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1 Highlights

- 2 • Given the average cell-cycle time, the minimum invasion speed is obtained when the cell-
3 cycle time is Dirac-delta distributed;
- 4 • We find the range of variability for the speed of mathematical models of cell migration
5 which adopt realistic hypoexponential cell-cycle time distributions;
- 6 • The maximum speed adopting hypoexponential distributions is obtained by using an ex-
7 ponentially distributed cell-cycle time;
- 8 • We find an analytical expression for the invasion speed of general Erlang cell-cycle time
9 distribu- tions.

The invasion speed of cell migration models with realistic cell cycle time distributions

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Abstract

Cell proliferation is typically incorporated into stochastic mathematical models of cell migration by assuming that cell divisions occur after an exponentially distributed waiting time. Experimental observations, however, show that this assumption is often far from the real cell cycle time distribution (CCTD). Recent studies have suggested an alternative approach to modelling cell proliferation based on a multi-stage representation of the CCTD.

In this paper we investigate the connection between the CCTD and the speed of the collective invasion. We first state a result for a general CCTD, which allows the computation of the invasion speed using the Laplace transform of the CCTD. We use this to deduce the range of speeds for the general case. We then focus on the more realistic case of multi-stage models, using both a stochastic agent-based model and a set of reaction-diffusion equations for

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29 the cells' average density. By studying the corresponding travelling wave solutions, we obtain
30 an analytical expression for the speed of invasion for a general N -stage model with identical
31 transition rates, in which case the resulting cell cycle times are Erlang distributed. We show
32 that, for a general N -stage model, the Erlang distribution and the exponential distribution
33 lead to the minimum and maximum invasion speed, respectively. This result allows us to
34 determine the range of possible invasion speeds in terms of the average proliferation time for
35 any multi-stage model.

36 *Keywords:* Cell migration, multi-stage model, cell cycle time distribution, invasion speed,
37 agent-based model, travelling wave.

38 1 Introduction

39 Cellular invasion is a process of fundamental importance in numerous morphogenetic and patho-
40 logical mechanisms. Important examples of processes in which cell migration plays a crucial role
41 include embryonic development [Gilbert, 2003, Keller, 2005], wound healing [Maini et al., 2004,
42 Deng et al., 2006] and tumour invasion [Hanahan and Weinberg, 2000].

43 Understanding how the properties of the individual cells contribute to the formation and
44 the propagation of the invasion wave is of fundamental importance. In fact, this can reveal the
45 micro-scale mechanisms that are responsible for a given phenomenological aspect, and hence
46 suggest effective therapeutic approaches to inhibit, or enhance, cell migration by interrupting
47 the cell cycle [Sadeghi et al., 1998, Gray-Schopfer et al., 2007, Haass and Gabrielli, 2017].

48 Despite the large variety of actions and interactions which cells can undergo, there are at
49 least two aspects of cells' behaviour which are essential in order for the invasion to take place.
50 These are cell motility and cell proliferation [Simpson et al., 2007, Mort et al., 2016]. If one
51 of these two aspects does not occur properly, the impact on the collective invasion is typically
52 evident and it can affect the success of the colonisation. For example, Mort et al. [2016] show

53 using joint experimental and a modelling approach that the failure of colonisation of the mouse
54 embryo by melanoblasts in Kit mutants is probably driven by reduced proliferation.

55 Extensive research has focused on the effect that cell motility and proliferation behaviours
56 have on the speed of the invasion, c . The common approach makes use of simple mathematical
57 models which typically take the form of a stochastic agent-based model (ABM) [Anderson and
58 Chaplain, 1998, Deutsch and Dormann, 2007] or a deterministic partial differential equation
59 (PDE) [Murray, 2007, Wise et al., 2008]. By computing the invasion speed of the model, either
60 analytically or numerically, it is possible to link the parameters which modulate the movement
61 and proliferation with the speed of invasion.

62 Many studies have investigated this link in more general contexts, beginning with the seminal
63 work of Fisher [1937] and Kolmogorov A. [1991], and more recently more complex models of
64 populations with multiple types or stages [Elliott and Cornell, 2012, Neubert and Caswell, 2000].
65 From these studies, it is well known that, when agents' motility is modelled as diffusion with
66 diffusion coefficient D and proliferation occurs at rates λ , the invasion speed is proportional to
67 the square root of the product of the rates, i.e. $c \propto \sqrt{D\lambda}$ [Fisher, 1937].

68 It is important to notice that the majority of the literature on the speed of invasion of trav-
69 elling waves is based on the assumption that proliferation events occur as independent Poisson
70 processes [Simpson et al., 2007, Mort et al., 2016]. In the context of cell migration, this is
71 equivalent to assuming that cells proliferate after an exponentially distributed random time.
72 However, experimental observations show that the cell cycle time distribution (CCTD) is typi-
73 cally non-monotonic and differs substantially from an exponential distribution (see Figure 1 (f)
74 for an example) [Golubev, 2016, Yates et al., 2017, Chao et al., 2018].

75 There is a vast literature regarding the appropriate representation of the CCTD [Csikász-
76 Nagy et al., 2006, Gérard and Goldbeter, 2009, Powathil et al., 2012]. One class of representa-

77 tions, known as multi-stage models (MSMs), have gained particular attention in several recent
78 studies [Yates et al., 2017, Vittadello et al., 2018, Chao et al., 2018]. The main idea of MSMs is to
79 partition the cell cycle into N sequential stages. As time evolves, each cell can transit from one
80 stage, i , to the next one, $i + 1$, after an exponentially distributed waiting time with parameter λ_i .
81 When a cell is found at the last stage, N , it can proliferate with rate λ_N , which leads the cell to
82 split into two daughter cells, both initialised at the first stage. The main motivation that makes
83 MSMs mathematically appealing is the Markov property of the exponentials which simplifies
84 both the analytical investigation of the model and its computational implementation. Moreover,
85 MSMs lead to CCTDs that are hypoexponential and hypoexponential distributions have been
86 shown to provide an excellent agreement with experimental data [Golubev, 2016, Yates et al.,
87 2017, Chao et al., 2018].

88 Despite the fact that there is some evidence to suggest that the cell cycle is a series of
89 uncoupled exponentially distributed phases [Chao et al., 2018], Yates et al. [2017] were at pains
90 to point out that the stages in their MSM do not correspond to the phases in the cell cycle, but
91 are tools which allow to fit the correct cell cycle distribution. Similarly, here, we are reticent
92 to link the N stages of our model to N realistic steps in biological cell cycles. Especially since,
93 when fitting to experimental data, different choices of N can give almost equally good agreement
94 to cell cycle distributions data. In particular, the *stages* of the MSMs should not be confused
95 with the biological *phases* of the cell cycle which, in general, are not exponentially distributed
96 (see Figure 1) [Chao et al., 2018].

97 Whilst previous studies have investigated MSMs extensively in the case of spatially uniform
98 scenarios [Yates et al., 2017], there is still little understanding about the effect which MSMs have
99 on invading waves of cells. In particular, it is not clear how, and to what extent, a multi-stage
100 representation of the CCTD can impact on the speed of invasion.

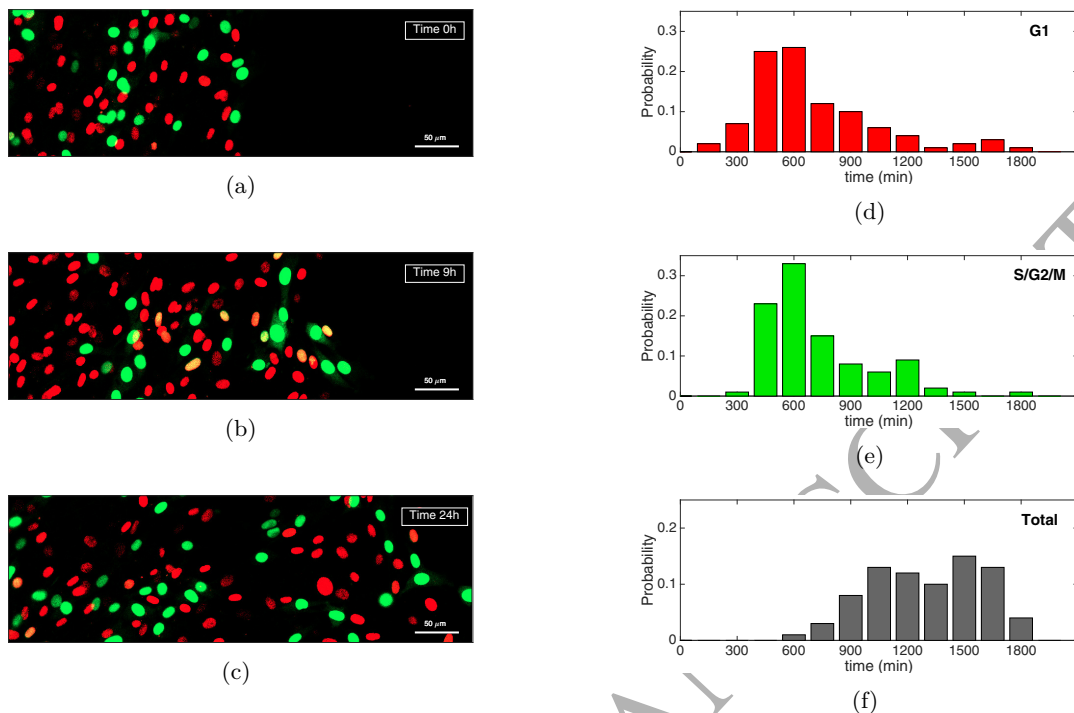


Figure 1: Panels (a-c): Mouse NIH-3T3 fibroblasts with Fucci2a status migrating into open space [Mort et al., 2014]. The Fucci2a system incorporates genetically encoded probes that highlight in red the nuclei of cells in the G1 phase and in green those of cells in one of the other phases, S/G2/M. Panels (d-f): experimental distributions of the time length of the G1 phase (panel (d)), S/G2/M phases (panel (e)) and total CCTD (panel (f)). Both the G1 and S/G2/M distributions show a clear non-monotonic trend, which indicates that they are not exponentially distributed. To capture both these non-monotonicities using a MSM for the CCTD, a minimum of four stages is required, two for each of the two phases.

The most recent progress on this was made by Vittadello et al. [2018]. In their work, the authors derive an analytical expression for the invasion speed of a 2-stage MSM in terms of the two rates of stage transition, λ_1, λ_2 , and the diffusion coefficient of cells, D :

$$c = \sqrt{2D \left[-\lambda_1 - \lambda_2 + \sqrt{\lambda_1^2 + 6\lambda_1\lambda_2 + \lambda_2^2} \right]}. \quad (1)$$

101 The findings of Vittadello et al. [2018] provide useful insights in the qualitative effect of the MSMs.

102 However a general expression for the invasion speed, as in equation (1), but for biologically

103 realistic MSMs, which typically have ten or more stages [Yates et al., 2017, Chao et al., 2018], is
104 not feasible analytically. Hence, there are important questions about the quantitative effect of
105 MSMs on the invasion speed which remain unanswered. In particular, the range of variability in
106 speed for a general N -stage MSM has yet to be studied.

107 To investigate the effect of incorporating a general CCTD into the invasion models, we follow
108 two distinct approaches. In the first part of the paper, we formulate a generalisation of the Fisher-
109 KPP equation in which the cell population is structured by age. By studying the traveling wave
110 solutions of the model, we derive an implicit equation for the speed of invasion in terms of the
111 Laplace transform of the CCTD. We obtain an expression for the minimum wave speed under
112 this model and show that for a general CCTD the invasion can be arbitrarily fast.

113 In the second part of the paper, we focus our attention on MSMs. We study a spatially
114 extended ABM which is designed to mimic cell invasion on a regular two-dimensional lattice. For
115 each agent, we implement a general N -stage MSM to simulate the stochastic waiting time before
116 the agent attempts to divide into two daughters. Through a mean-field closure approximation on
117 the average agent density, we derive a system of N reaction-diffusion PDEs which represents a
118 generalisation of the model of Vittadello et al. [2018]. By applying the front propagation method
119 [Van Saarloos, 2003] to the system of PDEs, we reduce the computation of the invasion speed
120 to an eigenvalue problem in terms of the rates of transition between consecutive stages, λ_i . We
121 use this result to study the case of identical transition rates, that corresponds to modelling the
122 CCTD as Erlang. In this case we provide the exact analytical expression for the speed. Finally,
123 we formulate a result for the maximum and minimum speed for a general N -stage MSM.

124 The paper is organised as follows. In Section 2 we define the age-structured model and we
125 derive the implicit equation for the invasion speed for general CCTD. In Section 3 we define two
126 MSMs: a stochastic ABM and the corresponding mean-field approximation. In Section 3.1 we

127 explain how to apply the front propagation method and we state the eigenvalue problem. We
 128 present our results on Erlang distributed cell cycle times and the general hypoexponential case in
 129 Section 3.2. We conclude in Section 4 with a brief discussion of this work and future challenges.

130 2 Age-Structured Model

131 The Fisher-KPP equation implicitly assumes Markov dynamics for the individual cells making
 132 up the population, implying a cell cycle time with an exponential distribution [Fisher, 1937].
 133 One way to adapt the model to allow for an arbitrary cell cycle time distribution is through the
 134 addition of age-structure. Cells have an associated age, denoted by a , which takes values in the
 135 positive real numbers and increases as time evolves. Cell motility is modelled as diffusion, with
 136 diffusivity D , and they proliferate with an age-dependent rate, $h(a)$.

We can write down a simple linear PDE for the density of cells with age a and spatial location
 x at time t , $C(a, x, t)$, as follows

$$\begin{aligned} \frac{\partial}{\partial t} C(a, x, t) &= -\frac{\partial}{\partial a} C(a, x, t) + D \frac{\partial^2}{\partial x^2} C(a, x, t) - h(a) C(a, x, t) \\ C(0, x, t) &= 2 \int_0^\infty h(s) C(s, x, t) ds. \end{aligned} \quad (2)$$

The function $h(s)$ is the hazard rate, related to the probability density function $f(s)$ of the age
 at which cells divide (i.e. the CCTD) via

$$h(s) = \frac{f(s)}{\int_s^\infty f(a) da}, \quad f(s) = h(s) \exp\left(-\int_0^s h(a) da\right). \quad (3)$$

137 The boundary condition for $C(0, x, t)$ in equation (2) gives the density of newborn cells as twice
 138 the total rate of cell division. Note that we have neglected from our formulation in system (2)
 139 any non-linear terms arising from crowding effects, as these are not relevant to the speed of the

140 front propagation. This model is a simple spatial adaptation of the McKendrick-Von Foerster
 141 equation for growing age-structured populations, and has been studied before [Webb and Webb,
 142 1985, Al-Omari and Gourley, 2002, Gabriel et al., 2012].

As our first result, we show that the speed of propagation for the model (2) is determined by the Laplace transform of the CCTD, defined by

$$\mathcal{L}\{f\}(s) = \int_0^{\infty} e^{-sa} f(a) da. \quad (4)$$

Theorem 1 If $\lim_{s \rightarrow \infty} \mathcal{L}\{f\}(s) < 1/2$ then the PDE (2) admits travelling wave solutions with propagation speed $c > 2\sqrt{D\lambda}$, where $\lambda > 0$ is the unique solution to

$$\mathcal{L}\{f\}(\lambda) = 1/2. \quad (5)$$

Proof. The system (2) is separable, hence we seek solutions of the form $C(a, x, t) = v(a)w(x-ct)$, corresponding to a travelling wave with speed c and internal age structure given by v . Inserting into (2) and rearranging, we find

$$c \frac{w'}{w} + D \frac{w''}{w} = \frac{v'}{v} + h. \quad (6)$$

The left-hand side here is a function only of $x - ct$, whilst the right-hand side is a function only of a . We thus determine that both are equal to a constant, say $-\lambda$. The w equation becomes

$$\lambda w + cw' + Dw'' = 0, \quad (7)$$

which is well-known as the linearisation of the Fisher-KPP equation, admitting travelling wave

solutions for all $c > 2\sqrt{D\lambda}$. The equation for v has solution

$$v(a) = v(0) \exp\left(-a\lambda - \int_0^a h(\alpha) d\alpha\right). \quad (8)$$

The boundary condition then gives us

$$1 = 2 \int_0^\infty h(a) \exp\left(-a\lambda - \int_0^a h(\alpha) d\alpha\right) da, \quad (9)$$

143 from which the definition of the hazard rate, equation (3), gives the result $1 = 2\mathcal{L}\{f\}(\lambda)$. Unique-
 144 ness of the solution (when one exists) follows from the monotonicity of the Laplace transform of
 145 a probability density. \square

We can use the previous result to investigate the range of speeds for an arbitrary CCTD with a given mean, $\bar{\mu}$. By using Jensen's inequality we have that for any positive supported f with mean $\bar{\mu}$

$$\mathcal{L}\{f\}(\lambda) \leq e^{-\lambda\bar{\mu}} = \mathcal{L}\{\delta_{\bar{\mu}}\}(\lambda), \quad (10)$$

where $\delta_{\bar{\mu}}$ is the Dirac delta function concentrated at $\bar{\mu} > 0$. From the monotonicity of the Laplace transform of a probability density, it follows that the minimum speed is obtained by using $f = \delta_{\bar{\mu}}$, which gives

$$c \geq 2\sqrt{\frac{D \ln 2}{\bar{\mu}}} \quad (11)$$

We now use Theorem 1 to show that there is no upper bound for the speed of invasion of a general CCTD with a given mean. Consider the set of probability density functions defined as

$$f_\varepsilon(x) = \frac{1}{2} \left(\delta_{\varepsilon\bar{\mu}} + \delta_{(2-\varepsilon)\bar{\mu}} \right), \quad (12)$$

where $\varepsilon \leq 1$. It follows immediately that each member of this set of functions have mean $\bar{\mu}$ and Laplace transform given by:

$$\mathcal{L}\{f_\varepsilon\}(\lambda) = \frac{1}{2} \left(e^{-\lambda\varepsilon\bar{\mu}} + e^{-\lambda(2-\varepsilon)\bar{\mu}} \right). \quad (13)$$

By substituting the expression (13) into equation (5) and rearranging, we obtain the implicit equation for λ given by

$$\lambda\varepsilon\bar{\mu} = -\ln \left(1 - e^{-2\lambda\bar{\mu}} \right). \quad (14)$$

146 The right-hand side of equation (14) is a strictly decreasing function of λ that converges to 0 as
 147 $\lambda \rightarrow \infty$. Therefore, we can always choose ε small enough so that the solution of equation (14) is
 148 arbitrarily large.

This demonstrates that, assuming that the CCTD is a general function with mean $\bar{\mu}$ and positive support, the range of possible invasion speeds is given by

$$c \in \left[2\sqrt{\frac{D \ln 2}{\bar{\mu}}}, \infty \right). \quad (15)$$

149 The result in Theorem 1 is important because it establishes the connection between a general
 150 CCTD and the corresponding invasion speed. However, for some particular classes of distribu-
 151 tions, solving equation (5) analytically can be challenging and the method of this Section does
 152 not provide any deeper insights. In particular, this is true for hypoexponential distributions,
 153 which are of special interest in the context of cell proliferation. In remaining part of the paper
 154 we further explore this class of distributions using a MSM of cell migration.

155 3 Multi-Stage Models

156 In this section we introduce the two MSM that we will use throughout the remainder of this
 157 paper. Firstly, we define a discrete ABM, in which the multi-stage representation of the CCTD
 158 is implemented as a stochastic feature of each cell at the microscale. Secondly, we introduce a
 159 system of deterministic PDEs describing the average cell density in a macroscopic manner.

160 **The ABM** We consider a continuous-time ABM on a two-dimensional regular square lattice,
 161 with a given spacing denoted by Δ . Each cell is modelled as a single agent which moves and
 162 proliferates. Volume exclusion is incorporated by allowing at most one agent to occupy a given
 163 lattice site.

164 Agents move according to a simple excluding random walk on the lattice. Each agent attempts
 165 a movement after an exponentially distributed waiting time with rate α . When this happens,
 166 a new position is chosen uniformly from one of the four nearest neighbouring sites and the
 167 movement takes place only if the selected site is empty. The event is aborted otherwise.

168 We implement cell proliferation using a MSM. We divide the cell cycle into N sequential
 169 stages. Agents at one of the first $N - 1$ stages, $i = 1, \dots, N - 1$, move to the next stage after
 170 an exponentially distributed waiting time of rate λ_i . Agents at the last stage, N , can attempt a
 171 proliferation event, after a further exponentially distributed waiting time of rate λ_N . In order to
 172 attempt a proliferation event, a target site is selected uniformly at random from one of the four
 173 nearest neighbouring sites. If such site is empty, a new first-stage agent is located on it, and the
 174 proliferating agent is returned to the first stage. If the target site is occupied, the proliferation
 175 event is aborted and the proliferating agent remains at the last stage¹.

¹Alternatively, we could choose to return the proliferating agent to the first stage every time an abortion occurs. This model has been studied in Yates et al. [2017] for homogeneously distributed agents. This modification does not substantially change our results. This is because our analysis of the speed of the wave front is based on low density regions, where abortion of events does not play an important role. For this reason, we decided to focus only on the stated version of the model.

176 We simulate the cell invasion by populating the first 10 columns of the lattice with agents at
 177 stages that are chosen uniformly at random. We impose zero flux boundary conditions on the
 178 x -direction and periodic boundary conditions on the y -direction. Agents are displaced uniformly
 179 at random in the vertical direction, so we can reduce the dimensionality of the problem by
 180 considering the average column density [Simpson et al., 2009].

181 **The PDE model** Here we define the continuous model for the average column density which
 182 will be the object of the invasion speed analysis.

We denote by $S_i(x, t)$ the density of i -stage agents in the column x at time t , averaged over multiple realisations of the ABM. Let $C(x, t)$ be the total density of column x at time t , i.e.

$$C(x, t) = \sum_{i=1}^N S_i(x, t). \quad (16)$$

By writing down the master equation of S_i , for $i = 1, \dots, N$ and taking the limit as $\Delta \rightarrow 0$, while keeping $\alpha\Delta^2$ constant, one can derive a system of reaction-diffusion PDEs for the column densities of the different stages:

$$\begin{cases} \frac{\partial S_1}{\partial t} &= D \frac{\partial}{\partial x} \left[(1-C) \frac{\partial S_1}{\partial x} + S_1 \frac{\partial C}{\partial x} \right] + 2\lambda_N(1-C)S_N - \lambda_1 S_1 \\ \frac{\partial S_i}{\partial t} &= D \frac{\partial}{\partial x} \left[(1-C) \frac{\partial S_i}{\partial x} + S_i \frac{\partial C}{\partial x} \right] + \lambda_{i-1} S_{i-1} - \lambda_i S_i & \text{for } i = 2, \dots, N-1 \\ \frac{\partial S_N}{\partial t} &= D \frac{\partial}{\partial x} \left[(1-C) \frac{\partial S_N}{\partial x} + S_N \frac{\partial C}{\partial x} \right] + \lambda_{N-1} S_{N-1} - \lambda_N(1-C)S_N, \end{cases} \quad (17)$$

183 where $D = \lim_{\Delta \rightarrow 0} \frac{\alpha\Delta^2}{4}$. Notice that other types of tessellations than the regular square lattice
 184 are common in the literature [Deutsch and Dormann, 2007, Simpson et al., 2018]. However, the
 185 model formulation and the corresponding mathematical analysis in these cases do not change
 186 substantially. For example, a detailed derivation for the three-stage model on an hexagonal

187 lattice can be found in Simpson et al. [2018]. Moreover, we should underline that the diffusivity
 188 of the cells in our model is independent of their stage which is not always true for real cells. We
 189 discuss this and other possible generalisations of the model in Section 4.

System (17) consists of a set of N reaction-diffusion PDEs with non-linearities in both the
 diffusion and the proliferation terms due to the effect of volume exclusion. Specifically, the term
 $(1 - C)$ accounts for the reduction in rate due to volume exclusion. Notice that by summing all
 the equations in (17), we obtain

$$\frac{\partial C}{\partial t} = D \frac{\partial^2 C}{\partial x^2} + \lambda_N (1 - C) S_N. \quad (18)$$

190 In other words, although the diffusion terms in each of the equations (17) are non-linear, the
 191 motility at the population-level is simple diffusion [Simpson et al., 2009]. Conversely, due to the
 192 dependence of equation (18) on S_N , it is not possible to obtain a closed PDE for the total agent
 193 density without further assumptions.

194 We conclude this section by showing a comparison of the two models in Figure 2. In the
 195 example, we choose realistic values of motility rate, α , and proliferation rate, $\bar{\lambda}$, as in Treloar
 196 et al. [2013] and Haass et al. [2014], respectively. We consider an ABM with five stages with
 197 increasing rates (the stage-to-stage transition rates are chosen to facilitate the visualisation of
 198 the different density profiles). In panels (a), (b) and (c) three successive snapshots are shown
 199 and the formation of the travelling wave appears clearly. As previously observed by Vittadello
 200 et al. [2018], due to the presence of volume exclusion, the travelling wave solutions of the N
 201 subpopulations of cells are of two qualitatively different types. The density profile of the first
 202 $N - 1$ subpopulations have the form of moving pulses located at the front of the total wave with
 203 the amplitude which depends on the rate of the corresponding stage. The profile of the last stage
 204 subpopulation, instead, appears as a moving wavefront which dominates the density at the back

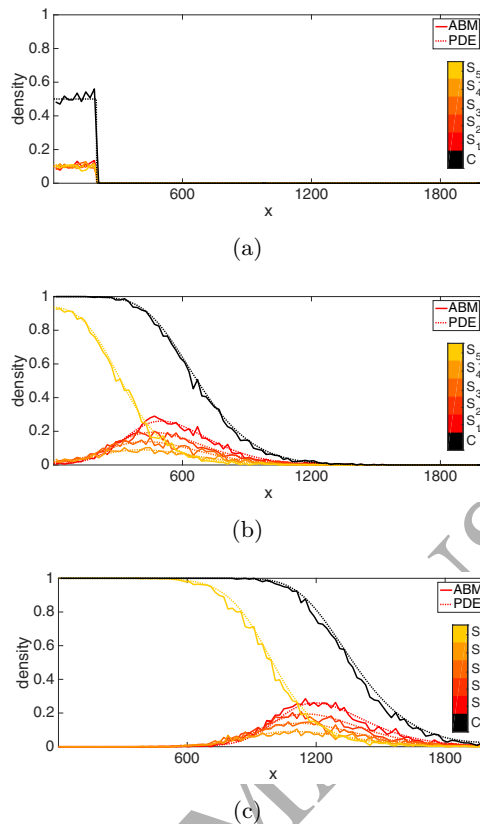


Figure 2: Comparison between the average column density for the ABM (full lines) and the PDE model (dotted lines) with a five-stage MSM. The panels show three snapshots of the evolution of the two models at time 0 (a), 150 (b) and 300 (c). In all cases, the profiles for the five different subpopulations are shown in different gradations of orange and the total density is plotted in black. The ABM profiles are obtained by averaging over 20 identically prepared simulations on a 2000×400 lattice. The other parameters of the models are $\Delta = 20$, $\alpha = 4$, $\bar{\lambda} = 0.0233$, $\lambda_1 = 0.15$, $\lambda_2 = 0.19$, $\lambda_3 = 0.25$, $\lambda_4 = 0.37$ and $\lambda_5 = 0.75$.

205 of the total wave.

206 The numerical solutions of the PDEs agree well with the average behaviour of the ABM.
 207 Therefore, we focus our attention on the the speed of the PDE model which we can investigate
 208 using an analytical approach (see Section 3.1).

209 It is important to note that the quantitative validity of our results on the PDE model will
 210 extend to the ABM only for the range of parameters which preserves the good agreement between

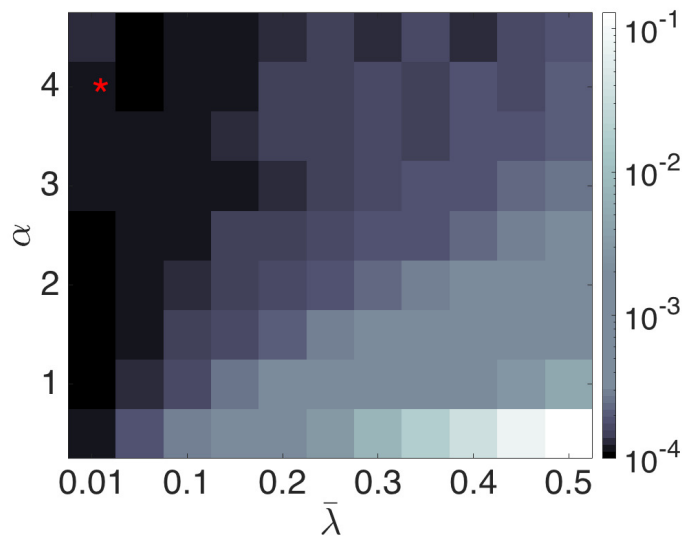


Figure 3: Histogram distance error (HDE) between column density profiles of ABM and the mean-field model for different parameters of movement and proliferation. The colour of each square denotes the HDE between the total density of the PDE and the ABM (averaging over 1000 simulations) as described in the legend. Dark squares represent small HDE which denotes good agreement between the two models, whereas light squares represent large HDE which denote a loss of agreement. The models are simulated on a 2000×400 domain with $\Delta = 20$ and the HDE is computed at the time when half of the domain in the mean-field model is occupied. All the ABMs are simulated with $N = 5$ and with transition rates proportional to the one of Figure 2. The red mark denotes the realistic parameter choice corresponding to Figure 2.

211 the two models. In Figure 3 we compare the total averaged column density profiles of the ABM
 212 and PDE for different parameters. The heat map shows the histogram distance error (HDE)²
 213 between ABM and the PDE model for different rates of movement and proliferation. When
 214 the rate of proliferation is large compared to the motility rate, the mean-field approximation
 215 loses its accuracy. This is a well known phenomenon which is caused by the presence of strong
 216 spatial correlations between occupied sites, induced by the proliferation [Middleton et al., 2014].
 217 Increasing the motility parameter tends to break up spatial correlations of neighbouring sites

²The HDE between two normalised histograms with values a_i and b_i at point i (i.e. $\sum a_i = \sum b_i = 1$) is defined as $HDE = \sum |a_i - b_i|/2$ [Cao and Petzold, 2006].

218 and, consequentially, to improve the accuracy of the mean-field approximation.

219 The results of Figures 2 and 3 confirm that for realistic choice of parameters ($\alpha \approx 4$ and
 220 $\bar{\lambda} \approx 0.02$ [Treloar et al., 2013, Haass et al., 2014]) the PDE model provides a good approximation
 221 of the ABM. This motivates us to focus our analysis on the continuum model. It is possible to
 222 derive more accurate descriptions in those cases where the agreement is lost using higher order
 223 moment closure schemes (see for example Baker and Simpson [2010], Markham et al. [2013]),
 224 but this is beyond the scope of this paper.

225 3.1 Wavespeed Analysis

226 In this Section we apply the front propagation method of Van Saarloos [2003] to system (17) to
 227 study the speed of invasion of the PDE model.

228

The system of equations (17) has two equilibria, an unstable empty state, $S_i(x, t) \equiv 0$ for
 $i = 1, \dots, N$, and a stable occupied state, $S_i(x, t) \equiv 0$ for $i = 1, \dots, N - 1$ and $S_N(x, t) \equiv 1$.
 Firstly we linearise the system about the unstable steady state, giving

$$\begin{cases} \frac{\partial S_1}{\partial t} = D \frac{\partial^2 S_1}{\partial x^2} + 2\lambda_N S_N - \lambda_1 S_1 \\ \frac{\partial S_i}{\partial t} = D \frac{\partial^2 S_i}{\partial x^2} + \lambda_{i-1} S_{i-1} - \lambda_i S_i \end{cases} \quad \text{for } i = 2, \dots, N . \quad (19)$$

We substitute

$$S_i(x, t) \propto \exp(-i\omega(k)t + ikx) ,$$

into equations (19), where i is the imaginary unit, $\omega(k)$ is the dispersion angular frequency of

the Fourier modes and k is the spatial wavenumber. Upon simplification, we obtain

$$\begin{cases} -i\omega(k) = -Dk^2 + 2\lambda_N - \lambda_1 \\ -i\omega(k) = -Dk^2 + \lambda_{i-1} - \lambda_i \quad \text{for } i = 2, \dots, N. \end{cases}$$

Following the front propagation method [Van Saarloos, 2003], the expression of the wave speed, c , is given by

$$c = \frac{\text{Im}[\omega(k^*)]}{\text{Im}[k^*]}, \quad (20)$$

where $k^* = iq$, with q real, and such that

$$\frac{d\omega}{dk}(k^*) = \frac{\text{Im}[\omega(k^*)]}{\text{Im}[k^*]}. \quad (21)$$

Notice that we can write down $i\omega(k)$ in the form

$$i\omega(k) = k^2 D - \rho, \quad (22)$$

where ρ is an eigenvalue of the matrix

$$\Lambda = \begin{bmatrix} -\lambda_1 & 0 & \dots & 0 & 2\lambda_N \\ \lambda_1 & -\lambda_2 & 0 & \dots & 0 \\ 0 & \lambda_2 & -\lambda_3 & \dots & 0 \\ \vdots & & \ddots & \ddots & \vdots \\ 0 & \dots & & \lambda_{N-1} & -\lambda_N \end{bmatrix}. \quad (23)$$

From expression (22) it follows that

$$\frac{d\omega}{dk}(k^*) = 2qD, \quad (24a)$$

$$\frac{\text{Im}[\omega(k^*)]}{\text{Im}[k^*]} = \frac{q^2D + \text{Re}[\rho]}{q}. \quad (24b)$$

By substituting equations (24) into (21), we obtain $q^2 = \text{Re}[\rho]/D$. Note that Λ is Metzler, and hence by Perron-Fobenius its rightmost eigenvalue is real. Hence, from (20), the wave speed of the invasion is given by

$$c = 2\sqrt{D\rho}, \quad (25)$$

where ρ is the maximum real eigenvalue of Λ , defined in terms of the characteristic polynomial of the matrix Λ , $\mathcal{P}_\Lambda(x)$, as follows

$$\rho(\Lambda) = \max\{x \in \mathbb{R} \mid \mathcal{P}_\Lambda(x) = 0\}. \quad (26)$$

229 This shows that the problem of finding the speed of invasion of the PDE model is equivalent to
230 computing the maximum real eigenvalue of the matrix Λ , $\rho(\Lambda)$.

231 3.2 Results

The characteristic polynomial of the matrix Λ can be computed directly from the matrix and it reads

$$\mathcal{P}_\Lambda(x) = \prod_{i=1}^N (\lambda_i + x) - 2 \prod_{i=1}^N \lambda_i. \quad (27)$$

232 In general, an analytical formula of the roots of the polynomial function $\mathcal{P}_\Lambda(x)$ is not available.
233 In this section we first consider the case of $\lambda_i = \lambda$ for $i = 1, \dots, N$ for which the maximum
234 eigenvalue $\rho(\Lambda)$ can be computed analytically. This corresponds to a special case of the general

235 hypoexponential distribution, known as the Erlang distribution. We conclude by proving a
 236 theorem in which we state the range of speed variability for the general hypoexponential CCTD.

The Erlang distribution Consider the case $\lambda_i = \lambda$ for $i = 1, \dots, N$, which corresponds the Erlang CCTD. Under this assumption, we can write down the characteristic equation of the matrix Λ , using formula (27), as

$$(\lambda + x)^N = 2\lambda^N. \quad (28)$$

The eigenvalues of Λ are then given by the solutions of equation (28) which are $x_j = \lambda (\xi^j \sqrt[N]{2} - 1)$ for $j = 1, \dots, N$, where $\xi = \exp(2\pi i/N)$ is the primitive N -th root of unity. Hence, we obtain that

$$\rho(\Lambda) = \lambda (\sqrt[N]{2} - 1). \quad (29)$$

By substituting the expression (29) into equation (25) we obtain the formula for the speed of invasion for the model with Erlang distribution

$$c = 2\sqrt{D\lambda (\sqrt[N]{2} - 1)}. \quad (30)$$

237 Notice that for $N = 1$, which corresponds to exponential CCTD, we recover the well known
 238 expression of the speed for the Fisher-KPP equation, $2\sqrt{D\lambda}$.

239 **The general case** For the case of a general hypoexponential distribution, there is no analytical
 240 formula for the expression of the maximum real eigenvalue of the matrix Λ . However, we find
 241 that the Erlang case and the exponential case, for which we do have the analytical formula of
 242 the speed, correspond to the lower and upper bound (respectively) for the speed of travelling
 243 waves with hypoexponential CCTD and a given total proliferation rate, $\bar{\lambda}$. This result follows

244 directly from the following theorem on the range of $\rho(\Lambda)$.

Theorem 2 Let $\rho(\Lambda)$ be defined by equation (26) as the maximum real eigenvalue of the matrix Λ . Then

$$\bar{\lambda}N \left(\sqrt[N]{2} - 1 \right) \leq \rho(\Lambda) < \bar{\lambda}, \quad (31)$$

245 where $\bar{\lambda} = \left(\sum_{i=1}^N 1/\lambda_i \right)^{-1}$.

A proof of Theorem 2 can be found in the appendix. It is immediate to interpret the result of the Theorem 2 in terms of invasion speeds. In particular, by using equation (25), together with the two inequalities (31), we deduce that the speed of the invasion of the PDE model with diffusion coefficient D and a general N -stage representation of the CCTD with total growth rate given by $\bar{\lambda}$, lies in the interval

$$c \in \left[2\sqrt{D\bar{\lambda}N \left(\sqrt[N]{2} - 1 \right)}, 2\sqrt{D\bar{\lambda}} \right). \quad (32)$$

We can generalise this result even further by taking the limit as $N \rightarrow \infty$ in the right-hand side of equation (32). Hence we obtain a general interval which holds for any multi-stage representation, regardless of the number of stages, which reads

$$c \in \left(2\sqrt{D\bar{\lambda} \ln 2}, 2\sqrt{D\bar{\lambda}} \right), \quad (33)$$

246 where we used $N \left(\sqrt[N]{2} - 1 \right) = \ln 2 + \mathcal{O}(N^{-1})$.

247 Notice that the lower bound of the interval (33) is equivalent to the lower bound for the
 248 general CCTD, obtained in (15) of Section 2. This can be intuitively understood by observing
 249 that, as we let number of stages of an hypoexponential distribution go to infinity while keeping the
 250 total rate, $\bar{\lambda}$, fixed, the variance of the distribution tends to zero. Consequently, the distribution

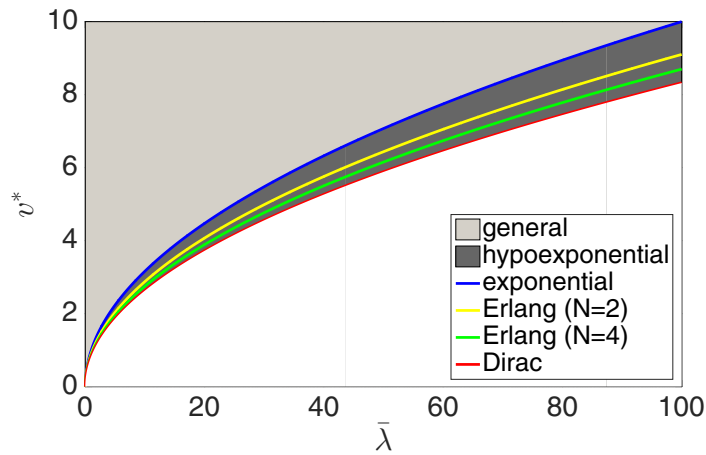


Figure 4: Illustration of the range of invasion speeds for a fixed mean proliferation rate and diffusion coefficient, $D = 1$. The two coloured regions represent the range of speed for a general CCTD. The dark grey subregion highlights the range of speeds for hypoexponential CCTDs. The global minimum speed is obtained by using the Dirac distribution (red line). The exponential CCTD (blue line) is the hypoexponential distribution which leads to maximum speed. There is no upper bound for the general case. Two examples of Erlang CCTDs with two stages (yellow line) and four stages (green line) are also shown.

251 converges to a Dirac function concentrated in the mean, $\bar{\mu} = \bar{\lambda}^{-1}$, which we have proved in
 252 Section 2 to be the distribution corresponding to the minimum invasion speed. In Figure 4 we
 253 summarise our findings about the range of invasion speed for different CCTD through a graphical
 254 representation.

255 4 Conclusion

256 In this work we investigated the quantitative effect of implementing a realistic CCTD into models
 257 of cell invasion. Firstly, we derived a general result from a generalised version of the Fisher-KPP
 258 equation. Then we investigated the case of MSMs by implementing a simple ABM of cells under-
 259 going undirected migration and proliferation by division, in which the time between successive
 260 divisions is modelled using a multi-stage representation (i.e. the CCTD is hypoexponential). By

261 studying a continuous version of the ABM, we connected the type of CCTD to the speed of the
 262 corresponding invasion.

263 The results indicate that, for a fixed mean division time, the minimum speed of invasion is
 264 obtained by the Dirac distribution, while there is no upper bound. In other words, the invasion
 265 can be, in general, infinitely fast. However, when we focus our attention to the case of MSMs,
 266 which are known to represent well the experimental CCTD, our analysis shows that the speed can
 267 vary in a bounded interval (see Figure 4). More precisely, we show that the maximum invasion
 268 speed is reached by adopting an exponential CCTD, which leads to the classic the Fisher-KPP
 269 model. On the other hand, the minimum speed is obtained by partitioning the CCTD into
 270 multiple exponential stages with identical rates, which corresponds to the case of Erlang CCTD.
 271 Finally, by considering the limiting case of infinitely many stages, we find that the infimum value
 272 of the speed for the class of hypoexponential CCTD coincides with the global minimum for a
 273 general CCTD.

274 The results indicate the invasion speed changes with the variance of the CCTD, i.e. decreasing
 275 the variance in the proliferation time distribution leads to slower invasion. We found that the
 276 maximum reduction in comparison to the classical formula for the Fisher-KPP model, is given
 277 by a multiplicative factor of $\sqrt{\ln 2} \approx 0.83$. Whilst interpreting this result in the context of
 278 experimental data is beyond the aim of this work, we want to stress that for number of stages
 279 $N \gg 1$, which is typically the case for experimentally observed distributions [Golubev, 2016,
 280 Yates et al., 2017, Chao et al., 2018], the speed converges to the lower bound of equation (33)
 281 with order given by $\mathcal{O}(N^{-1})$. This suggests that, with the only information of the mean of the
 282 CCTD (equivalently, the total rate), including the factor $\sqrt{\ln 2}$ in the formula for the speed leads
 283 to a more accurate estimation than the classic expression of Fisher-KPP.

284 In Section 3 we used a discrete ABM, but it is important to notice that alternative modelling

285 approaches might lead to different results. Although discrete space ABMs are widespread in
286 the literature [Deutsch and Dormann, 2007, Simpson et al., 2007, Cheeseman et al., 2014, Mort
287 et al., 2016, Simpson et al., 2018, Vittadello et al., 2018], a considerable number of studies focus
288 on lattice-free ABMs in which cells' positions are not constrained to a grid [Grima, 2008, Dyson
289 et al., 2012, Dyson and Baker, 2014, Middleton et al., 2014, Matsiaka et al., 2017]. Another
290 alternative approach, known as compartment-based model, consists in allowing multiple cells
291 occupying a single lattice site [Taylor et al., 2015, 2016, Cianci et al., 2017]. In the context of our
292 work, adopting alternative modelling regimes, such as lattice-free or compartment-based models,
293 would lead to different nonlinear factors in system (17). Since the analysis of the wave speed is
294 based on a linearisation of system (17), we believe that our results would still hold qualitatively.
295 However, a rigorous comparison of these modelling approaches is beyond the scope of this paper.

296 An important question that remains unanswered is the role of motility heterogeneity within
297 the cell cycle. Experimental studies have found that the motility of a cell can depend on its
298 cell cycle phase [Vittadello et al., 2018]. For example, during the mitotic phase, cells tend to
299 reduce their movement [Mort et al., 2016]. In order to investigate this phenomenon in the light
300 of the invasion speed, we could modify our model to allow different diffusion coefficients, D_i for
301 $i = 1, \dots, N$, for each stage in the system (17). Another aspect of the cell movement that can
302 vary within the cell cycle is the directional persistence. Our models do not incorporate directional
303 persistence of cells. However, it is possible to combine a MSM with existing models of directional
304 persistence [Codling et al., 2008, Gavagnin and Yates, 2018]. Unfortunately, the application of
305 the front propagation method of Van Saarloos [2003] (see Section 3.1) to these models leads to
306 a dead end and it may be necessary to study the problem using a different approach. We will
307 investigate this in future research.

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311 References

312 J. Al-Omari and S.A. Gourley. Monotone travelling fronts in an age-structured reaction-diffusion
313 model of a single species. *J. Math. Biol.*, 45(4):294–312, 2002.

314 A.R.A. Anderson and M.A.J. Chaplain. Continuous and discrete mathematical models of tumor-
315 induced angiogenesis. *Bull. Math. Biol.*, 60(5):857–899, 1998.

316 R.E. Baker and M.J. Simpson. Correcting mean-field approximations for birth-death-movement
317 processes. *Phys. Rev. E*, 82(4):041905, 2010.

318 Y. Cao and L. Petzold. Accuracy limitations and the measurement of errors in the stochastic
319 simulation of chemically reacting systems. *J. Comp. Phys.*, 212(1):6–24, 2006.

320 H.X. Chao, R.I. Fakhreddin, H.K. Shimerov, R.J. Kumar, G.P. Gupta, and J.E. Purvis. Evidence
321 that the cell cycle is a series of uncoupled, memoryless phases. *bioRxiv*, page 283614, 2018.

322 B.L. Cheeseman, D.F. Newgreen, and K.A. Landman. Spatial and temporal dynamics of cell
323 generations within an invasion wave: A link to cell lineage tracing. *J. Theor. Biol.*, 363:
324 344–356, 2014.

325 C. Cianci, S. Smith, and R. Grima. Capturing brownian dynamics with an on-lattice model of
326 hard-sphere diffusion. *Phys. Rev. E*, 95(5):052118, 2017.

327 E.A. Codling, M.J. Plank, and S. Benhamou. Random walk models in biology. *J. R. Soc.*
328 *Interface*, 5(25):813–834, 2008.

- 329 A. Csikász-Nagy, D. Battogtokh, K.C. Chen, B. Novák, and J.J. Tyson. Analysis of a generic
330 model of eukaryotic cell-cycle regulation. *Biophys. J.*, 90(12):4361–4379, 2006.
- 331 M. Deng, W.L. Chen, A. Takatori, Z. Peng, L. Zhang, M. Mongan, R. Parthasarathy, M. Sartor,
332 M. Miller, J. Yang, et al. A role for the mitogen-activated protein kinase kinase 1 in
333 epithelial wound healing. *Mol. Biol. Cell*, 17(8):3446–3455, 2006.
- 334 A. Deutsch and S. Dormann. *Cellular automaton modeling of biological pattern formation: char-*
335 *acterization, applications, and analysis*. Springer Science & Business Media, 2007.
- 336 L. Dyson and R.E. Baker. The importance of volume exclusion in modelling cellular migration.
337 *J. Math. Biol.*, 2014.
- 338 L. Dyson, P.K. Maini, and R.E. Baker. Macroscopic limits of individual-based models for motile
339 cell populations with volume exclusion. *Phys. Rev. E*, 86(3):031903, 2012.
- 340 E.C. Elliott and S.J. Cornell. Dispersal polymorphism and the speed of biological invasions.
341 *PLoS one*, 7(7):e40496, 2012.
- 342 R.A. Fisher. The wave of advance of advantageous genes. *Ann. Hum. Genet.*, 7(4):355–369,
343 1937.
- 344 P. Gabriel, S.P. Garbett, V. Quaranta, D.R. Tyson, and G.F. Webb. The contribution of age
345 structure to cell population responses to targeted therapeutics. *J. Theor. Biol.*, 311:19–27,
346 2012.
- 347 E. Gavagnin and C.A. Yates. Modeling persistence of motion in a crowded environment: The
348 diffusive limit of excluding velocity-jump processes. *Phys. Rev. E*, 97:032416, 2018.
- 349 C. Gérard and A. Goldbeter. Temporal self-organization of the cyclin/cdk network driving the
350 mammalian cell cycle. *Proc. Natl. Acad. Sci.*, 106(51):21643–21648, 2009.

- 351 S.F. Gilbert. The morphogenesis of evolutionary developmental biology. *Int. J. Dev. Biol.*, 47
352 (7-8):467, 2003.
- 353 A. Golubev. Applications and implications of the exponentially modified gamma distribution as
354 a model for time variabilities related to cell proliferation and gene expression. *J. Theor. Biol.*,
355 393:203–217, 2016.
- 356 V. Gray-Schopfer, C. Wellbrock, and R. Marais. Melanoma biology and new targeted therapy.
357 *Nature*, 445(7130):851, 2007.
- 358 R. Grima. Multiscale modeling of biological pattern formation. *Curr. Top. Dev. Biol.*, 81:435–460,
359 2008.
- 360 N.K. Haass and B. Gabrielli. Cell cycle-tailored targeting of metastatic melanoma: Challenges
361 and opportunities. *Exp. Dermatol.*, 26(7):649–655, 2017.
- 362 N.K. Haass, K.A. Beaumont, D.S. Hill, A. Anfosso, P. Mrass, M.A. Munoz, I. Kinjyo, and
363 W. Weninger. Real-time cell cycle imaging during melanoma growth, invasion, and drug
364 response. *Pigment Cell Melanoma Res.*, 27(5):764–776, 2014.
- 365 D. Hanahan and R.A. Weinberg. The hallmarks of cancer. *Cell*, 100(1):57–70, 2000.
- 366 R. Keller. Cell migration during gastrulation. *Curr. Opin. Cell Biol.*, 17(5):533–541, 2005.
- 367 Piscounov N. Kolmogorov A., Petrovskii I. A study of the diffusion equation with increase in the
368 amount of substance, and its application to a biological problem. In V.M. Tikhomirov, editor,
369 *Selected works of AN Kolmogorov*, pages 248–270. Springer, 1991.
- 370 P.K. Maini, D.L. S. McElwain, and D.I. Leavesley. Traveling wave model to interpret a wound-
371 healing cell migration assay for human peritoneal mesothelial cells. *Tissue Eng.*, 10(3-4):
372 475–482, 2004.

- 373 D.C. Markham, M.J. Simpson, P.K. Maini, E.A. Gaffney, and R.E. Baker. Incorporating spatial
374 correlations into multispecies mean-field models. *Phys. Rev. E*, 88(5):052713, 2013.
- 375 O.M. Matsiaka, C.J. Penington, R.E. Baker, and M.J. Simpson. Continuum approximations for
376 lattice-free multi-species models of collective cell migration. *J. Theor. Biol.*, 422:1–11, 2017.
- 377 A.M. Middleton, C. Fleck, and R. Grima. A continuum approximation to an off-lattice individual-
378 cell based model of cell migration and adhesion. *J. Theor. Biol.*, 359:220–232, 2014.
- 379 R.L. Mort, M.J. Ford, A. Sakaue-Sawano, N.O. Lindstrom, A. Casadio, A.T. Douglas, M.A.
380 Keighren, P. Hohenstein, A. Miyawaki, and I.J. Jackson. Fucci2a: a bicistronic cell cycle
381 reporter that allows Cre mediated tissue specific expression in mice. *Cell Cycle*, 13(17):2681–
382 2696, 2014.
- 383 R.L. Mort, R.J.H. Ross, K.J. Hainey, O.J. Harrison, M.A. Keighren, G. Landini, R.E. Baker, K.J.
384 Painter, I.J. Jackson, and C.A. Yates. Reconciling diverse mammalian pigmentation patterns
385 with a fundamental mathematical model. *Nat. Commun.*, 7, 2016.
- 386 J.D. Murray. *Mathematical biology: I. An introduction*, volume 17. Springer Science & Business
387 Media, 2007.
- 388 M.G. Neubert and H. Caswell. Demography and dispersal: calculation and sensitivity analysis
389 of invasion speed for structured populations. *Ecology*, 81(6):1613–1628, 2000.
- 390 G.G. Powathil, K.E. Gordon, L.A. Hill, and M.A.J. Chaplain. Modelling the effects of cell-cycle
391 heterogeneity on the response of a solid tumour to chemotherapy: biological insights from a
392 hybrid multiscale cellular automaton model. *J. Theor. Biol.*, 308:1–19, 2012.
- 393 H.M. Sadeghi, B. Seitz, S. Hayashi, L. LaBree, and P.J. McDonnell. In vitro effects of mitomycin-
394 c on human keratocytes. *J. Refract. Surg.*, 14(5):534–540, 1998.

- 395 M.J. Simpson, A. Merrifield, K.A. Landman, and B.D. Hughes. Simulating invasion with cellular
396 automata: connecting cell-scale and population-scale properties. *Phys. Rev. E*, 76(2):021918,
397 2007.
- 398 M.J. Simpson, K.A. Landman, and B.D. Hughes. Multi-species simple exclusion processes. *Phys.*
399 *A*, 388(4):399–406, 2009.
- 400 M.J. Simpson, W. Jin, S.T. Vittadello, T. Tambyah, J. Ryan, G. Gunasingh, N. Haass, and
401 S. McCue. Stochastic models of cell invasion with fluorescent cell cycle indicators. *Phys. A*,
402 510:375–386, 2018.
- 403 P.R. Taylor, C.A. Yates, M.J. Simpson, and R.E. Baker. Reconciling transport models across
404 scales: the role of volume exclusion. *Phys. Rev. E*, 92(040701), 2015.
- 405 P.R. Taylor, R.E. Baker, M.J. Simpson, and C.A. Yates. Coupling volume-excluding
406 compartment-based models of diffusion at different scales: Voronoi and pseudo-compartment
407 approaches. *J. R. Soc. Interface*, 13(120), 2016. ISSN 1742-5689.
- 408 K.K. Treloar, M.J. Simpson, P. Haridas, K.J. Manton, D.I. Leavesley, D.S. McElwain, and R.E.
409 Baker. Multiple types of data are required to identify the mechanisms influencing the spatial
410 expansion of melanoma cell colonies. *BMC Syst. Biol.*, 7(1):137, 2013.
- 411 W. Van Saarloos. Front propagation into unstable states. *Phys. Rep.*, 386(2-6):29–222, 2003.
- 412 S.T. Vittadello, S.W. McCue, G. Gunasingh, N.K. Haass, and M.J. Simpson. Mathematical
413 models for cell migration with real-time cell cycle dynamics. *Biophys. J.*, 114(5), 2018.
- 414 G.F. Webb and G. Webb. *Theory of nonlinear age-dependent population dynamics*. CRC Press,
415 1985.

416 S.M. Wise, J.S. Lowengrub, H.B. Frieboes, and V. Cristini. Three-dimensional multispecies
417 nonlinear tumor growth–I: model and numerical method. *J. Theor. Biol.*, 253(3):524–543,
418 2008.

419 C.A. Yates, M.J. Ford, and R.L. Mort. A multi-stage representation of cell proliferation as a
420 markov process. *Bull. Math. Biol.*, 79(12):2905–2928, 2017.

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421 Appendix

422 A Proof of Theorem 2

Proof. Let $\mu_i = 1/\lambda_i$ for every $i = 1, \dots, N$. By writing the characteristic equation $\mathcal{P}_\Lambda(x) = 0$ in terms of the parameters μ_i and upon rearranging, we obtain

$$\prod_{i=1}^N (\mu_i x + 1) = 2. \quad (\text{A.1})$$

We can write $\rho(\Lambda) = \rho(\mu_1, \dots, \mu_N) = \rho(\underline{\mu})$ as

$$\rho(\underline{\mu}) = \max \left\{ x \in \mathbb{R} \mid \prod_{i=1}^N (\mu_i x + 1) = 2 \right\}, \quad (\text{A.2})$$

423 for every $\underline{\mu} \in \{\mathbb{R}_{>0}\}^N$. It is easy to observe that $\rho(\underline{\mu})$ is a positive continuous function and we
 424 can extend the definition (A.2) to $\underline{\mu} \in \{\mathbb{R}_{\geq 0}\}^N \setminus \{(0, \dots, 0)\}$, by continuity.

Now fix $\bar{\lambda} = \left(\sum_{i=1}^N \mu_i\right)^{-1}$; without loss of generality we can take $\sum_{i=1}^N \mu_i = 1$, whence (31) becomes $N \left(\sqrt[N]{2} - 1\right) \leq \rho(\underline{\mu}) < 1$. The case of general $\bar{\lambda}$ follows by multiplying by rescaling factor. Since ρ is a continuous function, we aim to find the stationary points of $\rho(\underline{\mu})$ in the N -dimensional simplex:

$$\mathcal{U}_N = \left\{ (\mu_1, \dots, \mu_N) \in (0, 1]^N \mid \sum_{i=1}^N \mu_i = 1 \right\}. \quad (\text{A.3})$$

We apply the Lagrange multipliers method. Hence we study the Lagrangian function given by

$$\mathcal{L}(\mu_1, \dots, \mu_N, \sigma) = \rho(\underline{\mu}) + \sigma \left(\sum_{i=1}^N \mu_i - 1 \right). \quad (\text{A.4})$$

Throughout we adopt the notation $\mathcal{L}_j = \frac{\partial \mathcal{L}}{\partial \mu_j}$ and $\rho_j = \frac{\partial \rho}{\partial \mu_j}$. By imposing $\mathcal{L}_j = 0$ we obtain

$$\rho_j = -\sigma, \quad (\text{A.5})$$

for all $j = 1, \dots, N$. We can now differentiate equation (A.1) respect to μ_j , which gives us

$$0 = \sum_{i=1}^N \prod_{k \neq i} (1 + \mu_k \rho) (\rho \delta_{i,j} + \mu_i \rho_j) \quad (\text{A.6})$$

where $\delta_{i,j}$ denotes the Kronecker delta. If we multiply and divide each term of the right-hand side of equation (A.6) by $(1 + \mu_i \rho)$, we obtain

$$\begin{aligned} 0 &= \sum_{i=1}^N \frac{\rho \delta_{i,j} + \mu_i \rho_j}{1 + \mu_i \rho} \\ &= \frac{\rho}{1 + \mu_j \rho} + \rho_j \sum_{i=1}^N \frac{\mu_i}{1 + \mu_i \rho}, \end{aligned} \quad (\text{A.7})$$

By combining equations (A.5) and (A.7) we gain a condition on the coordinate μ_j of the stationary points, namely

$$\frac{\rho}{1 + \mu_j \rho} = \sigma \sum_{i=1}^N \frac{\mu_i}{1 + \mu_i \rho}. \quad (\text{A.8})$$

425 Notice that equation (A.8) holds for every $j = 1, \dots, N$ and it is independent of j , hence the
426 only stationary point of $\rho(\underline{\mu})$ in the simplex \mathcal{U}_N is the given by the centre $\underline{\mu}_N^* = (1/N, \dots, 1/N)$.

To conclude we need study the value of $\rho(\underline{\mu})$ on the boundary of the simplex, defined as

$$\partial \mathcal{U}_N = \left\{ (\mu_1, \dots, \mu_N) \in [0, 1]^N \mid \sum_{i=1}^N \mu_i = 1 \text{ and } \mu_j = 0, \exists j \in \{1, \dots, N\} \right\}. \quad (\text{A.9})$$

Let us consider the elements of $\partial \mathcal{U}_N$ with exactly n non-zero coordinates, with $n = 1, \dots, N - 1$.

Without loss of generality we can focus on the points of the form

$$(\mu_1, \dots, \mu_n, 0, \dots, 0) \in \partial\mathcal{U}_N, \quad (\text{A.10})$$

where $(\mu_1, \dots, \mu_n) \in \mathcal{U}_n$. Notice that the $\rho(\underline{\mu})$ is well defined in such points by continuity, as observed before. By repeating the Lagrange multiplier method in the sub-simplex \mathcal{U}_n , we find that the only stationary point of $\rho(\underline{\mu})$ of the form (A.10) is the one with $\mu_1 = \mu_2 = \dots = \mu_n$, i.e.:

$$\underline{\mu}_n^* = (\underbrace{1/n, \dots, 1/n}_n, 0, \dots, 0) \in \partial\mathcal{U}_N. \quad (\text{A.11})$$

427 This holds for every $n = 1, \dots, N - 1$, so we can write all the stationary points of $\rho(\underline{\mu})$ in $\partial\mathcal{U}_N$
 428 upon permutation of the coordinates in the form (A.11).

All the stationary points $\underline{\mu}_n^*$, for $n = 1, \dots, N$, correspond to an Erlang distribution for which we can compute the expression of ρ directly from the definition (A.2) as

$$\rho(\underline{\mu}_n^*) = n \left(\sqrt[n]{2} - 1 \right), \quad (\text{A.12})$$

for $n = 1, \dots, N$. The right-hand side of equation (A.12) is a decreasing function of n . We deduce that the centre of the simplex, $\underline{\mu}_N^* \in \mathcal{U}_N$, corresponds to the global minimum, i.e. for all $\underline{\mu} \in \mathcal{U}_N$

$$\rho(\underline{\mu}) \geq \rho(\underline{\mu}_N^*) = N \left(\sqrt[N]{2} - 1 \right). \quad (\text{A.13})$$

Finally, $\underline{\mu}_1^* \in \partial\mathcal{U}_N$ and all the points obtained by permuting its coordinates, correspond to

supremum points, i.e. for all $\mu \in \mathcal{U}_N$

$$\rho(\underline{\mu}) < \rho(\underline{\mu}_1^*) = 1. \quad (\text{A.14})$$

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