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1 **Background Levels of Dioxin-like Polychlorinated Biphenyls (dlPCBs), Polychlorinated,**
2 **Polybrominated and Mixed Halogenated Dibenzop-dioxins and Dibenzofurans**
3 **(PCDD/Fs, PBDD/Fs & PXDD/Fs) in sera of Pregnant Women in Accra, Ghana.**

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42 **Abstract**

43 Human exposure data on dioxins and dioxin-like compounds (DLCs) in Ghana are limited.
44 Based on health risks associated with dioxins and DLCs, the impact of maternal body burdens
45 on foetal exposure is significant. This is the first study that assesses polychlorinated,
46 polybrominated and mixed halogenated dibenzo-p-dioxins and dibenzofurans (PCDD/Fs,
47 PBDD/Fs and PXDD/Fs), and dioxin-like polychlorinated biphenyls (dlPCBs) in sera of
48 primiparous Ghanaians. Our sample selection includes 34 participants from two municipalities
49 (Accra and Tema), and explores contributions from environmental and dietary exposures using
50 questionnaire data. Sample preparation involved C₁₈ solid phase extraction, purification with
51 acidified silica and lipid removal cartridges, and detection with gas chromatography-
52 atmospheric pressure chemical ionization-tandem mass spectrometry. The calculated average
53 toxic equivalent concentration was 5.3 pg TEQ/g lw, with contributions from dlPCBs (1.25 pg
54 TEQ/g lw), PCDD/Fs (3.10 pg TEQ/g lw), PBDD/Fs (0.49 pg TEQ/g lw) and PXDD/Fs (0.50
55 pg TEQ/g lw). The calculated total TEQ concentration was lower than background TEQ
56 concentrations reported in sera of pregnant women globally. Positive correlations were
57 obtained for total dioxins and DLC concentrations with age and Body Mass Index (BMI).
58 Dietary intake of seafood and dairy products had a strong influence on PCDD/F and dlPCB
59 concentrations. Statistically significant differences were observed for dioxins and DLCs in
60 participants from Accra (in close proximity to Agbobloshie e-waste site) and Tema. Given the
61 significant TEQ contribution of PBDD/Fs and PXDD/Fs (~20%), it is essential to explore these
62 classes of dioxins and DLCs in future biomonitoring studies as they may pose health risks, and
63 add extra diagnostic information in source exposure investigations.

64

65 **Keywords:** Ghana; background concentrations; dioxins; dioxin-like compounds; serum;
66 pregnant women

67 **1.0 Introduction**

68 Dioxins and dioxin-like compounds (DLCs) are a class of persistent organic pollutants
69 (POPs) produced from combustion and industrial processes of brominated and/or chlorinated
70 organic compounds. The most toxic dioxins and DLCs include 12 polychlorinated biphenyls
71 (dioxin-like PCBs- dlPCBs), and 17 congeners of 2378-polychlorinated dibenzo-p-dioxins and
72 dibenzofurans (PCDD/Fs) (Van den Berg et al., 2006). Structurally related analogues- 2378-
73 polybrominated and mixed halogenated dibenzo-p-dioxins and dibenzofurans (PBDD/Fs and
74 PXDD/Fs, X=Br and Cl) exhibit similar toxicity profiles (Birnbaum et al., 2003; Olsman et al.,
75 2007; Van den Berg et al., 2006). This class of compounds bioaccumulate in the environment;
76 in humans, they bind with the aryl hydrocarbon receptor (AhR) and induce toxic effects (Mason
77 et al., 1987; Olsman et al., 2007).

78

79 For many decades, occupational, accidental (unintentional) and background exposure
80 to dioxins and DLCs have resulted in adverse health concerns including carcinogenic and
81 dermal defects, neurodevelopmental and reproductive health effects (Arisawa et al., 2005;
82 Hites, 2011; White and Birnbaum, 2009). Populations occupationally or accidentally exposed
83 to POPs usually possess higher body burdens. For background exposure populations,
84 concentrations are lower, can be variable for diverse cultural groups, vulnerable groups, and
85 can also be impacted by age and gender (Porta et al., 2008). Approximately 90% of human
86 background exposure to dioxins and DLCs arise from dietary intake of contaminated food
87 (Djien Liem et al., 2000). However, questions about critical windows of foetal and early-life
88 infant exposure to dioxins and DLCs, and their impacts on embryonic, infant developmental or
89 later life consequences, reflect on placental transfer of dioxins and DLCs in nutrients from
90 maternal blood and breastmilk during infancy (Caspersen et al., 2016; Needham et al., 2010;
91 Pryor et al., 2000; Schechter et al., 2006). Studies undertaken on maternal participants, have

92 highlighted epidemiological evidence of short and long-term effects of in-utero exposure.
93 These include associations between maternal exposure, maternal diet, maternal age, hormonal
94 disruptions in children, and impacts on estrogenic metabolism (Baba et al., 2018; Cao et al.,
95 2008; Lignell et al., 2016; Miyashita et al., 2018; Nakajima et al., 2017; Papadopoulou et al.,
96 2014; Wittsiepe et al., 2008). Additionally, investigative studies on foetal exposure to dioxins
97 and DLCs provide evidence of an association between maternal serum with foetal abortion,
98 birth defects and low birth weight (Guo et al., 1995; Nham Tuyet and Johansson, 2001;
99 Yamashita and Hayashi, 1985). Although higher concentrations of dioxins and DLCs have
100 been detected in placenta (Schechter et al., 1990; Schechter et al., 1996; Suzuki et al., 2005; Wang
101 et al., 2004), and umbilical cord blood (Koopman-Esseboom et al., 1994; Suzuki et al., 2005),
102 venous blood sample during pregnancy is considered the most representative to evaluate
103 maternal or foetal body burdens.

104

105 In the last decade, numerous studies have supported increasing evidence of adverse
106 effects of dioxins and DLCs in general and occupationally exposed populations (Fromme et
107 al., 2009; Hong et al., 2009; Mato et al., 2007; Patterson et al., 2008; Patterson et al., 2009).
108 Only a few studies on dioxins and DLCs have been documented in African populations. These
109 include populations in Egypt, South Africa, Morocco, Canary Island- Spain and Ghana (Adu-
110 Kumi et al., 2010b; Asante et al., 2011; Henríquez-Hernández et al., 2016a; Henríquez-
111 Hernández et al., 2016b; Luzardo et al., 2014; Pieters and Focant, 2014; van den Berg et al.,
112 2017; Wittsiepe et al., 2015). Records of non-dlPCBs have been reported in sera and breastmilk
113 (Asamoah et al., 2018; Asante et al., 2011; Darnerud et al., 2011; Ennaceur and Driss, 2010;
114 Hassine et al., 2012; Müller et al., 2017; Röllin et al., 2009) in some African countries.

115

116 Ghana is located along the southernmost part of the West coast of Africa, and has a
117 wide range of potential sources of POPs. These include a historical legacy of pesticide use,
118 along with more recent sources of emerging pollutants such as dioxins and DLCs from the
119 electronic waste site in Agbogbloshie. To date, the current knowledge on POPs in Ghana
120 dominantly relates to the presence of organochlorine pesticides (OCPs) and non-dlPCBs in
121 environmental matrices. A review on the state of POPs in Ghana emphasized concerns of an
122 absence of human biomonitoring studies on dioxins, DLCs and emerging contaminants (Bruce-
123 Vanderpuije et al., 2019a). The limited studies on human biomonitoring, undertaken in Ghana,
124 have identified health risks from infant dietary intake of OCPs and non-dlPCBs in breastmilk
125 in occupationally and non-occupationally exposed lactating mothers in farming, fishing and e-
126 waste communities in Ghana (Asamoah et al., 2018; Asante et al., 2011; Asante et al., 2013;
127 Ntow, 2001; Ntow et al., 2008). Pioneering studies on dioxins and DLCs in soil from e-waste
128 sites, sera of e-waste workers, breastmilk from lactating mothers, and edible lake fish in Ghana
129 point to possibilities of bioaccumulation in humans (Adu-Kumi et al., 2010a; Adu-Kumi et al.,
130 2010b; Tue et al., 2016; van den Berg et al., 2017; Wittsiepe et al., 2015). Two studies have
131 quantified infant exposure to PCDD/Fs and dlPCBs in milk from Ghanaian lactating mothers-
132 The Ghana Environmental Protection Agency, and a global survey completed by the World
133 Health Organization (WHO)/United Nations Environment Programme (UNEP) (Adu-Kumi et
134 al., 2010b; van den Berg et al., 2017). There are currently no background exposure studies that
135 have quantified the toxic equivalents, and/or assessed the risks of exposure of pregnant women
136 and foetuses to one of the most toxic class of POPs- dioxins and DLCs in sera of pregnant
137 women in Ghana. Based on health concerns of mother and foetus, the goal of this study is to
138 provide baseline concentration data and to quantify the background exposure levels of 17
139 congeners of PCDD/Fs, 12 congeners of dlPCBs, 7 congeners of 2378-substituted PBDD/Fs
140 and 7 congeners of 2378-substituted PXDD/Fs in 34 primiparous Ghanaians.

141

142 1.1 Study sites

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144 Samples were obtained from participants in Tema and Accra, two municipalities
145 located along the South coast of Ghana within the Greater Accra region. These sites are noted
146 to generate industrial pollution including organic pollutants (releases into air and water bodies)
147 from local manufacturing industries. In addition, Agbogbloshie e-waste site, situated in
148 industrial area in Accra, is noted for the release of organic pollutants from open-air burning of
149 scrap electronic waste. Figure 1 shows a description of the two study sites. Residential homes
150 of pregnant women (participants) were at average distances of 5.6 miles (Accra) and 12 miles
151 (Tema) away from the Agbogbloshie e-waste site.

152 It is important to note that participants in this study had no known occupational or
153 accidental exposure. This study provides important baseline data for background
154 concentrations of dioxins and DLCs within primiparous Ghanaians. The study further aims to
155 identify the differences in exposure levels to dioxins and DLCs within the two municipalities,
156 and to identify linkages with dietary patterns or local sources of pollution. A comparison of
157 findings from this study with existing global datasets for similar cohorts were made on
158 calculated toxic equivalent concentrations (TEQs) for PCDD/Fs and dlPCBs using the
159 WHO₂₀₀₅-Toxic Equivalency Factors (TEFs). Concentrations of PBDD/Fs and PXDD/Fs were
160 also quantified and the tentative TEQs calculated for the brominated and mixed halogenated
161 analytes were based on TEFs from their corresponding chlorinated analogues. In addition, other
162 types of PXDD/F and PCB analytes that are not considered as toxic as 2378-substituted dioxins
163 and DLCs were quantified, when identified.

164

2.0 Material and Methods

2.1 Participant Recruitment

The study was approved by the Ghana Health Service Ethics Review Committee, and conducted in accordance with ethical principles for medical research involving human subjects. Women in their eighth month of pregnancy were interviewed by research nurses during routine health check-ups at the clinics. Participants recruited from The Greater Accra Regional and Tema General Hospitals, voluntarily completed informed written consent forms and exposure assessment questionnaires prior to sample collection. Data on age, occupation, diet, cigarette smoking and alcohol intake and bodyweight were documented. Maternal venous blood (15 mL, n = 34) was collected in April 2017, from primiparous women in Accra (n = 17) and Tema (n = 17). Blood was collected into clear 15 mL Corning centrifuge tubes without anticoagulant; 5-6 mL serum was obtained by centrifugation in an Eppendorf Centrifuge 5810 at 4000 rpm/rcf for 10 min, within 24 hours of sample collection. Serum samples were stored at -20 °C prior to extraction and analysis.

2.2 Reagents and Chemicals

Distilled in glass grade organic solvents- n-hexane, toluene, nonane, acetonitrile, methanol and water were obtained from Caledon Laboratories Limited (Georgetown, Ontario, Canada). Formic acid, 88% analytical grade reagent and octadecyl non-encapped bonded silica C₁₈ cartridges (500 mg/6 mL) were obtained from Thermo Fisher Scientific. Captiva EMR-Lipid removal cartridges (600 mg/6 mL) were obtained from Agilent Technologies. For the separation of planar and co-planar PCDD/Fs, dlPCBs and PXDD/Fs (and PBDD/Fs) from non-planar compounds, ultra clean carbon mini-columns (2%) and re-usable glass column reservoirs (20 cm in length, 0.5 cm in diameter) from Cape Technologies were used. ¹³C-

191 labelled isotope and native PCDD/Fs, dlPCBs and PXDD/Fs standards were obtained from
192 Wellington Laboratories Inc. (Guelph, Ontario, Canada). Chromatographic separation column:
193 DB5-MS (5% diphenyl 95% dimethyl polysiloxane, 60 m x 0.25 mm ID x 10 µm film
194 thickness, J&W Scientific, CA, USA) was obtained from Agilent.

195

196 Preparation of calibration, recovery and injection standards for PCDD/Fs, PXDD/Fs,
197 PBDD/Fs and dlPCBs are described in detail in Section 1.2 in the Supplementary information.
198 PCDD/F, PBDD/F, dlPCB, and PXDD/F analytes analysed in this study are listed in Table S1.
199 All standards were prepared in nonane, except for recovery spiking solutions which were
200 prepared in methanol.

201

202 2.3 Sample Extraction

203

204 Sample extraction and clean-up steps used in this study were based on analytical
205 procedures from CDC with modifications (Centre for Diseases Control and Prevention, 2016).
206 Extraction was performed on serum samples using C₁₈ solid phase extraction (SPE) with
207 hexane, clean-up using Captiva-EMR lipid removal cartridge and acidified silica, and
208 fractionation on carbon column. Samples were analysed using capillary gas chromatography
209 with atmospheric pressure chemical ionization (APCI) and a triple quadrupole mass
210 spectrometer (GC-APCI-MS/MS, Xevo TQ-XS) from Waters Corporation, Manchester, UK.

211

212 The analytical procedure for method development is detailed in Section 1.3 in the
213 Supplementary information. For sample extraction, briefly, 2 g of serum was spiked with 5 µL
214 of 2 pg/µL label recovery mix- ¹³C₁₂-PCDD/Fs, ¹³C₁₂-dlPCBs and ¹³C₁₂-PXDD/Fs- to
215 determine extraction efficiency, matrix effects on recovery and enable quantitation by isotope

216 dilution mass spectrometry. Two millilitres of water was added for matrix dilution; 2 mL
217 formic acid for protein denaturation. Serum sample mix was vortexed and sonicated for 15 min
218 in between additions. C₁₈ cartridges were conditioned gravimetrically using two cartridge
219 volumes of methanol and water prior to loading serum mixture, and eluted at a flow rate of 0.6
220 mL/min. Serum culture tubes were rinsed with 2 x 5 mL H₂O and transferred onto C₁₈ cartridge
221 barrels. Cartridges were dried under vacuum pump suction for 1 hour to remove water.
222 Analytes were eluted from C₁₈ cartridge using 3 x 5 mL hexane and collected in clear EPA
223 vials, at a flow rate of 0.6 mL/min.

224

225 **2.4 Lipid removal Clean-up and Fractionation**

226

227 Extracts were evaporated to 500 µL; 3 mL acetonitrile was added. Extracts were loaded
228 onto Captiva-EMR Lipid removal cartridge and allowed to flow under gravity. Vials were
229 rinsed with 5 mL ACN and loaded onto EMR cartridge. Eluate was collected under gravity into
230 EPA vials. Extracts were evaporated to 1 mL.

231

232 For each sample, acidified silica cartridge was connected to a carbon column and
233 activated with 20 mL hexane. A 1 mL serum extract was loaded and the cartridge was rinsed
234 with 30 mL hexane. The acidified silica cartridge was replaced with a reusable glass column
235 reservoir. The carbon column was then inverted and eluted with 30 mL toluene. Eluate was
236 collected in 40 mL EPA vials, evaporated to 350 µL under low N₂, transferred into inset vials,
237 evaporated to dryness, and reconstituted with 10 µL of 1 pg/µL injection standard. A flow chart
238 of sample preparation steps is shown in Figure S1.

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2.5 Lipid measurement

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Five-hundred microlitres of each serum sample was used to determine the total lipid concentration. Enzymatic analysis of cholesterol and triglycerides was completed using a calibrated BT3000 chemistry auto analyzer (Biotechnica Instruments). Total lipids were calculated from the sum of the total cholesterol and triglycerides concentrations using the formula:

$$\text{Total lipids} = (2.27 \times \text{Total cholesterol}) + \text{Triglycerides} + 62.3 \text{ mg/dL} \quad \text{Equation 1}$$

247
248

Where total lipids, total cholesterol, triglycerides concentrations are reported as mg/dL.

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2.6 Instrumental Analysis

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The gold standard for measuring trace levels (femtogram) of POPs including PCDD/Fs has traditionally been with a gas chromatograph coupled to a double focusing magnetic deflection (sector)-high resolution mass spectrometer (GC-HRMS). However, gas chromatography with atmospheric pressure chemical ionization and tandem mass spectrometry (GC-APCI-MS/MS) has recently been used successfully to match the performance of the sector instrument for the analysis of PCBs, and PCDD/Fs (García-Bermejo et al., 2015; Geng et al., 2016; Megson et al., 2016; Organtini et al., 2015a; Organtini et al., 2015b; Stableski et al., 2018; van Bavel et al., 2015). Sample analyses was performed using GC-APCI-MS/MS. Method development was performed on a Q-Ion Mobility Spectrometry-ToF instrument (APGC Synapt G2-Si) operating in full scan mode. A 1 μL sample extract was injected on a DB5-MS (60m x 0.25mm x 0.1 μm) non-polar stationary phase column. Instrumental parameters and operating conditions are summarized in Table S2. The mass spectrometer was operated in positive ion mode, using multiple reaction monitoring. Four transitions (2

265 quantifiers and 2 qualifiers) were monitored for native and $^{13}\text{C}_{12}$ labelled dlPCBs, PCDD/Fs
266 and PXDD/Fs. For PCDD/Fs the loss of -COCl was monitored (Table S1a). For dlPCBs, the
267 loss of Cl_2 was monitored (Table S1b). Based on studies of Organtini et al. (2015a, b) and
268 Myers et al. (2012), and from the results of the method development, ions monitored for mixed
269 halogenated dibenzo-p-dioxins and dibenzofurans included native and label -COBrCl, -COBr,
270 $-(\text{CO})_2\text{BrCl}$, -COCl, -Br₂, and -Br (Myers et al., 2012; Organtini et al., 2015a; Organtini et al.,
271 2015b). The transitions, collision energies and isotope ratios are summarized in Table S1.

272

273 2.6.1 Quality assurance/ Quality Control

274

275 Analytes were quantified by isotope dilution using ions specified in Table S1. For non 2378-
276 PXDD/Fs and PBDD/Fs for which there were no ^{13}C labeled standards, a semi-quantitative
277 method was utilized in quantification. A detailed description on the methodology for reporting
278 linearity, quality control and method detection limits are explained in Section 1.5 in the
279 Supplementary Information. For linearity, the response obtained for a native, relative to its
280 corresponding label ^{13}C standard was linear for the range of calibration standards analyzed.
281 Calculated coefficient of determination for PCDD/F analytes was $R^2 \geq 0.998$ (except for
282 OCDD and OCDF: $R^2 = 0.984$), that for dlPCBs and PXDD/Fs were $R^2 \geq 0.995$ and $R^2 \geq 0.996$,
283 respectively. The percentage RSDs obtained for PCDD/Fs, dlPCBs and PXDD/Fs ranged
284 between 1.6 and 13.8%; this is in agreement with the acceptable 15% value (Centre for
285 Diseases Control and Prevention, 2006). Method validation was performed by analysis of
286 fortified serum NIST Standard Reference Material (SRM) 1958. Recoveries for NIST standard
287 ranged between 75% and 105% for PCDD/Fs, and 67.5% and 96.3% for dlPCBs. The results
288 for the SRM are presented in Supplementary Information-Section 1.3; data is presented in
289 Table S3a and 3b. Recoveries for isotopically labelled standards, spiked into serum prior to

290 extraction/clean-up, ranged between 47.9 and 120% for dlPCBs, 47.3 and 129.9% for
291 PXDD/Fs, and 41.8 and 140% for PCDD/Fs (recoveries obtained fell within the acceptable
292 EPA ranges (40-145%) (EPA, 2010), except for 4 samples, for which ¹³C-OCDD and ¹³C-
293 1,2,3,4,6,7,8-HpCDD ranged between 28.8 and 37%). The instrument limit of detection
294 (iLOD) was restricted by the lowest detectable standard concentration, and ranged between 5
295 and 100 fg/μL for PCDD/Fs and PXDD/Fs, and 5 fg/μL for dlPCBs. Where concentrations
296 were below the LOD, ½ LODs were assigned and used in the TEQ calculations.

297

298 The lipid adjusted sample serum concentrations of dioxins and DLCs are reported as pg/g lipid.
299 The Toxic Equivalent (TEQ) for each class of dioxins and DLCs was reported as pg WHO-
300 TEQ/g lipid weight (lw)(Van den Berg et al., 2006).

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302

2.6.2 Statistical Analyses

303

304 Dioxins and DLCs in sera were statistically analysed using the Minitab 18 software
305 package (Minitab, 2010), to determine differences in concentrations for the three groups of
306 analytes: PCDD/Fs, dlPCBs and PXDD/Fs. Spearman rank correlation and bivariate linear
307 regression were used to evaluate bivariate associative correlations between dioxins and DLC
308 concentrations, and factors such as age, gestational week, food consumed and body mass index
309 (BMI), and congener concentrations of PCDD/Fs, PXDD/Fs and dlPCBs, as well as total
310 concentration (PCDD/Fs + PXDD/Fs + dlPCBs). Multivariate statistical analysis (JMP[®],
311 Version 14.1. SAS Institute Inc., Cary, NC, 1989-2007) was used to assess congener-specific
312 distributions. Exploratory data analysis was conducted using Principal Component Analysis
313 (PCA) with Hierarchical Cluster Analysis (HCA) using both box-cox normalized and log-
314 normalized contaminant concentrations. This was performed to investigate if participants with

315 different characteristics (e.g. different geographical areas, dietary preferences and age) had a
316 distinctive chemical pattern.

317

318 The normality of data distribution was checked with Kolmogorov-Smirnov test. A log-
319 normal distribution for PCDD/F, PXDD/F and dlPCB concentrations for 34 pregnant women
320 dataset was identified. Because the serum concentration data was not normally distributed,
321 descriptive statistics for central tendency of the data was based on the geometric mean rather
322 than on arithmetic mean. The range and 95% confidence interval were used to describe the
323 data. In addition, the relative percentages of each congener to the total concentration was
324 evaluated for each class of dioxins and DLCs.

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3.0 Results and Discussion

3.1 Food consumption

Questionnaire responses on food consumption included major dietary intake of seafood, fish, meat and meat products, and dairy products; this can be located in Supplementary Information- Section 1.7. Seafood consumed included shrimps, clams, mussels, snails, squid, oysters, and lobsters. Fish types included: salmon, mackerel, tilapia, tuna, and dried herring. Dairy products frequently consumed included eggs and milk. Majority of the participants reported consumption patterns that were generally similar for both Tema and Accra municipalities.

3.2 Characteristics of pregnant women

The characteristics of the participants are presented in Table 1. All participants were primiparous non-smokers; the mean age was 25 yrs (range: 18-38 yrs). The mean gestational age was 31.9 weeks, and the average body weight was 75.7 kg. Almost 40% of participants had a BMI that exceeded 25 kg/m² (overweight), none was underweight, and 60% had healthy weight (range: 21.3-24.9 kg/m²). Fifty percent (17) of the subjects resided in Tema, and 50% (17) in Accra, Ghana.

3.3 Congener specific dioxins and DLCs identified

PCBs 105 and 118 were detected in all the participants. The remaining congeners (114, 126, 156, 157, 169 and 189) were regularly detected in participants with detection frequencies of 82.4 and 97.6%. For PCDD/Fs, analytes for which serum concentrations were consistently

362 above the LOD were 12378-PeCDF, 23478-PeCDF, 123478-HxCDF, 1234678-HpCDF,
363 OCDF, 123478-HxCDD, 123678-HxCDD, 123789-HxCDD, 1234678-HpCDD and OCDD.
364 Concentrations of chlorinated dioxins and PCBs have been widely reported in human serum;
365 however, much less congener specific data has been provided for mixed halogenated dioxins
366 and furans. This study provides an important baseline data showing that 8-B-234-CDF, 3-B-
367 278-CDF, 12-B-78-CDF and 4-B-2378-CDF were consistently identified in all 34 participants.
368 Seven congeners of 2378-PXDD/F and 7 congeners of 2378-PBDD/F were detected
369 intermittently. Concentrations of non-2378-PBDD/Fs and PXDD/Fs for 8-B-23-CDF, 7-B-23-
370 CDD, 2-B-78-CDD, 2-B-378-CDD, 23-B-78-CDF, 13-B-278-CDF, 1378-BDD, 2378-BDF,
371 1234-BDD, 12478-BDD and 23478-BDF were all below the LOD.

372

373 **3.4 Concentrations of dioxins and DLCs**

374

375 The baseline data obtained for both localities are presented in Table 2. The
376 concentrations of dioxins and DLCs in participants were within one or two orders of magnitude
377 where dlPCBs > PCDD/Fs > PBDD/Fs and PXDD/Fs. The total mean concentrations for
378 dlPCBs, PCDD/Fs, and PBDD/Fs (and PXDD/Fs) in Tema were: 59.3 pg/g lw, 52.6 pg/g lw,
379 and 2.47 pg/g lw, respectively. The mean concentrations in Accra were slightly higher than
380 that for Tema for dlPCBs, PCDD/Fs and PBDD/Fs (and PXDD/Fs); these were 96.1 pg/g lw,
381 71.8 pg/g lw, and 4.19 pg/g lw, respectively. Statistically significant differences were observed
382 between mean concentrations of dlPCBs in Tema and Accra with the exception of PCB 157
383 (p-value = 0.092). For PCDD/Fs, statistically significant differences were observed for 4 furan
384 congeners: 2378-TCDF, 1234678-HpCDF, 1234789-HpCDF and OCDF, for both groups of
385 participants. Concentrations of 7 of the 14 congeners of 2378-PXDD/Fs and PBDD/Fs (2-B-
386 378-CDD, 23-B-78-CDF, 23-B-78-CDD, 13-B-278-CDF, 2378-BDF, 12378-BDF, 12378-

387 BDD), were also statistically significantly higher in participants from Accra than Tema. Both
388 sites- Accra and Tema are along the coastal areas; however, the differences observed in dlPCBs
389 and the 4 sets of congeners may be explained by subtle dietary patterns as different food groups
390 have been shown to have an important influence on dlPCB and PCDD/F exposure. Other
391 possible reasons for the observed differences could be due to local factors such as combustion
392 processes from exposure sites: Agbobloshie e-waste site. Further research is required to
393 identify contributions from potential sources. Figure 2 presents a comparison of average
394 concentrations for dlPCBs, PCDD/Fs, and PBDD/Fs (with PXDD/Fs) for the two sites. Tables
395 2a, 2b and 2c show the baseline data obtained for the two sites for dlPCBs, PCDD/Fs, and
396 PBDD/Fs (and PXDD/Fs) respectively. It also shows p-values obtained from 2-sample t-test
397 (comparison), and their toxic equivalent concentrations (pg TEQ/g lw) for participants from
398 Tema and Accra.

399
400 Concentrations of PCB 118, 105 and 156 were significantly higher than other dlPCBs
401 in sera from both Tema and Accra (Table 2a). For PCDD/Fs, in comparison to other congeners
402 detected, 12378-PeCDF, OCDF, 1234678-HpCDD and OCDD were higher in both localities.
403 In addition, contributions from PCDDs were approximately twice that for PCDFs. Similarly,
404 for the sum of congeners of 2378-PBDD/Fs and 2378-PXDD/Fs, a higher total concentration
405 was observed in Accra in comparison to Tema. This could be attributed to greater releases of
406 PBDD/Fs, PXDD/Fs and PCDD/Fs into neighbourhoods from open-air burning of both
407 chlorinated and low molecular weight polybrominated diphenyl ethyl ether (PBDE) containing
408 e-waste materials in Agbobloshie, Accra.

409

410 **3.5 Total TEQ concentrations**

411

412 Tables 2 and S4 present a summary of the TEQ concentrations of dlPCBs, PCDD/Fs and
413 PBDD/Fs (and PXDD/Fs). The average total TEQ concentrations for participants in Accra and
414 Tema were 6.48 and 4.19 pg TEQ/g lw, respectively. Figure 3 presents a stacked bar chart of
415 TEQ data obtained for the four classes of dioxins and DLCs (dlPCBs, PCDD/Fs, PBDD/Fs and
416 PXDD/Fs) in each participant. Although PXDD/Fs (and PBDD/Fs) were generally present at
417 lower concentrations than for other dioxins and DLCs, they had a large impact on the overall
418 TEQ, accounting for approximately 20% of the total TEQ contribution in participants from
419 both Accra and Tema. This indicates the importance of mixed halogenated dioxins and furans
420 when undertaking human health risk assessments. The largest contributor to dlPCBs TEQ was
421 PCB 126 (the most toxic dlPCB) which accounted for approximately 25%. Large contributions
422 were also recorded for 12378-PeCDD (~25%) and for 2378-TCDD (~20%). The largest
423 contributions from the brominated and mixed halogenated congeners were 2378-BDD (~5%),
424 23-B-78-CDD (~4%) and 2-B-1378-CDD (~3%). Based on a comparison of relative potencies
425 of PBDD/Fs and PXDD/Fs, 2378-BDF and 3-B-278-CDD are of similar potency as 2378-
426 TCDD (Birnbaum et al., 2003) and this explains their contributions. However, the impact of
427 the TEF influences the congener contribution towards its toxicity; this explains why minimal
428 contributions were observed for OCDD and PCB 118, although these were the dominant
429 congeners detected.

430 To interpret TEQs observed, in a global context, our results were compared with
431 available TEQ data reported in maternal serum in literature (Table 3).

432

433

434 **3.6 Observed relationship between dioxins and DLCs and Ghanaian** 435 **maternal serum**

436

437 To detect associations between age, BMI and dioxins and DLCs, possible effects were
438 assessed from an examination of serum concentration data using Spearman rank correlation
439 and bivariate analysis. An explanation of results is presented in detail in Supplementary Section
440 1.6. There was statistically no significant difference between residents of Accra and Tema
441 participants in terms of age. From bivariate analysis, the main predictors that confirmed
442 statistically significant associations between mean concentrations of dioxins and DLCs, and
443 total TEQ (PCDD/Fs and dlPCBs) were age and BMI. Additionally, positive correlations
444 confirmed that an increase in BMI potentially induces an increase in dlPCBs, PCDD/Fs and
445 PXDD/Fs during pregnancy. Negative correlations were obtained for the association between
446 gestational week and dioxins and DLCs; an indication that gestational weeks do not necessarily
447 increase dioxins and DLCs. Respectively, the mean concentrations (pg WHO-TEQ/g lw) were
448 highest and lowest for age ranges of 28-38 years and 18-23 years. In overweight pregnant
449 women, mean concentrations of dioxins and DLCs were higher than for pregnant women of
450 normal weight. In addition, dioxin-like exposure was higher in sera of pregnant women who
451 resided in Accra in comparison to those from Tema (Table S4).

452

453 **3.7 Chemical signatures**

454

455 **3.7.1 Principal Component Analysis (PCA)**

456

457 Exploratory data analysis involving Principal Component Analysis (PCA) was used to
458 investigate how the relative proportions of dioxins and DLCs changed in different participants.
459 This was performed to determine if participants with different characteristics (e.g. different
460 geographical areas) had a distinct chemical signature. The data set was normalized for dlPCBs,
461 PXDD/Fs and PCDD/Fs by expressing the concentration of each congener as a percentage of
462 the combined sum of congeners within its class. A loading plot was constructed using PCA

463 (JMP[®], Version 14.1. SAS Institute Inc., Cary, NC, 1989-2007) to explain the observed
464 relationships between samples and their contributions. A consideration of all dioxins and DLC
465 data points showed that two demographics described the congener concentrations.
466 Demographic 1 (location), was influenced by PXDD/Fs, and demographic 2 (food type
467 frequently consumed) influenced both PCDD/Fs and dlPCBs. The results indicated that the
468 data was dominated by a few individuals with elevated PCB concentrations. When the data for
469 all dioxins and DLCs were assessed together, it appeared to be of little diagnostic value.
470 However, when certain classes of dioxins and DLCs were removed from the total data set, the
471 geographical locations of the participants were clearly separated. These results showed that
472 PXDD/F congeners were much better at identifying local differences, with PCB profiles also
473 showing some degree of relationship with diet.

474

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3.7.2 PCBs

477 A plot of the first two principal components shows the variances of normalized
478 concentrations for dlPCBs and PXDD/Fs for the 34 residents of Accra and Tema.
479 Complementary loading plots based on congener profiles that provide similarities in groupings
480 for contaminants observed in the PCA, for participants have also been shown in Figures 4 and
481 5. The first principal component accounted for 66.9% of the total variability in dlPCB
482 concentrations (Figure 4). These originated from the most abundant dlPCB congeners, and
483 showed a positive correlation towards participants who had lived in Accra during their
484 childhood, irrespective of their current place of habitation (Accra or Tema). The second
485 principal component accounts for 7.5% of the original variance of the data set, and was mostly
486 attributed to dlPCBs, and other non-dlPCBs. Serum samples for residents of Accra were
487 separated from those from Tema by the PCA. Group I consisted of samples R10, R12 and R13
488 (participants born and raised in Accra), and contained higher proportions of PCBs- 77, 81, 118,

489 167, 169, 114 and 194. Group II mostly consisted of participants born and raised in other parts
490 of the country who had settled in their matrimonial homes in Accra. These consisted of samples
491 R14-R17, R19 and R2. The concentrations in these samples were enriched with both dlPCBs
492 and non-dlPCBs: PCBs- 128, 206, 118, 105 and 123. The dietary pattern of the group consisted
493 mostly of fish and meat. Group III consisted of participants who had grown up in varying
494 localities of the country and later relocated to either Accra or Tema. These samples consisted
495 of T1-T19, and 9 of the samples from Accra (R1, R3, R5, R6, R8, R9, R11, R15 and R18). The
496 dietary patterns observed mostly consisted of fish, dairy products, seafood and meat. For the
497 majority of these concentrations, the patterns suggest that the source of dlPCBs detected in
498 serum could originate from differences in dietary intake.

499

500

3.7.3 PXDD/Fs and PBDD/Fs

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Using the Box-Cox normalized concentrations, exploratory data analysis using both
503 PCA and HCA was conducted. Results of the PCA and HCA are shown in Figure 5. For the
504 PCA, two factors accounted for 49.6% (component 1) and 13.3% (component 2) of the variance
505 explained (total 62.9%). Two groups were identified from the PCA clustering. Group I
506 contained 18 participants, some of whom had grown up in poor neighbourhoods in Nima
507 (Accra) and who are currently staying in Accra. From this group, 5 participants possessed a
508 signature that fits the pattern for Accra participants. The major congeners detected in this group
509 were 3-B-278-CDF, 2378-BDF and 1234678-BDD. The potential sources of these congeners
510 include combustion processes from vehicular emissions and biofuel (fuel/charcoal/firewood
511 burning for household and commercial use). In addition, potential sources could further be
512 attributed to the proximity of the Nima locality to the Agbogbloshie e-waste sites. However,
513 as highlighted in a previous study, combustion processes from such a neighbourhood have been
514 identified to contribute to biomass smoke and air-particle pollution in Nima- Accra (Zhou et

515 al., 2013). Group II consisted of 16 participants, the majority (n =13) of whom grew up in
516 Tema, and still reside in Tema. Commonly detected PXDD/F congeners in this group were 3-
517 B-278-CDF and 2378-BDF and 1234678-BDD. These results indicate the evidence that
518 PBDD/Fs (and PXDD/Fs) are potentially linked to the place of residence of an individual,
519 whereas PCBs are linked to dietary patterns.

520

521 **3.8 Data comparison with other studies**

522

523 In the absence of published data on exposure to dioxins and DLCs in sera of
524 primiparous Ghanaians, result comparisons for vulnerable populations with no known
525 exposure cannot be made. However, a comparison between this study and other studies
526 globally (Table 3), shows that the overall mean (5.3 pg WHO-TEQ/g lw) in sera of primiparous
527 Ghanaians is lower than most background concentrations reported in sera of pregnant women
528 in other parts of the world. Results from this study are also lower than median concentrations
529 (PCDDs + PCDFs) detected for occupationally exposed e-waste populations in Ghana: 6.18 pg
530 WHO-TEQ/g₂₀₀₅ lw (Wittsiepe et al., 2015); TEQ concentrations detected in the latter
531 (occupationally exposed e-waste workers) are still lower than TEQ concentrations reported in
532 sera of pregnant women globally. Our results [mean total TEQ concentrations [dlPCBs +
533 PCDD/Fs = 4.34 pg TEQ/g lw] are similar to the mean total TEQ concentrations [dlPCBs +
534 PCDD/Fs] detected in individual breastmilk of lactating mothers (n = 42, 6.07 pg TEQ/g lw)
535 in Accra in 2008 in studies of Adu-Kumi et al. (2010b), and from pooled samples from the
536 WHO/UNEP global survey monitoring program (n = 50, 5 pg TEQ/g lw)(Adu-Kumi et al.,
537 2010b; van den Berg et al., 2017). In a similar related study, higher concentrations of the seven
538 indicator PCBs (non-dlPCBs) were detected in breastmilk samples of lactating mothers
539 (occupationally exposed participants and residents: 4.43 ng/g lw) at the Agbogbloshie e-waste

540 site in comparison to breastmilk samples from Kwabenya (control group: 0.03 ng/g lw), in
541 Accra, Ghana (Asamoah et al., 2018). A high potential health risk estimated from the seven
542 indicator PCBs (including dlPCB 118) indicated significantly higher concentrations in
543 breastmilk from lactating mothers residing in Accra (sum of average PCBs: 82 ng/g lw, PCB
544 118: 3.0 ng/g lw) in comparison to participants from Kumasi (sum of average PCBs: 65 ng/g
545 lw, PCB 118: 2.6 ng/g lw) and Tamale (sum of average PCBs: 30 ng/g lw, PCB 118: 1.9 ng/g
546 lw). Thus, areas considered to be hotspots in Accra- such as the Agbogbloshie e-waste site and
547 heavy industrial areas can impact background concentrations of dioxins and dioxin-like
548 compounds (Asante et al., 2011).

549

550 In order to answer questions on why lower dioxins and DLC concentrations were
551 obtained in comparison to other countries, we considered studies in Ghana that had focused on
552 dietary intake of dioxins and DLCs, since 90% of human background exposure arises from
553 contaminated food (Djien Liem et al., 2000). Due to the absence of publications on estimated
554 daily exposure (dietary intake of PCDD/Fs, and dlPCBs) and exposures to PXDD/Fs and
555 PBDD/Fs for the general population of Ghana, it is not possible to make robust comparisons
556 with other countries. Relatively low concentrations of PCDD/Fs and dlPCBs have been
557 estimated in food (fish) in Ghana, in comparison to other industrialized countries (Adu-Kumi
558 et al., 2010a). Although the consumption of various foods influences the total TEQ for
559 PCDD/Fs and PCBs, there is only one study that has estimated concentrations in tilapia and
560 catfish from Lake Volta in Ghana (and may not necessarily be representative of the entire
561 country). Data/results from the study of Adu-Kumi et al. (2010a), however, serves as a baseline
562 for comparison with other countries. WHO-TEQ₂₀₀₅ concentration (wet weight) of PCDD/Fs
563 and dlPCBs in fish from Ghana: ~ 0.3 pg TEQ/g (Adu-Kumi et al., 2010a) was much lower
564 than that estimated in the Baltic Sea: 8 pg TEQ/g- salmon and prat and 12 pg TEQ/g- eel

565 (Szlinder-Richert et al., 2009), in fresh water or farmed fish from France: ~ 2.8 pg TEQ/g (Tard
566 et al., 2007), and in freshwater fish in South Korea: 1.3 pg TEQ/g (Kim et al., 2004). This
567 indicates that industrialized and developed countries may have a higher dietary intake of
568 contaminated foods- fish, meat, seafood containing PCDD/Fs and dlPCBs in comparison to
569 developing countries such as Ghana.

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590 **4.0 Conclusions**

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592 To our knowledge, this is the first study on background concentrations of dlPCBs,
593 PCDD/Fs, PBDD/Fs and PXDD/Fs in sera of primiparous Ghanaians. The results provide an
594 average value of 5.3 pg WHO-TEQ/g lw for a cohort of 34 individuals with no known
595 accidental or occupational exposure. This value was generally lower than TEQ concentrations
596 of dioxins and DLCs in other studies of sera from pregnant women in industrialized countries.
597 A breakdown of the results shows that %TEQ contributions in our cohort are predominantly
598 resulting from exposure to PCDD/Fs (57.9%), with significant contributions from dlPCBs
599 (23.4%), as well as PBDD/Fs and PXDD/Fs (18.8%). These percentages suggest that
600 substantial contributions from PCDDs (39.2%) are indicative of sources other than combustion
601 (potentially from dietary exposure). However, contributions from combustion processes (from
602 PBDD/Fs and PXDD/Fs) are also noted to influence the overall TEQ. The results of the
603 PBDD/F and PXDD/Fs data were significantly higher in participants living in close proximity
604 to both e-waste and heavy industrial areas in Accra, than for a group of cohorts in Tema.
605 Multivariate statistical analysis of the data was able to distinguish between participants from
606 the two municipalities using a chemical fingerprint generated with only PBDD/Fs and
607 PXDD/Fs. The results indicate that, over time, local sources of potential contamination
608 (industrial areas and Agbogbloshie e-waste site) may impact populations that visit, work or live
609 in close proximity to the site. We would recommend future studies to better establish the
610 sources of dioxins and DLCs in Ghana, and potential trends over time. We also recommend
611 that future biomonitoring studies on dioxins and DLCs include determination of PBDD/F and
612 PXDD/Fs as this study indicates that their total contribution is significant.

613

614

615 **Strengths**

616 The main strengths of the present study are the congener-specific analysis of toxic
617 dioxin-like contaminants- PCDD/Fs, dlPCBs, PXDD/Fs and PBDD/Fs in maternal serum,
618 using targeted analysis. To the best of our knowledge this is the first study, in Ghana, to
619 examine maternal and foetal exposure to background concentrations of dioxins and DLCs.

620

621 As this study was focused on providing trace level targeted analysis of dioxins and DLCs, it
622 was impossible to complete a non-targeted analysis to determine the presence of other classes
623 of POPs. Future monitoring studies, could focus on non-targeted analysis to identify the varied
624 classes of toxic contaminants vulnerable Ghanaians are exposed to.

625

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629

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634

635 **Financial Interest Declaration**

636 The authors declare no financial interests.

637

638 **Ethical Approval**

639 This study received ethical approval from the Ghana Health Service Ethics Review Committee

640 (ref: GHS-ERC 04/08/16) on February 16th, 2017.

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672 Adu-Kumi, S., Kawano, M., Shiki, Y., Yeboah, P., Carboo, D., Pwamang, J., Morita, M.,
673 Suzuki, N., 2010a. Organochlorine pesticides (OCPs), dioxin-like polychlorinated
674 biphenyls (dl-PCBs), polychlorinated dibenzo-p-dioxins and polychlorinated dibenzo
675 furans (PCDD/Fs) in edible fish from Lake Volta, Lake Bosumtwi and Weija Lake in
676 Ghana. *Chemosphere* 81, 675-684.

677 Adu-Kumi, S., Malisch, R., Kotz, A., Kypke, K., Asante, K., Takahashi, S., Tanabe, S.,
678 Takasuga, T., Clarke, E., Weber, R., 2010b. Levels of persistent organic pollutants
679 (POPs) in human breast milk samples from Ghana. *Organohalogen Compounds* 72,
680 1046-1049.

681 Arisawa, K., Takeda, H., Mikasa, H., 2005. Background Exposure to PCDDs/PCDFs/PCBs
682 and its Potential Health Effects: A Review of Epidemiologic Studies. *The Journal of*
683 *Medical Investigation* 52, 10-21.

684 Asamoah, A., Essumang, D.K., Muff, J., Kucheryavskiy, S.V., Sogaard, E.G., 2018.
685 Assessment of PCBs and Exposure Risk to Infants in Breast Milk of Primiparae and
686 Multiparae Mothers in an Electronic Waste Hot Spot and Non-hot Spot Areas in Ghana.
687 *Science of The Total Environment* 612, 1473-1479.

688 Asante, K.A., Adu-Kumi, S., Nakahiro, K., Takahashi, S., Isobe, T., Sudaryanto, A.,
689 Devanathan, G., Clarke, E., Ansa-Asare, O.D., Dapaah-Siakwan, S., Tanabe, S., 2011.
690 Human Exposure to PCBs, PBDEs and HBCDs in Ghana: Temporal Variation, Sources
691 of Exposure and Estimation of Daily Intakes by Infants. *Environment International* 37,
692 921-928.

693 Asante, K.A., Takahashi, S., Itai, T., Isobe, T., Devanathan, G., Muto, M., Agyakwah, S.K.,
694 Adu-Kumi, S., Subramanian, A., Tanabe, S., 2013. Occurrence of halogenated
695 contaminants in inland and coastal fish from Ghana: levels, dietary exposure
696 assessment and human health implications. *Ecotox Environ Safe* 94, 123-130.

697 Baba, T., Ito, S., Yuasa, M., Yoshioka, E., Miyashita, C., Araki, A., Sasaki, S., Kobayashi, S.,
698 Kajiwara, J., Hori, T., 2018. Association of Prenatal Exposure to PCDD/Fs and PCBs
699 with Maternal and Infant Thyroid Hormones: The Hokkaido Study on Environment and
700 Children's Health. *Science of The Total Environment* 615, 1239-1246.

701 Birnbaum, L.S., Staskal, D.F., Diliberto, J.J., 2003. Health Effects of Polybrominated Dibenzo-
702 p-dioxins (PBDDs) and Dibenzofurans (PBDFs). *Environment International* 29, 855-
703 860.

704 Bruce-Vanderpuije, P., Megson, D., Reiner, E.J., Bradley, L., Adu-Kumi, S., Gardella, J.A.,
705 2019a. The state of POPs in Ghana- A review on persistent organic pollutants:
706 Environmental and human exposure. *Environmental Pollution* 245, 331-342.

707 Cao, Y., Winneke, G., Wilhelm, M., Wittsiepe, J., Lemm, F., Fürst, P., Ranft, U., Imöhl, M.,
708 Kraft, M., Oesch-Bartlomowicz, B., Krämer, U., 2008. Environmental exposure to
709 dioxins and polychlorinated biphenyls reduce levels of gonadal hormones in newborns:
710 Results from the Duisburg cohort study. *International Journal of Hygiene and*
711 *Environmental Health* 211, 30-39.

712 Caspersen, I., Haugen, M., Schjølberg, S., Vejrup, K., Knutsen, H., Brantsaeter, A., Meltzer,
713 H., Alexander, J., Magnus, P., Kvalem, H., 2016. Maternal Dietary Exposure to Dioxins
714 and Polychlorinated Biphenyls (PCBs) is Associated with Language Delay in 3 year
715 old Norwegian Children. *Environment International* 91, 180-187.

716 Centre for Diseases Control and Prevention, 2006. Laboratory Procedure Manual: PCDDs,
717 PCDFs, cPCBs and ortho-substituted PCBs, in: Health, E. (Ed.).

718 Centre for Diseases Control and Prevention, 2016. Laboratory Procedure Manual:
719 Polychlorinated Dibenzo-p-dioxins and Furans (PCDD/F) and co-planar
720 Polychlorinated Biphenyls (cPCB) in Serum Matrix. Solid-Phase Extraction (SPE),
721 Purification by Acid-silica and Carbon, Analysis by Gas Chromatography Isotope
722 Dilution High Resolution Mass Spectrometry (GC-IDHRMS).

723 Darnerud, P.O., Aune, M., Larsson, L., Lignell, S., Mutshatshi, T., Okonkwo, J., Botha, B.,
724 Agyei, N., 2011. Levels of Brominated Flame Retardants and other Persistent Organic
725 Pollutants in Breast Milk Samples from Limpopo province, South Africa. *Science of
726 The Total Environment* 409, 4048-4053.

727 Djien Liem, A., Furst, P., Rappe, C., 2000. Exposure of Populations to Dioxins and Related
728 Compounds. *Food Additives & Contaminants* 17, 241-259.

729 Ennaceur, S., Driss, M.R., 2010. Serum Organochlorine Pesticide and Polychlorinated
730 Biphenyl Levels measured in Delivering Women from Different Locations in Tunisia.
731 *International Journal of Environmental Analytical Chemistry* 90, 821-828.

732 EPA, U., 2010. Method 1668C Chlorinated Biphenyl Congeners in Water, Soil, Sediment, and
733 Tissue by HRGC. HRMS.

734 Fromme, H., Albrecht, M., Boehmer, S., Büchner, K., Mayer, R., Liebl, B., Wittsiepe, J., Bolte,
735 G., 2009. Intake and Body Burden of Dioxin-like Compounds in Germany: The INES
736 Study. *Chemosphere* 76, 1457-1463.

737 García-Bermejo, Á., Ábalos, M., Sauló, J., Abad, E., González, M.J., Gómara, B., 2015. Triple
738 Quadrupole Tandem Mass Spectrometry: A Real Alternative to High Resolution
739 Magnetic Sector Instrument for the Analysis of Polychlorinated Dibenzo-p-dioxins,
740 Furans and Dioxin-like Polychlorinated Biphenyls. *Analytica Chimica Acta* 889, 156-
741 165.

742 Geng, D., Jogsten, I.E., Dunstan, J., Hagberg, J., Wang, T., Ruzzin, J., Rabasa-Lhoret, R., van
743 Bavel, B., 2016. Gas Chromatography/Atmospheric Pressure Chemical
744 Ionization/Mass Spectrometry for the Analysis of Organochlorine Pesticides and
745 Polychlorinated Biphenyls in Human Serum. *Journal of Chromatography A* 1453, 88-
746 98.

747 Guo, Y.L., Lambert, G.H., Hsu, C.-C., 1995. Growth Abnormalities in the Population Exposed
748 *In Utero* and Early Postnatally to Polychlorinated Biphenyls and Dibenzofurans.
749 *Environmental Health Perspectives* 103, 117.

750 Hassine, S.B., Ameer, W.B., Gandoura, N., Driss, M.R., 2012. Determination of Chlorinated
751 Pesticides, Polychlorinated Biphenyls, and Polybrominated Diphenyl Ethers in Human
752 Milk from Bizerte (Tunisia) in 2010. *Chemosphere* 89, 369-377.

753 Henríquez-Hernández, L.A., Boada, L.D., Pérez-Arellano, J.L., Carranza, C., Ruiz-Suárez, N.,
754 Sánchez, N.J., Valerón, P.F., Zumbado, M., Camacho, M., Luzardo, O.P., 2016a.
755 Relationship of Polychlorinated Biphenyls (PCBs) with Parasitism, Iron Homeostasis,
756 and other Health Outcomes: Results from a Cross-sectional Study on Recently arrived
757 African Immigrants. *Environmental Research* 150, 549-556.

758 Henríquez-Hernández, L.A., Luzardo, O.P., Arellano, J.L.P., Carranza, C., Sánchez, N.J.,
759 Almeida-González, M., Ruiz-Suárez, N., Valerón, P.F., Camacho, M., Zumbado, M.,
760 Boada, L.D., 2016b. Different Pattern of Contamination by Legacy POPs in Two
761 Populations from the same Geographical Area but with Completely Different
762 Lifestyles: Canary Islands (Spain) vs. Morocco. *Science of The Total Environment* 541,
763 51-57.

764 Hites, R.A., 2011. Dioxins: An Overview and History. *Environmental Science & Technology*
765 45, 16-20.

766 Hong, B., Garabrant, D., Hedgeman, E., Demond, A., Gillespie, B., Chen, Q., Chang, C.-W.,
767 Towey, T., Knutson, K., Franzblau, A., 2009. Impact of WHO 2005 revised toxic

768 equivalency factors for dioxins on the TEQs in serum, household dust and soil.
769 *Chemosphere* 76, 727-733.

770 Kim, B.-H., Lee, S.-J., Kim, H.-S., Chang*, Y.-S., 2004. Determination of Polychlorinated
771 Dibenzo-p-Dioxins and Dibenzofurans and Comparison of Extraction Methods for
772 Edible Freshwater Fish and Frogs in South Korea by using a High-Resolution GC/MS.
773 *Food Additives and Contaminants* 21, 700-710.

774 Koopman-Esseboom, C., Morse, D.C., Weisglas-Kuperus, N., Lutkeschipholt, I.J., Van Der
775 Paauw, C.G., Tuinstra, L.G., Brouwer, A., Sauer, P.J., 1994. Effects of dioxins and
776 polychlorinated biphenyls on thyroid hormone status of pregnant women and their
777 infants. *Pediatric research* 36, 468.

778 Lignell, S., Aune, M., Darnerud, P.O., Stridsberg, M., Hanberg, A., Larsson, S.C., Glynn, A.,
779 2016. Maternal Body Burdens of PCDD/Fs and PBDEs are associated with Maternal
780 Serum Levels of Thyroid Hormones in Early Pregnancy: A Cross-sectional Study.
781 *Environmental Health* 15, 55.

782 Luzardo, O.P., Boada, L.D., Carranza, C., Ruiz-Suárez, N., Henríquez-Hernández, L.A.,
783 Valerón, P.F., Zumbado, M., Camacho, M., Arellano, J.L.P., 2014. Socioeconomic
784 Development as a Determinant of the Levels of Organochlorine Pesticides and PCBs
785 in the Inhabitants of Western and Central African Countries. *Science of The Total*
786 *Environment* 497, 97-105.

787 Mason, G., Zacharewski, T., Denomme, M.A., Safe, L., Safe, S., 1987. Polybrominated
788 Dibenzo-p-dioxins and Related Compounds: Quantitative in vivo and in vitro Structure-
789 activity Relationships. *Toxicology* 44, 245-255.

790 Mato, Y., Suzuki, N., Katatani, N., Kadokami, K., Nakano, T., Nakayama, S., Sekii, H.,
791 Komoto, S., Miyake, S., Morita, M., 2007. Human Intake of PCDDs, PCDFs, and
792 Dioxin like PCBs in Japan, 2001 and 2002. *Chemosphere* 67, S247-S255.

793 Megson, D., Robson, M., Jobst, K.J., Helm, P.A., Reiner, E.J., 2016. Determination of
794 Halogenated Flame Retardants using Gas Chromatography with Atmospheric Pressure
795 Chemical Ionization (APCI) and a High-resolution Quadrupole Time-of-Flight Mass
796 Spectrometer (HRqTOFMS). *Analytical chemistry* 88, 11406-11411.

797 Minitab, 2010. Statistical Software [Computer Software]. State College, PA: Minitab Inc.

798 Miyashita, C., Araki, A., Mitsui, T., Itoh, S., Goudarzi, H., Sasaki, S., Kajiwara, J., Hori, T.,
799 Cho, K., Moriya, K., 2018. Sex-related Differences in the Associations between
800 Maternal Dioxin-like Compounds and Reproductive and Steroid Hormones in Cord
801 Blood: The Hokkaido Study. *Environment International* 117, 175-185.

802 Müller, M.H.B., Polder, A., Brynildsrud, O.B., Karimi, M., Lie, E., Manyilizu, W.B., Mdegela,
803 R.H., Mokiti, F., Murtadha, M., Nonga, H.E., Skaare, J.U., Lyche, J.L., 2017.
804 Organochlorine Pesticides (OCPs) and Polychlorinated Biphenyls (PCBs) in Human
805 Breast Milk and Associated Health Risks to Nursing Infants in Northern Tanzania.
806 *Environmental Research* 154, 425-434.

807 Myers, A.L., Mabury, S.A., Reiner, E.J., 2012. Analysis of mixed halogenated dibenzo-p-
808 dioxins and dibenzofurans (PXDD/PXDFs) in soil by gas chromatography tandem mass
809 spectrometry (GC-MS/MS). *Chemosphere* 87, 1063-1069.

810 Nakajima, S., Saijo, Y., Miyashita, C., Ikeno, T., Sasaki, S., Kajiwara, J., Kishi, R., 2017. Sex-
811 specific Differences in Effect of Prenatal Exposure to Dioxin-like Compounds on
812 Neurodevelopment in Japanese Children: Sapporo Cohort Study. *Environmental*
813 *Research* 159, 222-231.

814 Needham, L.L., Grandjean, P., Heinzow, B., Jørgensen, P.J., Nielsen, F., Patterson Jr, D.G.,
815 Sjödin, A., Turner, W.E., Weihe, P., 2010. Partition of environmental chemicals
816 between maternal and fetal blood and tissues. *Environmental Science & Technology*
817 45, 1121-1126.

818 Nham Tuyet, L.T., Johansson, A., 2001. Impact of Chemical Warfare with Agent Orange on
819 Women's Reproductive Lives in Vietnam: A Pilot Study. *Reproductive Health Matters*
820 9, 156-164.

821 Ntow, W.J., 2001. Organochlorine pesticides in water, sediment, crops, and human fluids in a
822 farming community in Ghana. *Archives of environmental contamination and*
823 *toxicology* 40, 557-563.

824 Ntow, W.J., Tagoe, L.M., Drechsel, P., Kelderman, P., Gijzen, H.J., Nyarko, E., 2008.
825 Accumulation of persistent organochlorine contaminants in milk and serum of farmers
826 from Ghana. *Environmental Research* 106, 17-26.

827 Olsman, H., Engwall, M., Kammann, U., Klempt, M., Otte, J., Van Bavel, B., Hollert, H., 2007.
828 Relative Differences in Aryl Hydrocarbon Receptor-mediated Response for 18
829 Polybrominated and Mixed Halogenated Dibenzo-p-dioxins and-Furans in Cell lines
830 from Four Different Species. *Environmental Toxicology and Chemistry* 26, 2448-2454.

831 Organtini, K.L., Haimovici, L., Jobst, K.J., Reiner, E.J., Ladak, A., Stevens, D., Cochran, J.W.,
832 Dorman, F.L., 2015a. Comparison of atmospheric pressure ionization gas
833 chromatography-triple quadrupole mass spectrometry to traditional high-resolution
834 mass spectrometry for the identification and quantification of halogenated dioxins and
835 furans. *Analytical chemistry* 87, 7902-7908.

836 Organtini, K.L., Myers, A.L., Jobst, K.J., Reiner, E.J., Ross, B., Ladak, A., Mullin, L., Stevens,
837 D., Dorman, F.L., 2015b. Quantitative Analysis of Mixed Halogen Dioxins and Furans
838 in Fire Debris Utilizing Atmospheric Pressure Ionization Gas Chromatography-Triple
839 Quadrupole Mass Spectrometry. *Analytical chemistry* 87, 10368-10377.

840 Papadopoulou, E., Kogevinas, M., Botsivali, M., Pedersen, M., Besselink, H., Mendez, M.A.,
841 Fleming, S., Hardie, L.J., Knudsen, L.E., Wright, J., Agramunt, S., Sunyer, J., Granum,
842 B., Gutzkow, K.B., Brunborg, G., Alexander, J., Meltzer, H.M., Brantsæter, A.L., Sarri,
843 K., Chatzi, L., Merlo, D.F., Kleinjans, J.C., Haugen, M., 2014. Maternal Diet, Prenatal
844 Exposure to Dioxin-like Compounds and Birth Outcomes in a European Prospective
845 Mother-Child Study (NewGeneris). *Science of The Total Environment* 484, 121-128.

846 Patterson, J., Donald G, Turner, W.E., Caudill, S.P., Needham, L.L., 2008. Total TEQ
847 Reference Range (PCDDs, PCDFs, cPCBs, mono-PCBs) for the US Population 2001-
848 2002. *Chemosphere* 73, S261-S277.

849 Patterson, J., Donald G, Wong, L.-Y., Turner, W.E., Caudill, S.P., DiPietro, E.S., McClure,
850 P.C., Cash, T.P., Osterloh, J.D., Pirkle, J.L., Sampson, E.J., 2009. Levels in the US
851 Population of those Persistent Organic Pollutants (2003- 2004) included in the
852 Stockholm Convention or in other Long-range Transboundary Air Pollution
853 Agreements. *Environmental Science & Technology* 43, 1211-1218.

854 Pieters, R., Focant, J.-F., 2014. Dioxin, Furan and PCB Serum Levels in a South African
855 Tswana Population: comparing the Polluting Effects of using Different Cooking and
856 Heating Fuels. *Environment International* 66, 71-78.

857 Porta, M., Puigdomènech, E., Ballester, F., Selva, J., Ribas-Fitó, N., Llop, S., López, T., 2008.
858 Monitoring Concentrations of Persistent Organic Pollutants in the General Population:
859 The International Experience. *Environment International* 34, 546-561.

860 Pryor, J.L., Hughes, C., Foster, W., Hales, B.F., Robaire, B., 2000. Critical Windows of
861 Exposure for Children's Health: The Reproductive System in Animals and Humans.
862 *Environmental Health Perspectives* 108, 491-503.

863 Röllin, H.B., Sandanger, T.M., Hansen, L., Channa, K., Odland, J.Ø., 2009. Concentration of
864 Selected Persistent Organic Pollutants in Blood from Delivering Women in South
865 Africa. *Science of The Total Environment* 408, 146-152.

- 866 Schecter, A., Päpke, O., Ball, M., 1990. Evidence for Transplacental Transfer of Dioxins from
867 Mother to Fetus: Chlorinated Dioxin and Dibenzofuran Levels in the Livers of Stillborn
868 Infants. *Chemosphere* 21, 1017-1022.
- 869 Schecter, A., Päpke, O., Harris, T.R., Tung, K., Musumba, A., Olson, J., Birnbaum, L., 2006.
870 Polybrominated Diphenyl Ether (PBDE) Levels in an Expanded Market Basket Survey
871 of US Food and Estimated PBDE Dietary Intake by Age and Sex. *Environmental Health
872 Perspectives* 114, 1515.
- 873 Schecter, A., Startin, J., Wright, C., Päpke, O., Ball, M., Lis, A., 1996. Concentrations of
874 Polychlorinated dibenzo-p-dioxins and dibenzofurans in Human Placental and Fetal
875 Tissues from the U.S. and in Placentas from Yu-Cheng Exposed Mothers.
876 *Chemosphere* 32, 551-557.
- 877 Stubleski, J., Kukucka, P., Salihovic, S., Lind, P.M., Lind, L., Kärman, A., 2018. A Method
878 for Analysis of Marker Persistent Organic Pollutants in Low-volume Plasma and Serum
879 Samples using 96-well plate Solid Phase Extraction. *Journal of Chromatography A*.
- 880 Suzuki, G., Nakano, M., Nakano, S., 2005. Distribution of PCDDs/PCDFs and Co-PCBs in
881 Human Maternal Blood, Cord Blood, Placenta, Milk, and Adipose tissue: Dioxins
882 showing High Toxic Equivalency Factor Accumulate in the Placenta. *Bioscience,
883 Biotechnology, and Biochemistry* 69, 1836-1847.
- 884 Szlinder-Richert, J., Barska, I., Usydus, Z., Ruczyńska, W., Grabic, R., 2009. Investigation of
885 PCDD/Fs and dl-PCBs in Fish from the Southern Baltic Sea during the 2002–2006
886 Period. *Chemosphere* 74, 1509-1515.
- 887 Tard, A., Gallotti, S., Leblanc, J.-C., Volatier, J.-L., 2007. Dioxins, Furans and Dioxin-like
888 PCBs: Occurrence in Food and Dietary Intake in France. *Food Additives and
889 Contaminants* 24, 1007-1017.
- 890 Tue, N.M., Goto, A., Takahashi, S., Itai, T., Asante, K.A., Kunisue, T., Tanabe, S., 2016.
891 Release of Chlorinated, Brominated and Mixed Halogenated Dioxin-related
892 Compounds to Soils from Open Burning of E-waste in Agbogboshie (Accra, Ghana).
893 *Journal of Hazardous Materials* 302, 151-157.
- 894 van Bavel, B., Geng, D., Cherta, L., Nacher-Mestre, J., Portolés, T., Ábalos, M., Sauló, J.,
895 Abad, E., Dunstan, J., Jones, R., Kotz, A., Winterhalter, H., Malisch, R., Traag, W.,
896 Hagberg, J., Ericson Jogsten, I., Beltran, J., Hernández, F., 2015. Atmospheric-Pressure
897 Chemical Ionization Tandem Mass Spectrometry (APGC/MS/MS) an Alternative to
898 High-Resolution Mass Spectrometry (HRGC/HRMS) for the Determination of
899 Dioxins. *Analytical chemistry* 87, 9047-9053.
- 900 Van den Berg, M., Birnbaum, L.S., Denison, M., De Vito, M., Farland, W., Feeley, M., Fiedler,
901 H., Hakansson, H., Hanberg, A., Haws, L., Rose, M., Safe, S., Schrenk, D., Tohyama,
902 C., Tritscher, A., Tuomisto, J., Tysklind, M., Walker, N., Peterson, R.E., 2006. The
903 2005 World Health Organization Reevaluation of Human and Mammalian Toxic
904 Equivalency Factors for Dioxins and Dioxin-Like Compounds. *Toxicological Sciences*
905 93, 223-241.
- 906 van den Berg, M., Kypke, K., Kotz, A., Tritscher, A., Lee, S.Y., Magulova, K., Fiedler, H.,
907 Malisch, R., 2017. WHO/UNEP global surveys of PCDDs, PCDFs, PCBs and DDTs in
908 human milk and benefit–risk evaluation of breastfeeding. *Archives of toxicology* 91,
909 83-96.
- 910 Wang, S.-L., Lin, C.-Y., Leon Guo, Y., Lin, L.-Y., Chou, W.-L., Chang, L.W., 2004. Infant
911 Exposure to Polychlorinated dibenzo-p-dioxins, dibenzofurans and biphenyls
912 (PCDD/Fs, PCBs)-correlation between Prenatal and Postnatal Exposure. *Chemosphere*
913 54, 1459-1473.

- 914 White, S.S., Birnbaum, L.S., 2009. An Overview of the Effects of Dioxins and Dioxin-like
915 Compounds on Vertebrates, as Documented in Human and Ecological Epidemiology.
916 Journal of Environmental Science and Health, Part C 27, 197-211.
- 917 Wittsiepe, J., Fobil, J.N., Till, H., Burchard, G.-D., Wilhelm, M., Feldt, T., 2015. Levels of
918 Polychlorinated Dibenzo-p-dioxins, Dibenzofurans (PCDD/Fs) and Biphenyls (PCBs)
919 in Blood of Informal E-waste Recycling Workers from Agbogbloshie, Ghana, and
920 Controls. Environment International 79, 65-73.
- 921 Wittsiepe, J., Schrey, P., Lemm, F., Eberwein, G., Wilhelm, M., 2008. Polychlorinated
922 Dibenzo-p-dioxins/Polychlorinated Dibenzofurans (PCDD/Fs), Polychlorinated
923 Biphenyls (PCBs), and Organochlorine Pesticides in Human Blood of Pregnant Women
924 from Germany. Journal of Toxicology and Environmental Health, Part A 71, 703-709.
- 925 Yamashita, F., Hayashi, M., 1985. Fetal PCB syndrome: Clinical Features, Intrauterine Growth
926 Retardation and Possible Alteration in Calcium Metabolism. Environmental Health
927 Perspectives 59, 41.
- 928 Zhou, Z., Dionisio, K.L., Verissimo, T.G., Kerr, A.S., Coull, B., Arku, R.E., Koutrakis, P.,
929 Spengler, J.D., Hughes, A.F., Vallarino, J., 2013. Chemical Composition and Sources
930 of Particle Pollution in Affluent and Poor Neighborhoods of Accra, Ghana.
931 Environmental Research Letters 8, 044025.

932