

ORIGINAL ARTICLE

Viruses causing lower respiratory symptoms in young children: findings from the ORChID birth cohort

Mohinder Sarna,^{1,2} Stephen B Lambert,^{2,3} Theo P Sloots,^{2,3} David M Whiley,^{2,4} Asma Alsaleh,⁵ Lebogang Mhango,^{2,3} Seweryn Bialasiewicz,^{2,3} David Wang,⁶ Michael D Nissen,^{2,3} Keith Grimwood,^{7,8} Robert S Ware^{1,2,9}

ABSTRACT

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¹School of Public Health. The University of Queensland, Brisbane, Australia ²UQ Child Health Research Centre, The University of Queensland, Brisbane, Australia ³Queensland Paediatric Infectious Diseases Laboratory, Children's Health Oueensland. Brisbane, Australia ⁴University of Queensland Centre for Clinical Research, The University of Queensland, Brisbane, Queensland, Australia ⁵Department of Botany and Microbiology, King Saud University, Riyadh, Saudi Arabia ⁶School of Medicine, Washington University in St Louis, St Louis, Missouri, USA ⁷School of Medicine and Menzies Health Institute Queensland, Griffith University, Brisbane, Australia ⁸Departments of Infectious Diseases and Paediatrics, Gold Coast Health, Gold Coast, Australia ⁹Menzies Health Institute Queensland, Griffith University, Brisbane, Queensland, Australia

Correspondence to

Ms. Mohinder Sarna, Centre for Children's Health Research, 62 Graham Street, South Brisbane, Queensland 4101, Australia; m.sarna@uq.edu.au

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To cite: Sarna M, Lambert SB, Sloots TP, et al. Thorax 2018;73:969–979. **Introduction** Viral acute respiratory infections (ARIs) cause substantial child morbidity. Sensitive molecularbased assays aid virus detection, but the clinical significance of positive tests remains uncertain as some viruses may be found in both acutely ill and healthy children. We describe disease-pathogen associations of respiratory viruses and quantify virus-specific attributable risk of ARIs in healthy children during the first 2 years of life.

Methods One hundred fifty-eight term newborn babies in Brisbane, Australia, were recruited progressively into a longitudinal, community-based, birth cohort study conducted between September 2010 and October 2014. A daily tick-box diary captured predefined respiratory symptoms from birth until their second birthday. Weekly parent-collected nasal swabs were batch-tested for 17 respiratory viruses by PCR assays, allowing calculation of virus-specific attributable fractions in the exposed (AFE) to determine the proportion of virus-positive children whose ARI symptoms could be attributed to that particular virus.

Results Of 8100 nasal swabs analysed, 2646 (32.7%) were virus-positive (275 virus codetections, 3.4%), with human rhinoviruses accounting for 2058/2646 (77.8%) positive swabs. Viruses were detected in 1154/1530 (75.4%) ARI episodes and in 984/4308 (22.8%) swabs from asymptomatic periods. Respiratory syncytial virus (AFE: 68% (95% CI 45% to 82%)) and human metapneumovirus (AFE: 69% (95% CI 43% to 83%)) were strongly associated with higher risk of lower respiratory symptoms.

Discussion The strong association of respiratory syncytial virus and human metapneumovirus with ARIs and lower respiratory symptoms in young children managed within the community indicates successful development of vaccines against these two viruses should provide substantial health benefits.

INTRODUCTION

Viral acute respiratory infections (ARIs) are a common cause of morbidity in children, either on their own or in synergy with bacterial pathogens.¹ Virus-positive ARIs early in life are associated with developing wheeze and asthma during childhood and adolescence.² Community-based studies from the 1950s to 1960s examining the aetiology of ARIs by traditional laboratory detection methods remain the template for understanding childhood respiratory virus epidemiology.³ Advances in molecular

Key messages

What is the key question?

What is the attributable risk for symptoms of acute respiratory infections (ARIs) when specific respiratory viruses are detected by PCR assays in nasal swab samples collected weekly from otherwise healthy young children?

What is the bottom line?

Attribution for ARIs by specific viruses was strongest for respiratory syncytial virus and human metapneumovirus in young children with lower respiratory tract infections, while among DNA viruses tested, only adenoviruses were significantly associated with an increased risk of symptoms.

Why read on?

The virus-specific attributable risk of ARIs is a population-level metric to inform causal inference in the aetiology of ARIs and provides quantitative estimates of the relative proportions of virus-associated ARIs in the community, yielding important insights for prioritising public health interventions.

diagnostics, including PCR testing, have improved virus detection in those with ARIs.⁴ However, this increased test sensitivity has also resulted in viruses being detected in asymptomatic individuals.^{4 5} The clinical significance of finding respiratory viruses is not always straightforward, and a better understanding of the proportion of infections that are symptomatic for individual viruses is needed.

The attributable fraction in the exposed (AFE) provides information about the percentage of cases among exposed individuals that can be attributed to the exposure. In our study, the AFE measures the percentage of ARIs in virus-positive children attributable to that particular virus. The AFE does not inform on the aetiology at an individual level, but is a key population-level metric for interpreting positive specimen results. It assists with estimation of the true burden of disease, thus guiding preventative strategies for respiratory infections, such as immunisation. Virus-specific AFE estimates for ARI in children are reported infrequently, with most studies reporting AFEs having important methodological limitations. Except for one community





Figure 1 Submission of nasal swabs and symptom diaries from participants in the Observational Research in Childhood Infectious Diseases study. Ct, cycle threshold; ERV-3, endogenous retrovirus-3.

study,⁵ these investigations recruited either highly selected hospitalised children or outpatients as study cases,⁶⁻⁸ or used controls from outpatient clinics.⁶ ⁷ A meta-analysis of 23 studies that calculated virus-specific AFEs included 22 studies employing hospital-based case ascertainment and only three using community-based controls.⁹ Other studies comparing symptomatic and asymptomatic groups calculated odds ratios (ORs),^{10–12} or relative risks,¹³ without calculating AFEs. Moreover, several studies focused solely either on specific viruses^{10 12} or outcomes.^{6 11–13}

Longitudinal, community-based studies employing sensitive molecular-based assays with regular and frequent sampling, irrespective of illness, are best suited to explore the relationship between respiratory virus detections and ARI symptoms, especially as they represent most ARI episodes. The aim of our study was to examine the disease-pathogen associations of respiratory viruses in infants participating in a community-based birth cohort project¹⁴ and to calculate their virus-specific AFE values.

METHODS

Study population

The Observational Research in Childhood Infectious Diseases (ORChID) project (clinicaltrials.gov: NCT01304914) is an observational, longitudinal community-based birth cohort study of ARI episodes in children until their second birthday.¹⁴ Participants were able to leave the study temporarily (such as during family holidays) and rejoin at a later date. It was conducted in the subtropical city of Brisbane, Australia (latitude 27° South, average monthly maximum temperature range 22°C–30°C, maximum rainfall in December-February, population 2.2 million) between September 2010 and October 2014. Parents consented antenatally for their child's participation. For enrolment, infants

needed to be healthy, without congenital abnormalities or underlying chronic disorders, and to be born at term (36–42 weeks). Recruitment was progressive over 2 years at one of two metropolitan hospitals (one private and one government-funded). Detailed methods of recruitment, study design and sample and data collection are described elsewhere.¹⁴ The Children's Health Queensland, Royal Brisbane and Women's Hospital, and The University of Queensland Human Research Ethics Committees approved the study.

Sociodemographic and clinical characteristics

Following recruitment, parents provided their sociodemographic and health history, pregnancy and birth details. Telephone interviews were conducted every 3 months to update immunisation, feeding and childcare attendance details. Childcare was categorised as formal (regulated care outside the child's home) or informal (non-regulated care provided by family or friends).¹⁵ The duration of exclusive breast feeding was from birth to the introduction of either any milk formula or solids.

Symptom data collection

Parents were trained to recognise respiratory symptoms and provided with diary cards designed in a tick-box format and listed with predefined respiratory symptoms or diagnoses to be completed daily and returned by regular mail monthly. The diary cards were modified from a prior study¹⁶ and used successfully elsewhere.¹⁷

Virus detection

Viruses were identified from parent-collected anterior nose swabs at the infant's birth, and weekly thereafter.¹⁸ Specimens were mailed

Table 1	Characteristics of children enrolled in the Observational
Research	in Childhood Infectious Diseases study

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Characteristic (n=158 unless stated)	Frequency (%) or mean (±SD)
Sex (male)	75 (47.5%)
Gestational age (weeks)	39.8 (±1.3)
Birth weight (g)	3530.8 (±430.4)
Delivery	
Vaginal	107 (67.7%)
Birth order (maternal)	
First born	106 (67.1%)
Parental mean age	
Mother (years) (n=157)	31.8 (±4.6)
Family history of asthma or eczema	
Mother	50 (31.6%)
Father	44 (27.8%)
Sibling	15 (9.5%)
First-degree relative with asthma or eczema	83 (52.5%)
Smoke exposure at birth	
Mother (n=156)	5 (3.2%)
Other householder (n=155)*	17 (11.0%)
Maternal educational status (n=157)	
Primary school	5 (3.2%)
High school	15 (9.6%)
Diploma/certificate	38 (24.2%)
University or higher university degree	99 (63.1%)
Household income (n=155)†	
Lowest quarter	0
Lower quarter	17 (11.0%)
Higher quarter	52 (33.5%)
Highest quarter	86 (55.5%)
Child immunisation status to 18 months (n=154) \ddagger	
Fully immunised	141 (91.6%)
Exclusive breast feeding	
At birth (n=149)	142 (95.3%)
At 3 months (n=142)	97 (68.3%)
At 6 months (n=133)	5 (3.8%)
Childcare	
At 6 months (n=133)	33 (24.8%)
At 9 months (n=123)	48 (39.0%)
At 12 months (n=115)	72 (62.6%)
At 15 months (n=110)	87 (79.1%)

*Includes two single parent households.

†Income categories based on Australian Bureau of Statistics income quartiles (quartile 1: <\$A26 000; quartile 2: \$A26 000-\$A67 499; quartile 3: \$A67 500-\$A114 999; quartile 4: ≥\$A115 000).</p>

±11/154 (7.1%) infants did not receive a full course of rotavirus vaccines because they presented for immunisation outside of required age limits; 2 children had chickenpox diagnosed before the varicella vaccine could be administered at the scheduled age of 18 months, but were up-to-date for all other vaccines; another 2 children withdrew early and information was unavailable on their later immunisations. Influenza vaccines are not part of the national Australian infant immunisation schedule. Only 7 children received the influenza vaccine in the first 2 years of life.

to the laboratory, taking a median of 3 (IQR 2–4) days, and where they were processed and stored at -80° C until further analysis.^{14 19}

Swabs were batch-tested for 17 respiratory viruses: human rhinovirus (HRV), influenza A and B (IFV-A, IFV-B), respiratory syncytial virus A and B (RSV-A, RSV-B), parainfluenza viruses 1–3 (PIV-1, PIV-2, PIV-3), human coronaviruses (HCoV) OC43, NL63, 229E and HKU1, human metapneumovirus (HMPV), adenovirus (AdV), human polyomaviruses WU (WU-PyV) and KI (KI-PyV) and human bocavirus-1 (HBoV-1), using previously validated real-time PCR assays.¹⁴ Details of sample collection, processing, PCR assay parameters, quality control measures and HRV typing into species A, B and C are outlined in the online supplementary text and E-table 1. Specimen quality was assessed by testing for a marker of human genomic DNA, endogenous retrovirus-3 (ERV-3).²⁰ Swabs with ERV-3 cycle-threshold (Ct) values >38 (including undetectable Ct values) were deemed to be of lesser quality and excluded from parts of the analysis.²¹

Definitions

New virus detection episode

When a new virus (different virus or new species/subtype of the same genus or species, respectively) was detected in a swab by PCR testing, or the same virus was detected after at least two negative intervening swabs, or the same virus was detected at least 30 days from the last positive swab.

Symptomatic new virus detection episode

When symptoms were reported 7 days before or 7 days after detecting a new virus.

ARI episode

ARI was defined as a new episode of respiratory symptoms of the upper and/or lower respiratory tracts demarcated by a window of ≥ 3 symptom-free days.^{17 22}

Upper respiratory tract infection

Parent-reported runny nose/nasal congestion, dry cough or doctor-diagnosed otitis media.²²

Lower respiratory tract infection

Parent-reported rattly breathing, moist cough, shortness of breath, wheeze or doctor-diagnosed pneumonia.²²

ARIs were subcategorised hierarchically as either a lower respiratory tract infection (LRTI) or upper respiratory tract infection (URTI), respectively.

Analysis

The incidence rate of single new virus detection episodes (VDEs) per 100 child-years was calculated using Poisson regression. To account analytically for the differing lengths of time children remained in the study, the natural logarithm of the number of swabs returned was included in the model as an offset. Swabs were removed from incidence rate calculations for individual viruses when the participant was not at risk of a new VDE (a positive consecutive swab following the initial positive swab, and two swabs after the last positive swab, as per our definition of a new VDE), or the swabs were of lesser quality (to avoid underestimating incidence rates).²¹ When calculating incidence rates for 'any virus', we assumed children were at risk of being positive for new viruses in every swab.

The virus-specific risk and AFE of single VDEs in cases of ARI and LRTI were estimated using a generalised linear model with binomial family and log link. Robust variance estimates were calculated using sandwich estimators to account for repeated measures within children. We analysed LRTIs separately as children with these infections are more likely to visit family physicians, present to hospital and receive antibiotics.²² The AFE was

Table 2Respiratory viruses detected in 8100 high-quality nasal swabs collected from 157 infants from the Observational Research in ChildhoodInfectious Diseases (ORChID) birth cohort in the first 2 years of life. Incidence rates/100 child-years presented overall, and for the first and second
years of life separately

	Virus-positive swabs	New virus detections N=1520*	New virus detection episodes/100 child-years (overall year mean)	New virus detection episodes/100 child-years (first year)	New virus detection episodes/100 child-years (second year)
Virus	n (%)	n (%)	Mean (95% CI)	Mean (95% CI)	Mean (95% CI)
All HRV†	2058 (25.4)	1012 (66.6)	1002 (942 to 1066)	770 (703 to 842)	1354‡ (1245 to 1472)
HRV-A	702 (8.7)	440 (28.9)	323 (295 to 355)	258 (224 to 297)	409 (361 to 464)
HRV-B	154 (1.9)	97 (6.4)	64 (53 to 78)	60 (45 to 79)	69 (52 to 92)
HRV-C	635 (7.8)	495 (32.6)	363 (332 to 396)	295 (259 to 336)	448 (398 to 505)
IFV-A	12 (0.1)	12 (0.8)	7.7 (4.4 to 13.6)	4.7 (1.8 to 12.5)	11.5 (5.7 to 22.9)
IFV-B	2 (0.0)	2 (0.1)	1.3 (0.3 to 5.2)	1.2 (0.2 to 8.3)	1.4 (0.2 to 10.1)
PIV-1	7 (0.1)	7 (0.5)	4.5 (2.2 to 9.5)	2.3 (0.6 to 9.3)	7.2 (3.0 to 17.2)
PIV-2	4 (0.0)	4 (0.3)	2.6 (1.0 to 6.9)	1.2 (0.2 to 8.3)	4.3 (1.4 to 13.3)
PIV-3	59 (0.7)	56 (3.7)	36.5 (28.1 to 47.4)	27.1 (18.0 to 40.8)	47.8 (34.0 to 67.3)
RSV-A	69 (0.9)	58 (3.8)	37.9 (29.3 to 49.0)	30.7 (20.9 to 45.1)	46.4 (32.8 to 65.6)
RSV-B	22 (0.3)	20 (1.3)	12.9 (8.3 to 20.1)	8.2 (3.9 to 17.2)	18.7 (10.9 to 32.2)
HCoV-OC43	51 (0.6)	42 (2.8)	27.3 (20.2 to 37.0)	18.8 (11.5 to 30.7)	39.1 (26.8 to 57.0)
HCoV-NL63	56 (0.7)	40 (2.6)	26.0 (19.1 to 35.5)	18.8 (11.5 to 30.7)	37.7 (25.6 to 55.3)
HCoV-229E	14 (0.2)	9 (0.6)	5.8 (3.0 to 11.2)	4.7 (1.8 to 12.5)	7.2 (3.0 to 17.2)
HCoV-HKU1	34 (0.4)	28 (1.8)	18.2 (12.5 to 26.3)	10.5 (5.5 to 20.3)	27.4 (17.5 to 43.0)
HMPV	27 (0.3)	23 (1.5)	14.9 (9.9 to 22.4)	12.9 (7.1 to 23.3)	17.2 (9.8 to 30.4)
AdV	104 (1.3)	80 (5.3)	52.6 (42.2 to 65.4)	47.6 (34.9 to 64.9)	58.3 (42.8 to 79.5)
WU-PyV	101 (1.2)	60 (3.9)	39.3 (30.5 to 50.6)	16.5 (9.8 to 27.8)	67.6 (50.6 to 90.2)
KI-PyV	168 (2.1)	84 (5.5)	55.7 (45.0 to 68.9)	51.6 (38.3 to 69.5)	60.4 (44.5 to 82.1)
HBoV-1	137 (1.7)	90 (5.9)	59.6 (48.5 to 73.3)	47.8 (35.1 to 65.2)	73.8 (55.9 to 97.4)
Any virus§	2646 (32.7)	1520 (18.8)	978 (930 to 1029)	794 (736 to 856)	1206 (1127 to 1290)
Codetections§	275 (3.4)	237 (15.6)	147 (130 to 168)	117 (96 to 142)	184 (155 to 219)

*Includes codetections.

†666 HRV (28.5%) positive samples could not be sequenced primarily because of low viral loads.

*The rate of HRV new VDEs and the mean incidence of HRV new VDEs may equal or exceed the total rate of all new VDEs depending on the denominator after all swabs with cycle-threshold values >38 are excluded.

\$Days at risk were not removed from the denominator as an infant could be at risk of being positive for other viruses in every swab.

HBoV-1, human bocavirus-1; HCoV, human coronavirus; HMPV, human metapneumovirus; HRV, human rhinovirus; IFV, influenza virus; KI-PyV, KI polyomavirus; PIV, parainfluenza virus; RSV, respiratory syncytial virus; VDE, virus detection episode; WU-PyV, WU polyomavirus.





calculated as $100\% \times (1-RR^{-1})$, where risk ratio (RR) is the proportion of children positive for a virus with ARI symptoms, divided by the proportion of children negative for the virus who have ARI symptoms (see online supplementary text 2). Only virus-negative swabs of lesser quality (Ct > 38) were excluded in these analyses of association, as swab quality of positive swabs did not modify the association between VDEs and ARI symptoms (see online supplementary E-table 2).

The association between potential risk factors (age, sex, type of delivery, gestational age at birth, season, family history of asthma or eczema, tobacco smoke exposure, household size, maternal education status, mode of feeding, category of childcare attendance) and symptomatic and asymptomatic virus detection was examined using mixed-effects logistic regression with the child entered as a random effect to account for repeated measurements within children. Both univariable and multivariable analyses were conducted. Breast feeding and childcare attendance were analysed as timevarying variables. Multivariable models included all the above risk factors with no forward or backward selection. Only virus-negative

 Table 3
 Acute respiratory infections and asymptomatic single new virus detections, risk ratios and attributable fractions by respiratory virus in 151

 children from the ORChID birth cohort (n=8202)

	ARI-associated single new virus detections N=3894 swabs	Asymptomatic single new virus detections N=4308 swabs	Risk ratio* (95% CI); P value	Attributable fraction in the exposed (%, 95% Cl)†
Virus	n (%)	n (%)		
Single new detections				
All HRV	679 (17.4)‡	328 (7.6)	1.5 (1.4 to 1.6); <0.001	34 (29 to 38)
HRV-A	289 (7.4)	122 (2.8)	1.5 (1.4 to 1.6); <0.001	34 (28 to 39)
HRV-B	49 (1.3)	41 (1.0)	1.2 (0.9 to 1.4); 0.17	14 (-6 to 30)
HRV-C	328 (8.4)	137 (3.2)	1.5 (1.4 to 1.7); <0.001	35 (30 to 40)
IFV-A	9 (0.2)	2 (0.0)	1.7 (1.3 to 2.3); <0.001	42 (23 to 57)
IFV-B	2 (0.1)	2 (0.0)	1.1 (0.4 to 2.8); 0.90	6 (- 150 to 64)
PIV-1	6 (0.2)	1 (0.0)	1.8 (1.3 to 2.5); <0.001	45 (26 to 59)
PIV-2	3 (0.1)	0 (0.0)	n/c	n/c
PIV-3	32 (0.8)	11 (0.3)	1.5 (1.3 to 1.9); <0.001	35 (21 to 46)
RSV-A	29 (0.7)	14 (0.3)	1.4 (1.2 to 1.7); <0.001	30 (16 to 42)
RSV-B	19 (0.5)	3 (0.1)	1.8 (1.5 to 2.2); <0.001	46 (35 to 54)
HCoV-OC43	26 (0.7)	8 (0.2)	1.6 (1.3 to 2.0); <0.001	38 (25 to 50)
HCoV-NL63	27 (0.7)	8 (0.2)	1.6 (1.3 to 2.0); <0.001	37 (21 to 49)
HCoV-229E	3 (0.1)	4 (0.1)	0.9 (0.4 to 2.2); 0.83	-10 (-161 to 54)
HCoV-HKU1	12 (0.3)	10 (0.2)	1.2 (0.8 to 1.7); 0.42	14 (- 24 to 40)
HMPV	12 (0.3)	4 (0.1)	1.6 (1.2 to 2.1); 0.001	37 (16 to 53)
AdV	28 (0.7)	16 (0.4)	1.4 (1.1 to 1.7); 0.008	29 (12 to 42)
WU-PyV	16 (0.7)	15 (0.3)	1.1 (0.8 to 1.6); 0.62	9 (- 31 to 36)
KI-PyV	23 (0.6)	34 (0.8)	0.9 (0.6 to 1.2); 0.37	–17 (–65 to 17)
HBoV-1	35 (0.9)	31 (0.7)	1.1 (0.9 to 1.4); 0.30	11 (- 12 to 29)
Any virus	961 (24.6)	491 (11.4)	1.5 (1.4 to 1.6); <0.001	35 (31 to 38)
New codetections§				
≥2 viruses	168 (4.3)	62 (1.4)	1.6 (1.4 to 1.7); <0.001	37 (30 to 42)

Statistically significant risk ratios and attributable fractions have been highlighted in bold.

*Adjusted for repeated measures within children using sandwich estimators.

 $\ensuremath{^+\text{CI}}$ for attributable fractions were calculated from CI for corresponding risk ratios.

Individual HRV species do not add up to 'all HRV' detections because of a combination of more than one HRV species detected within the same ARI episode, or single new virus detection episodes where HRV-positive specimens could not be sequenced.

§Includes 219 (160 symptomatic) new codetections and another 11 codetections with 2 and 3 viruses codetected, respectively where at least 1 codetected virus was a new detection.

ARI, acute respiratory infection; AdV, adenovirus; HBoV-1, human bocavirus-1; HCoV, human coronavirus; HMPV, human metapneumovirus; HRV, human rhinovirus; IFV, influenza virus; KI-PyV, KI polyomavirus; n/c, not calculable as there were no cases in the asymptomatic group; PIV, parainfluenza virus; RSV, respiratory syncytial virus; WU-PyV, WU polyomavirus.

lesser quality swabs were excluded in these analyses. As it was not possible to determine individual virus contributions to ARI symptoms during virus codetections, samples with more than one virus detected were analysed descriptively (see online supplementary E-table 3). Data were analysed using Stata V.12.1 (StataCorp, College Station, Texas, USA).

RESULTS

Cohort characteristics

Of 891 potential participants approached, 165 (18.5%) eligible singleton infants from 163 families were enrolled (two families enrolled two siblings) (figure 1). Seven were excluded subsequently; one from ineligibility (born <36 weeks) and six for failing to provide any swabs. Of the remaining children, 158 provided 11125 swabs (figure 1; 68.3% of maximum expected; median 84.5, range 1–104) from birth until their second birthday, and 154 infants provided 87547 child-days of

symptom diaries (77.9% of maximum expected days; median 726 (range 1–730 days)) with 67% followed until at least age 23 months (see online supplementary E-figure). One or more respiratory symptoms were recorded on 16877 (19.3%) days, and corresponded to 1641 discrete ARI episodes.

Demographic and social characteristics are reported in table 1. Compared with the general population of Brisbane and the state of Queensland, study children were from smaller families of advantaged backgrounds (see online supplementary E-table 4).

Virus detections

When calculating incidence rates, of the 11192 total swabs submitted by all children, 3025 (27.2%) lesser-quality swabs (346 virus-positive and 2679 virus-negative) were removed from the analyses, as were 67 swabs submitted after the child's second birthday, leaving 8100 swabs from 157 infants (figure 1). Overall,

Table 4	Lower respiratory tract infections an	d asymptomatic single new virus	detections, risk ratios a	nd attributable fractions by resp	iratory virus
in 151 chi	Idren from the ORChID birth cohort (n=5267 swabs)			

	LRTI-associated single new virus detections n=959 swabs	Asymptomatic single new virus detections n=4308 swabs	Risk ratio* (95% CI); P value	Attributable fraction in the exposed (%, 95% CI)†
Virus	n (%)	n (%)		
Single detections				
All HRV	126 (13.1)‡	328 (7.6)	1.1 (1.0 to 1.2); 0.22	8 (-5 to 20)
HRV-A	57 (6.0)	122 (2.8)	1.2 (1.0 to 1.5); 0.11	17 (-4 to 34)
HRV-B	8 (0.8)	41 (1.0)	0.8 (0.4 to 1.5); 0.43	-31 (-157 to 33)
HRV-C	68 (7.1)	137 (3.2)	1.3 (1.1 to 1.6); 0.01	22 (5 to 36)
IFV-A	2 (0.2)	2 (0.0)	1.6 (0.4, to 5.7); 0.50	36 (–131 to 82)
IFV-B	0 (0.0)	2 (0.0)	0.0 (0.0 to 0.0); <0.001	0 (0 to 0)
PIV-1	2 (0.2)	1 (0.0)	2.5 (0.8 to 7.8); 0.13	59 (–28 to 87)
PIV-2	1 (0.1)	0 (0.0)	n/c	n/c
PIV-3	10 (1.0)	11 (0.3)	2.0 (1.2 to 3.3); 0.006	50 (18 to 70)
RSV-A	15 (1.6)	14 (0.3)	3.0 (2.1 to 4.5); <0.001	67 (51 to 78)
RSV-B	8 (0.8)	3 (0.1)	3.2 (1.8 to 5.5); <0.001	68 (45 to 82)
HCoV-OC43	9 (0.9)	8 (0.2)	2.3 (1.4 to 3.8); 0.001	56 (29 to 73)
HCoV-NL63	7 (0.7)	8 (0.2)	1.7 (0.9 to 3.3); 0.10	42 (-10 to 70)
HCoV-229E	1 (0.1)	4 (0.1)	1.2 (0.2 to 7.7); 0.83	19 (–407 to 87)
HCoV-HKU1	2 (0.2)	10 (0.2)	0.8 (0.2 to 2.9); 0.71	-28 (-376 to 66)
HMPV	6 (0.6)	4 (0.1)	3.2 (1.7 to 6.1); <0.001	69 (43 to 83)
AdV	5 (0.5)	16 (0.4)	1.0 (0.4 to 2.5); 0.96	-2 (-156 to 59)
WU-PyV	5 (0.5)	15 (0.3)	1.4 (0.6 to 3.5); 0.48	28 (-80 to 71)
KI-PyV	8 (0.8)	34 (0.8)	1.2 (0.7 to 2.2); 0.55	17 (-53 to 56)
HBoV-1	10 (1.0)	31 (0.7)	1.3 (0.8 to 2.2); 0.33	24 (-30 to 55)
Any virus	217 (22.6)	491 (11.4)	1.4 (1.2 to 1.5); <0.001	27 (18 to 35)
New codetections§				
≥2 viruses	58 (6.0)	62 (1.4)	2.2 (1.7 to 3.0); <0.001	55 (41 to 66)

Statistically significant risk ratios and attributable fractions have been highlighted in bold.

*Adjusted for repeated measures within children using sandwich estimators.

 $\ensuremath{^+\text{CI}}$ for attributable fractions were calculated from CI $\,$ for corresponding risk ratios.

+Individual HRV species do not add up to 'all HRV' detections because of a combination of more than one HRV species detected within the same LRTI episode, or single new virus detection episodes where HRV-positive specimens could not be sequenced.

§Includes 110 (51 symptomatic) new codetections and another 10 codetections with 2 and 3 viruses codetected, respectively where at least one codetected virus was a new detection.

AdV, adenovirus; HBoV-1, human bocavirus-1; HCoV, human coronavirus; HMPV, human metapneumovirus; HRV, human rhinovirus; IFV, influenza virus; KI-PyV, KI polyomavirus; LRTI, lower respiratory tract infection; n/c, not calculable as there were no cases in the asymptomatic group;, PIV, parainfluenza virus; RSV, respiratory syncytial virus; WU-PyV, WU polyomavirus.

2646/8100 (32.7%) swabs were positive for respiratory viruses (table 2), of which 1520 were new VDEs.

The incidence of new VDEs increased in the first 9 months of life, with symptomatic episodes dominating after 6 months of age (figure 2). There was an overall incidence rate of 978 new VDEs per 100 child-years (95% CI 930 to 1029, table 2). More than one virus was detected in 275/8100 (3.4%) specimens. HRV was the most commonly detected virus (2058/2646 detections, 77.8% of all positive detections, mean rate of 1002 new VDEs per 100 child-years, 95% CI 942 to 1066), followed by HBoV-1 (60 new VDEs per 100 child-years, 95% CI 49 to 73) and KI-PyV (56 new VDEs per 100 child-years, 95% CI 45 to 69) (table 2).

Virus-positive ARIs and asymptomatic episodes

When considering virus-positive ARIs and asymptomatic episodes, we retained 346 virus-positive swabs with ERV-3 Ct values >38, but removed a further 244 swabs with missing

symptom data, to give a final dataset of 8202 swabs from 151 participants (3 participants submitted swabs of high or no Ct values or swabs with no corresponding symptom data before withdrawing early in the study) (tables 3-5).

Of 8202 swabs, 3894 (47.5%) were taken during symptomatic periods and 4308 (52.5%) from asymptomatic periods. Of all swabs taken any time during symptomatic periods, 1903/3894 (48.9%) were positive for a virus, and 1120/3894 (28.8%) were new VDEs. Overall, 984/4308 (22.8%) swabs taken during asymptomatic periods were positive for a virus, of which 562 (57.1%) were new VDEs and 491/562 (87.4%) were new single virus detections (tables 3 and 4).

Table 3 shows that for new VDEs involving only single viruses, HRV-A, HRV-C, IFV-A, PIV-3, RSV-A, RSV-B, HCoV-OC43, HCoV-NL63, HMPV, and AdV had statistically significant, positive AFE values, indicating they were significantly more often associated with ARI episodes than asymptomatic periods. In

Table 5 Number of children, c	child-months, vir	us-associated symp.	otomatic and as	ymptomatic episodes and a	ssociation between partic	ipant character	istics and acute respiratory	infections in 151 children
			Symptomatic r OR (95% Cl); P	iew VDE value		Asymptomati OR (95% CI); F	c new VDE Value	
Risk factor	Children (N)	Child-months (N)	Ľ	Unadjusted	Adjusted*	۲	Unadjusted	Adjusted
Age (months)								
0-<3	151	423.3	60	Referent rate	Referent rate	58	Referent rate	Referent rate
3-<6	142	398.0	92	1.6 (1.2 to2.3); 0.003	1.7 (1.2 to 2.4); 0.003	06	1.7 (1.2, to 2.3); 0.003	1.5 (1.0 to 2.2); 0.03
6-<12	135	715.4	354	3.5 (2.6 to 4.6); <0.001	3.6 (2.4 to 5.4); <0.001	146	1.5 (1.1 to 2.0); 0.01	1.2 (0.8 to 1.8); 0.46
12-<24	117	1250.6	627	3.5 (2.7 to 4.6); <0.001	3.1 (2.0 to 4.7); <0.001	270	1.6 (1.2 to 2.1); 0.001	1.2 (0.7 to 1.8); 0.54
Sex								
Male	74	1255.1	481	Referent rate	Referent rate	262	Referent rate	Referent rate
Female	77	1532.2	652	1.1 (1.0 to 1.4), 0.17	1.1 (0.9 to 1.3); 0.33	302	1.0 (0.8 to 1.2); 0.79	0.9 (0.7 to 1.2); 0.53
Type of delivery								
Vaginal	103	1876.3	802	Referent rate	Referent rate	387	Referent rate	Referent rate
Caesarean	48	910.9	331	0.8 (0.7 to 1.0); 0.12	0.8 (0.7 to 1.0); 0.09	177	0.9 (0.7 to 1.2); 0.64	1.0 (0.8 to 1.3); 0.94
Gestational age at birth								
Term (39w0d–41w6d)	117	2219.1	937	Referent rate	Referent rate	446	Referent rate	Referent rate
Late preterm/early term (36w0d-38w6d)	34	568.1	196	0.8 (0.7 to 1.1); 0.17	0.9 (0.7 to 1.2); 0.48	118	1.1 (0.8 to 1.5); 0.58	1.1 (0.8 to 1.5); 0.40
Season								
Summer	140	678.8	196	Referent rate	Referent rate	138	Referent rate	Referent rate
Autumn	138	702.0	290	1.4 (1.2 to 1.7); <0.001	1.4 (1.2 to 1.7); <0.001	135	0.9 (0.7 to 1.2); 0.64	1.0 (0.8 to 1.2); 0.75
Winter	138	706.6	350	1.7 (1.4 to 2.1); <0.001	1.6 (1.3 to 1.9); <0.001	142	1.0 (0.8 to 1.3); 0.93	0.9 (0.7 to 1.2); 0.67
Spring	142	6.99.9	297	1.5 (1.2 to 1.8); <0.001	1.4 (1.2 to 1.7); <0.001	149	1.1 (0.8 to 1.3); 0.68	1.1 (0.8 to 1.4); 0.56
Family history								
None present for asthma/eczema	73	1289.6	533	Referent rate	Referent rate	286	Referent rate	Referent rate
Either parent asthma/eczema	75	1447.3	582	1.0 (0.8 to 1.2); 0.93	1.0 (0.8 to 1.2); 0.85	264	0.8 (0.7 to 1.1); 0.16	0.9 (0.7 to 1.1); 0.22
First-degree relative asthma/ eczema†	78	1497.7	600	1.0 (0.8 to 1.2); 0.80	1.0 (0.8 to 1.2); 0.71	278	0.9 (0.7 to 1.1); 0.22	0.9 (0.7 to 1.1); 0.36
Tobacco smoke exposure								
No exposure	129	2383.4	973	Referent rate	Referent rate	472	Referent rate	Referent rate
Either parent smokes	17	307.4	127	1.0 (0.7 to 1.4); 0.86	0.9 (0.6 to 1.1); 0.29	75	1.2 (0.8 to 1.7); 0.42	1.1 (0.8 to 1.7); 0.51
Household size								
No other children in household	66	1864.9	723	Referent rate	Referent rate	350	Referent rate	Referent rate
More than one child in household	52	922.4	410	1.1 (0.9 to 1.4); 0.28	1.1 (0.9 to 1.4); 0.17	214	1.3 (1.0 to 1.7); 0.05	1.4 (1.0 to 1.7); 0.04
Maternal education status								
High school	14	246.7	113	1.1 (0.8 to 1.6); 0.56	1.0 (0.8 to 1.4); 0.81	62	1.4 (0.9 to 2.1); 0.11	1.4 (0.9 to 2.1); 0.17
								Continued

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Table 5 Continued								
			Symptomatic OR (95% Cl); P	new VDE V value		Asymptomatic OR (95% CI); P	: new VDE value	
Risk factor	Children (N)	Child-months (N)	=	Unadjusted	Adjusted*	Ē	Unadjusted	Adjusted
Diploma/certificate	36	650.2	245	0.9 (0.7 to 1.1); 0.37	0.9 (0.7 to 1.1); 0.25	149	1.2 (0.9 to 1.6); 0.29	1.2 (0.9 to 1.6); 0.15
University/higher university degree	95	1828.3	750	Referent rate	Referent rate	346	Referent rate	Referent rate
Mode of feeding								
Formula	142	2281.5	1039	Referent rate	Referent rate	484	Referent rate	Referent rate
Breastmilk	145	505.5	94	0.4 (0.3 to 0.5); <0.001	1.2 (0.9 to 1.7); 0.28	80	0.7 (0.6 to 0.9); 0.01	0.9 (0.6 to 1.3); 0.47
Childcare attendance								
No childcare	150	1423.6	423	Referent rate	Referent rate	242	Referent rate	Referent rate
Informal childcare only	47	389.5	117	1.2 (0.9 to 1.5); 0.13	0.8 (0.6 to 1.1); 0.16	93	1.3 (1.0 to 1.7); 0.10	1.3 (0.9 to 1.7); 0.16
Formal and/or informal childcare	90	973.8	593	2.2 (1.9 to 2.5); <0.001	1.5 (1.3 to 1.9); <0.001	229	1.4 (1.1 to 1.7); 0.002	1.4 (1.0 to 1.8); 0.03
Statistically significant risk ratios and an *Adjusted for age, sex, type of delivery, †Mother, father or sibling. OR, odds ratio, VDE, virus detection epis	ttributable fractic gestational age, ode.	ons have been highlighti season, family history, t	ed in bold. tobacco smoke exl	posure, household size, maternal (education status, mode of feedi	ng and childcare a	ittendance.	

contrast, HCoV-229E, HCoV-HKU1, WU-PyV, KI-PyV, and HBoV-1 were frequently detected in asymptomatic periods and had low point estimates for AFE, indicating they were less likely to have caused the ARI.

Of the total 1641 ARIs, 111 (6.8%) did not have swabs submitted during the ARI episode, including swabs taken 7 days before or 7 days after the last day of the illness. Of the remaining 1530 ARIs, 1154 (75.4%) ARIs (935 (81%) URTI, 219 (19.0%) LRTI) had ≥ 1 swabs with a virus detected during this period.

RSV and HMPV were more strongly associated with LRTI than asymptomatic detections (table 4), with higher risk ratios than other viruses. While significant, the association between LRTI and HRV-C, HCoV-OC43 and PIV-3 detection was weaker. Interestingly, codetected specimens had among the strongest AFE values for both ARIs and LRTIs (tables 3 and 4), with HRV the most frequently detected virus group, usually in combination with DNA viruses (see online supplementary E-table 3).

To investigate the sensitivity of results from including lesser quality swabs, we reanalysed the data by excluding 346 virus-positive swabs of lesser quality, and found minimal change to the AFEs calculated in tables 3 and 4 (see online supplementary E-table 2).

Viruses and association with risk factors

Independent risk factors associated with symptomatic rather than asymptomatic VDEs, included increasing age, particularly after 6 months, and virus infections during non-summer seasons (table 5). The regression model revealed childcare attendance was also independently associated with higher rates of VDEs, both symptomatic and asymptomatic, but no other risk factors were found to be associated with asymptomatic VDEs.

DISCUSSION

Healthy children in the ORChID community-based birth cohort experienced an incidence rate of 978 new VDEs per 100 childyears during the first 2 years of life, with HRV playing a dominant role. Overall, respiratory viruses were detected by PCR in regular weekly swabs at least once during 75% of ARIs, while in asymptomatic periods 23% of weekly nasal swabs were positive by PCR for respiratory viruses, accounting for 33% of all new VDEs. We demonstrated evidence of attribution for ARIs by RNA viruses, which included HRV (HRV-A, HRV-C), influenza (IFV-A), parainfluenza (PIV-1, PIV-3), RSV (RSV-A, RSV-B), HCoV (HCoV-OC43, HCoV-NL63) and HMPV; and AdV as the single DNA virus representative. All these viruses were detected significantly more often in children with an ARI than during asymptomatic periods and had statistically significant positive AFEs. The largest association was observed for RSV and HMPV, both of which were more strongly associated with LRTIs than other respiratory viruses. Increasing age ≥ 6 months, non-summer months and childcare attendance were independent risk factors for symptomatic VDEs. Age as a host risk factor for symptomatic infection is also noted in other studies,²³ and is presumably due to protection afforded by maternal transplacental and breastmilk antibodies in the very young.²⁴ Environmental risk factors of season and childcare attendance for ARIs may be related to the intensity and opportunity for exposure in these settings experienced by young children.^{10 25}

The virus-specific attribution of ARIs was quantified by identifying VDEs during asymptomatic periods and calculating the virus-specific AFE, an infrequently reported metric for helping to determine the relative causal roles of different viruses at a population level. A limitation of the AFE approach is that it cannot inform on the aetiology at an individual level: in individual cases it does not differentiate between those with a true positive test identifying the virus responsible for the ARI and those with a false positive result from asymptomatic virus shedding. Nevertheless, knowing the AFE for respiratory viruses can help inform clinicians on the likely clinical significance of detecting one or more viruses in nasal swab specimens taken from infants or young children during an ARI episode, by assigning the group aetiological fraction for viruses detected in that child. Importantly, AFE estimates can be applied to studies of ARIs in different settings to estimate the true burden of disease in the community, thus informing which viruses should be prioritised in future public health policies and interventions.

Most previous studies have overestimated AFE values by using ORs, rather than RRs, in calculations.^{5 6 8 9} ORs overestimate the effect size when the event is not rare, as occurs with ARIs in children. These studies are principally cross-sectional, hospital-based and often have poorly matched controls. Consequently, our estimates of AFE being lower than reported elsewhere is not surprising, and, we believe, are likely to provide more accurate estimates of the ARI and LRTI burden attributable to these viruses in the broader paediatric population.

The proportion of asymptomatic infections in our study highlights the risks of extrapolating hospital-based data from sick children to the community setting. Previously, the significance of detecting viruses by PCR was tempered by concerns that this may be from an incubating or unrelated subclinical infection or represent continued virus nucleic acid shedding from a recent ARI that has resolved. The intensive longitudinal specimen and symptom sampling from our cohort, as well as the definitions of VDEs and symptomatic VDEs used, helped identify nascent and subclinical infections. In this context, our observations emphasise the frequency of subclinical infections by respiratory viruses in young children, who when otherwise well, may act as important 'silent' reservoirs of infection. Similar observations and conclusions were reported recently in a household study involving both child and adult contacts of symptomatic cases of laboratory-confirmed influenza.²⁶ Nevertheless, in addition to influenza, established viruses, such as HRV, PIV-3, RSV, and HMPV are also important community respiratory pathogens, especially the latter two viruses, and potential candidates for vaccine and other therapeutic interventions.²⁷ This contrasts with the novel DNA viruses, HPyVs and HBoV-1, which have low AFEs calculated for both ARIs and LRTIs, despite their higher detection rates.^{28 29} However, as suggested by this cohort, synergy may exist between multiple viruses causing ARIs. Unexpectedly, this involved mostly HRV and DNA viruses, and this finding warrants further study.

Strengths and limitations

The 78% of expected child-days and 68% of expected swab returns was a very good retention rate given the intensive and prolonged nature of the study for participating families. Analysis of incidence rates of ARI in participants that withdrew from the study at different periods showed no clear association between time of dropout and ARI incidence. Regular parent-collected nasal swabs avoids the need for home visits by research staff or clinic attendance during an ARI, minimising possible biased estimates of ARI events and specimen availability from losing families failing to seek healthcare. Moreover, regular weekly nasal swab collections increase the likelihood of virus detections during an ARI episode rather than relying simply on results from a single specimen. Our previous work found that with sensitive PCR techniques, parent-collected nose or nose-throat swabs sent to the laboratory by standard mail had comparable sensitivity to nasopharyngeal swabs obtained by healthcare workers.¹⁸ ^{19 30} Longitudinal data collection also allows assessment of asymptomatic status by considering past and future illness history, as some viruses are detectable days before and after ARI symptoms develop,³¹ and are therefore less likely to yield false-positive findings.

There are also some important limitations to consider. To reduce the chances of false-negative test results,²¹ we excluded from analysis 2679 (23.9%) swabs where the internal control for human DNA, ERV-3, was either undetected or present at very low levels. We excluded both virus-positive and virus-negative swabs to avoid overestimating our incidence and prevalence rates. In addition, some viruses were detected rarely, in particular influenza, a finding reflected in other community studies,¹⁷ but nevertheless limiting our ability to provide precise AFE estimates for these agents. We also excluded codetections in individual virus assessments as multiple aetiological agents make individual contributions of each agent difficult to ascertain, but analysed them separately to look at the association patterns of individual viruses. Symptom information, other than doctor-diagnosed otitis media and pneumonia, captured by daily diaries was not validated. To maximise accuracy, parents were trained to recognise symptoms before commencing the study. While healthcare worker-validated symptoms would be ideal, a study of this scale is logistically challenging without parent participation. This study design has been used previously¹⁸ and with others have shown that parents can be trained to recognise symptoms of interest,¹⁸ often as reliably as healthcare professionals.^{32 33} Our rates of ARI, including those associated with wheeze, are comparable with other community studies.^{34 35}

Finally, as is common for these types of studies, families in our cohort were from more advantaged backgrounds and ARI episodes were predominantly of a mild-to-moderate nature.²² While many children in our study were first-born and from advantaged families, factors thought to decrease the rate of ARIs, secular changes in Australia have led to an increased proportion of children from these families attending childcare at an earlier age than other sectors of the population. A slightly higher proportion (80%-85%) of our cohort attended some form of care by the age of 15-24 months, compared with 74% of children in the Australian population, with childcare attendance an independent risk factor for ARIs.²⁵ Moreover, the incidence of ARIs in the ORChID cohort²² is comparable to other reports in this age group^{34 36 37} and RSV and HRV-associated ARIs approximate other community-based studies conducted in more temperate climates of Australia (Perth) and Europe.³⁸⁻⁴⁰ Our findings for this cohort remain valid and provide important estimates on community respiratory virus exposures and ARIs in Australian children in a subtropical, urban setting.

CONCLUSIONS

Respiratory viruses, particularly HRV, were detected with remarkable frequency in healthy Australian children in the first 2 years of life, with at least one-third of new VDEs unaccompanied by symptoms. With molecular methods used increasingly to detect respiratory viruses in young children, our study provides insights into supporting attribution for virus-specific ARIs in this age group

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managed within the community. The established RNA respiratory viruses and AdV were more strongly associated with symptomatic than asymptomatic infections, while no such associations were found for the newly described DNA viruses, HPyVs and HBoV-1. These data emphasise the high community levels of HRV circulating among children. Importantly, they also provide evidence to suggest that the current clinical trials of candidate RSV and HMPV vaccines seeking to protect infants against severe disease and hospitalisation might also deliver substantial health benefits for all young children within the community.²⁷

Correction notice This article has been corrected since it was published Online First. Figure 2 has been updated to include the missing figure legend.

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Contributors MS had full access to all of the data in the study and had the final responsibility for the decision to submit for publication.Study concept and design: KG, SBL, RSW, TPS, MDN, DW, DMW. Analysis and interpretation of data: MS, SBL, RSW, KG, TPS, AA, LM, SB. Drafting of the manuscript: MS, SBL, RSW, KG. Critical revision of the manuscript for important intellectual content: MS, RSW, TPS, DMW, AA, LM, SB, DW, MDN, SBL, KG. Statistical analysis: MS, RSW. Administrative and technical support: MS, RSW, SBL. Study supervision: SBL, RSW, KG.

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Competing interests After completion of data collection for this study, MDN became a full-time employee of GlaxoSmithKline, GlaxoSmithKline Vaccines Value Health Science, 150 Beach Road, Gateway West #7, Singapore 189720, Singapore. Other authors have no competing interest to declare.

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REFERENCES

- Brealey JC, Sly PD, Young PR, et al. Viral bacterial co-infection of the respiratory tract during early childhood. FEMS Microbiol Lett 2015;362.
- 2 Monto AS. Epidemiology of viral respiratory infections. Am J Med 2002;112:4–12.
- 3 Monto AS, Cavallaro JJ. The Tecumseh study of respiratory illness. II. Patterns of occurrence of infection with respiratory pathogens, 1965-1969. *Am J Epidemiol* 1971;94:280–9.
- 4 Jartti T, Söderlund-Venermo M, Hedman K, *et al*. New molecular virus detection methods and their clinical value in lower respiratory tract infections in children. *Paediatr Respir Rev* 2013;14:38–45.
- 5 Howard LM, Johnson M, Williams JV, et al. Respiratory viral detections during symptomatic and asymptomatic periods in young Andean children. *Pediatr Infect Dis J* 2015;34:1074–80.
- 6 Self WH, Williams DJ, Zhu Y, et al. Respiratory viral detection in children and adults: comparing asymptomatic controls and patients with community-acquired pneumonia. J Infect Dis 2016;213:584–91.
- 7 Pretorius MA, Tempia S, Walaza S, et al. The role of influenza, RSV and other common respiratory viruses in severe acute respiratory infections and influenza-like illness in a population with a high HIV sero-prevalence, South Africa 2012-2015. J Clin Virol 2016;75:21–6.
- 8 Singleton RJ, Bulkow LR, Miernyk K, et al. Viral respiratory infections in hospitalized and community control children in Alaska. J Med Virol 2010;82:1282–90.

- 9 Shi T, McLean K, Campbell H, et al. Aetiological role of common respiratory viruses in acute lower respiratory infections in children under five years: a systematic review and meta-analysis. J Glob Health 2015;5:010408–18.
- 10 Mack I, Kieninger E, Cangiano G, et al. Rhinovirus Infections and associated respiratory morbidity in Infants: a prospective cohort study. *Pediatr Infect Dis J* 2016;35:1069–74.
- 11 Chonmaitree T, Alvarez-Fernandez P, Jennings K, et al. Symptomatic and asymptomatic respiratory viral infections in the first year of life: association with acute otitis media development. *Clin Infect Dis* 2015;60:1–9.
- 12 Munywoki PK, Koech DC, Agoti CN, et al. Frequent asymptomatic respiratory syncytial virus infections during an epidemic in a rural Kenyan household cohort. J Infect Dis 2015;212:1711–8.
- 13 Rhedin S, Lindstrand A, Hjelmgren A, *et al*. Respiratory viruses associated with community-acquired pneumonia in children: matched case-control study. *Thorax* 2015;70:847–53.
- 14 Lambert SB, Ware RS, Cook AL, et al. Observational Research in Childhood Infectious Diseases (ORChID): a dynamic birth cohort study. BMJ Open 2012;2:e002134–40.
- 15 Australian Bureau of Statistics Australian social trends 4102.0. Child care. 2010. http://www.ausstats.abs.gov.au/ausstats/subscriber.nsf/LookupAttach/4102. 0Publication30.06.106/\$File/41020_ChildCare.pdf (accessed on 12 Feb 2017).
- 16 Belshe RB, Mendelman PM, Treanor J, et al. The efficacy of live attenuated, coldadapted, trivalent, intranasal influenzavirus vaccine in children. N Engl J Med 1998;338:1405–12.
- 17 Lambert SB, Allen KM, Druce JD, et al. Community epidemiology of human metapneumovirus, human coronavirus NL63, and other respiratory viruses in healthy preschool-aged children using parent-collected specimens. *Pediatrics* 2007;120:e92 9–e937.
- 18 Lambert SB, Allen KM, Nolan TM. Parent-collected respiratory specimens--a novel method for respiratory virus and vaccine efficacy research. *Vaccine* 2008;26:1826–31.
- 19 O'Grady KA, Torzillo PJ, Rockett RJ, et al. Successful application of a simple specimen transport method for the conduct of respiratory virus surveillance in remote Indigenous communities in Australia. *Trop Med Int Health* 2011;16:766–72.
- 20 Yuan CC, Miley W, Waters D. A quantification of human cells using an ERV-3 real time PCR assay. J Virol Methods 2001;91:109–17.
- 21 Alsaleh ÁN, Whiley DM, Bialasiewicz S, *et al*. Nasal swab samples and real-time polymerase chain reaction assays in community-based, longitudinal studies of respiratory viruses: the importance of sample integrity and quality control. *BMC Infect Dis* 2014;14:15–24.
- 22 Sarna M, Ware RS, Sloots TP, et al. The burden of community-managed acute respiratory infections in the first 2-years of life. *Pediatr Pulmonol* 2016;51:1336–46.
- 23 Byington CL, Ampofo K, Stockmann C, et al. Community surveillance of respiratory viruses among families in the Utah Better Identification of Germs-Longitudinal Viral Epidemiology (BIG-LoVE) study. *Clin Infect Dis* 2015;61:1217–24.
- 24 Duijts L, Jaddoe VW, Hofman A, et al. Prolonged and exclusive breastfeeding reduces the risk of infectious diseases in infancy. *Pediatrics* 2010;126:e18–e25.
- 25 Kamper-Jørgensen M, Benn CS, Simonsen J, *et al.* Clustering of acute respiratory infection hospitalizations in childcare facilities. *Acta Paediatr* 2010;99:877–82.
- 26 Ip DK, Lau LL, Leung NH, et al. Viral shedding and transmission potential of asymptomatic and paucisymptomatic influenza virus infections in the community. *Clin Infect Dis* 2017;64:736–42.
- 27 Tang JW, Lam TT, Zaraket H, et al. Global epidemiology of non-influenza RNA respiratory viruses:data gaps and a growing need for surveillance. Lancet Infect Dis 2017;17:320–326.
- 28 Rockett RJ, Bialasiewicz S, Mhango L, et al. Acquisition of human polyomaviruses in the first 18 months of life. Emerg Infect Dis 2015;21:365–7.
- 29 Broccolo F, Falcone V, Esposito S, et al. Human bocaviruses: possible etiologic role in respiratory infection. J Clin Virol 2015;72:75–81.
- 30 Lambert SB, Whiley DM, O'Neill NT, et al. Comparing nose-throat swabs and nasopharyngeal aspirates collected from children with symptoms for respiratory virus identification using real-time polymerase chain reaction. *Pediatrics* 2008;122:e615–20.
- 31 Lessler J, Reich NG, Brookmeyer R, et al. Incubation periods of acute respiratory viral infections: a systematic review. Lancet Infect Dis 2009;9:291–300.
- 32 Chang AB, Gaffney JT, Eastburn MM, et al. Cough quality in children: a comparison of subjective vs. bronchoscopic findings. *Respir Res* 2005;6:3–13.
- 33 Bekhof J, Reimink R, Bartels IM, et al. Large observer variation of clinical assessment of dyspnoeic wheezing children. Arch Dis Child 2015;100:649–53.
- 34 von Linstow ML, Holst KK, Larsen K, et al. Acute respiratory symptoms and general illness during the first year of life: a population-based birth cohort study. *Pediatr Pulmonol* 2008;43:584–93.
- 35 van der Gugten AC, Uiterwaal CS, van Putte-Katier N, et al. Reduced neonatal lung function and wheezing illnesses during the first 5 years of life. Eur Respir J 2013;42:107–15.
- 36 Kusel MM, de Klerk N, Holt PG, et al. Occurrence and management of acute respiratory illnesses in early childhood. J Paediatr Child Health 2007;43:139–46.
- 37 Hartert TV, Carroll K, Gebretsadik T, et al. The Tennessee children's respiratory initiative: objectives, design and recruitment results of a prospective cohort study

investigating infant viral respiratory illness and the development of asthma and allergic diseases. *Respirology* 2010;15:691–9.

- 38 Kusel MM, de Klerk NH, Holt PG, et al. Role of respiratory viruses in acute upper and lower respiratory tract illness in the first year of life: a birth cohort study. *Pediatr Infect Dis J* 2006;25:680–6.
- 39 van der Zalm MM, Uiterwaal CS, Wilbrink B, et al. Respiratory pathogens in respiratory tract illnesses during the first year of life: a birth cohort study. Pediatr Infect Dis J 2009;28:472–6.
- 40 Legg JP, Warner JA, Johnston SL, *et al*. Frequency of detection of picornaviruses and seven other respiratory pathogens in infants. *Pediatr Infect Dis J* 2005;24:611–6.