

Glycogen synthase kinase-3 is a pivotal mediator of cancer invasion and resistance to therapy

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journal or publication title	Cancer Science
volume	107
number	10
page range	1363-1372
year	2016-10-01
URL	http://hdl.handle.net/2297/46504

doi: 10.1111/cas.13028

Supplementary References (SR)

- SR1. Hiscox S, Morgan L, Barrow D, Dutkowski C, Wakeling A, Nicholson RI. Tamoxifen resistance in breast cancer cells is accompanied by an enhanced motile and invasive phenotype: inhibition by gefitinib ('Iressa', ZD1839). *Clin Exp Metastasis* 2004;**21**:201-12.
- SR2. Hiscox S, Jiang WG, Obermeier K, et al. Tamoxifen resistance in MCF7 cells promotes EMT-like behaviour and involves modulation of β -catenin phosphorylation. *Int J Cancer* 2006;**118**:290-301.
- SR3. Li QQ, Xu JD, Wang WJ, et al. Twist1-mediated adriamycin-induced epithelial-mesenchymal transition relates to multidrug resistance and invasive potential in breast cancer cells. *Clin Cancer Res* 2009;**15**:2657-65.
- SR4. Acharyya S, Oskarsson T, Vanharanta S, et al. A CXCL1 paracrine network links cancer chemoresistance and metastasis. *Cell* 2012;**150**:165-78.
- SR5. Yang AD, Fan F, Camp ER, et al. Chronic oxaliplatin resistance induces epithelial-to-mesenchymal transition in colorectal cancer cell lines. *Clin Cancer Res* 2006;**12**:4147-53.
- SR6. Shah AN, Summy JM, Zhang J, Park SI, Parikh NU, Gallick GE. Development and characterization of gemcitabine-resistant pancreatic tumor cells. *Ann Surg Oncol* 2007;**14**:3629-37.
- SR7. Wang Z, Li Y, Kong D, et al. Acquisition of epithelial-mesenchymal transition phenotype of gemcitabine-resistant pancreatic cancer cells is linked with activation of the notch signaling pathway. *Cancer Res* 2009;**69**:2400-7.
- SR8. Arumugam T, Ramachandran V, Fournier KF, et al. Epithelial to mesenchymal transition contributes to drug resistance in pancreatic cancer. *Cancer Res* 2009;**69**:5820-8.
- SR9. Zhang Y, Wei J, Wang H, et al. Epithelial mesenchymal transition correlates with CD24⁺CD44⁺ and CD133⁺ cells in pancreatic cancer. *Oncol Rep* 2012;**27**:1599-605.
- SR10. Wang X, Ling MT, Guan XY, et al. Identification of a novel function of TWIST, a bHLH protein, in the development of acquired taxol resistance in human cancer cells. *Oncogene* 2004;**23**:474-82.
- SR11. Kajiyama H, Shibata K, Terauchi M, et al. Chemoresistance to paclitaxel induces epithelial mesenchymal transition and enhances metastatic potential for epithelial ovarian carcinoma cells. *Int J Oncol* 2007;**31**:277-83.
- SR12. Rosanò L, Cianfrocca R, Spinella F, et al. Acquisition of chemoresistance and EMT phenotype is linked with activation of the endothelin A receptor pathway in ovarian carcinoma cells. *Clin Cancer Res* 2011;**17**:2350-60.
- SR13. Marín-Aguilera M, Codony-Servat J, Reig Ò, et al. Epithelial-to-mesenchymal transition mediates docetaxel resistance and high risk of relapse in prostate cancer. *Mol Cancer Ther* 2014;**13**:1270-84.
- SR14. Sun L, Yao Y, Liu B, et al. MiR-200b and miR-15b regulate chemotherapy-induced epithelial-mesenchymal transition in human tongue cancer cells by targeting BMI1. *Oncogene* 2012;**31**:432-45.

- SR15. Munson JM, Fried L, Rowson SA, et al. Anti-invasive adjuvant therapy with imipramine blue enhances chemotherapeutic efficacy against glioma. *Sci Transl Med* 2012;**4**:127ra36.
- SR16. Qian LW, Mizumoto K, Urashima T, et al. Radiation-induced increase in invasive potential of human pancreatic cancer cells and its blockade by a matrix metalloproteinase inhibitor, CGS27023. *Clin Cancer Res* 2002;**8**:1223-7.
- SR17. Tsukamoto H, Shibata K, Kajiyama H, Terauchi M, Nawa A, Kikkawa F. Irradiation-induced epithelial-mesenchymal transition (EMT) related to invasive potential in endometrial carcinoma cells. *Gynecol Oncol* 2007;**107**:500-4.
- SR18. Wild-Bode C, Weller M, Rimmer A, Dichgans J, Wick W. Sublethal irradiation promotes migration and invasiveness of glioma cells: implications for radiotherapy of human glioblastoma. *Cancer Res* 2001;**61**:2744-50.
- SR19. Goetze K, Scholz M, Taucher-Scholz G, Mueller-Klieser W. The impact of conventional and heavy ion irradiation on tumor cell migration *in vitro*. *Int J Radiation Biol* 2007;**83**:889-96.
- SR20. Ogata T, Teshima T, Kagawa K, et al. Particle irradiation suppresses metastatic potential of cancer cells. *Cancer Res* 2005;**65**:113-20.
- SR21. Buck E, Eyzaguirre A, Barr S, et al. Loss of homotypic cell adhesion by epithelial-mesenchymal transition or mutation limits sensitivity to epidermal growth factor receptor inhibition. *Mol Cancer Ther* 2007;**6**:532-41.
- SR22. Lucio-Eterovic AK, Piao Y, de Groot JF. Mediators of glioblastoma resistance and invasion during antivascular endothelial growth factor therapy. *Clin Cancer Res* 2009;**15**:4589-99.
- SR23. Keunen O, Johansson M, Oudin A, et al. Anti-VEGF treatment reduces blood supply and increases tumor cell invasion in glioblastoma. *Proc Natl Acad Sci U S A* 2011; **108**:3749-54.
- SR24. Piao Y, Liang J, Holmes L, Henry V, Sulman E, de Groot JF. Acquired resistance to anti-VEGF therapy in glioblastoma is associated with a mesenchymal transition. *Clin Cancer Res* 2013;**19**:4392-403.
- SR25. Meijer L, Flajolet M, Greengard P. Pharmacological inhibitors of glycogen synthase kinase 3. *Trends Pharmacol Sci* 2004;**25**:471-80.
- SR26. Phukan S, Babu VS, Kannoji A, Hariharan R, Balaji VN. GSK3 β : role in therapeutic landscape and development of modulators. *Br J Pharmacol* 2010;**160**:1-19.
- SR27. Eldar-Finkelman H, Martinez A. GSK-3 inhibitors: preclinical and clinical focus on CNS. *Front Mol Neurosci* 2011;**4**:32.
- SR28. Osolodkin DI, Paltulin VA, Zefirov NS. Glycogen synthase kinase 3 as an anticancer drug target: novel experimental findings and trends in the design of inhibitors. *Curr Pharmacol Design* 2013;**19**:665-79.
- SR29. Bhat R, Xue Y, Berg S, et al. Structural insights and biological effects of glycogen synthase kinase 3-specific inhibitor AR-A014418. *J Biol Chem* 2003;**278**:45937-45.
- SR30. Li J, Mizukami Y, Zhang X, Jo WS, Chung DC. Oncogenic K-ras stimulates Wnt signaling in colon cancer through inhibition of GSK-3 β . *Gastroenterology* 2005;**128**:1907-18.

- SR31. Thiel A, Heinonen M, Rintahaka J, et al. Expression of cyclooxygenase-2 is regulated by glycogen synthase kinase-3 β in gastric cancer cells. *J Biol Chem* 2006;**281**:4564-9.
- SR32. Watson RL, Spalding AC, Zielske SP, et al. GSK3 β and β -catenin modulate radiation cytotoxicity in pancreatic cancer. *Neoplasia* 2010;**12**:357-65.
- SR33. Ben-Josef E, George A, Regine WF, et al. Glycogen synthase kinase 3 beta predicts survival in resected adenocarcinoma of the pancreas. *Clin Cancer Res* 2015;**21**:5612-8.
- SR34. Beurel E, Kornprobst M, Blivet-Van Eggelpoël MJ, et al. GSK-3 β inhibition by lithium confers resistance to chemotherapy-induced apoptosis through the repression of CD95 (Fas/APO-1) expression. *Exp Cell Res* 2004;**300**:354-64.
- SR35. Beurel E, Kornprobst M, Blivet-Van Eggelpoël MJ, Cadoret A, Capeau J, Desbois-Mouthon C. GSK-3 β reactivation with LY294002 sensitizes hepatoma cells to chemotherapy-induced apoptosis. *Int J Oncol* 2005;**27**:215-22.
- SR36. Huang KT, Huang YH, Li P, et al. Correlation between tuberous sclerosis complex 2 and glycogen synthase kinase 3 beta levels, and outcomes of patients with hepatocellular carcinoma treated by hepatectomy. *Hepatol Res* 2014;**44**:1142-50.
- SR37. Qiao G, Le Y, Li J, Wang L, Shen F. Glycogen synthase kinase-3 β is associated with the prognosis of hepatocellular carcinoma and may mediate the influence of type 2 diabetes mellitus on hepatocellular carcinoma. *PLoS One* 2014;**9**:e105624.
- SR38. Song CL, Tang H, Ran LK, et al. Sirtuin 3 inhibits hepatocellular carcinoma growth through the glycogen synthase kinase-3 β /BCL2-associated X protein-dependent apoptotic pathway. *Oncogene* 2016;**35**:631-41.
- SR39. Wang L, Lin HK, Hu YC, Xie S, Yang L, Chang C. Suppression of androgen receptor-mediated transactivation and cell growth by the glycogen synthase kinase 3 β in prostate cells. *J Biol Chem* 2004;**279**:32444-52.
- SR40. Jiang Y, Dai J, Zhang H, et al. Activation of the Wnt pathway through AR79, a GSK3 β inhibitor, promotes prostate cancer growth in soft tissue and bone. *Mol Cancer Res* 2013;**11**:1597-610.
- SR41. Cai G, Wang J, Xin X, Ke Z, Luo J. Phosphorylation of glycogen synthase kinase-3 β at serine 9 confers cisplatin resistance in ovarian cancer cells. *In J Oncol* 2007;**31**:657-62.
- SR42. Zhai Y, Iura A, Yeasmin S, et al. MSX2 is an oncogenic downstream target of activated WNT signaling in ovarian endometrioid adenocarcinoma. *Oncogene* 2011;**30**:4152-62.
- SR43. Pastorino JG, Hoek JB, Shulga N. Activation of glycogen synthase kinase 3B disrupts the binding of hexokinase II to mitochondria by phosphorylating voltage-dependent anion channel and potentiates chemotherapy-induced cytotoxicity. *Cancer Res* 2005;**65**:10545-54.
- SR44. Dong J, Peng J, Zhang H, et al. Role of glycogen synthase kinase 3 β in rapamycin-mediated cell cycle regulation and chemosensitivity. *Cancer Res* 2005;**65**:1961-72.
- SR45. Farago M, Dominguez I, Landesman-Bollag E, et al. Kinase-inactive glycogen synthase kinase 3 β promotes Wnt signaling and mammary tumorigenesis. *Cancer Res* 2005;**65**:5792-801.

- SR46. Wang Y, Lam JB, Lam KS, et al. Adiponectin modulates the glycogen synthase kinase-3 β / β -catenin signaling pathway and attenuates mammary tumorigenesis of MDA-MB-231 cells in nude mice. *Cancer Res* 2006;**66**:11462-70.
- SR47. Soto-Cerrato V, Viñals F, Lambert JR, Kelly JA, Pérez-Tomás R. Prodigiosin induces the proapoptotic gene NAG-1 via glycogen synthase kinase-3 β activity in human breast cancer cells. *Mol Cancer Ther* 2007;**6**:362-9.
- SR48. Ding Q, He X, Hsu JM, et al. Degradation of Mcl-1 by β -TrCP mediates glycogen synthase kinase 3-induced tumor suppression and chemosensitization. *Mol Cell Biol* 2007;**27**:4006-17.
- SR49. Ding Q, He X, Xia W, et al. Myeloid cell leukemia-1 inversely correlates with glycogen synthase kinase-3 β activity and associates with poor prognosis in human breast cancer. *Cancer Res* 2007;**67**:4564-71.
- SR50. Mora-Santos M, Limon-Mortes MC, Limón-Mortés MC, et al. Glycogen synthase kinase-3 β (GSK3 β) negatively regulates PTTG1/human securin protein stability, and GSK3 β inactivation correlates with securin accumulation in breast tumors. *J Biol Chem* 2011;**286**:30047-56.
- SR51. Dembowy J, Adissu HA, Liu JC, Zacksenhaus E, Woodgett JR. Effect of glycogen synthase kinase-3 inactivation on mouse mammary gland development and oncogenesis. *Oncogene* 2015;**34**:3514-26.
- SR52. Li J, Xing M, Zhu M, et al. Glycogen synthase kinase 3 β induces apoptosis in cancer cells through increase of survivin nuclear localization. *Cancer Lett* 2008;**272**:91-101.
- SR53. Kao SH, Wang WL, Chen CY, et al. GSK3 β controls epithelial-mesenchymal transition and tumor metastasis by CHIP-mediated degradation of Slug. *Oncogene* 2014;**33**:3172-82.
- SR54. Koo J, Yue P, Gal AA, Khuri FR, Sun SY. Maintaining glycogen synthase kinase-3 activity is critical for mTOR kinase inhibitors to inhibit cancer cell growth. *Cancer Res* 2014;**74**:2555-68.
- SR55. Leis H, Segrelles C, Ruiz S, Santos M, Paramio JM. Expression, localization, and activity of glycogen synthase kinase 3 β during mouse skin tumorigenesis. *Mol Carcinog* 2002;**35**:180-5.
- SR56. Ma C, Wang J, Gao Y, et al. The role of glycogen synthase kinase 3 β in the transformation of epidermal cells. *Cancer Res* 2007;**67**:7756-64.
- SR57. Liu Q, Mier JW, Panka DJ. Differential modulatory effects of GSK-3 β and HDM2 on sorafenib-induced AIF nuclear translocation (programmed necrosis) in melanoma. *Mol Cancer* 2011;**10**:115.
- SR58. Li Z, Tan F, Thiele CJ. Inactivation of glycogen synthase kinase-3 β contributes to brain-derived neurotrophic factor/TrkB-induced resistance to chemotherapy in neuroblastoma cells. *Mol Cancer Ther* 2007;**6**:3113-21.
- SR59. Shine B, McKnight R, Leaver L, Geddes JR. Long-term effects of lithium on renal, thyroid, and parathyroid function: a retrospective analysis of laboratory data. *Lancet* 2015;**386**:461-8.
- SR60. Tolosa E, Litvan I, Höglinger GU, et al. A phase 2 trial of the GSK-3 inhibitor tideglusib in progressive supranuclear palsy. *Mov Disord* 2014;**29**:470-8.

- SR61. Höglinger GU, Huppertz HJ, Wagenpfeil S, et al. Tideglusib reduces progression of brain atrophy in progressive supranuclear palsy in a randomized trial. *Mov Disord* 2014;**29**:479-87.
- SR62. del Ser T, Steinwachs KC, Gertz HJ, et al. Treatment of Alzheimer's disease with the GSK-3 inhibitor tideglusib: a pilot study. *J Alzheimers Dis* 2013;**33**:205-15.
- SR63. Lovestone S, Boada M, Dubois B, et al. A phase II trial of tideglusib in Alzheimer's disease. *J Alzheimers Dis* 2015;**45**:75-88.
- SR64. Zamek-Gliszczynski MJ, Abraham TL, Alberts JJ, et al. Pharmacokinetics, metabolism, and excretion of the glycogen synthase kinase-3 inhibitor LY2090314 in rats, dogs, and humans: a case study in rapid clearance by extensive metabolism with low circulating metabolite exposure. *Drug Metab Dispos* 2013;**41**:714-26.
- SR65. Gray JE, Infante JR, Brail LH, et al. A first-in-human phase I dose-escalation, pharmacokinetic, and pharmacodynamic evaluation of intravenous LY2090314, a glycogen synthase kinase 3 inhibitor, administered in combination with pemetrexed and carboplatin. *Invest New Drugs* 2015;**33**:1187-96.

Supplementary Table S1. Previous studies reporting the putative tumor suppressor roles of GSK3 β .

Cancer type	Species	Summary of results	Effect of GSK3 β inhibition on tumor cells*	Ref. No.
Colon	human	Stimulation of Wnt signaling by mutant K-ras ^{Val12} was associated with inhibition of GSK3 β activity in Caco-2 cancer cells.	Not examined	SR30
Stomach	human	Inhibition of GSK3 β activity by pharmacological inhibitors induced expression of COX-2 mRNA and protein as well as the enzyme activity in TMK-1 and MKN-28 cancer cells.	Not examined	SR31
Pancreas	human	LiCl, GSK3 β -siRNA or a kinase-dead mutant GSK3 β transfection resulted in radioresistance of PANC-1 and BxPC-3 cancer cells, which was associated with stabilization of β -catenin and expression of its target gene.	GSK3 β inhibition resulted in radio-resistance and its overexpression in radio-sensitization in cancer cells.	SR32
	human	Pancreatic cancer patients with higher expression of GSK3 β in the tumors had a reduced risk of dying of pancreatic cancer.	Not examined	SR33
Liver	human	LiCl and SB-415286 repressed chemotherapeutic drugs induction of HepG2 cell apoptosis by inhibiting CD95 expression and caspase-8 activity and by disrupting nuclear GSK3 β -p53 complexes.	GSK3 β inhibitors render the cancer cells insusceptible to etoposide and camptothecin.	SR34
	human	PI3K inhibitor LY294002 sensitized HepB3 cells to etoposide and camptothecin by enhancing the expression of DR4 and DR5 and by decreasing pGSK3 β ^{S9} .	No direct effect was examined. SB-415286 repressed the chemosensitizing effect by LY294002 in the cancer cells.	SR35
	human	Decreased TSC2 and GSK3 β expression in HCC tumors was significantly correlated with advanced clinico-pathological characteristics and poor prognosis of the patients.	Not examined.	SR36
	human	Overexpression of pGSK3 β ^{S9} in HCC tumors was significantly associated with the presence of type 2 DM and with poor prognosis of the patients.	Not examined.	SR37

	human	Ectopic expression of SIRT3 (a class III histone deacetylase) inhibited proliferation and inhibited apoptosis in HCC cells, which was associated with deacetylation of GSK3 β and decreased pGSK3 β ^{S9} .	No direct effect was examined. GSK3 β inhibitor reversed the SIRT3-induced proliferation inhibition and apoptosis in cancer cells.	SR38
Prostate	human	Transfection of wild-type and constitutively active mutant GSK3 β repressed AR-mediated transactivation in cancer cells.	No direct effect was examined. Transfection of kinase-dead mutant GSK3 β showed little effect on the AR transactivation in the cancer cells. LiCl abolished AR transactivation by GSK3 β .	SR39
	human	A pharmacological GSK3 β inhibitor, AR79, promotes cancer cell proliferation in soft tissue and bone in mice by dephosphorylation and stabilization of β -catenin.	GSK3 β inhibitor promotes the cancer cell proliferation in mice.	SR40
Ovary	human	Level of pGSK3 β ^{S9} but not total GSK3 β and pGSK3 β ^{Y216} was higher in cisplatin-resistant derivative of cancer cells than the parental cells.	No direct effect was examined. LiCl counteracted cisplatin-induced apoptosis in both parental and resistant cancer cells.	SR41
	human	Inhibition of GSK3 β by SB-216763 increased MSX2 oncogenic factor via activation of β -catenin signaling in endometrioid cancer cells.	Not examined.	SR42
Uterine cervix (HeLa cells)	human	Inhibition of Akt enhances doxorubicin- or paclitaxel-induced apoptosis in cancer cells, which was associated with decrease in the level of pGSK3 β ^{S9} and the binding of hexokinase II to mitochondria.	No direct effect was examined. GSK3 β siRNA reversed the effect of Akt inhibitor on chemosensitivity of the cancer cells.	SR43
Breast	human	GSK3 β inhibitors (LiCl, SB-216763 and SB-415286) decreased rapamycin-induced down regulation of cyclin D1, but not inhibit cell cycle G1 arrest in cancer cells. Rapamycin enhances paclitaxel-induced cytotoxicity in GSK3 β wild-type but GSK3 β -null cancer cells.	No direct effect was examined. GSK3 β inhibition reversed rapamycin-induced down regulation of cyclin D1 expression in cancer cells.	SR44
	mouse	Transgenic mice overexpressing kinase-inactive GSK3 β under the control of the mouse mammary tumor virus-long terminal repeat developed mammary tumors with overexpression of β -catenin and cyclin D1.	Not examined.	SR45

	human	Adiponectin attenuated cancer cell proliferation by suppression of Akt phosphorylation and pGSK3 β ^{S9} in association with accumulation and activation of β -catenin.	No direct effect was examined. LiCl reversed the effect of adiponectin in cancer cells.	SR46
	human	Therapeutic effect of prodigiosin, a bacterial metabolite, against cancer cells was associated with increased expression of NAG-1 via Akt dephosphorylation (inactivation).	No direct effect was examined. GSK3 β inhibition with AR-A014418 reversed the effect of prodigiosin against the cancer cells.	SR47
	human	GSK3 β phosphorylates Mcl-1 (proto-oncoprotein) for β -TrCP-mediated ubiquitination and proteasomal degradation in cancer cells.	Not examined.	SR48
	human	Expression of Mcl-1 was correlated with pGSK3 β ^{S9} in multiple cancer cell lines and primary cancer samples, and was significantly linked with poor prognosis of human breast cancer.	Not examined.	SR49
	human	GSK3 β phosphorylates securin to promote its degradation via β -TrCP. A significant correlation between securin accumulation and pGSK3 β ^{S9} was observed in breast cancer tissues.	Not examined. Level of tumor pGSK3 β ^{S9} was correlated with Ki-67 proliferative index and tumor grades in breast cancer.	SR50
	mouse	Genetic deletion of GSK3 in mammary epithelial cells resulted in β -catenin activation and induced intraepithelial neoplasia that progressed to development of adenosquamous carcinoma. Mammary-specific knockout of GSK3 and β -catenin induced adenocarcinoma.	Not examined.	SR51
Lung	human	Constitutively active mutant GSK3 β transfected in A549 cells binds to survivin, resulting in G1 cell-cycle arrest, apoptosis and sensitization to doxorubicin.	Dominant-negative mutant GSK3 β and LiCl increased survivin expression, leading to cell-cycle progression and resistance to apoptosis.	SR52
	human	The level of pGSK3 β ^{S9} was associated with expression of Slug, a transcriptional repressor of E-cadherin, in cancer cells and non-small cell lung cancer. GSK3 β -	Not examined.	SR53

		mediated phosphorylation of Slug facilitated Slug protein degradation.		
	human	Expression of a constitutively active GSK3 β sensitized cancer cells to mTOR inhibitors. Higher basal levels of GSK3 β activity in cancer cell lines correlated with more efficacious responses to the inhibitors.	No direct effect was examined. Pharmacologic inhibition and genetic depletion of GSK3 β antagonized the effects of mTOR inhibitors against cancer cells.	SR54
Skin	mouse	The level of pGSK3 β ^{S9} was higher and that of pGSK3 β ^{Y216} was lower in the later stage of chemically-induced two-stage skin carcinogenesis mouse model.	Not examined.	SR55
	mouse	The level of pGSK3 β ^{S9} in skin carcinoma was weaker than normal skin. However, its level in TPA-mediated transformation-sensitive epidermal cells was higher than the transformation-resistant cells.	No direct effect was examined. Overexpression of wild-type and constitutively active mutant GSK3 β in the TPA-mediated transformation-resistant epidermal cells suppressed EGF- and TPA-mediated anchorage-independent growth in soft agar and tumorigenicity in nude mice.	SR56
Melanoma	human	A multikinase inhibitor sorafenib activates GSK3 β via inhibition of its upstream kinases and alters subcellular localization of p53 to induce apoptosis in B-raf mutant melanoma cells.	No direct effect was examined. GSK3 β shRNA reversed and constitutively active mutant GSK3 β facilitated the effect of sorafenib against tumor cells.	SR57
Neuroblastoma	human	BDNF activation of TrkB induced the Akt-dependent pGSK3 β ^{S9} , resulting in its inactivation. Treatment of neuroblastoma cells with inhibitors of GSK3 β , LiCl, GSK3 β inhibitor VII, kenpaullone, or a GSK3 β -siRNA resulted in a 15% to 40% increase in neuroblastoma cell survival after treatment with etoposide or cisplatin.	GSK3 β inhibition enhanced the survival of neuroblastoma cells after cytotoxic treatment.	SR58

*Direct effect of pharmacological GSK3 β inhibitors and/or genetic depletion of GSK3 β expression (e.g., RNA interference) or its activity (e.g., recombinant kinase-dead form) on tumor cell survival, proliferation, invasive ability and susceptibility to therapy.

Abbreviations: AR, androgen receptor; BDNF, brain-derived neurotropic factor; DM, diabetes mellitus; DR4, 5, death receptor 4, 5; EGF, epidermal growth factor; GSK3 β , glycogen synthase kinase 3 β ; HCC, hepatocellular carcinoma; LiCl, lithium chloride (classical but not specific

GSK3 β inhibitor); Mcl-1, myeloid cell leukemia-1; mTOR, mammalian target of rapamycin; MSX2, msh homeobox 2; NAG-1, nonsteroidal anti-inflammatory drug activated gene 1; pGSK3 β ^{S9}, GSK3 β phosphorylated at seine 9 residue (inactive form); pGSK3 β ^{Y216}, GSK3 β phosphorylated at tyrosine 216 residue (active form); PI3K, phosphatidylinositol 3-kinase; shRNA, short hairpin RNA; siRNA, small interfering RNA; SIRT3, sirtuin 3; TPA, 12-*O*-tetradecanoylpholbor-13-acetate; β -TrCP, β -transducin repeats-containing protein; TrkB, tyrosine kinase receptor B; TSC2, tuberous sclerosis protein 2;

Supplementary Table S2. Clinical trials of GSK3 β inhibitors for treatment of diseases

GSK3 β inhibitor (Company)	Disease	Trial ID and phase	Combined regimen	URL (access date: July 5, 2016)	Reference
AZD-1080 (AstraZeneca)	Alzheimer's disease	Phase I	none	https://ja.scribd.com/doc/851553/AstraZeneca-Therapy-R-D-Pipeline-Summary-December-7-2007	
NP031112/tideglusive (Noscira SA)	Progressive supranuclear palsy	NCT01049399 Phase IIb	none	https://clinicaltrials.gov/ct2/show/NCT01049399	SR60,61
	Alzheimer's disease	NCT01350362 Phase II	none	https://clinicaltrials.gov/ct2/show/NCT01350362	SR62,63
LY2090314 (Eli Lilly)	Acute leukemia	NCT01214603 Phase II	none	https://clinicaltrials.gov/ct2/show/NCT01214603	
	Metastatic pancreatic cancer	NCT01632306 Phase I/II	Gemcitabine, FOLFOX, or Gemcitabine + nab-paclitaxel	https://clinicaltrials.gov/ct2/show/NCT01632306	
	Advanced or metastatic solid cancer	NCT01287520 Phase I	Pemetrexed + carboplatin	https://clinicaltrials.gov/show/NCT01287520	SR64,65
CLOVA cocktail*	Advanced pancreatic cancer	UMIN000005095 Phase I/II	Gemcitabine	https://upload.umin.ac.jp/cgi-open-bin/ctr/ctr.cgi?function=brows&action=brows&type=summary&recptno=R000006032&language=E	
	Recurrent glioblastoma	UMIN000005111 Phase I/II	Temozolomide	https://upload.umin.ac.jp/cgi-open-bin/ctr/ctr.cgi?function=brows&action=brows&type=summary&recptno=R000002506&language=E	*Furuta T, et al.

Abbreviations: CLOVA, combined cimetidine, lithium chloride, olanzapine and valproate regimen; FOLFOX, combined folate, 5-fluorouracil and oxaliplatin regimen; SR, supplementary reference No.

*Furuta T, et al., unpublished data