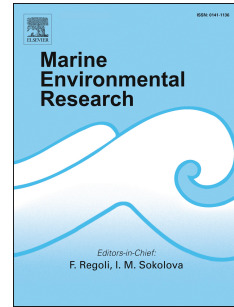


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Condition of pteropod shells near a volcanic CO₂ vent region

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Abstract

Natural gradients of pH in the ocean are useful analogues for studying the projected impacts of Ocean Acidification (OA) on marine ecosystems. Here we document the *in situ* impact of submarine CO₂ volcanic emissions (CO₂ vents) on live shelled-pteropods (planktonic gastropods) species *Creseis conica* in the Gulf of Naples (Tyrrhenian Sea, Mediterranean). Since the currents inside the Gulf will likely drive those pelagic calcifying organisms into and out of the CO₂ vent zones, we assume that pteropods will be occasionally exposed to the vents during their life cycle. Shell degradation and biomass were investigated in the stations located within and nearby the CO₂ vent emission in relation to the variability of sea water carbonate chemistry. A relative decrease in shell biomass (22%), increase in incidence of shell fractures (38%) and extent of dissolution were observed in *Creseis conica* collected in the Gulf of Naples compared to those from the Northern Tyrrhenian Sea (control stations). These results suggest that discontinuous but recurrent exposure to highly variable carbonate chemistry could consistently affect the characteristic of the pteropod shells.

28 **Key words**

29 Ocean Acidification, Mediterranean, calcification, pteropods

30

31 **1. Introduction**

32 Marine ecosystems are increasingly influenced by decreasing seawater pH and carbonate
33 chemistry changes resulting from oceanic absorption of anthropogenic CO₂, a process now
34 well known as Ocean Acidification (OA) (Feely et al., 2004). Calcifying organisms are
35 particularly susceptible to OA because perturbations in the seawater carbonate system,
36 including changes in H⁺ and CO_{2(aq)}, can reduce their ability to synthesize and/or maintain
37 calcium carbonate skeletons and shells. In efforts to understand the implications of these
38 changes on marine organisms, shallow submarine volcanic CO₂ vents have been identified as
39 useful analogues for studying the prospective impacts of Ocean Acidification on marine
40 ecosystems (Hall-Spenser et al., 2008) since the water surrounding the CO₂ vent naturally
41 lowers the pH of the water column (Williams et al., 1992).

42 Identifying the natural response of marine organisms to OA is a difficult task in laboratory
43 conditions since the behaviour of the organism is constrained and the feeding environment is
44 poorly simulated (Howes et al., 2015). However, the combination of laboratory experiments
45 with the assessment of naturally acidified environmental gradients (such as CO₂ vent
46 environments), can provide further insights into the threshold pH affecting the performance
47 of vulnerable marine species (Basso et al., 2015). Volcanic CO₂ vents have been widely used
48 as a proxy for future OA conditions by numerous authors showing the negative response of
49 the higher pCO₂ conditions to which benthic organisms have commonly been exposed for
50 their entire life span (i.e. Ricevuto et al., 2012; Milazzo et al., 2014; Langer et al., 2014).

51 Marine volcanic CO₂ vents are abundant in the Mediterranean Sea, especially around Italy
52 (Dando et al., 1999). Recent studies in the Gulf of Naples (Tyrrhenian Sea, Italy), on the
53 impact of CO₂ vents on marine benthic organisms inhabiting shallow coastal waters, showed

54 a shift from benthic calcareous communities to communities lacking scleractinian coral (Hall-
55 Spenser et al., 2008). Furthermore, settlement and colonization by mollusks and microfauna
56 decreased at the acidified stations (Ricevuto et al., 2012; Milazzo et al., 2014). In the same
57 region, the natural pH gradient negatively affected the growth and survival in bivalves *Pinna*
58 *nobilis* (Ricevuto et al., 2012) while the patellogastropod limpet *Patella caerulea* was able to
59 counteract the low pH induced shell corrosion by the addition of aragonitic shell layers
60 (Langer et al., 2014).

61 With specific reference to the hydrological features, the Gulf of Naples is characterised by
62 the presence of two main water masses typical of the southern Tyrrhenian Sea: the Modified
63 Atlantic Water (MAW) and the Levantine Intermediate Water (LIW) (Uttieri et al., 2011).
64 Even if in the study area the water masses are essentially the same as for the southern
65 Tyrrhenian Sea, the presence of CO₂ submarine emissions alters the carbonate chemistry
66 nature of the water masses. The presence of natural submarine gas emissions was suggested
67 by (Sacchi et al., 2005). More recently (Passaro et al., 2014, 2016), detected and mapped the
68 gas discharge (dominated by CO₂) at the seafloor of the Gulf of Naples and suggested that the
69 occurrence of CO₂ vents in this area could be linked to the interaction between volcanic
70 related seafloor morphologies and the main, North East striking faults present in the area,
71 (i.e., Vesuvian fault).

72 However, all the CO₂ vent related studies have been mainly focused on the response of the
73 coastal benthic ecosystem, while the impact of these natural pH gradients on the planktonic
74 calcifying population has not been explored. Unlike sessile benthic organisms, pelagic
75 species can move in and out of waters surrounding the CO₂ vents and experience a
76 pronounced variability of pCO₂ conditions over time. This mobility makes it difficult to
77 quantify the exposure of pelagic organisms to high pCO₂ levels. However, a recent study on

78 corals found that repetitive exposure to high pCO₂ conditions may cause greater responses
79 within certain organisms than exposure to static OA (Roleda et al., 2015).

80 Euthecosome pteropods (planktonic shelled gastropods) have been identified as indicator for
81 OA (OSPAR/ICES advisor group, 2015); as their thin shells are made of aragonite, a
82 metastable form of biogenic CaCO₃ (Mucci, 1983), shelled pteropods are extremely sensitive
83 to changes in marine carbonate chemistry. These organisms have been widely studied for OA
84 effects, both in simulated OA conditions in the lab and in the field where high pCO₂ levels
85 already occur. Short-term lab experiments (up to a month), examining the impact of exposure
86 to high pCO₂, document pteropod shell dissolution, lowered shell calcification, altered
87 metabolism, behavior, gene expression and decreased survivorship (i.e. Manno et al., 2007;
88 Comeau et al., 2010; Lischka and Riebesell, 2012; Moya et al., 2016). In the field, changes in
89 pteropod species community composition, geographical distribution and presence of shell
90 dissolution have been observed as a result of co-variation of natural high CO₂ and low
91 dissolved oxygen across a frontal system in the Southern California Current (Bednaršek et al.,
92 2014; 2015) and within an upwelling region in the Scotia Sea (Bednaršek et al., 2012b). Maas
93 et al. (2016) suggested that natural environmental exposure to low pH and oxygen influences
94 pteropod metabolic sensitivity in the Oxygen Minimum Zone in the North Atlantic.

95 Here we present our observations of pteropods collected around the CO₂ vent region in the
96 Gulf of Naples (Tyrrhenian Sea). We aim to assess the condition of pteropod shells (in terms
97 of biomass and dissolution) to episodic exposure to high pCO₂ in the presence of volcanic
98 CO₂ vents. We focus on the species *Creseis conica* (*C. conica*) which are common and
99 distributed in tropical and subtropical water masses worldwide.

100 This study documents, for the first time, the impact of natural CO₂ volcanic emissions on live
101 pteropods extracted directly from the natural environment. In particular the present work adds
102 new insight to the in situ response of pteropods *C. conica* to recurrent exposure to critical

103 carbonate chemistry environments. This study also highlights the importance of including
104 CO₂ vent regions within a long term monitoring program to investigate the potential ability of
105 pteropods to persist in a high CO₂ ocean.

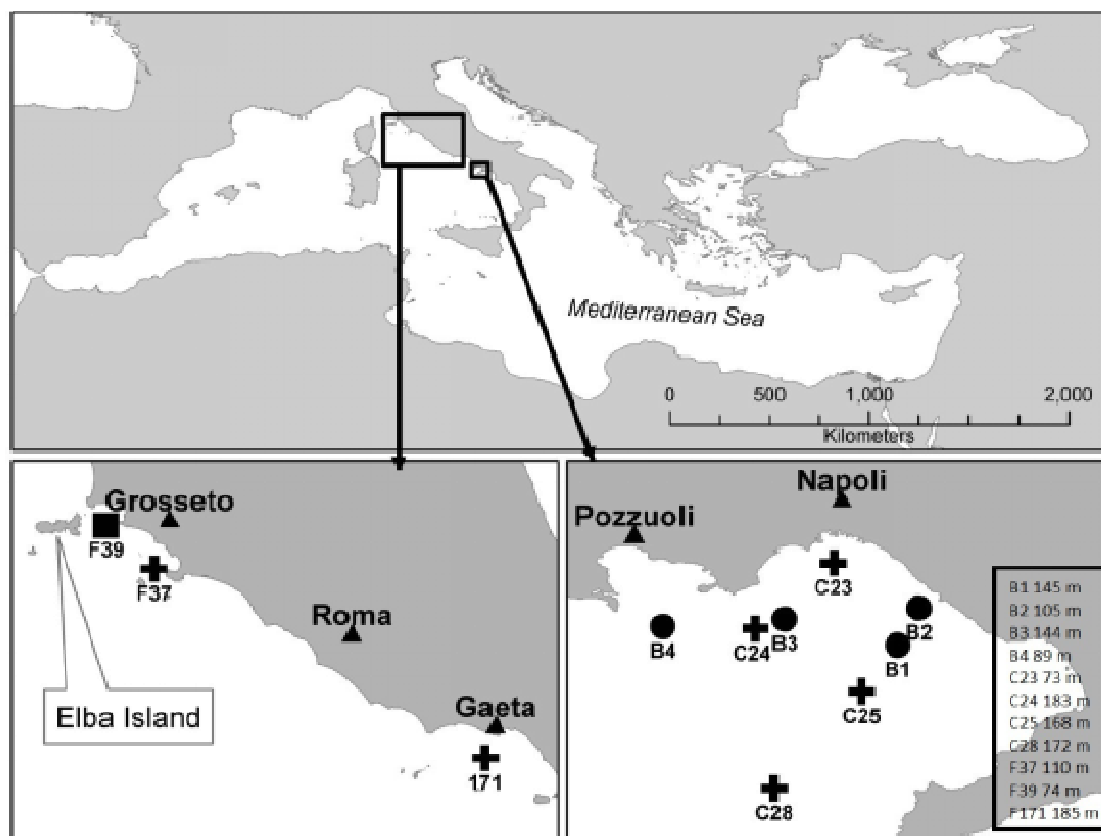
106 **2. Methods**

107

108 **2.1 Study region**

109 This study was performed within the framework of the Medias (Mediterranean International
110 Acoustic Survey) project in the Tyrrhenian and Ligurian seas. All the samples and data were
111 collected on August 2015 during an oceanographic cruise in the Tyrrhenian Sea on board of
112 the R/V “G. Dallaporta”. A total of 8 stations were sampled in the Gulf of Naples
113 characterized by on site (4 stations, group “B”) and nearby (4 stations, group “C”) presence
114 of natural submarine volcanic CO₂ emissions. Since currents inside the gulf will likely drive
115 the pteropods in (B stations) and out (C stations) the CO₂ vent zones, we assume that those
116 organisms will be periodically exposed to the vents during their life. In addition, more
117 stations (3 stations, group “F”) were sampled outside of the Gulf of Naples, in the northern
118 Tyrrhenian Sea (Fig 1), where no CO₂ vents have been identified, to provide a control suite of
119 samples (Fig 1).

120 Stations characterized by natural gas emissions were identified during a previous
121 oceanographic survey in the same area (Passaro et al., 2014) by means of the Simrad EK60
122 Scientific Echosounder. Such instrumentation is typically used for estimating biomass and
123 distributions of small pelagic fish species in many areas of the Mediterranean Sea (Bonanno
124 et al., 2014) but also readily identify plumes of bubbles derived from CO₂ vents at the
125 seafloor.



126

127

128 **Fig 1** Sampling station positions in the Northern Tyrrhenian Sea and in the Gulf of
 129 **Naples (Mediterranean Sea)**. Stations B1-B2-B3-B4 are characterized by the presence of
 130 natural submarine volcanic CO₂ emissions. Each station depth is indicated. See Passaro et al.,
 131 2014; 2016 for a detailed map of CO₂ vents emission points.

132 **2.2 Hydrology and Carbonate Chemistry measurements**

133 Full depth hydrological casts were acquired across all the stations using an SBE 9/11 Plus
 134 CTD, with temperature, oxygen, conductivity and fluorometer sensors. The probes were
 135 calibrated before the cruise at Sea-Bird Electronics in Kempton, Germany. The collected
 136 downcast data were quality-checked and processed using the Seasoft-Win32 software. The
 137 overall accuracies are within 0.001°C for temperature, 0.001 sm⁻¹ for conductivity, and
 138 0.015% of full scale for pressure. Raw fluorescence values were converted to Chl a biomass
 139 (µg*l⁻¹) using the factory calibration.

140 Discrete Total Alkalinity (TA) and Dissolved Inorganic Carbon (DIC) samples were
141 collected at different depths of the water column using a carousel equipped with Niskin
142 bottles and then poisoned with HgCl_2 (2% saturated solution) to prevent biological alteration.
143 Seawater TA and DIC were measured by potentiometer titration, employing the open-cell
144 procedure. The precision for TA was $\pm 2.0 \text{ mmol kg}^{-1}$ and 4 mmol kg^{-1} for DIC. Data accuracy
145 was confirmed by regular analyses of Certified Reference Materials (Scripps Institution of
146 Oceanography). Carbonate saturation states of aragonite (Ω_{ar}) were indirectly calculated
147 from TA and DIC data using the CO2SYS software (Lewis and Wallace, 1998), with
148 carbonate dissociation constants by (Mehrbach et al., 1973) refitted by (Dickson et al., 1997)
149 and sulfate dissociation constants by (Dickson et al., 1990). Note that for logistical reasons no
150 chemistry samples, were collected at station F39.

151 **2.3 Pteropod collection and investigation**

152 On board, living pteropods were collected from near bottom depth (ranging between 65 m
153 and 170 m) to the surface by a Bongo-40 zooplankton net (200 μm mesh size). Sampling took
154 place over one time at each station during the day time. The volume of sea water sampled was
155 measured by General Oceanics mechanical flow-meters attached to the ring net. Samples
156 were stored for 3 weeks within buffered formalin solution and kept at 4°C . pH was measure
157 in all the samples, at the beginning and the end of the storing period to ensure that the state of
158 the shells were not affected by the preservation technique. After three weeks from the
159 collection, pteropod species were identified and counted using a light microscope Olympus
160 SZX16. Pteropod abundance within the water column was calculated as individuals per cubic
161 meter ($\text{Ind} \cdot \text{m}^{-3}$).

162 Investigation of shell morphology and shell biomass was determined only for the dominant
163 pteropod species *C. conica*. Shell morphology was performed using a Scanning Electron
164 Microscope (SEM). The number of individuals analysed for SEM ranged between 10 or 20

165 for each station (except for station B2 where we analysed only 5 organisms) depending on the
166 availability of specimens. Only individuals with similar shell size (juveniles ranging between
167 280 μm -320 μm) were selected to facilitate comparison between different groups assuming
168 same life stage has similar susceptibility to high pCO_2 level. Before SEM imaging,
169 individuals were carefully washed with DI water to remove salt on the shell and then air-
170 dried for 24h. The shells outer organic layer (periostracum) was not removed. We
171 acknowledge that the exclusion (Bednaršek et al. 2016) or inclusion (Peck et al., 2016b) of
172 periostracum for evaluate shell dissolution is still in debate. However our rationale for not
173 removing the periostracum prior to imaging shells follows previous studies (Peck et al., 2015)
174 showing that the removal of the organic outer layer, which also has an intra-crystalline matrix
175 (Marin et al., 1996), can expose crystals in a way which could be mis-interpreted as shell
176 dissolution.

177 Shell degradation was evaluated by applying a semi-quantitative index of dissolution
178 (Gerhardt et al., 2001; Lischka et al., 2011; Manno et al., 2012). This Dissolution Index is
179 represented by six preservation stages (from 0 = best preservation to 5 = highest degree of
180 dissolution), determined by: shell surface lustre (whether lustrous or dull); shell damage
181 (surface with shell corrosion and/or perforation of at least one layer of aragonite). For each
182 station, we calculated the % of shell falling in four dissolution levels: no corrosion
183 (transparency, preservation stage 0); low corrosion (opacity with small sign of dissolution,
184 preservation stage 1-2); high corrosion (periostracum and the first aragonite prismatic are
185 partially missing, preservation stage 3-4); damage (presence of perforation, preservation stage
186 5).

187 Shell surface was inspected for the presence or absence of fracture zone (i.e., resulting from
188 in situ mechanical damage) and represented as % shells presenting fractures to the total
189 shells. To discriminate between “natural fractures” and fractures due to mechanical damage

190 from the net and collection processing, we only considered the “historical fractures” where it
191 appears that the animal has built up shell material to weld the shell back together (Peck et al.
192 2018).

193 For the measurement of shell biomass (carbonate content expressed as $\mu\text{g CaCO}_3$),
194 individuals were heated to 550°C for 5 h to eliminate organic matter content and the ashes
195 (representing the remains of the shells) weighed using a Toledo microbalance. The ash
196 weight can be considered an indirect estimate of CaCO_3 content. As for SEM investigation, to
197 allow us to estimate shell biomass difference between groups, we only used individuals with
198 similar shell diameter (juveniles, $302\mu\text{m} \pm 11$, for a total of 76 specimens) and presenting the
199 best shell condition within each group (31, F; 30 C; 15, B specimens).

200 **2.4 Data analysis**

201 For each station, the average values of carbonate chemistry parameters (pH, TA, Ω_{ar} , DIC,
202 pCO_2) were computed together with total abundance, shell biomass, shell dissolution level,
203 and percentage of fractured shells. Temperature, salinity, Chl a and oxygen values recorded at
204 the same depth of carbonate chemistry measurements were extracted from CTD profiles to
205 obtain the average hydrological conditions at each station. Obtained data matrix was then
206 used in the statistical analysis. The pairwise correlation between all above-mentioned factors
207 was computed by using Spearman correlation coefficient. PCA was used to investigate the
208 presence of pattern of variables (that could be interpreted as “processes”) as well as to best
209 explain the variation observed among stations. The differences among the identified groups
210 of stations were assessed with parametric statistical tests (namely ANOVA and t-test
211 according to the numbers of groups). If serious violations in the assumption required by
212 parametric tests were identified, the non-parametric alternative were used (Kruskal-Wallis
213 ANOVA and Wilcoxon rank sum test).

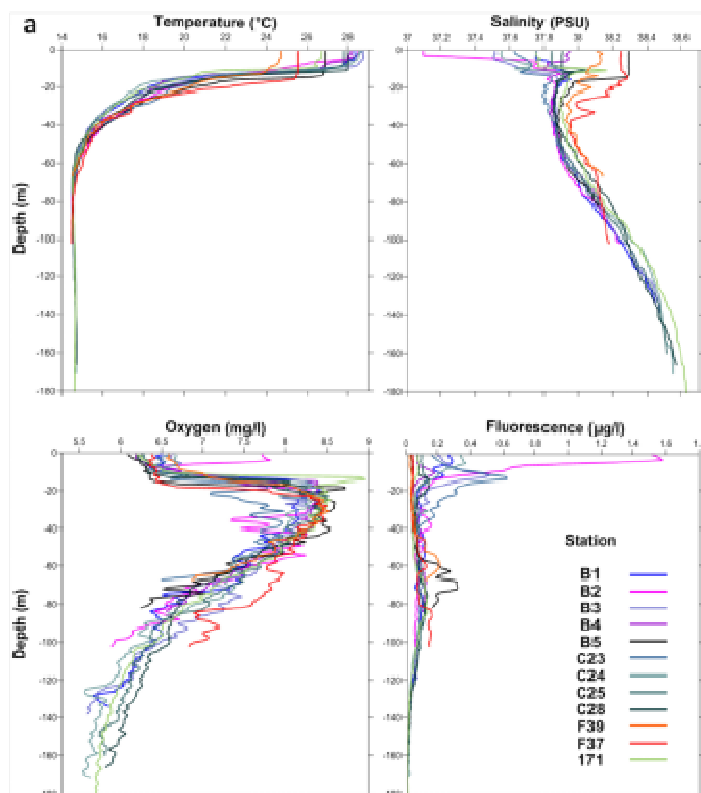
214 Shannon diversity index was used to characterize the pteropods biodiversity in the stations.

215 All statistical analysis were carried out in R statistical environment (R Core Team, 2018).

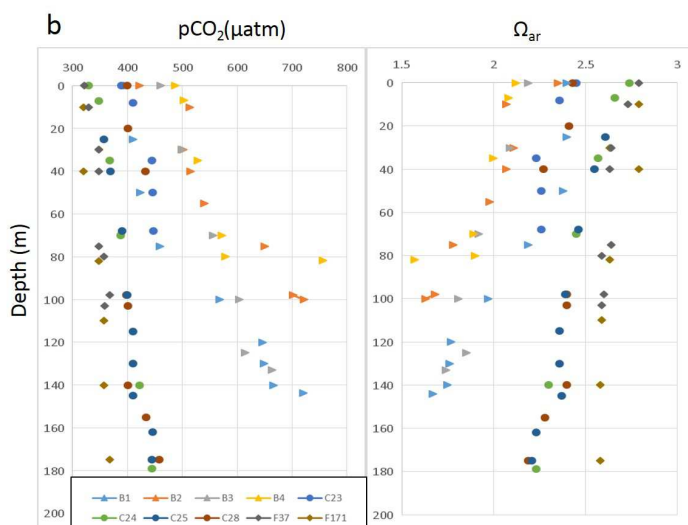
216 **3. RESULTS**

217 **3.1 Hydrology profiles**

218 At all stations mean surface and bottom temperature ranged between 25.6 °C and 28.2 °C
219 and between 14.45 °C and 14.67 °C respectively (Fig 2a). Surface salinity was strongly
220 influenced by the river outflow with values ranging from 37.09 to 38.25. The salinity
221 minimum due to the presence of the Modified Atlantic Water (MAW) was typically
222 positioned between 30 and 45 m (Fig 2a). Oxygen concentration exhibited a similar profile at
223 all stations except B2 (Fig 2a) where higher surface oxygen values were mainly influenced by
224 the Sarno river outflow. Fluorescence values ranged between 0.01 and 1.58 $\mu\text{g}\cdot\text{l}^{-1}$. The
225 higher values were recorded in the Gulf of Naples and in particular in the B2, B3 and C23
226 stations (Fig. 2a). Dataset of the hydrological parameters is available in Table 1 in S1_Table.



227



228

229 **Fig 2 Vertical hydrological and carbonate chemistry profiles.** a) Temperature, salinity,
 230 oxygen ($\text{mg}\cdot\text{l}^{-1}$), Chl a ($\mu\text{g}\cdot\text{l}^{-1}$) and b) pCO_2 (μatm) and Ω_{ar} (aragonite saturation state) at
 231 the sampling stations (F control, C nearby vents and B vents station).

232 3.2 Carbonate chemistry

233 Significant differences were recorded among the three groups of stations for both Ω_{ar}
 234 (ANOVA, $F(2,7)=101.4$, $p<0.001$) and pCO_2 (ANOVA, $F(2,7)=240.7$, $p<0.05$). In particular,

235 although seawater was not under-saturated with respect to the aragonite at any of the stations
236 (i.e., $\Omega_{ar}>1$ at all stations), Ω_{ar} and pCO_2 values in the Gulf of Naples (stations B and C) were
237 respectively significantly lower and higher than at control site (stations F). Differences in
238 carbonate chemistry were also evaluated by grouping the B and C stations (Gulf of Naples)
239 and comparing such group with the stations outside the gulf (control). Obtained results
240 showed that stations outside the gulf were significantly different from the B+C group
241 ($\Omega_{ar}; t_{(7)}=5.78; p<0.05$ and $pCO_2; t_{(7)}=4.7, p<0.05$).

242 Dataset of carbonate chemistry is available in Table 2 in S1_Table.

243 **3.3 Difference in pteropod abundance and “shell fitness” between** 244 **stations**

245 Pteropod abundance was significantly different between the three groups of stations (K-W
246 ANOVA, $H_{(2)}= 7.13, p<0.05$). In particular, pteropod abundance was significantly lower at
247 group C (40%) and B (82%) stations than the control stations, group F.

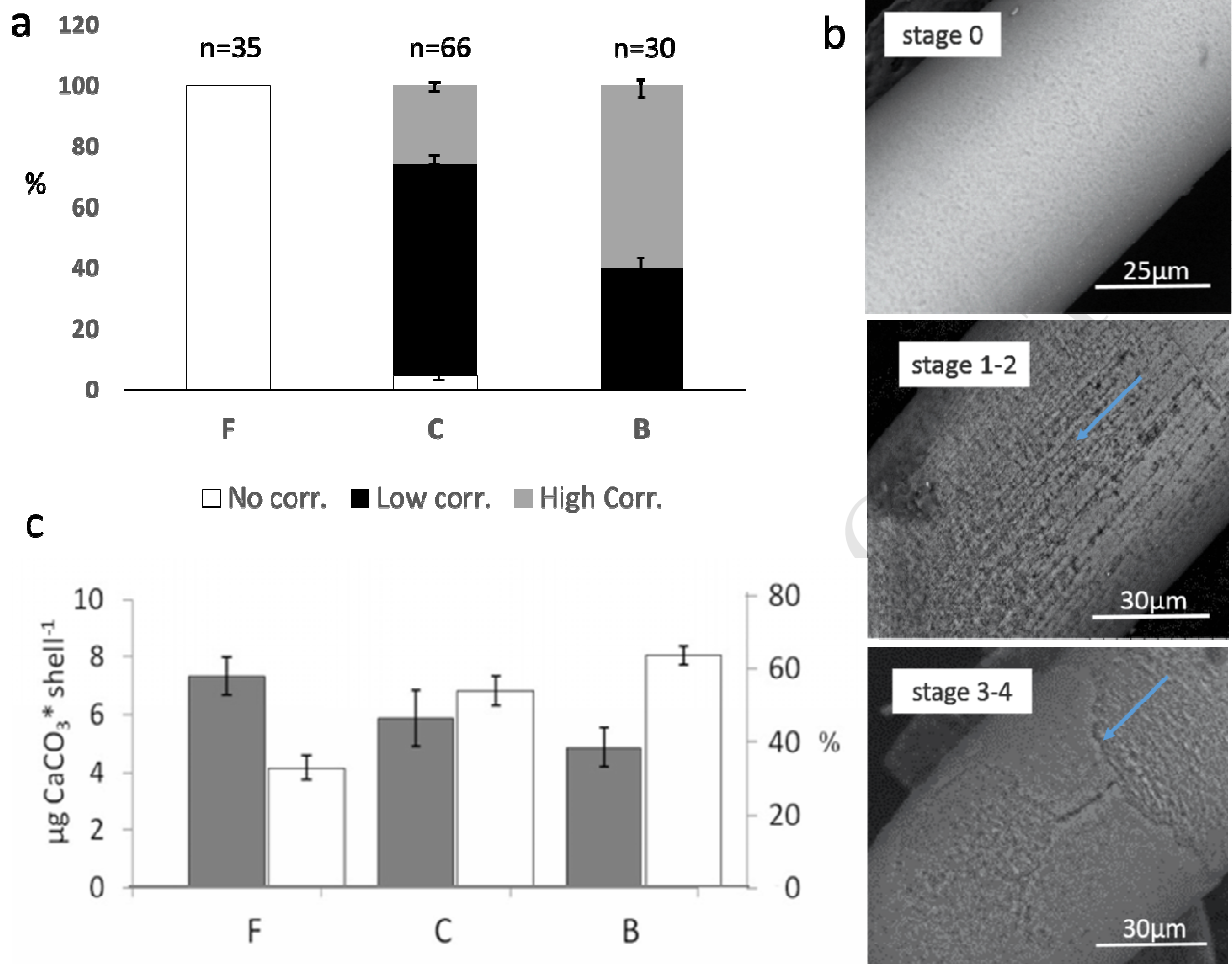
248 Pteropod diversity was significantly different (Shannon diversity index, (K-W ANOVA, $H_{(2)}=$
249 $6.76, p<0.05$) between the three groups of stations also. In particular, diversity was
250 significantly higher at the control stations outside the Gulf of Naples (group F, 100% of
251 identified species) than in the stations of the groups B and C. Dataset of pteropod relative
252 abundance is available in Table 3 in S1_Table.

253 The state of *C. conica* shell condition is presented in (Fig 3a). ANOVA test showed that the
254 percentage of shells presenting no signs of corrosion, low corrosion and high corrosion were
255 significant different among the considered groups (No Corr: $F(2,8)=9284, p<0.05$; Low Corr:
256 K-w: $H(2)=9.07, p<0.05$; High Corr: $F(2,8)=102.6, p<0.05$). Pteropods collected from the
257 CO_2 vent stations (group B) presented a significantly higher degree of dissolution than
258 pteropods collected from stations C and F. In particular all pteropods collected in group F had

259 a well preserved and transparent shells (stage 0). Conversely, within group B, 60% of shells
260 showed stage 4 levels of dissolution. Shell dissolution (even if moderate) was also observed
261 in the group C, with 70% of shells exhibiting opacity and dullness (stage 3). SEM pictures in
262 the Fig 3b are representative of the different *C. conica* shell dissolution stages observed. We
263 did not observe evidence of shell perforation (stage 5) in any specimens.

264 Significant differences among the three groups were evidenced also in terms of incidence of
265 shell fractures ($F(2, 8)=51, p<0.001$) and biomass. In particular, at stations within group B
266 the highest incidence of shell fractures and the lowest biomass was recorded. Significant
267 differences were also recorded between C and F stations, the latter presenting the lowest
268 incidence of shell fractures and the highest biomass. Dataset of pteropod shell biomass,
269 fractures and dissolution are available in Table 4, 5 and 6 in S1_Table.

270 Comparing the B+C stations against the F group, the presence of significant differences
271 between the Gulf of Naples (B+C) and the Control station (F) were confirmed (No Corr:
272 $t_{(7)}=103.97, p<0.05$; Low Corr: $t_{(7)}=9.26, p<0.05$; High Corr: $t_{(7)}=6.32, p<0.05$; incidence of
273 shell fractures: $t_{(7)}=7.8, p<0.05$; biomass: $t_{(7)}=7.35, p<0.05$).



274

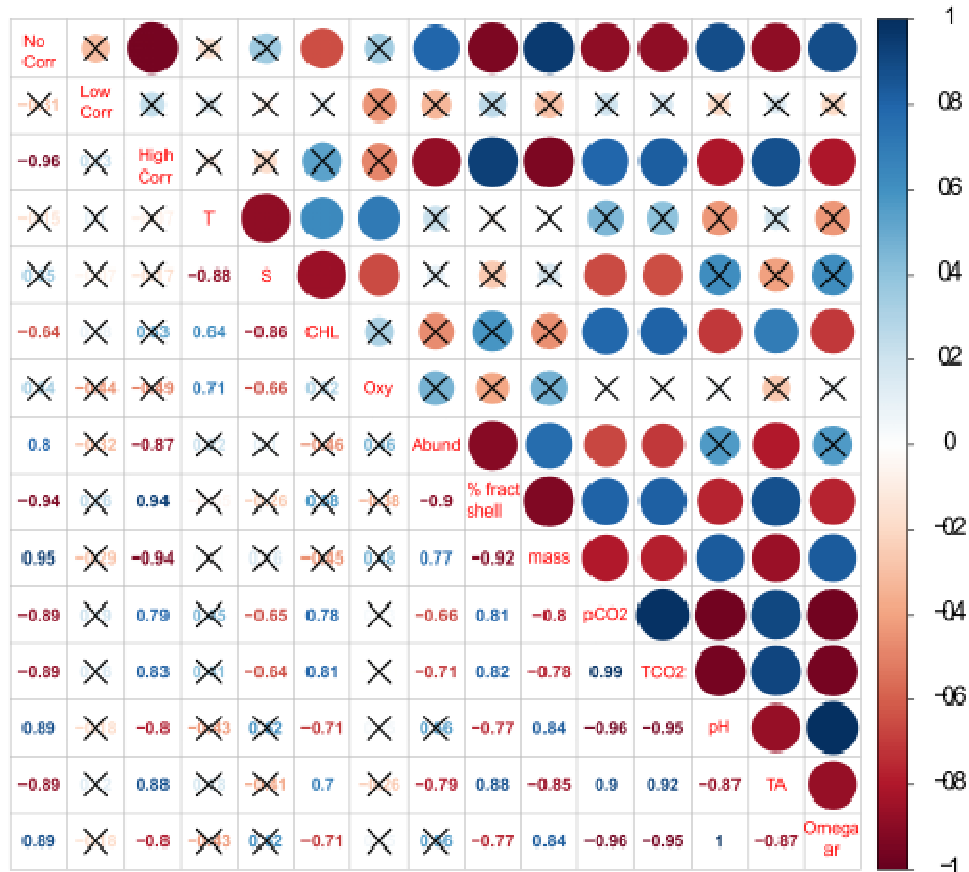
275 **Fig 3 Difference in pteropod abundance and “shell fitness” between stations** a) Shell
 276 dissolution level (%) of *C. conica* collected at the group B (vent stations), C and station F; b)
 277 SEM images showing different levels of dissolution for *C. conica* shells. The top image
 278 shows a detail of *C. conica* shell in perfect condition (stage 0, mainly found in the group F);
 279 in the middle *C. conica* shell lustreless with sign of dissolution (stage 2-3, mainly found in
 280 group C and B); on the bottom *C. conica* shell with high dissolution where the periostracum
 281 and the first aragonite prismatic are partially missing (stage 4, mainly found in the group B);
 282 c) shell biomass (grey histogram, $\mu\text{g CaCO}_3 \cdot \text{shell}^{-1}$) and shell presenting fractures (white
 283 histogram, %) of *C. conica*

284 **3.4 Relationships with carbonate chemistry and hydrology**

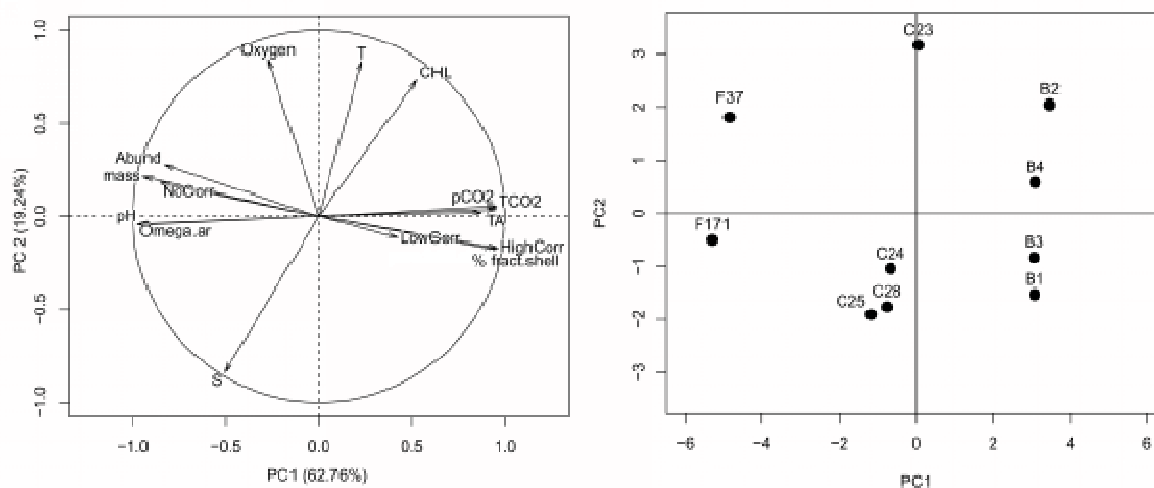
285 The pairwise correlation analysis (Fig 4a) showed the presence of strong correlations between
286 carbonate chemistry and some parameters related to the condition of pteropods (such as the
287 lowest and the highest dissolution levels, the biomass and the percentage of shell fractures).
288 In particular, the absence of dissolution was positively related to high Ω_{ar} values. Conversely,
289 the highest level of corrosion was negatively related to Ω_{ar} . The abundance and the shell
290 biomass were found positively correlated with Ω_{ar} while the opposite was true for the
291 percentage of shell presenting fractures. No significant differences were found among the
292 three groups of stations in terms of temperature, salinity and oxygen, evidencing the presence
293 of comparable hydrological conditions..

294 PCA analysis further confirm the relationships observed in correlation analysis, providing a
295 more clear picture of the factors driving differences among the three groups of stations (Fig.
296 4b,c). The first two PCA axis explained 82% of the total variance. In particular, the 1st PC
297 axis was significantly (see Table 7 in S1_Table) related to Ω_{ar} , and pCO₂, absence of
298 corrosion (No Corr) and higher corrosion, as well as to abundance, percentage of shell with
299 fractures and biomass. Such patterns evidenced that stations having lower values on the first
300 PCA axis were characterized by higher abundance, biomass, Ω_{ar} and lower shell dissolution
301 (No Corr.) as well as by lower pCO₂, percentage of shell with fractures and lower proportion
302 of pteropods shell characterized by higher degree of corrosion. As the 1st axis accounts for
303 62.76% of the total variance it is clear that most of the variability among stations is related to
304 pteropods and carbonate chemistry parameters. In this context all the B stations were
305 clustered on the right side of the 1st PC axis, while the F stations showed the lowest values
306 with respect to such axis. C stations were mainly found in intermediate position along the 1st
307 PC axis evidencing the presence of intermediate conditions between B and F stations in terms
308 of the parameters related to the 1st PC. Regarding the 2nd PC axis, it was found strongly
309 related only to the hydrological parameters. Also, as it accounts for a much lower proportion

310 of the total variance, the weak effect of hydrological condition in driving the differences
 311 among the stations was confirmed. Dispersion of stations along the 2nd PC axis is much lower
 312 than the one along the 1st PC, and the observed differences along the 2nd PC are mainly due to
 313 local factors, such as distance from the coast (leading to higher Chl a) or the presence of
 314 freshwaters input (it is the case of C23 and F37 stations).



315
 316 **Fig. 4a: Pairwise Spearman correlation plot among the considered variables.** Correlation
 317 values are reported in the lower triangular matrix. In the upper triangular matrix a graphical
 318 representation of correlation values is reported (higher the correlation, bigger the circle; blue
 319 and red colours indicate positive and negative correlations respectively). The "X" symbol is
 320 used to mark the non-significant ($p > 0.05$) correlations.. Note: no corrosion=% shell
 321 presenting preservation stage 0; low corrosion=% shell presenting preservation stage 1-2;
 322 high corrosion = % shell presenting preservation stage 3-4; DIC =TCO₂.



323

324 **Fig 4b: PCA of the considered variables.** Visual representation of the correlation among

325 environmental, chemistry, biological factors variables and PCs (left panel) and distribution

326 of the stations along the 1th and 2nd PCs space (right panel). Note corr = corrosion;327 TCO₂=DIC328 **4. DISCUSSION**329 **4.1 Pteropod shell fitness around CO₂ vents**330 This study documents, for the first time, the impact of natural CO₂ volcanic emissions on live

331 pteropods extracted directly from the natural environment. We illustrated that the decrease of

332 Ω_{ar} , associated with the presence of CO₂ vents, can alter the chemical environment for333 planktonic calcifying organisms in the vicinity. In particular, *in situ* shell dissolution and

334 change in shell biomass were the predominant features observed in the live pteropods

335 collected in the Gulf of Naples (in the station located within and nearby the CO₂ vent

336 discharge) compare to pteropods collected in the control stations. Unfortunately, so far there

337 are no studies on seasonal variability of the carbonate chemistry in this region, however,

338 the pH difference between the CO₂ vent stations and the controls is higher than the natural

339 seasonal variability of the Liguria coastal site (Howes et al., 2015), located on the border

340 with the Tyrrhenian Sea.

341 The difficulty in investigating pelagic organisms along a “natural gradient” is determining the
342 residence-times of populations within the CO₂ vent stations, so as to parameterise the
343 duration of their exposure to the stressor. However, pteropods may perform diel and/or
344 seasonal vertical migration, spending part of their time under low Ω_{ar} (nearby the CO₂ vent
345 source at the bottom) and part in the more saturated waters at the surface (Bednaršek et al.,
346 2012; Manno et al., 2016). In particular, Creseidae (such as *C. conica*) seems to perform diel
347 vertical migration (Be and Gilmer, 1977; Hsueh, 1995) with a vertical distribution >100 m
348 (van der Spoel, 1967; Be and Gilmer, 1977). Consequently, despite we do not have
349 information on the vertical distribution of pteropods at the time of collection, we can assume
350 that the organisms collected at the CO₂ vent stations (group B, emission depth ranging from
351 89m to 145m) will have been daily exposed to pCO₂ fluctuation.

352 Another challenge is that pteropods are also not static spatially and will likely move around
353 and outside the Gulf of Naples transported by currents. Two different water inflow and
354 outflow regimes are present in the Gulf of Naples, with a tendency towards stagnation inside
355 the basin during spring and summer and a more effective water renewal mechanism in fall
356 and winter under NE winds (Cianelli et al., 2015). In particular, Mazzocchi et al. (2012)
357 outlined that the only few species representative of the coastal area dominate the zooplankton
358 assemblage in summer owing to coastal retention (Cianelli et al., 2015). Thus, given a growth
359 rate of about 0.33 mm per month (Bednaršek et al., 2012; Well, 1976) and a mean diameter
360 of pteropod shell investigated in this study of 302 μ m, it is likely that specimens were retained
361 in the Gulf of Naples from the very beginning of their life cycle and within the same water
362 mass condition. As Bednaršek et al., 2012 and Well, 1976 reported similar values of shell
363 growth rates on pteropods collected from very different regions (i.e. Scotia Sea and West
364 India respectively), we assume that the used growth rate can be representative for pteropods
365 collected in the present study.

366 Uttieri et al., 2011 using a model simulations of particle transport (in the summer period)
367 demonstrate the presence of a scarce renewal of coastal waters, both over short (i.e., 48 h)
368 and long (i.e., 1 month) periods. The authors found that the residence times was very high,
369 with particles remaining in the deployment area on average for more than 15 days. This
370 simulation confirms that pteropods will spend a relevant amount of time in the station of
371 collection before to be moved back and forward around the Gulf. Thus, although we do not
372 have information about the resident time of pteropods in the Gulf of Naples, we can assume
373 that pteropods collected in August (this study) have likely been trapped in the Gulf and have
374 experienced intermittent CO₂ vent impact for months. The presence of impacted pteropod
375 shell in the stations (group C) not directly located on the CO₂ vent discharge, confirm the role
376 of currents within the Gulf of Naples, driving the pelagic calcifiers inside and outside the CO₂
377 vent emissions. Conversely, sessile benthic calcifiers (as for result of their sedentary
378 behaviour) experience shell degradation only when directly located around the CO₂ vents
379 (i.e., Hall-Spenser et al., 2014, Basso et al., 2015; Milazzo et al. 2014).

380 The variability nature of the CO₂ vent system over the time is a key factor in the
381 interpretation of the observed negative impact on pteropods shell and in part explains the high
382 level of shell dissolution despite the presence of oversaturated seawater ($\Omega_{ar}>1$). We are
383 aware that our data are not representative of the carbonate chemistry condition over time and
384 a detailed survey throughout the year will be an important next step. However, Passaro et al.
385 (2016) found that in the Gulf of Naples bubble plumes generated at the CO₂ vent are
386 highly variable: from a continuous, dense bubble-flux to short-lived phenomena. In
387 particular, the authors found the pH values above a shallow CO₂ vent emission (75 m
388 depth) decrease from 8.4 (at 70 m depth) to 7.8 (at the bottom). Unfortunately, the authors
389 did not provide Ω_{ar} values but the pH values at the bottom are lower than the pH we
390 observed near the CO₂ vent emissions and it could likely correspond to lower Ω_{ar} than the

391 values observed in our study. Therefore, periodical exposition to critical low Ω_{ar} values may
392 drive the dissolution state of pteropod shells.

393 Overall, we suggest that pteropods around the CO₂ vents in the Gulf of Naples, are
394 negatively impacted when periodically exposed to high spatial and temporal variability in
395 Ω_{ar} . Evidence of impact on pteropod shell dissolution has already been reported in the field.
396 Within an upwelling system, Bednaršek et al. (2014), observed higher levels of shell
397 dissolution (up to preservation stage 5) than the present study. This can reflect either the
398 higher magnitude as well as time of exposure of pteropods in this region compared to the
399 Gulf of Naples. Further, the absence of additive environmental stressors in the Gulf of Naples
400 such as variability in oxygen and nutrient concentration could also partially explain the lower
401 impact on the shell compared to the upwelling system.

402 In the future targeted research, focused on the investigation of vertical distribution and
403 migration of pteropods in the CO₂ vent regions, will be crucial to improve our understanding
404 on the potential ability of these organisms to avoid water depths with critical carbonate
405 saturation state. It will be important to use Lagrangian modelling studies (to track pteropods
406 across temporal and spatial scales) since in addition to intensity and duration of exposure
407 (Manno et al., 2012; 2016), the impact of CO₂ vents on pteropods is likely to be also a
408 function of the recovery time between the exposures itself. Lagrangian particle tracking
409 models coupled with hydrodynamic models are particularly efficient tools to examine the role
410 played by various physical processes and to study transport processes over an entire basin to
411 simulate zooplankton dispersion and distribution at different scales (e.g. Speirs et al., 2006;
412 Lett et al., 2007).

413 **4.2 Impact on the pteropod shell biomass**

414 We found that shell biomass was significantly lower in pteropods living within the Gulf of
415 Naples compared to those in the Northern Tyrrhenian Sea. Only individuals of the similar

416 length (juveniles, $302 \mu\text{m} \pm 11$) were used to measure shell biomass in order to compare the
417 different groups. The decrease in shell biomass suggests that calcification was lower than
418 dissolution and in turn the shell biomass decrease. This was more the case of the individuals
419 presenting high level of shell dissolution (group B) where likely the dissolution exceed the
420 calcification. However, shell biomass of individuals in the group C, which present manly
421 shells with opacity and/or low level of dissolution was still significantly lower, suggesting
422 that the lower shell biomass was a common features of pteropods in the Gulf of Naples
423 (compared to the shell biomass values of the control stations outside the Gulf). We are aware
424 that other environmental factors can play a role in pteropods shell growth (e.g. temperature
425 and salinity) (Lalli and Gilmer, 1989), however differences in salinity and temperature
426 between the stations during the summer were within the natural seasonal variability of the
427 Tyrrhenian Sea and well within the pteropods' tolerance window (Lalli and Gilmer, 1989).
428 Further, incubation experiment of pteropods under a range of salinity (Manno et al., 2012)
429 and temperature (Lischka and Riebsell, 2012) show that those parameters have to change
430 quite considerably before a negative effect is detectable (i.e., shell growth, behaviour,
431 survival). Similarly, the potential role of temperature on shell dissolution was excluded
432 because previous works found that under manipulate water condition increasing in
433 temperature not leads to dissolution on pteropods (Lischka et al. 2011, Gardner et al. 2018).
434 Food availability may also play a critical role in determining the shell growth because food
435 supply is required to support the metabolic processes facilitating bio-calcification as well as
436 the resistance of calcifiers to adverse condition such as OA (Ramajo et al., 2016). Particulate
437 food availability to pteropods, as inferred indirectly from average Chl a fluorescence in each
438 station was not significantly different between the three groups of stations, suggesting
439 pteropods were not limited by food availability in the region around the CO₂ vents.

440 Evidence of change in shell morphology in response to change in carbonate chemistry
441 associated with shallow-water CO₂ vents has already been observed in benthic molluscs (i.e.,
442 Langer et al., 2014, Garilli et al., 2015). Garilli et al. (2015) show that benthic gastropod
443 species (*Cyclope neritea* and *Nassarius corniculus*) adapted to acidified seawater ($\Omega_{\text{ar}}=0.68$)
444 were smaller than those found in normal pH conditions (8.1) while Langer et al., (2014)
445 found that the patellogastropod limpet *Patella caerulea* counteracted the induced shell
446 dissolution in the CO₂ vent waters ($\Omega_{\text{ar}}=3.01$) by enhanced production of internal aragonite
447 shell layers. Incubation experiments on the Mediterranean pteropod, *Creseis acicula*, reported
448 a 30% decrease in calcification with a decrease in Ω_{ar} from 3.3 to 2.0 (Comeau et al., 2012).
449 Moya et al. (2016) show that pteropod *Heliconoides inflatus* exhibited a 50% decrease in
450 gross calcification when exposed to waters of $\Omega_{\text{ar}} = 2$ (compare to control condition $\Omega_{\text{ar}} =$
451 2.9).

452 Our results provide *in situ* evidence that shifts away from an organisms optimum Ω_{ar} values
453 can significantly affect calcification despite waters remaining oversaturated. In support our
454 observation, pteropod shells collected within sediment traps became significantly lighter over
455 recent decades as Ω_{ar} decreased (Robert et al., 2011). A decrease in the shell thickness of
456 modern (2000+, $\Omega_{\text{ar}}=4.0$) tropical pteropod *D. longirostris* compared to 1960s ($\Omega_{\text{ar}}=3.5$)
457 samples has been observed (Roger et al., 2012). Further, Howes et al. (2017) compared the
458 difference in shell thickness of pteropod samples (*Cavolinia inflexa* and *Styliola subula*)
459 collected in the Tyrrhenian Sea with archived samples from 1910's. The authors observed
460 that shell thickness from modern pteropods ($\Omega_{\text{ar}}=3.4$) was significantly less than from
461 individuals collected on 1910's ($\Omega_{\text{ar}}=3.88$) (despite they state those decrease in shell
462 thickness should be treated with caution). Comparison with the present study and Howes et
463 al. (2017), both made in the Tyrrhenian Sea, highlights the relevance of using natural
464 environmental gradients to forecast the impact of high pCO₂ on marine organisms as spatial

465 change (natural variability of the carbonate chemistry, associated to CO₂ vents) can be a
466 substitute for time (100's older vs. modern samples, Howes et al., 2017). Further short time
467 experimental studies (up 29 days), where pteropods were incubated at undersaturated Ω_{ar}
468 levels, found a decrease of calcium carbonate precipitation and shell diameter, respectively
469 up 28% (Comeau et al., 2010) and 12 % (Lischka et al., 2012) confirming the relevance of
470 short episodic exposure in natural environments.

471 We observe an inverse relationship between shell biomass and the incidence of shells
472 presenting fractures, indicating that fractures are most commonly found in shells with low
473 biomass i.e., thinner/low density shells. Assuming predation pressure is comparable across all
474 sites, we consider that thin shells found at station B are more fragile and therefore more prone
475 to fracture than the more robust, high biomass shells. Although the effectiveness of the
476 periostracum for pteropods is a matter of debate (Peck et al., 2016b; Bednaršek et al. 2016).
477 Peck et al. (2016a) indicated that the shells of healthy, living pteropods are only susceptible
478 to dissolution of the shell where the periostracum has been breached and the aragonite
479 beneath is exposed to undersaturated waters. The susceptibility of the thin, fragile shells of
480 pteropods at the CO₂ vent stations to fracturing increases the incidence of aragonite being
481 exposed beneath the damaged periostracum. The consequence of increased incidence of
482 mechanical damage to the shell and exposure to undersaturated waters is consistent with our
483 observation of heightened incidence of shell dissolution.

484 In this study, *C. conica* (as well as the total pteropod assemblage) in the Gulf of Naples were
485 lower in abundance compared to those collected in the control stations (Northern Tyrrhenian
486 Sea). Due to the highly patchy distribution of pteropod abundance and sampling collection
487 being limited to one time, the impact of CO₂ vents on pteropod survivorship can only be
488 speculative and any interpretation have to be evaluated with extremely caution. However, it is
489 likely that the observed increasing in shell degradation and decreasing in shell biomass could

490 contribute to increasing in pteropod mortality (because affecting shell buoyancy, defence
491 against predator etc.). Bednaršek et al. (2014) also observed a relationship between shell
492 dissolution and decrease in pteropod abundance within the upwelling system and suggested
493 that increased dissolution combined with increased shell fragility could potentially induce
494 pteropod population decline.

495 Marine organisms have the potential to adapt to changes in ocean pH and adaptation
496 potential can be inferred from existing genetic diversity related to patterns of local adaptation
497 across present gradients in environmental pH (Stilmann and Paganini, 2015). Even if not
498 explored in this study, the decrease in shell biomass of pteropods as potential local adaptation
499 to natural low saturation state of CaCO_3 , is an interesting matter for future investigation. At
500 the high latitudes, for example, due to the natural lower saturation state of CaCO_3 , shell-
501 building materials are more difficult to extract from seawater and calcifying organisms
502 present thinner shells than individuals living at medium and low latitudes (Grauss et al.
503 1974). Understanding the persistence of populations of marine organisms in future altered
504 environments requires first an understanding of extant phenotypic plasticity under realistic
505 environmental conditions and the potential for adaptation (Stilmann and Paganini 2015). CO_2
506 vent regions might help to improve our understanding to predict if pteropod populations
507 possess adequate genetic variation to adapt to forecasted environmental change. Future long
508 term monitoring of the in situ population dynamics as well as study on phenotypic plasticity
509 and genetic variation across natural small scale gradients (such as CO_2 vent) will be crucial to
510 understanding the plasticity- adaptive-defence of this organism to persist in a more acidified
511 ocean over short (< 10 year) to medium (10–100 year) temporal scales.

512

513 **Appendix A. Supplementary data**

514 **S1 Table Hydrology and carbonate chemistry variables and pteropod dataset of all the**
515 **stations.** 1) Temperature, salinity, oxygen and fluorescence; 2) pH, Ω_{ar} , Total Alkalinity
516 (TA), Dissolved Inorganic ; 3) pteropods abundance and species contribution; 4) shell
517 biomass and length ; 5) shell fractures; 6) shell dissolution; 7) Correlation statistics

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Highlights

- 1- *in situ* shell dissolution and change in shell biomass were the predominant features observed in the live pteropods collected within and nearby CO₂ vent regions.
- 2- Low pteropod biomass shells (collected nearby the CO₂ vents) were more fragile and therefore more prone to fracture than the more robust, high biomass shells (collected in the control stations).
- 3- In the Gulf of Naples, intermittent shifts away from optimum Ω_{ar} values can significantly affect pteropod calcification despite waters remaining oversaturated.