## **Serveur Académique Lausannois SERVAL serval.unil.ch**

# **Author Manuscript Faculty of Biology and Medicine Publication**

**This paper has been peer-reviewed but does not include the final publisher proof-corrections or journal pagination.**

Published in final edited form as:

**Title:** Innate receptors for adaptive immunity. **Authors**: Michallet MC, Rota G, Maslowski K, Guarda G **Journal**: Current opinion in microbiology **Year**: 2013 Jun **Volume**: 16 **Issue**: 3 **Pages**: 296-302 **DOI:** [10.1016/j.mib.2013.04.003](http://dx.doi.org/10.1016/j.mib.2013.04.003)



Université de Lausanne Faculté de biologie et de médecine

#### INNATE RECEPTORS FOR ADAPTIVE IMMUNITY

Marie-Cécile Michallet<sup>1</sup>, Giorgia Rota<sup>2</sup>, Kendle Maslowski<sup>2</sup>, and Greta Guarda<sup>2,3</sup>

<sup>1</sup> Centre International de Recherche en Infectiologie (CIRI) INSERM U1111 - CNRS UMR5308 Université de Lyon 1, ENS de Lyon 21 Avenue Tony Garnier 69365 LYON cedex 07 - France

<sup>2</sup> Department of Biochemistry Chemin des Boveresses 155 1066 Epalinges - Switzerland

<sup>3</sup> to whom correspondence should be addressed: Greta Guarda Department of Biochemistry Chemin des Boveresses 155 1066 Epalinges - Switzerland Phone +41-21-692-5708 Fax +41-21-692-5705 E-mail: Greta.Guarda@unil.ch

#### **Summary**

Pattern recognition receptors (PRRs) are commonly known as sensor proteins crucial for the early detection of microbial or host-derived stress signals by innate immune cells. Interestingly, some PRRs are also expressed and functional in cells of the adaptive immune system. These receptors provide lymphocytes with innate sensing abilities; for example B cells express Toll-like receptors, which are important for the humoral response. Strikingly, certain other NOD-like receptors are not only highly expressed in adaptive immune cells, but also exert functions related specifically to adaptive immune system pathways, such as regulating antigen presentation. In this review, we will focus particularly on the current understanding of PRR functions intrinsic to B and T lymphocytes; a developing aspect of PRR biology.

#### **Highlights**

- PRRs are expressed by cells of the innate, but also of the adaptive immune system
- Certain PRRs endow B and T cells with innate sensing abilities
- Other PRRs evolved to fulfill functions related to the adaptive immune system

#### **Keywords**

T cell; B cell; TLR; NLR; RLR

#### **Introduction**

Pattern recognition receptors (PRRs) are defined as key sensors involved in detecting pathogens or danger signals and initiating inflammatory processes. The engagement of PRRs in innate immune cells, such as dendritic cells, is also crucial for indirect instruction of adaptive immune responses. While these aspects of PRR signaling are well understood, less is known about PRRs that are expressed by adaptive immune cells. Here, we will discuss evidence supporting a function of Tolllike, retinoic acid-inducible gene I (RIG-I)-like, or NOD-like receptors (TLRs, RLRs, and NLRs, respectively) intrinsic to B lymphocytes, conventional and regulatory T cells.

#### **TOLL-LIKE RECEPTORS IN B AND T CELLS**

The TLR family was the first identified among PRRs and is therefore the most characterized. TLRs are transmembrane glycoproteins that bind to a wide range of pathogen- and danger-associated molecular patterns (PAMPs and DAMPs). Thirteen mammalian TLRs have been identified; ten functional receptors in humans and twelve in mice [1]. While TLR10 and TLR11-13 are exclusively expressed in humans and mice, respectively, TLR1-9 are shared by both species [2]. These receptors are typically expressed in innate immune cells, but analyses at the mRNA level in human and mouse have demonstrated TLR expression in all peripheral blood leukocytes including B and T cells [3-7]. Such studies have provided a rationale for examining a cell-intrinsic function of TLRs in adaptive immune cells.

#### *TLRs in B cells*

The current understanding as to how TLRs modulate B lymphocyte activation, antigen presentation, proliferation, class switch recombination, and antibody production is comprehensively reviewed elsewhere [8,9]. Therefore, we will focus on a few selected studies investigating these aspects.

Several reports have described the expression of TLRs in different mouse and human B cell subsets and their regulation by cytokines as well as signaling from the B cell receptor [6-8,10,11]. TLR1, TLR2, TLR4, TLR6, TLR7 and TLR9 are expressed in most murine B cell subsets, including naïve B cells, but at varying levels [10], suggesting a subset-specific sensitivity to diverse TLR agonists. Similar data were obtained in human B cells, although constitutive TLR expression in humans is

most prominent among memory B cells, which has been suggested to play an important role in the maintenance of serological memory [7,12,13].

The first *in vivo* evidence for a B cell-autonomous role of TLRs in the regulation of humoral responses came from Pasare and Medzhitov [14]. Performing transfer experiments of B cells deficient for the TLR signaling adaptor *Myd88* or for *Tlr4*, they showed that TLR signaling in B cells is mandatory for the generation of optimal T cell-dependent antibody responses. Notably, the role of TLRs in B cells has been corroborated in B cell-specific *Myd88-*deficient mice, which showed impaired humoral response upon immunization with virus-like particles delivering TLR9 ligands [15]. However, as MyD88 is an adaptor shared also by the IL-1 receptor family, the use of conditional TLR-deficient animals would further strengthen these results.

The implications of these studies are highly relevant not only in the context of antiviral antibody-mediated responses, but also for vaccine development and understanding autoimmunity more broadly [8,16].

#### *TLRs in T cells*

Murine and human T cells express several functional TLRs, which are regulated depending on T cell activation status [6,11]. The first evidence of a functional effect of TLR ligands on T cell physiology was on clonal expansion. Bendigs and collaborators demonstrated that CpG-DNA treatment enhanced antigen-mediated proliferation of murine T cells, whereas no effect was observed on naive cells [17]. Furthermore, in murine  $CD4^+$  T cells, TLR9 engagement was shown to be important for cell survival through the activation of mitogen-activated protein kinases (MAPKs) and NF-κB [18,19].

Several studies demonstrated a costimulatory effect of TLR2 ligands on antigenmediated proliferation of  $CD4^+$  and  $CD8^+$  T cells of human and murine origin [20-23]. TLR2 is well expressed on mouse cytotoxic  $CDS^+$  T cells and human memory  $CD4^+$ T cells [20,22]. Cooperation of TLR2 and TLR5 engagement was also described to enhance activated  $CD8<sup>+</sup> T$  cell expansion [24]. Furthermore, homeostatic proliferation of memory  $CD4^+$  T cells was increased by ligands of TLR2, TLR5 or TLR7/8, suggesting a role for TLR engagement in long-term maintenance of memory T cells [20,25].

Concerning effector functions and cytokine production, TLR2 signaling was shown to be essential for promoting the production of interleukin (IL)-17 and interferon (IFN)-γ by effector CD4<sup>+</sup> T cells [26,27], but also chemokine (C-C motif) ligand 3 (CCL3), and CCL4 [28]. TLR2 or TLR3 engagement on T cell receptor (TCR)-activated  $CDS^+$  T cells also enhanced IFN- $\gamma$  secretion [21,22,29].

Finally, a dual effect of TLR ligands was described on the regulatory T cell (Treg) compartment. TLR8 ligands were shown to reverse the suppressive function of human T regulatory cells [30], while it was reported that LPS might induce proliferation and enhance suppressive activity of murine regulatory cells [31].

Given the high expression of many TLRs among APCs, and the sensitivity of T cells to APC-derived activating signals, it is important to remember that a small number of contaminating APCs could confound the analysis. Nonetheless, an effect of TLRs on T cell physiology at the intrinsic level is nowadays supported by several independently conducted and well-controlled experiments, clearly demonstrating that TLR agonists affect expansion, differentiation, or activity of effector/memory T cells as well as the regulatory T cell population. However, the impact of TLR ligands on other T-cell subsets, as for instance Th17 or Th22, still awaits further investigation *in vitro* and particularly *in vivo*. Such studies may open up potential therapeutic applications in vaccination or autoimmune diseases [32].

#### **RIG-I-LIKE RECEPTORS IN T LYMPHOCYTES**

The RLR family members RIG-I and melanoma differentiation-associated gene 5 (MDA5) act as cytoplasmic RNA sensors inducing type I IFN responses. The third family member, laboratory of genetics and physiology 2 (LGP2), is instead a positive regulator of their signaling (see Table 1) [33]. RLR signaling is mediated by the downstream adaptor mitochondrial antiviral-signaling protein (MAVS), which is required for IFN induction.

Both, RIG-I and MDA5, are highly expressed in T lymphocytes [6]. Interestingly, stimulation of Tregs with Encephalomyocarditis virus has been shown to decrease Treg inhibitory function in an MDA5-dependent manner, as demonstrated by the use of knockout cells [34]. However, no additional insights were provided on the mechanism underlying this phenomenon.

A recent paper showed the involvement of LGP2 in cytotoxic T cell survival upon West Nile virus infection, an outcome that was surprisingly independent of MAVS. [35]. This was achieved by down-modulating sensitivity to Fas-mediated apoptosis. Given that RLRs specifically detect RNAs, it is however still unclear whether LGP2 mediates T cell survival only upon infection by RNA viruses [35].

Despite the expression of RLRs in T cells, there is little evidence that these lymphocytes efficiently produce type I IFN in response to RNA. Accordingly, emerging data show that RLRs in T cells can fulfill functions unrelated to classical MAVS-mediated signaling; a novel aspect that deserves further investigation.

#### **NLRs IN ADAPTIVE IMMUNE CELLS**

NLRs are intracellular proteins involved in diverse immune processes [36-40]. In this review, we will focus on family members that are expressed by adaptive immune cells (Table 2).

#### *Inflammasome-forming NLRs*

Upon detection of stress signals, certain NLRs assemble into complexes called "inflammasomes" [37]. Inflammasomes trigger the cleavage of caspase-1, which proteolytically activates interleukin (IL)-1β and mediates an inflammatory cell death called "pyroptosis" [37].

Although inflammasomes have mainly been described in myeloid cells, caspase-1 and the adaptor protein ASC (apoptosis speck protein with CARD) are also expressed in lymphocytes [6,11]. This supports the possibility that these multiprotein platforms can also be formed in adaptive immune cells. Indeed, a very interesting study suggested formation of an inflammasome in T cells upon abortive HIV infection, with viral DNA being the trigger [41], though the sensor inducing inflammasome assembly has not been identified.

Indeed, certain inflammasome-forming NLRs are expressed in lymphoid cells. For instance, NLR family, pyrin domain containing 1a (NLRP1a) is expressed in common myeloid and lymphoid progenitors [6,36,37,42-45]. This NLR has recently been shown to induce pyroptosis upon stresses such as chemotherapy or infection, prolonging cytopenia in both myeloid and lymphoid compartments, therefore supporting the possibility that an inflammasome is formed in lymphoid precursor cells [42]. In addition, human NLRP1 is highly expressed in T and B cells, though its function in lymphocytes remains unexplored [6,43,46-48].

The prototypical inflammasome-forming NLR, NLRP3, is however barely detectable in T cells at the steady-state [6,43,44,49,50], although it could be upregulated upon activation, which is the case in B cells, particularly following Ctype lectin stimulation [51]. In B cells, Nlrp3 is involved in activation and immunoglobulin production downstream of C-type lectin stimulation. Though the molecular details of this phenomenon remain elusive, it was found to be independent of MyD88, suggesting that it was not mediated by IL-1 receptor signaling [51].

#### *NOD1 and NOD2*

NOD1 and NOD2 activate NF-κB and MAPK pathways upon sensing peptidoglycans [36,38]. Whilst the function of NOD1 and NOD2 has largely been explored in innate immune and mucosal epithelial cells, NOD1 and, to a lesser extent, NOD2 are expressed by cells of the adaptive immune system [6,43,44,47,50,52]. Stimulation of B and  $CD8<sup>+</sup>$  T cells with a NOD1 agonist improved antigen receptordriven proliferation, and the use of *Nod1-/-* T cells nicely demonstrated the specificity of this effect [44,47].

Similar to the LGP2 effect in T cells, NOD2 stimulation increased survival of Tregs by decreasing sensitivity to Fas-mediated apoptosis [50]. Furthermore, *Nod2-/* conventional T cells produced less IL-2, effector cytokines, and showed reduced nuclear accumulation of the NF-κB family member c-Rel in the context of *Toxoplasma gondii* infection [53]. However, these data were not substantiated in a later study [52], reminiscent of the debated T cell-autonomous role of receptorinteracting serine/threonine-protein kinase 2, the kinase acting downstream of NOD1 and NOD2 [44,54]. Therefore, future work is required to clarify these discrepancies.

#### *Signaling and transcription regulatory NLRs*

NLR family, CARD domain containing 3 (NLRC3) is a poorly studied NLR, predominantly expressed in T and NK lymphocytes [6,55]. An early study suggested a negative role in T cell activation because NLCR3 transcript abundance decreased upon TCR triggering and NLRC3 overexpression impaired TCR-induced NF-κB signaling [55]. However, a recent report demonstrated experimental artifacts can be generated using NLRC3 overexpression assays, suggesting caution should be taken when interpreting such overexpression studies [56]. Nonetheless, an increase in TLRdriven NF-κB activation was shown in macrophages derived from *Nlrc3*-deficient mice, though T lymphocytes were not investigated in this study [57].

One of the most exciting areas of PRR function in adaptive immunity is in transcriptional regulation. Whilst they belong to NLRs, CIITA and NLRC5 act as transcriptional regulators of major-histocompatibility complex class II (MHCII) and class I (MHCI), respectively [39,40,58]. In humans and in mouse, CIITA deficiency causes a lack of MHCII, leading to severe immunodeficiency [39]. CIITA expression is virtually restricted to antigen-presenting cells (APCs) and in humans also to recently activated  $CD4^+$  T cells [6,39]. MHCII expression is crucial for the homeostasis and the activity of helper T cells, and its expression specifically by B lymphocytes is essential for the maturation of the humoral response.

Under homeostatic conditions, NLRC5 is highly expressed in lymphocytes, predominantly in T cells [6,40,58]. Accordingly, *Nlrc5* deficiency caused a dramatic defect in MHCI expression in T cells and an intermediate phenotype in B cells, and a milder defect was observed in innate APCs. Notably, reduced MHCI levels on *Nlrc5- /-* lymphocytes facilitated evasion from cytotoxic T cell-mediated surveillance, while *Nlrc5<sup>-/-</sup>* B cells were defective in priming  $CD8^+$  T cell responses [40,58].

A prerequisite for the activation of several NLRs, as well as for RLRs, is the internalization of their specific stimuli. It is currently unclear how this is achieved by T lymphocytes, which are considered non-phagocytic cells. However, NLRs such as CIITA and NLRC5 acquired regulatory functions that are independent from a DAMP or PAMP type of ligand, delineating a novel and fascinating evolution of their activity (as schematically illustrated in Figure 1).

#### **Concluding Remarks**

Although detailed studies into the role of PRRs in adaptive immunity are relatively few, there is growing evidence to suggest an important function for innate receptors in adaptive immunity. On the one hand, B and T lymphocytes can be clearly endowed with innate immune-sensing properties, often integrating antigen-receptor signaling. This is illustrated by the example of B cells, where TLR engagement is important for the development of antibody responses, or by emerging data supporting formation of an inflammasome platform in lymphoid cells.

On the other hand, the dependency on a ligand and the signaling pathways activated downstream of PRRs in lymphoid cells can differ from what we have learned from innate immune cells (Figure 1). Some PRRs even fulfill their functions constitutively, as in the case of CIITA and NLRC5. In this evolution from PAMP/DAMP-inducible to constitutive function, several questions remain to be answered. Is a ligand required for the function of these NLRs or for LGP2 in T cells? The former of these are active even in the absence of infectious or inflammatory signals, so what could be the nature of the ligand or 'activator'? Indeed, the inaptitude of T cells to phagocytose suggests that an agonist, if existing, would be of endogenous origin. Could there be a 'modern surrogate' that has evolved in place of the innate pathogen- or danger-associated pattern? To some extent, this is reminiscent of the intensively investigated mechanism leading to the activation of NLRP3 [45]. Though NLRP3 activity depends on DAMPs or PAMPs, the wide spectrum of activating stimuli suggests that NLRP3 itself is unlikely to interact directly with them, leading to the hypothesis that a secondary 'event' or 'messenger' is mediating inflammasome assembly.

With regard to this idea, the study of PRRs in B and T cells might unveil aspects of their activity, which could inform studies on PRRs in the innate immune system, and *vice versa*. Further understanding of classical and novel roles of PRRs in lymphocytes could ultimately open new avenues for therapeutic targeting of the adaptive immune system.

#### **Acknowledgements**

We thank F. Martinon, S. Chelbi, and K. Ludigs, UNIL, Lausanne, Switzerland, for critical reading of the manuscript. Studies in the group of GG are funded by the Swiss National Science Foundation (PP00P3\_139094) and the European Research Council (ERC-2012-StG310890). MCM is supported by the Institut National de la Santé et de la Recherche Médicale and Université de Lyon I. KM is supported by an Australian National Health and Medical Research Council Overseas Biomedical Training Fellowship (ID 1013515).

The authors declared no conflicts of interest.

#### **Tables and Figure Legends**

#### **Table 1. RLRs in adaptive immune cells**

The current knowledge on RLR expression in adaptive immune cells, their accepted function, and their specific role in T or B cells are summarized.



#### **Table 2. NLRs in adaptive immune cells**

This table focuses on NLRs, which have been reported to be expressed in T or B cells. Their established function and their adaptive cell-intrinsic roles are summarized.





### **Figure 1. PRRs in adaptive immune cells: from microbial sensing to constitutive functions**

This figure covers the spectrum of possible activities played by PRRs in B and T lymphocytes, referring to their mode of activation and downstream signaling cascade. As depicted on the left-hand side, PRRs can act in their canonical way in cells of the adaptive immune system; that is, their activity is induced by PAMPs or DAMPs, and triggers innate signaling pathways such as NF-κB. This is well exemplified by TLRs in B cells.

Conversely, the NLRs CIITA and NLRC5 fulfill an 'atypical' function, acting as transcriptional regulators of MHCs, the key molecules for adaptive immune responses. Moreover, these NLRs transactivate MHC expression also constitutively, indicating that they evolved activities, which are independent of pathogen-derived or danger signals. Moving from innate to adaptive, from inducible to constitutive, the function of PRRs reveals exciting evolutionary paths.

#### **References**

- 1. McGettrick AF, O'Neill LA: Toll-like receptors: key activators of leucocytes and regulator of haematopoiesis. *Br J Haematol* 2007, 139:185-193.
- 2. Chaturvedi A, Pierce SK: How location governs toll-like receptor signaling. *Traffic*  2009, 10:621-628.
- 3. Gururajan M, Jacob J, Pulendran B: Toll-like receptor expression and responsiveness of distinct murine splenic and mucosal B-cell subsets. *PLoS One* 2007, 2:e863.
- 4. Pietschmann K, Beetz S, Welte S, Martens I, Gruen J, Oberg HH, Wesch D, Kabelitz D: Toll-like receptor expression and function in subsets of human gammadelta T lymphocytes. *Scand J Immunol* 2009, 70:245-255.
- 5. Zarember KA, Godowski PJ: Tissue expression of human Toll-like receptors and differential regulation of Toll-like receptor mRNAs in leukocytes in response to microbes, their products, and cytokines. *J Immunol* 2002, 168:554-561.
- 6. Wu C, Orozco C, Boyer J, Leglise M, Goodale J, Batalov S, Hodge CL, Haase J, Janes J, Huss JW, 3rd, et al.: BioGPS: an extensible and customizable portal for querying and organizing gene annotation resources. *Genome Biol* 2009, 10:R130.
- 7. Hornung V, Rothenfusser S, Britsch S, Krug A, Jahrsdorfer B, Giese T, Endres S, Hartmann G: Quantitative expression of toll-like receptor 1-10 mRNA in cellular subsets of human peripheral blood mononuclear cells and sensitivity to CpG oligodeoxynucleotides. *J Immunol* 2002, 168:4531-4537.
- 8. Browne EP: Regulation of B-cell responses by Toll-like receptors. *Immunology*  2012, 136:370-379.
- 9. Meyer-Bahlburg A, Rawlings DJ: Differential impact of Toll-like receptor signaling on distinct B cell subpopulations. *Front Biosci* 2012, 17:1499-1516.
- 10. Genestier L, Taillardet M, Mondiere P, Gheit H, Bella C, Defrance T: TLR agonists selectively promote terminal plasma cell differentiation of B cell subsets specialized in thymus-independent responses. *J Immunol* 2007, 178:7779-7786.
- 11. Heng TS, Painter MW: The Immunological Genome Project: networks of gene expression in immune cells. *Nat Immunol* 2008, 9:1091-1094.
- 12. Bernasconi NL, Onai N, Lanzavecchia A: A role for Toll-like receptors in acquired immunity: up-regulation of TLR9 by BCR triggering in naive B cells and constitutive expression in memory B cells. *Blood* 2003, 101:4500-4504.
- 13. Lanzavecchia A, Bernasconi N, Traggiai E, Ruprecht CR, Corti D, Sallusto F: Understanding and making use of human memory B cells. *Immunol Rev* 2006, 211:303-309.
- 14. Pasare C, Medzhitov R: Control of B-cell responses by Toll-like receptors. *Nature* 2005, 438:364-368.
- 15. Hou B, Saudan P, Ott G, Wheeler ML, Ji M, Kuzmich L, Lee LM, Coffman RL, Bachmann MF, DeFranco AL: Selective utilization of Toll-like receptor and MyD88 signaling in B cells for enhancement of the antiviral germinal center response. *Immunity* 2011, 34:375-384.<br>•• By using a B cell-specific *MyD88*-knockout mouse, the authors show that this

adaptor is required for optimal antibody responses upon certain immunization regimens.

- 16. Meyer-Bahlburg A, Rawlings DJ: B cell autonomous TLR signaling and autoimmunity. *Autoimmun Rev* 2008, 7:313-316.
- 17. Bendigs S, Salzer U, Lipford GB, Wagner H, Heeg K: CpGoligodeoxynucleotides co-stimulate primary T cells in the absence of antigenpresenting cells. *Eur J Immunol* 1999, 29:1209-1218.
- 18. Gelman AE, Zhang J, Choi Y, Turka LA: Toll-like receptor ligands directly promote activated CD4+ T cell survival. *J Immunol* 2004, 172:6065-6073.
- 19. Gelman AE, LaRosa DF, Zhang J, Walsh PT, Choi Y, Sunyer JO, Turka LA: The adaptor molecule MyD88 activates PI-3 kinase signaling in CD4+ T cells and enables CpG oligodeoxynucleotide-mediated costimulation. *Immunity* 2006, 25:783-793.
- 20. Komai-Koma M, Jones L, Ogg GS, Xu D, Liew FY: TLR2 is expressed on activated T cells as a costimulatory receptor. *Proc Natl Acad Sci U S A* 2004, 101:3029-3034.
- 21. Asprodites N, Zheng L, Geng D, Velasco-Gonzalez C, Sanchez-Perez L, Davila E: Engagement of Toll-like receptor-2 on cytotoxic T-lymphocytes occurs in vivo and augments antitumor activity. *FASEB J* 2008, 22:3628-3637.
- 22. Cottalorda A, Verschelde C, Marcais A, Tomkowiak M, Musette P, Uematsu S, Akira S, Marvel J, Bonnefoy-Berard N: TLR2 engagement on CD8 T cells lowers the threshold for optimal antigen-induced T cell activation. *Eur J Immunol* 2006, 36:1684-1693.
- 23. Sobek A, Gustafsson O: Latitudinal fractionation of polychlorinated biphenyls in surface seawater along a 62 degrees N-89 degrees N transect from the southern Norwegian Sea to the North Pole area. *Environ Sci Technol* 2004, 38:2746-2751.
- 24. McCarron M, Reen DJ: Activated human neonatal CD8+ T cells are subject to immunomodulation by direct TLR2 or TLR5 stimulation. *J Immunol* 2009, 182:55-62.
- 25. Caron G, Duluc D, Fremaux I, Jeannin P, David C, Gascan H, Delneste Y: Direct stimulation of human T cells via TLR5 and TLR7/8: flagellin and R-848 upregulate proliferation and IFN-gamma production by memory CD4+ T cells. *J Immunol* 2005, 175:1551-1557.
- 26. Reynolds JM, Pappu BP, Peng J, Martinez GJ, Zhang Y, Chung Y, Ma L, Yang XO, Nurieva RI, Tian Q, et al.: Toll-like receptor 2 signaling in CD4(+) T lymphocytes promotes T helper 17 responses and regulates the pathogenesis of autoimmune disease. *Immunity* 2010, 32:692-702.<br>
• Nice study showing a CD4<sup>+</sup> T cell-intrinsic role for TLR2 in promoting Th17

development.

- 27. Imanishi T, Hara H, Suzuki S, Suzuki N, Akira S, Saito T: Cutting edge: TLR2 directly triggers Th1 effector functions. *J Immunol* 2007, 178:6715-6719.
- 28. Biswas A, Banerjee P, Biswas T: Porin of Shigella dysenteriae directly promotes toll-like receptor 2-mediated CD4+ T cell survival and effector function. *Mol Immunol* 2009, 46:3076-3085.
- 29. Tabiasco J, Devevre E, Rufer N, Salaun B, Cerottini JC, Speiser D, Romero P: Human effector CD8+ T lymphocytes express TLR3 as a functional coreceptor. *J Immunol* 2006, 177:8708-8713.
- 30. Peng G, Guo Z, Kiniwa Y, Voo KS, Peng W, Fu T, Wang DY, Li Y, Wang HY, Wang RF: Toll-like receptor 8-mediated reversal of CD4+ regulatory T cell function. *Science* 2005, 309:1380-1384.
- 31. Caramalho I, Lopes-Carvalho T, Ostler D, Zelenay S, Haury M, Demengeot J: Regulatory T cells selectively express toll-like receptors and are activated by lipopolysaccharide. *J Exp Med* 2003, 197:403-411.
- 32. Mills KH: TLR-dependent T cell activation in autoimmunity. *Nat Rev Immunol*  2011, 11:807-822.
- 33. Loo YM, Gale M, Jr.: Immune signaling by RIG-I-like receptors. *Immunity* 2011, 34:680-692.
- 34. Anz D, Koelzer VH, Moder S, Thaler R, Schwerd T, Lahl K, Sparwasser T, Besch R, Poeck H, Hornung V, et al.: Immunostimulatory RNA blocks suppression by regulatory T cells. *J Immunol* 2010, 184:939-946.
- 35. Suthar MS, Ramos HJ, Brassil MM, Netland J, Chappell CP, Blahnik G, McMillan A, Diamond MS, Clark EA, Bevan MJ, et al.: The RIG-I-like Receptor LGP2 Controls CD8(+) T Cell Survival and Fitness. *Immunity* 2012.<br>
•• Description of an intrinsic role for LGP2 in promoting West Nile virus-specific

CD8+ T cell survival independently of MAVS.

- 36. Kufer TA, Sansonetti PJ: NLR functions beyond pathogen recognition. *Nat Immunol* 2011, 12:121-128.
- 37. Strowig T, Henao-Mejia J, Elinav E, Flavell R: Inflammasomes in health and disease. *Nature* 2012, 481:278-286.
- 38. Chen G, Shaw MH, Kim YG, Nunez G: NOD-like receptors: role in innate immunity and inflammatory disease. *Annu Rev Pathol* 2009, 4:365-398.
- 39. Reith W, LeibundGut-Landmann S, Waldburger JM: Regulation of MHC class II gene expression by the class II transactivator. *Nat Rev Immunol* 2005, 5:793- 806.
- 40. Kobayashi KS, van den Elsen PJ: NLRC5: a key regulator of MHC class Idependent immune responses. *Nat Rev Immunol* 2012, 12:813-820.
- 41. Doitsh G, Cavrois M, Lassen KG, Zepeda O, Yang Z, Santiago ML, Hebbeler AM, Greene WC: Abortive HIV infection mediates CD4 T cell depletion and inflammation in human lymphoid tissue. *Cell* 2010, 143:789-801.<br>•• Study showing induction of the pro-inflammatory response through caspase-1
- activation in  $CD4<sup>+</sup>$  T cells upon HIV infection.

42. Masters SL, Gerlic M, Metcalf D, Preston S, Pellegrini M, O'Donnell JA, McArthur K, Baldwin TM, Chevrier S, Nowell CJ, et al.: NLRP1 Inflammasome Activation Induces Pyroptosis of Hematopoietic Progenitor Cells. *Immunity* 2012, 37:1009-1023.<br>
•• Demonstration of NLRP1a role to induce pyroptosis in hematopoietic progenitor

cells upon hematopoietic stress.

43. Lech M, Avila-Ferrufino A, Skuginna V, Susanti HE, Anders HJ: Quantitative expression of RIG-like helicase, NOD-like receptor and inflammasomerelated mRNAs in humans and mice. *Int Immunol* 2010, 22:717-728. •

Useful analysis of mRNA expression of different NLRs and RLRs in human and mouse tissues, monocytes, and T lymphocytes.

44. Mercier BC, Ventre E, Fogeron ML, Debaud AL, Tomkowiak M, Marvel J, Bonnefoy N: NOD1 Cooperates with TLR2 to Enhance T Cell Receptor-Mediated Activation in CD8 T Cells. *PLoS One* 2012, 7:e42170.

" Study showing that NOD1 stimulation in  $CD8<sup>+</sup>$  T cells increases TCR-driven proliferation and effector functions, also cooperating with TLR2.

- 45. Schroder K, Tschopp J: The inflammasomes. *Cell* 2010, 140:821-832.
- 46. Sanz C, Calasanz MJ, Andreu E, Richard C, Prosper F, Fernandez-Luna JL: NALP1 is a transcriptional target for cAMP-response-element-binding protein (CREB) in myeloid leukaemia cells. *Biochem J* 2004, 384:281-286.
- 47. Petterson T, Jendholm J, Mansson A, Bjartell A, Riesbeck K, Cardell LO: Effects of NOD-like receptors in human B lymphocytes and crosstalk between NOD1/NOD2 and Toll-like receptors. *J Leukoc Biol* 2011, 89:177-187. • This work investigates NLR expression in human B lymphocytes and shows that

stimulation with NOD1 agonist enhances BCR-induced activation.

- 48. Petterson T, Mansson A, Riesbeck K, Cardell LO: Nucleotide-binding and oligomerization domain-like receptors and retinoic acid inducible gene-like receptors in human tonsillar T lymphocytes. *Immunology* 2011, 133:84-93.
- 49. Guarda G, Zenger M, Yazdi AS, Schroder K, Ferrero I, Menu P, Tardivel A, Mattmann C, Tschopp J: Differential Expression of NLRP3 among Hematopoietic Cells. *J Immunol* 2011, 186:2529-2534.
- 50. Rahman MK, Midtling EH, Svingen PA, Xiong Y, Bell MP, Tung J, Smyrk T, Egan LJ, Faubion WA, Jr.: The pathogen recognition receptor NOD2 regulates human FOXP3+ T cell survival. *J Immunol* 2010, 184:7247-7256.
- 51. Kumar H, Kumagai Y, Tsuchida T, Koenig PA, Satoh T, Guo Z, Jang MH, Saitoh T, Akira S, Kawai T: Involvement of the NLRP3 inflammasome in innate and humoral adaptive immune responses to fungal beta-glucan. *J Immunol* 2009, 183:8061-8067.
- 52. Caetano BC, Biswas A, Lima DS, Jr., Benevides L, Mineo TW, Horta CV, Lee KH, Silva JS, Gazzinelli RT, Zamboni DS, et al.: Intrinsic expression of Nod2 in CD4+ T lymphocytes is not necessary for the development of cell-mediated immunity and host resistance to Toxoplasma gondii. *Eur J Immunol* 2011, 41:3627-3631.
- 53. Shaw MH, Reimer T, Sanchez-Valdepenas C, Warner N, Kim YG, Fresno M, Nunez G: T cell-intrinsic role of Nod2 in promoting type 1 immunity to Toxoplasma gondii. *Nat Immunol* 2009, 10:1267-1274.
- 54. Nembrini C, Reissmann R, Kopf M, Marsland BJ: Effective T-cell immune responses in the absence of the serine/threonine kinase RIP2. *Microbes Infect*  2008, 10:522-530.
- 55. Conti BJ, Davis BK, Zhang J, O'Connor W, Jr., Williams KL, Ting JP: CATERPILLER 16.2 (CLR16.2), a novel NBD/LRR family member that negatively regulates T cell function. *J Biol Chem* 2005, 280:18375-18385.
- 56. Ling A, Soares F, Croitoru DO, Tattoli I, Carneiro LA, Boniotto M, Benko S, Philpott DJ, Girardin SE: Post-transcriptional inhibition of luciferase reporter assays by the Nod-like receptor proteins NLRX1 and NLRC3. *J Biol Chem*  2012, 287:28705-28716.
- 57. Schneider M, Zimmermann AG, Roberts RA, Zhang L, Swanson KV, Wen H, Davis BK, Allen IC, Holl EK, Ye Z, et al.: The innate immune sensor NLRC3 attenuates Toll-like receptor signaling via modification of the signaling adaptor TRAF6 and transcription factor NF-kappaB. *Nat Immunol* 2012.
- 58. Staehli F, Ludigs K, Heinz LX, Seguin-Estevez Q, Ferrero I, Braun M, Schroder K, Rebsamen M, Tardivel A, Mattmann C, et al.: NLRC5 Deficiency

Selectively Impairs MHC Class I- Dependent Lymphocyte Killing by

Cytotoxic T Cells. *J Immunol* 2012.<br>•• By using a knockout mouse, this work demonstrates that NLRC5 drives MHCI transcription in particular in lymphocytes.