

# 博士論文

## Geographic variation and genetic structure of teak (*Tectona grandis*) in Myanmar revealed by cpSNP and nrSSR markers

(葉緑体 SNP と核 SSR マーカーで明らかにされたミャンマーに  
おけるチーク (*Tectona grandis*) の地理変異と遺伝構造)

**Thwe Thwe Win**  
(トウエ トウエ ウィン)

**The University of Tokyo**

## Contents

Acknowledgement	1
Chapter 1 Introduction	3
1. Introduction	3
1.1 Teak ( <i>Tectona grandis</i> )	3
1.2 Teak in Myanmar	5
1.3 Genetic information of native teak	6
1.4 Problem statements	7
1.5 Objectives	8
Chapter 2 Genetic diversity of teak in its native region	13
2.1 Introduction	13
2.2 Materials and Methods	15
2.2.1 Sampling design and DNA extraction	18
2.2.2 Molecular genotyping	19
2.2.3 Statistical analysis	20
2.3 Results	26
2.4 Discussion	26
2.5 Conclusion	28
Chapter 3 Development of chloroplast single nucleotide polymorphism markers of teak	28
3.1 Introduction	28
3.2 Materials and methods	30
3.2.1 DNA extraction	30
3.2.2 DNA sequencing for the development of chloroplast	

Markers	30
3.3 Results	32
3.3.1 Chloroplast polymorphism	32
3.4 Discussion	32
Chapter 4 Geographic variation pattern of Myanmar teak revealed by newly developed cpSNP and nrSSR markers	39
4.1 Introduction	39
4.2 Materials and methods	40
4.2.1 Study site and sampling design	40
4.2.2 DNA extraction, chloroplast SNP and nuclear microsatellite genotyping	44
4.2.3 Data analysis	46
4.3 Results	47
4.3.1 Geographic variation revealed by cpSNP markers	47
4.3.2 Genetic structure revealed by nrSSR	53
4.3.3 Geographic variation and genetic differentiation	61
4.4 Discussion	67
4.4.1 Geographic variation revealed by chloroplast SNP markers	67
4.4.2 Genetic diversity and genetic structure revealed by nrSSR markers	68
4.4.3 Comparison between findings from cpSNP and nrSSR analysis	69
4.5 Conclusion	71

Chapter 5 Comparison of genetic composition between alien and native teak in Myanmar using simple sequence repeat (SSR) markers	73
5.1 Introduction	73
5.2 Materials and methods	74
5.2.1 Sampling	74
5.2.2 Genotyping	77
5.2.3 Statistical Analysis	77
5.3 Results	78
5.4 Discussion	87
Chapter 6 General Discussion	89
Summary	93
References	97

## Acknowledgement

I am very first indebted to my parents, U San Thein and Daw Win Sein for giving me a human life, looking after till now and guiding how to survive in life. I'd like to gratitude Dr. Kurunobu, Dr. Nagame for favoring me to start my study at The University of Tokyo. I deep in thank Dr. Yuji Ide for the kind supports to my studies. My special thanks also go to Dr. Susumu Goto and his family, my supervisor for valuable advice and supports for my master and doctorate studies. Deepest thanks convey to Dr. Atsushi Watanabe from Kyushu University, his wife and Dr. Tomonori Hirao from Forest Tree Breeding Center for their great contribution and supports to do the experiments at FTBC, and care when I stayed in Hitachi and encouragement whenever I faced the difficulties. My sincere thanks go to Dr. Wataru Ishizuka for lending his hands for both studying and my personal affairs since the beginning of my stay in Japan. It is grateful for Dr. Makoto Takahashi, Dr. Hiroshi Iwata and Dr. Sakaguchi, for teaching me concerned with data analysis. I'd like to thank the MEXT from Japan for the financial assistance for studying and living cost in Japan for six years. My heartfelt thanks go to Takayuki Sato and his wife and Akira Sakai and his family for their care a family member during my stay in Japan. Dr. C. Lian and K. Ushiyama are acknowledged for their advices and reviewing my dissertation.

Authorized persons from The Republic of Myanmar, particularly from the Ministry of Environmental Conservation and Forestry are specially thanked for giving the permission to study in abroad. I'd like to convey my deepest thanks to the Staff from Forest Tree Breeding Center from Japan for helping me during my study and from the Ministry of Forestry for collecting samples. I deeply appreciate Ma Khin Moe Kyi, my best friend and Ma Soe Soe New, lovely junior for favoring me

without vexation whenever I make a request. I also thank to Myanmar friends for trying together to fix up with new environment and Japanese friends for giving a hand to face the difficulties in your land.

My special heartfelt thanks go to my brothers' families, Ko Zaw Naing Oo and Ma Khin Nyein Chan, Ko Win Thein Oo and Ma Toe Toe Lwin, sisters, Ma Nu Nu Win and Ma Swe Swe Win and relatives, U Aung Moe and Ma Htay Htay Myint, and my lovely nieces Dr. May Phyu Moe, Aye Chan Moe and May Myo Myat Khin, nephews, Ent Zin Ko and Ye Wint Ko for your caring our beloved mother, being patient to me and physical and mental supports to see my dream in real. Your kindness refreshes and enhances me when I am away from you all. At last but not least, heartfelt thanks go to my beloved Ko Zaw Zaw Toe, my fiancé, for your encouragement, kindness and special care. Everyone who favors and contributes to my study is deeply appreciated though I left to express the individual's name.

## Chapter 1

### Introduction

Deforestation rates significantly increased across tropical Asia and the highest level of deforestation experienced in Southeast Asia during 1990s (Wright, 2005, Miettinen et al. 2011). Four countries of teak native regions, Thai and Laos included in Southeast Asia (Fig 1.1). Due to the increasing demand, teak plantations were widely established in the world including its indigenous countries and stands third position of planted species. Thus, both conservation and breeding programs are urgently required to save the natural genetic resources of teak and to provide the genetically improved materials for teak plantations. Information on genetic variation is important to maintain the natural population as evolutionarily viable units which are adaptable to changing environments in the long term (Zuo et al. 2010). Genetic information particularly phylogeographic variation and genetic structure are indispensable for effectively and efficiently implementing the conservation and breeding programs for target species through formulating the useful strategies.

#### 1.1 Teak

The study species, teak (*Tectona grandis*) is one of the most economically important tropical deciduous timber species. It belongs to family Lamiaceae and genus *Tectona*. Teak requires a high light intensity for its growth and development and it has been classified as a pioneer species. Teak is angiosperm, diploid, monoecious and allogamous species (Gill, et al., 1983; Mathew, et al., 1987; Kertadikara and Prat, 1994). It is primarily an outcrossing and insect pollinated species, but self-pollination is possible. No self-pollinated flowers develop into

mature fruits, although many of the fruits develop to different sizes before they abort (Tangmitchroen and Owens, 1997).

Teak (*Tectona grandis* L.f) has been regarded as one of the world's most precious tropical tree species due to precious timber qualities (Pandey and Brown, 2000; Kaosa-ard, 2003) and increasing demand in the world market. It is recognized worldwide as the most important wood for multipurpose particularly for ship building and furniture. The unique qualities of teak such as durability, ease of seasoning without splitting and cracking, workability, beautiful color and grain, and resistance to termite, fungus and weathering, etc. make increasing its demand and endangered species (Gyi and Tint, 1998; Kaosa-ard, 1998). Timber color of teak varies with four types, golden yellow, light brown, dark brown and black stripe (Fig 1.2).

The recorded latitudinal limits of its natural range are between 9° to 26° N latitude and 73° to 104° E longitude (White, 1991). Its natural distribution is limited to a discontinuous range in South and Southeast Asia from the Indian subcontinent to Myanmar, Thailand and Laos (Khanduri, et al., 2008). The total amounts of teak bearing forest is ca 27.9 million ha and out of them 8,900,000 ha in India (Tewari, 1992), 2,500,000 ha in Thailand (Kaosa-ard, 1991), and 16,000 ha in Laos (Anon, 1992). The rest of area about 16,517, 700 ha are in Myanmar (Pengduoang, 1991).

Natural populations of teak in its native countries are nearly to be depleted through over-exploitation, illegal cutting and other factors such as the transformation of land-use systems. Teak logging from the natural forest has been banned in its native regions in the late 1980s except Myanmar (Pandey and Brown, 2000). Logging from natural forests in India, Thailand and Laos has been banned since the



late 1980s. Teak is now a threatened species so conservation effort is urgently needed to safeguard the genetic resources of teak from degraded natural teak forests. Teak plantations were widely planted not only in native regions but also Asia, Africa and Central America to supply the high demand of teak. Therefore, conservation and breeding program should be balanced to retain the irreplaceable natural genetic resource of teak and to produce the genetically qualified planting materials for the commercial plantations. Genetic diversity and genetic variation are key components of the stability of forest resources (Rajora, et al., 2000). It is therefore important to evaluate the genetic diversity and genetic divergence of natural populations in native countries to facilitate conservation efforts aimed at maintaining species' genetic resources.

## 1.2 Teak in Myanmar

Myanmar is geographically situated between latitudes 09° 32' to 28°31' N and longitudes 92°10' to 101° 11' E. In Myanmar, natural teak forests occurs within 25°30'N and 10°00' N latitude and widely distributed from the sea level to 1000 m elevation (Fig 1.3). Today, beautiful natural teak stands can be seen only in Myanmar where is home to the best quality of teak. Myanmar has the largest area of natural teak forests and its timber quality is declared as the best in the market. In Myanmar, teak grows throughout the Shan state except over limited elevation and extends beyond the frontier into Thailand and Laos on the east. In the northwest it does not extend beyond the western watershed of the Ayeyarwady and Chindwin rivers; in the southwest it occurs on the west bank of the Ayeyarwady into the foothills of Rhakhin Yomas in decreasing abundance to approximately 18° N latitude. It does not occur

abundantly in the dry zone of central Myanmar, or in the tidal regions of the delta area.

The forests in Myanmar are classified into six major forest types. Teak naturally occurs in major three types of forests; the semi-evergreen forests, mixed deciduous forests (Moist upper mixed deciduous forests, dry upper mixed deciduous forests, and lower mixed deciduous forests) and deciduous dipterocarp or Indaing forests (Fig 3). The composition of teak in natural forests varies with forest types from 4-12%. Teak is usually found as scattered individuals or in small groups with little or no regeneration present in the semi-evergreen forest (Kermode, 1964). In the lower mixed deciduous forest, teak may be found gregariously or in patches with a large girth and height and fluted trees while teak with cleaner and straight boles in the moist upper mixed deciduous forest. Small size and poor quality of teak grows in Indaing forest in Myanmar.

### 1.3 Genetic information of native teak

Genetic studies on teak populations in its native countries of India, Thailand and Laos have been conducted using plant materials derived from international provenance trials established in the early 1970s (Keiding et al., 1986, Kjaer et al., 1995) and natural populations. Previous population genetic studies have used various DNA markers such as allozymes (Kertadikara and Prat, 1995, Kjaer and Seigismund, 1996), amplified fragment length polymorphisms (AFLP) (Shrestha, et al., 2005, Fofana et al., 2013), inter simple sequence repeats (ISSR) (Narayanan, et al., 2007), and simple sequence repeats (SSR) (Fofana, et al., 2008, 2009; 2013; Minn, et al., 2014). SSRs are arguably the most informative of these marker types due to their

hyper-polymorphic nature and co-dominance (Powell, et al. 1996).

Molecular markers of teak showed the genetic variation of teak. AFLP markers showed large genetic variation in natural teak provenances within India and also natural populations from native regions; India, Thai and Laos teak (Shrestha, et al., 2005; Sreekanth, et al. 2012) and higher genetic divergence of Indian teak at isozyme variation (Kertadikara and Prat, 1995; Kjaer and Seigismund, 1996). Fofana, et al. (2009, 2013) found that the southern Indian populations possessed the highest genetic diversity, followed by the northern Indian, Thailand and Laotian teak populations. Similar results were obtained using AFLP markers (Shrestha, et al., 2005). Significant geographic variation pattern of Myanmar teak was recently detected among southern and northern populations (Minn, et al., 2014). However, comparison of genetic diversity of teak in its native areas; India, Myanmar, Thai and Laos, has not investigated yet.

#### 1.4 Problem statements

Economically and ecologically important tree species, teak have suffered from particularly severe pressure because of selective logging of phenotypically superior trees. Diminishing the area of natural teak forest in native regions might be resulted in decreased in genetic diversity, disrupted gene flow and genetically isolated tree populations. Thus, conservation and breeding programs are urgently needed for teak.

Although knowledge of the genetic variation of extant populations over the entire range of natural distribution is essential for the conservation of genetic resources (Neale and Kremer, 2011), genetic information is very limited for

Myanmar teak among its native regions. Without genetic information of Myanmar teak, it is impossible to discover the genetic center of the teak in the world. Tropical forests were decreased due to various reasons including the Myanmar forests. Within two decades natural teak forest area in Myanmar was drastically diminished, but genetic information resulted from very few studies of teak is insufficient to apply to conservation or breeding purposes. Large scale area of teak plantation was widely established using seeds from various sources of Myanmar teak and also from alien teak without knowing their genetic background that poses a risk of genetic disturbance to natural genetic resources. There was no clear instruction for seed transfer to plantation sites. For commercial teak plantation, breeding program is important to produce the genetically improved planting materials, while focusing on the conservation of natural genetic resource of teak. Thus, geographic variation and genetic structure of teak needed to understand for balancing conservation and breeding processes.

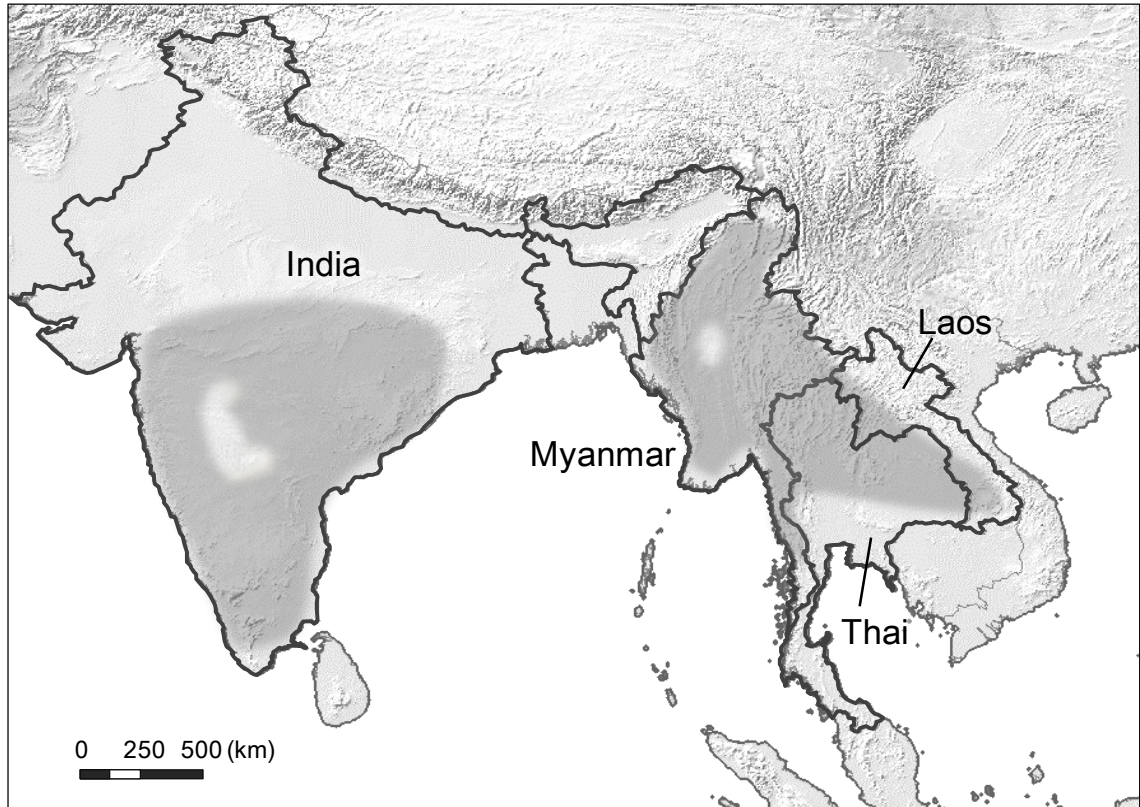
### 1.5 Objectives

1. To evaluate the genetic diversity of Myanmar teak comparing with other native teak
2. To reveal phylogeographic variation and genetic structure of Myanmar teak
3. To formulate seed transfer guidelines and designate zones for conservation and breeding
4. To elucidate genetic component of alien teak planted in Myanmar by private companies

5. To retain the natural genetic resources of Myanmar teak and prevent from genetic disturbance of alien teak

6. To provide the genetic information of Myanmar teak for conservation

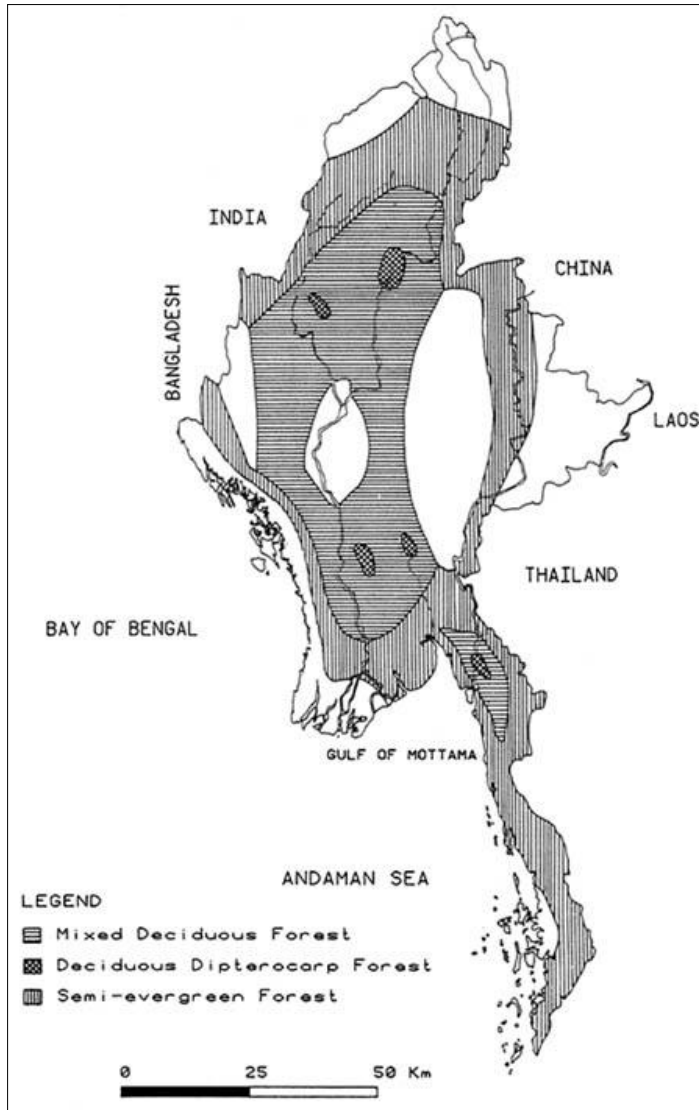
To complete the objectives of this study, first, level of genetic diversity of Myanmar teak among native teak was investigated using nuclear SSR markers by the comparison with that of other native teak in chapter 2. After developing cpSNP marker for teak in chapter 3, phylogeographic variation and genetic structure of Myanmar were investigated using newly developed cpSNP and nrSSR markers in chapter 4. Genetic components of alien teak planted in Myanmar were elucidated using nrSSR makers in chapter 5. In chapter 6, the findings from this study were finally discussed to retain the natural genetic resources of Myanmar teak through balancing conservation and breeding activities and to alarm the genetic disturbance of alien teak which was genetically different from Myanmar teak.



**Figure 1.1 Natural distribution of teak in its native regions, India, Myanmar, Laos and Thailand.** The shaded areas show natural distribution of teak in each respective country.



**Figure 1.3 Color variation of teak**



**Figure 1.2 Natural teak bearing forests in Myanmar (FD-Myanmar)**



## Chapter 2

### Genetic diversity of teak in its native region

#### 2.1 Introduction

Teak (*Tectona grandis* L.f) has been regarded as one of the world's most precious tropical tree species because it provides premium timber with a number of very desirable properties including high durability, strength and workability (Pandey and Brown, 2000; Kaosa-ard, 2003). Its natural distribution is limited to a discontinuous range in South and Southeast Asia from the Indian subcontinent to Myanmar, Thailand and Laos (Khanduri, et al., 2008). Natural populations of teak in its native countries have decreased through over-exploitation, illegal cutting and other factors such as the transformation of land-use systems, so that logging from natural forests was banned in the late 1980s in India, Thailand and Laos but not in Myanmar (Pandey and Brown, 2000). Teak is now a threatened species and conservation effort is urgently needed to safeguard the genetic resources of teak from degraded natural teak forests. Genetic diversity and genetic variation are key component of the stability of forest resources (Rajora, et al., 2000). It is therefore important to evaluate the genetic diversity and genetic divergence of natural populations in native countries to facilitate conservation efforts aimed at maintaining species' genetic resources.

Genetic studies on teak populations in its native countries of India, Thailand and Laos have been conducted using plant materials derived from international provenance trials established in the early 1970s (Keiding, et al., 1986, Kjaer, et al.,

1995) and natural populations. Previous population genetic studies have used various DNA markers such as allozymes (Kertadikara and Prat, 1995, Kjaer and Seigismund, 1996), amplified fragment length polymorphisms (AFLP) (Shrestha, et al., 2005, Fofana et al., 2013), inter simple sequence repeats (ISSR) (Narayanan, et al., 2007), and simple sequence repeats (SSR) (Fofana, et al., 2008, 2009; 2013; Minn, et al., 2014). SSRs are arguably the most informative of these marker types due to their hyper-polymorphic nature and co-dominance (Powell, 1996) and therefore useful for elucidating the spatial structure of genetic diversity and the demographic patterns of variation which have resulted from migration (Neale and Ingvarsson, 2008) and drift as well as through evolutionary history.

In the previous studies, large genetic variation was observed in natural teak provenances and higher genetic divergence of Indian teak at isozyme variation (Kertadikara and Prat, 1995; Kjaer and Seigismund, 1996). Fofana, et al. (2009, 2013) found that the southern Indian populations possessed the highest genetic diversity, followed by the northern Indian, Thailand and Laotian teak populations. Similar results were obtained using AFLP markers (Shrestha, et al., 2005). Significant geographic variation pattern of Myanmar teak was recently detected among southern and northern populations (Minn, et al., 2014). However, comparison of genetic diversity of teak in its native areas has not investigated yet. Therefore, this study was conducted to figure out the genetic diversity of Myanmar teak by comparing that of Indian, Thailand, and Laotian teak.

## 2.2 Materials and Methods

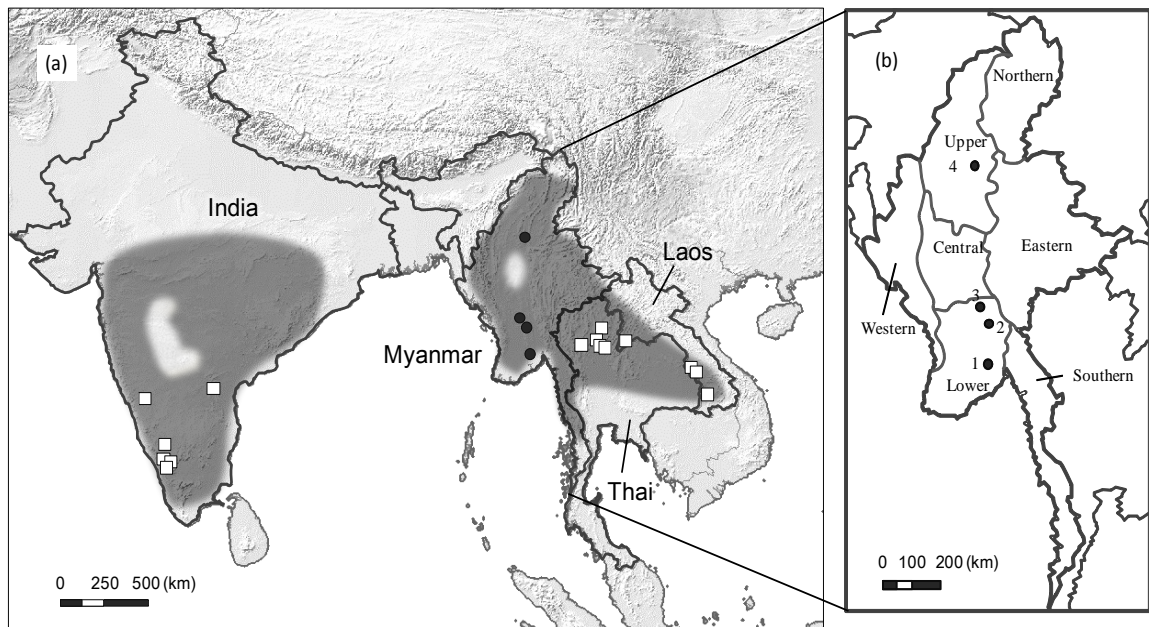
### 2.2.1 Sampling design and DNA extraction

A total of 128 leaf samples from four natural populations were used to investigate the genetic diversity of Myanmar teak (Fig 2.1, Table 2.1). Those samples were collected from a provenance trial established at Pyinmana, Myanmar in 2007 and the collected samples represented natural populations with seed sources from Bago, Phyu, Oktwin and Kanbalu. From this provenance trial, fresh leaves were collected and dried overnight at 80 °C and stored in silica gel at room temperature.

Total DNA was extracted following the method of Shiraishi and Watanabe (1995). Approximately 100 mg of leave sample was frozen in liquid nitrogen and ground in a homogenizer. Each homogenized sample was mixed with 1 ml of CTAB (hexadecyltrimethylammonium bromide) buffer (100 mM Tris-HCl, pH 9.0, 20 mM EDTA, 2% CTAB), with 0.1% beta-mercaptoethanol added immediately prior to use. The mixture was incubated at 65 °C for 1 hr and centrifuged for 10 min at 12 000 xg; 600 µl of the supernatant was then transferred to a 1.5 ml microcentrifuge tube. The supernatant was mixed twice with phenol/chloroform/isoamyl alcohol (25:24:1) and centrifuged for 10 min at 12 000 xg. DNA was precipitated from the aqueous phase by adding 0.1 volume of 3 M sodium acetate and 2.5 volumes of ethanol. The precipitate was washed twice with 70% ethanol and dissolved in water. Extracted DNA was further purified using the DNeasy Plant Mini kit (Qiagen).

**Table 2.1 Geographic and climatic information of four natural population of teak in Myanmar**

No.	Population Name	Seed source of provenance trial	N	Latitude	Longitude	Altitude (meter)	Sampling Site
1	Bago	Natural	32	18° 7'N	96° 4'E	134	Provenance trial
2	Phyu	Natural	32	18°28'N	96°20'E	399	Provenance trial
3	Oktwin	Natural	32	18°55'N	96° 1'E	245	Provenance trial
4	Kanbalu	Natural	32	23°30'N	95°52'E	274	Provenance trial



**Figure 2.1 (a) Maps of the distribution of teak in India, Myanmar, Laos and Thailand and (b) the locations of the ten sampled populations of teak in Myanmar.** In (a), open squares indicate the locations of the teak populations from a previous study (Fofana, et al., 2009) and closed circles represent Myanmar teak populations. In (b) the shaded area shows the natural distribution of teak in its native regions.

### 2.2.2 Molecular genotyping

Fifteen microsatellite markers (Verhaegen, et al., 2005) were used to compare the genetic diversity of natural populations of teak from Myanmar with that of teak from India, Thailand and Laos (Fofana, et al., 2009). To compare the genetic diversity of Myanmar teak with other teak from its native regions, we must use the same number of markers. We modified the locus CIRAD4TeakH09 based on the sequence obtained from Genbank as it could not depict the clear amplification of peaks. The modified forward and reverse primer sequences of CIRAD4TeakH09 are 5'-CTGTGCCTTCTAGTTGCCAGCGCAAGAGCTGAAAGCAACC-3' and 5'-GGCCGTTAGCACTCCATTTA -3'. The microsatellite genotyping was conducted with four fluorescent dyes detected using multiple-tailed primers to allow simultaneous genotyping of four different microsatellite loci (Missiaggia and Grattapaglia, 2006). For PCR, we used the QIAGEN multiplex PCR kit with 2xQIAGEN multiplex PCR master mix (final concentration, 1x), a 0.25  $\mu$ M concentration of each set of primer, 2.5  $\mu$ L of distilled water, and 2  $\mu$ L of DNA for a total volume of 10  $\mu$ L. The florescent universal tail primers, T7 terminator primer (FAM-5'-ATGCTAGTTA TTGCTCAGCGG-3'), reverse complement of BGH-R primer (VIC-5'-CTGTGCCT TCTAGTTGCCAGC-3'), reverse complement of pCold-R primer (NED-5'-TTGGGTGCAATGAGAATGCG-3') and pCold TF-F1 primer (PET-5'-CACTTTCAACGAGCTGATG-3') were developed (Hirao et al., unpublished) based on the TAKARA universal primers (TAKARA Shuzo, Japan). These oligo tails were added to the 5' end of forward primers of developed teak microsatellite markers to complement the sequences of different loci in the PCR reaction. PCR amplifications

were carried out in a PTC-200 thermocycler (MJ Research) using the multiplex-touchdown-PCR protocol (QIAGEN Multiplex PCR kit, QIAGEN): denaturing at 94°C for 15 min, an initial 10 cycles of denaturing at 94°C for 30 s, annealing at 55°C for 90 s with a decrease of 0.5°C per cycle, and an extension at 72°C for 1 min with the annealing temperature of the remaining 20 cycles set at 50°C for 90 s. After a final extension at 72°C for 10 min was used to ensure complete amplification, the products were stored at 4°C. A 1 µL aliquot of the PCR product was mixed with 11.7 µL of Hi-Di™ formamide (Applied Biosystems) including 0.3 µL of Genescan-500 size standard (Applied Biosystems). After denaturing the mixed products at 95°C for 5 min, they were examined using electrophoresis on an ABI 3130xl Genetic Analyzer (Applied Biosystems, USA) and their fragment lengths were assayed using GeneMapper software (Applied Biosystems).

### 2.2.3 Statistical analysis

The following genetic diversity parameters for each locus over the four natural populations of Myanmar: the number of alleles ( $A$ ), allelic richness ( $R$ ), observed heterozygosity ( $H_O$ ), expected heterozygosity ( $H_E$ ), and fixation indices; genetic differentiation among populations ( $F_{ST}$ ) and inbreeding coefficient ( $F_{IS}$ ) were computed. To compare the genetic diversity of Myanmar teak with other teak from its native regions, the genetic diversity parameters;  $R$ ,  $H_E$  and  $F_{ST}$  were measured for each natural population across 15 loci. Samples of each natural population were randomly excluded to reduce to the minimum sample size of population from Fofana et al., 2009 for the calculation of allelic richness due to rarefaction method (Leberg,

2002). Weighted average values of  $H_E$  and  $R$  of populations from each country were used for the comparison of genetic diversity of teak from each native country as accurate as possible and calculated as following. The sample of each population was divided by total sample size of each country and multiplied by  $H_E$  or  $R$  values of correspondent population. Then average  $H_E$  or  $R$  of all populations from each country was calculated. We tested the significance of the differences in the  $R$  and  $H_E$  between Myanmar teak and Indian, Thailand and Laos teak populations using permutation tests with 3 000 permutations.

### 2.3 Results

The number of alleles at each locus from the four natural populations varied from 7 (CIRAD4TeakDa12) to 20 (CIRAD3TeakB02 and CIRAD1TeakH10) with an average of 13. The mean allelic richness was 8.41 and ranged from 3.94 (CIRAD1TeakG02) to 14.14 (CIRAD1TeakH10). Average expected heterozygosity was 0.611 with a range from 0.177 (CIRAD1TeakG02) to 0.851 (CIRAD1TeakH10). Seven of fifteen loci showed significant  $F_{IS}$  values with minimum and maximum  $F_{IS}$  values observed at CIRAD4TeakH09 (-0.203) and CIRAD3TeakE06 (0.311), respectively, (Table 2.2).



**Table 2.2 Genetic information of 15 SSR markers across four natural populations of Myanmar teak**

Locus Name	$N$	$A$	$R$	$H_o$	$H_o$	$H_E$	$F_{ST}$	$P$ -value
CIRAD1TeakA06	127	10	6.89	0.614	0.614	0.650	0.079	0.186 (NS)
CIRAD1TeakB03	127	15	10.21	0.788	0.788	0.755	0.128	0.864 (NS)
CIRAD1TeakF05	128	12	8.07	0.391	0.391	0.572	0.056	0.001 (*)
CIRAD1TeakG02	127	7	3.94	0.173	0.173	0.211	0.095	0.040 (*)
CIRAD1TeakH10	128	20	14.14	0.820	0.820	0.851	0.047	0.192 (NS)
CIRAD2TeakB07	128	18	8.83	0.477	0.477	0.574	0.090	0.001 (*)
CIRAD2TeakC03	116	14	10.08	0.827	0.827	0.799	0.086	0.826 (NS)
CIRAD3TeakA11	128	14	9.41	0.664	0.664	0.758	0.036	0.002 (*)
CIRAD3TeakB02	128	20	12.91	0.695	0.695	0.730	0.093	0.141 (NS)
CIRAD3TeakDa09	126	8	5.59	0.313	0.313	0.375	0.093	0.012 (*)
CIRAD3TeakE06	127	12	8.64	0.487	0.487	0.693	0.062	0.001 (*)
CIRAD3TeakF01	128	13	9.35	0.641	0.641	0.722	0.074	0.009 (*)
CIRAD4TeakDa12	128	7	4.03	0.367	0.367	0.338	0.055	0.910 (NS)
CIRAD4TeakF02	128	9	6.55	0.547	0.547	0.564	0.111	0.329 (NS)
CIRAD4TeakH09	127	12	7.50	0.660	0.660	0.546	0.084	0.999 (NS)
Mean	127	13	8.408	0.564	0.564	0.609	0.079	

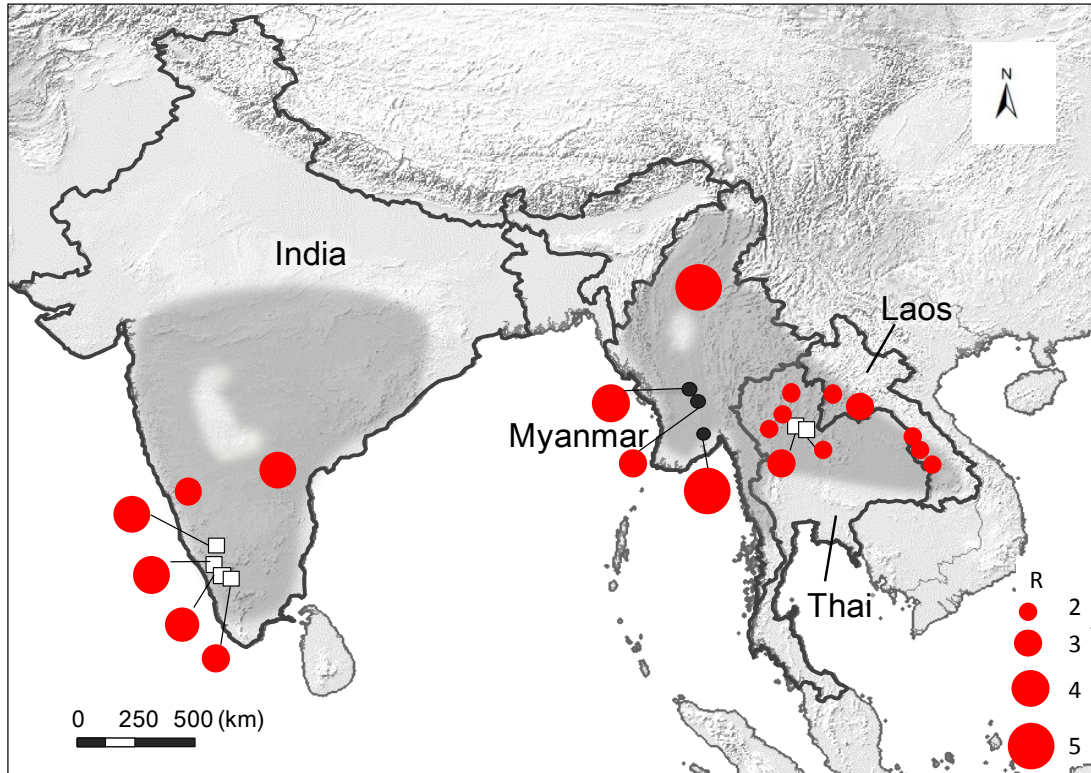
$N$ : number of samples,  $A$ : mean number of alleles,  $R$ : allelic richness,  $H_o$ : the observed heterozygosity,  $H_E$ : the expected heterozygosity,  $F_{ST}$ : genetic differentiation among populations,  $P$  values for the HWE test, (NS) means non-significant, (\*) Significance threshold at 5 % and (\*\*) Significance threshold at 1 %.

Genetic diversity parameters calculated from 15 loci for Myanmar natural teak were  $R = 4.91$ ,  $H_E = 0.609$ , and  $F_{ST} = 0.079$ . The weighted average values of the expected heterozygosity and allelic richness of six natural populations from India, five from Thailand and five from Laotian teak obtained from Fonfana, et al., (2009) were calculated and compared with Myanmar teak (Table 2.3). Allelic richness of Myanmar teak was significantly higher than that of Indian, Thai and Laotian teak (Fig 2.2). However, expected heterozygosity of Myanmar teak was significantly lower than that of Indian teak, but significantly higher than that of Thai and Laotian teak (Fig. 2.3).

**Table 2.3 Statistical comparison of genetic diversity estimates between Myanmar teak and Indian, Thai and Laotian teak**

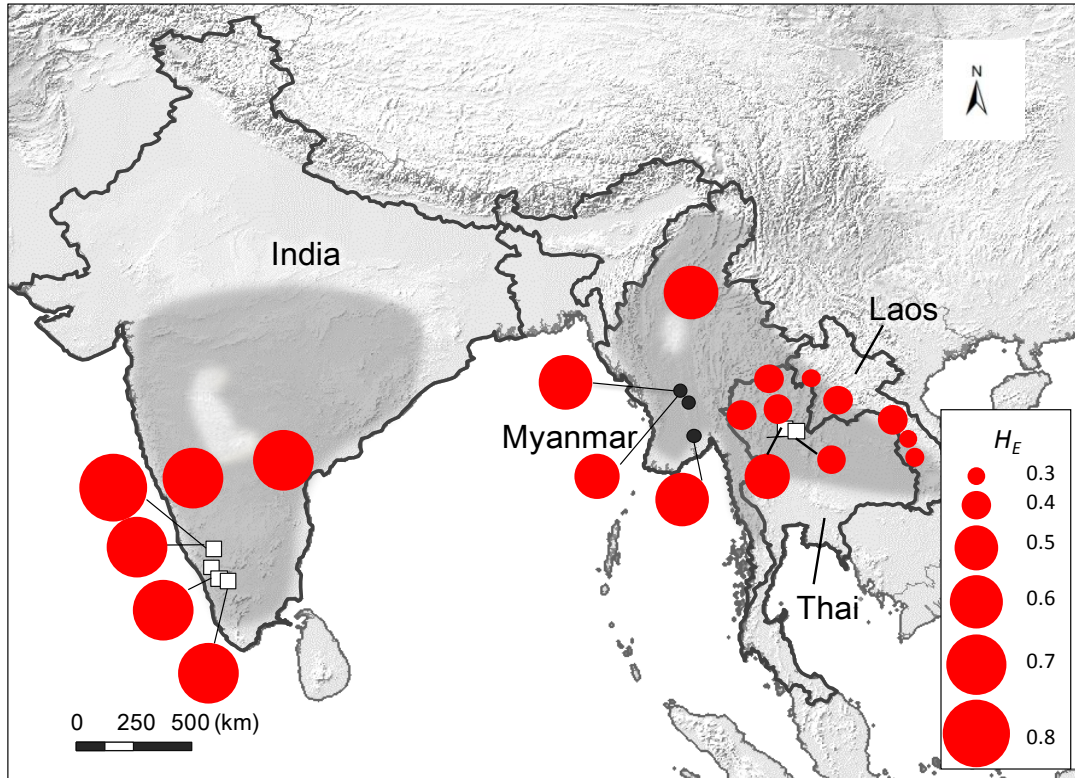
Country	No. of populations	$N$	$R$ ( $p$ -value)	$H_E$ ( $p$ -value)	$F_{ST}$	Reference
Myanmar	4	128 (32)	4.91	0.609	0.079	This study
South India	6	71 (7 - 22)	4.20 (0.03)	0.748 (0.004)	0.030	Fofana et al. 2009
North Thai	5	46 (5 - 13)	2.68 (0.003)	0.450 (0.016)	0.120	Fofana et al. 2009
Laos	5	39 (5 - 13)	2.14 (0.002)	0.356 (0.002)	0.050	Fofana et al. 2009

$N$ : number of samples (numbers in parenthesis indicate the range among different populations),  $R$ : weighted average of allelic richness,  $H_E$ : weighted average of expected heterozygosity.  $p$ : probabilities in  $R$  and  $H_E$  using 3,000 permutations.  $F_{ST}$ : genetic differentiation among populations. (\*) Significance threshold at 5 % and (\*\*) Significance threshold at 1 %.



**Fig. 2.2 Distribution of genetic diversity parameters of teak for allelic richness.**

The diameter of the circles is proportionate to the level of allelic richness or expected heterozygosity and numbers indicate values. Parameters for Myanmar were calculated in this study and those for Indian, Thai and Laotian populations are from Fofana et al. (2009).



**Fig 2.3 Distribution of genetic diversity parameters of teak for expected heterozygosity.**

## 2.4 Discussion

Our results of high allelic richness and the expected heterozygosity of teak in Myanmar compared to other countries (except for the expected heterozygosity in India) does not support our hypothesis that Myanmar teak has the highest genetic diversity among the four native countries. However, genetic diversity of Myanmar teak is significantly higher than that of Thailand and Laotian teak. Genetic diversity is expected to be lower in small isolated populations, such as Thailand and Laos, as a consequence of bottlenecks, founder effects, and inbreeding (Lammi, et al., 1999). Finding in this study is consistent with the summarizing of the previous studies that genetic diversity of teak is decreasing with the eastward direction; from south India, north India, Myanmar, Thai and Laotian teak. However, natural teak forests cover a much larger area in Myanmar which therefore has higher genetic diversity and a moderate level of genetic differentiation compared to those in other teak native regions (Table 2.3). Both population divergence and diversity are important for conservation because they contribute to total species diversity (Petit, et al., 1998). Thus, Myanmar teak populations with high genetic diversity and moderate genetic differentiation among populations would be an important global genetic resource.

## 2.5 Conclusion

For conservation, more attention should be given to genetic diversity, allelic richness and genetic divergence (Petit, et al., 1998, Steven, 2004; Shrestha, et al., 2005). We found that teak populations from Myanmar possessed high genetic diversity, the highest allelic richness and moderate genetic divergence compared to

other native countries. Genetic resources of Myanmar teak should therefore be a priority for *in situ* conservation programs. However, Myanmar and Indian teak might be future prospective for understanding geographic patterns in the genetic structure of teak.

## Chapter 3

Development of chloroplast single nucleotide polymorphism (cpSNP)  
markers of teak

This chapter is going to publish in soon.



## Chapter 4

Geographic variation pattern of Myanmar teak revealed by newly developed cpSNP and nrSSR markers

This chapter is under the process of submission.

## Chapter 5

Comparison of genetic composition between alien and native teak in Myanmar using simple sequence repeat (SSR) markers

Chapter 5 is also in preparation for publication.

## Chapter 6

### General Discussion

Among four native countries of teak, the highest genetic diversity was observed in India teak. This finding was congruent with Hasen, et al., 2015. At the age of the inversion of Myanmar central Basin, ca 10 million years ago, the opening of the Andaman Sea affected sharply bending northeastward the India relative to Myanmar motion (Bertrand and Rangin, 2003). The tectonic movement and the level of genetic diversity of teak from native regions indicated that teak might have migrated into eastward direction from India to Myanmar. Anyhow, natural teak forest in India was nearly to be depleted since the late 1980s. In the long term, reduction in population size may decrease the genetic variation (Ledig, 1992) that is important for adaptation to environmental changes. Moreover, genetic erosion has occurred in natural teak in India due to uncontrolled logging and unrestricted movement of planting materials (Ansari, et al., 2012). Myanmar teak with high genetic diversity is therefore important as natural gene resources of teak in the world. Moreover, Myanmar has the largest area of natural teak forests with the best timber quality. Thus, Myanmar teak should be concentrated not only for conservation but also for the production of genetically improved materials through breeding and tree improvement program.

Knowledge of variation patterns in intraspecific chloroplast DNA (cpDNA) is useful to examine the numerous aspects of evolutionary genetics including migration patterns and rates, drift, and population structure (Golden and Bain, 2000). Among three regions, tranK-rps16 was described as the best choice for molecular studies because of its phylogenetically informative character (Powell et al. 1996).

The region psbK-psbI has the highest discrimination power and is useful for species identification (Zuo et al. 2010). The regions developed cpSNP markers, psbI-psbK, trnK-rps16 and rpl16 regions The developed three cpSNP markers are applicable to population and phylogeographic study of teak. Anyhow, the better resolution of cpSNP markers of teak should be increased to see the clear picture of geographic structure. Furthermore, those chloroplast SNP markers might cross amplify in related species; *Tectona hamiltonia* and *Tectona philipino*.

It was unclear for weak geographic variation of Myanmar teak for cpSNP markers. Generation time is one of the biological factors influence on rates of nucleotide sequence evolution (Wu and Li, 1985). A prolonged or severe demographic bottleneck in recent times might have resulted in low haplotype diversity and nucleotide diversity (Avise, 2000). The widespread distribution of common haplotype indicated ancient population bottleneck (Liu, et al., 2012). Myanmar teak possessed low haplotype diversity and widely distribution of common haplotype (H1). Thus, severe bottleneck after glaciations or rapid expansion of founder populations or long life span of teak or human interference to natural populations might account for weak cpDNA variation of Myanmar teak. Only the genetic data is insufficient to distinguish between natural dispersal and migration (Gong, et al., 2008). Fossil data and genetic data are required to confirm the human propagation, natural dispersal and migration of teak.

Clear genetic structure of Myanmar teak for nrSSR markers showed limited pollen flow of teak. Both economical and ecological traits are largely varied for different provenances (Keiding, et al., 1986, Kjaer, et al., 1999, Monteuuis, et al., 2011). Three major zones are proposed to designate based on genetic structure of teak revealed by

nrSSR markers. In addition to the knowledge of genetic structure, natural climatic and physiographic divisions should be considered for designating the boundaries of seed zones (Ledig, 1992). The boundary should therefore be made between upper and lower regions due to the difference in climatic conditions. However, provenance test should be conducted to designate profoundly the seed zones, breeding zones or plantation zones. High genetic diversity within populations of Myanmar teak indicated its importance for conservation and breeding purposes. The populations with high level of genetic diversity have the adaptability the capacity of rapid adaptive changes (Lefevre, et al., 2004). Thus, HMB, TDG, POL and KTA should be conserved not only for retaining the natural genetic resources of Myanmar teak but also for breeding programs to use as raw populations.

Teak is an important source of tropical timber and planted not only in its native region but also outside of its natural distribution. Teak plantation stands on the third position of world plantation to supply the demand of timber market because of decreasing the capacity of natural teak bearing forest. Expanding the ranges of species of economic value may lead to genetic divergence and mixing divergent populations will contaminate local gene pools and homogenizing species structure (Ledig, 1992). Gene flow of foreign genes into natural populations, exotic or genetically modified plants by hybridization and introgression can cause genetic pollution (Linacre and Ades, 2003). Secondary evolution can occur through the hybridization between the indigenous species and related exotic populations (Lefevre, et al., 2004). Furthermore, non-local tree cross with native populations may increase the genetic diversity of next generation but with negative consequences for local adaptation (Ledig, 1992). Thus, to retain the natural genetic resource of Myanmar

teak and also to supply the demand, the best way is balancing conservation and breeding program of teak. Timber quality of Myanmar teak is famous for the best timber quality in the world. Conserved populations with high genetic diversity and genetic divergence can be use as breeding population that satisfied major economic needs. Therefore, instead of introducing the alien teak for plantations, producing the genetically superior quality of planting materials should be concentrated by implementing the breeding program for Myanmar teak.

## Summary

The tropical deciduous and semi ever-green tree species, teak, is one of the most economically important tree species. It naturally occurs in India, Myanmar, Thai and Laos. Genetic information of teak from its native regions has been investigated using molecular markers and they showed south India teak has the highest genetic diversity followed by teak from North India, Thai and Laos. About 60% of the total natural forest area occurs in Myanmar that is the largest area of natural teak forest. Few study for genetic diversity of Myanmar teak has been conducted but no comparison between Myanmar teak and from its indigenous countries has been reported. The same markers used in the previous study were applied for evaluating the level of genetic diversity of Myanmar teak to compare with that of teak from other native countries in chapter 2. As the results, Myanmar teak has significantly lower genetic diversity than that of India teak, but significantly higher than that of Thai and Laos teak.

Natural teak forest in Myanmar drastically diminished due to over logging, illegal cutting and transforming landuse systems, therefore conservation of Myanmar teak is urgently needed to retain the natural genetic resources of teak in the world. Furthermore, teak plantation was widely established at about 43 countries including its native countries, Myanmar. Knowledge of the genetic variation of extant populations over the entire range of their distribution is therefore essential for the conservation of genetic resources. Microsatellite markers (nrSSRs), which are highly polymorphic, are useful for elucidating the spatial genetic structure and the demographic patterns of variation which have resulted from migration and drift as well as through evolutionary history. Chloroplast markers are also useful for

phylogeographic studies and gene conservation, because chloroplast genomes, which are haploid, are maternally inherited in angiosperms and hence transmitted by seeds. Nevertheless, no chloroplast markers for teak have been developed yet. Thus, cpSNP markers for teak have been developed to determine phylogeographic structure of Myanmar teak. After sequencing about one third of complete genome of teak about 43,734 bp, three cpSNP markers of teak were developed to study the geographic variation of teak in Myanmar.

Geographically genetic structure of Myanmar teak was examined using total 480 individuals of 20 natural populations from five regions representing almost natural teak forests in Myanmar and two types of molecular markers; three newly developed cpSNP markers and 10 nrSSR markers in chapter 4. The combined studied of cpSNP and nrSSR markers suggested there are at least four genetic resources of Myanmar teak. Randomized distribution of four haplotypes showed by cpSNP markers did not depicted clear geographic structure of Myanmar teak. On the other hand, four genetic clusters of 20 natural populations depicted by nrSSR markers suggested clear geographic genetic structure of Myanmar teak. The putative genetic boundaries of 20 populations suggested at least three zones such as planting or seed zones can be designated based on combined cpSNP and nrSSR data. Of 20 populations, four populations with their high contribution to total genetic diversity were found to be prioritized for conservation.

Teak plantation in Myanmar has been started using local seeds since 1700 to replenish the degraded natural forests. A couple of years ago, private sectors were allowed to establish teak plantation at deforested area or some were around natural teak forests. No seed guideline of teak is formulated in Myanmar. Therefore, seeds



from wherever available were used for teak plantation without considering their genetic component. Moreover, teak plantation established by private companies used alien teak from Indonesia, China and Costa Rica without information on genetic background. To prevent genetic disturbance for Myanmar natural teak, genetic component of recently established teak plantation by private sectors were investigated using 10 nrSSR markers and compared with that of natural teak and old teak plantation. Higher genetic diversity and less genetic differentiation among populations of recently established teak plantation supported the assumption of various seeds sources used for those plantations. Alien teak showed low genetic diversity and significant level of genetically differentiated from Myanmar teak especially Indonesian teak.

At last, gene conservation and afforestation strategy for Myanmar teak were discussed based on findings obtained in this study. Among four native countries of teak, Myanmar with the largest natural teak forests and high genetic diversity may be genetic core of teak in the world. The current four genetic resources of Myanmar teak should be retained not to be deteriorated by genetic erosion by designating the planting zones or seeds zone based on geographic genetic structure of Myanmar teak. Alien teak introduced to Myanmar for planting purpose should be restricted. Seeds from alien teak should be avoided for the establishment of next teak plantation in Myanmar because those seeds may be products of outbreeding between alien teak and Myanmar teak with high genetic divergence. Instead of using the alien teak, genetically improved planting materials, Myanmar teak should be focused on producing the planting materials through breeding and tree improvement programs. In doing so, retaining natural genetic resources of Myanmar and supplying the high

demand of teak can be implemented. Genetic information of Myanmar teak observed in this study may take a part of role for the conservation of natural genetic resource of teak.

## References:

- Anon. (1992) Forestry fact sheets. Forest Resources Division, Forest Department, Myanmar
- Ansari, S.A., Narayanan, C., Wali, S.A., Kumar, R., Shukla, N. and Rahangdale, S.K. (2012) ISSR markers for analysis of molecular diversity and genetic structure of Indian teak (*Tectona grandis* L.f) populations. *Annals of forest research* 55: 11-23.
- Antao, T., Lopes, A., Lopes R.J., Beja-Pereira, A., Luikart, G. (2008) Lositan: A workbench to detect molecular adaptation based on Fst-outlier method. *BMC Bioinformatics* 9:323-328. doi:10.1186/1471-2105-9-323
- Asmussen, C.B. and Chase, M.W. (2001) Coding and noncoding plastid DNA in palm systematic. *American journal of botany* 88:1103-1117.
- Aoki, K., Suzuki, T., Hsu, T.W. and Murakami, N. (2004) Phylogeography of the component species of broad-leaved evergreen forests in Japan, based on chloroplast DNA variation. *Journal of Plant Research* 117: 77-94.
- Avise, J. (2000) *Phylogeography. The history and formation of species.* Harvard University press. Cambridge, Massachusetts London, England.
- Bertrand, G. and Rangin, C. (2003) Tectonics of the Western margin of the Shan plateau (Central Myanmar): implication for the India-Indonesia oblique convergence since the Oligocene. *Journal of Asian Earth Science* 21:1139-1157.
- Birky, C.W. (1995) Uniparental inheritance of mitochondrial and chloroplast genes: mechanisms and evolution. *Proceedings of the National Academy of Sciences* 92:11331-11338.
- Bryan, G.J., McNicoll, J., Ramsay, G., Meyer, R.C. and Jong, W.S.D. (1999) Polymorphic simple sequence repeat markers in chloroplast genome of *Solanaceous*

- plants. *Theoretical Applied Genetic*. 99:859-867.
- Bryant, R.L. (1997) *The political ecology of forestry in Burma, 1824-1994*. London, UK: C. Hurst and Co.:257 pp
- Buiteveld, J. and Koelewijn, H. (2006) CpDNA haplotype variation reveals strong human influence on oak stands of the Veluwe forest in The Netherlands. *Forest Ecology and Management* 228: 160-167.
- Clark, P.U., Dyke, A.S., Shakun, J.D., Carlson, A.E., Clark, J., Wohlfarth, W. and McCabe, A.M. (2009) The last glacial maximum. *Science* 325:710-714.
- Clement, M., Posada, D. and Crandall, K.A. (2000) TCS: a computer program to estimate gene genealogies. *Molecular Ecology* 9:1657-1659.
- Clerc-Blain, J., Starr, J., Bull, R.D. and Saarela, J.M. (2010) A regional approach to plant DNA barcoding provides high species resolution of sedges (*Carex* and *Kobresia*, Cyperaceae) in the Canadian Arctic Archipelago. *Molecular Ecology Resources* 10:69-91.
- Desplanque, B., Viard, F., Bernar, J., Forcioli, D., Saumitou-Laprade, P., Cuguen, J. and Van, D.H. (2000) The linkage disequilibrium between chloroplast DNA and mitochondrial DNA haplotypes in *Beta vulgaris* ssp. *Maritima* (L.): the usefulness of both genomes for population genetic studies. *Mol. Ecol.* 9:141-154.
- Downie, S.R. and Palmer, J.D. (1992) Use of chloroplast DNA rearrangement in reconstruction plant phylogeny. In Soltis et al. [eds.], *Molecular systematic of plants*, 1-13. Chapman and Hall, New York, New Yourk, USA.
- Duran, C., Appleby, N., Vardy, M., Imelfort, M., Edwards, D. and Batley, J. (2009) Single nucleotide polymorphism discovery in barley using autoSNPdb. *Plant Biotechnology Journal* 7:326-333.

Evanno, G., Regnaut, S. and Goudet, J. (2005) Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Molecular Ecology* 14:2611-2620.

Excoffier, L. and Lischer, H.E. (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular ecology resources* 10: 564-567.

Falconer, D.S. (1981) *Introduction to Quantitative Genetics*, 2<sup>nd</sup> ed. Longman, New York.

Fofana, I.J., Lidah, Y.J., Diarrassouba, N., N'guetta, S.P.A., Sangare, A. and Verhaegen, D. (2008) Genetic structure and conservation of teak (*Tectona grandis*) plantations in Cote d'Ivoire, revealed by site specific recombinase (SSR). *Tropical Conservation Science* 3:279-292.

Fofana, I.J., Ofori, D., Poitel, M. and Verhaegen, D. (2009) Diversity and genetic structure of teak (*Tectona grandis* L.f) in its natural range using DNA microsatellite markers. *New Forests* 37:175-195.

Fofana, I.J., Silue, S., Diarrassouba, N., Kadio, A.A. and Sangare, A. (2013) Comparative analyses of amplified fragment length polymorphism (AFLP) and simple sequences repeat (SSR) in genetic diversity of teak (*Tectona grandis* L.f). *International Journal of Advance Agricultural Research* 1:114-123.

Ford, C.S., Ayres, K.L., Toomey, N., Haider, N., van Alphen Stahl, J., Kelly, L.J., Wikstrom, N., Hollingsworth, P.M., Duff, R.J., Hoot, S.B., Cowan, R.S., Chase, M.W. and Wilkinson, M.J. (2009) Selection of candidate coding DNA barcoding regions for use on land plants. *Botanical Journal of Linnean Society* 159:1-11

Fujii, N., Tomaru, N., Okuyama, K., Koike, T., Mikami, K. and Ueda, K. (2002)

Chloroplast DNA phylogeography of *Fagus crenata* (Fagaceae) in Japan. *Plant System Evolution* 232:21-33.

Gaut, B.S. (1998). Molecular clocks and nucleotide substitution rates in higher plants. In M.K. Hech, R.J. MacIntyre, and M.T. Clegg [eds.], *Evolutionary biology*, vol. 30, 93-120. Plenum Press, New York, New York, USA.

Gill, B.S., Bedi, Y.S. and Bir, S.S. (1983) Cytopalynological studies in woody members of family Verbenaceae from north-west and central India. *J. India Botany Soc.* 62:235-244.

Golden, J. L. and Bain, J. F. (2000) Phylogeographic patterns and high levels of chloroplast DNA diversity in four *Packera* (Asteraceae) species in southwestern Alberta. *Evolution* 54: 1566-1579.

Gong, W., Chen, C., Dobes, C., Fu, C.X. and Koch, M.A. (2008) Phylogeography of a living fossil: Pleistocene glaciations forced *Ginkgo biloba* L. (Ginkgoaceae) into two refuge areas in China with limited subsequent postglacial expansion. *Molecular Phylogenetics and Evolution* 48:1094-1105.

Goudet, J. (2001) FSTAT, version 2.9.3, A program to estimate and test gene diversities and fixation indices. *Journal of Heredity* 86: 485-486 Lausanne University, Lausanne, Switzerland.

Gupta, P.K. and Kumar, A. (1976) Estimating of potential germinability of teak (*Tectona grandis* L.f.) fruits from twenty three Indian sources by cutting test. *Indian Forester* 102:808-813.

Gunaga, R.P., Surendran, T. and Prabhu, H.N. (2013) Morphological Variation and Delineation of Teak (*Tectona grandis* L. F.) Clones of Kerala through Leaf Character: Implication for Seed Orchard Management. *Mysore J. Agric, Sci.*, 47: 202-205.

Gyi, K.K. and Tint K. (1998) Management status of natural teak forests in Teak for future. In: Masakazu K, Kevin W (eds) Teak for the future, Dharmasarn, Bangkok, pp27-48.

Hadden, R.L. (2008) The Geology of Burma (Myanmar): An Annotated Bibliography of Burma's Geology, Geography and Earth Science, DTIC Document. Pp312. Accession Number: ADA487552.

Hamza N.B. (2010) Cytoplasmic and nuclear DNA markers as powerful tools in populations' studies and in setting conservation strategies. African Journal of Biotechnology 9:4510-4515.

Han, J.E., Kook-Hyun, C., Nemoto, T. and Byoung-Hee, C. (2010). Phylogenetic analysis of eastern Asian and eastern American disjunct *Lespedeza* (Fabaceae) inferred from nuclear ribosomal ITS and plastid region sequences. Botanical journal of the Linnean Society 164:221-235.

Hasen, O.K., Changtragoon, S., Ponooy, B., Kjaer, E.D., Minn, Y., Finkeldey, R., Nielsen, K.B. and Graudal, L. (2015) Genetic resources of teak (*Tectona grandis* Linn.f.) – strong genetic structure among natural populations. Tree Genetics and Genomes 11:802-818. DOI 10.1007/s11295-014-0802-5.

Hebel, I., Haas, R. and Dounavi, A. (2006) Genetic variation of common ash (*Fraxinus excelsior* L.) populations from provenance regions in southern Germany by using nuclear and chloroplast microsatellites. Silvae Genetica 55: 38-43.

Hewitt, G. (2004) Genetic consequences of climatic oscillations in the Quaternary. Philosophical Transactions of the Royal Society of London. Series B: Biological Science 359:183-195.

Hufford, K.M, and Mazer, S.J. (2003) Plant ecotypes: genetic differentiation in the

- age of ecological restoration. *Trends in Ecology and Evolution* 18:147–155.
- Jennings, N.R., Faratin, P., Lomuscio, A.R., Parsons, S., Wooldridge, M. and Sierra, C. (2001) Automated negotiation: Prospects, methods and challenges. *Group Decision and Negotiation* 10:199-215.
- Kaosa-ard, A. (1991) Country report in Thailand, Paper presented at the China/ESCAP/FAO regional seminar on Teak, Guangzhou, China.
- Kaosa-ard, A. (1998) Overview of problems in teak plantation establishment.
- Kaosa-ard, A. (2003) Teak breeding and improvement strategies. Proceeding of the Second Regional Seminar on teak, 29 May-3 June, 1995, Yangon, Myanmar. Pp 61-82.
- Keiding, H., Wellendorf, H. and Lauridsen, E.B. (1986) Evaluation of an international series of teak provenance trials. DANIDA Forest Seed Centre, pp 81.
- Kermode, C.W.D. (1964) Some aspects of silviculture in Burma. Central press, Yangon.
- Kertakikara, A.W.S. and Prat, D. (1994) Genetic structure and mating system in teak (*Tectona grandis* L.f.) provenances. *Silvae Genetica* 44:104-110.
- Kertadikara, A.W.S. and Prat, D. (1995) Isozyme variation among teak (*Tectona grandis* L.f) provenances. *Theoretical and Applied Genetic* 90:803-810.
- King, R.A. and Ferris, C. (1998) Chloroplast DNA phylogeography of *Alnus glutinosa* (L.) Gaertn. *Molecular Ecology* 7:1151-1161.
- Khanduri, V.P., Lalnundanga, and Vanlalremkimi, J. (2008) Growing stock variation in different teak (*Tectona grandis*) forest stands of Mizoram, India. *Journal of Forestry Research* 19:204-208.
- Kjaer, E.D., Lauridsen, E.B. and Wellendorf, H. (1995) Second evaluation of an



international series of teak provenance trials. DANIDA Forest Seed Centre, [www.sl.life.ku.dk](http://www.sl.life.ku.dk).

Kjaer, E.D. and Siegismund, H.R. (1996) Allozyme diversity in two Tanzanian and two Nicaraguan landraces of teak (*Tectona grandis* L.). *Forest Genetic* 1:45-52.

Kjaer, E.D., Kajornsrichon, S. and Lauridsen, E.B. (1999) Heartwood, calcium and silica content in five provenance teak (*Tectona grandis* L.). *Silvae Genetica* 48:1-3.

Knapp, E. E. and Rice, K.J. (1994) Starting from seed: genetic issues in using native grasses for restoration. *Restoration and Management Notes* 12:40–45.

Korpelainen, H. (2004) The evolutionary processes of mitochondrial and chloroplast genomes differ from those of nuclear genomes. *Naturwissenschaften* 91:505–518.

Lammi, A., Siikamaki, P. and Mustajarvi, K. (1999) Genetic diversity, population size, and fitness in central and peripheral populations of a rare plant *Lychnis viscaria*. *Conservation Biology* 13:1069-1078.

Leberg, P.L. (2002) Estimating allelic richness: Effects of sample size and bottlenecks. *Molecular Ecology* 11:2445-2449.

Ledig, T. (1992) Human impacts on genetic diversity in forest ecosystems. *OIKOS* 63:87-106.

Lefevre, F., Fady, B., Fallour-Rubio, D., Ghosn, D. and Bariteau, M. (2004) Impact of founder populations, drift and selection on the genetic diversity of recently translocated tree population. *Heredity* 93:542-550.

Linacre, N.A. and Ades, P.K. (2003) Estimating isolation distances for genetically modified trees in plantation forestry. *Ecological modeling* 179:247-257.

Liu, H.-Z., Takeichi, Y., Kamiyha, K. and Harada, K. (2012) Phylogeography of *Quercus phillyraeoides* (Fagaceae) in Japan as revealed by chloroplast DNA

variation. Journal of forest research: 1-10.

Lowe, A.J., Harris, D., Dormontt, E. and Dawson, I.K. (2010) Testing putative African tropical forest refugia using chloroplast and nuclear DNA phylogeography. *Tropical Plant Biology* 3:50-58.

Manni, F., Guérard, E. and Heyer E. (2004) Geographic patterns of (genetic, morphologic, linguistic) variation: how barriers can be detected by “Monmonier’s algorithm”. *Human Biology* 76: 173-190.

Mathew, G., Mathew, P.K. and Mohanadas (1987) Preliminary studies on insects visitors to teak (*Tectona grandis* L.f.) inflorescence in Kerala, India. *Indian Forest*. 11361-64.

Maung, T.M. and Yamamoto, M. (2008) Exploring the socio-economic situation of plantation villagers: a case study in Myanmar Bago Yoma. *Small-scale Forestry* 7: 29-48.

McKay, J.K., Christian, C.E. and Harrison, C.E. (2005) How local is local?- A review of practical and conceptual issue in the genetic of restoration. *Restoration Ecology* 13:432-440.

Miettinen, J., Shi, C. and Liew, S.C. (2011) Deforestation rates in insular Southeast Asia between 2000 and 2010. *Global Change Biology* 17:2261-2270.

Minn Y., Prinz, K. and Finkeldey, R. (2014) Genetic variation of teak (*Tectona grandis* Linn.) in Myanmar revealed by microsatellite markers. *Tree Genetics and Genomes* DOI 10.1007/s11295-014-0772-7.

Missiaggia, A. and Grattapaglia, D. (2006) Plant microsatellite genotyping with 4-color fluorescent detection using multiple-tailed primers. *Genetic and Molecular Research* 5: 72-78.

- Monteuuis, O., Goh, D.K.S., Garcia, C., Alloysius, D., Gidiman, J., Bacilieri, R. and Chaix, G. (2011) Genetic variation of growth and tree quality traits among 42 diverse genetic origins of *Tectona grandis* planted under humid tropical conditions in Sabah, East Malaysia. *Tree Genetics & Genomes* 7: 1263-1275.
- Montalvo, A. M., Williams, S. L., Rice, S.L., Buchmann, S.L., Cory, C., Handel, S.N., Nabhan, G.P., Primack, R. and Robichaus, R.H. (1997). Restoration biology: a population biology perspective. *Restoration ecology* 5: 277-290.
- Montalvo, A.M. and Ellstrand, N.C. (2000) Transplantation of the Subshrub *Lotus scoparius*: Testing the Home-Site Advantage Hypothesis. *Conservation biology* 14: 1034-1045.
- Morgenstern, E.K. (1996) *Geographic Variation in Forest Trees*. UBC Press, Vancouver, 209 pp, ISBN 0-7748-0579-X
- Narayanan, C., Wali, S.A., Shukla, N., Kumar, R., Mandal, A.K. and Ansari, A. (2007) RAPID and ISSR markers for molecular characterization of teak (*Tectona grandis*) plus trees. *Journal of Tropical Forest Science* 19:218-225.
- Neale, D.B. and Ingvarsson, P.K. (2008) Population, quantitative and comparative genomics of adaptation in forest trees. *Current Opinion in Plant Biology* 11:149-155.
- Neale, D.B. and Kremer, A. (2011) Forest tree genomics: growing resources and applications. *Nature Reviews Genetics* 12:111-122.
- Newton, A., Allnutt, T., Gillies, A., Lowe, A. and Ennos, R. (1999) Molecular phylogeography, intraspecific variation and the conservation of tree species. *Trends in Ecology and Evolution* 14:140-145
- Nicolosi, E., Deng, Z.N., Gentile, A., Malfa, S.L. and Tribulato, G.C. (2000) Citrus phylogeny and genetic origin of important species as investigated by molecular

marker. *Theoretical Applied Genetic* 100:1155-1166.

Ohtani, M., Kondo, T., Tani, N., Ueno, S., Lee, L.S., Ng, K.S., Muhammad, N., Finkeldey, R., Naiem, M., Indrioko, S., Kamiya, K., Harada, K., Diway, B., Khoo, E., Kawamura, K. and Tsumura, Y. (2013) Nuclear and chloroplast DNA phylogeography reveals Pleistocene divergence and subsequent secondary contact of two genetic lineages of the tropical rainforest tree species *Shorea leprosula* (Dipterocarpaceae) in South- East Asia. *Molecular Ecology* 22:2264-2279.

Palmer, J.D., Jansen, R.K., Michaels, H.J., Chase, M.W. and Manhart, J.R. (1988) Chloroplast DNA variation and plant phylogeny. *Annals of the Missouri Botanical Garden* 75:1180-1206. <http://www.nal.usda.gov/>

Pakkad, G., Ueno, S. and Yoshimaru, H. (2008) Genetic diversity and differentiation of *Quercus semiserrata* Roxb. in northern Thailand revealed by nuclear and chloroplast microsatellite markers. *Forest Ecology and Management* 255:1067-1077.

Pandy, D. and Brown, C. (2000) Teak: a global overview. *Unasylva* 201(51):3-13.

Peakall, R. and Smouse, P.E. (2012) GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Bioinformatics Application Notes* 28:2537-2539. doi:10.1093/bioinformatics/bts460

Perum, P. (1993) Teak in Indonesia. FORSPA publication No. 4: Teak in Asia.

Pengduoang, V. (1991) Teak in Laos. Country Report. Paper presented at the China/ESCAP/FAP regional seminar on teak, Guangzhou, China

Petit, R.J., Mousadik, A.E. and Pons, O. (1998) Identifying populations for conservation on the basis of genetic markers. *Conservation Biology* 12:844-855.

Petit, R.J., Aguinagalde, I., Beaulieu, J.L., Bittkau, C., Brewer, S., Cheddadi, R., Ennos, R., Fineschi, S., Grivet, D, Lascoux M, Mohanty A, Muller-Starck G,

Demesure-Musch B, Palme, Martin, A., J.P., Rendell, S. and Vendramin, G.G. (2003) Glacial refugia: hotspots but not melting pots of genetic diversity. *Science* 300:1563-1565.

Petit, R.J., Duminil, J., Fineschi, S., Hampe, A., Salvini, D. and Vendramin, G.G. (2005) Comparative organization of chloroplast, mitochondrial and nuclear diversity in plant populations. *Molecular Ecology* 14:689-701.

Petit, R.J. and Vendramin, G.G. (2007) Phylogeography of organelle DNA in plants: an introduction. In Weiss S, Ferrand N. eds. *Phylogeography of southern European Refugia*. Springer, pp.23-97.

Piry, S., Luikart, G. and Cornuet, J.M. (1999) Bottleneck: a computer program for detecting recent reduction in the effective population size using allele frequency data. *Journal of Heredity* 90: 502–503.

Pons, O. and Petit, R.J. (1996) Measuring and testing genetic differentiation with ordered versus unordered alleles. *Genetics* 144:1237-1245.

Powell, W., Morgante, M., Andre, C., Hanafey, M., Vogel, J., Tingey, S. and Rafalski, A. (1996) The comparison of RFLP, RAPD, AFLP and SSR (microsatellite) markers for germplasm analysis. *Molecular Breeding* 2:225-238.

Pritchard, J.K., Stephens, M. and Donnelly, P. (2000) Inference of population structure using multilocus genotype data. *Genetics* 155:945-959. Priya, B.P. and Bhat, K.M. (1999) Influence of rainfall, irrigation and age of the growth periodicity and wood structure in teak (*Tectona grandis*). *IAWA journal* 20:181-192.

Rajora, O, M. Rahman, Buchert, G.P. and Dancik, B.P. (2000). Microsatellite DNA analysis of genetic effects of harvesting in old growth eastern white pine (*Pinus strobus*) in Ontario, Canada. *Molecular Ecology* 9: 339-348.

- Rauch, E.M. and Bar-Yam, Y. (2005) Estimating the total genetic diversity of a spatial field population from a sample and implications of its dependence on habitat area. *Proceedings of the National Academy of Sciences of the United States of America USA* 102:9826-9829.
- Rinna J., Thong, H.L., Leong, L.S., Judy, L. and Laura, S. (2014) Integrating genetic factors into management of tropical Asian production forests: A review of current knowledge. *Forest Ecology and Management* 315:191-201.
- Rosane, G.C., Grattapaglia, D. and John, D.H. (2003) Evidence for multiple maternal lineages of *Caryocar brasiliense* populations in the Brazilian Cerrado based on the analysis of chloroplast DNA sequences and microsatellite haplotype variation. *Molecular Ecology* 12: 105-115.
- Saw, E.D. (2003) Sustainable management of teak forests in Myanmar. *Proceeding of the International Conference on Quality Timber Products of Teak from Sustainable Forest Management, Peechi, India, 2-5 December 2003. Pp 135-142*
- Sakaguchi, S., Qiu, Y.X., Liu, Y.H., Qi, X.S., Kim, S.H., Han, J., Takeuchi, Y., Worth, J.R.P., Yamasaki, M., Sakurai, S. and Isagi, Y. (2012) Climate oscillation during the Quaternary associated with landscape heterogeneity promoted allopatric lineage divergence of a temperate tree *Kalopanax septemlobus* (Araliaceae) in East Asia. *Molecular Ecology* 21: 3823-3838.
- Saltonstall, K. (2002) Cryptic invasion by a non-native genotype of the common reed, *Phragmites australis*, into North America. *Proceedings of the National Academy of Sciences* **99**: 2445-2449.
- Samuel, R., Kathriarachchi, H., Hoffman, R., Barfuss, M.H.J., Wudarck, K.J., Davis, C.C. and Chase, M.W. (2005) Molecular phylogenetics of Phyllanthaceae: Evidence

from plastid MATK and nuclear PHYC sequences. *American Journal of Botany* 92:132-141.

Schneider, S., Roessli, D. and Excoffier, L. (2000) Arlequin ver. 2.000. A software for population genetics data analysis Genetics and Biometry Laboratory, University of Geneva, Switzerland. Pp.111. <http://anthro.unige.ch/arlequin>

Shaw, J., Kickey, E.B., Beck, J.T., Farmer, S.B., Liu, W., Miller, J., Siripun, K.C., Winder, C.T., Schilling, E.E. and Small, R.L. (2005) The tortoise and the hare II: Relative utility of 21 noncoding chloroplast DNA sequences for phylogenetic analysis. *American Journal of Botany* 92:142-166.

Shaw, J., Lickey, E.B., Schilling, E.E. and Small, R.L. (2007) Comparison of whole chloroplast genome sequences to choose noncoding regions for phylogenetic studies in angiosperms: The tortoise and the hare III. *American Journal of Botany* 94:275-288.

Shen, L., Chen, X., Zhang, X., Li, Y., Fu, C. and Qiu, Y. (2004) Genetic variation of *Ginkgo biloba* L. (Ginkgoaceae) based on cpDNA PCR-RFLPs: inference of glacial refugia. *Heredity* 94:396-401.

Shiraishi, S. and Watanabe, A. (1995) Identification of chloroplast genome between *Pinus densiflora* Sieb. et Zucc. and *P. thunbergii* Parl. based on the polymorphism in *rbcL* gene. *Journal of Japanese Forest Society* 77:429-436 (in Japanese with English summary).

Shrestha, M.K., Volkaert, H. and Straeten, D.V.D. (2005) Assessment of genetic diversity in *Tectona grandis* using amplified fragment length polymorphism markers. *Canadian Journal Forest Research* 35:1017-1022.

Soltis, D.E., Matthew, A.G., Strenge, D.D. and Soltis, P.E. (1997) Chloroplast DNA

intraspecific phylogeography of plants from the Pacific Northwest of North America. *Plant Systematics and Evolution* 206:353-373.

Srrekanth, P.M., Balasundaran, M., Nazeem, P.A. and Suma, T.B. (2012) Genetic diversity of nine natural *Tectona grandis* L.f. populations of the Western Ghats in Southern India. *Conserv Genet.* DOI 10.1007/s10592-012-0383-5.

Steven, T.K. (2004) Counting alleles with rarefaction: Private alleles and hierarchical sampling designs. *Conservation Genetics* 5:539-543.

Swell, M.M., Parks, C.R., and Chase, M.W. (1996) Intraspecific chloroplast DNA variation and biogeography of North American *Liriodendron* L. (Mangoliaceae). *Evolution* 50:1147-1154.

Tanah, L.H., Lee, S.L., Ng, K.K., Lee, C.T., Bhassu, S. and Othman, R.Y. (2013). Phylogeographical Pattern and Evolutionary History of an Important Peninsular Malaysian Timber Species, *Neobalanocarpus heimii* (Dipterocarpaceae). *Journal of Heredity* 104:115-126.

Tangmitcharoen, S. and Owens, J.N. (1997) Floral biology, pollination, pistil receptivity, and pollen tube growth of teak (*Tectona grandis* Linn f.) *Annals of Botany* 79:227-241.

Tarayre, M., Pierre, S.L., Cuguen, J., Couver, D. and Thompason, J.D. (1997) The spatial genetic structure of cytoplasmic (cpDNA) and nuclear (allozyme) markers within and among populations of the gynodioecious *Thymus vulgaris* (Labiatae) in southern France. *American Journal of Botany* 84:1675-1684.

Tewari, D.N. (1992) A monograph on teak (*Tectona grandis* Linn.f).

Verhaegen, D., Ofori, D., Fofana, I.J., Poitel, M. and Vaillant, A. (2005) Development and characterization of microsatellite markers in *Tectona grandis*



(Linn.f). *Molecular Ecology* 5:945-947.

Vogel, M., Banfer, G., Moog, U. and Weising, K. (2003) Development and characterization of chloroplast microsatellite markers in *Macaranga* (Euphorbiaceae). *Genome* 46:845-857.

White, K.J. (1991) Teak: some aspects of research and development. Publication 1991/17. FAO Regional office for Asia and the Pacific (RAPA). Bangkok.

White, T.L., Adams, W.T. and Neale, D.B. (2007) *Forest Genetics*. CABI publishing, Cambridge, 682pp, ISBN 978-0-85199-083-5

Wolfe, K.H., Li, W.H. and Sharp, P.M (1987) Rates of nucleotide substitution vary greatly among plant mitochondrial, chloroplast, and nuclear DNAs. *Proceeding of National Academic Science USA*. 84:9054-9058.

Wolfe, K.H., Mordern, C.W. and Palmer, J.D. (1992) Function and evolution of a minimal plastid genome from a non-photosynthetic parasitic plant. *Proceeding of the National Academy of Science, USA* 89:10648-10652.

Wright, S.J. (2005) Tropical forests in a changing environment. *Trends in Ecology and Evolution* 20:553-560.

Wu, C.I. and Li, W.H. (1985) Evidence for higher nucleotide substitution in Rodent than in man. *Evolution* 82:1741-1745.

Zuo, Y., Chen, Z., Kondo, K., Funamoto, T., Wen, J. and Zhou, S. (2010) DNA barcoding of *Panax* species. *Planta Medica* 77:182-187.

## List of tables

Table 2.1 Geographic and climatic information of four natural populations of teak in Myanmar	16
Table 2.2 Genetic information of 15 SSR markers across four natural populations of Myanmar teak	21
Table 2.3 Statistical comparison of genetic diversity estimates between Myanmar teak and Indian, Thai and Laotian teak	23
Table 3.1 List of (58) walking primers used for finding chloroplast polymorphism in teak from Myanmar	34
Table 3.2 Locus-specific and extension primers used for SnaPshot genotyping	37
Table 4.1 Location and sample size of 20 natural populations of teak from Myanmar and sample size collected from each population	42
Table 4.2 Polymorphism sites and cpDNA haplotypes based on SnaPshot analysis	50
Table 4.3 Statistical summary of the diversity revealed using cpSNP and nrSSR markers for 20 teak populations	55
Table 4.4 Results of AMOVA analysis for cpSNP and nrSSR markers	66
Table 5.1 Genetic diversity parameters and fixation index of each population and each group by 10 nrSSR markers	80
Table 5.2 Pairwise genetic differentiation between populations	82
Table 5.3 Hierarchical analysis of AMOVA for two groups; alien teak and native teak	86

## List of figures

Figure 1.1 Natural distribution of teak in its native regions, India, Myanmar, Laos and Thailand	10
Figure 1.2 Natural teak bearing forests in Myanmar	11
Figure 1.3 Color variation of teak	12
Figure 2.1 (a) Maps of the distribution of teak in India, Myanmar, Laos and Thailand and (b) the locations of the four sampled populations of teak in Myanmar	17
Fig. 2.2 Distribution of genetic diversity parameter of teak for allelic richness	24
Fig 2.3 Distribution of genetic diversity parameter of teak for expected heterozygosity	25
Fig 3.1 Complete genome of teak	38
Figure 4.1 Location of 20 natural populations of teak in Myanmar	43
Figure 4.2 Haplotype network detected in Myanmar teak	51
Figure 4.3 Haplotype distribution in 20 natural populations of Myanmar teak.	52
Figure 4.4 Genetic clusters of 20 populations from five regions of Myanmar revealed at nrSSR markers	56
Figure 4.5 Genetic structure of Myanmar teak revealed at nrSSR markers	57
Figure 4.6 Scattergram of allelic richness and nucleotide diversity of 20 natural populations	58
Fig 4.7 Scattergram of the nucleotide diversity and the expected heterozygosity of 20 populations	59

Fig 4.8 Contribution to total genetic diversity of each population due to genetic diversity and genetic divergence of population	60
Figure 4.9 Putative genetic boundaries of teak estimated by (a) cpSNP, (b) nrSSR, and (c) cpSNP and nrSSR markers	62
Fig 4.10 Zonation of teak in Myanmar	63
Fig 4.11 Isolation by distance analysis for cpSNP markers	64
Fig 4.12 Isolation by distance analysis for nrSSR markers	65
Figure 5.1 Location of sampled populations from teak private plantations, old plantations and natural populations	76
Fig 5.2 Scatter plot of individual based principal coordinate analysis for alien teak and native teak	83
Fig 5.3 Scatter plot of population based principal coordinate analysis for alien teak and native teak	84
Fig 5.4 Proportion of genetic component of each population from alien teak and native teak revealed by 10 SSRs	85