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Review Article

Coordinate action of membrane-type matrix metalloproteinase-1 (MT1-MMP) and MMP-2 enhances pericellular proteolysis and invasion

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Membrane-type matrix metalloproteinase-1 (MT1-MMP) mediates cleavage of not only MMP-2/gelatinase A for activation, but also a variety of substrates including type I collagen (reviewed in *Cancer Sci* 2005; 96: 212–7). MMP-2 activation involves tissue inhibitor of MMP (TIMP)-2 as a bridging molecule between MT1-MMP and pro-MMP-2. Thus, net activity of MT1-MMP and MMP-2 is regulated in a complex manner depending on TIMP-2 concentration. During invasive growth of tumor cells in type I collagen matrix, MT1-MMP initiates denaturation of collagen into gelatin, which is subsequently digested further by MMP-2 adjacent to MT1-MMP. Coordinate action of MT1-MMP and MMP-2 may facilitate pericellular proteolysis, and enhance not only tumor invasion/migration but also cell growth. Tetraspanins as binding proteins of MT1-MMP regulate MT1-MMP subcellular localization and compartmentalization, leading to efficient MMP-2 activation and proteolysis coupled with cellular function. (*Cancer Sci* 2010; 101: 843–847)

The major physiological role of matrix metalloproteinases (MMPs) is for homeostatic regulation of the extracellular environment, not simply to degrade matrix as their name suggests. MMPs are overexpressed in various human malignancies.^(1,2) The expression of MMPs is specifically regulated at the level of transcription. However, the catalytic activity of these proteinases is further controlled at additional levels, including activation of the pro-enzyme and inhibition of the active enzyme by specific inhibitors.^(3–5) Because MMPs including MMP-2 (gelatinase A/72-kDa type IV collagenase) are secreted as an inactive zymogen, activation is another key regulatory step for MMP function *in vivo*. The molecular environments in tumors appear conducive to MMP activation. Activated MMP-2 was specifically observed in a variety of tumor tissues, suggesting the presence of pro-MMP-2 activator(s) in tumor tissues.^(6–8) This led to the identification of the first membrane type-MMP (MT1-MMP) as an activator of pro-MMP-2 expressed on the surface of tumor cells.⁽⁸⁾ In addition to pro-MMP-2 cleavage for activation, MT1-MMP was shown to cleave variety of substrates, not only extracellular matrix (ECM) macromolecules, but also various biologically important molecules (reviewed in references^{9,10}).

Interaction of MT1-MMP with TIMP

To date, six MT-MMPs have been identified by cDNA cloning, and the proteolytic activation of pro-MMP-2 was shown experimentally for all MT-MMPs.^(11–14) MT-MMPs are unique in that they contain a furin recognition motif between the pro- and catalytic domains, and are processed by proprotein convertases such

as furin in the secretory pathway.⁽¹⁵⁾ In contrast to secretory form of MMPs, MT-MMPs are presented to the cell surface in active form. MT1-MMP is inhibited by endogenous inhibitors TIMP-2, -3, and -4, but not by TIMP-1.^(16,17) This TIMP-1 insensitive nature is common to all MT-MMPs except for MT4-MMP.⁽¹⁸⁾ MT1-MMP causes a rapid auto-degradation in the absence of its inhibitor to produce a 44-kDa form.⁽¹⁹⁾ Due to rapid turnover rate of active MT1-MMP, its concentration on cell surface is very low, and the active form of MT1-MMP predominantly localizes on the cell surface by forming a complex with the inhibitor (Fig. 1). TIMP-2 is the most common inhibitor of MT1-MMP, and MT1-MMP/TIMP-2 complex serves as a receptor for pro-MMP-2. Administration of the synthetic reversible inhibitor of MMP induces accumulation of MT1-MMP on the cell surface, which exerts enzyme activity at a burst by the removal of the inhibitor.⁽²⁰⁾

Implications from MMP-deficient mice

MT1-MMP is the most common physiological activator of pro-MMP-2, and the MT1-MMP-deficient mouse produces only a faint level of active MMP-2.^(21,22) MT1-MMP-null mice have severe defects in skeletal development and angiogenesis and die within several weeks after birth, suggesting an essential role for MT1-MMP in the process of angiogenesis and bone growth.^(21,22) This phenotype is much more severe than that seen in the MMP-2-deficient mice, which have smaller stature, blunted tumor-related angiogenesis, and abnormal lung alveolarization.⁽²³⁾ A hereditary deficiency in MMP-2 has been described in Middle Eastern families who develop a vanishing bone disorder.^(24,25) Thus, MT1-MMP functionality significantly exceeds MMP-2 activation in development. However, MT1-MMP and MMP-2 double-knockout mice show more severe phenotype than MT1-MMP-deficient mice, and die immediately after birth with respiratory failure, abnormal blood vessels, and immature muscle fibers reminiscent of central core disease.⁽²⁶⁾ This indicates that residual MMP-2 activation in MT1-MMP-null mice contributes to prolonged survival by partly complicating MT1-MMP functions in development. MMP-2 degrades types IV and V collagens, gelatins, laminin, fibronectin, and elastin.^(27,28) From the analysis of MMP-2-deficient mice, MMP-2 does not seem to be indispensable in development, although it partially complicates MT1-MMP. However, MMP-2 plays critical roles in pathological conditions including tumor growth.⁽²³⁾ For instance, angiogenesis and tumor growth are substantially suppressed in MMP-2 deficient mice.⁽²³⁾ MMP-2-null

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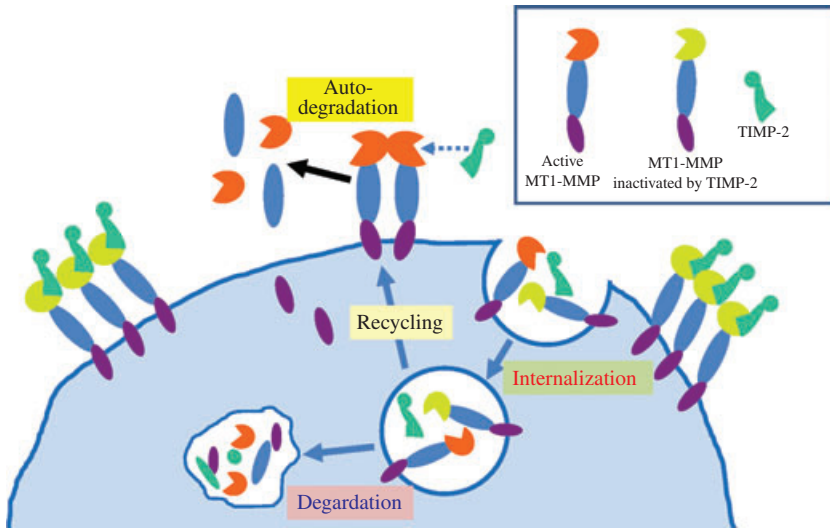


Fig. 1. Fate of membrane-type matrix metalloproteinase-1 (MT1-MMP) on the cell surface. MT1-MMP on cell surface rapidly turns over by auto-degradation or clathrin-dependent internalization. The internalized MT1-MMP is recycled or degraded in lysosome. MT1-MMP inactivated by tissue inhibitor of MMP (TIMP)-2 avoids auto-degradation, and accumulates on the cell surface. Accumulated MT1-MMP/TIMP-2 complex serves as a receptor for pro-MMP-2. It should be noted that concentration of active MT1-MMP on cell surface is quite low due to rapid auto-degradation, and only TIMP-2-inactivated MT1-MMP accumulates.

mice (also MMP-9-null mice) have extremely low incidences of cardiac rupture after acute myocardial infarction.^(29,30)

Mechanism of MMP-2 activation by MT1-MMP

Accumulated lines of evidence have demonstrated that MMP-2 is always activated in tumor tissues, in which MT1-MMP is expressed.^(8,31,32) Furthermore, MT1-MMP expression and MMP-2 activation levels are well co-related and closely associated with invasiveness and malignancy of tumors, suggesting that MT1-MMP is as critical a factor for tumor invasion and metastasis as MMP-2 activator.^(31,32) However, due to the complex MMP-2 activation process, the enzyme activity of MMP-2 and its possible contribution to tumor malignancy remain unclear.

The MT1-MMP mechanism of pro-MMP-2 activation has been well studied and partially defined.⁽³³⁻³⁵⁾ The suggested model implicates TIMP-2 as a bridging molecule, tethering pro-MMP-2 through binding between the COOH-terminal ends of pro-MMP-2 and TIMP-2 and binding between the NH₂-terminal ends of MT1-MMP and TIMP-2 (Fig. 2). The propeptide

of pro-MMP-2 is cleaved by an adjacent TIMP-2-free MT1-MMP between Asn37 and Leu38, generating an activated intermediate form that is further processed to the fully activated form by an intermolecular auto-cleavage when an intermediate form is present at a sufficiently high concentration at the cell surface. Pro-MMP-2 activation was expected to occur only at low TIMP-2 concentrations relative to MT1-MMP, which would permit the availability of active MT1-MMP to process the proMMP-2 bound in the ternary complex.^(19,34-39) Actually MT1-MMP auto-degradation is suppressed in the presence of TIMP-2, and MT1-MMP/TIMP-2 complex accumulates on cell surface.⁽⁴⁰⁾ The accumulated MT1-MMP/TIMP-2 complex serves as a receptor for pro-MMP-2, and the receptor-bound pro-MMP-2 is cleaved for activation by TIMP-2-free active MT1-MMP. In order to induce efficient pro-MMP-2 processing, the molar ratio of MT1-MMP/TIMP-2 complex to TIMP-2-free MT1-MMP is quite high (Fig. 3). Thus, MT1-MMP cannot cleave other direct substrates at the TIMP-2 level that induces efficient pro-MMP-2 processing.^(40,41) Furthermore, MMP-2 activated by MT1-MMP at TIMP-2 concentration to induce most efficient pro-MMP-2 processing is inactive. The TIMP-2 concentration to produce

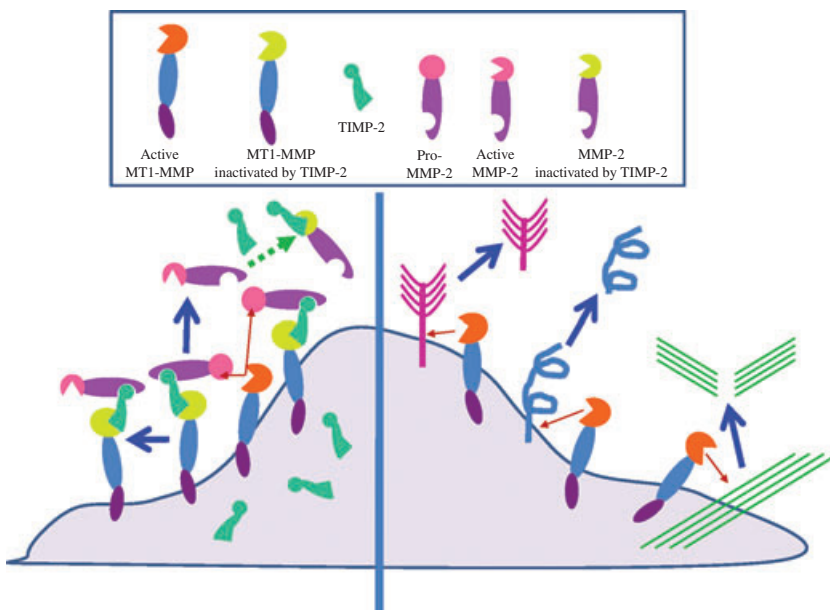


Fig. 2. Tissue inhibitor of MMP (TIMP)-2 concentration dictates substrate choice of membrane-type matrix metalloproteinase-1 (MT1-MMP). Left, MT1-MMP forms a complex with TIMP-2, and the complex serves as a receptor for pro-MMP-2. Pro-MMP-2, which binds to MT1-MMP/TIMP-2 complex is then processed by the neighboring TIMP-2-free MT1-MMP. The processed MMP-2 is inactivated by TIMP-2 depending on its concentration. Right, under TIMP-2-free condition, MT1-MMP directly cleaves its own substrates, such as extracellular matrix components, cell-surface molecules and so on.

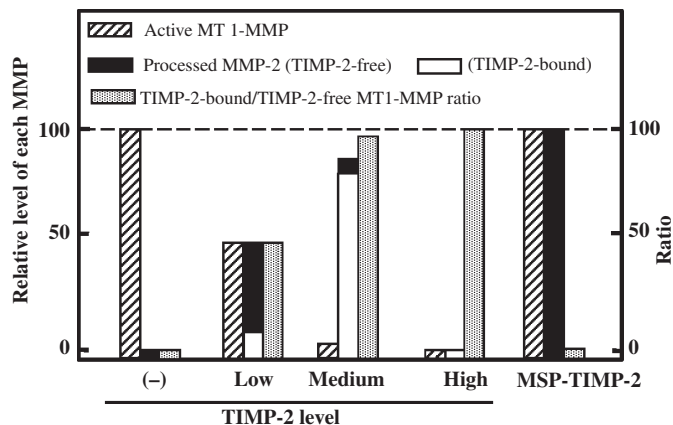


Fig. 3. Schematic illustration of tissue inhibitor of MMP (TIMP)-2-regulated membrane-type matrix metalloproteinase-1 (MT1-MMP) and MMP-2 activities. The level of active MT1-MMP and the ratio between MT1-MMP of TIMP-2 complex and TIMP-2-free (active) in the cells expressing MT1-MMP and different levels of TIMP-2 are schematically illustrated. The level of MMP-2 processed by these cells is also shown, which includes TIMP-2-free and TIMP-2-bound forms. In the absence of TIMP-2, MT1-MMP activity is the highest, and decreases proportionally to the TIMP-2 level. MMP-2 activated under a low TIMP-2 level is free from TIMP-2, and shows the highest activity. MMP-2 is processed most effectively at medium level of TIMP-2, in which most of MT1-MMP serves as a receptor for pro-MMP-2 by binding with MT1-MMP and processed MMP-2 is inactivated by TIMP-2. MMP-2 processing is blocked when MT1-MMP is all bound by a high level of TIMP-2. MSP-TIMP-2 does not affect MT1-MMP activity, and generates a maximum level of processed MMP-2 which is free from TIMP-2.

TIMP-2-free active MMP-2 is restricted to a narrow range at a low TIMP-2 concentration.^(41,42)

The above observation and interpretation were confirmed by using an artificial receptor for pro-MMP-2 (MSP-TIMP-2), which exposes the TIMP-2 COOH-terminal domain on the cell surface (Fig. 4).⁽⁴¹⁾ Since MSP-TIMP-2 does not have an MMP-inhibition function, the MT1-MMP level required to generate active MMP-2 with MSP-TIMP-2 is much lower than that with TIMP-2. This indicates that in the process of TIMP-2-mediated pro-MMP-2 activation by MT1-MMP, the majority of MT1-MMP functions as a receptor for pro-MMP-2 by making a complex with TIMP-2. Only a minor part of MT1-MMP is free from TIMP-2 and cleaves pro-MMP-2 in a tri-molecular complex. Thus, MT1-MMP is unable to cleave direct substrates effectively at TIMP-2 concentration to induce the most efficient pro-MMP-2 processing. COS-1 cells synthesize relatively higher concentration of TIMP-2 than 293T cells,⁽¹⁴⁾ and MT1-MMP exogenously expressed in COS-1 cells processes pro-MMP-2 more efficiently than those in 293T cells, but cleaves direct sub-

strates less effectively. Furthermore, MMP-2 activated by MT1-MMP in COS-1 cells shows a poor enzyme activity (H.Sato, Y. Okada and M. Seiki, unpublished data, 1994).

MT1-MMP turns over very rapidly due to autolysis if not inactivated/stabilized by TIMP-2. The time course of MT1-MMP production and its binding to TIMP-2 on the cell surface is a critical factor; however, this still remains unsolved. The cell-surface concentration of MT1-MMP stabilized by TIMP-2 is regulated by clathrin-dependent internalization (Fig. 1). Furthermore, cell-surface MT1-MMP is regulated temporally and spatially on the cell surface through the interaction with various binding molecules including tetraspanins, which will be discussed below. It should be noticed that such a dynamic cell-surface event may not be interpreted simply in a static stoichiometry, and further analysis is required.

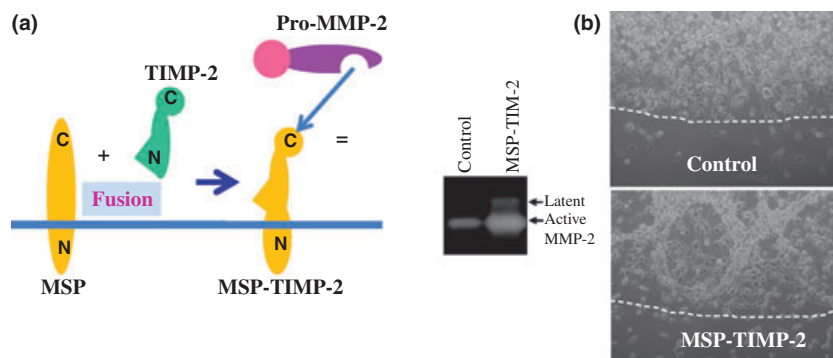
TIMP-2-independent MMP-2 activation

Miyamori *et al.* reported that tight junction protein claudin mediates the TIMP-2-independent MMP-2 activation pathway by not only MT1-MMP but by other MT-MMPs.⁽¹⁴⁾ Claudin recruits MT-MMP and MMP-2 at a tight junction to achieve elevated focal concentrations, and consequently enhances activation of pro-MMP-2. Among MT-MMP family members, only MT1-MMP can accumulate on the cell surface through the complex formation with TIMP-2, and serves as a receptor for pro-MMP-2 (T. Takino and H. Sato, unpublished data, 2001). An artificial receptor for pro-MMP-2 (MSP-TIMP-2) substitutes for MT-MMP/TIMP-2 complex, and induces activation by all MT-MMPs except for MT4-MMP.⁽⁴¹⁾ MT2-MMP was shown to induce pro-MMP-2 activation in cells derived from TIMP-2-deficient mice.^(43,44) Pro-MMP-2 associates with TIMP-2-deficient cells through the hemopexin domain, and is processed by MT2-MMP.⁽⁴³⁾ Pro-MMP-2 receptor in TIMP-2-deficient cells has not been identified, which cooperates with only MT2-MMP but not MT1-MMP for activation. In any case, all MT-MMPs can functionally cleave pro-MMP-2 for activation, but only MT1-MMP/TIMP-2 complex serves as an efficient receptor for pro-MMP-2 activation (T. Takino and H. Sato, unpublished data, 2001).

Coordinate action of MMP-2 and MT1-MMP

The HT1080 fibrosarcoma cell line expresses MT1-MMP, and produces active MMP-2. MT1-MMP plays a pivotal role in invasive growth in collagen gel, but the role of MMP-2 still remains unclear. Expression of an artificial receptor for pro-MMP-2 MSP-TIMP-2 in HT1080 cells enhances activation of MMP-2 on the cell surface, and results in drastic degradation of collagen gel, which never takes place by overexpression of MT1-MMP or MMP-2 alone (Fig. 4b).⁽⁴¹⁾ A possible explanation for intensive digestion of collagen gel is that MT1-MMP as

Fig. 4. Artificial receptor for pro-matrix metalloproteinase-2 (MMP-2) (MSP-TIMP-2). (a) MSP-TIMP-2 consists of the transmembrane domain of type II transmembrane protein mosaic serine protease (MSP) and tissue inhibitor of MMP (TIMP)-2. Pro-MMP-2 binds to the C-terminus of MSP-TIMP-2. (b) Left, MSP-TIMP-2-expressing HT1080 cells generate more active MMP-2 than mock cells. Right, MSP-TIMP-2-expressing cells were cultured in collagen gel, which caused intensive degradation of collagen gel. Dotted line, border of collagen gel. Magnification, $\times 100$.



a collagenase cleaves type I collagen to initiate denaturation into gelatin, which is subsequently digested to pieces by MMP-2 gelatinase. Unlike TIMP-2, MSP-TIMP-2 induces gelatinase (MMP-2) activity without loss of collagenase (MT1-MMP) activity, and its coordinate action may result in the excessive melting of collagen gel. TIMP-2 regulates the balance between MT1-MMP and MMP-2 activity depending on its concentration. HT1080 cells produce a low level TIMP-2, and express both active MT1-MMP and MMP-2 as shown in Figure 4, which may coordinately contribute to the invasive character of HT1080 cells. In fact, down-regulation of either MT1-MMP or MMP-2 expression suppresses invasive growth of HT1080 cells in collagen gel (T. Takino and H. Sato, unpublished data, 2004).

MMP-2 activated through MT1-MMP/TIMP-2 complex as a receptor remains on the receptor. MT1-MMP stimulates integrin-mediated extracellular signal-regulated kinase (ERK) activation by enhancing the turnover rate of focal adhesions on fibronectin or collagen.^(45,46) MMP-2 activated by MT1-MMP co-operates with MT1-MMP for ERK activation.⁽⁴¹⁾ Recently, Kenny *et al.* showed, using small-interfering RNA and neutralizing antibody targeting MMP-2, that MMP-2 plays an essential role in tumor growth and metastasis in nude mice model in ovarian cancer cells.⁽⁴⁷⁾ MMP-2 enhances peritoneal adhesion of ovarian cancer cells through cleavage of fibronectin and vitronectin into small fragments, and increases the binding of cells to these fragments and their receptors, $\alpha 5\beta 1$ and $\alpha V\beta 3$ integrin.

Tetraspanins as regulators of MT1-MMP. Tetraspanins are attracting attention as binding proteins of MT1-MMP, which regulate subcellular localization and compartmentalization of MT1-MMP and consequent MT1-MMP activities.^(48,49) Tetraspanins of integral membrane proteins with four predicted transmembrane domains comprise a large group of ubiquitously expressed proteins (33 human members). Direct association of MT1-MMP with a tetraspanin family member CD63 was first reported by Takino *et al.*⁽⁵⁰⁾ CD63 was shown to mediate internalization of MT1-MMP from the cell surface to lysosomes, and MT1-MMP undertakes lysosomal proteolysis. Among tetraspanin family members, CD151 in endothelial cells and CD9, CD81, and TSPAN12 in cancer cells were reported to be associ-

ated with regulation of MT1-MMP functions.^(48,49) They support MT1-MMP cell-surface expression and functions including ECM degradation and MMP-2 activation. The CD151-mediated functional association of MT1-MMP with integrin $\alpha 3\beta 1$ regulates MT1-MMP membrane compartmentalization, subcellular localization, and cell-surface concentration as a result in endothelial cells.⁽⁴⁸⁾ Since MT1-MMP plays an essential role in angiogenesis, CD151 may contribute to endothelial homeostasis through the regulation of MT1-MMP. CD9, CD81, and TSPAN12 also associate with cell surface molecules including integrins,⁽⁵¹⁾ which may form specialized membrane structures for invasion (e.g. lamellipodia, filopodia, and invadopodia) with MT1-MMP.⁽⁵²⁾ Tetraspanin-regulated cell surface localization of MT1-MMP increases the local concentration for focal proteolysis within the pericellular environment and leads to efficient ECM degradation and MMP-2 activation. Further study will be necessary to elucidate physiological and pathological roles of tetraspanin-MT1-MMP interaction and functional variations among tetraspanin family members in terms of MT1-MMP regulation.

Conclusion

TIMP-2 dictates substrate choice of MT1-MMP, either direct cleavage of substrate or pro-MMP-2 activation. In the absence of TIMP-2, MT1-MMP mediates only direct cleavage of substrates, but not pro-MMP-2 activation. At an appropriately low concentration of TIMP-2, both MT1-MMP and MMP-2 activated by MT1-MMP can exert enzyme activities. Coordinate action of them induces more efficient tissue degradation and consequent invasive growth compared with that by MT1-MMP or MMP-2 alone. MMP-2 was shown to play a pivotal role in invasive growth of ovarian cancer,⁽⁴⁷⁾ in which MT1-MMP should be involved in not only MMP-2 activation but also ECM degradation, and cooperate with MMP-2. Broad-spectrum synthetic inhibitors targeting MMP active sites suffered from harmful side effects in clinical testing. Targeting MMP-2 or MMP-2 activation step may not be excessively toxic, and is promising for the suppression of tumor malignancy.

References

- Birkedal HH, Moore WG, Bodden MK *et al.* Matrix metalloproteinases: a review. *Crit Rev Oral Biol Med* 1993; **4**: 197–250.
- Stetler-Stevenson WG, Aznavoorian S, Liotta LA. Tumor cell interactions with the extracellular matrix during invasion and metastasis. *Annu Rev Cell Biol* 1993; **9**: 541–73.
- Matrisian LM. Metalloproteinases and their inhibitors in matrix remodeling. *Trends Genet* 1990; **6**: 121–5.
- Curry TJ, Mann JS, Estes RS, Jones PB. Alpha 2-macroglobulin and tissue inhibitor of metalloproteinases: collagenase inhibitors in human preovulatory ovaries. *Endocrinology*, 1990; **127**: 63–8.
- Zhu C, Woessner JJ. A tissue inhibitor of metalloproteinases and α -macroglobulins in the ovulating rat ovary: possible regulators of collagen matrix breakdown. *Biol Reprod* 1991; **45**: 334–42.
- Brown PD, Bloxidge RE, Stuart NSA, Gatter KC, Carmichael J. Association between expression of activated 72-Kilodalton gelatinase and tumor spread in non-small-cell lung carcinoma. *J Natl Cancer Inst* 1993; **85**: 574–8.
- Azzam HS, Arand G, Lippman ME, Thompson EW. Association of MMP-2 activation potential with metastatic progression in human breast cancer cell lines independent of MMP-2 activation. *J Natl Cancer Inst* 1993; **85**: 1758–64.
- Sato H, Takino T, Okada Y *et al.* A matrix metalloproteinase expressed on the surface of invasive tumour cells. *Nature* 1994; **370**: 61–5.
- Sato H, Takino T, Miyamori H. Roles of membrane-type matrix metalloproteinase-1 in tumor invasion and metastasis. *Cancer Sci* 2005; **96**: 212–7.
- Itoh Y. MT1-MMP: A Key Regulator of Cell Migration in Tissue. *IUBMB Life* 2006; **58**: 589–96.
- Will H, Hinzmann B. cDNA sequence and mRNA tissue distribution of a novel human matrix metalloproteinase with a potential transmembrane segment. *Eur J Biochem* 1995; **231**: 602–8.
- Takino T, Sato H, Shinagawa A, Seiki M. Identification of the second membrane-type matrix metalloproteinase (MT-MMP-2) gene from a human placenta cDNA library. *J Biol Chem* 1995; **270**: 23013–20.
- Velasco G, Cal S, Merlos-Suarez A *et al.* Human MT6-matrix metalloproteinase: identification, progelatinase A activation, and expression in brain tumors. *Cancer Res* 2000; **60**: 877–82.
- Miyamori H, Takino T, Kobayashi Y *et al.* Claudin promotes activation of pro-matrix metalloproteinase-2 mediated by membrane-type matrix metalloproteinases. *J Biol Chem* 2001; **276**: 28204–11.
- Sato H, Kinoshita T, Takino T *et al.* Activation of a recombinant membrane type 1-matrix metalloproteinase (MT1-MMP) by furin and its interaction with tissue inhibitor of metalloproteinases (TIMP)-2. *FEBS Lett* 1996; **393**: 101–4.
- Will H, Atkinson SJ, Butler GS, Smith B, Murphy G. The soluble catalytic domain of membrane type 1 matrix metalloproteinase cleaves the propeptide of progelatinase A and initiates autolytic activation. Regulation by TIMP-2 and TIMP-3. *J Biol Chem* 1996; **271**: 17119–23.
- Bigg HF, Morrison CJ, Butler GS *et al.* Tissue inhibitor of metalloproteinases-4 inhibits but does not support the activation of gelatinase A via efficient inhibition of membrane type 1-matrix metalloproteinase. *Cancer Res* 2001; **61**: 3610–8.
- Kolkenbrock H, Essers L, Ulbrich N, Will H. Biochemical characterization of the catalytic domain of membrane-type 4 matrix metalloproteinase. *Biol Chem* 1999; **380**: 1103–8.
- Hernandez-Barrantes S, Toth M, Bernardo MM *et al.* Binding of active (57 kDa) membrane type 1-matrix metalloproteinase (MT1-MMP) to tissue inhibitor of metalloproteinase (TIMP)-2 regulates MT1-MMP processing and Pro-MMP-2 activation. *J Biol Chem* 2000; **275**: 12080–9.
- Toth M, Bernardo MM, Gervasi DC *et al.* Tissue inhibitor of metalloproteinase (TIMP)-2 acts synergistically with synthetic matrix metalloproteinase (MMP) inhibitors but not with TIMP-4 to enhance the

- (Membrane type 1)-MMP-dependent activation of pro-MMP-2. *J Biol Chem* 2000; **275**: 41415–23.
- 21 Holmbeck K, Bianco P, Caterina J *et al*. MT1-MMP-deficient mice develop dwarfism, osteopenia, arthritis, and connective tissue disease due to inadequate collagen turnover. *Cell* 1999; **99**: 81–92.
 - 22 Zhou Z, Apte SS, Soininen R *et al*. Impaired endochondral ossification and angiogenesis in mice deficient in membrane-type matrix metalloproteinase I. *Proc Natl Acad Sci U S A* 2000; **97**: 4052–7.
 - 23 Itoh T, Tanioka M, Yoshid H, Yoshioka T, Nishimoto H, Itohar S. Reduced angiogenesis and tumor progression in gelatinase A-deficient mice. *Cancer Res* 1998; **58**: 1048–51.
 - 24 Martignetti JA, Al Aqeel AA, Al Sewairi W *et al*. Mutation of the matrix metalloproteinase 2 gene (MMP2) causes a multicentric osteolysis and arthritis syndrome. *Nat Genet* 2001; **28**: 261–5.
 - 25 Vu TH. Don't mess with the matrix. *Nat Genet* 2001; **28**: 202–3.
 - 26 Oh J, Takahashi R, Adachi E *et al*. Mutations in two matrix metalloproteinase genes, MMP-2 and MT1-MMP, are synthetic lethal in mice. *Oncogene* 2004; **23**: 5041–8.
 - 27 Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. *Annu Rev Cell Dev Biol* 2001; **17**: 463–516.
 - 28 Okada Y. Proteinases and matrix degradation. In: Ruddy S, Harris ED Jr, Sledg CB, eds. *Kelley's Textbook of Rheumatology*. Philadelphia, PA, USA: W.B. Saunders Company, 2001; 55–72.
 - 29 Hayashidani S, Tsutsui H, Ikeuchi M *et al*. Targeted deletion of matrix metalloproteinase-2 attenuates early left ventricular rupture and late remodeling after experimental myocardial infarction. *Am J Physiol Heart Circ Physiol* 2003; **285**: H1229–35.
 - 30 Heymans S, Lutun A, Nuyens D *et al*. Inhibition of plasminogen activators or matrix metalloproteinases prevents cardiac rupture but impairs therapeutic angiogenesis and causes cardiac failure. *Nat Med* 1999; **5**: 1135–42.
 - 31 Ueno H, Nakamura H, Inoue M *et al*. Expression and tissue localization of membrane-type 1, 2, and 3 matrix metalloproteinases in human invasive breast carcinoma. *Cancer Res* 1997; **57**: 2055–60.
 - 32 Tokuraku M, Sato H, Murakami S, Okada Y, Watanabe Y, Seiki M. Activation of the precursor of gelatinase A/72 kDa type IV collagenase/MMP-2 in lung carcinomas correlates with the expression of membrane-type matrix metalloproteinase (MT-MMP) and with lymph node metastasis. *Int J Cancer* 1995; **64**: 355–9.
 - 33 Howard EW, Banda MJ. Binding of tissue inhibitor of metalloproteinases 2 to two distinct sites on human 72-kDa gelatinase: Identification of a stabilization site. *J Biol Chem* 1991; **266**: 17972–7.
 - 34 Strongin AY, Collier I, Bannikov G *et al*. Mechanism of cell surface activation of 72-kDa type IV collagenase. Isolation of the activated form of the membrane metalloprotease. *J Biol Chem* 1995; **270**: 5331–8.
 - 35 Kinoshita T, Sato H, Okada A *et al*. TIMP-2 promotes activation of progelatinase A by membrane-type 1 matrix metalloproteinase immobilized on agarose beads. *J Biol Chem* 1998; **273**: 16098–103.
 - 36 Butler GS, Butler MJ, Atkinson SJ *et al*. The TIMP2 membrane type 1 metalloproteinase 'receptor' regulates the concentration and efficient activation of progelatinase A. A kinetic study. *J Biol Chem* 1998; **273**: 871–80.
 - 37 Zucker S, Drews M, Conner C *et al*. Tissue inhibitor of metalloproteinase-2 (TIMP-2) binds to the catalytic domain of the cell surface receptor, membrane type 1-matrix metalloproteinase 1 (MT1-MMP). *J Biol Chem* 1998; **273**: 1216–22.
 - 38 Cao J, Sato H, Takino T, Seiki M. The C-terminal region of membrane type matrix metalloproteinase is a functional transmembrane domain required for pro-gelatinase A activation. *J Biol Chem* 1995; **270**: 801–5.
 - 39 Atkinson SJ, Crabbe T, Cowell S *et al*. Intermolecular autolytic cleavage can contribute to the activation of progelatinase A by cell membranes. *J Biol Chem* 1995; **270**: 30479–85.
 - 40 Kudo T, Takino T, Miyamori H *et al*. Substrate choice of membrane-type 1 matrix metalloproteinase is dictated by tissue inhibitor of metalloproteinase-2 levels. *Cancer Sci* 2007; **98**: 563–8.
 - 41 Nishida Y, Miyamori H, Thompson EW, Takino T, Endo Y, Sato H. Activation of Matrix Metalloproteinase-2 (MMP-2) by Membrane Type-1 Matrix Metalloproteinase through an Artificial Receptor for ProMMP-2 Generates Active MMP-2. *Cancer Res* 2008; **68**: 9096–104.
 - 42 Bernardo MM, Fridman R. TIMP-2 (tissue inhibitor of metalloproteinase-2) regulates MMP-2 (matrix metalloproteinase-2) activity in the extracellular environment after pro-MMP-2 activation by MT1 (membrane type 1)-MMP. *Biochem J* 2003; **374**: 739–45.
 - 43 Morrison CJ, Butler GS, Bigg HF *et al*. Cellular Activation of MMP-2 (Gelatinase A) by MT2-MMP Occurs via a TIMP-2-independent Pathway. *J Biol Chem* 2001; **276**: 47402–10.
 - 44 Morrison CJ, Overall CM. TIMP Independence of Matrix Metalloproteinase (MMP)-2 Activation by Membrane Type 2 (MT2)-MMP Is Determined by Contributions of Both the MT2-MMP Catalytic and Hemopexin C Domains. *J Biol Chem* 2006; **281**: 26528–39.
 - 45 Takino T, Miyamori H, Watanabe Y *et al*. Membrane type 1 matrix metalloproteinase regulates collagendependent mitogen-activated protein/extracellular signal-related kinase activation and cell migration. *Cancer Res* 2004; **64**: 1044–9.
 - 46 Takino T, Saeki H, Miyamori H *et al*. Inhibition of membrane-type 1 matrix metalloproteinase at cellmatrix adhesions. *Cancer Res* 2007; **67**: 11621–9.
 - 47 Kenny HA, Kaur S, Coussens LM, Lengyel E. The initial steps of ovarian cancer cell metastasis are mediated by MMP-2 cleavage of vitronectin and fibronectin. *J Clin Invest* 2008; **118**: 1367–79.
 - 48 Yañez-Mó M, Barreiro O, Gonzalo P *et al*. MT1-MMP collagenolytic activity is regulated through association with tetraspanin CD151 in primary endothelial cells. *Blood* 2008; **112**: 3217–26.
 - 49 Lafleur MA, Xu D, Hemler ME. Tetraspanin Proteins Regulate Membrane Type-1 Matrix Metalloproteinase-dependent Pericellular Proteolysis. *Mol Biol Cell* 2009; **20**: 2030–40.
 - 50 Takino T, Miyamori H, Kawaguchi N *et al*. Tetraspanin CD63 promotes targeting and lysosomal proteolysis of membrane-type 1 matrix metalloproteinase. *Biochem Biophys Res Com* 2003; **304**: 160–6.
 - 51 Berditchevski F. Complexes of tetraspanins with integrins: more than meets the eye. *J Cell Sci* 2001; **114**: 4143–51.
 - 52 Hemler ME. Tetraspanin functions and associated microdomains. *Nat Rev Mol Cell Biol* 2005; **6**: 801–11.