



Edinburgh Research Explorer

The Functional Role of Spontaneously Opening GABAA **Receptors in Neural Transmission**

Citation for published version:

O'Neill, N & Sylantyev, S 2019, 'The Functional Role of Spontaneously Opening GABAA Receptors in Neural Transmission' Frontiers in Molecular Neuroscience, vol. 12. DOI: 10.3389/fnmol.2019.00072

Digital Object Identifier (DOI):

10.3389/fnmol.2019.00072

Link:

Link to publication record in Edinburgh Research Explorer

Document Version:

Publisher's PDF, also known as Version of record

Published In:

Frontiers in Molecular Neuroscience

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh Has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh Has made every reasonable effort to ensure that Edinburgh Research Explorer
The University of Edinburgh Research Re content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.







The Functional Role of Spontaneously Opening GABA_A Receptors in Neural Transmission

Nathanael O'Neill and Sergiy Sylantyev*

Center for Clinical Brain Sciences, University of Edinburgh, Edinburgh, United Kingdom

Ionotropic type of γ-aminobutyric acid receptors (GABAARs) produce two forms of inhibitory signaling: phasic inhibition generated by rapid efflux of neurotransmitter GABA into the synaptic cleft with subsequent binding to GABAARs, and tonic inhibition generated by persistent activation of extrasynaptic and/or perisynaptic GABAARs by GABA continuously present in the extracellular space. It is widely accepted that phasic and tonic GABAergic inhibition is mediated by receptor groups of distinct subunit composition and modulated by different cytoplasmic mechanisms. Recently, however, it has been demonstrated that spontaneously opening GABA_ARs (s-GABA_ARs), which do not need GABA binding to enter an active state, make a significant input into tonic inhibitory signaling. Due to GABA-independent action mode, s-GABAARs promise new safer options for therapy of neural disorders (such as epilepsy) devoid of side effects connected to abnormal fluctuations of GABA concentration in the brain. However, despite the potentially important role of s-GABAARs in neural signaling, they still remain out of focus of neuroscience studies, to a large extent due to technical difficulties in their experimental research. Here, we summarize present data on s-GABAARs functional properties and experimental approaches that allow isolation of s-GABAARs effects from those of conventional (GABA-dependent) GABAARs.

OPEN ACCESS

Edited by:

Andrea Barberis, Istituto Italiano di Tecnologia, Italy

Reviewed by:

Tija Jacob, University of Pittsburgh, United States Paul Andrew Davies, Tufts University School of Medicine, United States

*Correspondence:

Sergiy Sylantyev s.sylantyev@ed.ac.uk orcid.org/0000-0002-1358-0601

> Received: 22 January 2019 Accepted: 08 March 2019 Published: 28 March 2019

Citation:

O'Neill N and Sylantyev S (2019) The Functional Role of Spontaneously Opening GABA_A Receptors in Neural Transmission. Front. Mol. Neurosci. 12:72. doi: 10.3389/fnmol.2019.00072 Keywords: GABA-A receptor, GABA-independent inhibition, phasic conductance, tonic conductance, G-proteins

INTRODUCTION

Ionotropic receptors of γ -aminobutyric acid (GABA receptors of type A, GABA_Rs) are the main receptor type that generates inhibitory interneuronal signaling in the brain. The classical form of GABA_R-induced inhibitory signal is phasic inhibition: a short synchronized opening of GABA_Rs in a synapse, generated by the binding of GABA released from a presynaptic terminal. However, there is an alternative form of inhibition: charge transfer through continuously active GABA_Rs, or tonic inhibition, detected in peripheral nervous system in the 1970s (Brown, 1979) but documented for the central nervous system only in the 1990s (Otis et al., 1991; Brickley et al., 1996). The classical view is that tonic inhibition is generated in response to GABA, which is continuously present in the extracellular space of neural tissue due to spillover from synapses or release from astroglia and/or neurogliaform cells (Farrant and Nusser, 2005; Kozlov et al., 2006; Oláh et al., 2009). This implies the generation of a continuous inhibitory tone mainly by perisynaptic and extrasynaptic GABA_Rs, since the vast majority of transporters which perform reverse uptake of GABA are localized in synapses or in their immediate vicinity (Minelli et al., 1996; Chiu et al., 2002; Conti et al., 2004). Hence, the magnitude of tonic GABA_Rs-delivered current is considered to be regulated by the

1

availability of extracellular GABA, and by the quantity of GABA_ARs at an extrasynaptic surface of a given neuron (Glykys and Mody, 2007). Later research, however, revealed that a significant part of tonic inhibition mediated by GABA_ARs is independent of GABA binding, i.e., it is delivered by spontaneously opening GABA_ARs (s-GABA_ARs). s-GABA_ARs in that study were shown to be insensitive to the competitive GABA antagonist SR-95531 (SR), but could be suppressed by the GABA_AR open channel blocker picrotoxin (PTX), and, to the less extent, by competitive GABA antagonist bicuculline (BIC; McCartney et al., 2007).

In the last few decades, studies of GABA_ARs-mediated tonic currents have attracted a considerable interest, and have described a functional role of this form of inhibition in a number of brain areas; in particular, its important input into neural excitability, synaptic plasticity, neurogenesis and network oscillations (Mody and Pearce, 2004; Farrant and Nusser, 2005; Glykys and Mody, 2007). Since our understanding of underlying mechanisms is still far from excellent, the newly discovered type of tonic conductance delivered *via* s-GABA_ARs promises a conceptual breakthrough in the field. Nevertheless, despite the phenomenon of GABA-independent gating of GABA_ARs being reported in numerous publications (Neelands et al., 1999; Birnir et al., 2000; Maksay et al., 2003; Miko et al., 2004), until recently the functional role of s-GABA_ARs in living neural tissue has remained beyond the focus of neuroscience research.

In this article, we try to summarize the data available to date on s-GABA_ARs function in neural transmission and to discuss perspective directions for further studies which should clarify the role of s-GABA_ARs under normal conditions and in pathology.

FUNCTIONAL PROPERTIES OF s-GABARS

s-GABARs: Problem of the Isolation of GABA-Independent Effects

One of the main factors which prevent a detailed study of s-GABA_ARs functioning is a lack of specific pharmacological

tools: the independence of s-GABA_ARs gating from GABA binding makes impossible the use of competitive GABA antagonists for selective s-GABA_ARs silencing, whereas allosteric modulators such as benzodiazepines display a lack of specificity, tuning both GABA-dependent and GABA-independent effects (Bianchi and Macdonald, 2001; McCartney et al., 2007; Gerak, 2009).

Hence, to clarify the input of s-GABAARs into a given effect, differences in molecular mechanisms of SR- and PTX-induced GABAARs silencing have been used. SR is a competitive antagonist and thus negates GABAAR activity induced by GABA binding (i.e., it acts on conventional GABAARs); in contrast, PTX binds inside the GABAAR ion channel, and thus blocks all open channels, independently of the presence of GABA binding (i.e., it acts on both conventional GABAARs and s-GABAARs). Therefore, conventional GABAAR activity can be assessed as the change in the given effect obtained in the control vs. after application of SR, whereas s-GABAAR activity can be measured as the change in the effect obtained after SR application vs. after subsequent application of SR+PTX (Wlodarczyk et al., 2013)-see Figure 1. SR is a "silent" competitor for the GABA-binding site, i.e., it does not display inverse agonist properties. Obviously, competitive antagonists such as BIC, which display inverse agonism, cannot be used for the quantitative assessment of s-GABAARs effects: BIC was shown not only to suppress synaptic events as SR does but also to induce an outward shift of holding current (Wlodarczyk et al., 2013).

s-GABARs Single-Channel Properties

The obvious step in the biophysical characterization of different subgroups of ionotropic receptors is a dissection of single-channel properties, such as electrical conductance, opening frequency and average open time. Single-channel recordings have repeatedly demonstrated similar or very close conductance values for s-GABA_ARs and conventional GABA_ARs (Mathers, 1985; Neelands et al., 1999; Birnir et al., 2000;

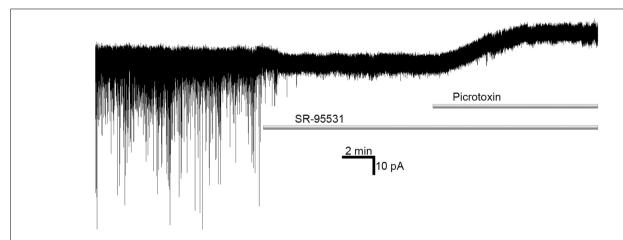


FIGURE 1 | Competitive γ-aminobutyric acid (GABA) antagonist SR-95531 suppresses spontaneous GABA-ergic synaptic signaling, but does not affect tonic conductance; on the contrary, open-channel blocker picrotoxin applied after SR-95531 shuts spontaneously opening GABA-receptors (s-GABA_ARs), revealing the amount of inhibitory current passing through s-GABA_ARs independently of GABA binding.

O'Neill and Sylantyev, 2018a,b) thus making this parameter hardly applicable for distinguishing between two receptor subtypes. Similarly, the dependence of GABAARs opening frequency on the concentration of GABA, makes this parameter inapplicable for discrimination of effects of s-GABAARs and conventional GABAARs in single-channel recordings. In contrast, the average open time was found to be significantly lower for s-GABAARs than for conventional GABAARs. This generates a two-peak distribution of opening time values under physiological conditions when free GABA is present in extracellular space (O'Neill and Sylantyey, 2018a). Earlier observations demonstrated that the two-peak Gaussian distribution of average open times is a characteristic feature of GABAARs of at least three different subunit compositions (Mortensen et al., 2010). It is important to note that the mode values for shorter durations in that work were found to be similar, irrespective of the agonist's type and concentration, thus representing an agonist-independent input. This suggests that: (i) s-GABAARs activity is a common element of integral GABAAR response; and (ii) that s-GABAARs represent a functionally similar receptor subgroup composed of receptors of various subunit compositions.

Another method of distinguishing between s-GABA_ARs and conventional GABA_ARs at a level of single-channel effects may potentially develop from the recent observation about the ability of benzodiazepine flurazepam to modulate GABA-dependent and GABA-independent GABA_AR gating *via* different molecular mechanisms (Jatczak-Śliwa et al., 2018).

s-GABARs Input Into Tonic Conductance

Overall, charge transfer with phasic events mediated by GABA_ARs (and induced by GABA binding) compared to that delivered by tonic conductance through GABA_ARs, displays a ratio of more than 9/1 (Cope et al., 2005; O'Neill and Sylantyev, 2018a). Taking into account that GABA-induced tonic current was found to be negligible under physiological concentrations of extracellular GABA, whereas under these conditions s-GABA_ARs generated a significant amount of tonic current (Wlodarczyk et al., 2013), s-GABA_ARs should be considered as a potential key element in the generation of lasting inhibitory tone and, in a wider context, in inter-neuronal crosstalk.

Tonic inhibition has been widely accepted to be a strong modulator of action potential (AP) generation (Hamann et al., 2002; Bonin et al., 2007), AP firing patterns (Häusser and Clark, 1997) and the coincidence detection time window for synaptic inputs (Tang et al., 2011). Experiments on s-GABAARs have readily confirmed their significant input into the regulation of the following phenomena: the modulation of AP generation (O'Neill and Sylantyev, 2018b), firing patterns (Botta et al., 2015; O'Neill and Sylantyev, 2018a), neurons' rheobase, and the time window of coincidence detection of excitatory inputs (O'Neill and Sylantyev, 2018a).

s-GABA_ARs Input Into Phasic Conductance

Several classical studies have demonstrated that $GABA_ARs$ of specific subunit compositions (e.g., δ - $GABA_ARs$) which may be

responsible for a lion's share of tonic current (Nusser and Mody, 2002; Stell et al., 2003; Mortensen et al., 2010) are localized exclusively at the extrasynaptic membrane (Nusser et al., 1998; Wei et al., 2003). However, if s-GABAARs are a functionally similar group of receptors of different subunit composition (see "s-GABARs Single-Channel Properties" section), their absence in synapses would be highly doubtful. This, in turn, raises a question as to how (and whether) s-GABAARs modify synaptic (phasic) GABA-ergic inhibitory responses (inhibitory post-synaptic currents, IPSCs). In truth, recent studies have demonstrated their significant input into IPSC decay kinetics: s-GABAARs introduced a slow element of decay profile (O'Neill and Sylantyev, 2018a), probably due to their higher potency to GABA (Yeung et al., 2003) and/or modified receptor efficacy.

It was shown earlier that GABAAR-generated IPSC may contain fast and slow components with different sensitivities to GABA competitive antagonists, which resembles the functional profile of s-GABAARs (Kapur et al., 1997). In this research, the generation of fast and slow components of whole-cell IPSC was attributed to different cell regions: dendritic and somatic, respectively. On the other hand, later direct recordings of s-GABAARs activity confirmed a significant input of this receptor subtype into both whole-cell IPSCs (which are generated in synapses), and into IPSCs evoked in nucleated membrane patches, i.e., generated by GABAARs localized at a neural cell soma (O'Neill and Sylantyev, 2018a). On top of that, a significant input of δ-GABAARs into IPSCs was recently demonstrated (Sun et al., 2018), which confirms once again both the synaptic and extrasynaptic localization of GABAARs which display high tonic activity.

Intracellular Regulatory Mechanisms of s-GABA_ARs Activity

The particular intracellular mechanisms which are used by neural cells to modulate the activity of GABAARs are still far from being completely understood; however, it has long been established that direct phosphorylation is of major importance (Brandon et al., 2002). It was shown that GABAARs functions can be modulated differentially (potentiated or suppressed) depending on the receptor subunit composition, the type of neuron, et cetera by cAMP-dependent protein kinase A (PKA), tyrosine kinase Src and PKC: refer to Brandon et al. (2002) for review. In particular, GABAAR-mediated tonic inhibitory currents were shown to be downregulated by PKC Bright and Smart, 2013, whereas PKA was found to enhance this type of inhibition (Carlson et al., 2016). In addition, GABAARs effects were repeatedly shown to be modulated by G-protein-coupled receptors via G-proteins of different types (Cai et al., 2002; Wang et al., 2002) which are, in turn, tightly connected to the regulation of PKC and PKA activity (Neves et al., 2002). Hence, the clarification of impact on s-GABAARs function delivered by intracellular regulatory factors (specifically, by various kinases and G-proteins), is one of the key steps needed for understanding and predicting s-GABAARs functional input into a neural transmission.

To date, there is little data on this. It has been demonstrated that in dentate gyrus granule cells of hippocampus PKC regulates tonic GABA-dependent inhibitory conductance but has no significant impact on the GABA-independent effects of s-GABAARs (O'Neill and Sylantyev, 2018b). However, at a longer time scale it was repeatedly shown that PKC and Ca²⁺/calmodulin-dependent protein kinase II increase tonic inhibition in hippocampus and amygdala due to enhanced phosphorylation and membrane insertion of β3-containing GABAARs (Saliba et al., 2012; Modgil et al., 2017) and α4-containing GABAARs; this PKC action can be potentiated by neurosteroids such as THDOC (Abramian et al., 2010, 2014; Romo-Parra et al., 2015). In turn, s-GABAARs-mediated tonic inhibition in dentate gyrus granule cells is controlled by G-proteins: non-specific block of G-proteins by pertussis toxin decreases the tonic current via the reduction of the s-GABAARs opening frequency (O'Neill and Sylantyey, 2018b).

In contrast to PKC, activation of PKA was found to increase the tonic current through $\alpha 4\beta 3\delta$ and, to a lesser extent, $\alpha 4\beta 3\gamma 2L$ -GABA_ARs in absence of GABA due to upregulation of single-channel opening frequency. Addition of GABA to an ambient solution, however, gradually decreased the sensitivity of GABA_ARs of both subunit compositions to modulation by PKA; such a modulation became insignificant when GABA concentration reached micromolar values (Tang et al., 2010).

It is important to note, however, that a significant part of GABA-independent s-GABA_ARs activity was found to be out of the control of any soluble cytoplasmic factors. GABA-independent openings of GABA_ARs were recorded from outside-out patches excised from dentate gyrus granule cells somata: in this preparation, all cytoplasmic signaling chains are surely destroyed (O'Neill and Sylantyev, 2018b). However, anchored kinases that modulate ionotropic receptors (Brandon et al., 2003; Carnegie and Scott, 2003) may still be responsible for at least a part of the s-GABA_ARs activity observed in outside-out patches.

CONCLUSIONS AND FURTHER RESEARCH DIRECTIONS

To date, there have been only a few publications highlighting the functional properties of s-GABAARs in living neurons. This imposes obvious limitations on conclusions in terms of the applicability for different brain regions and types of neurons. Nevertheless, the significant input of s-GABAARs into the modulation of output signal generation and into the integration of input signaling in a given neuron, suggests that s-GABAAR activity is one of the key actors that regulate neural inhibition.

Indeed, the relative importance of GABA-independent s-GABA_ARs signaling in a given region of the brain depends critically on the native concentration of GABA in the extracellular space. Different groups report *in vivo* concentrations varying by more than an order of magnitude: from less than 100 (Wlodarczyk et al., 2013) or 200 (Glaeser and Hare, 1975) nM to units of micromoles (Tossman et al., 1986; Takagi et al., 1993). Moreover, there may be local inhomogeneities of GABA concentrations due to cell-specific

differences in the distribution and/or activity of GABA transporters and the elements of the GABA synthesis system. This was indirectly confirmed by the observation that the silencing of GAD-65 activity reduces tonic inhibitory currents in interneurons, but not in the pyramidal neurons of the hippocampal CA1 area (Song et al., 2011). A recent study on the hippocampus has demonstrated that at a GABA concentration of ~100 nM, the amount of GABA-induced tonic current (which can be suppressed by SR) is close to statistical noise (see example at Figure 1), and negligible when compared to that through GABA-independent openings of s-GABAARs (Wlodarczyk et al., 2013); on the contrary, SR has been shown to reveal a huge amount of tonic GABA-dependent current in thalamus (Cope et al., 2005). These data suggest that the relative impact of s-GABAARs into neural signaling varies widely, depending on the particular brain region and cell type. To the best of our knowledge, previous articles that discuss lower EC50 values (i.e., higher potency) of extrasynaptic GABAARs in vivo do not consider spontaneous channels and how they influence such measurements. This fact enforces the importance of the work on s-GABAARs pharmacology for an understanding of biophysical phenomena in living neurons.

The important question regarding s-GABAARs is whether or not these receptors represent a convergent group with similar functional properties, or if they share common receptor subunit(s). Numerous studies have attributed the majority (up to 75%) of GABA_AR-delivered tonic inhibition to δ-containing GABA_ARs (Stell et al., 2003), which are abundant at extrasynaptic membranes (Nusser et al., 1998) but have been also found in synapses where they make a significant input into phasic inhibition (Sun et al., 2018), and in perisynaptic loci (Wei et al., 2003). The remaining portion of tonic inhibition is, to a large extent but not fully, produced by receptors containing the α5-subunit (Farrant and Nusser, 2005). Furthermore, the agonist-independent GABAAR openings were observed under similar conditions for receptors of three different subunit compositions (Mortensen et al., 2010). In addition, the observation that mutations in $\alpha 1$ and $\beta 2$ subunits modulate spontaneous GABAARs gating (Baptista-Hon et al., 2017) prevents us from ruling out these subunits as potential alternative candidates to be involved in the formation of s-GABAARs. Combined with the facts of the GABA-independent tonic activity of $\alpha 4$ -GABA_ARs (Tang et al., 2010) and spontaneous openings of $\alpha 2\beta 1\epsilon$ -GABAARs which contribute to the baseline currents in whole-cell recordings (Wagner et al., 2005), the abovementioned data on GABA-independent activity suggest that GABA-independent inhibition is of poly-subtype origin, with a substantial part inherent in the non- δ - and non- α 5containing receptors.

In view of numerous subunits and subunit compositions of GABA_AR which demonstrate spontaneous gating, the obvious question is: are there GABA_ARs subtype(s) which do not demonstrate GABA-independent activity? The existence of such GABA_ARs was suggested by the study showing that, in contrast to the $\alpha 2\alpha 1\epsilon$ receptor, responses of $\alpha 2\beta 1$ and $\alpha 2\beta 1\gamma 2$ -GABA_ARs do not produce a "baseline overshoot" associated with spontaneous openings (Wagner et al., 2005).

Therefore, data collected to date suggest revision of two traditional views, now common in fundamental neuroscience: (i) that tonic inhibitory conductance is generated by ambient GABA (due to proven significance of s-GABA_ARs input); and (ii) that tonic and phasic inhibition are mediated by different GABA_ARs subtypes (due to growing evidence that typical extrasynaptic GABA_ARs can make a significant contribution into IPSCs *via* a synaptic and/or perisynaptic presence).

It has been demonstrated that a scarcity of $\alpha 1$ subunit is correlated with resistance to anti-epileptic drugs (Bethmann et al., 2008), whereas increased α1-GABAAR expression in the hippocampus suppresses the development of temporal lobe epilepsy (TLE; Raol et al., 2006). Apart from that, it was shown that phasic GABA-ergic inhibition is lowered in TLE, whereas tonic GABA-ergic conductance remains intact (Palma et al., 2007; Pavlov et al., 2011), making tonic GABA-ergic current a perspective target for TLE treatment. The classical paradigm, where extracellular GABA triggers tonic GABA-ergic current, implies that the most effective therapeutic approach is to increase the concentration of GABA in the cerebrospinal fluid, and thus augment inhibitory conductance. However, this approach was repeatedly found to be ineffective (Cohen et al., 2002; Glykys et al., 2009) or even one that leads to epileptogenesis (Palma et al., 2006; Cope et al., 2009) due to various side effects. These side effects impose limitations on the clinical use of specific antiepileptic drugs that increase the concentration of GABA in cerebrospinal fluid (Sander and Hart, 1990; Leppik, 1995). In contrast, the modulation of s-GABAARs in GABA-independent manner promises an alternative for TLE treatment through the regulation of tonic conductance without the need to interfere with extracellular GABA concentration, thus avoiding the afore mentioned side effects.

Apart from the potential of $\alpha 1$ -GABA_ARs for TLE treatment, $\alpha 5$ -GABA_ARs (which also display GABA-independent activity) were found to be a perspective target for schizophrenia treatment (Lodge and Grace, 2011). Taking into account similar concentration of GABA found *in vivo* in the brains of schizophrenic patients and of a control group (Tayoshi et al., 2010), and the well-established fact that changes in tonic GABA-ergic inhibition are involved in the generation of schizophrenia symptoms (Damgaard et al., 2011), these data suggest a potentially important role of drugs targeting s-GABA_ARs in the suppression of schizophrenia development, since action through s-GABA_ARs in GABA-independent manner eliminates the need to modify GABA concentration in cerebrospinal fluid.

Another clinical implication of s-GABA $_A$ Rs rises from the fact that sedative and analgesic effects of gaboxadol (THIP) are mediated exclusively by $\alpha 4$ -containing GABA $_A$ Rs (Chandra et al., 2006), that demonstrate GABA-independent activity.

AUTHOR CONTRIBUTIONS

NO and SS contributed to the conception and design of the article. NO received data displayed at the figure and analyzed literature connected to the topic, contributed to manuscript revision. SS wrote the manuscript.

FUNDING

This work was supported by the Rosetrees Research Grant A-1066.

REFERENCES

- Abramian, A. M., Comenencia-Ortiz, E., Vithlani, M., Tretter, E. V., Sieghart, W., Davies, P. A., et al. (2010). Protein kinase C phosphorylation regulates membrane insertion of GABA_A receptor subtypes that mediate tonic inhibition. J. Biol. Chem. 285, 41795–41805. doi: 10.1074/jbc.m110.149229
- Abramian, A. M., Comenencia-Ortiz, E., Modgil, A., Vien, T. N., Nakamura, Y., Moore, Y. E., et al. (2014). Neurosteroids promote phosphorylation and membrane insertion of extrasynaptic GABA_A receptors. *Proc. Natl. Acad. Sci.* USA 111, 7132–7137. doi: 10.1073/pnas.1403285111
- Baptista-Hon, D. T., Gulbinaite, S., and Hales, T. G. (2017). Loop G in the GABAA receptor $\alpha 1$ subunit influences gating efficacy. *J. Physiol.* 595, 1725–1741. doi: 10.1113/jp273752
- Bethmann, K., Fritschy, J. M., Brandt, C., and Löscher, W. (2008). Antiepileptic drug resistant rats differ from drug responsive rats in GABA_A receptor subunit expression in a model of temporal lobe epilepsy. *Neurobiol. Dis.* 31, 169–187. doi: 10.1016/j.nbd.2008.01.005
- Bianchi, M. T., and Macdonald, R. L. (2001). Agonist trapping by GABA_A receptor channels. *J. Neurosci.* 21, 9083–9091. doi: 10.1523/jneurosci.21-23-09083.2001
- Birnir, B., Everitt, A. B., Lim, M. S., and Gage, P. W. (2000). Spontaneously opening GABA_A channels in CA1 pyramidal neurones of rat hippocampus. *J. Membr. Biol.* 174, 21–29. doi: 10.1007/s002320001028
- Bonin, R. P., Martin, L. J., MacDonald, J. F., and Orser, B. A. (2007). α5GABA_A receptors regulate the intrinsic excitability of mouse hippocampal pyramidal neurons. J. Neurophysiol. 98, 2244–2254. doi: 10.1152/jn.00482.2007
- Botta, P., Demmou, L., Kasugai, Y., Markovic, M., Xu, C., Fadok, J. P., et al. (2015).
 Regulating anxiety with extrasynaptic inhibition. *Nat. Neurosci.* 18, 1493–1500.
 doi: 10.1038/nn.4102

- Brandon, N., Jovanovic, J., and Moss, S. (2002). Multiple roles of protein kinases in the modulation of gamma-aminobutyric acid_(A) receptor function and cell surface expression. *Pharmacol. Ther.* 94, 113–122. doi: 10.1016/s0163-7258(02)00175-4
- Brandon, N. J., Jovanovic, J. N., Colledge, M., Kittler, J. T., Brandon, J. M., Scott, J. D., et al. (2003). A-kinase anchoring protein 79/150 facilitates the phosphorylation of GABA_A receptors by cAMP-dependent protein kinase via selective interaction with receptor β subunits. *Mole. Cell. Neurosci.* 22, 87–97. doi: 10.1016/s1044-7431(02)00017-9
- Brickley, S. G., Cull-Candy, S. G., and Farrant, M. (1996). Development of a tonic form of synaptic inhibition in rat cerebellar granule cells resulting from persistent activation of GABA_A receptors. *J Physiol.* 497, 753–759. doi: 10.1113/jphysiol.1996.sp021806
- Bright, D. P., and Smart, T. G. (2013). Protein kinase C regulates tonic GABA_A receptor-mediated inhibition in the hippocampus and thalamus. *Eur. J. Neurosci.* 38, 3408–3423. doi: 10.1111/ejn.12352
- Brown, D. A. (1979). Extrasynaptic GABA systems. Trends Neurosci. 2, 271–273. doi: 10.1016/0166-2236(79)90107-3
- Cai, X., Flores-Hernandez, J., Feng, J., and Yan, Z. (2002). Activity-dependent bidirectional regulation of GABA_A receptor channels by the 5-HT₍₄₎ receptormediated signalling in rat prefrontal cortical pyramidal neurons. *J. Physiol.* 540, 743–759. doi: 10.1113/jphysiol.2001.013391
- Carlson, S. L., Bohnsack, J. P., Patel, V., and Morrow, A. L. (2016). Regulation of extrasynaptic GABA_A α4 receptors by ethanol-induced protein kinase, A, but not protein kinase C activation in cultured rat cerebral cortical neurons. *J. Pharmacol. Exp. Ther.* 356, 148–156. doi: 10.1124/jpet.115.228056
- Carnegie, G. K., and Scott, J. D. (2003). A-kinase anchoring proteins and neuronal signaling mechanisms. Genes Dev. 17, 1557–1568. doi: 10.1101/gad.1095803

- Chandra, D., Jia, F., Liang, J., Peng, Z., Suryanarayanan, A., Werner, D. F., et al. (2006). GABA_A receptor α 4 subunits mediate extrasynaptic inhibition in thalamus and dentate gyrus and the action of gaboxadol. *Proc. Natl. Acad. Sci. U S A* 103, 15230–15235. doi: 10.1073/pnas.0604304103
- Chiu, C. S., Jensen, K., Sokolova, I., Wang, D., Li, M., Deshpande, P., et al. (2002). Number, density and surface/cytoplasmic distribution of GABA transporters at presynaptic structures of knock-in mice carrying GABA transporter subtype 1-Green fluorescent protein fusions. J. Neurosci. 22, 10251–10266. doi: 10.1523/jneurosci.22-23-10251.2002
- Cohen, I., Navarro, V., Clemenceau, S., Baulac, M., and Miles, R. (2002). On the origin of interictal activity in human temporal lobe epilepsy in vitro. Science 298, 1418–1421. doi: 10.1126/science.1076510
- Conti, F., Minelli, A., and Melone, M. (2004). GABA transporters in the mammalian cerebral cortex: localization, development and pathological implications. *Brain Res. Rev.* 45, 196–212. doi: 10.1016/j.brainresrev.2004.03. 003
- Cope, D. W., Hughes, S. W., and Crunelli, V. (2005). GABAA receptormediated tonic inhibition in thalamic neurons. *J Neurosci.* 25, 11553–11563. doi: 10.1523/jneurosci.3362-05.2005
- Cope, D. W., Di Giovanni, G., Fyson, S. J., Orbán, G., Errington, A. C., Lorincz, M. L., et al. (2009). Enhanced tonic GABA_A inhibition in typical absence epilepsy. *Nat. Med.* 15, 1392–1398. doi: 10.1038/nm.2058
- Damgaard, T., Plath, N., Neill, J. C., and Hansen, S. L. (2011). Extrasynaptic GABA_A receptor activation reverses recognition memory deficits in an animal model of schizophrenia. *Psychopharmacology* 214, 403–413. doi: 10.1007/s00213-010-2039-9
- Farrant, M., and Nusser, Z. (2005). Variations on an inhibitory theme: phasic and tonic activation of GABA_A receptors. *Nat. Rev. Neurosci.* 6, 215–229. doi: 10.1038/nrn1625
- Gerak, L. R. (2009). Selective changes in sensitivity to benzodiazepines, and not other positive GABA_A modulators, in rats receiving flunitrazepam chronically. *Psychopharmacology* 204, 667–677. doi: 10.1007/s00213-009-1497-4
- Glaeser, B. S., and Hare, T. A. (1975). Measurement of GABA in human cerebrospinal fluid. *Biochem. Med.* 12, 274–282. doi: 10.1016/0006-2944(75)90129-5
- Glykys, J., and Mody, I. (2007). Activation of GABA_A receptors: views from outside the synaptic cleft. *Neuron* 56, 763–770. doi: 10.1016/j.neuron.2007. 11.002
- Glykys, J., Dzhala, V. I., Kuchibhotla, K. V., Feng, G., Kuner, T., Augustine, G., et al. (2009). Differences in cortical versus subcortical GABAergic signaling: a candidate mechanism of electroclinical uncoupling of neonatal seizures. Neuron 63, 657–672. doi: 10.1016/j.neuron.2009.08.022
- Hamann, M., Rossi, D. J., and Attwell, D. (2002). Tonic and spillover inhibition of granule cells control information flow through cerebellar cortex. *Neuron* 33, 625–633. doi: 10.1016/s0896-6273(02)00593-7
- Häusser, M., and Clark, B. A. (1997). Tonic synaptic inhibition modulates neuronal output pattern and spatiotemporal synaptic integration. *Neuron* 19, 665–678. doi: 10.1016/s0896-6273(00)80379-7
- Jatczak-Śliwa, M., Terejko, K., Brodzki, M., Michaowski, M. A., Czyzewska, M. M., Nowicka, J. M., et al. (2018). Distinct modulation of spontaneous and GABA-evoked gating by flurazepam shapes cross-talk between agonist-free and liganded GABA_A receptor activity. Front. Cell. Neurosci. 12:237. doi: 10.3389/fncel.2018.00237
- Kapur, A., Pearce, R. A., Lytton, W. W., and Haberly, L. B. (1997). GABA_A-mediated IPSCs in piriform cortex have fast and slow components with different properties and locations on pyramidal cells. *J. Neurophysiol.* 78, 2531–2545. doi: 10.1152/jn.1997.78.5.2531
- Kozlov, A. S., Angulo, M. C., Audinat, E., and Charpak, S. (2006). Target cell-specific modulation of neuronal activity by astrocytes. *Proc. Natl. Acad.* Sci. U S A 103, 10058–10063. doi: 10.1073/pnas.0603741103
- Leppik, I. E. (1995). Tiagabine: the safety landscape. *Epilepsia* 36, S10–S13. doi: 10.1111/j.1528-1157.1995.tb06009.x
- Lodge, D. J., and Grace, A. A. (2011). Hippocampal dysregulation of dopamine system function and the pathophysiology of schizophrenia. *Trends Pharmacol.* Sci. 32, 507–513. doi: 10.1016/j.tips.2011.05.001
- Maksay, G., Thompson, S. A., and Wafford, K. A. (2003). The pharmacology of spontaneously open $\alpha 1~\beta 3~\epsilon$ GABA_A receptor-ionophores. *Neuropharmacology* 44, 994–1002. doi: 10.1016/s0028-3908(03)00116-3

- Mathers, D. A. (1985). Spontaneous and GABA-induced single channel currents in cultured murine spinal cord neurons. *Can. J. Physiol. Pharmacol.* 63, 1228–1233. doi: 10.1139/y85-203
- McCartney, M. R., Deeb, T. Z., Henderson, T. N., and Hales, T. G. (2007).
 Tonically active GABA_A receptors in hippocampal pyramidal neurons exhibit constitutive GABA-independent gating. *Mol. Pharmacol.* 71, 539–548. doi: 10.1124/mol.106.028597
- Miko, A., Werby, E., Sun, H., Healey, J., and Zhang, L. (2004). A TM2 residue in the β1 subunit determines spontaneous opening of homomeric and heteromeric γ-aminobutyric acid-gated ion channels. *J. Biol. Chem.* 279, 22833–22840. doi: 10.1074/jbc.m402577200
- Minelli, A., DeBiasi, S., Brecha, N. C., Zuccarello, L. V., and Conti, F. (1996). GAT-3, a high-affinity GABA plasma membrane transporter, is localized to astrocytic processes, and it is not confined to the vicinity of GABAergic synapses in the cerebral cortex. *J. Neurosci.* 16, 6255–6264. doi: 10.1523/jneurosci.16-19-06255.
- Modgil, A., Parakala, M. L., Ackley, M. A., Doherty, J. J., Moss, S. J., and Davies, P. A. (2017). Endogenous and synthetic neuroactive steroids evoke sustained increases in the efficacy of GABAergic inhibition via a protein kinase C-dependent mechanism. *Neuropharmacology* 113, 314–322. doi: 10.1016/j. neuropharm.2016.10.010
- Mody, I., and Pearce, R. A. (2004). Diversity of inhibitory neurotransmission through GABA_A receptors. *Trends Neurosci.* 27, 569–575. doi: 10.1016/j.tins. 2004.07.002
- Mortensen, M., Ebert, B., Wafford, K., and Smart, T. G. (2010). Distinct activities of GABA agonists at synaptic- and extrasynaptic-type GABA_A receptors. *J. Physiol.* 588, 1251–1268. doi: 10.1113/jphysiol.2009.182444
- Neelands, T. R., Fisher, J. L., Bianchi, M., and Macdonald, R. L. (1999). Spontaneous and γ-aminobutyric acid (GABA)-activated GABA_A receptor channels formed by ε subunit-containing isoforms. *Mol. Pharmacol.* 55, 168–178. doi: 10.1124/mol.55.1.168
- Neves, S. R., Ram, P. T., and Iyengar, R. (2002). G protein pathways. *Science* 296, 1636–1639. doi: 10.1126/science.1071550
- Nusser, Z., and Mody, I. (2002). Selective modulation of tonic and phasic inhibitions in dentate gyrus granule cells. J. Neurophysiol. 87, 2624–2628. doi: 10.1152/jn.2002.87.5.2624
- Nusser, Z., Sieghart, W., and Somogyi, P. (1998). Segregation of different GABA_A receptors to synaptic and extrasynaptic membranes of cerebellar granule cells. *J. Neurosci.* 18, 1693–1703. doi: 10.1523/jneurosci.18-05-01693.1998
- O'Neill, N., and Sylantyev, S. (2018a). Spontaneously opening GABA_A receptors play a significant role in neuronal signal filtering and integration. *Cell Death Dis.* 9:813. doi: 10.1038/s41419-018-0856-7
- O'Neill, N., and Sylantyev, S. (2018b). Feature Article: selective modulation of tonically active GABA_A receptor functional subgroups by G-proteins and protein kinase C. *Exp. Biol. Med.* 243, 1046–1055. doi: 10.1177/1535370218
- Oláh, S., Füle, M., Komlósi, G., Varga, C., Báldi, R., Barzó, P., et al. (2009).Regulation of cortical microcircuits by unitary GABA-mediated volume transmission. *Nature* 461, 1278–1281. doi: 10.1038/nature08503
- Otis, T. S., Staley, K. J., and Mody, I. (1991). Perpetual inhibitory activity in mammalian brain slices generated by spontaneous GABA release. *Brain Res.* 545, 142–150. doi: 10.1016/0006-8993(91)91280-e
- Palma, E., Amici, M., Sobrero, F., Spinelli, G., Di Angelantonio, S., Ragozzino, D., et al. (2006). Anomalous levels of Cl- transporters in the hippocampal subiculum from temporal lobe epilepsy patients make GABA excitatory. Proc. Natl. Acad. Sci. U S A 103, 8465–8468. doi: 10.1073/pnas.06029 79103
- Palma, E., Roseti, C., Maiolino, F., Fucile, S., Martinello, K., Mazzuferi, M., et al. (2007). GABA_A-current rundown of temporal lobe epilepsy is associated with repetitive activation of GABA_A "phasic" receptors. *Proc. Natl. Acad. Sci. U S A* 104, 20944–20948. doi: 10.1073/pnas.0710522105
- Pavlov, I., Huusko, N., Drexel, M., Kirchmair, E., Sperk, G., Pitkänen, A., et al. (2011). Progressive loss of phasic, but not tonic, GABA_A receptor-mediated inhibition in dentate granule cells in a model of post-traumatic epilepsy in rats. *Neuroscience* 194, 208–219. doi: 10.1016/j.neuroscience.2011.07.074
- Raol, Y. H., Lund, I. V., Bandyopadhyay, S., Zhang, G., Roberts, D. S., Wolfe, J. H., et al. (2006). Enhancing GABA_A receptor alpha 1 subunit levels in hippocampal dentate gyrus inhibits epilepsy development in an animal model of temporal

- lobe epilepsy. J. Neurosci. 26, 11342–11346. doi: 10.1523/jneurosci.3329-06.2006
- Romo-Parra, H., Blaesse, P., Sosulina, L., and Pape, H. C. (2015). Neurosteroids increase tonic GABAergic inhibition in the lateral section of the central amygdala in mice. J. Neurophysiol. 113, 3421–3431. doi: 10.1152/jn.00045.2015
- Saliba, R. S., Kretschmannova, K., and Moss, S. J. (2012). Activity-dependent phosphorylation of GABA_A receptors regulates receptor insertion and tonic current. EMBO J. 31, 2937–2951. doi: 10.1038/emboj.2012.109
- Sander, J. W., and Hart, Y. M. (1990). Vigabatrin and behaviour disturbances. Lancet 335:57. doi: 10.1016/0140-6736(90)90190-g
- Song, I., Savtchenko, L., and Semyanov, A. (2011). Tonic excitation or inhibition is set by GABA_A conductance in hippocampal interneurons. *Nat. Commun.* 2:376. doi: 10.1038/ncomms1377
- Stell, B. M., Brickley, S. G., Tang, C. Y., Farrant, M., and Mody, I. (2003). Neuroactive steroids reduce neuronal excitability by selectively enhancing tonic inhibition mediated by delta subunit-containing GABA_A receptors. *Proc. Natl. Acad. Sci. U S A* 100, 14439–14444. doi: 10.1073/pnas.2435457100
- Sun, M. Y., Shu, H. J., Benz, A., Bracamontes, J., Akk, G., Zorumski, C. F., et al. (2018). Chemogenetic isolation reveals synaptic contribution of δ GABA_A receptors in mouse dentate granule neurons. *J. Neurosci.* 38, 8128–8145. doi: 10.1523/jneurosci.0799-18.2018
- Takagi, K., Ginsberg, M. D., Globus, M. Y., Dietrich, W. D., Martinez, E., Kraydieh, S., et al. (1993). Changes in amino acid neurotransmitters and cerebral blood flow in the ischemic penumbral region following middle cerebral artery occlusion in the rat: correlation with histopathology. J. Cereb. Blood Flow Metab. 13, 575–585. doi: 10.1038/jcbfm.1993.75
- Tang, X., Hernandez, C. C., and Macdonald, R. L. (2010). Modulation of spontaneous and GABA-evoked tonic α4β3δ and α4β3γ2L GABA_A receptor currents by protein kinase A. J. Neurophysiol. 103, 1007–1019. doi: 10.1152/jn. 00801.2009
- Tang, Z. Q., Dinh, E. H., Shi, W., and Lu, Y. (2011). Ambient GABA-activated tonic inhibition sharpens auditory coincidence detection via a depolarizing shunting mechanism. J. Neurosci. 31, 6121–6131. doi: 10.1523/jneurosci.4733-10.2011
- Tayoshi, S., Nakataki, M., Sumitani, S., Taniguchi, K., Shibuya-Tayoshi, S., Numata, S., et al. (2010). GABA concentration in schizophrenia patients

- and the effects of antipsychotic medication: A proton magnetic resonance spectroscopy study. *Schizophr. Res.* 117, 83–91. doi: 10.1016/j.schres.2009.
- Tossman, U., Jonsson, G., and Ungerstedt, U. (1986). Regional distribution and extracellular levels of amino acids in rat central nervous system. *Acta Physiol. Scand.* 127, 533–545. doi: 10.1111/j.1748-1716.1986.tb07938.x
- Wagner, D. A., Goldschen-Ohm, M. P., Hales, T. G., and Jones, M. V. (2005). Kinetics and spontaneous open probability conferred by the ε subunit of the GABA_A receptor. J. Neurosci. 25, 10462–10468. doi: 10.1523/jneurosci.1658-05.2005
- Wang, X., Zhong, P., and Yan, Z. (2002). Dopamine D4 receptors modulate GABAergic signaling in pyramidal neurons of prefrontal cortex. J. Neurosci. 22, 9185–9193. doi: 10.1523/jneurosci.22-21-09185.2002
- Wei, W., Zhang, N., Peng, Z., Houser, C. R., and Mody, I. (2003). Perisynaptic localization of delta subunit-containing GABA_A receptors and their activation by GABA spillover in the mouse dentate gyrus. *J. Neurosci.* 23, 10650–10661. doi: 10.1523/jneurosci.23-33-10650.2003
- Wlodarczyk, A. I., Sylantyev, S., Herd, M. B., Kersanté, F., Lambert, J. J., Rusakov, D. A., et al. (2013). GABA-independent GABA_A receptor openings maintain tonic currents. *J. Neurosci.* 33, 3905–3914. doi: 10.1523/jneurosci. 4193-12.2013
- Yeung, J. Y., Canning, K. J., Zhu, G., Pennefather, P., MacDonald, J. F., and Orser, B. A. (2003). Tonically activated GABA_A receptors in hippocampal neurons are high-affinity, low-conductance sensors for extracellular GABA. *Mol. Pharmacol.* 63, 2–8. doi: 10.1124/mol.63.1.2

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 O'Neill and Sylantyev. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.