, 40 mg of dried feces was minced and extracted in 800 μ l of 75% ethanol at ~50°C for 2 h. The extract was centrifuged, and 100- μ l samples of supernatant were diluted to 400 μ l with a 25% PBS solution for assay (2). Concentrations of total bile acids were determined using the Total Bile Acid Test Wako (Wako).

Analysis of serum triglyceride and cholesterol levels

Serum triglyceride and cholesterol levels were determined by LabAssayTM Triglyceride (Wako), LabAssayTM Cholesterol (Wako).

SUPPLEMENTARY FIGURES

Supplementary Figure 1. Gene expression profiles and concentration of bile acid cholesterol and triglyceride in OCL mice

Ten-week-old DMP-1-hHB-EGF Tg mice and wild-type mice as littermate controls were injected with DT (50 μ g/kg body weight). DT-injected Tg mice are indicated as OCL mice. DT-injected wild-type mice are indicated as control (Cont) mice. A) Photograph of representative kidney in Cont and OCL mice. White adipose tissue around the kidney (mg)/body weight (g) (n=6/group)

(B) and (C) At 5 days after DT injection, total RNA of the liver and distal intestine (late jejunum and ileum) were isolated from OCL and Cont mice. Expression levels of bile acid-related genes were examined using real-time PCR. Data were normalized to GAPDH and pooled from three independent experiments.

B) Liver: Cyp7a1, cholesterol 7 α -hydroxylase; FXR, nuclear receptor farnesoid X receptor; Cyp8b1, sterol 12 α -hydroxylase; SHP, small heterodimer partner; FGFR4, fibroblast growth factor receptor 4; β -klotho; NTCP, Na⁺-taurocholate transporting polypeptide.

C) Distal intestine: IBABP, ileal bile acid binding protein; ASBT, apical sodium-dependent bile acid transporter; FXR; SHP; OST- α , organic solute transporter alpha; OST- β , organic solute transporter beta; FGF15, fibroblast growth factor 15; UGT1a1, UDP glucuronosyltransferase family 1 member A1; UGT1a6, UDP glucuronosyltransferase family 1 member A6; UGT1a7c, UDP glucuronosyltransferase 1 family, polypeptide A7C; UGT2b34, UDP

glucuronosyltransferase 2 family, polypeptide B34. The bar graphs are presented as arithmetic means \pm SEM (n=6/group). Two-tail unpaired t test ***P*<0.01, **P*<0.05 vs Cont

D) At 5 days after DT injection, the epithelial cells of mouse small intestine and serum were collected. The feces were collected for 24 h using a metabolic cage. Serum, small intestine and feces bile acid levels were measured. Serum cholesterol and serum triglyceride levels were measured. The bar graphs are presented as arithmetic means \pm SEM (n=6/group). Two-tail unpaired t test ***P*<0.01, **P*<0.05 vs Cont

Supplementary Figure 2.

Gene expression profiles and bile acid cholesterol and triglyceride concentrations in high Pi diet-fed mice.

Ten-week-old wild-type mice as littermates were fed the Control Pi diet (CP) or the High Pi diet (HP) for 1 week. A) Photograph of a representative kidney from CP-fed and HP-fed mice. White adipose tissue around kidney (mg) / body weight (g) (n=6/group)

(B) and (C) Total RNA of the liver and distal intestine (late jejunum and ileum) were isolated from CP and HP mice. Expression levels of bile acid-related genes were examined using real-time PCR. Data were normalized to GAPDH and pooled from three independent experiments. (n=6/group)

B) Liver: Cyp7A1, cholesterol 7 α -hydroxylase; FXR, nuclear receptor farnesoid X receptor; Cyp8b1, sterol 12 α -hydroxylase; SHP, small heterodimer partner; FGFR4, fibroblast growth factor receptor 4; β -klotho; NTCP, Na⁺-taurocholate transporting polypeptide.

C) Distal intestine: IBABP, ileal bile acid binding protein; ASBT, apical sodium-dependent bile acid transporter; FXR; SHP; OST- α , organic solute transporter alpha; OST- β , organic solute transporter beta; FGF15, fibroblast growth factor 15; UGT1a1, UDP glucuronosyltransferase family 1 member A1; UGT1a6, UDP glucuronosyltransferase family 1 member A6; UGT1a7c, UDP glucuronosyltransferase 1 family, polypeptide A7C; UGT2b34, UDP glucuronosyltransferase 2 family, polypeptide B34.

D) Epithelial cells of mouse small intestine and serum were collected. Serum and small intestine bile acid levels were measured. Serum cholesterol and serum triglyceride levels were measured. The bar graphs are presented as arithmetic means \pm SEM (n=6/group). Two-tail unpaired t test ***P*<0.01, **P*<0.05. (CP VS HP)

Supplemental Table 1. Primer sequences for real-time RT-PCR

Gene -Bone	Forward primer	Reverse primer	Product size (bp)
DMP1	5'-GGCTGTCCTGTGCTCTCCCAG-3'	5'-GGTCACTATTTGCCTGTGCCTC-3'	159
E11/GP38	5'-CAGTGTTGTTCTGGGTTTTGG-3'	5'-GGGGTCACAATATCATCTTCA-3'	91
MEPE	5'-GTGAATGACGCCAGAGGGC -3'	5'-TGTCTTCATTCGGCATTGG-3'	98
Cbfa1	5'-CACTGCCACCTCTGACTTCT -3'	5'-GCTCTCAGTGAGGGATGAAA -3'	127
ALP	5'-CAGGGTACACCATGATCTCACC-3'	5'-CGCCCATACCATCTCCCAGG -3'	180
Osteocalcin	5'-GAGGACCATCTTTCTGCTCAC-3'	5'-CCAAGGTAGCGCCGGAGTCTG- 3'	153
FGE23	5'- ACTTGTCGCAGAAGCATC-3'	5'-GTGGGCGAACAGTGTAGAA-3'-3'	144
SOST	5'-GGAATGATGCCACAGAGGTCA -3'	5'-CCCGGTTCATGGTCTGGTTT -3'	81
Phey	5'-GTGCATCTACCAACCAGATACG-3'	5'-TCTGTTCCCCAAAAGAAAGG-3'	67
Osteonontin	5'-GTGAAAGTGACTGATTCTGGCAGC-3'	5'-CATCATCGTCGTCCATGTGGTCAT-3'	214
Col 1o1		5' TGGGACAGTCCAGTTCTTCAT 3'	111
Gene -Liver	Forward	Poverse	144
			100
	5 - COURTOGROARGAARCTOCA-3		109
			121
			09
			02
Сурарт			84
SHP	5-CGATCUTUTUCACUCAGATG-3		102
β-Klotho	5-IGGGGAGICACIGAGICIGI-3	5'-CATACAGGTGAGGATCGGTAAAC-3	/6
Gene-intestine	Forward	Reverse	
TRPV6	5'-CTCCTCATTGCCATGATGGG -3'	5'-GGCCACAACCTGTGCTCTCCAG-3'	81
CaBP D9k	5'-CCTGCAGAAATGAAGAGCATTTT-3'	5'-CTCCATCGCCATTCTTATCCA-3'	172
PMCA1b	5'-CGCCATCTTCTGCACCATT-3'	5'-CAGCCATTGCTCTATTGAAAGTTC -3'	109
Claudin2	5'-ATACTACCCTTTAGCCCTGACCGAGA-3'	5'-CAGTAGGAGCACACATAACAGCTACCAC-3'	95
Claudin12	5'-CAGACCAGTGTGTACTCAGACTTTCTACCC-3'	5'-GAAGCAACATACTGACTGTCTCCTGACG-3'	109
Claudin15	5'-CATCTTTGAGAACCTGTGGTACAGC-3'	5'-GATGGCGGTGATCATGAGAGC-3'	130
OST-α	5'- ATGCATCTGGGTGAACAGAA -3'	5'-GAGTAGGGAGGTGAGCAAGC-3'	134
OST-B	5'-GACCACAGTGCAGAGAAAGC-3'	5'-CTTGTCATGACCACCAGGAC-3'	142
ASBT	5'-GGAACTGGCTCCAATATCCTG-3'	5'-GTTCCCGAGTCAACCCACAT-3'	146
FGF15	5'-GAGGACCAAAACGAACGAAA-3'	5'-ACGTCCTTGATGGCAATCG-3'	71
IBABP	5'-GGTCTTCCAGGAGACGTGAT-3'	5'-ACATTCTTTGCCAATGGTGA-3'	144
UGT1a1	5'-ATGGCTTTCTTCTCCGGAAT-3'	5'-CAGAAAAAGCCCCTATCCC-3'	116
UGT1a6	5'-CACCGGAACTAGACCATCGAA-3'	5'-GCATCATCACCATCGGAACTC-3'	164
UGT1a7c	5'-TGCAATGGAGTTCCGATGGT-3'	5'-CTGGAGAGGCGCATGATGTT-3'	188
LIGT2b34	5'-GGAGAATGCCATGCGGTTAT-3'	5'-CTGCCACACGAAGATGCTTG-3'	122
Gene_kidney	Forward	Reverse	122
Not2a	5'-AGAGCCCTTCACAAGACTCATCAT-3'	5'-TACCCTGGACATAGAAGTGGAAGC-3'	148
Not2c	5'-TGAAGAACGCTGACCAACTGA-3'		137
Dit2		5'-AAACGTGACCGTCATTCCTC-3'	140
1a(OH)260	5'-GAGCAAACTCCAGGAAGCAG-3'	5'-TCAGCAATCATCAGCAGAGG-3'	11/
24(OH)250	5'-TGGGAAGATGATGGTGACCC_3'	5'-TCGATGCAGGGCTTGACTG -3'	114
a klotho			68
	5'_TGCAGACATCAAGAAGGTTGA_3'	5'_CCCCACTCATCCAACCTC_3'	102
	5' CTTCCTTCTTACCCCTTCAAC 2'		102
IRPV3			104
			/4
			122
FGFR3	5-GUAUAAUUTGGAUTAUTAUAAG-3	5-CAGUGTAAAGATUTUUUAGAG-3	145
			116
GAPDH	5-CIGCACCACCAACIGCIIAGC-3	5-UATUUAUAGTUTTUTUUGUTU-3	110



Supplementary Figure 1



Supplementary Figure 2