Electrohydrodynamic atomization of Balangu (*Lallemantia royleana*) seed gum for the fast-release of *Mentha longifolia* L. essential oil: characterization of nano-capsules and modeling the kinetics of release

4

5

Hassan Rezaeinia¹, Behrouz Ghorani¹, Bahareh Emadzadeh¹& Nick Tucker²

1 Department of Food Nanotechnology, Research Institute of Food Science and Technology (RIFST), Km 12
 Mashhad-Quchan Highway, Mashhad, Iran, P.O.Box : 91895-157-356

2 University of Lincoln, School of Engineering, Brayford Pool, Lincoln, LN6 7TS, United Kingdom (UK)

9 10

8

11 Abstract

The aim of this study is to optimize encapsulation of Mentha longifolia L. essential oil into 12 13 Balangu (Lallemantia royleana) seed gum nano-capsules, to increase their utility as flavoring 14 and bioactive agents in foods and beverages. Essential oil emulsions with Balangu seed gum 15 (0.25 and 0.5% w/w) and various polyvinyl alcohol (PVA) concentrations (0.5, 1 and 2%) 16 combined with Tween-20 (0.06, 0.08 and 0.1%) were electrosprayed. Increasing the 17 concentration of PVA increased the emulsion viscosity and improved both loading capacity 18 (77.56 to 84.68%) and encapsulation efficiency (81.54 to 87.82%) of the essential oil within 19 the structure of the Balangu gum nano-capsules. Field emission scanning electron 20 microscopy (FESEM) indicated that by increasing the amount of the gum (from 0.25 to 0.5%) 21 and PVA (from 1 to 2%), the process could be made to produce nanofibers. The Mentha 22 longifolia L. essential oil was entrapped in nanostructures without any chemical interaction 23 with encapsulant material, this was demonstrated by Fourier transform infrared spectroscopy 24 and differential scanning calorimetry. The release mechanisms and kinetics of loaded Mentha 25 longifolia L. essential oil were evaluated in different simulated food models (aqueous, acidic, 26 alcoholic or alkalic and oily food models) and release profiles data were fitted to first order, 27 Kopcha, Korsmeyer-Peppas, and Peppas-Sahlin models. The essential oil release profiles fitted 28 well to the Peppas-Sahlin model for a range of simulated foods. The release mechanism of the 29 essential oil from the nanostructure of the Balangu seed gum is mainly controlled by the Fickian 30 diffusion phenomenon.

Keywords: Electrohydrodynamic atomization, Balangu seed gum, nano-capsule, *Mentha longifolia* L, kinetics of release, fast release.

34 **1. Introduction**

35 The term "nutraceutical" is a portmanteau of nutrition and pharmaceutical and refers to foods 36 containing bioactive compounds found which in addition to the nutritional characteristics are 37 claimed to improve human health by means of biochemical properties such as antioxidant 38 activity and radical scavenging (Zlotogorski, et al., 2013b), with various effects being claimed 39 such as anti-cancer properties (Zlotogorski, et al., 2013a), and improvement to oral diseases 40 (McClements & Xiao, 2017). There are several compounds in both natural and processed foods 41 for which nutraceutical properties are claimed including carotenoids, flavonoids, 42 curcuminoids, phytosterols and certain fatty acids (Gupta, 2016).

43 Mentha longifolia L. is a medicinal and aromatic herb which belongs to Lamiaceae family 44 (Mahmoudi, 2014). The essential oil of *Mentha longifolia* L. is obtained from various parts of 45 the plant and has many applications in the food, pharmaceutical, and hygiene industries. Since 46 ancient times, the leaves, flowers, and stems of this plant have been used to prepare herbal teas 47 and dairy products (Gulluce, et al., 2007; Mahmoudi, 2014). The essential oil of Mentha 48 *longifolia* L. has both good flavor and odor, and consequently is used as a flavoring and aroma 49 agent in various food products. The principal components of the essential oils have been 50 identified in previous studies, and include pulegone, carvone, limonene, 1,8-cineole, menthone 51 and piperitenone oxide (Gulluce, et al., 2007; Mahmoudi, 2014; Mkaddem, et al., 2009). The 52 essential oil has been demonstrated to show antioxidant activity, antimicrobial activity against 53 a wide range of microorganisms, and therapeutic properties: and has hence been considered as 54 an additive in beverages, confectionery, chewing gum, and dairy products (Dhifi, Litaiem, 55 Jelali, Hamdi, & Mnif, 2011; Golestan, Seyedyousefi, Kaboosi, & Safari, 2016).

56 There are various limiting factors for the application of nutraceuticals in foods and 57 subsequently, functional food production. These factors include a low water solubility index 58 (Gleeson, Ryan, & Brayden, 2016; Murugesan & Orsat, 2012), and chemical and biochemical 59 instability when temperature, pH, are varied, in addition to vulnerability to enzyme attack (L. Chen, Remondetto, & Subirade, 2006). They may also have undesirable effects on food flavors 60 61 and textures, coupled with poor-bioavailability (McClements, 2015; McClements, Decker, 62 Park, & Weiss, 2009). It is therefore necessary to design treatments to overcome these 63 constraints to enable the use of nutraceuticals in food systems. Nanoencapsulation is one of the 64 more successful protection methods; the bioactive compounds are entrapped within a nanoscale 65 protective shell (Bhushani, Kurrey, & Anandharamakrishnan, 2017). Nano-capsules have a higher surface to volume ratio compared to larger encapsulating structures, thus having higher 66 67 solubility, improved encapsulation efficiency, more bioavailability and a better controlled 68 release of the entrapped components (Aditya, Espinosa, & Norton, 2017; Pereira, et al., 2018; 69 Prakash, et al., 2018). The encapsulation process of bioactive compounds is carried by various methods such as dispersion and freeze drying (H. Chen & Zhong, 2015), emulsifying (Y. Chen, 70 71 et al., 2018), spray drying (Otálora, Carriazo, Iturriaga, Nazareno, & Osorio, 2015) and 72 coacervation (Joaquín Gomez-Estaca, Comunian, Montero, Ferro-Furtado, & Favaro-73 Trindade, 2016). Damage to the payload constituents contained within the structure of the 74 capsules can occur when conventional encapsulation methods (spray drying, freeze drying) are 75 used and this may be accompanied by untimely and incomplete release. These limitations are 76 not commonly observed when electrohydrodynamic (EHD) processing (i.e. electrospinning 77 and electrospraying) is used (Alehosseini, Ghorani, Sarabi-Jamab, & Tucker, 2017; Jahangiri, 78 et al., 2014; Peltonen, Valo, Kolakovic, Laaksonen, & Hirvonen, 2010).

EHD is a recent approach for nanoencapsulation of bioactive and nutraceutical compounds which has found application in the food and pharmaceutical industries (J Gomez-Estaca, Balaguer, Gavara, & Hernandez-Munoz, 2012). The process involves pumping the feed solution through a fine nozzle or spinneret and spraying it from the spinneret using an electric field for motive power. The nanofibers or nano-capsules are collected on the nearest earthed 84 surface. If a nanofiber is formed, the process is referred to as electrospinning, and if the process leads to nano-capsule production, it is called electrohydrodynamic atomization (EHDA) or 85 electrospraying (Ghorani & Tucker, 2015). The process is carried out at ambient temperature 86 87 (Deng, Kang, Liu, Feng, & Zhang, 2017) and results in the production of both very fast, and 88 burst release systems (Bock, Dargaville, & Woodruff, 2014). Recently, the electrospraying 89 technique for nano-capsule formation has been used for the stabilization of food bioactive 90 components such as lycopene (Rocio Pérez-Masiá, Lagaron, & Lopez-Rubio, 2015), β-91 carotene (Gómez-Mascaraque, Perez-Masiá, González-Barrio, Periago, & López-Rubio, 92 2017)D-limonene (Khoshakhlagh, Koocheki, Mohebbi, & Allafchian, 2017) and green tea 93 catechins (Bhushani, et al., 2017).

94 The choice of an appropriate encapsulation material is a critical issue in the process. A wide 95 range of synthetic food grade polymers and biopolymers are used in the electrospraying of bioactive and nutraceutical compounds, viable microorganisms (Librán, Castro, & Lagaron, 96 97 2017) and enzymes (Yaghoobi, Majidi, ali Faramarzi, Baharifar, & Amani, 2017). However, 98 researchers are always looking for new materials - natural biopolymers are particularly popular, 99 as they are highly acceptable to consumers. Balangu (Lallemantia royleana) is a medicinal 100 plant grown in European and Middle East countries especially Iran, it produces quantities of a 101 viscous gummy material when its seeds are soaked in water (Najafi, Hosaini, Mohammadi-102 Sani, & Koocheki, 2016; Razavi, Cui, & Ding, 2016). Balangu seed gum is a flexible polymer 103 with a high molecular weight giving it the ability to form edible films with high thermal 104 stability, good oxygen, and moisture permeability, water solubility and thixotropic behavior; it 105 could, therefore, be considered as an appropriate encapsulant agent (Razavi, et al., 2016; 106 Sadeghi-Varkani, Emam-Djomeh, & Askari, 2018). However, to the best of our knowledge, 107 the development of electrosprayed capsules for Balangu seed gum has not been yet been 108 described.

109 Accordingly, in this study, we developed electrosprayed Balangu seed gum nano-capsules 110 containing Mentha longifolia L. essential oil. The feasibility of nano-capsule production and 111 the effects of varying the properties of the gum encapsulant, the active agent, the addition of 112 surfactants and the effect of these variations on morphological characteristics were evaluated. 113 The structural properties, loading capacity and the encapsulation efficiency of nano-capsules were also studied. In addition, the kinetics and the mechanisms of the release of essential oil 114 115 from the fabricated structures in various representative model food systems; namely aqueous, 116 alcohol based, acidic or alkali, and oily foodstuffs were modeled using first order, Kopcha, 117 Korsmeyer-Peppas, and Peppas-Sahlin empirical equations.

118

119 **2.** Materials and Methods

120 **2.1. Materials**

121 The *Mentha longifolia* L. essential oil was kindly provided by Exir Gol Sorkh Co., Ltd. (Iran).
122 Polyvinyl alcohol (PVA) (Mw = 77,000-79,000 Da, 98% hydrolyzed) and Hexane (HPLC
123 grade) were purchased from Sigma-Aldrich Company (USA). Tween 20 (HLB = 16.7), ethanol
124 and acetic acid were obtained from Merck (Germany). All chemicals were used without further
125 purification.

126 **2.2. Methods**

127 **2.2.1 Extraction of Balangu seed gum**

The extraction of Balangu seed gum was performed based on a modification of the method described by Razavi et al. (2016). Briefly, Balangu seeds were cleaned manually to remove all impurities. The seed gum was then hydrated: the seeds being soaked in distilled water with a seed to water mass ratio of 1:30 and the suspension placed in a water bath at a constant temperature of 85°C for 150 minutes. The mixture was poured slowly and at a constant rate into the extractor (Pars Khazar, JC-700P Juicer, Iran). The extraction process was repeated twice for each batch. For the purification process, the extracted Balangu seed gum solution was
mixed with ethanol at a ratio of 1 to 4. Finally, the purified Balangu gum was dried in a freeze
dryer.

137 **2.2.2. Solution preparation**

138 To prepare the sample solutions for the electrospraying process, gum solutions (0.25 and 0.5% 139 (w/v)) were hydrated overnight at 4°C. Tween-20 polysorbate-type nonionic surfactant (at 0.06, 0.08 and 0.1% based on gum weight) was added to the gum solutions to improve 140 141 sprayability. To improve the nano-capsule production efficiency, PVA was also added at levels 142 of 0.5 and 1% (w/v); higher levels of PVA led to nanofiber formation which was not the aim 143 of this research. Therefore, two levels of PVA (0.5 and 1%) were used in the final emulsions. Finally, Mentha longifolia L. essential oil (0.015 g based on gum weight) was inserted in the 144 145 emulsion systems as the oil phase, and as the bioactive and flavoring compound. To prepare 146 the oil-in-water emulsions, the coarse emulsions were first prepared by mixing with a 147 magnetic stirrer at 300rpm for 10 min. The coarse emulsions were then homogenized using an 148 Ultra-Turrax homogenizer (model T-25, IKA Instruments, Germany) at a speed of 13000rpm 149 for 3 min in an iced water bath.

150 **2.2.3. Electrospraying process**

151 All stages of the electrospraying process were performed under constant conditions. For this 152 purpose, 10ml of each emulsion was drawn into a plastic syringe connected to a blunt-ended 153 Luer Lock metal syringe needle (Gauge-21, nominal outer diameter 0.8192mm, and nominal inner diameter 0.514mm, Sigma-Aldrich). The syringe was then mounted into a triple-head 154 155 syringe pump that was connected to a high voltage power supply (ES-Lab RN/X, ANSTCO, 156 Iran). The process conditions were fixed at 1mL/h pump flow rate, 25kV spinning voltage, a 157 needle tip-to-collector distance of 150mm, in an environment of 25 ± 3 °C and a relative 158 humidity of $22\pm 2\%$.

159 **2.2.4. Surface tension**

A tensiometer (Krüss® K100 tensiometer, Germany) was used to determine the surface tension at 20°C of each sample based on the Wilhelmy plate method. The instrument was calibrated using distilled water (71.64mN/m). The results were obtained from three replicates and the average data was reported (Zaeim, Sarabi-Jamab, Ghorani, Kadkhodaee, & Tromp, 2018).

164 **2.2.5. Emulsion viscosity**

165 The flow behavior parameters were determined using a Brookfield viscometer (Brookfield 166 DVIII Ultra, Brookfield Engineering Laboratories, Stoughton, MA, USA) equipped with 167 anSC4-27 spindle at 25°C. Flow curves were acquired at shear rates of $1-82s^{-1}$. All 168 measurements were performed in triplicate. The shear stress and shear rate data were analyzed 169 using SlideWrite Plus Graphics Software (version 7.01, USA). The flow behavior data from 170 the samples was approximated to a power law model (Eq. 1).

171
$$\sigma = k \cdot \gamma^n$$
 Eq. 1

172 Where σ , *k*, γ , and n are shear stress (Pa), consistency coefficient (Pa.sⁿ), shear rate (s⁻¹) and 173 flow behavior index (dimensionless), respectively.

174

175 **2.2.6. Morphology of nano-capsules**

176 The morphology of the electrosprayed Balangu seed gum nano-capsules was observed using 177 Field emission scanning electron microscopy (FESEM), MIRA3, using a 178 TESCAN, Czech Republic). The Balangu seed gum nano-capsules were 179 coated with gold using a sputter coater (Q150R Rotary-Pumped Sputter Coater, Quorum Technologies Ltd., UK) for 150s at 20mA and the micrographs were observed at an 180 181 accelerating potential of 15kV. One hundred nano-capsules prepared from optimal treatment 182 were selected as a sample to determine capsule size using Image-Pro Plus software (Version183 7.3).

184 **2.2.7. Photography of electrospray jet modes**

A high-speed digital camera (DFK 22BUC03, The Imaging Source Company, Germany)
equipped with a zoom lens was used to determine the jet mode formed at the tip of the needle.
The pictures were recorded as the electrospraying process became stable (H.-H. Kim, Kim, &
Ogata, 2011).

189 **2.2.8. Fourier-transform infrared (FTIR)**

The FTIR spectra were used to analyze the interaction between Balangu seed gum, PVA and the *Mentha longifolia* L. essential oil in the nano-capsules. Electrosprayed samples were mixed with KBr and pressed into pellets. The FTIR spectra were recorded in the wave number range 4000-400 cm⁻¹ using an FTIR spectrometer (Bruker Alpha FTIR, US) (Khoshakhlagh, *et al.*, 2017).

195

196 **2.2.9. Thermal analysis**

197 The thermal properties of pure PVA and Balangu seed gum, free *Mentha longifolia* L. essential 198 oil and nano-capsules PVA/gum/essential oil were tested by Differential Scanning Calorimetry 199 (DSC, Mettler Toledo, Switzerland). Nominal 15mg samples were placed in an aluminum pan 200 and heated from 0 to 400°C at a 10°C/min heating rate under a nitrogen atmosphere with a flow 201 rate of 30 mL/min (Santos, *et al.*, 2014).

202

203 **2.2.10.** Loading capacity and encapsulation efficiency

204 The encapsulation efficiency of the essential oil in Balangu nano-capsules was calculated using

205 the method described by (Wang, et al., 2017). 5mg of the nano-capsules were first washed with

1ml of distillated water to remove the surface essential oil, and then dissolved in 1ml of 50% ethanol aqueous solution for 24h. The concentration of the essential oil was measured by recording the absorbance at 281nm using a spectrophotometer (UNICO-2100, USA). The encapsulation efficiency (EE) of the essential oil in Balangu nano-capsules was determined using the following equation (Eq. 2):

211

$$EE\% = (W_1 \div W_2) \times 100$$
 Eq. 2

Where W_1 and W_2 are the weight of essential oil in a certain weight of nano-capsules and weight of the essential oil in the feed solution, respectively.

Also, the loading capacity (LC) of Balangu nano-capsules was calculated by Eq. 3
(Khoshakhlagh, *et al.*, 2017):

216
$$LC\% = (W_1 - W_3 / W_4) \times 100$$
 Eq. 3

Where W_3 is the amount of free essential oil for a certain weight of nano-capsules and W_4 is the weight of nano-capsules.

219

220 **2.2.11. Release kinetics of the essential oil in food models**

To explain the *Mentha longifolia* L. essential oil release profile from Balangu seed gum nanocapsules, the release kinetics in food models were fitted to first-order, Kopcha, Korsmeyer-Peppas and Peppas-Sahlin empirical models. The release kinetics of the essential oil were simulated in aqueous (distilled water), acidic (3% acetic acid), alcoholic or alkali (10% ethanol) and oily (50% ethanol) food models according to the EU Commission regulation 10/2011 EU (10/2011/EC) (Atay, et al., 2018).

227 The first order model (Eq. 4), which is a mathematical model describes the release of the loaded

compounds from porous structures (Costa & Lobo, 2001).

230

$\ln M_t = \ln M_0 - K_1 \times t$ Eq. 4

Where M_t is the amount of essential oil released at time t, M_0 is the amount of essential oil released at time 0 and K_1 is release constant of the first order model (Desai, Singh, Simonelli, & Higuchi, 1966).

The Kopcha model (Eq. 5) is based on the release of bioactive components from a deliverysystem by diffusion or erosion mechanisms.

236 $Mt = A \times t^{0.5} + B \times t$ Eq. 5

Where M_t is the amount of essential oil released at time t, A is the diffusion rate constant and B is the erosion rate constant. The A/B ratio is used to predict the dominant mechanism in the release of essential oil from the delivery system structure. If the diffusion is the dominant mechanism, the ratio of A/B would be >1 and if the release rate is governed by erosion, the ratio would be <1. If both mechanisms are involved in the release of essential oil, A/B = 1 (Kopcha, Lordi, & Tojo, 1991).

In the cases that the main mechanism of release is a combination of Fickian (diffusion) and non-Fickian transfer, the Korsmeyere-Peppas equation (Eq. 6) would be the best simple semiempirical model for explaining the release profile (Mehrgan & Mortazavi, 2010).

- 246
- 247 $M_t/M_{\infty} \times 100 = kt^n \qquad \qquad \text{Eq. 6}$

Where M_t and M_{∞} are the amount of essential oil released at time t and the initial mass of essential oil loaded in the nano-capsules, K is the release kinetic constant and n is the release exponent. The release mechanism is determined by the n value. For spherically shaped capsules, values n \leq 0.43 indicate a diffusion (Fickian) mechanism, values 0.43<n<0.85 represent non-Fickian diffusion and values $n \ge 0.85$ up to 1 indicate erosion mechanism of the delivery system(Lee, *et al.*, 2006).

The Peppas–Sahlin model (Eq. 7) is used to evaluate the mechanism of Fickian and non-Fickian
transfer from the structure of delivery systems.

256
$$M_t/M_{\infty} = k_1 t^m + k_2 t^{2m}$$
 Eq. 7

Where k_1 is the diffusion rate (Fickian) constant, k_2 is the erosion rate constant and m is the purely Fickian diffusion exponent for a system of any configuration which exhibits controlled release. If the ratio of $k_1/k_2>1$, the bioactive profile release is described mostly by diffusion and if $k_1/k_2<1$, the loaded component release is described mostly by erosion. Also, if the ratio of $k_1/k_2=1$, both Fickian diffusion and erosion mechanisms are involved (Peppas & Sahlin, 1989).

262 **2.2.12. Statistical analysis**

Data were analyzed using a one-way analysis of variance (ANOVA) and a Duncan's Multiple Range test for a statistical significance $P \le 0.05$, using the IBM SPSS statistics software (version 22.0, IBM Corp., USA). All experiments were performed in duplicate, and data were presented as mean \pm standard deviation (SD) values.

267

268 **3. Results and discussions**

269 **3.1. Surface tension**

Surface tension is one of the factors that affect the electrospraying process and the morphological characteristics of the nano-capsules produced (Deng, *et al.*, 2017). Aqueous solutions with high surface tension require some modifications to the electrospraying process or feed formulation (Okutan, Terzi, & Altay, 2014). In terms of process variables, the applied voltage can be increased to overcome increased surface tension in the feed and to eject the polymer solution towards the collector, but the level of voltage is ultimately limited due breakdown of the insulation of the device (K. Kim, Kim, & Shim, 2017). Therefore, it may be
appropriate to use surfactants in the feed formulation that will reduce surface tension and hence
the need for elevated voltages (Stephansen, García-Díaz, Jessen, Chronakis, & Nielsen, 2016).
In this study, Tween-20 was used as an emulsifier giving the side benefit of reducing the surface
tension of pure Balangu seed gum in solution.

281 The surface tension values of the pure Balangu seed gum and the various Oil/Water (O/W) 282 emulsions containing Mentha longifolia L. essential oil, Tween-20 and various PVA 283 concentrations are shown in Table 1. By increasing the level of Balangu seed gum to 0.5%, the 284 surface tension of pure distilled water (71.64mN/m) dropped to 60.77mN/m. This reduction of 285 surface tension as gum concentration is increased is probably due to the reduction of the amount 286 of water per unit volume. Also, the addition of Tween-20 (0.06 to 0.1%) in the PVA free 287 samples reduced the surface tension in both gum ratios:0.25% (39.36 to 38.18mN/m) and 0.5% 288 (35.02 to 31.70mN/m). Emulsifiers, due to their amphiphilic property are rapidly absorbed to 289 the air/solution interface and reduce the surface or interfacial tension of the solution (Rocío 290 Pérez-Masiá, Lagaron, & López-Rubio, 2014). Increasing the PVA concentration (from 0.5 to 291 2%) at constant Balangu gum and Tween-20 values resulted in significantly lower surface 292 tensions (Table.1). Okutan et al. (2014) reported that the surface tension of the polymer solution 293 significantly decreased from 36.24 to 34.91mN/m when polymer concentration was increased 294 from 7 to 20%.

295

Table 1. The surface tension of pure Balangu seed gum and various Oil/Water emulsions.

297

3.2. Morphology of electrosprayed nano-capsules

EHD is one of the most commonly used methods in transforming a wide range of biopolymersand synthetic polymers into nano-fiber and -capsule forms. Different morphologies can be

301 obtained by EHD processing of polymer solutions depending on the process parameters, 302 solution properties, and environmental conditions. In process-based changes, the spinning is 303 influenced by the applied voltage, volume feed rate, distance from the needle to collector and 304 needle diameter (Okutan, et al., 2014; Rocío Pérez-Masiá, et al., 2014). The solution property-305 based approaches include polymer concentration, the ratio of polymer(s) in solution, solvent 306 type, polymeric solution viscosity, surface tension, electrical conductivity, and presence of an 307 emulsifier. Furthermore, environmental conditions such as temperature and relative humidity 308 influence the ability to spin (Alehosseini, et al., 2017; Ghorani, Alehosseini, & Tucker, 2017; 309 Stijnman, Bodnar, & Tromp, 2011).

Capsules are easier to handle and disperse compared to equivalent nanofibers, and so are
preferred for food applications (Gómez-Mascaraque, Lagarón, & López-Rubio, 2015).
Therefore, the production of Balangu nano-capsules was optimized to obtain individual
capsules without the co-production of fibers.

314 Initially, 0.25 and 0.5% pure Balangu gum solutions were electrosprayed. The results showed 315 that the pure gum did not have enough sprayability and no droplets were produced due to the high surface tension of the solutions (Fig. 1) (Liu, et al., 2017). As can be seen in Fig. 1, with 316 317 a high surface tension solution (Table 1), the jet does not form at the tip of the needle and large 318 pendant drops fall from the spinneret. When the surface tension is very high the applied electric 319 field is insufficiently strong to produce a spray or fiber from the solution (Alehosseini, *et al.*, 320 2017). Tween-20 was added at 0.06, 0.08 and 0.1% (based on gum weight) as an emulsifier to reduce the surface tension (Table 1), and initiate spraying. 321

As shown in Fig. 2, increasing the emulsifier level from 0.06 to 0.08% at both 0.25 and 0.5% gum concentrations (Fig. 2A, 2B, 2D, and 2E), enhanced the production of nano-capsules, but increasing the amount of emulsifier from 0.08 to 0.1% reduced the density of nano-capsules on the collector (Fig. 2C and 2F). The high surface tension of aqueous solutions prevents the formation of Taylor cone and subsequently, reduces the sprayability of the polymers (Zhao, Sun, Shao, & Xu, 2016). Also, the hydrophilic nature and poor surface activity of Balangu gum are probably the other important factors in reducing the sprayability of this gum, which justifies the use of emulsifiers. Based on these results, Tween-20 was used at a level of 0.08% for further study. To improve the rate of nano-capsules production, PVA was used at 0.5, 1 and 2% as an adjunct spinning polymer.

332 Fig.3A-F shows the results of the effect of variation of the gum (0.25 and 0.5%) and PVA (0.5, 333 1 and 2%) concentration on the morphological properties of Balangu seed gum nano-capsules. 334 In samples containing 0.25% gum, by increasing the PVA concentration from 1 to 2%, the 335 process made nanofibers instead of nano-capsules (Fig. 3B to 3C) this is due to the reduction 336 in repulsive forces in the charged polymeric solution (Eatemadi et al., 2016). Also, the use of 337 0.5% gum with various levels of PVA (0.5% to 2%) (Fig. 3D, 3E and 3F) to preferentially form 338 nanofibers is probably due to the high viscosity of the 0.5% gum solution indicating a high 339 level of molecular scale entanglement in the polymer chains (Koski, Yim, & Shivkumar, 340 2004).

341 Spraying or spinning gums faces two major limitations. First, most of the gums produce a high 342 viscosity in low concentrations, and second, they provide strong shear thinning properties and 343 as a very high shear force is applied to the polymeric solution at the tip of the needle, it may 344 make spinning problematic (Stijnman, et al., 2011) (Okutan, et al., 2014). Therefore, the critical 345 overlap concentration and shear thinning properties are the determinant factors in a successful 346 EHD process. From examination of the results shown in Fig. 3, the nano-capsules prepared 347 from 0.25% Balangu seed gum emulsion, 1% PVA, 0.08% Tween-20, and Mentha longifolia 348 L. essential oil were selected as the optimal treatment (Fig. 4) and subsequent experiments

349	were performed using this formulation, to efficiently produce nano-capsules. The optimal
350	treatment gave an average capsule size of 96.53±3.41 nm.
351 352 353 354 355 356 357 358 359	 Fig. 1. The surface tension of pure Balangu seed gum at the tip of the needle. Fig. 2. FESEM images of Balangu seed gum (0.25 and 0.5%) nano-capsules with different levels of Tween-20 (0.06, 0.08 and 0.1% based on gum weight): A) 0.25 gum with 0.06% Tween-20, B) 0.25 gum with 0.08% Tween-20, C) 0.25 gum with 0.1% Tween-20, D) 0.5 gum with 0.06% Tween-20, E) 0.5 gum with 0.08% Tween-20 and F) 0.5 gum with 0.1% Tween-20.
360 361 362 363	Fig. 3 . FESEM images of Balangu seed gum (0.25 and 0.5%) nano-capsules with different levels of PVA (0.5, 1 and 2%): A) 0.25 gum with 0.5% PVA, B) 0.25 gum with 1% PVA, C) 0.25 gum with 2% PVA, D) 0.5 gum with 0.5% PVA, E) 0.5 gum with 1% PVA and F) 0.5 gum with 2% PVA.
364 365 366 367 368 369	Fig. 4. FESEM images of Balangu seed gum nano-capsules under the optimal conditions (0.25% Balangu seed gum, 1% PVA, 0.08% Tween-20 and the <i>Mentha longifolia</i> L. essential oil).
370	3.3. Viscosity
371	The flow behavior parameters and the curve of apparent viscosity versus shear rate of the pure
372	gum solutions and the emulsions based on gum and gum with PVA are shown in Table 2 and
373	Fig. 4, respectively. The analysis of the rheograms showed shear thinning (pseudoplastic)
374	behavior for all the solutions. The power law model was the better-fitting model in than
375	Herschel-Bulkley ($R^2 > 0.99$) and the flow behavior index was below 0.608. Most biopolymers
376	do show a pseudoplastic behavior. Ma, Du, Yang, & Wang (2017) have also shown that the
377	blended film-forming solution of Tara gum and PVA exhibited a shear-thinning behavior at
378	0.1 to 100 1/s shear rates.
379	Fig. 5 shows the viscosity-shear rate changes for different solution/emulsion samples.

Fig. 5 shows the viscosity-shear rate changes for different solution/emulsion samples. Increasing the Balangu seed gum concentration from 0.25 to 0.5% in the samples resulted in higher viscosity values (or consistency coefficient (*k*), Table 2). Since the gums will provide a high viscosity at low concentrations, increasing the concentration of Balangu seed gum from 0.25 to 0.5% dramatically increases apparent viscosity. As a result, due to high viscosity and
high surface tension in the pure gum, especially at a concentration of 0.5% pure gum, there
was no possibility of electrospraying.

Also, increasing the amount of Tween-20 up to 0.08% increased the viscosity and the *k* value of the solutions, but its higher amount (0.08 to 0.1%) significantly (p<0.05) decreased the viscosity of the solutions (Table 2). Although increasing the emulsifier up to 0.08% increased the viscosity, surface tension (high surface tension is one of the limiting factors in the electrospraying process) decreased, so the 0.08% concentration of the Tween-20 was selected as the optimum value.

392 Surfactants can produce hydrophobic and electrostatic interactions, and promote hydrogen 393 bonding. Therefore, they increase the interaction between polymer chains and result in higher 394 viscosities. However, higher concentrations of surfactants have a modulating role and may 395 therefore reduce the viscosity of the solution (Kriegel, Kit, McClements, & Weiss, 2009).

As shown in Fig. 5, increasing the PVA level from 0.5 to 2% in O/W emulsions, improves the
viscosity of the feed solutions.

Increasing the PVA concentration will result in higher interactions and entanglements between the polymer and the biopolymer chains and thus increase the viscosity (Zhou, *et al.*, 2017). It is apparent that this phenomenon offsets the negative effects of the hydrocolloid

401 pseudoplasticity under the applied process. A change in morphology from asymmetric capsules
402 with low density to denser and more regular capsules occurred as the PVA concentration was
403 increased (Fig. 3 A-B). Fibers were obtained for the samples at a PVA concentration of 2%
404 (Fig 3 C-F), while the typical pseudo-spherical capsules of the electrospraying process, with a
405 few residual fibrils, were produced at 0.5 and 1% gum/PVA concentrations.

407 Table 2. Rheological parameters of pure Balangu seed gum and various O/W emulsions in the power law model.
409
410 Fig. 5. The apparent viscosity of Balangu seed gum/PVA O/W emulsions (the codes in the legend refer to the row numbers of Table 2).
412

413

414 **3.4. Evaluation of electrospray jet modes**

415 The mode of jet formation at the tip of the spinneret is influenced by the behavior and 416 characteristics of the polymeric solution and the process conditions. Under constant process 417 conditions (namely, voltage, distance and pump flow rate), only the feed properties affect the 418 jet mode (Enavati, Chang, Bragman, Edirisinghe, & Stride, 2011). The various jet modes 419 observed during this study are shown in Fig. 6. The best jet mode in the electrospray process 420 is the cone-jet mode. This is the most stable form of the jet in which the polymer solution will 421 spray well (Prajapati & Patel, 2010). The cone-jet mode was observed during the 422 electrospraying of the emulsion prepared from 0.25% Balangu seed gum emulsion, 1% PVA 423 and, 0.08% Tween-20 containing the Mentha longifolia L. essential oil. This mode is 424 influenced by the emulsion characteristics such as viscosity and surface tension. The cone-jet 425 mode was observed during the electrospraying of emulsions 0.25% Balangu seed gum, 426 0.5% PVA, 0.08% Tween-20 and the Mentha longifolia L. essential oil and 0.25% Balangu 427 seed gum, 2%PVA, 0.08% Tween-20 and the essential oil, however, cone jets formed 428 instantaneously and were unstable. During the electrospraying of these two samples, dripping 429 (dripping and micro-dripping) and the spindle formation (single and multi-spindle) modes were 430 also observed. This is probably due to high viscosity and also interactions between the highly 431 charged but tiny droplets (H.-H. Kim, et al., 2011) in the sample containing 2% PVA (Fig.6). 432 In emulsions of 0. 5% Balangu seed gum, 0.5% PVA, 0.08% Tween-20, 0.5% Balangu seed

433 gum, 1%PVA, 0.08% Tween-20 and 0.5% Balangu seed gum, 2%PVA, 0.08% Tween-20

434 containing the *Mentha longifolia* L. essential oil the process proceeded to produce nanofibers 435 (beaded fibers), multi-jet and precession modes were also observed. These modes are probably due to the increased viscosity and the reduced surface tension (leading conditions toward the 436 437 production of nanofibers) (Drosou, Krokida, & Biliaderis, 2017) that were observed in the 438 emulsions (Sections 3-1 and 3-3). In the pure Balangu seed gum solution (0.25% and 0.5%) 439 oscillating-jet mode was observed due to the high surface tension and viscosity (Table 1-2). 440 Additionally, in all PVA-free emulsions, intermittent precession and micro-dripping modes 441 were observed.

- 443 **Fig. 6.** Images of real and simulated needles with types of jet modes are formed at the tip of the needle
- 445

442

446 **3.5. FTIR analysis**

The FTIR analysis was used for studying possible interactions between Balangu seed gum, 447 448 PVA, essential oil and Tween-20 in the electrospray nano-capsules (Fig. 7). The spectrum of Balangu seed gum gave a very broad absorbance peak at 3406 cm⁻¹ that was related to the 449 450 stretching vibration of O-H groups, hydrogen bonds of Balangu gum molecules as well as the 451 existence of water molecules connected to the gum chains. The absorption bands at 2924, 1609, 1423, 1374,1315 and 1057cm⁻¹ were attributed to C-H, C-OO asymmetric stretching, C-OO 452 453 symmetric stretching, C-O and C-O-C stretching, respectively (Fig. 7) (Farhadi, 2017). 454 The spectrum of PVA, showed the specific bands of O-H, C-H stretching vibration, C=O, C-

- 455 H₂bending, C-O and C-C groups at 3388, 2939, 1739, 1441, 1377, 1265, 1093, 848 cm⁻¹,
- 456 respectively (Fig. 3) (Li, Kanjwal, Lin, & Chronakis, 2013).
- 457 The FTIR spectrum of Tween-20 is shown in Fig. 7. The broad peak around 3396 cm⁻¹ is
- 458 defined as H-bonded O-H stretching vibration. Absorptions at 2925 and 2869 cm⁻¹ indicate
- 459 the C–H alkane. The peaks at 1734 and 1641 are assigned to the C=O vibration and C=C

bending vibrations of the alkenes are present at 1458 and 1350 cm⁻¹(García-Benjume, EspitiaCabrera, & Contreras-García, 2009). A very sharp peak at 1107cm⁻¹ is attributed to the C-O
stretching vibration of many esters, ether, hydroxyl groups, and also the bending vibration of
C-C bonds(Khoshakhlagh, *et al.*, 2017).

464 Pure *Mentha longifolia* L. essential oil spectra show characteristic peaks at 2953 and 2925 (C465 H stretching), 1682 (N-H bending), 1455 (CH₂ bending), 1286 (C-O-C), 1130 (C-O-C
466 stretching) and 937 cm⁻¹ (C-H bending) (Fig. 7).

467 The FTIR spectrum of the Balangu seed gum (0.25% w/v) nano-capsules loaded by the *Mentha* 468 *longifolia* L. essential oil containing 0.5 and 1% (w/v) PVA is shown in Fig. 7. As the amount 469 of Tween-20 and the essential oil is lower than the Balangu gum and PVA concentrations in 470 the structure of nano-capsules, most of their small peaks were integrated or vanished entirely.

471 The wave number of nano-capsules containing 0.5% and 1% PVA is a little different (Fig. 7). 472 However, the waves of the samples prepared by 1% PVA are wider and higher than capsules 473 containing 0.5% PVA. Thus, Fig. 7shows that capsules containing 1% PVA have more 474 hydrogen bonds, probably due to the hydrogen bonds of hydroxyl groups and other functional 475 groups of capsule constituents. The peaks around 840-850 (C-C vibration) and 1100-1110 cm⁻ 476 ¹ (C-O vibration) were also strengthened in the structure of both capsules which express the 477 successful encapsulation of the Mentha longifolia L. essential oil within the electrosprayed 478 nano-capsules of Balangu gum and PVA. The overall observations indicate that there has been 479 no adverse reaction between the constituents of the nano-capsules and the loaded essential oil, 480 so the essential oil has been successfully trapped physically. Similarly, Khoshakhlagh et al. 481 (2017) reported that the D-limonene encapsulation in the structure of Alyssum homolocarpum 482 seed gum nano-capsules reinforces the peaks related to C-C and C-O vibration bonds.

483

Fig.7. FTIR spectra of 0.25% Balangu seed gum/0.5% PVA nano-capsules (0.5PVA); 0.25% Balangu seed gum/1% PVA nano-capsules (1PVA); pure Balangu seed gum (Gum); *Mentha longifolia* L.
essential oil (EO); pure PVA (PVA); and pure Tween-20 (Tween-20).

488

489 **3.6. Thermal analysis**

490 The thermal properties of different samples of pure PVA and Balangu seed gum, free Mentha 491 longifolia L. essential oil and nano-capsules PVA/gum/essential oil (the optimal nanocapsules) were analyzed by Differential Scanning Calorimetry (DSC). In the pure PVA 492 493 thermogram, an endothermic peak in the range of 187 to 211° C with a melting transition (T_m) 494 of 199°C can be seen (Fig. 8). The T_m represents the melting temperature in PVA. The DSC 495 thermogram of PVA also has an endothermic peak between 298.3 to 322.5°C (centered at 496 310.6°C) which is related to the complete decomposition of the sample (Santos, et al., 2014). 497 The pure Balangu seed gum DSC curve shows a glass transition temperature (T_g) at 95.5±5°C 498 and a relatively wide peak in about 291.5 to 345°C (centered at 315.6°C) which is assigned to 499 the decomposition temperature of the pure gum (Fig. 8). The thermal properties of the free 500 Mentha longifolia L. essential oil were also evaluated by DSC thermograms. The DSC curve 501 of the essential oil showed two endothermic peaks at 29.1 and 169.4°C which are related to the 502 evaporation temperature and the complete decomposition of the essential oil. These results 503 confirm the heat sensitivity and volatile nature of the Mentha longifolia L. essential oil.

504 DSC thermogram of the optimal nano-capsules shows only an endothermic peak around 505 223.5°C. As can be seen in this curve, the evaporation peak of the essential oil has disappeared 506 which approves the successful encapsulation of the essential oil in the complex structure. In 507 addition, the comparison of the pure samples thermogram with the complex thermogram does 508 not indicate any additional peak which is an indication of no interaction between Balangu seed 509 gum/PVA/*Mentha longifolia* L. essential oil (Khoshakhlagh, *et al.*, 2017).

510

Fig 8. DSC thermograms for Balangu seed gum (Gum), PVA, *Mentha longifolia* L. essential oil (EO) and optimum electrosprayed nano-capsule.

- 513
- 514

515 **3.7. Loading capacity and encapsulation efficiency**

The samples that resulted in nano-capsules were further evaluated for essential oil loading capacity and encapsulation efficiency tests (Fig. 3 A-B). Table 3 shows the encapsulation efficiency and loading capacity of *Mentha longifolia* L. essential oil in the Balangu seed gum-PVA electrosprayed nano-capsules. By increasing the PVA concentration from 0.5 to 1%, the encapsulation efficiency and loading capacity of *Mentha longifolia* L. essential oil in electrosprayed Balangu seed gum-PVA nano-capsules increases from 81.54 to 87.82% and 77.56 to 84.68%, respectively.

This behavior is probably related to the higher viscosity of the emulsion containing 0.25% Balangu, 1% PVA, and *Mentha longifolia* L. essential oil than the emulsion sample containing 0.25% Balangu, 0.5% PVA, and the essential oil (Table 2 and Figure 5). The higher viscosity in the emulsion system will result in an improved stability, and will increase the loading capacity and encapsulation efficiency (Yeo & Park, 2004).

528 PVA and Balangu gum form hydrogen-bonded water molecules through the hydroxyl groups 529 in their structure. This leads to the formation of a hydrated layer at the surface of the droplets 530 and subsequently, an increase in the encapsulation efficiency and loading capacity will occur 531 (Song, et al., 2008). Bhushani et al. (2017) investigated the efficiency of the electrospray 532 method for encapsulation of green tea catechins. They reported that the encapsulation 533 efficiency of zein nano-capsules ranged from 86.84 to 97.45 %. However, Khoshakhlagh et al. 534 (2017) reported loading capacity and encapsulation efficiency were between 9.21 to 20.13% 535 and 74.93 to 93.24%, respectively, for D-limonen in electrosprayed Alyssum homolocarpum

- seed gum nano-capsules. These differences are probably related to the emulsification method,encapsulant materials, and process conditions.
- 538

Table 3. Encapsulation efficiency and loading capacity of different electrosprayed Balangu seed
 gum/PVA nano-capsules

542

543 **3.8. Release kinetics of the essential oil in food models**

544 The nano-capsules prepared from 0.25% Balangu seed gum emulsion containing 1% PVA, 545 0.08% Tween-20, and the *Mentha longifolia* L. essential oil had the highest loading capacity 546 and encapsulation efficiency (Table 3). So, this sample was chosen as the best candidate for 547 examining essential oil release. The in vitro release kinetics of the Mentha longifolia L. 548 essential oil from an optimal sample is shown in Fig. 9. The amount of essential oil released at 549 different times was measured at 281 nm. As illustrated in Fig. 9, the Mentha longifolia L. 550 essential oil release profile was a function of the type of food model. In all the model food 551 models, the essential oil had an explosive and immediate release in the first 3 minutes. After 3 552 minutes, their release was continued gradually at a gentle gradient, for 60, 120, 180 and 180 553 minutes for distilled water, 10% ethanol, 50% ethanol and 3% acetic acid media, respectively. 554 As the results show, the highest release from the nano-capsules was obtained in distillated 555 water, 10% ethanol, 50% ethanol and 3% acetic acid media, respectively.

It is believed that surface erosion, disintegration, diffusion, and desorption are the mechanisms involved in the release of bioactive compounds and drug from nano-capsules and microcapsules (Hariharan, *et al.*, 2006). Therefore, to determine the mechanism of release of *Mentha longifolia* L. essential oil from Balangu seed gum nano-capsules, the release profile within various food model systems was fitted with different kinetic equations. In our study, first-order, Kopcha, Korsmeyer-Peppas and Peppas-Sahlin models were used to evaluate the release behavior of the essential oil. The constants and the coefficient of determination (\mathbb{R}^2) of each model are shown in Table 4. Concerning the \mathbb{R}^2 values, the first-order and the Kopcha models are not suitable for determining the release behavior of the essential oil from the nanocapsules structure. The Peppas-Sahlin model with an \mathbb{R}^2 over 0.9945 was chosen as the appropriate model for explaining the release kinetics of the *Mentha longifolia* L. essential oil.

In the Peppas-Sahlin model, K_1 and K_2 are diffusion and erosion constants, respectively. As the ratio of K_1 to K_2 was greater than 1 so the *Mentha longifolia* L. essential oil release was mostly governed by diffusion mechanism in all food models studied (Peppas, *et al.*, 1989). However, the power of the Korsmeyer-Peppas equation (n) was lower than 0.43 (between 0.09939 and 0.1163), which indicates a Fickian mechanism of release (Lee, *et al.*, 2006).

As shown in Table 4, in the Kopcha model, the ratio of A/B is greater than 1, which indicates the predominance of the diffusion phenomenon (Fickian behavior) in the release of the *Mentha longifolia* L. essential oil in all media. Also, the comparison of diffusion (k_1) and erosion constants (k_2) in the Peppas-Sahlin model (Table 4) shows that the release mechanism of the essential oil from the structure of nano-capsules of the Balangu seed gum is mainly governed by the Fickian diffusion phenomenon since the ratio k_1/k_2 is greater than 1.

Fig. 9 shows that the highest amount of the essential oil is released in distilled water media, followed by 10% ethanol and 50% ethanol media. The lowest release rate is observed in a medium containing 3% acetic acid. Probably since Balangu seed gum and PVA are both watersoluble and have high solubility in water, so Balangu seed gum/PVA nano-capsules have a high solubility and swelling degree in distilled water, which results in the more and faster release of the essential oil in the distilled water media. However, due to the insolubility of these two polymers in alcohol and acetic acid (note that the gum is sparingly soluble in acetic acid),

585	the swelling and solubility of the nano-capsules decrease in higher alcohol-containing and
586	acetic acid media, swelling and solubility being ultimately lower than in distilled water media.
587	
588 589	Table 4. Kinetics constant of the <i>Mentha longifolia</i> L. essential oil release profile in different food models
590	
591	
592 593	Fig.9 . The cumulative release profile of the <i>Mentha longifolia</i> L. essential oil in different aqueous food stimulants.
594	
595	
596	
597	

598 **4.** Conclusion

599 In this study, EHDA or electrospraying process of Balangu seed gum/PVA nano-capsules 600 loaded by Mentha longifolia L. essential oil was investigated. FESEM examination indicated that the emulsion containing 0.25% Balangu seed gum, 1% PVA, 0.08% Tween-20, and 601 Mentha longifolia L. essential oil could be considered as the optimal formulation for nano-602 603 capsule production. FTIR spectra and DSC indicated that no undesirable interactions were 604 occurred between Balangu seed gum/PVA and loaded Mentha longifolia L. essential oil. The 605 Peppas-Sahlin model was chosen as the best model for predicting the essential oil release 606 profile in simulated aqueous foods. Release kinetics of *Mentha longifolia* L. essential oil in 607 simulated media followed a Fickian diffusion mechanism. The results showed a burst release 608 of Mentha longifolia L. essential oil within the first 3 min, followed by sustained release for a 609 further 180min. The results of this study showed that Balangu seed gum could be considered 610 as a fruitful natural source for production of nano-capsules containing Mentha Longifolia L. 611 essential oil. Concerning the observed release mechanism, these nano-capsules would be a

- 612 good choice for fast- flavor release systems (the system is under study and development by the
- 613 authors).

- 615 5. References
- Aditya, N., Espinosa, Y. G., & Norton, I. T. (2017). Encapsulation systems for the delivery of
 hydrophilic nutraceuticals: Food application. *Biotechnology advances*, *35*(4), 450-457.
- Alehosseini, A., Ghorani, B., Sarabi-Jamab, M., & Tucker, N. (2017). Principles of
 electrospraying: A new approach in protection of bioactive compounds in foods. *Critical reviews in food science and nutrition*, 1-18.
- Atay, E., Fabra, M. J., Martínez-Sanz, M., Gomez-Mascaraque, L. G., Altan, A., & LopezRubio, A. (2018). Development and characterization of chitosan/gelatin electrosprayed
 microparticles as food grade delivery vehicles for anthocyanin extracts. *Food Hydrocolloids*, 77, 699-710.
- Bhushani, J. A., Kurrey, N. K., & Anandharamakrishnan, C. (2017). Nanoencapsulation of
 green tea catechins by electrospraying technique and its effect on controlled release and
 in-vitro permeability. *Journal of Food Engineering*, *199*, 82-92.
- Bock, N., Dargaville, T. R., & Woodruff, M. A. (2014). Controlling microencapsulation and
 release of micronized proteins using poly (ethylene glycol) and electrospraying.
 European Journal of Pharmaceutics and Biopharmaceutics, 87(2), 366-377.
- 631 Chen, H., & Zhong, Q. (2015). A novel method of preparing stable zein nanoparticle
 632 dispersions for encapsulation of peppermint oil. *Food Hydrocolloids*, 43, 593-602.
- Chen, L., Remondetto, G. E., & Subirade, M. (2006). Food protein-based materials as
 nutraceutical delivery systems. *Trends in Food Science & Technology*, 17(5), 272-283.
- Chen, Y., Shu, M., Yao, X., Wu, K., Zhang, K., He, Y., Nishinari, K., Phillips, G. O., Yao, X.,
 & Jiang, F. (2018). Effect of zein-based microencapsules on the release and oxidation
 of loaded limonene. *Food Hydrocolloids*.
- Costa, P., & Lobo, J. M. S. (2001). Modeling and comparison of dissolution profiles. *European journal of pharmaceutical sciences*, 13(2), 123-133.
- Deng, L., Kang, X., Liu, Y., Feng, F., & Zhang, H. (2017). Effects of surfactants on the
 formation of gelatin nanofibres for controlled release of curcumin. *Food chemistry*,
 231, 70-77.
- Desai, S. J., Singh, P., Simonelli, A. P., & Higuchi, W. I. (1966). Investigation of factors
 influencing release of solid drug dispersed in inert matrices ii quantitation of
 procedures. *Journal of pharmaceutical sciences*, 55(11), 1224-1229.
- 646 Dhifi, W., Litaiem, M., Jelali, N., Hamdi, N., & Mnif, W. (2011). Identification of a new
 647 chemotye of the plant Mentha aquatica grown in Tunisia: chemical composition,
 648 antioxidant and biological activities of its essential oil. *Journal of Essential Oil Bearing*649 *Plants*, 14(3), 320-328.
- Drosou, C. G., Krokida, M. K., & Biliaderis, C. G. (2017). Encapsulation of bioactive
 compounds through electrospinning/electrospraying and spray drying: A comparative
 assessment of food-related applications. *Drying Technology*, *35*(2), 139-162.
- Enayati, M., Chang, M.-W., Bragman, F., Edirisinghe, M., & Stride, E. (2011).
 Electrohydrodynamic preparation of particles, capsules and bubbles for biomedical engineering applications. *Colloids and Surfaces A: Physicochemical and Engineering Aspects, 382*(1-3), 154-164.

- Farhadi, N. (2017). Structural elucidation of a water-soluble polysaccharide isolated from
 Balangu shirazi (Lallemantia royleana) seeds. *Food Hydrocolloids*, 72, 263-270.
- García-Benjume, M., Espitia-Cabrera, M., & Contreras-García, M. (2009). Hierarchical macro mesoporous structures in the system TiO2–Al2O3, obtained by hydrothermal synthesis
 using Tween-20® as a directing agent. *Materials Characterization, 60*(12), 1482-1488.
- Ghorani, B., Alehosseini, A., & Tucker, N. (2017). Nano-capsule formation by electrospinning.
 In *Nanoencapsulation Technologies for the Food and Nutraceutical Industries* (pp. 264-319): Elsevier.
- 665 Ghorani, B., & Tucker, N. (2015). Fundamentals of electrospinning as a novel delivery vehicle 666 for bioactive compounds in food nanotechnology. *Food Hydrocolloids*, *51*, 227-240.
- Gleeson, J. P., Ryan, S. M., & Brayden, D. J. (2016). Oral delivery strategies for nutraceuticals:
 Delivery vehicles and absorption enhancers. *Trends in Food Science & Technology*, 53,
 90-101.
- Golestan, L., Seyedyousefi, L., Kaboosi, H., & Safari, H. (2016). Effect of M entha spicata L.
 and M entha aquatica L. essential oils on the microbiological properties of fermented
 dairy product, kashk. *International journal of food science & technology*, *51*(3), 581587.
- Gomez-Estaca, J., Balaguer, M., Gavara, R., & Hernandez-Munoz, P. (2012). Formation of
 zein nanoparticles by electrohydrodynamic atomization: Effect of the main processing
 variables and suitability for encapsulating the food coloring and active ingredient *curcumin. Food Hydrocolloids, 28*(1), 82-91.
- 678 Gomez-Estaca, J., Comunian, T., Montero, P., Ferro-Furtado, R., & Favaro-Trindade, C.
 679 (2016). Encapsulation of an astaxanthin-containing lipid extract from shrimp waste by
 680 complex coacervation using a novel gelatin–cashew gum complex. *Food*681 *Hydrocolloids*, 61, 155-162.
- Gómez-Mascaraque, L. G., Lagarón, J. M., & López-Rubio, A. (2015). Electrosprayed gelatin
 submicroparticles as edible carriers for the encapsulation of polyphenols of interest in
 functional foods. *Food Hydrocolloids*, 49, 42-52.
- Gómez-Mascaraque, L. G., Perez-Masiá, R., González-Barrio, R., Periago, M. J., & López Rubio, A. (2017). Potential of microencapsulation through emulsion-electrospraying to
 improve the bioaccesibility of β-carotene. *Food Hydrocolloids*, 73, 1-12.
- Gulluce, M., Sahin, F., Sokmen, M., Ozer, H., Daferera, D., Sokmen, A., Polissiou, M.,
 Adiguzel, A., & Ozkan, H. (2007). Antimicrobial and antioxidant properties of the
 essential oils and methanol extract from Mentha longifolia L. ssp. longifolia. *Food chemistry*, 103(4), 1449-1456.
- 692 Gupta, R. C. (2016). Nutraceuticals: efficacy, safety and toxicity: Academic Press.
- Hariharan, S., Bhardwaj, V., Bala, I., Sitterberg, J., Bakowsky, U., & Kumar, M. R. (2006).
 Design of estradiol loaded PLGA nanoparticulate formulations: a potential oral delivery
 system for hormone therapy. *Pharmaceutical research*, 23(1), 184-195.
- Jahangiri, A., Davaran, S., Fayyazi, B., Tanhaei, A., Payab, S., & Adibkia, K. (2014).
 Application of electrospraying as a one-step method for the fabrication of triamcinolone
 acetonide-PLGA nanofibers and nanobeads. *Colloids and Surfaces B: Biointerfaces, 123*, 219-224.
- Khoshakhlagh, K., Koocheki, A., Mohebbi, M., & Allafchian, A. (2017). Development and
 characterization of electrosprayed Alyssum homolocarpum seed gum nanoparticles for
 encapsulation of d-limonene. *Journal of colloid and interface science*, 490, 562-575.
- Kim, H.-H., Kim, J.-H., & Ogata, A. (2011). Time-resolved high-speed camera observation of
 electrospray. *Journal of Aerosol Science*, 42(4), 249-263.
- Kim, K., Kim, J., & Shim, H. (2017). Fiber formation model for PVP (polyvinyl pyrrolidone)
 electrospinning. I. Critical voltage. *Fibers and Polymers*, 18(3), 493-501.

- Kopcha, M., Lordi, N. G., & Tojo, K. J. (1991). Evaluation of release from selected
 thermosoftening vehicles. *Journal of pharmacy and pharmacology*, 43(6), 382-387.
- Koski, A., Yim, K., & Shivkumar, S. (2004). Effect of molecular weight on fibrous PVA
 produced by electrospinning. *Materials Letters*, 58(3-4), 493-497.
- Kriegel, C., Kit, K., McClements, D. J., & Weiss, J. (2009). Electrospinning of chitosan-poly
 (ethylene oxide) blend nanofibers in the presence of micellar surfactant solutions.
 Polymer, 50(1), 189-200.
- Lee, S., Kim, M., Kim, J., Park, H., Woo, J., Lee, B., & Hwang, S. J. (2006). Controlled delivery of a hydrophilic drug from a biodegradable microsphere system by supercritical anti-solvent precipitation technique. *Journal of microencapsulation*, 23(7), 741-749.
- Li, X., Kanjwal, M. A., Lin, L., & Chronakis, I. S. (2013). Electrospun polyvinyl-alcohol
 nanofibers as oral fast-dissolving delivery system of caffeine and riboflavin. *Colloids and Surfaces B: Biointerfaces, 103*, 182-188.
- Librán, C., Castro, S., & Lagaron, J. (2017). Encapsulation by electrospray coating atomization
 of probiotic strains. *Innovative food science & emerging technologies*, *39*, 216-222.
- Liu, X., Baldursdottir, S. G., Aho, J., Qu, H., Christensen, L. P., Rantanen, J., & Yang, M.
 (2017). Electrospinnability of poly lactic-co-glycolic acid (PLGA): the role of solvent
 type and solvent composition. *Pharmaceutical research*, *34*(4), 738-749.
- Ma, Q., Du, L., Yang, Y., & Wang, L. (2017). Rheology of film-forming solutions and physical
 properties of tara gum film reinforced with polyvinyl alcohol (PVA). Food
 Hydrocolloids, 63, 677-684.
- Mahmoudi, R. (2014). Effect of Mentha longifolia L. Essential oil on physicochemical
 properties of the bio-ayran. *Journal of Essential Oil Bearing Plants*, 17(1), 56-66.
- McClements, D. J. (2015). Enhancing nutraceutical bioavailability through food matrix design.
 Current Opinion in Food Science, *4*, 1-6.
- McClements, D. J., Decker, E. A., Park, Y., & Weiss, J. (2009). Structural design principles
 for delivery of bioactive components in nutraceuticals and functional foods. *Critical reviews in food science and nutrition*, 49(6), 577-606.
- McClements, D. J., & Xiao, H. (2017). Designing food structure and composition to enhance
 nutraceutical bioactivity to support cancer inhibition. In *Seminars in cancer biology*:
 Elsevier.
- Mehrgan, H., & Mortazavi, S. A. (2010). The release behavior and kinetic evaluation of
 diltiazem HCl from various hydrophilic and plastic based matrices. *Iranian Journal of Pharmaceutical Research*, 137-146.
- Mkaddem, M., Bouajila, J., Ennajar, M., Lebrihi, A., Mathieu, F., & Romdhane, M. (2009).
 Chemical composition and antimicrobial and antioxidant activities of Mentha (longifolia L. and viridis) essential oils. *Journal of food science*, *74*(7), M358-M363.
- Murugesan, R., & Orsat, V. (2012). Spray drying for the production of nutraceutical ingredients—a review. *Food and Bioprocess Technology*, 5(1), 3-14.
- Najafi, M. N., Hosaini, V., Mohammadi-Sani, A., & Koocheki, A. (2016). Physical stability,
 flow properties and droplets characteristics of Balangu (Lallemantia royleana) seed
 gum/whey protein stabilized submicron emulsions. *Food Hydrocolloids*, 59, 2-8.
- Okutan, N., Terzi, P., & Altay, F. (2014). Affecting parameters on electrospinning process and
 characterization of electrospun gelatin nanofibers. *Food Hydrocolloids*, *39*, 19-26.
- Otálora, M. C., Carriazo, J. G., Iturriaga, L., Nazareno, M. A., & Osorio, C. (2015).
 Microencapsulation of betalains obtained from cactus fruit (Opuntia ficus-indica) by
 spray drying using cactus cladode mucilage and maltodextrin as encapsulating agents.
 Food chemistry, 187, 174-181.

- Peltonen, L., Valo, H., Kolakovic, R., Laaksonen, T., & Hirvonen, J. (2010). Electrospraying,
 spray drying and related techniques for production and formulation of drug
 nanoparticles. *Expert opinion on drug delivery*, 7(6), 705-719.
- Peppas, N. A., & Sahlin, J. J. (1989). A simple equation for the description of solute release.
 III. Coupling of diffusion and relaxation. *International journal of pharmaceutics*, 57(2), 169-172.
- Pereira, M. C., Oliveira, D. A., Hill, L. E., Zambiazi, R. C., Borges, C. D., Vizzotto, M.,
 Mertens-Talcott, S., Talcott, S., & Gomes, C. L. (2018). Effect of nanoencapsulation
 using PLGA on antioxidant and antimicrobial activities of guabiroba fruit phenolic
 extract. *Food chemistry*, 240, 396-404.
- Pérez-Masiá, R., Lagaron, J. M., & Lopez-Rubio, A. (2015). Morphology and stability of edible
 lycopene-containing micro-and nano-capsules produced through electrospraying and
 spray drying. *Food and Bioprocess Technology*, 8(2), 459-470.
- Pérez-Masiá, R., Lagaron, J. M., & López-Rubio, A. (2014). Surfactant-aided electrospraying
 of low molecular weight carbohydrate polymers from aqueous solutions. *Carbohydrate polymers, 101*, 249-255.
- Prajapati, B. G., & Patel, M. (2010). A technology update: Electro spray technology.
 International Journal of Pharmaceutical Sciences Review and Research, 1(1), 12-13.
- Prakash, B., Kujur, A., Yadav, A., Kumar, A., Singh, P. P., & Dubey, N. (2018).
 Nanoencapsulation: An efficient technology to boost the antimicrobial potential of plant essential oils in food system. *Food Control*, 89, 1-11.
- Razavi, S. M. A., Cui, S. W., & Ding, H. (2016). Structural and physicochemical characteristics
 of a novel water-soluble gum from Lallemantia royleana seed. *International journal of biological macromolecules*, 83, 142-151.
- Sadeghi-Varkani, A., Emam-Djomeh, Z., & Askari, G. (2018). Physicochemical and
 microstructural properties of a novel edible film synthesized from Balangu seed
 mucilage. *International journal of biological macromolecules, 108*, 1110-1119.
- Santos, C., Silva, C. J., Büttel, Z., Guimarães, R., Pereira, S. B., Tamagnini, P., & Zille, A.
 (2014). Preparation and characterization of polysaccharides/PVA blend nanofibrous
 membranes by electrospinning method. *Carbohydrate polymers*, *99*, 584-592.
- Song, X., Zhao, Y., Hou, S., Xu, F., Zhao, R., He, J., Cai, Z., Li, Y., & Chen, Q. (2008). Dual
 agents loaded PLGA nanoparticles: systematic study of particle size and drug
 entrapment efficiency. *European Journal of Pharmaceutics and Biopharmaceutics*,
 69(2), 445-453.
- Stephansen, K., García-Díaz, M., Jessen, F., Chronakis, I. S., & Nielsen, H. M. (2016).
 Interactions between surfactants in solution and electrospun protein fibers: effects on release behavior and fiber properties. *Molecular pharmaceutics*, 13(3), 748-755.
- Stijnman, A. C., Bodnar, I., & Tromp, R. H. (2011). Electrospinning of food-grade
 polysaccharides. *Food Hydrocolloids*, 25(5), 1393-1398.
- Wang, H., Hao, L., Wang, P., Chen, M., Jiang, S., & Jiang, S. (2017). Release kinetics and
 antibacterial activity of curcumin loaded zein fibers. *Food Hydrocolloids*, 63, 437-446.
- Yaghoobi, N., Majidi, R. F., ali Faramarzi, M., Baharifar, H., & Amani, A. (2017). Preparation,
 optimization and activity evaluation of PLGA/streptokinase nanoparticles using
 electrospray. Advanced pharmaceutical bulletin, 7(1), 131.
- Yeo, Y., & Park, K. (2004). Control of encapsulation efficiency and initial burst in polymeric
 microparticle systems. *Archives of pharmacal research*, 27(1), 1.
- Zaeim, D., Sarabi-Jamab, M., Ghorani, B., Kadkhodaee, R., & Tromp, R. H. (2018).
 Electrospray-assisted drying of live probiotics in acacia gum microparticles matrix.
 Carbohydrate polymers, 183, 183-191.

- Zhao, J., Sun, Z., Shao, Z., & Xu, L. (2016). Effect of surface-active agent on morphology and
 properties of electrospun PVA nanofibres. *Fibers and Polymers*, *17*(6), 896-901.
- Zhou, Y., Liang, K., Zhang, C., Li, J., Yang, H., Liu, X., Yin, X., Chen, D., Xu, W., & Xiao,
 P. (2017). Photocrosslinked methacrylated chitosan-based nanofibrous scaffolds as
 potential skin substitute. *Cellulose*, 24(10), 4253-4262.
- Zlotogorski, A., Dayan, A., Dayan, D., Chaushu, G., Salo, T., & Vered, M. (2013a).
 Nutraceuticals as new treatment approaches for oral cancer–I: Curcumin. *Oral oncology*, 49(3), 187-191.
- Zlotogorski, A., Dayan, A., Dayan, D., Chaushu, G., Salo, T., & Vered, M. (2013b).
 Nutraceuticals as new treatment approaches for oral cancer: II. Green tea extracts and
 resveratrol. *Oral oncology*, 49(6), 502-506.
- 816
- 817

818 Highlights:

- Electrospray-assisted fabrication of Balangu seed gum nano-capsules was optimized;
- The effects of processing parameters on morphology and properties were studied;
- *Mentha longifolia L.* essential oil was electro-encapsulated in the nano-capsules;
- The types of jet-modes were reported for electrospraying of Balangu seed gum;
- Kinetic modeling ws applied to essential oil release from Balangu nano-capsules;
- Different release behaviors of *Mentha longifolia L*. essential oil were studied.