

# Effect of eutrophication on the distribution of arsenic species in eutrophic and mesotrophic lakes

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1 **Effect of Eutrophication on the Distribution of Arsenic**  
2 **Species in Eutrophic and Mesotrophic Lakes**

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26 **Keywords:** Arsenic speciation, As(V+III), DMAA, Hidden arsenic, Eutrophication, Lake  
27 waters.

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29

30 **Abstract:**

31 Effects of eutrophication on arsenic speciation were studied in eutrophic Lake Kiba  
32 and mesotrophic Lake Biwa, Japan. By combining hydride generation atomic absorption  
33 spectrometry with ultraviolet irradiation, inorganic, methyl and ultraviolet-labile fractions of  
34 arsenic were determined. In both Lakes, inorganic species (As(V+III)) dominated over other  
35 forms of arsenic all the year round. Most of methylarsenic fraction was dimethylarsinic acid  
36 (DMAA), and the concentration of monomethylarsonic acid (MMAA) was below the  
37 detection limit. Measurements of size-fractionated arsenic concentrations in water column  
38 indicate that most of the DMAA was distributed in truly dissolved fraction (<10 kDa), while  
39 ultraviolet-labile fractions were distributed in particulate (>0.45  $\mu\text{m}$ ) and colloidal (10 kDa –  
40 0.45  $\mu\text{m}$ ) fractions. Arsenic speciation in eutrophic Lake Kiba fluctuated greatly by seasonal  
41 changes. The ultraviolet-labile fractions were observed with the increase of DMAA from  
42 May to October, and they disappeared with the decrease of DMAA in January. In  
43 mesotrophic Lake Biwa, the ultraviolet-labile fractions of arsenic were not influenced as  
44 much as those in eutrophic Lake Kiba. On the other hand DMAA concentration was higher in  
45 Lake Biwa compared to that in Lake Kiba. The results suggest that the biosynthesis of  
46 complex organoarsenicals was enhanced by eutrophication, and the arsenic speciation would  
47 be influenced by the balance of biological processes in natural waters.

48

49

## 50 Introduction

51 Arsenic exists in a variety of chemical forms in natural waters and sediments.  
52 Arsenate, ( $\text{AsO}(\text{OH})_3$ ; As(V)) is the thermodynamically stable state in oxic waters, while  
53 arsenite ( $\text{As}(\text{OH})_3$ ; As(III)) is predominant in reduced redox potential conditions ([Andreae](#)  
54 [1986](#); [Cullen and Reimer 1989](#)). Biological processes also reduce the oxidation state of  
55 arsenic in surface waters ([Andreae 1986](#); [Cullen and Reimer 1989](#)).

56 The metabolism of arsenic by aquatic organisms results in the occurrence of  
57 thermodynamically unstable arsenite and methylarsenic compounds in natural waters. The  
58 inorganic forms (As(V) and As(III)) and the methylated forms (methylarsonic acid  
59  $\text{CH}_3\text{AsO}(\text{OH})_2$ ; MMAA(V) and dimethylarsinic acid  $(\text{CH}_3)_2\text{AsO}(\text{OH})$ ; DMAA(V)) are the  
60 main species of arsenic in natural waters ([Cullen and Reimer 1989](#)). The bulk of the total  
61 dissolved arsenic is inorganic species in seawater ([Peterson and Carpenter 1983](#)) and in  
62 freshwater ([Seyler and Martin 1989](#); [Kuhn and Sigg 1993](#)). Although the predominant form  
63 of methylarsenicals is consistently DMAA(V) followed by MMAA(V), the existence of  
64 methylarsenic(III) species in the environment has also been reported in literatures ([Sohrin et](#)  
65 [al. 1997](#); [Hasegawa et al. 1994, 1996](#)).

66 Previously, the speciation of arsenic in natural waters was determined by hydride  
67 generation followed by atomic absorption spectrophotometry ([Braman et al. 1977](#); [Andreae](#)  
68 [1977](#)). Arsenosugars and arsenobetaine can not be detected by the conventional hydride  
69 generation technique. [Howard and Comber \(1989\)](#) discovered and defined hidden arsenic in  
70 coastal water, which had not been detected previously by hydride generation atomic  
71 absorption spectrometry. [Hasegawa et al. \(1999\)](#) also reported the presence of  
72 organoarsenicals other than methylarsenicals in natural waters. [Hasegawa et al. \(1999\)](#)  
73 classified hidden arsenic into different fractions based on their photochemical degradation

74 ability. This hidden arsenic in natural waters is predicted to be related to the arsenic  
75 speciation and biological production in organisms.

76 In natural waters, the cycling of arsenic species would depend on the bioactivity of  
77 organisms (Cullen and Reimer 1989; Sanders 1980). Microorganisms produce  
78 methylarsenicals (MMAA and DMAA) in natural waters (Sanders and Riedel 1993), which  
79 exhibit seasonal cycle with maximum concentrations of methylarsenicals in summer (Sohrin  
80 et al. 1997; Hasegawa et al. 1999; Howard et al. 1995). Methylarsenicals was supposed to be  
81 produced by phytoplankton and organisms of higher trophic levels as a detoxification  
82 mechanism (Edmonds and Francesconi 1987). Recent studies propose accidental occurrences  
83 of methylarsenicals in nature (Please inset references here). Sanders and Riedel (1993)  
84 observed correlation between As(III)/ methylarsenicals and Chlorophyll-*a* concentrations  
85 and/or phytoplankton density. Howard et al. (1995) reported that the seasonal change in  
86 DMAA concentration is correlated with temperature but not with Chlorophyll-*a*  
87 concentrations and/or phytoplankton density. The bulk of other organoarsenicals are also  
88 found in organisms (Maeda 1994). The arsenosugars are usually found in algae and  
89 arsenobetaine is the predominant form in marine animals (Edmonds and Francesconi 1987;  
90 Francesconi and Kuehnelt 2002). The degradation and mineralization of organoarsenic  
91 compounds are supposed to be mostly depended on bacterial activities, which influence the  
92 arsenic cycling in aquatic environment (Kaise et al. 1985; Maki et al. 2005).

93 Eutrophication is a process whereby water bodies receive excess nutrients that  
94 stimulate excessive growth of phytoplankton, periphyton attached algae, nuisance plants and  
95 weeds. Eutrophication enhances not only the growth of phytoplankton but also the bacterial  
96 activities in the water column. On the other hand, mesotrophic lakes are lakes with an  
97 intermediate level of productivity, greater than oligotrophic lakes, but less  
98 than eutrophic lakes. These lakes are commonly clear water lakes with beds of submerged

99 aquatic plants and medium levels of nutrients. In the present experiment, we studied the  
100 distribution and speciation of arsenic in eutrophic and mesotrophic lakes. Determination of  
101 arsenic species, including the hidden arsenic, was performed by hydride generation atomic  
102 adsorption spectrometry using ultraviolet irradiation. The changes of hidden arsenic fractions  
103 in the water column were also studied to determine the influence of biological activity in  
104 arsenic speciation. Finally, the effects of eutrophication on arsenic speciation and distribution  
105 in natural waters have been discussed.

106

## 107 **Experimental**

### 108 **Sample collection and pretreatment**

109 Field investigations were carried out from May, 2006 to January, 2007 in Lake Kiba  
110 and Lake Biwa, Japan. Lake Biwa is the largest Lake in Japan with a surface area of 616 km<sup>2</sup>,  
111 and an average depth of 44 m (Sohrin et al. 1997). The northern basin of the lake is located  
112 near rural area, and is thought to be mesotrophic because of higher density of phytoplankton  
113 (2500 cells/ml in 1993) at the center of the basin (DCPLB 1995). On the other hand, the  
114 surface area of Lake Kiba is about 1.26 km<sup>2</sup> with an average depth of 2.2 m, and located in  
115 Hokuriku area, Japan. The concentrations of Chl-*a* in Lake Kiba is higher than that in Lake  
116 Biwa, and the dissolve oxygen (DO) concentration in the Lake is comparatively less than that  
117 in Lake Biwa. Based on phytoplankton density, Chl-*a* and nutrient concentrations in the  
118 water column, Lake Kiba and Lake Biwa is classified as eutrophic and mesotrophic,  
119 respectively.

120 The samples were collected within 0.5 m of the water surface. For analysis of arsenic  
121 and nutrients, the samples were filtered with 0.45 μm (HA type, Millipore) and 10 kDa  
122 (Minitan-S, Millipore) filters immediately after collection. Both filtered and unfiltered

123 samples were acidified to pH 2.0 by the addition of 1.0 M hydrochloric acid (HCl), and  
124 stored in refrigerator until analysis.

125

## 126 **Reagents**

127 Stock solutions ( $10^{-2}$  M) for the determination of arsenic compounds were prepared  
128 by dissolving the corresponding sodium salts ((CH<sub>3</sub>)<sub>3</sub>AsO<sub>3</sub>Na<sub>2</sub> was prepared by Quick's  
129 method (Hasegawa et al. 1994), and NaAsO<sub>2</sub>, Na<sub>2</sub>HAsO<sub>4</sub> and (CH<sub>3</sub>)<sub>2</sub>AsO<sub>2</sub>Na were obtained  
130 from Nacalai Tesque, Japan) in 0.1 M sodium hydroxide. These stock solutions were  
131 standardized by using inductively coupled plasma atomic emission spectrometry (ICP-AES,  
132 Optima 3300XL, Perkin Elmer) after decomposition to As(V). They were diluted to the  
133 desired concentrations just before use. Sodium borohydride (Kanto Chemical, Japan) was  
134 used for hydride generation. A 3% (w/v) sodium borohydride solution, stabilized in  $10^{-2}$  M  
135 sodium hydroxide solution, was prepared daily. Other reagents were of analytical grade or  
136 better.

137

## 138 **Arsenic analysis**

### 139 **Inorganic and methylarsenicals**

140 Analysis of inorganic and methylarsenicals was performed by a modified technique of  
141 hydride generation method (CT-HG-AAS), using an apparatus and materials similar to those  
142 described in previous paper (Hasegawa et al. 1994). In this technique, arsenic species were  
143 reduced to their corresponding arsines with sodium borohydride, trapped in U-tube with  
144 liquid nitrogen, and sequentially evolved into a heated quartz T-tube mounted in the atomic  
145 absorption spectrometer. To measure As(V+III), MMAA and DMAA, 3 mL of 0.1 M EDTA  
146 and 3 mL of 5.0 M HCl were added to 30 mL of the sample in the reaction vessel. In arsenite  
147 determination, 5 mL of 0.5 M potassium hydrogen phthalate buffer solution was added to 30

148 mL of the sample with an initial pH of 4. The detection limits were 0.11 nM and 0.14 nM for  
 149 As(V+III) and MMAA, respectively (3 times the standard deviation of the blank), and the  
 150 precision of five replicate determinations were 2.1% for inorganic arsenic and 5.1% for  
 151 DMAA (a relative standard deviation) at 1.0 nM with a 30 ml sample size.

152

### 153 **Ultraviolet irradiation**

154 Ultraviolet photolytic decomposition was accomplished by 400 W high-pressure  
 155 mercury lamp (Sigemi, AHH-400S) in a 3-chamber reaction vessel constructed from quartz  
 156 (Hasegawa et al. 1999). Samples were acidified to pH 2.0 using 1.0 M HCl, and introduced  
 157 into the outer-chamber of the reaction vessel that was capped with natural rubber septum.  
 158 They were then irradiated with a 400 W high-pressure mercury lamp mounted in the center-  
 159 chamber. During irradiation, cooling water was circulated into the mid-chamber from a  
 160 constant temperature bath. Aliquots were taken at selected time intervals. Arsenic analysis of  
 161 the digests was performed with CT-HG-AAS as described above.

162

### 163 **Speciation of organoarsenic species**

164 Organoarsenic species can be classified into different fractions according to their  
 165 lability to the photochemical degradation; UV-As and UV-DMAA (Hasegawa et al. 1999).  
 166 The decomposition of UV-As and UV-DMAA to As(V) followed the steps as described  
 167 below-



170 , where UV-DMAA was stepwise decomposed to As(V) through DMAA(V) by ultraviolet  
 171 irradiation with a time. The UV-As was transformed to As(V) directly. A flow chart of the  
 172 calculation is shown in Fig. 1. The UV-As and UV-DMAA were estimated from the



173 concentration changes of As(V+III) and DMAA during the ultraviolet irradiation by a non-  
174 linear least-squares computation, respectively (Figs. 2, 3).

175

## 176 **Results and Discussions:**

### 177 **Photolysis of hidden arsenic species in natural waters**

178 In the present experiment, irradiation test was performed for a time period of 0-12 hrs  
179 (Figs. 2, 3). Water samples were collected from surface level (0 m depth) of Lake Kiba on  
180 May 25, 2006. The unfiltered and filtered samples of Lake Kiba initially contained both  
181 As(V+III) and DMAA. The concentrations of As(V+III) in unfiltered and filtered samples  
182 were  $5.7\pm 0.5$  and  $4.2\pm 0.2$  nM, respectively, whereas DMAA concentrations were  $1.9\pm 0.1$  and  
183  $2.0\pm 0.8$  nM, respectively. Immediately after irradiation, both As(V+III) and DMAA  
184 concentrations in the unfiltered samples rapidly increased and attained equilibrium in 3-4 h.  
185 On the other hand, there was no increase of As(V+III) in filtered samples after irradiation,  
186 though DMAA increased a little (Fig. 3). The concentration of MMAA in both unfiltered and  
187 filtered samples was bellowing the instrumental limit of detection ( $< 0.14$  nM).

188 Water samples were collected from Lake Biwa on July 31, 2006, and irradiated for a  
189 time period of 0-12 hrs. The initial concentrations of As(V+III) and DMAA in Lake Biwa  
190 were 10.1 and 3.5 nM in unfiltered samples, and 9.5 and 3.3 nM in filtered samples,  
191 respectively. The DMAA concentrations in both unfiltered and filtered samples increased  
192 immediately after irradiation and then decreased gradually. On the other hand, the  
193 concentrations of As(V+III) in both unfiltered and filtered samples remained almost  
194 unchanged with the increase of irradiation time.

195 Other samples collected from natural waters also showed similar changes in arsenic  
196 speciation during irradiation though they varied in their concentrations. Hasegawa et al.  
197 (1999) observed similar changes of arsenic speciation in estuarine waters. Determination of

198 hidden arsenic fraction in seawater after photolytic decomposition has been described by  
199 [Castro et al. \(2007\)](#). [Howard and Comber \(1989\)](#) showed that irradiation of coastal seawater  
200 from a short-arc mercury lamp gave a large increases in As(V), MMAA and DMAA  
201 concentrations at natural pH. Both unfiltered and filtered samples were acidified to pH 2.0  
202 during the irradiation by the addition of 1.0 M HCl solution. [Howard and Comber \(1989\)](#)  
203 reported no increase of DMAA in acidified samples (about 0.011 M HCl) during ultraviolet  
204 irradiation, though [Hasegawa et al. \(1999\)](#) observed an increase of DMAA concentrations  
205 during ultraviolet irradiation. From their observations, [Hasegawa et al. \(1999\)](#) proposed that  
206 the inhibition effect of ultraviolet irradiation on DMAA concentration in acidified sample (as  
207 reported by [Howard and Comber \(1989\)](#)) may depend on the wavelength of ultraviolet light,  
208 rather than the acidity. The possibility of adsorption of arsenic on the particles, and of  
209 complexation of arsenic by organic substances is very low. Because the samples were  
210 acidified using 1.0 M HCl, and therefore, no changes of arsenic concentrations were observed  
211 in the samples at this very low pH.

212 [Brockbank et al. \(1988\)](#) reported that methylarsenic was demethylated during  
213 irradiation by 254 nm ultraviolet light without a digestion reagent. [Hasegawa et al. \(1999\)](#)  
214 also observed that DMAA gradually decreased after 4 h of irradiation by the high pressure  
215 mercury light. In the present study, it was observed that the DMAA concentration in  
216 unfiltered samples was increased rapidly within 3-4 h of irradiation though the concentration  
217 decreased gradually with the increase of irradiation time. It would be due to the  
218 decomposition of DMAA into As(V).

219

## 220 **Fractional distribution of arsenic speciation in Lake Kiba**

221 Dissolved and particulate fractions of arsenic were estimated by filtration through  
222 0.45  $\mu\text{m}$  membrane filters. Tangential flow filtration system with 10 kDa pore size was used

223 to divide the dissolved fractions of arsenic species (<0.45  $\mu\text{m}$ ) into colloidal (10 kDa - 0.45  
224  $\mu\text{m}$ ) and truly dissolved (<10 kDa) fractions. Hidden arsenic was determined in filtered and  
225 unfiltered samples after irradiation with a 400 W high-pressure mercury lamp. Data of arsenic  
226 species distribution in Lake Kiba during May 25, 2006 to January 17, 2007 are summarized  
227 in [Fig. 4 and Table 1](#).

228 In July,  $4.74 \pm 0.13$  nM of As(III) was recorded in filtered samples (0.45  $\mu\text{m}$   
229 membrane filter) whereas the As(V) concentration was  $1.95 \pm 0.17$  nM. In May, the  
230 concentration of As(III) was also higher than that of As(V). During October 26, 2006 and  
231 January 17, 2007 the concentrations of As(III) were lower than those of As(V) ([Table 1](#)).

232 In all seasons, total concentrations of arsenic were higher in the unfiltered samples  
233 compared to those in filtered samples, and the inorganic species of arsenic (As(V+III))  
234 dominate over DMAA and other arsenic fractions ([Fig. 4](#)). The concentrations of inorganic  
235 species (As(V+III)) were higher in the unfiltered samples compared to those in the filtered  
236 samples, except in July. The results imply that As(V) and As(III) were distributed in both the  
237 dissolved and particulate fractions. On the other hand, DMAA concentrations were  
238 comparable between the filtered and the unfiltered samples all the year round, which indicate  
239 that most of the DMAA consists of the truly dissolved fraction. The other methylarsenicals  
240 were totally absent or bellow the instrumental limit of detection. The UV-As and UV-DMAA  
241 in the filtered samples were lower than those in the unfiltered samples ([Fig. 4](#)). The results  
242 indicate that the UV-As and UV-DMAA were distributed mainly in the particulate fraction,  
243 and partially in the truly dissolved fraction.

244

#### 245 **Seasonal variations of arsenic species in Lake Kiba**

246 Arsenic species in water column of Lake Kiba fluctuated greatly during May 2006 to  
247 January 2007 by the seasonal variation. From May to October, 0.9-2.1 nM of DMAA was

248 detected though MMAA was less than 0.14 nM (Fig. 4). The high concentrations of DMAA  
249 might be due to the conversion of inorganic arsenic into organoarsenic compounds by aquatic  
250 organisms. Some organisms such as fungi and plankton uptake inorganic arsenic, and excrete  
251 DMAA in freshwater (Cullen and Reimer 1989; Maeda 1994; Hasegawa et al. 2001). A  
252 substantial amount of hidden arsenic, UV-As and UV-DMAA were also detected in the water  
253 column during May to October (Fig. 4). On the other hand, about 90% of the total arsenic  
254 was As(V+III) in January. The amount of UV-As was about 2-10%. The UV-As(V+III), UV-  
255 DMAA and other organoarsenicals were totally absent or below the instrumental limit of  
256 detection (Fig. 4).

257 In the present study, it was observed that the concentrations of DMAA and UV-  
258 degradable fractions of arsenic in water column of Lake Kiba were higher during summer  
259 (May to October) compared to those in winter (January), when the ratios of  
260 phosphate/As(V+III) were low. During summer, phosphate concentrations ranged between  
261 0.05-0.10  $\mu\text{M}$ , which were much lower than those in winter (0.31  $\mu\text{M}$ ) (Table 2). The  
262 behavior of DMAA has been reported to be consistent in other geographical areas (Anderson  
263 and Bruland 1991), and in laboratory experiments (Hasegawa et al. 2001). Howard et al.  
264 (1995) reported that DMAA is more frequently observed at higher water temperatures than  
265 As(III). Sohrin et al. (1997) reported that the DMAA concentrations were well correlated  
266 with water temperature in Lake Biwa, Japan whereas As(III) was not. Hasegawa (1996) also  
267 reported that the concentrations of DMAA follow the rise of water temperature in estuarine  
268 waters. Bright et al. (1994) demonstrated that microbes from anaerobic lake sediments  
269 methylate arsenic, and DMAA is produced by bacterial methylation in the suspended solids,  
270 which is subsequently released into the water column. Thus, bacterial methylation is  
271 responsible for the increase of DMAA concentration in surface water. Higher concentrations  
272 of DMAA might also be resulted from the long-term accumulation of DMAA excreted by

273 phytoplankton. However, Hasegawa (1996) reported that the concentrations of DMAA and  
274 chlorophyll-*a* in estuarine waters were not correlated significantly ( $r = 0.181$ ,  $n = 205$ ).

275 Our results also indicate that UV-As and UV-DMAA concentrations increased when  
276 the phosphate conditions were low (Fig. 4 and Table 2). The result suggests that the  
277 production of UV-As and UV-DMAA might increase with the increase of biological  
278 activities in proportion to the water temperature.

279

### 280 **Comparison of Lake Kiba and Lake Biwa:**

281 Concentration ranges of arsenic species, nitrate, phosphate, ammonium and  
282 chlorophyll-*a* in surface waters of Lake Kiba and Lake Biwa are presented in Table 3. The  
283 concentrations of chlorophyll-*a* have been considered as crude indicator of the net primary  
284 productivity, which is influenced by both phytoplankton productivity and grazing pressure in  
285 lake waters. The concentrations of nitrate, phosphate and ammonium in Lake Kiba were  
286 higher than those in Lake Biwa (Table 3). On the basis of chlorophyll-*a* and nutrient  
287 concentrations in water column, Lake Kiba is classified as eutrophic, and Lake Biwa as  
288 mesotrophic.

289 The concentrations of DMAA and MMAA in Lake Biwa were much higher than  
290 those in Lake Kiba. The DMAA concentrations ranged between <0.11-2.5 nM and 1.0-4.5  
291 nM in Lake Kiba and Lake Biwa, respectively. On the other hand, MMAA concentration in  
292 Lake Kiba was below the instrumental limit of detection though its concentrations ranged  
293 between 0.14-0.50 nM in Lake Biwa. The concentrations of UV-As and UV-DMAA were  
294 higher in Lake Kiba than those in Lake Biwa (Fig. 5). In Lake Kiba, most of the UV-As and  
295 the UV-DMAA were in particulate (>0.45  $\mu\text{m}$ ) and colloidal (10 kDa – 0.45  $\mu\text{m}$ ) fractions,  
296 while As(V+III) and DMAA were distributed mainly in truly dissolved fraction (<10 kDa).

297

**298 Degradation of DMAA by microorganisms:**

299 Water samples were collected from Lake Kiba and Lake Biwa on July 31 and 29,  
300 2006, respectively, and spiked with 1000 nM DMAA. The samples were then incubated for  
301 80 days under dark conditions and observed the changes of arsenic species. In the spiked  
302 samples of Lake Kiba, the DMAA was quantitatively degraded to As(V+III) within 45 days  
303 of incubation. The production of MMAA was below 0.14 nM during the experiment. On the  
304 other hand, it took about 80 days for the complete degradation of DMAA in water samples of  
305 Lake Biwa. The rates of DMAA degradation were 12.6 and 22.3 nM/day in Lake Biwa and  
306 Lake Kiba, respectively. Thus, degradation of DMAA would be related to the trophic  
307 condition of the lakes and the DMAA degradation would be higher in eutrophic lakes  
308 compared to that in mesotrophic lakes. Microbial degradation (mostly by bacterial activity) of  
309 DMAA into inorganic arsenic species was reported by [Maki et al. 2006](#). Methylarsenic  
310 species in the lake water was assumed to be converted into inorganic arsenic species by some  
311 anaerobic microbial reactions. The degradation of organoarsenic compounds is also predicted  
312 to be mostly depended on bacterial activities, which influence the arsenic cycles in the  
313 aquatic system ([Kaise et al. 1985](#); [Maki et al. 2005](#)). Thus, eutrophication would play an  
314 important role in the degradation of DMAA in lake waters.

315

**316 Effects of eutrophication on arsenic speciation:**

317 In freshwater systems, the proportions of arsenic species vary with the scope of  
318 anthropogenic input and biological activity. In this study, we investigated the distribution of  
319 arsenic species in mesotrophic and eutrophic lakes in relation to the biological activity. A  
320 number of freshwater organisms have been reported to contain organoarsenic compounds.  
321 [Hasegawa et al. \(2001\)](#) reported the direct production of methylarsenicals in several strains of  
322 phytoplankton. As(V) is biotransformed to organoarsenic compounds in freshwater food

323 chains (Maeda 1994). The decrease of total arsenic concentration and relative increase of  
324 methylarsenicals with the trophic level augmentation is observed in most of the food chains  
325 (Kuehnelt and Goessler 2003). The recent development of HPLC-ICP-MS for the  
326 determination of arsenic species has revealed the constituents and the behavior of arsenic,  
327 including complex organoarsenic compounds such as arsenosugars and arsenobetaine in  
328 freshwater organisms (Schaeffer et al. 2006).

329 In freshwater organisms, methylarsenic species, especially DMAA is the major  
330 organoarsenic compounds (Francesconi and Kuehnelt 2002). The increase of DMAA in water  
331 column of eutrophic Lake Kiba was observed from May to October in the present study (Fig.  
332 4). Similar trend of DMAA distribution in relation to the seasonal variations was reported in  
333 other lakes (Sandars and Riedel 1993). The source of DMAA could be the direct production  
334 of phytoplankton, or the decomposition of organic matter containing complex organoarsenic  
335 compounds by microorganisms or sunlight. Anderson and Bruland (1991) denied the direct  
336 phytoplankton excretion of DMAA because of the lack of correlation between chlorophyll-*a*  
337 and DMAA in the field studies. The photochemical degradation by sunlight does not  
338 contribute to the production of DMAA in lake waters, which suggests the degradation of  
339 complex organoarsenic compounds by microbial activity would be the possible reason for  
340 DMAA production (Hasegawa et al., 1999).

341 The concentrations of UV-As and UV-DMAA was correlated with that of DMAA in  
342 Lake Kiba. The UV-As, UV-DMAA and DMAA appeared from May to October though they  
343 disappeared in January (Fig. 4). The production of UV-As and UV-DMAA would be related  
344 to the biological activity as the similar manner of DMAA. Most of the UV-As and UV-  
345 DMAA is supposed to be derived from organic matter as the concentrations of As(V+III) and  
346 DMAA did not increase in the acidified unfiltered samples. The decrease of UV-As and UV-  
347 DMAA concentrations in filtered and ultra-filtered samples suggests that these fractions of

348 arsenic species were distributed in the colloidal and particulate fractions, which comprise  
349 organic and inorganic matter. [Bright et al. \(1996\)](#) also suggested that the hidden arsenic  
350 species or complex organoarsenic compounds such as arsenosugars might be absorbed tightly  
351 to organic matter. The degradation behavior of UV-As and UV-DMAA during the ultraviolet  
352 irradiation implies that the structures of UV-DMAA have DMAA fragments. Although  
353 DMAA and As(V+III) could be released from the particles of organic matter by ultraviolet  
354 irradiation, the UV-As and UV-DMAA fractions would mainly consist of complex  
355 organoarsenic compounds that were synthesized in phytoplankton and other freshwater  
356 organisms ([Kuehnelt and Goessler 2003](#)). [Koch et al. \(1999\)](#) reported the presence of  
357 arsenoriboses in microbial mats and green algae. Oxo- and thio-arsenosugars have been  
358 identified in several freshwater mussels and fishes as an important arsenic constituent, and  
359 arsenobetaine as a minor concentration ([Schaeffer et al. 2006](#); [Schmeisser et al. 2004](#)).

360         The concentrations of UV-As and UV-DMAA in eutrophic Lake Kiba were higher  
361 than those in mesotrophic Lake Biwa, and were correlated with DMAA concentration ([Fig. 4](#)).  
362 It can be elucidated by the eutrophication condition of the lakes. The eutrophication increased  
363 the microbial biomass and biosynthesis of complex organoarsenic compounds in the entire  
364 reservoir, which results in the degradation of DMAA and other organoarsenic compounds.  
365 Moreover, the degradation rate of DMAA was higher in eutrophic Lake Kiba than that in  
366 mesotrophic Lake Biwa. The result suggests direct transformation of As(V) into  
367 methylarsenicals or other organoarsenic compounds by biota, which in turn is degraded to  
368 DMAA and arsenate. The composition of arsenic species in Lake Kiba and Lake Biwa,  
369 shown in [Fig. 5](#), would attribute to the balance of biological processes; the metabolism of  
370 phytoplankton, grazing pressure due to zooplankton, and the decomposition of organic matter  
371 by microbial communities.



372 Total concentration of arsenic in Lake Kiba was higher in summer compared to winter  
373 (Fig. 4). This might be due to the release of As(V+III) from the anoxic sediments. The  
374 increase of As(V+III) concentration was also correlated with the water temperature. Because,  
375 reductive dissolution of iron and manganese oxides decrease the adsorptive surfaces of  
376 sediment particles and release As(V+III) (Crececius 1975). The dissolved arsenite  
377 (thermodynamically stable species in anoxic waters) is not as strongly adsorbed as arsenate  
378 (Leckie et al. 1984). The increase of arsenic concentration strongly correlates with the  
379 reductive dissolution of iron and manganese oxides (Crececius 1975).

380 Compared to filtered samples, higher concentrations of As(V+III) in unfiltered  
381 samples of Lake Kiba suggests that the dissolved fractions of arsenate were transformed into  
382 particulate fractions by adsorption or coprecipitation on iron and manganese oxides.  
383 Takamatsu et al. (1985) also reported the adsorption of arsenate on manganese oxides in  
384 manganese-rich lakes. The DMAA is converted almost completely to arsenate after winter  
385 mixing, and removed from the water column to the sediments (Anderson and Bruland 1991).  
386 This was because arsenate is particle reactive.

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Table 1: Distribution of arsenic species in filtered water of Lake Kiba. Water samples were collected from the surface level during May 2006 to January 2007, and filtered through 0.45  $\mu\text{m}$  membrane filters.

Sampling time	Concentrations (nM)					
	Arsenate	Arsenite	MMAA	DMAA	UV-As	UV-DMAA
May 25, 2006	1.41 $\pm$ 0.12	1.99 $\pm$ 0.20	<sup>§</sup> N.D.	1.72 $\pm$ 0.14	0.96 $\pm$ 0.12	0.94 $\pm$ 0.10
Jul. 31, 2006	1.95 $\pm$ 0.17	4.74 $\pm$ 0.13	<sup>§</sup> N.D.	0.95 $\pm$ 0.23	0.40 $\pm$ 0.11	1.20 $\pm$ 0.08
Oct. 26, 2006	2.83 $\pm$ 0.32	2.42 $\pm$ 0.17	<sup>§</sup> N.D.	2.08 $\pm$ 0.15	0.60 $\pm$ 0.09	2.10 $\pm$ 0.13
Jan. 17, 2007	2.14 $\pm$ 0.04	1.51 $\pm$ 0.08	<sup>§</sup> N.D.	<sup>§</sup> N.D.	0.20 $\pm$ 0.04	<sup>§</sup> N.D.

<sup>§</sup>N.D. “Not detected” because the concentrations were bellow the instrumental limit of detection.

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Table 2: Water temperature, nitrate and phosphate concentrations in Lake Kiba from May 2006 to Jan. 2007

Elements	May 25, 2006	Jul. 31, 2006	Oct. 26, 2006	Jan. 17, 2007
Water temperature (°C)	19.20	27.5	18.20	6.90
Chl. a ( $\mu\text{gL}^{-1}$ )	47.70	33.00	50.00	7.50
$\text{NO}_3\text{-N}$ ( $\mu\text{M}$ )	1.08	1.34	3.03	14.27
$\text{NO}_2\text{-N}$ ( $\mu\text{M}$ )	0.02	0.02	0.13	0.50
$\text{NH}_4\text{-N}$ ( $\mu\text{M}$ )	0.40	1.17	2.91	8.04
$\text{PO}_4\text{-P}$ ( $\mu\text{M}$ )	<sup>§</sup> N.D.	0.05	0.10	0.31
DOC ( $\mu\text{gL}^{-1}$ )	3.02	3.22	3.21	1.65

<sup>§</sup>N.D. “Not detected” because the concentrations were bellow the instrumental limit of detection.

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533 Table 3: Comparison in elemental concentration ranges between eutrophic Lake Kiba and

534 mesotrophic Lake Biwa.

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Elements	Lake Kiba*	Lake Biwa**
As(V) (nM)	2.3-7.6	2.0-6.0
As(III) (nM)	§ N.D. -4.1	1.0-3.0
MMAA (nM)	§ N.D.	§ N.D.-0.5
DMAA (nM)	§ N.D. -2.5	1.0-4.5
NO <sub>3</sub> -N (µM)	0.4-57	<0.7-14
NH <sub>4</sub> -N (µM)	0.3-19	<0.7-1.4
PO <sub>4</sub> -P (µM)	0.1-0.3	<0.032
Chl- <i>a</i> (µg/L)	3.0-89	0.8-10

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537 § N.D. “Not detected” because the concentrations were bellow the instrumental limit  
538 of detection.

539 \* Concentration range during the period of Apr. 2004 to Mar. 2006.

540 \*\* Concentration range during the period of Feb. 1993 to Dec. 1994.

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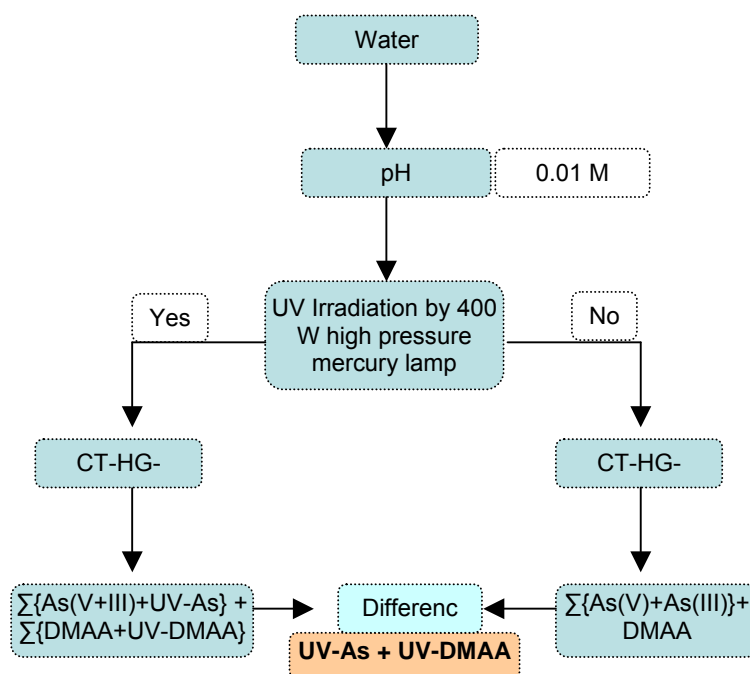
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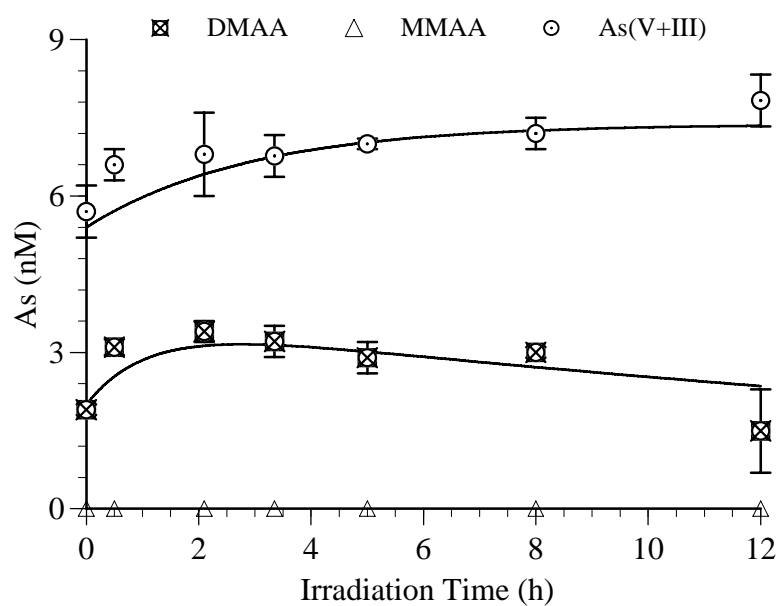
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552 Fig. 1: Schematic diagram showing the protocol for the determination of ultraviolet-labile  
553 fractions (UV-As and UV-DMAA) of arsenic in Lake water following UV-Irradiation.

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568 Fig. 2: Effect of irradiation time with a 400 W high-pressure mercury lamp on arsenic  
569 speciation in unfiltered water sample. Samples were collected from surface level of  
570 Lake Kiba on May 25, 2006. Error bars represent mean  $\pm$  S.D.

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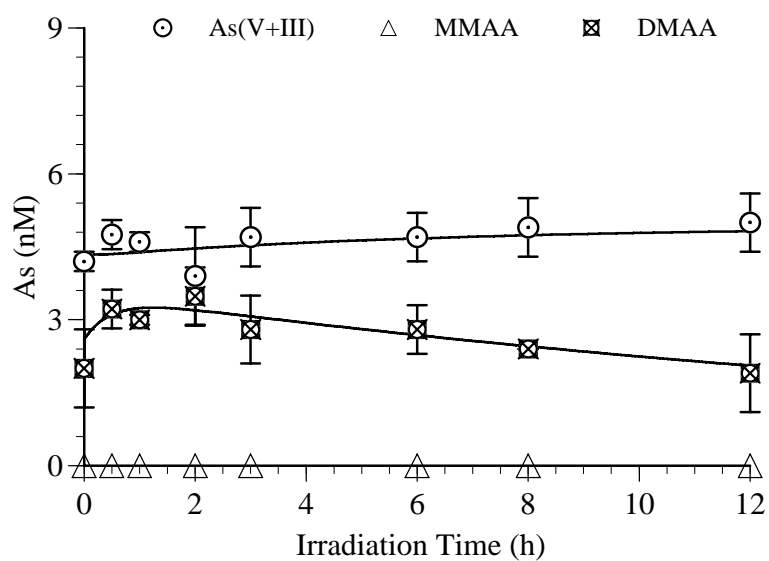
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585 Fig. 3: Effect of irradiation time with a 400 W high-pressure mercury lamp on arsenic  
586 speciation in filtered water sample. Samples were collected from surface level of Lake  
587 Kiba on 25 May, 2006. Error bars represent mean  $\pm$  S.D.

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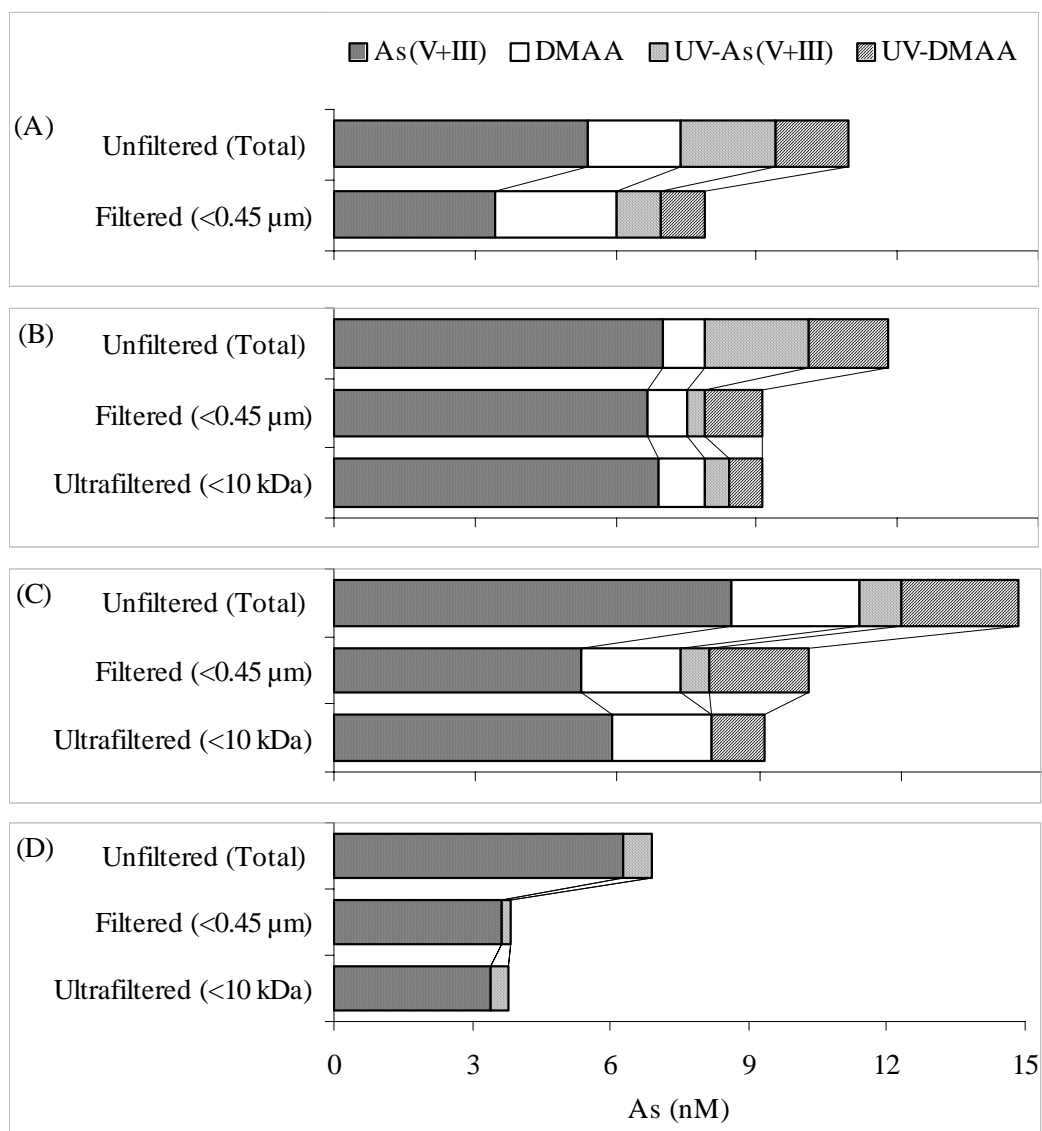
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602 Fig. 4: Concentration and distribution of arsenic species in surface water (depth 0 m) of Lake

603 Kiba collected from 25 May, 2006 (A), 31 Jul., 2006 (B), 26 Oct., 2006 (C) and 17

604 Jan., 2007 (D).

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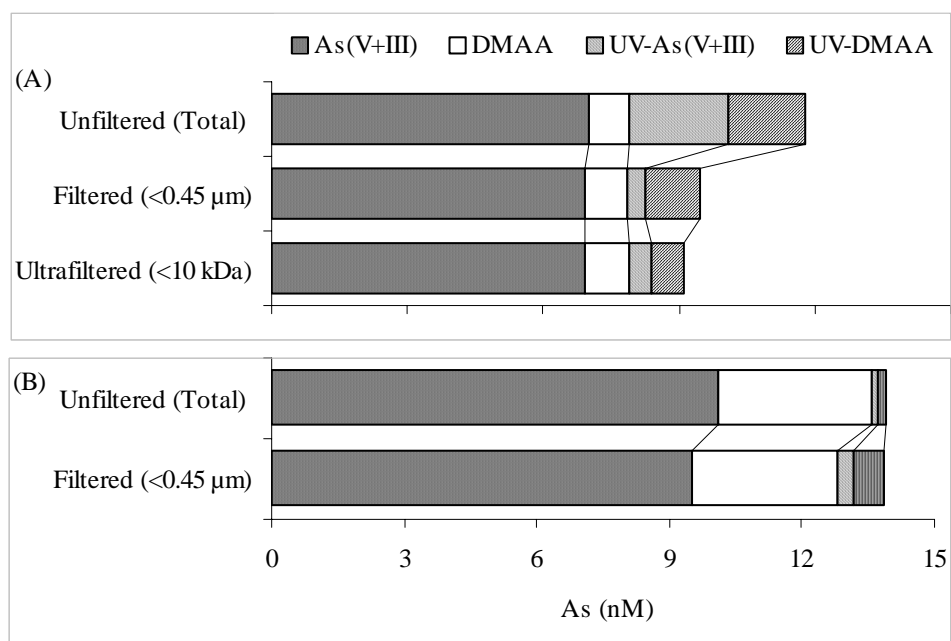
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612 Fig. 5: Mean concentration and distribution of arsenic in surface water (depth 0 m) of Lake

613 Kiba (A), and Lake Biwa (B) collected on 31 Jul., 2006 and 29 Jul., 2006,

614 respectively.