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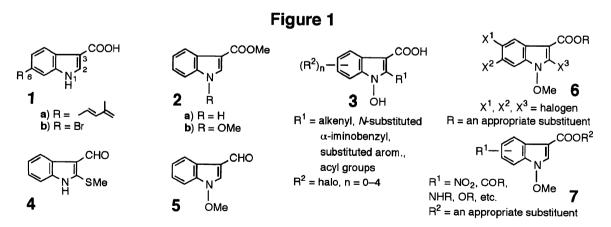
SYNTHESIS OF ANALOGS OF WASABI PHYTOALEXIN (METHYL 1-METHOXYINDOLE-3-CARBOXYLATE) 1

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Abstract — Syntheses of wasabi phytoalexin analogs, such as 6-bromo-5-iodo, 2-bromo-5-iodo, 6-nitro, 5-chloroacetyl, and 6-chloroacetyl congeners, are reported. An interesting effect of the 1-methoxy group on the regioselectivity of electrophilic substitution reactions on indole nucleus is observed.

Indoles having a carboxy or a formyl group at the 3-position, including 1-hydroxy- or 1-methoxyindole derivatives, exhibit marked physiological activities in spite of their simple chemical structures.² Compounds (1-3)^{2a-d} are a few representative examples (Scheme 1). Brassicanal A³ (4) and 1-methoxy-indole-3-carbaldehyde^{3a} (5) are additional examples isolated by Takasugi³ and co-workers as phytoalexins of plant family, *Cruciferae*. Recently, Pedras⁴ and co-workers determined methyl 1-methoxy-indole-3-carboxylate (2b) as a phytoalexin of Wasabi (*Wasabia japonica*, syn. *Eutrema wasabi*).

We have been much interested in determining the effect of 1-methoxy and 1-hydroxy groups on both the chemical reactivities⁵ and biological activities⁶ of indole compounds, aiming at preparing our own biologically active substances. From these points of view, we needed various kinds of analogs⁷ of wasabi phytoalexin like **6** and **7** as shown in general formula.

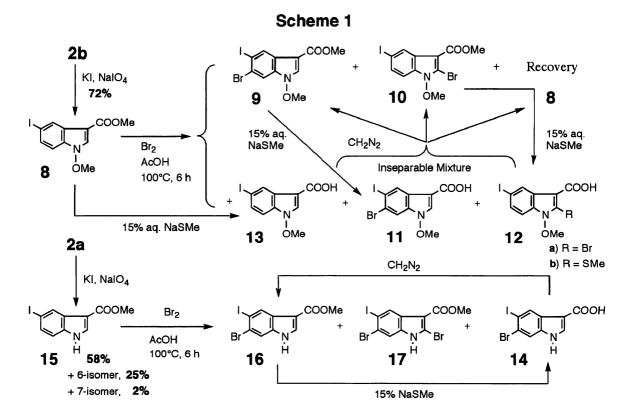


In the previous papers, ⁷ we established simple synthetic method for **2b** and its regioselective iodination to give methyl 5-iodo-1-methoxyindole-3-carboxylate ^{7a} (8). Based on these results, synthesis of methyl 6-bromo-5-iodo-1-methoxyindole-3-carboxylate (9), a representative of 6, seemed to be easy. In fact, however, bromination of 8 gave complex mixtures of products under various reaction conditions.

Relatively clean reaction took place with Br_2 (1.3 mol eq.) in AcOH at 100° C to provide **9**, methyl 2-bromo-5-iodo-1-methoxyindole-3-carboxylate (**10**), 6-bromo- (**11**), 2-bromo-5-iodo-1-methoxyindole-3-carboxylic acid (**12a**), and 5-iodo-1-methoxyindole-3-carboxylic acid (**13**) in 17, 33, 6, 5, and 7% yields, respectively, together with 16% yield of unreacted **8**. Since the carboxylic acids (**11**, **12a**, and **13**) were inseparable, the mixture was methylated with diazomethane (CH_2N_2) to give **9**, **10**, and **8** in 6, 5, and 7% yields, respectively. Assuming that the methylation proceeded in a quantitative yield, the yields of **11**, **12a**, and **13** in the initial bromination products were estimated. Pure compound (**13**) was obtained in 38% yield by reacting **8** with 15% aq. NaSMe. Under similar reaction conditions, **9** provided **11** and **14** in 45 and 38% yields, respectively. In the same reaction of **10**, the expected compound (**12a**) was not produced at all, instead nucleophilic substitution reaction by methyl sulfide occurred at the 2-position to afford **12b** in 83% yield. As a result, pure **12a** has not been obtained yet.

On the other hand, bromination (1.3 mol eq.) of methyl 5-iodoindole-3-carboxylate (15) in AcOH at 100° C provided methyl 6-bromo- (16), 2,6-dibromo-5-iodoindole-3-carboxylate (17), and 6-bromo-5-iodoindole-3-carboxylic acid (14) in 48, 8, and 17% yields, respectively. In the same reaction, when 0.85 mol eq. of bromine was employed, 16 and 14 were produced in 56 and 18% yields, respectively, without formation of 17. The structure of 14 was confirmed by the direct comparison with the sample obtained in 92% yield by the treatment of 16 with 15% NaSMe under reflux. Furthermore, methylation of 14 with CH₂N₂ in MeOH provided 16 in 90% yield.

Above results exhibited that the bromination of 15 proceeds regioselectively at the 6-position, while the



introduction of methoxy group into the 1-position of **15** alters the reaction site and tends to favor the 2-po- sition. These observations are quite interesting considering the initial iodination of **2a** and **2b**. Thus, **2b** produced **8** exclusively in 72% yield, ^{7a} while **2a** afforded **15** together with 6- and 7-iodo isomers in 58, 25, and 2% yields. ^{7a} Further reaction examples would be needed for clarifying the effect of 1-methoxy group on the regioselectivity of electrophilic substitution reactions.

We next tried the preparation of methyl 1-methoxy-6-nitroindole-3-carboxylate (18b, Scheme 2) as a common intermediate for 7, because the nitro group can easily be transformed into other functional groups such as amino, halogen, hydroxy, etc. According to our procedure, 8 1-methoxy-6-nitroindole (19) was prepared from an industrial raw material, 2,3-dihydroindole, in three steps in 71% overall yield. Subsequent Vilsmeier reaction of 19 with POCl₃ and N,N-dimethylformamide provided a 94% yield of 1-methoxy-6-nitroindole-3-carbaldehyde (20), an analog of daikon phytoalexin^{3a} (5). Oxidation of 20 to 1-methoxy-6-nitroindole-3-carboxylic acid (18a) was achieved in 92% yield by the treatment with NaClO₂⁹ in the presence of 2-methyl-2-butene. Methylation of 18a with CH₂N₂ in MeOH provided the desired 18b in 99% yield.

Scheme 2

For the preparation of wasabi phytoalexin analogs having an acyl group, direct acylation of $\bf 2b$ was examined. Thus, Friedel-Crafts acylation using chloroacetyl chloride and $AlCl_3$ in nitrobenzene at $100^{\circ}C$ provided cleanly methyl 5-chloroacetyl- ($\bf 21a$) and 6-chloroacetyl-1-methoxyindole-3-carboxylate ($\bf 22a$) in 52 and 20% yields, respectively, and the ratio of $\bf 21a$ to $\bf 22a$ was 2.5:1. The acylated position of $\bf 21a$ is determined by the presence of C4-H signal at 8.80 ppm (d, $\it J=1.7$ Hz) in its $\it ^1H$ -NMR spectrum, while the corresponding C4-H signal of $\bf 22a$ is observed at 8.26 ppm (d, $\it J=8.5$, 0.7 Hz) proving the 6-substituted indole structure.

Similar results were already reported by Nakatsuka¹⁰ and co-workers in the chloroacetylation of methyl 1-methylindole-3-carboxylate (23). In their experiment, however, 21b and 22b were obtained as an

inseparable mixture. Based on the ratio of the following Baeyer-Villiger oxidation products, the ratio of **21b** to **22b** could be estimated to be 3:1. Similar Friedel-Crafts acylation of indoles having an electron withdrawing group has been reported by several researchers. ¹¹ Comparing these results, it would be safe to say that the introduction of a methoxy group into the 1-position prefers the 6-substitution to 5-substitution in Friedel-Crafts acylation.

In conclusion, introduction of a methoxy group into the 1-position is found to influence significantly the regiochemistry of electrophilic substitution reactions on indole nucleus. With key building blocks such as 9, 10, 15, 18, 21a, and 22a in hand, preparations of various kinds of wasabi phytoalexin analogs are now in progress to conduct structure-activity relationship study.

EXPERIMENTAL

Melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. IR spectra were determined with a Horiba FT-720 spectrophotometer, and ¹H-NMR spectra with a JEOL GSX-500 spectrometer with tetramethylsilane as an internal standard. MS spectra were recorded on a JEOL SX-102A spectrometer. Column chromatography was performed on silica gel (SiO₂, 100-200 mesh, from Kanto Chemical Co., Inc.) throughout the present study.

Methyl 6-Bromo- (9), 2-Bromo-5-iodo-1-methoxyindole-3-carboxylate (10), 6-Bromo-(11), 2-Bromo- (12a), and 5-iodo-1-methoxyindole-3-carboxylic acid (13) from Methyl 5-Iodo-1-methoxyindole-3-carboxylate (8) — A solution of Br₂ (3.8 mL, 0.76 mmol) in AcOH was added to a solution of 8 (192.7 mg, 0.58 mmol) in AcOH (10mL) and the mixture was heated at 100°C for 6 h with stirring under Ar atmosphere. After addition of H₂O under ice cooling, the whole was extracted with CHCl₃-MeOH (9:1, v/v). The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave an oil, which was column-chromatographed repeatedly on SiO₂ with CHCl₃-hexane (1:1, v/v) and then with CHCl 3 to give **10** (80 mg, 33%), **9** (40.7 mg, 17%), 8 (30.1 mg, 16%), and a mixture fraction (11, 12a, and 13, 45.6 mg) in the order of elution. 9: mp 142.5—143.0°C (colorless fine needles, recrystallized from CHCl₃-hexane). IR (KBr): 1695, 1517, 1446, 1205 cm⁻¹. ¹H-NMR (CDCl₃) δ: 3.91 (3H, s), 4.14 (3H, s), 7.79 (1H, s), 7.90 (1H, s), 8.69 (1H, s). MS m/z: 410 (M+: Br⁸¹), 408 (M+: Br⁷⁹). Anal. Calcd for C₁₁H₉NO₃BrI: C, 32.22; H, 2.21; N, 3.42. Found: C, 32.28; H, 2.22; N, 3.43. 10: mp 158—159°C (colorless fine needles, recrystallized from CHCl₃-hexane). IR (KBr): 1722, 1498, 1195, 1178, 1016 cm⁻¹. ¹H-NMR (CDCl₃) δ: 3.96 (3H, s), 4.14 (3H, s), 7.22 (1H, d, J=8.5 Hz), 7.59 (1H, dd, J=8.5, 1.5 Hz), 8.50 (1H, d, J=1.5 Hz). MS m/z: 410 (M+: Br⁸¹), 408 (M+: Br⁷⁹). Anal. Calcd for C₁₁H₉NO₃BrI·1/2H₂O: C, 31.53; H, 2.17; N, 3.34. Found: C, 31.21; H, 2.19; N, 3.28. Pure samples, (11) and (13), were obtained from 9 and 8, respectively, by the reaction with 15% aq. NaSMe as shown below.

Separation of 11, 12a, and 13 as Methylated Products, (9), (10), and (8) — Excess CH₂N₂ in Et₂O was added to a solution of the mixture fraction (11, 12a, and 13, 45.6 mg), obtained in the above experiment, in MeOH (5.0 mL) and the whole was stirred at rt for 10 min. The solvent was evaporated under reduced pressure to leave an oil, which was column-chromatographed on SiO₂ with CHCl₃-hexane (1:2, v/v) to give 10 (11.9 mg, 5%), 9 (14.0 mg, 6%), and 8 (12.6 mg, 7%).

6-Bromo-5-Iodo-1-methoxyindole-3-carboxylic acid (11) and 14 from 9 — Excess 15% aq. NaSMe (8 mL) was added to a solution of 9 (51.8 mg, 0.13 mmol) in MeOH (3 mL) and the mixture was refluxed for 2 h with stirring under Ar atmosphere. After addition of H₂O under ice cooling, the whole was made acidic (pH 1) by adding 8% HCl and extracted with EtOAc. The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave a solid, which was column-chromatographed on SiO₂ with CHCl₃-MeOH-AcOH (46:1:0.1, v/v) to give 11 (22.5 mg, 45%) and 14 (17.8 mg, 38%) in the order of elution. 11: mp 275—280°C (decomp, colorless fine needles, recrystallized from CHCl₃-MeOH). IR (KBr): 1658, 1520, 1217 cm⁻¹. ¹H-NMR (5% CD₃OD in CDCl₃) δ: 4.14 (3H, s), 7.80 (1H, s), 7.93 (1H, s), 8.71 (1H, s). MS m/z: 397 (M+: Br⁸¹), 395 (M+: Br⁷⁹). Anal. Calcd for C₁₀H₇NO₃BrI: C, 30.33; H, 1.78; N, 3.54. Found: C, 30.54; H, 1.85; N, 3.52.

5-Iodo-1-methoxy-2-methylthioindole-3-carboxylic acid (12b) from 10 — Excess 15% aq. NaSMe (2 mL) was added to a solution of 10 (23.8 mg, 0.06 mmol) in MeOH (2 mL) and the mixture was refluxed for 1 h with stirring under Ar atmosphere. After addition of H₂O under ice cooling, the whole was made acidic (pH 3) by adding 8% HCl and extracted with EtOAc-MeOH (9:1, v/v). The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave a solid, which was column-chromatographed on SiO₂ with CHCl₃-MeOH-AcOH (46:1:0.5, v/v) to give 12b (17.4 mg, 83%). 12b: mp 167—168°C (decomp, colorless fine needles, recrystallized from CHCl₃-hexane). IR (KBr): 1668, 1492, 1205 cm⁻¹. ¹H-NMR (CDCl₃) δ: 2.68 (3H, s), 4.16 (3H, s), 7.21 (1H, dd, *J*=8.5, 0.7 Hz), 7.61 (1H, dd, *J*=8.5, 1.5 Hz), 8.62 (1H, dd, *J*=1.5, 0.7 Hz). High-Resolution MS *m*/z:: Calcd for C₁₁H₁₀NO₃IS: 362.9426. Found: 362.9429.

5-Iodo-1-methoxyindole-3-carboxylic acid (13) from 8 — Excess 15% aq. NaSMe (5.0 mL) was added to a solution of 8 (32.2 mg, 0.9 mmol) in MeOH (2 mL) and the mixture was refluxed for 1 h with stirring under Ar atmosphere. After addition of H₂O under ice cooling, the whole was made acidic (pH 3) by adding 8% HCl and extracted with EtOAc-MeOH (95:5, v/v). The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave a solid, which was column-chromatographed on SiO₂ with CHCl₃-MeOH-AcOH (46:1:0.1, v/v) to give 13 (11.7 mg, 38%). 13: mp 211—213°C (colorless fine needles, recrystallized from CHCl₃-hexane). IR (KBr): 1670, 1525, 1220 cm⁻¹. ¹H-NMR (DMSO- d_6) δ : 4.13 (3H, s), 7.42 (1H, d, J=8.6 Hz), 7.58 (1H,dd, J=8.6, 1.7 Hz), 8.35 (1H, s), 8.39 (1H, d, J=1.7 Hz). MS m/z: 317 (M⁺). Anal. Calcd for C₁₀H₈NO₃I·1/2 H₂O: C, 36.83; H, 2.78; N, 4.30. Found: C, 36.95; H, 2.57; N, 4.35.

Methyl 6-Bromo- (16), 2,6-Dibromo-5-iodoindole-3-carboxylate (17), and 6-Bromo-5-iodoindole-3-carboxylic acid (14) from 15 — [Method 1] A solution of Br₂ (2.2 mL, 0.43 mmol) in AcOH was added to a solution of 15 (100.8 mg, 0.33 mmol) in AcOH (5.0 mL), and the mixture was heated at 100°C for 6 h with stirring under Ar atmosphere. After addition of H ₂O under ice cooling, the whole was extracted with CHCl₃-MeOH (9:1, v/v). The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave an oil, which was column-chromatographed repeatedly on SiO₂ with acetone-benzene (1:9, v/v) and then with CH₂Cl₂-hexane (2:1, v/v) to give 17 (12.7 mg, 8%), 16 (61.5 mg, 48%), and 14 (20.5 mg, 17%) in the order of elution. 16:

mp 262.5—263°C (colorless prisms, recrystallized from MeOH). IR (KBr): 3270, 1677, 1520, 1441, 1198, 1053 cm⁻¹. 1 H-NMR (DMSO- d_6) δ : 3.82 (3H, s), 7.88 (1H, s), 8.12 (1H, s), 8.52 (1H, s), 12.10 (1H, br s, disappeared on addition of D₂O). MS m/z: 381 (M⁺: Br⁸¹), 279 (M⁺: Br⁷⁹). Anal. Calcd for C₁₀H₇NO₂BrI: C, 31.61; H, 1.86; N, 3.69. Found: C, 31.74; H, 1.73; N, 3.74. **17**: mp 275—277°C (colorless fine needles, recrystallized from CHCl₃-MeOH). IR (KBr): 3226, 1681, 1504, 1430, 1382, 1062 cm⁻¹. 1 H-NMR (DMSO- d_6) δ : 3.85 (3H, s), 7.74 (1H, s), 8.47 (1H, s), 13.05 (1H, br s, disappeared on addition of D₂O). MS m/z: 460 (M⁺: 2 x Br⁸¹), 458 (M⁺: Br⁸¹, Br⁷⁹), 456 (M⁺: 2 x Br⁷⁹). Anal. Calcd for C₁₀H₆NO₂Br₂I·1/4H₂O: C, 25.92; H, 1.41; N, 3.02. Found: C, 25.80; H, 1.44; N, 2.95. **14**: mp 224—226°C (decomp, brown prisms, recrystallized from EtOAc-MeOH). IR (KBr): 3249, 1637, 1531, 1186 cm⁻¹. 1 H-NMR (DMSO- d_6) δ : 7.85 (1H, s), 8.02 (1H, s), 8.54 (1H, s), 11.96 (1H, br s, disappeared on addition of D₂O). MS m/z: 367 (M⁺: Br⁸¹), 365 (M⁺: Br⁷⁹). Anal. Calcd for C₉H₅NO₂BrI·1/2H₂O: C, 28.82; H, 1.61; N, 3.74. Found: C, 28.68; H, 1.51; N,3.67.

[Method 2] A solution of Br₂ in AcOH (1.5 mL, 0.30 mmol) was added to a solution of 15 (108.3 mg, 0.36 mmol) in AcOH (5.0 mL), and the mixture was heated at 100°C for 6 h with stirring under Ar atmosphere. After the same work-up and column-chromatography as described in the [Method 1], 16 (76.9 mg, 56%) and 14 (23.8 mg, 18%) were obtained.

Methyl 6-Bromo-5-iodoindole-3-carboxylate (16) from 14 — Excess CH₂N₂ in Et₂O was added to a solution of 14 (13.0 mg, 0.036 mmol) in MeOH (10 mL) and the mixture was stirred at rt for 30 min. The solvent was evaporated under reduced pressure to leave a solid, which was column-chromatographed on SiO₂ with CHCl₃-MeOH (95:5, v/v) to give 16 (12.1 mg, 90%).

6-Bromo-5-iodoindole-3-carboxylic acid (14) from 16 — Excess 15% aq. NaSMe (8.0 mL) was added to a suspension of 16 (50.9 mg, 0.13 mmol) in MeOH (3 mL) and the mixture was refluxed for 2 h with stirring under Ar atmosphere. After addition of H₂O under ice cooling, the whole was made acidic (pH 3) by adding 8% HCl and extracted with CHCl₃-MeOH (9:1, v/v). The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave a solid, which was column-chromatographed on SiO₂ with CHCl₃-MeOH (95:5, v/v) to give 14 (45.3 mg, 92%).

1-Methoxy-6-nitroindole-3-carboxylic acid (18a) from 20 — A solution of sodium chlorite (1.186 g, 13.1 mmol) and sodium dihydrogen phosphate (1.535 g, 9.84 mmol) in H₂O (6 mL) was added to a solution of 20 (144.4 mg, 0.66 mmol) and 2-methyl-2-butene (3.0 mL) in DMF (10 mL), and the mixture was stirred at rt for 1.5 h. The whole was made acidic (pH 3) by adding 8% HCl under ice cooling, and extracted with EtOAc. The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave a solid, which was column-chromatographed on SiO₂ with CHCl₃-MeOH-AcOH (46:1:0.1, v/v) to give 18a (143.2 mg, 92%). 18a: mp 232.5—233°C (pale yellow fine needles, recrystallized from CHCl₃-hexane). IR (KBr): 1682, 1516, 1348, 1221 cm⁻¹. ¹H-NMR (DMSO-d₆) δ: 4.23 (3H, s), 8.11 (1H, dd, J=9.0, 2.0 Hz), 8.24 (1H, d, J=9.0 Hz), 8.46 (1H, d, J=2.0 Hz), 8.77 (1H, s). MS m/z: 236 (M⁺). Anal. Calcd for C₁₀H₈N₂O₅: C, 50.85; H, 3.41; N, 11.86. Found: C, 50.64; H, 3.40; N, 11.83.

Methyl 1-Methoxy-6-nitroindole-3-carboxylate (18b) from 18a — Excess CH₂N₂ in Et₂O was added to a suspension of 18a (11.1 mg, 0.047 mmol) in MeOH (2.0 mL) and the mixture was

stirred at rt for 15 min. After evaporation of the solvent under reduced pressure, the residue was column-chromatographed on SiO₂ with CHCl₃-hexane (2:1, v/v) to give **18b** (11.6 mg, 99%). **18b**: mp 155.5—156°C (pale yellow fine needles, recrystallized from CHCl₃-hexane). IR (KBr): 1716, 1514, 1352, 1213 cm⁻¹. 1 H-NMR (CDCl₃) δ : 3.94 (3H, s), 4.23 (3H, s), 8.16 (1H, dd, J=9.0, 2.0 Hz), 8.18 (1H, s), 8.28 (1H, d, J=9.0 Hz), 8.43 (1H, d, J=2.0 Hz). MS m/z: 250 (M⁺). Anal. Calcd for C₁₁H₁₀N₂O₅·1/8H₂O: C, 52.33; H, 4.09; N, 11.10. Found: C, 52.21; H, 4.05; N, 11.11.

1-Methoxy-6-nitroindole-3-carbaldehyde (20) from 1-Methoxy-6-nitroindole (19) — POCl₃ (0.97 mL, 10.4 mmol) was added to dry DMF (5.8 mL, 63 mmol) at 0°C and stirring was continued for 15 min. A solution of 19 (1.00 g, 5.2 mmol) in dry DMF (15 mL) was added to the resultant mixture at 0°C. After stirring at rt for 7 h, H₂O was added to the reaction mixture at 0°C and the whole was made basic (pH 10) by adding sat. aq. NaHCO₃. After stirring at rt for 30 min maintaining pH 10 by adding sat. aq. NaHCO₃, the whole was extracted with EtOAc. The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave yellow solid, which was column-chromatographed on SiO₂ with EtOAc-hexane (1:2, v/v) to give 20 (1.08 g, 94%). 20: mp 180—182°C (yellow prisms, recrystallized from CHCl₃-hexane). IR (KBr): 1664, 1653, 1508, 1342 cm⁻¹. ¹H-NMR (CDCl₃) δ: 4.28 (3H, s), 8.14 (1H, s), 8.22 (1H, dd, J=8.8, 2.0 Hz), 8.43 (1H, d, J=8.8 Hz), 8.45 (1H, d, J=2.0 Hz), 10.02 (1H, s). *Anal*. Calcd for C₁₀H₈N₂O₄: C, 54.55; H, 3.66; N, 12.72. Found: C, 54.32; H, 3.61; N, 12.54.

Methyl 5-Chloroacetyl- (21a) and Methyl 6-Chloroacetyl-1-methoxyindole-3-carboxylate (22a) from 2b — Anhydrous AlCl₃ (392.7 mg, 2.94 mmol) and then a solution of chloroacetyl chloride (166.1 mg, 1.47 mmol) in nitrobenzene (0.4 mL) was added successively to a solution of 2b (100.5 mg, 0.49 mmol) in nitrobenzene (1 mL) and the mixture was heated at 100°C for 15 min with stirring. After addition of H₂O with stirring under ice cooling, the solvent was evaporated under reduced pressure. The residue was made basic (pH 11) by adding 8% NaOH and extracted with CHCl₃. The extract was washed with brine, dried over Na₂SO₄, and evaporated under reduced pressure to leave an oil, which was column-chromatographed repeatedly on SiO2 with CH2Cl2 and CHCl3 to give 22a (26.9 mg, 20%) and **21a** (71.3 mg, 52%) in the order of elution. **21a**: mp 138-139 °C (pale brown prisms, recrystallized from CHCl₃-hexane). IR (KBr): 1700, 1610, 1218, 1178 cm⁻¹. ¹H-NMR (CDCl₃) δ: 3.95 (3H, s), 4.18 (3H, s), 4.84 (2H, s), 7.55 (1H, dd, J=8.7, 1.0 Hz), 8.01 (1H, dd, J=8.7, 1.7 Hz), 8.03 (1H, s), 8.80 (1H, d, J=1.7 Hz). MS m/z: 283 (M+: Cl³⁷), 281 (M+: Cl³⁵). Anal. Calcd for C₁₃H₁₂NO₄Cl·1/8H₂O: C, 54.99; H, 4.35; N, 4.96. Found: C, 54.83; H, 4.29; N, 4.84. **22a**: mp 156—158 °C (colorless prisms, recrystallized from CHCl₃-hexane). IR (KBr): 1695, 1616, 1437, 1387, 1197 cm⁻¹. 1 H-NMR (CDCl₃) δ : 3.93 (3H, s), 4.21 (3H, s), 4.79 (2H, s), 7.85 (1H, dd, J=8.5, 1.5 Hz), 8.12 (1H, s), 8.16 (1H, dd, J=1.5, 0.7 Hz), 8.26 (1H, dd, J=8.5, 0.7 Hz). High Resolution MS m/z: Calcd. for C₁₃H₁₂NO₄Cl: 283.0425 (M⁺: Cl³⁷) and 281.0455 (M⁺: Cl³⁵). Found: 283.0427 and 281.0427.

REFERENCES AND NOTES

1. a) This report is Part 115 of a series entitled "The Chemistry of Indoles". b) Part 114: K. Yamada, F.

- Yamada, and M. Somei, Heterocycles, 2002, 57, 1231.
- a) N. Zelikovitch, Z. Eyal, and Y.Kashman, Phytopathology, 1992, 82, 275; b) W. R. Schleigh and T. R. Welter, Eur. Pat. Appl. Ep 470,665 (Chem. Abstr., 1992, 116, 250503n). c) J. Jensen, U. Anthoni, C. Christophersen, and P. H. Nielsen, 1993, J. Nat. Prod., 56, 1553; d) M. Sakurai, J. Kawano, and N. Nakanishi, J. Antibiotics, 2001, 54, 628; e) L. M. Levy, G. M. Gabrera, J. E. Wright, and A. M. Seldes, Phytochemistry, 2000, 54, 941; f) J. -F. Hu, D. Wunderlich, I. Sattler, A. Härtl, I. Papastavrou, S. Grond, S. Grabley, X. -Z. Feng, and R. Thiericke, J. Antibiot., 2000, 53, 944.
- 3. a) M. Takasugi, K. Monde, N. Katsui, and A. Shirota, Symposium Papers, The 29th Symposium on The Chemistry of Natural Products, Sapporo, Aug. 1987, p. 629; b) M. Takasugi, K. Monde, N. Katsui, and A. Shirata, *Bull. Chem. Soc. Japan*, 1988, **61**, 285; M. Somei, T. Kawasaki, Y. Fukui, F. Yamada, T. Kobayashi, H. Aoyama, and D. Shinmyo, *Heterocycles*, 1992, **61**, 1877; M. Takasugi, *Kagaku to Seibutu*, 1993, **31**, 22 and references cited therein.
- 4. M. S. C. Pedras and J. L. Sorensen, Phytochemistry, 1998, 49, 1959.
- 5. Review: M. Somei, *Heterocycles*, 1999, **50**, 1157.
- M. Somei, K. Yamada, M. Hasegawa, M. Tabata, Y. Nagahama, H. Morikawa, and F. Yamada, Heterocycles, 1996, 43, 1855; M. Somei, K. Noguchi, R. Yamagami, Y. Kawada, K. Yamada, and F. Yamada, ibid., 2000, 53, 7.
- 7. a) M. Somei, A. Tanimoto, H. Orita, F. Yamada, T. Ohta, *Heterocycles*, 2001, **54**, 425. The yield of **8** from **2b** was improved to 72%. b) F. Yamada, K. Yamada, H. Takeda, and M. Somei, *ibid.*, 2001, **55**, 2361.
- 8. K. Yamada, T. Kawasaki, Y. Fujita, and M. Somei, Heterocycles, 2001, 55, 1151.
- 9. A. Inada, Y. Nakamura, and Y. Morita, *Chemistry Lett.*, 1980, 1287; M. Somei and Y. Saida, *Heterocycles*, 1985, 23, 3113; M. Somei, Y. Saida, T. Funamoto, and T. Ohta, *Chem. Pharm. Bull.*, 1987, 35, 3146.
- 10. S. Nakatsuka, O. Asano, K. Ueda, and T. Goto, Heterocycles, 1987, 26, 1471.
- 11. T. Hino, Y. Torisawa, and M. Nakagawa, *Chem. Pharm. Bull.*, 1982, **30**, 2349; M. Tani, T. Aoki, S. Ito, S. Matsumoto, M. Hideshima, K. Fukushima, R. Nozawa, T. Maeda, M. Tashiro, Y. Yokoyama, and Y. Murakami, *ibid.*, 1990, **38**, 3261 and references cited therein.

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