

# **UNIVERSITI PUTRA MALAYSIA**

CHARACTERIZATION OF Ralstonia solanacearum RACE 2 BIOVAR 1 ASSOCIATED WITH MOKO DISEASE OF BANANA IN PENINSULAR MALAYSIA

# DZARIFAH MOHAMED ZULPERI

FP 2015 65



## CHARACTERIZATION OF *Ralstonia solanacearum* RACE 2 BIOVAR 1 ASSOCIATED WITH MOKO DISEASE OF BANANA IN PENINSULAR MALAYSIA



By

DZARIFAH MOHAMED ZULPERI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

January 2015

## COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Doctor of Philosophy

### CHARACTERIZATION OF *Ralstonia solanacearum* RACE 2 BIOVAR 1 ASSOCIATED WITH MOKO DISEASE OF BANANA IN PENINSULAR MALAYSIA

By

#### DZARIFAH MOHAMED ZULPERI

January 2015

#### Chairman: Associate Professor Kamaruzaman Sijam, PhD

#### Faculty: Agriculture

Moko disease caused by Ralstonia solanacearum race 2 biovar 1 (R. solanacearum R2Bv1) is a major disease affecting banana (Musa spp.) production. Although local reports suggested that this disease is widespreading in Malaysia, characterization of R. solanacearum strains associated with Moko disease in this country has not been done. This study was conducted to isolate, identify and characterize R. solanacearum R2Bv1 of Moko-causing strains in Peninsular Malaysia. During March 2011 to June 2012, 170 banana plants associated with Moko disease and adjacent soil samples were collected in 12 different locations of five outbreak states in Peninsular Malaysia comprising Kedah, Selangor, Pahang, Negeri Sembilan and Johor with disease incidence exceeding 80 % in some severely affected plantations. All 197 isolates produced fluidal colonies that were white to pink coloration after incubation at 24 to 48 hours at 29 °C on Kelman's TZC agar medium and were divided into two defined colony type, the B and SFR types. These isolates appeared as Gram-negative rods after Gram-stain, and positive for potassium hydroxide (KOH), Kovacs oxidase, catalase and lipase activity on Tween 80 solution tests. In biovar determination, only 30 isolates displayed characteristics of biovar 1 R. solanacearum, which was negative for utilization of disaccharides and hexose Tobacco hypersensitivity assay revealed all isolates elicited alcohols. hypersensitive response (HR) at 12 h after infiltration, suggesting that they were of race 2. In preliminary pathogenicity study, all 30 isolates were virulence towards three Moko most affected local banana cultivars namely Musa paradisiaca cv. Nipah, Musa paradisiaca cv. Tanduk and Musa acuminata cv. Berangan cultivars with diverse degrees of virulence; highly virulent, moderately virulent and weakly virulent with isolate NS-N1 as the most virulent, while isolates Ked-KN4 and Ked-KN5 were classified as weakly virulent. Musa paradisiaca cv. Nipah (ABB triploid) significantly exhibited the highest degree of severity to R. solanacearum, followed by Musa paradisiaca cv. Tanduk (AAB triploid) and Musa acuminata cv. Berangan (AAA triploid). Moreover, statistical results revealed

there were relationships between geographical origins of isolates and their severity, with the most and the lowest severity was related to isolates from Johor and Negeri Sembilan. This study represents the first evidence on the introduction of R. solanacearum biovar 1 associated with Moko disease of banana in Peninsular Malaysia. Partial 16S rDNA sequence analyses disclosed that all 30 isolates of R. solanacearum biovar 1 were clustered to the published R. solanacearum biovar 1 related to Moko-causing strains from the Philippines (MOD5 and R633) with 91 % Bayesian posterior probability support and completely different from Ralstonia syzygii (R. syzygii, S444E), blood disease bacterium (T520) and the outgroup strain, Xanthomonas spp. (55485). Meanwhile, phylogenetic analyses further demonstrated that all strains were grouped with 100 % posterior probability support to the published R. solanacearum race 2 insertion sequence gene, ISRso19 (AF450275). Phylotypespecific multiplex PCR (Pmx-PCR) showed all strains belonged to phylotype II displaying a 372 bp amplicon. Phylogenetic analyses of endoglucanase (egl) sequences clustered all 30 strains into phylotype II/4, together with the reference sequences strains from Peru (UW129, UW162 and UW163) and Colombia (UW070). Bioinformatics analysis of pooled rep-PCR fingerprinting method defined two major groups; cluster 1 (sub-group A and B) and cluster 2 (sub-group C), with 35 % average similarity coefficient within these two clusters. The subgroups in cluster 1 were represented by strains from Kedah, Selangor, Negeri Sembilan and Johor; while cluster 2 sub-group was represented exclusively by strains of Pahang. This is indeed the first time that genetic diversity of R. solanacearum R2Bv1 has been characterized in this country, where rep-PCR technique clearly distinguished clonal lineages of Moko-causing strains in Peninsular Malaysia. These findings provide constructive documentations on R. solanacearum R2Bv1 in Malaysia, since banana has been identified as the second most important commercial fruit crop with a high economic value in this country.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

#### PENCIRIAN KE ATAS *Ralstonia solanacearum* RAS 2 BIOVAR 1 DIKAITKAN DENGAN PENYAKIT MOKO PISANG DI SEMENANJUNG MALAYSIA

Oleh

#### DZARIFAH MOHAMED ZULPERI

Januari 2015

## Pengerusi: Professor Madya Kamaruzaman Sijam, PhD

#### Fakulti: Pertanian

Penyakit Moko yang disebabkan oleh bakterium Ralstonia solanacearum ras 2 biovar 1 (R. solanacearum R2Bv1) adalah penyakit yang memberi kesan utama ke atas pengeluaran pisang (Musa spp.) di dunia. Walaupun terdapat laporan menyatakan penyakit ini semakin menular di Malaysia, pencirian ke atas strain R. solanacearum yang dikaitkan dengan penyakit Moko di negara ini belum pernah dijalankan. Oleh itu, kajian ini dijalankan untuk memencil, mengenalpasti dan mencirikan strain *R. solanacearum* R2Bv1 daripada penyakit Moko pokok pisang di Semenanjung Malaysia. Pada bulan Mac 2011 hingga Jun 2012, sebanyak 170 pokok pisang dengan simptom penyakit Moko dan sampel tanah berdekatan telah disampel secara rawak di 12 lokasi berbeza di lima negeri di Semenanjung Malavsia yang terdiri dari Kedah, Selangor, Pahang, Negeri Sembilan dan Johor, dengan simptom penyakit melebihi 80 % di ladang-ladang yang terjejas. Keseluruhan 197 isolat menghasilkan koloni berfluidal berwarna putih ke merah jambu selepas inkubasi selama 24 hingga 48 jam pada 29 ° C di atas agar TZC Kelman yang dibahagikan kepada dua jenis koloni iaitu B dan SFR. Kesemua isolat adalah Gram-negatif rod serta positif bagi ujian biokimia berikut; kalium hidroksida (KOH), Kovacs oxidase, catalase dan aktiviti lipase di dalam Tween 80. Dalam penentuan biovar, hanya 30 isolat R. solanacearum memaparkan ciriciri biovar 1, iaitu negatif terhadap penggunaan disakarida dan hexose alkohol. hipersensitiviti tembakau mendedahkan bahawa kesemua strain Uiian menghasilkan tindakbalas hipersensitif (HR) pada 12 jam selepas inokulasi. Penyaringan patogenisiti oleh keseluruhan 30 isolat ke atas tiga jenis kultivar pisang tempatan yang paling terjejas akibat penyakit Moko iaitu Musa paradisiaca cv. Nipah, Musa paradisiaca cv. Tanduk dan Musa acuminata cv. Berangan menghasilkan darjah virulen berbeza iaitu; sangat virulen, sederhana virulen dan kurang virulen dengan isolat NS-N1 sebagai yang paling virulen, manakala isolat Ked-KN4 dan Ked-KN5 dikelaskan sebagai paling kurang Musa paradisiaca cv. Nipah (ABB triploid) secara signifikan virulen. menghasilkan tahap kerentanan tertinggi terhadap penyakit Moko, diikuti oleh Musa paradisiaca cv. Tanduk (AAB triploid) dan Musa acuminata cv. Berangan

(AAA triploid) sebagai yang paling resistan terhadap penyakit Moko. Analisis statistik juga menunjukkan terdapat hubungan antara kedudukan geografi isolat dan tahap virulen, iaitu yang paling virulen adalah strain dari Johor dan paling kurang virulen adalah isolat dari Negeri Sembilan. Kajian ini adalah bukti kemasukan R. solanacearum biovar 1 yang dikaitkan dengan penyakit Moko pisang di Semenanjung Malaysia. Analisis jujukan 16S rDNA separa menunjukkan bahawa kesemua 30 isolat R. solanacearum biovar 1 menyamai strain-strain R. solanacearum biovar 1 rujukan penyakit Moko dari Filipina (MOD5 and R633) dengan 91 % sokongan kebarangkalian posterior Bayesian. Analisis filogenetik membuktikan kesemua strain telah dikelompokkan bersama gen rujukan bagi R. solanacearum ras 2, ISRso19 dengan 100 % sokongan kebarangkalian posterior. Multipleks PCR berfilotip khusus (Pmx-PCR) pula menghasilkan 372 bp amplikon yang menunjukkan kesemua strain adalah dalam kumpulan filotip II. Analisis filogenetik ke atas jujukan-jujukan endoglucanase (eql) membuktikan kesemua 30 strain berada dalam kumpulan filotip II seguevar 4, bersamaan dengan strain rujukan dari Peru (UW129, UW162 and UW163) dan Colombia (UW070). Analisis bioinformatik data PCR-berkelompok menghasilkan dua kumpulan utama; kelompok 1 (sub-kumpulan A dan B) dan kelompok 2 (subkumpulan C), dengan 35 % nilai koefisien. Sub-kumpulan kelompok 1 diwakili strain dari Kedah, Selangor, Negeri Sembilan dan Johor; manakala subkumpulan kelompok 2 diwakili hanya strain dari Pahang. Ini merupakan kali pertama pencirian kepelbagaian genetik itu R. solanacearum R2Bv1 dilaporkan di Semenanjung Malaysia. Oleh kerana pisang telah dikenal pasti sebagai tanaman buah-buahan komersial kedua terpenting di Malaysia, penemuan daripada kajian ini dapat menyediakan dokumentasi konstruktif ke atas R. solanacearum R2Bv1 di negara ini.

### ACKNOWLEDGEMENTS

In the first place I would like to thank both of my parent, and family for their effort and support throughout my education. Their encouragement, love and care were the main source of power that prompted me to accomplish this thesis.

I would like to express sincerest gratitude to my supervisor, Associate Professor Dr. Kamaruzaman Sijam for giving me the opportunity to work on this project. I earnestly appreciate his guidance, advice, understanding and patience throughout my graduate study, particularly in the research work and writing. I also would like to thank my committee members, Associate Professor Dr. Zainal Abidin Mior Ahmad and Associate Professor Dr. Yahya Awang for their valuable advices and suggestions on conducting my research. My utmost appreciation to all officers from the Department of Agriculture of Malaysia for allowing me to sample diseased bananas in plantations and orchards, and for providing assistance during my samplings. I appreciate the help from the staffs of Department of Plant Protection especially Ms. Junaina Jaafar, Mr. Yusoff Mohd Yassin and Mr. Johari Mohd Sarikat. I am also thankful for the significant efforts and contributions of my department colleagues and labmates; Erneeza Mohd Hata, Aisyah Azman, Zafri Wahab, Tavga Sulaiman Rashid, Roslina Sulaiman, Norhayati Madiha, Norhayu Asib and Siti Izera Ismail to this project.

Finally, I would like to thank all my friends for their generous support and help. For the three wonderful persons who have been with me through many difficulties in my life; Zarirah Zulperi, Ina Salwany Md Yasin and Murni Marlina Abd Karim, you are all irreplaceable! I certify that a Thesis Examination Committee has met on 20 January 2015 to conduct the final examination of Dzarifah Mohamed Zulperi on her thesis entitled "Characterization of *Ralstonia solanacearum* Race 2 Biovar 1 Associated with Moko Disease of Banana in Peninsular Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Pertanian Malaysia (P.U. (A) 106) 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Ganesan a/I Vadamalai, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

## Jugah b Kadir, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

# Shuhaimi bin Musta<mark>fa, PhD</mark>

Professor Halal Products Research Institute Universiti Putra Malaysia (Internal Examiner)

## Jeffrey b Jones, PhD

Professor Plant Pathology Department University of Florida (External Examiner)



**ZULKARNAIN ZAINAL, PhD** Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 26 February 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Kamaruzaman Sijam, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Zainal Abidin Mior Ahmad, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

#### Yahya Awang, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

> BUJANG BIN KIM HUAT, PhD Professor and Dean

School of Graduate Studies Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No:	

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of the thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of Chairman of Supervisory Committee:

Signature:

Name of Member of Supervisory Committee:

Signature:

Name of Member of Supervisory Committee:

## TABLE OF CONTENTS

	<b>-</b>	Page
ABSTRAC ABSTRAK		iii
	EDGEMENTS	V
APPROVA		vi
DECLARA		viii
LIST OF FI		xvi
LIST OF T	ABLES	xix
LIST OF A	BRREVIATIONS	xiii
CHAPTER		1
1	INTRODUCTION	1
	1.1 Background of the Study 1.2 Statement of the Problem	1
	1.3 Significance of the Study	2 2
	1.4 Objective of the Study	2
	1.4 Objective of the Study	2
2	LITERATURE REVIEW	5
-	2.1 History and Global Production of Banana	5
	2.2 Banana Cultivation in Malaysia	9
	2.3 Bacterial Diseases of Banana	11
	2.4 Moko Disease	12
	2.4.1 Distribution and Epidemiology of Moko Disease	12
	2.4.2 Symptoms of Moko Disease	14
	2.4.3 Prevalence of Moko Disease	17
	2.4.4 Management and Control of Moko Disease	17
	2.5 Ralstonia solanacearum	17
	2.5.1 History and Biology of Ralstonia solanacearum	17
	2.5.2 Pathogenesis of Ralstonia solanacearum	20
	2.5.3 Virulence Factors of Ralstonia solanacearum	23
	2.6 Diversity of Ralstonia solanacearum Species Complex	
	2.6.1 Races	26
	2.6.2 Biovars	26
	2.6.3 Molecular Characterization and Phylogenetic	26
	Analyses 2.7 Genetic Diversity of <i>Ralstonia solanacearum</i>	26 28
	2.7 Genetic Diversity of <i>Raistonia</i> solariacearum 2.7.1 rep-PCR as a Fingerprinting Method in	20
	Deciphering Genetic Diversity of <i>Ralstonia</i>	
	solanacaerum	28
3	ISOLATION, IDENTIFICATION AND PHENOTYPIC	
	CHARACTERIZATION OF Ralstonia solanacearum	
	ASSOCIATED WITH MOKO DISEASE OF BANANA IN	
	PENINSULAR MALAYSIA	31
	3.1 Introduction	31
	2.2 Matarials and Mathads	20

3.2 Materials and Methods323.2.1 Samples Collection32

	3.2.2 Isolation of Pure Ralstonia solanacearum of Moko D	isease 35
	3.2.3 Biochemical Characterization of Ralstonia solanacea	arum
	3.2.4 Biovar Determination of <i>Ralstonia solanacearum</i> of Disease	35 Moko 36
	3.2.5 Hypersensitivity Assay of <i>Ralstonia solanacearum</i> B of Moko Disease	
	3.2.6 Preliminary Study on Pathogenicity of Ralstonia	
	solanacearum Biovar 1 in Banana Explants	38
	3.2.6.1 Disease Severity Assessment	39
	3.2.6.2 Statistical Analysis	39
	3.3 Results	41
	3.3.1 Field Symptoms of Moko Disease	41
	3.3.2 Isolation of and Morphological Characterization of	
	Ralstonia solanacearum of Moko Disease	42
	3.3.3 Biochemical Characterization of Ralstonia solanace	arum
		45
	3.3.4 Biovar Determination of Ralstonia solanacearum of M	Noko
	Disease	45
	3.3.5 Hypersensitivity Assay of Ralstonia solanacearum	
	Biovar 1 of Moko Disease	48
	3.3.6 Preliminary Study on Pathogenicity of Ralstonia	
	solanacearum Biovar 1 in Banana Explants	49
	3.4 Discussion	53
4	MOLECULAR CHARACTERIZATION AND PHYLOGENY OF	=
	Ralstonia solanacearum BIOVAR 1 STRAINS OF MOKO	
	DISEASE IN PENINSULAR MALAYSIA	61
	4.1 Introduction	61
	4.2 Materials and Methods	62
	4.2.1 Samples Collection	62
	4.2.2 Bacterial DNA Extraction	64
	4.2.3 Qualification and Quantification of DNA	64
	4.2.4 Species-specific PCR Amplification of Ralstonia	-
	solanacearum Biovar 1 Strains	64
	4.2.5 Race-specific PCR Amplification of Ralstonia solana	cearum
	Biovar 1 Strains	65
	4.2.6 Phylotype-specific Multiplex PCR Amplification of Ra	
	solanacearum Race 2 Biovar 1 Strains	65
	4.2.7 Endoglucanase (egl) Gene Amplification of Ralstonia	
	solanacearum Race 2 Biovar 1 Strains	66
	4.2.8 Detection of PCR Products	66
	4.2.9 Purification of the PCR Products	66
	4.2.10 DNA Sequencing and Sequence Alignment	67
	4.2.11 Phylogenetic Analyses	67

	4.3 Results	70
	4.3.1 Species-specific PCR Amplification of <i>Ralstonia</i>	70
	solanacearum Biovar 1 Strains	70
	4.3.2 Race-specific PCR Amplification of <i>Ralstonia</i>	74
	solanacearum Biovar 1 Strains	74
	4.3.3 Phylotype-specific Multiplex PCR Amplification of R	
	solanacearum Race 2 Biovar 1 Strains	, 78
	4.3.4 Endoglucanase (egl) Gene Amplification of Ralston	
	solanacearum Race 2 Biovar 1 Strains	78
	4.4 Discussion	83
5	GENETIC DIVERSITY OF THE MOKO-CAUSING STRAIN,	
	Ralstonia solanacearum RACE 2 BIOVAR 1 IN PENINSUL	.AR
	MALAYSIA AS REVEALED BY rep-PCR FINGERPRINTING	G
	METHOD	91
	5.1 Introduction	91
	5.2 Materials and Methods	92
	5.2.1 Samples Collection	92
	5.2.2 Bacterial DNA Extraction	92
	5.2.3 REP-, ERIC- and BOX-PCR Amplifications	92
	5.2.4 Data Analysis and Dendrogram Construction	94
	5.3 Results	94
	5.3.1 REP PCR Amplification	94
	5.3.2 ERIC PCR Amplification	97
	5.3.4 BOX PCR Amplification	100
	5.3.5 Pooled REP-, ERIC- and BOX PCR Amplifications	103
	5.3.6 Statistical Analysis	105
	5.4 Discussion	106
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS	
·	FOR FUTURE RESEARCH	109
REFEREN	CES	119
APPENDI		136
BIODATA OF STUDENT 139		
LIST OF PUBLICATIONS 140		

## LIST OF FIGURES

Figu	re	Page
	Major banana exports countries from 2006 to 2008 (adopted from , 2009).	11
2.2:	Distribution map of <i>Ralstonia solanacearum</i> race 2 worldwide in 2013.	19
2.3:	Symptoms and sign of Moko disease on banana plantain.	21
2.4:	From inside to outside: Core Genome, Dispensable Genome, Specific Genome at the phylotype level.	30
2.5:	Virulence mechanism of <i>Ralstonia solanacearum</i> -host interaction (adopted from Abramovitch <i>et al.</i> , 2006).	33
2.6:	The <i>Ralstonia</i> solanacearum infectious cycle (adopted from Genin, 2010).	33
2.7:	Diagrammatic representation of the hierarchical classification scheme (adopted from Fegan, 2006).	42
2:8:	Relative capability of DNA-based methods to resolve bacteria at different taxonomic levels (adopted from Louws <i>et al.</i> , 1999).	45
3.1:	Sampling areas in Peninsular Malaysia where the samples were collected.	49
3.2:	Pictures postulated the possible outcome of hypersensitivity assay of <i>Ralstonia solanacearum</i> isolates according to their races.	56
3.3:	Symptoms of Moko disease observed on banana plantains in Keratong, Pahang of Peninsular Malaysia.	60
3.4:	Symptoms of Moko disease and sign of the bacterial existence observed on various banana plantains in Peninsular Malaysia.	61
3.5:	Colony morphology of <i>Ralstonia solanacearum</i> isolated from infected sampling areas across Peninsular Malaysia.	64
3.6:	Tobacco infiltration assay of <i>Ralstonia solanacearum</i> isolates.	69
3.7:	Wilt symptoms induced by a <i>Ralstonia solanacearum</i> biovar 1 isolate, NS-N1 at four weeks after infiltration.	74

3.8: Gene-for-gene interaction specifies plant disease resistance.	83
4.1: PCR amplification of total genomic DNA from 30 Malaysian Ralstonia solanacearum biovar 1 strains based on 16S rDNA region with primers L 10 and R 1541, each producing a ~1400 bp amplicon.	100
4.2: Phylogenetic tree construsted from a Bayesian analysis of the partial 16S rDNA gene sequences using MrBayes version 3.2.0.	101
4.3: PCR amplification of total genomic DNA from 30 Malaysian Ralstonia solanacearum biovar 1 strains using race-specific primers.	104
4.4: Phylogenetic tree construsted from a Bayesian analysis of the IS <i>Rso19</i> gene sequences using MrBayes version 3.2.0.	105
4.5: Phyloptype-specific multiplex PCR amplification of total genomic DNA from 30 Malaysian <i>Ralstonia solanacearum</i> race 2 biovar 1 strains used in this study.	109
4.6: PCR amplification of total genomic DNA from 30 Malaysian Ralstonia solanacearum race 2 biovar 1 strains using Endo- F/Endo-R primers, each raising a ~750 bp amplicon.	110
4.7: Phylogenetic tree construsted from a Bayesian analysis of the partial <i>egl</i> sequences using MrBayes version 3.2.0.	111
5.1: REP-PCR amplification of 30 genomic DNA from <i>Ralstonia solanacearum</i> R2Bv1 strains isolated from various states in Malaysia.	132
5.2: Dendrogram on diversity of 30 strains of <i>Ralstonia</i> solanacearum R2Bv1 based on REP-PCR amplification.	133
5.3: ERIC-PCR amplification of 30 genomic DNA from Ralstonia solanacearum R2Bv1 strains isolated from various states in Malaysia.	135
5.4: Dendrogram on diversity of 30 strains of <i>Ralstonia solanacearum</i> R2Bv1 based on ERIC-PCR amplification.	136
5.5: BOX-PCR amplification of 30 genomic DNA from <i>Ralstonia solanacearum</i> R2Bv1 strains isolated from various states in Malaysia.	138

5.6:	Dendrogram on diversity of 30 strains of Ralstonia solanacearum	139
	R2Bv1 based on BOX-PCR amplification.	

5.7: Dendrogram on diversity of 30 strains of *Ralstonia solanacearum* 141 R2Bv1 based on pooled rep-PCR amplification data.



## LIST OF TABLES

Table Page		
2.1: Global production statistic of banana and plantain in 2003 (adopted from FAO, 2003).	10	
2.2: Number of cultivars under the different species and types of edible bananas (adopted from Valmayor, 2000).	10	
2.3: The hectarage, production and value of production of major fruit crops in Malaysia during 2011 (adopted from DOA Malaysia, 2012).	13	
2.4: The hectarage of cultivated banana areas in various states in Malaysia within 2007 to 2011 (adopted from DOA Malaysia, 2012).	13	
2.5: Major banana diseases and their causal pathogens (based from Jones, 2000).	16	
2.6: Characteristics of <i>Ralstonia solanacearum</i> strains associated with Moko disease (adopted from Jones, 2000).	38	
2.7: Tests for biovar determination in <i>Ralstonia solanacearum</i> (adopted from EPPO/CABI, 2004).	39	
3.1: Total banana plants and soil samples collected from different locations area in Peninsular Malaysia from March 2011 to June 2012.	50	
3.2: Disease incidence of Moko disease in infected sampling areas across Peninsular Malaysia and distribution of colony types of suspected <i>Ralstonia solanacearum</i> isolates obtained.	63	
3.3: Morphological, biochemical and biovar characterization on 30 isolates of <i>Ralstonia solanacearum</i> from Peninsular Malaysia.	68	
3.4: Susceptibility of <i>Musa paradisiaca</i> cv. Nipah, <i>Musa paradisiaca</i> cv. Tanduk and <i>Musa acuminata</i> cv. Berangan to 30 <i>Ralstonia solanacearum</i> isolates based on disease severity indexes at four weeks after infiltration and the symptoms produced.	73	
4.1: Sources of isolation, sampling areas and banana varieties of 30 isolates of <i>Ralstonia solanacearum</i> biovar 1 used in this study.	89	

4.2: List of primers used in this study.

4.3: Origin and characterization of <i>Ralstonia</i> solanacearum biovar strains from Malaysia based on partial 16S rDNA gene sequenci and reference strains used in this study.	
4.4: Origin and characterization of <i>Ralstonia solanacearum</i> race biovar 1 strains from Malaysia based on IS <i>Rso19</i> ge sequencing and reference strains used in this study.	
4.5: Origin and characterization of <i>Ralstonia solanacearum</i> race biovar 1 strains from Malaysia and reference strains used in the study based on phylotype classification and sequevar group.	
5.1: List of rep-PCR primers used in this study.	142
5.2: ANOVA results on total number of observed bands among thr fingerprinting (REP-, ERIC- and BOX-PCR) methods used in t study.	
5.3: ANOVA results among locations of origin for the total number observed bands with REP-, ERIC-, BOX- and pooled rep-PC data.	

 $\mathcal{O}$ 

## LIST OF ABBREVIATIONS

% °C bp CABI DNA DOA EDTA FAO g h kb L M Mb min ml Mb min ml mm mM ng nm OD PCR rpm sec TAE Taq T <sub>M</sub> U UV VV VV VV VV VV VV	percent degree celcius base pair Commonwealth Agricultural Bureaux International deoxyribonucleic acid Department of Agriculture ethylene-diamine-tetraacetic acid Food and Agriculture Organization gram hour kilobase pair liter molar megabase pair milliter
V	voltan/volt
hđ	microgram
μg/ml	microgram per mililiter
μΪ	microliter
μM	micromolar
μm	micronmeter





## **CHAPTER 1**

## INTRODUCTION

## 1.1 Background of the Study

Ralstonia solanacearum, the causal agent of bacterial wilt, is a soil-borne plant pathogen with a worldwide distribution that afflicts economically important crops and ornamentals (Lin et al., 2014; Denny, 2006; Agrios, 2005). This aerobic, Gram-negative organism, formerly known as Pseudomonas solanacearum is commonly encountered in tropical and subtropical areas. Its immense phenotypic and genotypic diversity contributes to its status as a major plant pathogen (Fegan and Prior, 2006; Agrios, 2005). R. solanacearum infects more than 200 plant species in 50 botanical families; among its hosts are tobacco (Nicotiana tabacum), tomato (Lycopersicon esculentum), potato (Solanum tuberosum) and banana (Musa spp.) (Alvarez et al., 2010; Hayward, 1994). Bacterial wilts of banana, known commonly as Moko, Bugtok and blood disease, are incited by distinct subgroups of the R. solanacearum species complex (RSSC) and pose a major threat to dessert and cooking banana production (Fegan and Prior, 2006).

Moko disease has been acknowledged as one of the remarkable major diseases threatening banana cultivation worldwide. After its first outbreak in Trinidad in the late 1890s, this disease caused by *Ralstonia solanacearum* (*R. solanacearum*) race 2 became endemic in several regions of Central and South America (Jones, 2002). In Jamaica as an example, Moko disease has become a devastating disease attacking banana plantains with estimated annual loss of about USD 5.8 million. This disease has been a major concern for banana growers in the Amazon region of Brazil where it has been the major production constraint for banana yields (Netto and Nutter, 2005).

The presence of Moko disease pathogen in Asia was first detected in the Philippines of Mindanao region (Eyres and Hammond, 2001). To date, the emergence and widespread of Moko disease has been identified in several countries in Asia, Africa, North America, Central America, South America, parts of the Caribbean Islands and Australia continents (EPPO, 2013).

## **1.2 Statement of the Problem**

In Malaysia, the suspected outbreak of Moko disease was primary recognized in Muar, Johor in 2007 (Mokhtarud-din and William, 2011). Earlier findings revealed that tropical condition with a temperate climate like Malaysia was even more conducive for the growth of R. solanacearum and development of this disease in the infected region (Denny, 2006; Hayward, 1991). This vital situation on the epidemic of Moko disease has further diminished little enthusiasm of farmers on banana industry since the disease is amongst the most serious fruit diseases in the country where it widespread rapidly, retards banana plant growth, causes critical yield losses and can rigorously impact the banana growth sector. As banana has been recognized as one of the fruit types for special attention under the implemented Economic Transfer Programme (ETP) by Malaysian government, constant occurrences of this disease have been the most important and major constraint to the production of bananas, resulting to loss of yield and areas that are gradually becoming unsuitable for the production of the crops (Mokhtarud-din and William, 2011; Tengku Abdul Malik et al., 2011; Nik Hassan, 2003).

## 1.3 Significance of the Study

As banana (*Musa* spp.) remains the second most important economic-driven fruit crops in Malaysia for both local and export markets, scrutinizing records on the current status of Moko disease is of significant importance. Up to this point, none of the disease occurrences have been well documented in Malaysia since the first suspected outbreak in 2007. The results of our study will be an important pioneer documentation of Moko disease of banana in Malaysia. Taking this matter into serious account, our study would be a major platform on generating details documentation of Moko disease and its causal pathogen *R. solanacearum* race 2 biovar 1 in banana fruit crops in Malaysia by using combination of phenotypic characterization and molecular phylogenomics approaches.

### 1.4 Objective of the Study

Our study was carried out with the following objectives:

- 1. To isolate, identify and characterize *R. solanacearum* of Mokocausing strains in Peninsular Malaysia by using phenotypic characteristics.
- 2. To investigate genetic relationships and diversity of *R. solanacearum* race 2 biovar 1 strains of Moko disease via molecular characterization and phylogenetic analyses.

The output from this research perhaps may improve and increase efficiency in the development of accurate molecular diagnostic tests for detection and identification of *R. solanacearum* race 2 biovar 1. Indeed, the data obtained will be useful for quarantine purposes and suppression of Moko disease spread, thus bettering the banana industry in Malaysia.



#### REFERENCES

- Abramovitch, R.B., Anderson, J.C. and Martin, G.B. (2006). Bacterial elicitation and evasion of plant innate immunity. *Nature Reviews Molecular Cell Biology*, 7: 601-611.
- Adam, G. (2006). Significant Ways to Spread Plant Virus Diseases in Agricultural Ecosystems: Is Agroterrorism Possible? *Virus Diseases and Crop Biosecurity*, 9: 45-54.
- Agrios, G.N. (2005). Plant Pathology. 5th eds, Academic Press: New York, USA.
- Ahmed, N.N., Islam, M., Hossain, M.A., Meah, M.B. and Hossain, M.M. (2013). Determination of Races and Biovars of *Ralstonia solanacearum* Causing Bacterial Wilt Disease of Potato. *Journal of Agricultural Science*, 5: 86-93.
- Aldag, R. (1987). Simple and Diversified Crop Rotations-Approach and Insight into Agroecosystems. *Potentials and Limitations of Ecosystem Analysis Ecological Studies*, 61: 100-114.
- Albuquerque, G.M., Santos, L.A., Félix, K.C., Rollemberg, C.L., Silva, A.M., Souza, E.B., Cellier, G., Prior, P. and Mariano, R.R. (2014). Moko disease-causing strains of *Ralstonia solanacearum* from Brazil extend known diversity in paraphyletic phylotype II. *Phytopathology*, (http://dx.doi.org/10.1094/PHYTO-12-13-0334-R).
- Aley, E.F.L.G.P. and Elphinstone, J. (1995). Culture media for *Ralstonia solanacearum* isolation, identification and maintenance. *Fitopatologia*, 30: 126-130.
- Altschul, S.F., Gish, W., Miller, W., Myers, E.W. and Lipman, D.J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, 215: 403-410.
- Álvarez, B., Biosca, E.G. and López, M.M. (2010). On the life of *Ralstonia* solanacearum, a destructive bacterial plant pathogen. *Current Research, Technology and Education Topics in Applied Microbiology* and *Microbial Biotechnology*, 1: 267-279.
- Anderson, P.K., Cunningham, A.A., Patel, N.G., Morales, F.J., Epstein, P.R. and Daszak, P. (2004). Emerging infectious diseases of plants: pathogen pollution, climate change and agrotechnology drivers. *Trends in Ecology and Evolution*, 19: 535-544.
- Arshiya, M., Suryawanshi, A., More, D. and Baig, M.M.V. (2014). Repetitive PCR based detection of Genetic Diversity in *Xanthomonas axonopodis*

pv citri Strains. Journal of Applied Biology and Biotechnology, 2: 017-022.

- Azhar, M. and Heslop-Harrison, J.S. (2008). Genomes, diversity and resistance gene analogues in *Musa* species. *Cytogenetic and Genome Research*, 121: 59-66.
- Bonas, U. and Van den Ackerveken, G. (1999). Gene-for-gene interactions: bacterial avirulence proteins specify plant disease resistance. *Current Opinion in Microbiology*, 2: 94-98.
- Buddenhagen, I. and Kelman, A. (1964). Biological and physiological aspects of bacterial wilt caused by *Pseudomonas solanacearum*. *Annual Review of Phytopathology*, 2: 203-230.
- Buddenhagen, I.W. (1986). Bacterial wilt revisited. In Bacterial Wilt Disease in Asia and the South Pacific, ed. G. J. Persley, pp. 126-143.
- Buddenhagen, I.W. (1986). Disease susceptibility and genetics in relation to breeding of bananas and plantains. *Banana and Plantain Breeding Strategies*, 21: 95-109.
- Buddenhagen, I.W. (1994). Banana diseases caused by bacteria. In Compendium of Tropical Fruit Diseases (RC Ploetz, GA Zentmyer, WT Nishijima, KG Rohrbach, & HD Ohr, eds.). APS Press, St Paul, Minnesota, 15-17.
- Buddenhagen, I.W., Sequeira, L. and Kelman, A. (1962). Designation of Races in *Pseudomonas solanacearum*. *Phytopathology*, 52: 726.
- Castillo, J.A. and Greenberg, J.T. (2007). Evolutionary dynamics of *Ralstonia solanacearum. Applied Environmental Microbiology*, 73: 1225-1238.
- Cellier, G. and Prior, P. (2010). Deciphering phenotypic diversity of *Ralstonia solanacearum* strains pathogenic to potato. *Phytopathology*, 100: 1250-1261.
- Cellier, G., Remenant, B., Chiroleu, F., Lefeuvre, P. and Prior, P. (2012). Phylogeny and population structure of brown rot-and Moko diseasecausing strains of *Ralstonia solanacearum* phylotype II. *Applied and Environmental Microbiology*, 78: 2367-2375.
- Chandrashekara, K.N., Kumar, M.K. and Saroja, S. (2012). Aggressiveness of *Ralstonia solanacearum* isolates on Tomato. *Journal of Experimental Sciences*, 3: 5-9.
- Chaudhry, Z. and Rashid, H. (2011). Isolation and Characterization of *Ralstonia solanacearum* from Infected Tomato Plants of Soan Skesar Valley of Punjab. *Pakistan Journal of Botany*, 43: 2979-2985.

- Claverie, J.M. and Notredame, C. (2003). Bioinformatics for dummies. JW Wiley Inc., New York, NY.
- Cook, R.J. and Weller, D.M. (2004). *In* Defense of Crop Monoculture. *In* Proceedings of the 4th International Crop Science Congress, 26<sup>th</sup> Sep– 1<sup>st</sup> Oct 20, Brisbane, Australia.
- Cook, D., Barlow, E. and Sequeira, L. (1989). Genetic diversity of *Pseudomonas solanacearum*: detection of restriction fragment length polymorphisms with DNA probes that specify virulence and the hypersensitive response. *Molecular Plant-Microbe Interactions*, 1: 113-121.
- Cooke, B.M. (2006). Disease assessment and yield loss. In The Epidemiology of Plant Diseases (pp. 43-80). Springer Netherlands.
- Cruz, L., Eloy, M., Quirino, F., Oliveira, H. and Tenreiro, R. (2012). Molecular epidemiology of *Ralstonia solanacearum* strains from plants and environmental sources in Portugal. *European Journal of Plant Pathology*, 133: 687-706.
- Davey, M.W., Van den Bergh, I., Markham, R., Swennen, R. and Keulemans, J. (2009). Genetic variability in *Musa* fruit provitamin A carotenoids, lutein and mineral micronutrient contents. *Food Chemistry*, 115: 806-813.
- de Bruijn, F.J. (1992). Use of repetitive (repetitive extragenic palindromic and enterobacterial repetitive intergeneric consensus) sequences and the polymerase chain reaction to fingerprint the genomes of *Rhizobium meliloti* isolates and other soil bacteria. *Applied and Environmental Microbiology*, 58: 2180-2187.
- Deberdt, P., Guyot, J., Coranson-Beaudu, R., Launay, J., Noreskal, M., Rivière, P., Vigne, F., Laplace, D., Lebreton, L. and Wicker, E. (2014). Diversity of *Ralstonia solanacearum* in French Guiana Expands Knowledge of the "Emerging Ecotype". *Phytopathology*, 104: 586-596.
- Denny, T. (2006). Plant pathogenic *Ralstonia* species. In *Plant-associated bacteria* (pp. 573-644). Springer Netherlands.
- Dhanalakshmi, S. and Stephan, R. (2014). Low Cost Media Options for the Production of Banana (*Musa paradisiaca* L.) through Plant Tissue Culture. *Journal of Academia and Industrial Research (JAIR)*, 2: 509.

DOA Malaysia. (2012). Fruit crops statistic of Malaysia.

Drummond, A.J. and Rambaut, A. (2007). BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology*, 7: 214.

- Eden-Green, S.J. (1994). Diversity of *Pseudomonas solanacearum* and related bacteria in South East Asia: New directions for Moko disease. In Bacterial wilt: The disease and its causative agent, Pseudomonas solanacearum, A.C Hayward and G.L Hartman eds., CAB International: UK. pp. 25-34.
- Elphinstone, J.G. (2005). The current bacterial wilt situation: a global overview. In *Bacterial Wilt Disease and the Ralstonia solanacearum Species Complex* (Allen, C., Prior, P. and Hayward, A.C., eds), pp. 9– 28. St Paul, MN: APS Press.
- Engering, A., Hogerwerf, L. and Slingenbergh, J. (2013). Pathogen-hostenvironment interplay and disease emergence. *Emerging Microbes and Infections*, 2: e5.
- Englberger, L., Schierle, J., Marks, G.C. and Fitzgerald, M.H. (2003). Micronesian banana, taro, and other foods: newly recognized sources of provitamin A and other carotenoids. *Journal of Food Composition and Analysis*, 16: 3-19.
- Englberger, L., Wills, R.B., Blades, B., Dufficy, L., Daniells, J.W. and Coyne, T. (2006). Carotenoid content and flesh color of selected banana cultivars growing in Australia. *Food and Nutrition Bulletin*, 27: 281-291.
- EPPO. (2013). Distribution map of *Ralstonia solanacearum* race 2 worldwide. PQR database. Paris, France: European and Mediterranean Plant Protection Organization. <u>http://www.eppo.org</u> (Date of accessed: February 23<sup>rd</sup> 2014).
- EPPO/CABI. (2004). Data Sheets on Quarantine Pests Ralstonia solanacearum. Prepared by CABI and EPPO for the EU under Contract 90/399003.
- Eyres, N. and Hammond, N. (2001). Moko disease *Ralstonia solanacearum* (Race 2, Biovar 1). Department of Agriculture and Food Factsheet, Government of Western Australia.
- Fahy, P.C. and Hayward, A.C. (1983). Media and methods for isolation and diagnostic tests.
- FAO. (2003). Food and Agricultural Organization. Rome, Italy (<u>http://www.fao.org</u>).
- FAO. (2009). Food and Agricultural Organization. Rome, Italy (<u>http://www.fao.org/es/ESC/en</u>).
- Fegan, M. (2006). Bacterial wilt of banana–diagnostics manual draft copy. In Plant Health Australia and the CRC for tropical plant protection, St Lucia, Australia.

- Fegan, M. and Prior, P. (2006). Diverse members of the *Ralstonia* solanacearum species complex cause bacterial wilts of banana. *Australasian Plant Pathology*, 35: 93-101.
- Fegan, M., Allen, C., Prior, P. and Hayward, A.C. (2005). Bacterial wilt diseases of banana: evolution and ecology. In *Bacterial wilt disease and the Ralstonia solanacearum species complex*, 379-386.
- Fegan, M., Prior, P., Allen, C. and Hayward, A.C. (2005). How complex is the "*Ralstonia solanacearum* species complex"?. *Bacterial wilt disease and the Ralstonia solanacearum species complex*, 449-461.
- Fonseca, N.R., Guimarães, L.M.S., Hermenegildo, P.S., Teixeira, R.U., Lopes, C.A. and Alfenas, A.C. (2013). Molecular characterization of *Ralstonia solanacearum* infecting *Eucalyptus* spp. in Brazil. *Forest Pathology*, 44: 107-116
- French, E. and Sequeira, L. (1970). Strains of *Pseudomonas solanacearum* from Central and South America: a comparative study. *Phytopathology*, 60: 506-12.
- French, E.R. (1985). Interaction between strains of *Ralstonia* solanacearum, its hosts and the environment. In *Bacterial wilt disease* en Asia and the South Pacific. ACIAR Proceedings, 13: 99-104.
- Genin, S. (2010). Molecular traits controlling host range and adaptation to plants in *Ralstonia solanacearum*. *New Phytologist*, 187: 920-928.
- Genin, S. and Boucher, C. (2002). *Ralstonia solanacearum*: secrets of a major pathogen unveiled by analysis of its genome. *Molecular Plant Pathology*, 3: 111-118.
- Genin, S. and Boucher, C. (2004). Lessons Learned from the Genome Analysis of *Ralstonia solanacearum*. *Annual Review of Phytopathology*, 42: 107–34.
- Genin, S. and Denny, T.P. (2012). Pathogenomics of the *Ralstonia solanacearum* species complex. *Annual Review of Phytopathology*, 50: 67-89.
- Ghini, R., Bettiol, W. and Hamada, E. (2011). Diseases in tropical and plantation crops as affected by climate changes: current knowledge and perspectives. *Plant Pathology*, 60: 122-132.
- Gouy, M., Guindon, S. and Gascuel, O. (2010). SeaView version 4: a multiplatform graphical user interface for sequence alignment and phylogenetic tree building. *Molecular Biology and Evolution*, 27: 221-224.

- Gram, C. (1884). The differential staining of *Schizomycetes* in tissue sections and in dried preparations. *Fortschritte der Medizin*, 2: 185-9.
- Gund, S.V. (2010). Development of a Scar Marker for the Detection of *Ralstonia solanacearum* (Smith) Yabuuchi (Doctoral dissertation, University of Agricultural Sciences, Dharwad, India).
- Hall, B.G. (2008). *Phylogenetic trees made easy: a how-to manual* (Vol. 547). Sunderland: Sinauer Associates.
- Hallam, D. (1995). The world banana economy. In *Bananas and Plantains* (pp. 509-533). Springer: Netherlands.
- Hayward, A.C. (1964). Characteristics of *Pseudomonas solanacearum*. *Journal of Applied Bacteriology*, 27: 265-277.
- Hayward, A.C. (1985). Bacterial wilt caused by *Pseudomonas* solanacearum in Asia and Australia: an overview. In Bacterial wilt disease in Asia and the South Pacific: proceedings of an international workshop, ed. G.J. Persley, PCARRD, pp. 15-24.
- Hayward, A.C. (1991). Biology and epidemiology of bacterial wilt caused by *Pseudomonas solanacearum. Annual Review of Phytopathology*, 29: 65-87.
- Hayward A.C. (1994). The hosts of Pseudomonas solanacearum. In Bacterial wilt: the disease and its causative agent, *Pseudomonas solanacearum*, eds. A.C. Hayward and G.L. Hartman, Wallingford: CAB International pp. 9.
- Hayward, A.C. (2000). *Ralstonia solanacearum*. In Encyclopedia of Microbiology, ed. J. Lederberg, San Diego, CA: Academic Press, pp. 32-42.
- Hayward, A.C. (2006). Fruit rots of banana caused by *Ralstonia solanacearum* race 2: questions of nomenclature, transmission and control. *InfoMusa*, 15: 7-10.
- Hayward, A.C. and Hartman, G.L. (1994). Bacterial wilt: the disease and its causative agent, *Pseudomonas solanacearum*. CAB International: UK.
- He, L.Y., Sequeira, L. and Kelman, A. (1983). Characteristics of strains of *Pseudomonas solanacearum* from China. *Plant Disease*, 67: 1357-1361.
- Hong, J.C., Norman, D.J., Reed, D.L., Momol, M.T. and Jones, J.B. (2012). Diversity among *Ralstonia solanacearum* strains isolated from the southeastern United States. *Phytopathology*, 102: 924-936.

- Horita, M. and Tsuchiya, K. (2001). Genetic diversity of Japanese strains of *Ralstonia solanacearum. Phytopathology*, 91: 399-407.
- Horita, M., Tsuchiya, K. and Ooshiro, A. (2005). Characteristics of *Ralstonia solanacearum* biovar N2 strains in Asia. *Journal of Phytopathology*, 153: 209-213.
- Horita, M., Suga, Y., Ooshiro, A. and Tsuchiya, K. (2010). Analysis of genetic and biological characters of Japanese potato strains of *Ralstonia solanacearum. Journal of General Plant Pathology*, 76: 196-207.
- Horita, M., Tsuchiya, K., Suga, Y., Yano, K., Waki, T., Kurose, D. and Furuya, N. (2014). Current classification of *Ralstonia solanacearum* and genetic diversity of the strains in Japan. *Journal of General Plant Pathology*, 1-11.
- http://reg.upm.edu.my/spk\_upm/OPR-TPU-web/BUKU PANDUAN (pdf)/OPR-TPU-BP-STA-11, Panduan Tanaman Pisang.pdf (Date of accessed: February 20<sup>th</sup> 2014).
- http://www.cabi.org/isc/?compid=5&dsid=44999&loadmodule=datasheet&p age=481&site=144 (Date of accessed: February 23<sup>rd</sup> 2014).
- Isenberg, H.D. (1992). Clinical microbiology procedures handbook.
- Ivey, M.L.L., Gardener, B.B.M., Opina, N. and Miller, S.A. (2007). Diversity of *Ralstonia solanacearum* infecting eggplant in the Philippines. *Phytopathology*, 97: 1467-1475.
- Izadiyan, M. and Taghavi, S.M. (2011). Genotypic Diversity of Iranian strains of *Ralstonia solanacearum*. *Phytopathologia Mediterranea*, 50: 236-244.
- Jeong, E.L. and Timmis, J.N. (2000). Novel insertion sequence elements associated with genetic heterogeneity and phenotype conversion in *Ralstonia solanacearum. Journal of Bacteriology*, 182: 4673-4676.
- Ji, P, Allen, C., Sanchez-Perez, A., Yao, J., Elphinstone, J.G., Jones, J.B. and Momol, M.T. (2007). New diversity of *Ralstonia solanacearum* strains associated with vegetable and ornamental crops in Florida. *Plant Disease*, 91: 195-203.
- Jones, D.R. (2000). Introduction to banana, abaca and enset. In Diseases of banana, abaca and enset, ed. Jones, D.R, pp. 1-31. CABI Publishing: London, UK.
- Jones, D.R. (2002). Risk of spread of banana diseases in international trade and germplasm exchange. In Conferencia Magistral, 105-113.

- Jyothi, H.K. and Santhosha, H.M. (2012). Recent advances in breeding for bacterial wilt (*Ralstonia solanacearum*) resistance in tomato-review. *Current Biotica*, 6: 370-398.
- Katoh, K., Misawa, K., Kuma, K.I. and Miyata, T. (2002). MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research*, 30: 3059-3066.
- Kawasaki, B.T., Hurt, E.M., Mistree, T. and Farrar, W.L. (2008). Targeting cancer stem cells with phytochemicals. *Molecular* Intervention 8: 174-184.
- Keen, N.T. (1990). Gene-for-gene complementarity in plant-pathogen interactions. *Annual Review of Genetics*, 24: 447-463.
- Kelman, A. (1954). The relationship of pathogenicity of *Pseudomonas* solanacearum to colony appearance in a tetrazolium medium. *Phytopathology*, 44:693–695.
- Khakvar, R. (2009). Diversity and characterization of *Ralstonia solanacearum* strains in Peninsular Malaysia. *PhD thesis*. Faculty of Agriculture, Universiti Putra Malaysia, Serdang.
- Khakvar, R., Kamaruzaman, S., Wong, M.Y., Radu, S., Jones, J. and Thong, K.L. (2008). Genomic diversity of *Ralstonia solanacearum* strains isolated from banana farms in West Malaysia. *Plant Pathology Journal*, 7: 162-167.
- Khakvar, R., Sijam, K., Jones, J. and Thong, K.L. (2011). Comparative diversity analysis of *Ralstonia solanacearum* strains in solanaceae farms. In: *Proceedings of International Conference on Life Science and Technology*, 3: 38-42.
- Khakvar, R., Sijam, K., Yun, W.M., Radu, S. and Lin, T.K. (2008). Improving a PCR-Based Method for Identification of *Ralstonia solanacearum* in Natural Sources of West Malaysia. *American Journal of Agricultural & Biological Science*, 3: 490-493.
- Kovacs, N. (1956). Identification of *Pseudomonas pyocyanea* by the oxidase reaction. *Nature*, 178: 703.
- Kranz, J. (1988). Measuring plant disease. In Experimental Techniques in Plant Disease Epidemiology (pp. 35-50). Springer Berlin Heidelberg.
- Kumar, A., Sarma, Y.R. and Anandaraj, M. (2004). Evaluation of genetic diversity of *Ralstonia solanacearum* causing bacterial wilt of ginger using REP-PCR and PCR-RFLP. *Current Science*, 87: 1555-1561.
- Larkin M.A., Blackshields G., Brown N.P., Chenna R., McGettigan P.A., McWilliam H., Valentin F., Wallace I.M., Wilm A., Lopez R., Thompson

J.D., Gibson T.J. and Higgins D.G. (2007). ClustalW and ClustalX version 2. *Bioinformatics*, 23: 2947-2948.

- Lee, Y. and Khor, C.N. (2003). A Novel Insertion Sequence, IS*Rso19*, Isolated from *Ralstonia solanacearum* and Its Application to Race Differentiation. *Plant Pathology Bulletin*, 12: 57-64.
- Lee, Y. and Khor, C.N. (2003). A Novel Insertion Sequence, ISR*so19*, Isolated from *Ralstonia solanacearum* and Its Application to Race Differentiation. *Plant Pathology Bulletin*, 12: 57-64.
- Lee, Y.A., Fan, S.C., Chiu, L.Y. and Hsia, K.C. (2001). Isolation of an Insertion Sequence from *Ralstonia solanacearum* Race 1 and Its Potential Use for Strain Characterization and Detection. *Applied and Environmental Microbiology*, 67: 3943-3950.
- Lelliott, R.A. and Stead, D.E. (1987). *Methods for the diagnosis of bacterial diseases of plants*. Blackwell Scientific Publications.
- Lemessa, F. and Zeller, W. (2007). Isolation and characterisation of *Ralstonia solanacearum* strains from *Solanaceae* crops in Ethiopia. *Journal of Basic Microbiology*, 47: 40-49.
- Lemessa, F. and Zeller, W. (2007). Pathogenic characterisation of strains of *Ralstonia solanacearum* from Ethiopia and influence of plant age on susceptibility of hosts against *R. solanacearum*. *Journal of Plant Diseases and Protection*, 114: 241-249.
- Lemessa, F., Zeller, W. and Negeri, D. (2010). Genetic diversity among strains of *Ralstonia solanacearum* from Ethiopia assessed by repetitive sequence-based polymerase chain reaction (rep-PCR). *EJAST*, 1: 17-26.
- Li, J.G., Liu, H.X., Cao, J., Chen, L.F., Gu, C., Allen, C. and Guo, J.H. (2010). PopW of *Ralstonia solanacearum*, a new two-domain harpin targeting the plant cell wall. *Molecular Plant Pathology*, 11: 371-381.
- Lin, C.H., Tsai, K.C., Prior, P. and Wang, J.F. (2014). Phylogenetic relationships and population structure of *Ralstonia solanacearum* isolated from diverse origins in Taiwan. *Plant Pathology*, 63: 1395-1403.
- Louws, F.J., Fulbright, D.W., Stephens, C.T. and De Bruijn, F.J. (1994). Specific genomic fingerprints of phytopathogenic *Xanthomonas* and *Pseudomonas* pathovars and strains generated with repetitive sequences and PCR. *Applied and Environmental Microbiology*, 60: 2286-2295.
- Louws, F.J., Rademaker, J.L.W. and De Bruijn, F.J. (1999). The three Ds of PCR-based genomic analysis of phytobacteria: diversity, detection, and disease diagnosis. *Annual Review of Phytopathology*, 37: 81-125.

- Lozano, J.C. and Sequeira, L. (1970). Differentiation of races of *Pseudomonas solanacearum* by a leaf infiltration technique. *Phytopathology*, 60: 833-838.
- Mansfield, J., Genin, S., Magori, S., Citovsky, V., Sriariyanum, M., Ronald, P., ... and Foster, G.D. (2012). Top 10 plant pathogenic bacteria in molecular plant pathology. *Molecular Plant Pathology*, 13(6): 614-629.
- Marques, E., Uesugi, C.H., Ferreira, M.A. and Rezende, D.V.D. (2012). Characterization of isolates of *Ralstonia solanacearum* biovar 2, pathogenic to *Eucalyptus* "urograndis" hybrids. *Tropical Plant Pathology*, 37: 399-408.
- Meng, F. (2013). *Ralstonia Solanacearum* Species Complex and Bacterial Wilt Disease. *Journal of Bacteriology and Parasitology*, 4: 1-9.
- Meng, F. (2013). The Virulence Factors of the Bacterial Wilt Pathogen *Ralstonia solanacearum. Journal of Plant Pathology and Microbiology*, 4: 2.
- Mengiste, T. (2012). Plant Immunity to Necrotrophs. Annual Review of *Phytopathology*, 50: 267-294.
- Milgroom, M.G. and Peever, T.L. (2003). Population biology of plant pathogens: the synthesis of plant disease epidemiology and population genetics. *Plant Disease*, 87: 608-617.
- Miller, R.N., Bertioli, D.J., Baurens, F.C., Santos, C.M., Alves, P.C., Martins, N.F., Togawa, R.C., Souza, M.T. and Pappas, G.J. (2008). Analysis of non-TIR NBS-LRR resistance gene analogs in *Musa acuminata* Colla: Isolation, RFLP marker development, and physical mapping. *BMC Plant Biology*, 8: 15.
- Minitab 16 Statistical Software. (2010). (Computer software). State College, PA: Minitab, Inc. (<u>www.minitab.com</u>).
- Mohapatra, D., Mishra, S. and Sutar, N. (2010). Banana and its by-product utilization: an overview. *Journal of Scientific and Industrial Research*, 69: 323-329.
- Mohapatra, B.R., Broersma, K. and Mazumder, A. (2007). Comparison of five rep- PCR genomic fingerprinting methods for differentiation of fecal *Escherichia coli* from humans, poultry and wild birds. *FEMS Microbiology Letters*, 277: 98-106.
- Mokhtarud-din, H. and William, R. (2011). Status of banana cultivation and disease incidences in Malaysia. In: Abstract of the Workshop on Integrated Approaches in Banana Disease Management, 22<sup>th</sup> March 2011, Serdang, Malaysia, pp. 5.

- Molina, A. (2006). Managing bacterial wilt/fruit rot disease of banana in Southeast Asia. In Developing a regional strategy to address the outbreak of banana *Xanthomonas* wilt in East and Central Africa. INIBAP, Montpellier, France, 26-31.
- Morse, S.M. (1995). Factors in the emergence of infectious diseases. *Emerging Infectious Disease*, 1: 7-15.
- Muiru, W.M., Koopmann, B., Tiedemann, A.V., Mutitu, E.W. and Kimenju, J.W. (2010). Use of repetitive extragenic palindromic (REP), enterobacterial repetitive intergenic consensus (ERIC) and BOX sequences to fingerprint *Exserohilum turcicum* isolates. *Journal of Applied Biosciences*, 30: 1828-1838.
- Nesmith, W.C. and Jenkins, S.F. (1979). A selective medium for the isolation and quantification of *Pseudomonas solanacearum* from soil. *Phytopathology*, 69: 6.
- Netto, R.A.C. and Nutter, F.W.J. (2005). Use of GPS and GIS Technologies to Map the Prevalence of Moko Disease of Banana in the Amazonas Region of Brazil. In Bacterial Wilt and the *Ralstonia solanacearum* species complex, 412-416.
- Nik Hassan, N.M.M. (2003). Banana R&D in Malaysia: Updates and highlights. In Proceedings of the 21st BAPNET Steering Committee Meeting, Jakarta, Indonesia, 75-79.
- Norman, D.J., Zapata, M., Gabriel, D.W., Duan, Y.P., Yuen, J.M., Mangravita-Novo, A. and Donahoo, R.S. (2009). Genetic diversity and host range variation of *Ralstonia solanacearum* strains entering North America. *Phytopathology*, 99: 1070-1077.
- Nouri, S., Bahar, M. and Fegan, M. (2009). Diversity of *Ralstonia solanacearum* causing potato bacterial wilt in Iran and the first record of phylotype II/biovar 2T strains outside South America. *Plant Pathology*, 58: 243-249.
- Ocho, F.L. (2006). Biochemical, Pathological and Genetic Characterization of Strains of *Ralstonia Solanacearum* (Smith) from Ethiopia and Biocontrol of *Ralstonia Solanacearum* with Bacterial Antagonists (PhD dissertation).
- Parker, I.M. and Gilbert, G.S. (2004). The evolutionary ecology of novel plant-pathogen interactions. *Annual Review of Ecology, Evolution and Systematics*, 35:675–700.
- Pasanen, T., Koskela, S., Mero, S., Tarkka, E., Tissari, P., Vaara, M. and Kirveskari, J. (2014). Rapid Molecular Characterization of *Acinetobacter baumannii* Clones with rep-PCR and Evaluation of Carbapenemase

Genes by New Multiplex PCR in Hospital District of Helsinki and Uusimaa. *PloS One*, 9: e85854.

- Peeters, N., Guidot, A., Vailleau, F. and Valls, M. (2013). *Ralstonia solanacearum*, a widespread bacterial plant pathogen in the post-genomic era. *Molecular Plant Pathology*, 14: 651-662.
- Pillay, M., Ude, G. and Kole, C. (2012). Classical genetics and traditional breeding in Musa. *Genetics, Genomics, and Breeding of Bananas*, 34-39.
- Poussier, S. and Luisetti, J. (2000). Specific detection of biovars of *Ralstonia solanacearum* in plant tissues by nested-PCR-RFLP. *Journal of Plant Pathology*, 106: 255-265.
- Poussier, S., Prior, P., Luisetti, J., Hayward, C. and Fegan, M. (2000). Partial Sequencing of the *hrpB* and Endoglucanase Genes Confirms and Expands the Known Diversity within the *Ralstonia solanacearum* Species Complex. *Systematic and Applied Microbiology*, 23: 479-486.
- Poussier, S., Trigalet-Demery, D., Vandewalle, P., Goffinet, B., Luisetti, J. and Trigalet, A. (2000). Genetic diversity of *Ralstonia solanacearum* as assessed by PCR-RFLP of the *hrp* gene region, AFLP and 16S rRNA sequence analysis, and identification of an African subdivision. *Microbiology*, 146: 1679-1692.
- Prasannakumar, M.K., Chandrashekara, K.N., Deepa, M., Vani, A. and Khan, A.N.A. (2013). Finger printing of *Ralstonia solanacearum* isolates by Rep-PCR and RAPD. *Pest Management In Horticultural Ecosystems*, 18: 179-187.
- Price, N.S. (1995). The origin and development of banana and plantain cultivation. In *Bananas and plantains* (pp. 1-13). Springer: Netherlands.
- Prieto Romo, J., Morales Osorio, J.G. and Salazar Yepes, M. (2012). Identification of new hosts for *Ralstonia solanacearum* (Smith) race 2 from Colombia. *Revista de Protección Vegetal*, 27: 151-161.
- Prior, P. and Fegan, M. (2004). Recent developments in the phylogeny and classification of *Ralstonia solanacearum*. In *International Symposium on Tomato Diseases* 695 (pp. 127-136).
- Prior, P., Fegan, M., Allen, C. and Hayward, A.C. (2005). Diversity and molecular detection of *Ralstonia solanacearum* race 2 strains by multiplex PCR. In Bacterial wilt disease and the *Ralstonia solanacearum* species complex, 405-414.
- Rahman, M.F., Islam, M.R., Rahman, T. and Meah, M.B. (2013). Biochemical Characterization of *Ralstonia solanacerum* Causing

Bacterial Wilt of Brinjal in Bangladesh. *Progressive Agriculture*, 21: 9-19.

- Rambaut, A. and Drummond, A. (2009). FigTree v1.4. (<u>http://tree.bio.ed.ac.uk/software/figtree/</u>).
- Raymundo, A.K., Orlina, M.E., Lavina, W.A. and Opina, N.L. (2005). Comparative genome plasticity of tomato and banana strains of *Ralstonia solanacearum* in the Philippines. *In* Bacterial Wilt Disease and the *Ralstonia solanacearum* Species Complex, eds. C. Allen, P. Prior and A.C. Hayward, APS Press, pp. 387-393.
- Remenant, B., Coupat-Goutaland, B., Guidot, A., Cellier, G., Wicker, E., Allen, C., .... and Prior, P. (2010). Genomes of three tomato pathogens within the *Ralstonia solanacearum* species complex reveal significant evolutionary divergence. *BMC Genomics*, 11: 379.
- Remenant, B., de Cambiaire, J.C., Cellier, G., Jacobs, J.M., Mangenot, S., Barbe, Vallanet, D., Medigue, C., Fegan, M., Allen, C. and Prior, P. (2011). *Ralstonia syzygii*, the blood disease bacterium and some Asian *R. solanacearum* strains form a single genomic species despite divergent lifestyles. *PLoS One*, 6: e24356.
- Rodrigues, L.M., Destéfano, S.A., Diniz, M.C.T., Comparoni, R. and Rodrigues Neto, J. (2011). Pathogenicity of Brazilian strains of *Ralstonia solanacearum* in *Strelitzia reginae* seedlings. *Tropical Plant Pathology*, 36: 409-413.
- Ronquist, F. and Huelsenbeck, J.P. (2003). MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics*, 19: 1572-1574.
- Rouhrazi, K. and Khodakaramian, G. (2014). Genetic fingerprinting of Iranian *Xanthomonas campestris* pv. *campestris* strains inducing black rot disease of crucifers. *European Journal of Plant Pathology*, 139: 175-184.
- Sahilah, A.M., Tosiah, S., Radu, S., Ooi, W.L., Queen, C.Y.K., Jeffrey, L.S.H., Chua, S.C. and Senawi, M.T. (2005). Typing of *Ralstonia solanacearum* isolated from tomato by antibiotic susceptibility, plasmid profiling and PCR-based techniques of RAPD and ERIC. *Journal of Tropical Agriculture and Food Science*, 33: 73-82.
- Saile, E., McGarvey, J.A., Schell, M.A. and Denny, T.P. (1997). Role of extracellular polysaccharide and endoglucanase in root invasion and colonization of tomato plants by *Ralstonia solanacearum*. *Phytopathology*, 87: 1264-1271.

- Salanoubat, M., Genin, S., Artiguenave, F., Gouzy, J., Mangenot, S., Arlat, M., ... and Boucher, C. A. (2002). Genome sequence of the plant pathogen Ralstonia solanacearum. *Nature*, 415: 497-502.
- Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989). In *Molecular cloning: a laboratory manual*, 2<sup>nd</sup> Edition, New York: Cold Spring Harbor Laboratory Press.
- Sands, D.C. (1990). Physiological criteria-determinative tests. *Methods in Phytobacteriology*, 133-143.
- Santana, B.G., Lopes, C.A., Alvarez, E., Barreto, C.C., Allen, C. and Quirino, B.F. (2012). Diversity of Brazilian biovar 2 strains of *Ralstonia* solanacearum. Journal of General Plant Pathology, 78: 190-200.
- Schaad, N.W., Jones, J.B. and Chun, W. (2001). Laboratory Guide for Identification of Plant Pathogenic Bacteria. Minnesota, USA, APS Press.
- Schell, M.A. (2000). Control of virulence and pathogenicity genes of *Ralstonia solanacearum* by an elaborate sensory network. *Annual Review of Phytopathology*, 38: 263-292.
- Seal, S.E., Jackson, L.A., Young, J.P.W. and Daniels, M.J. (1993). Differentiation of *Pseudomonas solanacearum*, *Pseudomonas syzygii*, *Pseudomonas pickettii* and the blood disease bacterium by partial 16S rRNA sequencing: construction of oligonucleotide primers for sensitive detection by polymerase chain reaction. *Journal of General Microbiology*, 139: 1587-1594.
- Sequeira, L. (1998). Bacterial wilt: the missing element in international banana improvement programs. *In* Bacterial Wilt Disease: Molecular and Ecological Aspects, eds. P. Prior, C. Allen and J. Elphinstone, Springer-Verlag, pp. 6-14.
- Simmonds, N.W. (1986). Classification and Breeding of Bananas. *In* Banana Plantain and Breeding Strategies. Proceedings of an International Workshop, 23<sup>rd</sup> October 1986, Cairns, Australia, pp. 69-73.
- Someya, S., Yoshiki, Y. and Okubo, K. (2002). Antioxidant compounds from bananas (*Musa Cavendish*). *Food Chemistry*, 79: 351-354.
- Stander, E.I.M., Hammes, P.S. and Beyers, E.A. (2013). Survival of *Ralstonia solanacearum* biovar 2 in soil under different cropping systems. *South African Journal of Plant and Soil*, 20: 176-179.
- Staskawicz, B.J., Ausubel, F.M., Baker, B.J., Ellis, J.G. and Jones, J.D. (1995). Molecular genetics of plant disease resistance. *Science-New York Then Washington*, 661-661.

- Stefan Van Dongen, T. and Winnepenninckx, B. (1996). Multiple UPGMA and neighbor-joining trees and the performance of some computer packages. *Molecular Biology and Evolution*, 13: 309-313.
- Stover, R.H. (1972). Banana, plantain and abaca disease. *Commonwealth Mycological Institute.* Kew, Survey. England. pp. 316.
- Stuiver, M.H. and Custers, J.H. (2001). Engineering disease resistance in plants. *Nature*, 411: 865-868.
- Sulaiman, S.F., Yusoff, N.A.M., Eldeen, I.M., Seow, E.M., Sajak, A.A.B. and Ooi, K.L. (2011). Correlation between total phenolic and mineral contents with antioxidant activity of eight Malaysian bananas (*Musa* sp.). *Journal of Food Composition and Analysis*, 24: 1-10.
- Sun, J., Chu, Y.F., Wu, X. and Liu, R.H. (2002). Antioxidant and antiproliferative activities of common fruits. *Journal of Agricultural and Food Chemistry*, 50: 7449-7454.
- Suslow, T.V., Schroth, M.N. and Isaka, M. (1982). Application of a rapid method for Gram differentiation of plant pathogenic and saprophytic bacteria without staining. *Phytopathology*, 72: 917-918.
- Taghavi, M., Hayward, C., Sly, L.I. and Fegan, M. (1996). Analysis of the phylogenetic relationships of strains of *Burkholderia solanacearum*, *Pseudomonas syzygii*, and the blood disease bacterium of banana based on 16S rRNA gene sequences. *International Journal of Systematic Bacteriology*, 46: 10-15.
- Tans-Kersten, J., Brown, D. and Allen, C. (2004). Swimming motility, a virulence trait of *Ralstonia solanacearum*, is regulated by FlhDC and the plant host environment. *Molecular Plant-Microbe Interactions*, 17: 686-695.
- Tengku Ab. Malik, T.M., Rozeita, L., Maimun, T. and Umi Kalsum, B. (2011). Status of banana diseases research in Malaysia. In: Abstract of the Workshop on Integrated Approaches in Banana Disease Management, 22<sup>th</sup> March 2011, Serdang, Malaysia, pp. 5.
- Thwaites, R, Eden-Green, S.J. and Black, R. (2000). Disease Caused by Bacteria. In Diseases of Banana, Abaca and Enset, ed. D.R Jones, CABI Publishing: UK, pp. 213-239.
- Thwaites, R., Mansfield, J., Eden-Green, S. and Seal, S. (1999). RAPD and rep PCR-based fingerprinting of vascular bacterial pathogens of *Musa* spp.. *Plant Pathology*, 48: 121-128.
- Tsujimoto, S., Nakaho, K., Adachi, M., Ohnishi, K., Kiba, A. and Hikichi, Y. (2008). Contribution of the type II secretion system in systemic

infectivity of *Ralstonia solanacearum* through xylem vessels. *Journal of General Plant Pathology*, 74: 71-75.

- Valmayor, R.V. (2000). *Banana cultivar names and synonyms in Southeast Asia*. Bioversity International.
- Vandamme, P. and Peeters, C. (2014). Time to revisit polyphasic taxonomy. *Antonie van Leeuwenhoek*, 106: 57-65.
- Versalovic, J., Schneider, M., De Bruijn, F.J. and Lupski, J.R. (1994). Genomic fingerprinting of bacteria using repetitive sequence-based polymerase chain reaction. *Methods in Molecular and Cellular Biology*, 5: 25-40.
- Villa, J.E., Tsuchiya, K., Horita, M., Opina, N. and Hyakumachi, M. (2005). Phylogenetic relationships of *Ralstonia solanacearum* species complex strains from Asia and other continents based on 16S rDNA, endoglucanase, and *hrpB* gene sequences. *Journal of General Plant Pathology*, 71: 39-46.
- Vinatzer, B.A. (2012). "Listening In" on How a Bacterium Takes Over the Plant Vascular System. *mBio*, 3: e00269-12.
- Wall, M.M. (2006). Ascorbic acid, vitamin A, and mineral composition of banana (*Musa* sp.) and papaya (*Carica papaya*) cultivars grown in Hawaii. *Journal of Food Composition and Analysis*, 19: 434-445.
- Wardlaw, C.W. (1972). Banana diseases: Including plantains and abaca. Longman Group Limited: London.
- Wicker, E., Grassart, L., Coranson-Beaudu, R., Mian, D., Guilbaud, C., Fegan, M. and Prior, P. (2007). *Ralstonia solanacearum* strains from Martinique (French West Indies) exhibiting a new pathogenic potential. *Applied and Environmental Microbiology*, 73: 6790-6801.
- Wicker, E., Grassart, L., Coranson-Beaudu, R., Mian, D. and Prior, P. (2009). Epidemiological evidence for the emergence of a new pathogenic variant of *Ralstonia solanacearum* in Martinique (French West Indies). *Plant Pathology*, 58: 853-861.
- Winstead, N.N. and Kelman, A. (1952). Inoculation techniques for evaluating resistance to *Pseudomonas solanacearum*. *Phytopathology*, 42: 628-634.
- Woods, A.C. (1984). Moko Disease: Atypical Symptoms Induced by Afluidal Variants of *Pseudomonas solanacearum* in Banana Plants. *Phytopathology*, 74: 972-976.

- Xu, J., Pan, Z.C., Prior, P., Xu, J.S., Zhang, Z., Zhang, H., Zhang, L.Q., He, L.Y and Feng, J. (2009). Genetic diversity of *Ralstonia solanacearum* strains from China. *European Journal of Plant Pathology*, 125: 641-653.
- Yabuuchi, E., Kosako, Y., Oyaizu, H., Yano, I., Hotta, H., Hashimoto, Y., Takayuki E. and Michio A. (1992). Proposal of *Burkholderia* gen. nov. and transfer of seven species of the genus *Pseudomonas* homology group II to the new genus, with the type species *Burkholderia cepacia* (Palleroni and Holmes 1981) comb. nov. *Microbiology and Immunology*, 36: 1251-1275.
- Yabuuchi, E., Kosako, Y., Yano, I., Hotta, H. and Nishiuchi, Y. (1995). Transfer of Two *Burkholderia* and An *Alcaligenes* Species to *Ralstonia* Gen. Nov. *Microbiology and Immunology*, 39: 897-904.
- Yao, J. and Allen, C. (2006). Chemotaxis is required for virulence and competitive fitness of the bacterial wilt pathogen *Ralstonia solanacearum. Journal of Bacteriology*, 188: 3697-3708.
- Yao, J. and Allen, C. (2007). The plant pathogen *Ralstonia solanacearum* needs aerotaxis for normal biofilm formation and interactions with its tomato host. *Journal of Bacteriology*, 189: 6415-6424.
- Zhang, C.X., Ho, S.C., Chen, Y.M., Fu, J.H., Cheng, S.Z. and Lin, F.Y. (2009). Greater vegetable and fruit intake is associated with a lower risk of breast cancer among Chinese women. *International journal of cancer*, 125: 181-188.
- Zhang, P., Whistler, R.L., BeMiller, J.N. and Hamaker, B.R. (2005). Banana starch: production, physicochemical properties, and digestibility—a review. *Carbohydrate Polymers*, 59: 443-458.