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2 Impact of NOM Character on Copper Adsorption by

3 Trace Ferric Hydroxide from Iron Corrosion in

4 Water Supply System

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ABSTRACT

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The fate of trace concentrations of cupric sulphate (< 0.4 mg/L) dosed into chloraminated distribution systems to inhibit nitrification has been shown in this study to be controlled by at least two crucial factors: the character of natural organic matter (NOM) and iron hydroxide corrosion products present at low concentrations (< 2 mg/L). This research quantified the removal of Cu(II) ions added into waters containing trace Fe(OH)₃ flocs and the effect of NOM of different character on this removal. The dominant dissolved copper species in NOM-containing waters were found to be Cu(II)-NOM complexes. Both intramolecular chelation and intermolecular complexation can occur, with the latter occurring preferentially and resulting in the aggregation of smaller organic molecules to form larger molecules. The presence of ferric hydroxide flocs when Cu(II) ions were added into NOM-containing waters was shown to result in removal of Cu(II) ions, presumably as Cu(II)-NOM complexes. This removal was through adsorption processes obeying Freundlich isotherms, although the presence of larger NOM molecules and heterogeneous copper species (e.g. Cu(OH)2(s) and CuO(s)) appeared to shield smaller Cu(II)-NOM complexes from adsorption to some extent. For the strategy of inhibition of nitrification in distribution systems by the addition of Cu(II) ions, complexation of Cu(II) ions by NOM and adsorption of Cu(II)-NOM complexes by ferric hydroxide flocs released from pipe walls pose significant operational challenges to maintaining the concentration of Cu(II) ions through the distribution system.

- 41 **Key Words:** Chloramine; Adsorption; Copper; Corrosion; Distribution system; Inhibition; Nitrification;
- 42 NOM

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1. INTRODUCTION

Maintaining an adequate disinfectant residual at water distribution extremities is required by the 44 guidelines of the environmental protection agency. Chloramine is chosen especially when challenging 45 conditions are encountered in distribution systems, including aging pipes, unexpected chlorine loss due 46 to inefficient organics removal in water treatment or other causes such as disinfectant by-products. The 47 Goldfields and Agricultural Water Supply Scheme (G&AWSS) in Western Australia is currently 48 applying chloramine mainly because the authorities met with incidences of Naegleria fowleri when 49 chlorine was used as disinfectant in their above ground pipe line system which is about 530 km end to 50 end with branches of pipeline totaling about 7000 km and experiences temperatures from 8 - 45°C [1]. 51 N. fowleri is an amoeba and has been responsible for a number of fatal infections linked to public water 52 supplies. While chloramine provided a better alternative, it met with another challenge. Nitrification, a 53 microbial process serially converting ammonia to nitrite, has significantly reduced the stability of 54 disinfectant. Therefore, it is important to find solution to stabilize the chloramine. 55 More than 0.1 mg-Cu(II)/L was found to inhibit nitrifying bacteria [2]. The use of cupric sulphate 56 (CuSO₄) in lakes, reservoirs and other managed bodies remains the most effective algicidal treatment [3, 57 4, 5]. The World Health Organization (WHO) recommends a copper concentration of 1 mg/L as an 58 aesthetic standard for drinking water [6]. A novel method of dosing cupric sulphate (up to 400 µg-59

Cu(II)/L) directly into drinking water distribution systems to inhibit the activity of nitrifying bacteria [7] and protect chloramine residuals is being trialed in the G&AWSS. Effective inhibition and the desired copper concentration were achieved when copper dosing was practiced in a pilot reservoir. However, dissolved Cu(II) concentrations were found to decrease with distance along the cement-lined cast iron pipeline, limiting the effectiveness of this approach. Therefore, it is important to understand the fate of copper in the distribution system and devise approaches to maintain its concentration in the pipe line. The likely species of copper in natural water bodies have been extensively studied. Wagemann and Barica [8] claimed that only 5% of total copper could be found as free Cu²⁺ in natural water systems. Snoeyink and Jenkins [9] summarized the status of various inorganic copper compounds being in equilibrium with tenorite (CuO) in carbonate-buffered water, controlled by the partial pressure of CO₂. The majority of copper existing in natural water bodies is believed to be in the form of Cu-NOM complexes [10]. Edwards and Nicolle [11] reported the effects of NOM on copper corrosion by-product release and found that NOM can interfere with the formation of a solid scale layer of Cu(OH)2 and dramatically increase the soluble copper concentration in water. Hullebusch et al. [12] suggested a proportional relationship between organic copper compounds and dissolved organic carbon (DOC). Gamble et al. [13] noted the aggregation mechanism adopted by Cu(II) when binding multiple molecules.

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The solubility of Cu(II) is reported to be controlled by an equilibrium between the soluble copper complex and metastable solid phases, such as cupric hydroxide (Cu(OH)₂) and tenorite (CuO) [14]. The

size of $Cu(OH)_2$ particles varies with pH. At pH 8, the pH of water in the G&AWSS, the size of $Cu(OH)_2$ particulates was reported to be about 400 nm [15]. Our laboratory experiments have also shown that both $Cu(OH)_2$ and CuO particles can be removed by 0.2 μ m membrane filters at pH >7.5 [16], such that these filters were chosen to remove particulate copper formed in the current research.

Our previous research using differential absorbance spectroscopy showed that NOM in Mundaring Weir water (the feed water for the G&AWSS) was deficient in phenolic chromophores which are the major constituents of the hydrophobic fraction of NOM [17]. Gamble et al. [18] suggested that phenolic chromophores contribute to salicylic-acid type of bidentate chelating sites, which are believed to preferentially chelate with Cu²⁺ and form relatively stable chelates. Controversially, one striking finding revealed that the removal of the hydrophobic acid fraction from the surface water had little effect on Cu binding, suggesting that the binding affinities of Cu to phenolic sites are weak [19].

When copper is dosed into an iron pipe distribution system, its fate could be affected by iron corrosion. The vulnerability of a distribution system to corrosion depends on the chemical properties of the water delivered (e.g. pH, alkalinity, dissolved oxygen, total dissolved solids) and its physical characteristics (temperature, velocity), as well as the nature of pipe materials [20]. McNeill and Edwards [21] did a comprehensive review of such various important factors, including dissolved oxygen (DO), pipe age, biological activity, disinfectants, temperature, and dissolved copper, as they affect iron pipe corrosion. Generally, the corrosion rate increases with the increased DO concentration [22]. Zhang and Edwards [23] reported that cast iron can also reduce nitrite/nitrate to ammonia, indicating iron pipe

that might stimulate microbiologically influenced corrosion (MIC) [24]. Disinfectant residuals in general increase the corrosion rate [25]. Both chloramine and nitrification are present in the G&AWSS. When copper sulphate is dosed into the G&AWSS to inhibit nitrification, a series of potential redox reactions could be involved [9]. These reactions are listed in Table 1, arranged in order of oxidizing capacity: O₂>NO₃->Cu²⁺>Fe²⁺. Either in bulk water under aerating conditions or nitrification, both of which are considered possible aquatic conditions in the distribution system, direct reaction between Cu^{2+} and exposed iron, can be overruled by the presence of stronger oxidizing agents (O₂ and NO₃). Chlorine and chloramine are also strong oxidizing agents which can compete with Cu²⁺ to react with iron. Our earlier study [26] and studies by others [8, 10] found little dissolved copper existing in natural water in the form of cupric ions, as most of the copper existed as Cu-NOM complexes. This finding further excludes the possible reaction between free cupric ions and iron. Therefore, the effect of iron pipe corrosion on dissolved Cu(II) in the context of natural waters literally refers to the interaction between the dominant dissolved Cu(II)-NOM and trace iron corrosion products released through corrosion. Dissolved Cu(II) removal by iron salts has been extensively documented. Ridge and Sedlak [27] reported that Cu²⁺ and Cu-EDTA complexes up to 13 mg-Cu(II)/L were noticeably removed by hydrous

ferric oxide (HFO) in wastewater treatment. Iron oxides have also been found to be good adsorbents to

remove high concentrations of cupric ions and ammonia-complexed copper in wastewater [28]. Since

corrosion caused by the products of nitrification. Nitrification can also increase the growth of bacteria

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dissolved copper of high concentrations can be efficiently removed by iron salts in wastewater treatment processes, it raised a concern that the solubility of dosed copper salts could, on the other hand, be jeopardized by iron salts released into distribution systems due to iron pipe corrosion, even if iron salts are present at low concentrations.

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Iron corrosion scales include goethite (a-FeOOH), lepidocrocite (g-FeOOH), magnetite (Fe₃O₄), siderite (FeCO₃), ferrous hydroxide (Fe(OH)₂), ferric hydroxide (Fe(OH)₃), ferrihydrite (5Fe₂O₃•9H₂O), green rusts (e.g. Fe₄^{II}Fe₂^{III}(OH)₁₂CO₃) and calcium carbonate (CaCO₃). A high concentration of readily soluble Fe(II) content is reported to be present inside scales [28, 29]. On one hand, ferrous ions can be released into bulk water during corrosion and oxidized to ferric particles. Sarin et al. [30] reported that iron is released to bulk water primarily in the ferrous form. The ferrous ions are converted into ferrous solids (e.g. Fe(OH)₂), which may then be converted to ferric solids (e.g. Fe(OH)₃) after reaction with oxygen or other oxidising agents such as chloramine which are consistently present in the drinking water [20]. On the other hand, NOM existing in the distribution system can form complexes with released ferrous or ferric ions. Jones et al. [31] studied the dissociation kinetics of Fe(III)-NOM complexes and reported the stability constant K for Fe(III)-NOM complexation at pH 8.0 within a range of 9.90~10.34, which is significantly smaller than the stability constant K of 10^{2.4} for the formation of $Fe(OH)^{2+}$ or $10^{2.3}$ for the formation of $Fe(OH)_{3(s)}$ [9]. The stability constant for Fe(II)-NOM complexation is less than those for the Fe(III)-NOM complexation by two to four orders of magnitude [32]. Therefore, the complexation between Fe(II) or Fe(III) and organic matter is insignificant or

negligible compared to the strong stability of Fe(OH)₃ formed in the drinking water at pH 8.0 short after ferrous ions released into bulk water due to corrosion. Therefore, pre-formed ferric hydroxide flocs were chosen to be the representative of the major corrosion products which are stable and available in a relatively long period to react with dissolved Cu(II) in bulk water.

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Based on these previous studies, Cu(II)-NOM complexes are unanimously acknowledged as the major soluble copper forms in natural water, though the structure and preference of copper chelation with certain fractions of specific organic matter remains unresolved. High concentrations of cupric ions and complexed copper (up to 500 mg/L) in wastewater can be removed by adsorption to iron oxide media and filtration. However, from the perspective of protecting low concentrations of dissolved Cu(II) in distribution systems (e.g. as a nitrification inhibitor), little research has been conducted on dissolved Cu(II) removal by low concentration (< 2 mg/L) iron pipe corrosion products released from corroded or damaged pipes in distribution systems like the G&AWSS. Preliminary experiments using water from Mundaring Weir showed adsorption of dissolved Cu(II) by Fe(OH)₃ flocs present at trace concentrations, with the adsorption obeying Langmuir or Freundlich adsorption isotherms [26]. However, the relationships between NOM character and its distinct impact on copper-organic chelation and dissolved Cu(II) removal by trace Fe(OH)3 flocs remain unknown. Furthermore, the fate of dissolved Cu(II) in water containing nitrifying bacteria is also of interest. This research aims to quantify the removal of dosed Cu(II) in bulk water containing trace Fe(OH)₃ flocs and NOM of different characters. The impact of NOM characteristics on dissolved Cu(II) removal was elucidated using molecular weight distribution detected by UV_{254} .

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2. MATERIALS AND METHODS

2.1 Selection, Sampling and Preparation of Water Samples

Selected samples and the experimental protocol are given in Figure 1. To evaluate the effect of NOM of different characters on Cu(II) removal, three different waters were selected. Mundaring raw water (MRW) was selected as this is the source water for the G&AWSS. Pre-coagulated Mundaring raw water (MCW) was prepared as comparison with MRW to evaluate how coagulable NOM and its absence can affect the character of Cu-NOM complexes and thus their removal by Fe(OH)₃ flocs. As coagulation preferentially removes hydrophobic fraction mainly containing aquatic humic material with high MW [33], the majority of NOM contained in MCW would be uncoagulable and of small molecular weight. The organic Cu(II) complexes formed between dissolved Cu(II) and these uncoagulable NOM were also expected to be of smaller molecular size than those formed in MRW and hence be protected from removal by ferric flocs. A synthetic water largely containing a hydrophobic fraction was prepared with commercial humic acid and termed humic acid bulk water (HAW) in comparison with MRW which is known to mainly contain hydrophilic organic compounds [17]. In order to study the effect of nitrification observed in the distribution system on organic Cu(II) complexation and its removal by ferric flocs, a severely nitrified water (NW) sample was collected from the laboratory

reactors fed with Mundaring water. Severe nitrification is defined as the condition when nitrite concentration was more than 0.1 mg-N/L and chloramine decay is severely accelerated [34].

Depending on the copper dose, addition of copper was found to form particles in some experiments and hence some sub-samples of MRW were filtered through a 0.2µm membrane filter to remove the effect of preformed Cu(II)-containing particles. This filtration was not done for HAW and NW because their preparation step already involved filtration.

Mundaring Raw Water (MRW): Regular sample collection from the outlet of Mundaring Weir, the source water for the G&AWSS, has been conducted by our group from July 2008 to the present. The quality of Mundaring water has been found to vary only slightly in terms of pH (7.6~8.1), DOC (2.4~3.0 mg-C/L) and UV₂₅₄ absorbance (0.031~0.038 cm⁻¹). A sample for this study was collected in July 2009. The raw water was filtered through a 0.45 μm membrane to remove suspended solids or particles before copper dosing.

Post-Coagulated Mundaring Water (MCW): Ferric chloride was employed as the coagulant to remove coagulable NOM in MRW. Coagulation experiments showed that maximum DOC removal was achieved by adding a coagulant dose (concentration in the jar) of 40 mg-FeCl₃/L at pH 5~5.5. Kastl et al. [35] observed similar doses and pH resulting in maximum DOC removal in many waters. A jar tester was used to control the coagulation process. The stirring speed was set at 200 rpm and ferric chloride was dosed and the speed was continued for the next 2 minutes. The speed was set at 20 rpm and continued for another 20 minutes. During coagulation, the pH of the bulk water was adjusted and

maintained at 5~5.5 using a HACH40d pH meter. HCl (1 N) and NaOH (1 N) were used to adjust pH. After the coagulation, bulk water was kept intact for sedimentation and then filtered through 0.45 μm and 0.2 μm polycarbonate membranes consecutively to remove flocs. NaOH (1 N) was titrated into the filtrate to increase the pH to 7.8±0.2. The concentration of dissolved Fe(II) or Fe(III) in the filtrate was determined using atomic absorption spectroscopy in SGS Pty. Ltd (WA) and found to be <0.05 mg/L.

Humic Acid Water (HAW): Humic acid water was prepared by diluting stock HA solution (3 g-C/L) with MilliQ water (18 MΩ/cm, DOC<100 ppb). Humic acid was acquired from Sigma Aldrich®, containing 20% ash. HA solid was dissolved in MilliQ water first then centrifuged at 4000 rpm for 5 minutes to separate ash and other insoluble particles from the solution. The supernatant was withdrawn and filtered through 0.45 μm membrane to further remove insoluble impurities. The final filtrate was treated as the stock HA standard solution (3 g-C/L). This solution was diluted to give HAW the following characteristics: 2.5 ± 0.1 mg-C/L DOC and had UV Abs=0.248/cm (254 nm, 10 cm quartz cell). This concentration of DOC was chosen to be as equal as possible to the DOC concentration in MRW.

Nitrified Water (NW): Two identical reactor systems, each assembled in series with five 20 L reactors (R1 to R5), were set up in the laboratory. Automatic flow rate and temperature control were installed for the reactors. Chloraminated Mundaring water was fed into the reactor, with mass ratio of (Cl:NH₃-N) 4.5 to 1 maintained in the 25-litre feeding tank. In the start-up period, chlorine was maintained at about 1mg/L. To expedite nitrification and to obtain the DS inoculums, chloraminated water collected from G&AWSS was added into the reactors except R1. The chloramine concentration

was gradually increased up to 2.5 mg/L in the feeding tank. Water (20 L) was fed into the system continuously every day to gain retention time of 20±2 hrs. Water temperature was maintained at 20±2°C in the reactors R1~R3 whereas 23±2°C was maintained in R4 and R5 in order to achieve accelerated microbial activities. By varying hydraulic conditions, temperature and chloramine residuals, nitrification occurring in the distribution system can be simulated in the laboratory. In this research, NW was collected from R4. NW contained 0.10 mg/L NH₃-N, 0.2 mg/L NO₂-N and 0.1 mg/L NO₃-N. The levels of these inorganic nitrogen products indicated that severe nitrification occurred in NW [33].

2.2 Analysis of Soluble Copper and Iron Species

Soluble Cu(II), Fe(II) and Fe(III) concentrations in the source waters and bulk water samples prepared in the laboratory were measured by Atomic Absorption Spectroscopy (AAS) by a commercial laboratory before any experiment commenced. Total Cu(II) concentrations were found to be less than 20 µg/L before addition of copper and the dissolved Fe(II) and Fe(III) concentrations were below 0.05 mg/L before ferrous or ferric salt addition.

In the experiments, copper concentrations were analyzed after conversion of all forms of copper to soluble copper(II) form and then by the bicinchoninate spectrophotometric method (Hach method 8506; HACH DR2800). This method had a measuring range of 0.04~5 mg/L with ±20 µg/L accuracy. Samples were digested using nitric acid (1:1) at pH 4~6 for total copper measurement. For measuring dissolved copper concentrations, samples were filtered through a 0.2 µm PC (polycarbonate) membrane prior to measuring total Cu concentration in the filtrate. In order to minimise interaction between the sample and

membrane and to prevent speciation changes, the volume of the filtrate was chosen to represent approximately 50% of the volume of the raw sample [36]. For instance, to obtain 50 mL filtrate, 100 mL sample water was added and 50 mL was filtered through the filter paper.

2.3 Dissolved Organic Carbon Measurement and Molecular Weight Distribution Analysis

Water samples were filtered through a $0.45\mu m$ membrane before the analysis of UV_{254} absorbance and dissolved organic carbon (DOC) concentration. UV_{254} absorbance was measured at 254 nm in a 10 cm quartz cell using a Helios UV/V Spectrophotometer. DOC concentrations were analysed using a GE 5310C TOC analyser.

The apparent molecular weight distribution of the DOC detected by UV₂₅₄ in the samples was analysed by high performance size exclusion chromatography (HPSEC) according to the method of Allpike et al. [37] and Warton et al. [38], except that an Agilent 1100 Series HPLC system was used and that polystyrene sulfonate standards were used for molecular weight calibration. SEC was performed using a TSK G3000SWxl (TOSOH Biosep, 5 μm resin) column and an Agilent 1100 HPLC instrument with diode array detection at 254 nm. The column had an internal diameter of 7.8 mm and a length of 30 cm, with a void volume of 5.5 mL, determined with dextran blue. The eluent used was a 20 mM phosphate buffer (1.36 g/L KH₂PO₄ and 3.58 g/L Na₂HPO₄.12H2O) at a pH of 6.85. The sample volume was 100 μL and the flow rate was 1 mL/minute. Samples were first filtered through a 0.45 μm nylon filter. The system was calibrated using a combination of polystyrene sulfonate standards (840 Da,

1290 Da, 3610 Da, 6520 Da, 15200 Da, 81800 Da). The calibration curve was linear ($R^2 = 0.991$) over the apparent MW range tested.

2.4: Preparation of Fe(OH)₃ flocs

The method of preparing an Fe(OH)₃ suspension and its particle size distribution analysis are presented in the Supplementary Information (SI).

2.5 Cu(II) Solubility and Speciation

Copper solubility was determined in bulk waters under various conditions: MilliQ water (DOC<100 μ g/L, 18 $M\Omega$ /cm), CaCO₃ buffered Milli-Q water (50 mg-CaCO₃/L) and MRW. Copper solubility can be affected by the partial pressure of CO₂ contacting the water surface [9], and therefore two experiments representing open and closed systems were conducted to investigate Cu(II) solubility in each of these. In the open systems, the bulk water surface was open to the atmosphere (Log (P_{CO2}) = -3.50) without capping the container during the course of the experiment. The closed system was simulated by purging CO₂ from the bulk waters using nitrogen gas during the experiments and excluding air from the samples during storage. An aliquot (1 mL) was withdrawn from the CuSO₄ stock solution (1 g-Cu(II)/L) and added into the three types of bulk water (1 L), to attain a total Cu(II) concentration of 1 mg-Cu(II)/L. The dissolved Cu(II) concentration was then measured at 2, 4, 24 and 48 hours. The experimental results were compared with the desktop results modelled using MINEQL+®

(Version 4.6, chemical equilibrium modeling system) to verify possible Cu(II) species and their proportions.

2.6 Dissolved Cu(II) Removal by Trace Ferric Hydroxide Flocs

2.6.1 General Experimental Procedure

MRW, MCW, NW and HAW were used for the experiments. In each type of bulk water, different initial Cu(II) concentrations (250, 400 and 1000 μ g-Cu(II)/L) were made in duplicates, by dosing appropriate stock solution of copper sulphate.

To investigate dissolved Cu(II) removal by Fe(OH)₃ flocs from each bulk water sample and their duplicates, various amounts of Fe(OH)₃ suspension were added into each subsample containing a known dissolved Cu(II) concentration. The appropriate Fe(III) concentration was achieved by adding aliquots of Fe(OH)₃ stock suspension (1 g-Fe(III)/L). Tested Fe(III) concentrations were 0.1, 0.3 0.5, 1.0 and 2 mg-Fe(III)/L. A jar tester was used to stir (60 rpm for 30 mins) bulk water during and after the addition of ferric salts. After the jar test, the samples were left for sedimentation for 4 hours (4 hours was sufficient to reach equilibrium between dissolved Cu(II) species and Fe(OH)₃ flocs, see Figure 2S in SI). During this time, the pH of each sample was monitored and adjusted to around pH 7.8-8.2, by adding HCl solution (1 N) or NaOH solution (1 N), as necessary. Subsamples were then filtered through 0.2 µm polycarbonate membranes and the Cu(II) concentration remaining in the filtrate was measured.

The Cu(II) removal was calculated as the difference between the dissolved Cu(II) in the bulk water before and after Fe(III) treatment.

2.6.2 Method to Investigate the Effect of Pre-formed Cu(II) Particles on Dissolved Cu(II) Removal

In order to evaluate the effect of the pre-formed Cu(II) particles (e.g. Cu(OH)₂, CuO_(s)) on dissolved Cu(II) removal by Fe(OH)₃, the above post-Cu(II)-dosed samples were divided into two groups. In one group (MRW and MCW), pre-formed particles were allowed to remain in bulk water during Fe(OH)₃ treatment. In the other (MWF, NW and HAW), pre-formed Cu(II) particles were removed by filtration through a 0.2 µm polycarbonate membrane prior to Fe(OH)₃ addition.

3. RESULTS AND DISCUSSION

3.1 Water Quality Characteristics of the Water Samples

The water quality characteristics of the four bulk water samples are presented in Table 2. The apparent molecular weight (AMW) profiles detected by UV₂₅₄ from size exclusion chromatographic analysis (SEC-UV₂₅₄) of each sample are shown in Figure 2. The major NOM fraction(s) derived from AMW analysis are described in Table 3.

The DOC concentration of MRW, HAW and NW were similar, and MCW had a relatively low DOC concentration after removal of some of the DOC through coagulation. Despite having a slightly lower

DOC concentration than MRW, HAW had a much higher SUVA₂₅₄ than MRW (1.35 vs. 10.33 L mg⁻¹

m⁻¹, respectively (Table 2)), indicating substantially higher aromatic character in the HAW NOM [39]. MRW contained material with molecular weight less than 3000 Da (Figure 2) with majority of DOC having an AMW distribution between 2000-3000 Da. For ease of discussion, NOM in Mundaring water was classified as two fractions: coagulable (removed by coagulation, i.e. the NOM present in MRW but not in MCW) and uncoagulable (NOM_{uc}; NOM remaining post-coagulation in MCW). The majority of the 2000-3000 Da material in MRW was removed by coagulation, leaving DOC with AMW 1000-2000 Da being the predominant fraction in MCW (Table 3). This is consistent with previous reports of coagulation removing predominantly higher MW NOM [37, 38, 40]. In the NW sample, representing MRW subject to nitrification, there was a relative increase in one lower AMW fraction, centered on 500 Da (Figure 2). Two peaks between 900~1500 Da in MRW were not present in the chromatogram of NW, possibly indicating microbiological use of this low-medium AMW material and production of lower AMW material. This also supports our earlier observation that soluble microbial products are formed in the nitrifying waters [41,42] The much higher UV₂₅₄ response for DOC in the HAW sample compared to the MRW sample is consistent with the much higher SUVA₂₅₄ measured for the former sample. The AMW profiles suggest that NOM in HAW was comprised of much higher MW components (2-10 kDa) [43] than NOM in MRW (1-3 kDa) and clearly show that there is a substantial difference in the character of these two NOM types.

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3.2 Cu(II) Solubility and Speciation in the Bulk Waters under Various Aqueous Conditions

Table 4 summarizes the copper species (both particulate and dissolved forms) under different aqueous conditions.

According to Table 4, free cupric ions (Cu²⁺) are negligible in bulk water at pH>7.5. In equilibrium with CuO(s) in an open system, Cu(II)-inorganic compounds can be neglected. Therefore, the dominant dissolved copper species in NOM-containing bulk waters are Cu-NOM complexes. For instance, in MRW, 840 μg/L dissolved Cu(II) was found to be in the form of Cu(II)-NOM complexes. The remaining copper (160 μg/L) was presumably removed in the filtration step as copper particulates, possibly comprised of Cu(OH)₂ and CuO.

Figure 3 shows the dissolved Cu(II) concentration measured in MRW and MCW 24 hours after dosing with an aqueous copper sulphate solution to achieve an initial Cu concentration of 250, 400 and 1000 μg/L in the jar. The dissolved Cu(II) concentrations were almost equal in MRW and MCW when the initial copper concentration was 250 μg/L and 400 μg/L, with only 20~30 μg/L and 50~80 μg/L, respectively, found to be in the particulate form. However, when the initial Cu(II) concentration was increased to 1000 μg/L, the different solubility of copper in these two waters became evident. In MRW, the majority (840 μg/L) of the initial copper concentration was maintained as dissolved Cu-NOM, but in MCW, only 340 μg/L existed as dissolved Cu(II)-NOM, with 660 μg/L Cu consisting of particulate forms. This indicates that coagulable NOM (NOM_c) in MRW contributed significantly to increasing the copper solubility by forming Cu(II)-NOM_c (Cu(II)-coagulable-NOM complex) when the initial copper concentration was relatively higher. However, at lower initial copper concentrations (250 μg/L and 400

 μ g/L), uncoagulable NOM (NOM_{uc}) remaining in MCW was still capable of forming Cu-NOM_{uc} (Cu-uncoagulable-NOM complexes) at similar concentrations to those found in MRW. In other words, dosed copper preferentially complexed with NOM_{uc} in MRW until the uncoagulable fraction was saturated. Further increased initial copper concentration up to 1 mg/L started binding with the coagulable fraction.

3.3 Removal of Dissolved Cu(II) by Addition of Fe(OH)3 Flocs at Low Concentrations

Figure 4 (A, B, C, E and F) show the removal of dissolved Cu(II) by Fe(OH)₃ flocs in MRW, MWF,

NW, MCW and HAW respectively.

Addition of the flocs at a concentration of 2 mg-Fe/L effectively removed most of the dissolved Cu(II) from Mundaring water samples (MRW and MWF) as well as NW when the initial copper(II) concentrations were 250 or 400 µg/L. In the samples with initial Cu(II) concentration of 400 µg-Cu(II)/L, the addition of ferric hydroxide flocs (2 mg-Fe/L) resulted in less than 100 µg/L dissolved Cu(II) remaining (Figure 4A, B, C and E). Since the use of MINEQL® had established that the majority of copper in these samples existed as Cu(II)-NOM complexes, it is likely that the Fe(OH)₃ floc was, in fact, removing Cu(II)-NOM complexes.

The relative capacity of NOM to stabilize soluble Cu(II) was found to be stronger in HAW than in NW or MRW with higher initial concentrations of dissolved Cu(II) and less removal of dissolved Cu(II) by Fe(OH)₃ flocs observed in HAW (Figure 4F). For instance, only 20µg/L dissolved Cu(II) was removed by 2 mg-Fe(III)/L flocs when initial copper concentration was at 250µg/L. These observations

can be attributed to the distinct character of HAW NOM which gave higher $SUVA_{254}$ (Table 2) and contained more large MW components as evidenced in the AMW profile (Figure 2). Detailed discussion is presented in 3.5.

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The laboratory results (Figure 4A, B, C and E) were tested against known adsorption isotherms, Freundlich, Langmuir and BET. Freundlich isotherms ($\frac{Cu(II)removal}{Fe(III)Conc} = K_F \times FinalCu(II)^{1/n}$) were found to be capable of representing the adsorption of dissolved Cu (II), presumably as Cu(II)-NOM, in MRW, MCW (except 1 mg-Cu/L) and NW. Best fit parameters for the Freundlich isotherms are shown in Table 5. Adsorption phenomena generally occurred between dissolved Cu(II) and Fe(OH)₃ flocs, as shown by the well-matched Freundlich isotherm curves in Figure 4. Considering that "K_F" and "n" reflect the characteristics of adsorbent (Fe(OH)₃ flocs) and adsorbate (Cu(II)-NOM) [44], it is reasonable to see similar K_F and n values for MRW (K_F=0.003, n=1.19) and MWF (K_F=0.003, n=1.16) due to their common NOM composition. Higher initial Cu(II) concentrations were found in NW than in MRW or MWF as more Cu(II) may have been complexed with NOM in NW (Figure 4C) than in MRW or MWF. A direct comparison of Cu(II) removal between MWF and NW is given in Figure 4D. Slightly less dissolved Cu(II) was removed from NW than MWF for the case of 250 and 400 µg-Cu(II)/L initial Cu(II) concentration while Cu(II) removal was similar in both waters for the case of initial 1000 µg-Cu(II)/L. Considering the measurement error of ±20 µg/L, the difference is not significant. However, the same Freundlich isotherm parameters adopted for MRW or MWF could not be fit to NW data. Instead, different "K_F" and "n" values were calculated for NW (K_F=0.0002, n=0.73). "K_F" and "n"

values are interpreted in Freundlich Adsorption Isotherm in terms of fundamental kinetics and diffusion based properties [44]. Skopp [44] suggested that "K_F" is related to the diffusion coefficient of an adsorption-desorption dynamics and "n" reflects the probability distribution for a molecule to access adsorption sites. The different K_F and n values are believed to be related to soluble microbial products from nitrification [41, 42], which were chelated with dissolved copper and had a different character from Cu-NOM complexes formed in MRW or MWF.

Figure 4E shows the removal of dissolved Cu(II) from MCW which contains only uncoagulable NOM (NOM_{uc}). Therefore, Figure 4E can be regarded as the removal of Cu(II)-NOM_{uc}. Despite the difference in DOC concentration between MRW (2.6 mg-C/L) and MCW (0.9 mg-C/L), similar trends (MCW "n=1.17" and " K_F =0.0028") of Cu(II) removal were observed when initial Cu(II) concentration was 250 or 400 μ g-Cu(II)/L, indicating that the same type and number of Cu(II)-NOM complexes (i.e. mostly Cu(II)-NOM_{uc}) may have formed. Much less removal found in MCW than MRW for 1000 μ g-Cu(II)/L initial copper case can be explained by the effect of pre-formed Cu(II) particles.

3.4 Effect of Pre-formed Cu(II) Particles on Dissolved Cu(II) Removal

Some deviation of the experimental data from the Freundlich isotherm was observed in MRW (Figure 4A). It shows the impact of pre-formed Cu-containing particles (CuO and Cu(OH)₂) on dissolved Cu(II) removal. When copper is added to bulk water it potentially forms either particulate or stays as dissolved form. Particulate copper can be easily removed by filtering through 0.2 µm filter

paper. If the particulates are removed prior to Fe(OH) $_3$ flocs addition (MWF), in comparison with MRW, one can easily find the impact of particulates on dissolved Cu(II) removal by ferric flocs. In the case of MWF (Figure 4B), 160 µg-Cu(II)/L as Cu(II) particulates had been removed from the sample with 1000 µg-Cu(II)/L initial Cu(II) concentration. In this experiment, 350 µg-Cu(II)/L remained in MWF after 1 mg/L Fe(OH) $_3$ treatment, while 410 µg-Cu/L was measured in MRW in which these preformed copper particulates remained. This indicates that the presence of these copper particulates prevents removal of some of the dissolved Cu(II). However, one could note that the pre-filtration step had little effect on dissolved Cu(II) removal at the two lower initial Cu(II) cases (250 and 400 µg-Cu(II)/L) due to negligible Cu(II) particles formed at low initial copper cases.

This impact from the particles became obvious when initial copper concentration of 1000 μ g-Cu(II)/L was practiced in MCW (Figure 4E). Only around 350 μ g-Cu/L remained in MCW at 0 mg-Fe(III)/L addition. Reduced Cu(II) complexing sites on the lower concentration of NOM_{uc} must only have been available to complex with around 350 μ g-Cu(II)/L, with the remaining copper presumably forming precipitates such as Cu(OH)₂ and CuO_(s). When Fe(OH)₃ floc was added, the presence of this large proportion of Cu-containing particles severely interfered with dissolved Cu(II) removal. With 2 mg-Fe/L, only 60 μ g/L Cu-NOM_{uc} was removed. This dramatic change in the dissolved Cu(II) removal could be due to preferential adsorption of the large copper-based precipitates on the Fe(OH)₃ flocs.

3.5 Cu(II)-NOM Chelation and Its Impact on Cu(II)-NOM Removal by Fe(OH)₃ Flocs

During the course of the experiments on dissolved Cu(II) removal by Fe(OH)₃ flocs, the analysis of AMW distributions were undertaken on MRW and HAW. The samples were respective filtrates collected 24 hours after the copper (400 µg-Cu(II)/L) addition, and 4 hours after the addition of Fe(OH)₃ floc (0.5 mg-Fe/L) (Figure 5A and B, respectively). The DOC concentration and UV₂₅₄ absorbance were also measured after copper dosing. The DOC concentration of MRW and HAW remained unchanged before and after copper dosing. However, the UV₂₅₄ absorbance increased in both MRW and HAW after copper dosing (Figure 6A and B, respectively). In the MRW samples, addition of copper did not change the DOC concentration of the water samples (DOC concentration of 2.6±0.1 mg/L before and after copper addition). However, a concentration of 400 µg/L copper increased the UV₂₅₄ absorbance of MRW from 0.036 cm⁻¹ to 0.046 cm⁻¹ (Figure 6A), equivalent to a 28% increase in relative UV₂₅₄ absorbance or SUVA₂₅₄ (since the DOC remained unchanged) (Figure 6B). Apart from the SUVA₂₅₄ increase, the addition of copper slightly shifted the distribution of MW to a higher range (2000-4000 Da) (Figure 5A). The change in both SUVA₂₅₄ and AMW distribution reflected a restructuring of the DOC after complexing or chelating with copper ions, perhaps indicating a change in the type of organic matter from hydrophilic to hydrophobic through intermolecular bidentate chelation between smaller NOM molecules. The removal of Cu-NOM by 0.5 mg-Fe/L Fe(OH)₃ was also concentrated on a narrow spectrum between 2500 Da and 3500 Da. According to our previous work [17], the hydrophobic fraction of NOM, which is generally rich in phenolic chromophores, is depleted in Mundaring water. Therefore, the change of

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AMW distribution after Fe(OH)₃ adsorption provided another evidence showing that the dosed copper in MRW chelated with hydrophilic fraction and converted them to hydrophobic molecules which were readily removed by the flocs.

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By contrast, only 4.7% increase in UV₂₅₄ absorbance was measured in HAW when the initial copper concentration was 400 µg/L (Figure 6B) and the DOC concentration remained unchanged. In addition, little change in the AMW profile of DOC detected by UV₂₅₄ in HAW was caused by copper dosing (Figure 5B) since the DOC which was chelated with copper only accounted for a small proportion of the whole UV₂₅₄-detected DOC in HAW. However, after the addition of Fe(OH)₃ flocs, compared with the corresponding situation in MRW, the DOC detected by UV₂₅₄ in HAW was considerably reduced over a more extensive AMW range (2000-6000 Da), indicating that Fe(OH)₃ flocs removed the components of large MW in HAW, since coagulation with ferric salts is believed to readily and preferentially remove hydrophobic fraction mainly containing aquatic humic material with high MW [33]. In addition, little dissolved Cu(II) removal by 0.5 mg/L Fe(OH)₃ as discussed previously (Figure 4F) again indicated that dosed Cu(II) preferentially chelated with small MW components or the hydrophilic fraction in HAW and consequently was shielded from Fe(OH)₃ adsorption by large MW components or the hydrophobic fraction.

Gamble et al. [13] reported two general types of bidentate chelating sites for dissolved Cu(II): a salicylate type and a dicarboxylate type (Figure 7). The dicarboxylate type could be both intramolecular and intermolecular, depending on the functional groups available. According to the shift in UV_{254} -

detected DOC in MRW to higher AMW upon addition of copper(II), intermolecular dicarboxylate chelation may be mainly responsible for copper binding in MRW, resulting in the aggregation of small molecules, while salicylate chelation likely occurred in HAW in which the NOM may contain more salicylate type binding sites. In either MRW or HAW, copper preferentially chelated with small organic molecules or the hydrophilic fraction and converted them (or part of them) to hydrophobic complexes.

4. CONCLUSIONS

Soluble Cu(II) ions used to inhibit nitrification in natural waters mainly exist in the form of various organic Cu(II) complexes. Acting as a bridging ion, copper appears to have the capacity to aggregate smaller organic molecules to form larger organic molecules via intermolecular complexation. Based on the evidence provided by the changes in SUVA₂₅₄ and AMW distribution profiles of different NOM-containing waters, Cu(II) ions are believed to preferentially form complexes with small molecules, rather than large molecules. Trace ferric hydroxide flocs, which could be naturally present or released from aging or corroded iron pipes, demonstrated varying capacity of removing Cu-NOM subject to different aqueous conditions. Organic Cu(II) removal by ferric hydroxide flocs in Mundaring water was found to be governed by an adsorption process obeying a Freundlich isotherm, while less organic Cu(II) removal was observed in a solution of commercial humic acid due to the predominance of large MW DOC in the humic acid, which shields small organic Cu(II) complexes from adsorption. Insoluble

copper species (e.g CuO_(s) and Cu(OH)_{2(s)}) can affect adsorption of dissolved copper on ferric hydroxide flocs to differing extents, depending on their relative proportion.

For the strategy of inhibition of nitrification in distribution systems by the addition of Cu(II) ions, the impact of the character of NOM in the distributed water on the fate of dissolved copper ions is therefore of crucial importance. Iron pipe corrosion offers a potential explanation for the significant loss of Cu(II) in the distribution system, in comparison to the successful pilot-scale reservoir study where release of ferric hydroxide flocs would have been minimal.

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SUPPORTING INFORMATION AVAILABLE

Details of Fe(OH)₃ suspension preparation and dynamic process of Cu(II) removal by Fe(OH)₃ flocs.

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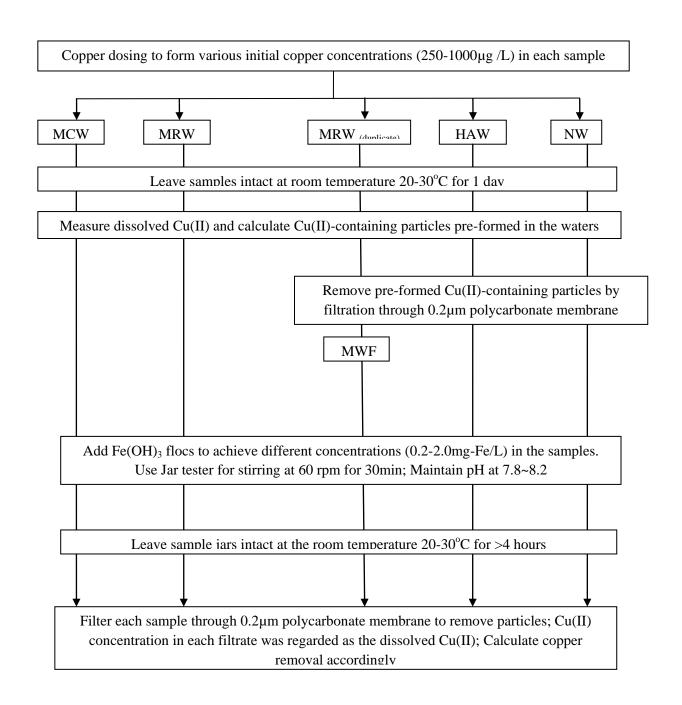


Figure 1: Flowchart of Cu(II) Removal Experiment Procedure

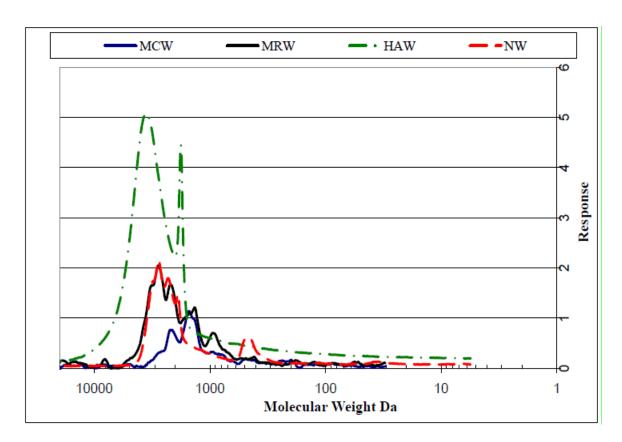


Figure 2: Apparent Molecular Weight Distributions Detected by UV_{254} in MRW, MCW, HAW and NW Bulk Water Samples

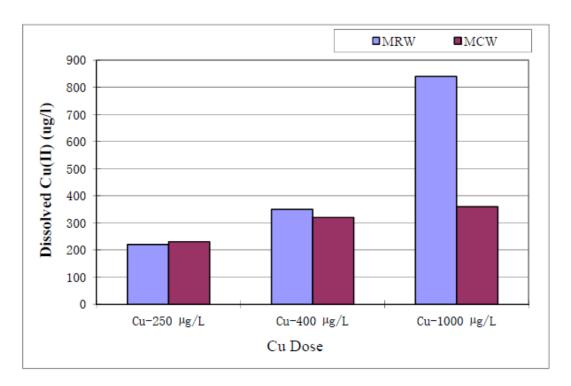


Figure 3: Copper(II) Solubility in MRW and MCW at Various Initial Copper Concentrations (Dissolved copper was measured 24 hours after copper dose) Measurement error in copper concentrations is $\pm 20 \mu g$ -Cu(II)/L.

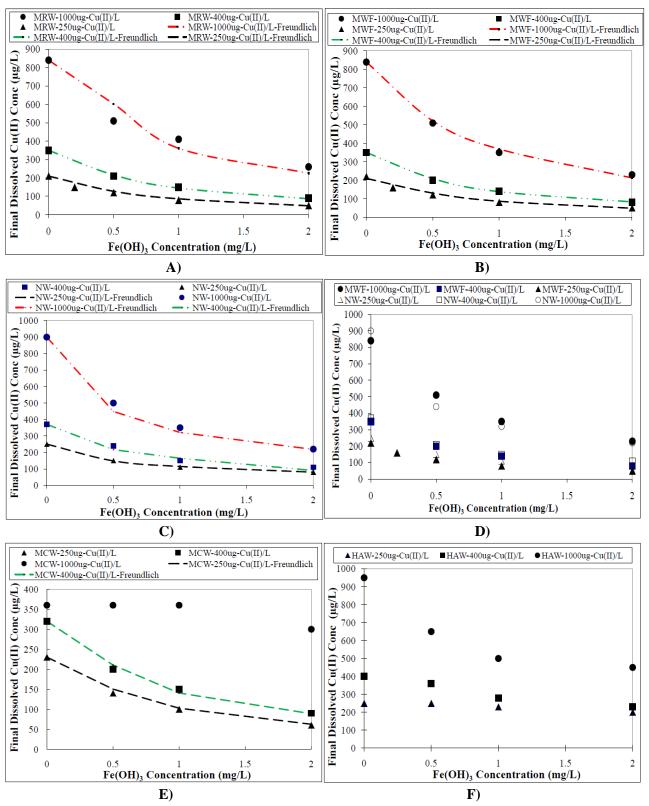


Figure 4: Dissolved Cu(II) concentrations remaining in bulk waters 4 hours after addition of Fe(OH)₃ flocs dosed to reach concentrations as marked in x-axis. A) MRW; B) MWF; C) NW; D) Comparison of dissolved Cu(II) removal between MWF and NW; E) MCW; F) HAW; In MWF, NW and HAW Fe(OH)₃ was dosed after filtering particulates formed by copper addition in the previous step.

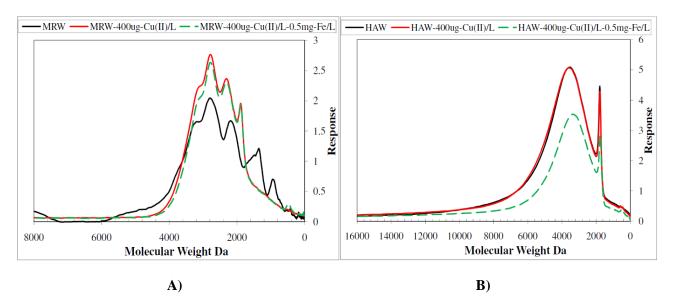


Figure 5: AMW distributions of UV_{254} -detected DOC in A) MRW, in the MRW filtrate 24 hours after addition of Cu(II) (400 μ g-Cu(II)/L) and in the MRW filtrate 4 hours following further addition of $Fe(OH)_3$ floc (0.5mg-Fe/L); and B) HAW, in the HAW filtrate 24 hours after addition of Cu(II) (400 μ g-Cu(II)/L) and in the HAW filtrate 4 hours following further addition of $Fe(OH)_3$ floc (0.5mg-Fe/L).

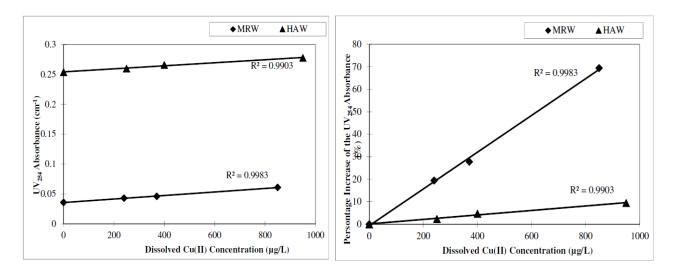


Figure 6: Change of UV_{254} Absorbance Vs Copper Concentration A) Absolute UV_{254} Abs Vs Copper concentration B) Relative Change (%) of UV_{254} Absorbance Vs Copper Concentration

Salicylate chelation

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O \\
R'
\end{array}$$
 $\begin{array}{c}
O \\
C \\
C \\
O \\
\end{array}$

Dicarboxylate chelation

Figure 7: Copper Chelation with Two Types of Bidentate Chelating Sites

Table 1: Potential Redox Reactions Considered in G&AWSS (Snoeyink and Jenkins, 1980)

Reaction	Standard Electrode Potentials at 25°C (Volt)		
$O_2(ag) + 4H^+ + 4e^- = 2H_2O$	+1.27		
$NO_3^- + 2H^+ + 2e^- = NO_2^- + H_2O$	+0.84		
$Cu^{2+} + 2e^{-} = Cu_{(s)}$	+0.34		
$Fe^{2+} + 2e^{-} = Fe_{(s)}$	-0.44		

Table 2: Water Quality Characteristics of the Bulk Water Samples

Water Samples	рН	UV_{254}	DOC	SUVA	Total concentration of dissolved Fe(II) or Fe(III)	
	-	cm ⁻¹	mg-C/L	$L \cdot mg^{-1} \cdot m^{-1}$	mg/L	
MRW	7.9±0.2	0.035±0.002	2.6±0.1	1.35	< 0.02	
MCW	7.9±0.2	0.010±0.002	0.90±0.1	1.11	< 0.05	
HAW	7.9±0.2	0.248±0.002	2.4±0.1	10.33	< 0.02	
NW	7.9±0.2	0.026±0.002	2.5±0.1	1.04	< 0.02	

Table 3: The NOM Fraction(s) Dominating in Water Samples (Derived from Figure 1)

Water Samples	AMW Range (Da)	Major NOM Fraction
MCW	1000~2000	Uncoagulable
MRW	1000~3000	Coagulable + uncoagulable
HAW	2000~10000	More aromatic groups
NW	~500 + 2000~3000	Similar to MRW + smaller molecules produced through nitrification

Table 4: The Major Cu(II) Species under Various Aqueous Conditions after Dosing an Aqueous CuSO₄ Solution into Each Bulk Water to Achieve An Initial Concentration of 1000 μ g-Cu(II)/L

Bulk Water Conditions	Intermediate ¹ / Equilibrated ² Copper Species	MINEQL® Calculation	Laboratory Data
Milli-Q water, pH=6.3±0.1, Open system	Cu ²⁺	Cu ²⁺ =1 mg/L	Dissolved copper was measured at 0.98 mg/L 1 day after Cu dose
Milli-Q water, pH=7.5±0.1, Open system	Cu(OH) ₂ / CuO	Cu ²⁺ =3 μg/L + CuO _(s)	Dissolved copper (<20 µg/L) was measured both 2 hrs and 2 days after Cu dose
CaCO ₃ buffered solution, pH=7.5±0.1, Open system	CuCO ₃ °, CuOH ⁺ and Cu(OH) ₂ / only CuO	Total soluble Cu=9 μg/L + CuO _(s)	100 µg/L and 10 µg/L dissolved Cu were found 2 hrs and 1 day after Cu dose respectively, the latter occurs when the system is in equilibrium with CuO
CaCO ₃ buffered solution, pH=7.5±0.1, Closed system	Cu ²⁺ , CuCO ₃ °, CuOH ⁺ and Cu(OH) ₂	$Cu^{2+}=30 \mu g/L,$ $CuOH^{+}=30 \mu g/L,$ $CuCO_{3}^{\circ}=48\mu g/L$ $+Cu(OH)_{2(s)}$	150 μg/L dissolved Cu was measured 4 hours after Cu dose. The system may be in equilibrium with Cu(OH) _{2(s)}
MRW pH=7.9±0.1, DOC=2.6 mg/L, Open system	CuCO ₃ °, CuOH ⁺ , Cu(OH) ₂ and Cu- NOM / CuO and Cu-NOM	*Dependant on availability of binding ligands of MRW NOM	950 µg/L and 840 µg/L dissolved Cu was found 2 hrs and 1 day after Cu dose respectively. The latter occurs when the system reaches equilibrium with both CuO and Cu-NOM complexes

Note:

Intermediate¹ Copper Forms: possible Cu species existing in bulk water only 2 hours after dosing CuSO₄.

Equilibrated² Copper Forms: possible Cu species existing in bulk water 1 day after copper dose

Table 5: The Parameters Adapted in Freundlich Adsorption Isotherms

Variables and Constants	Valid Cu(II) Concentration	\mathbf{K}_{F}	n	R^2
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^{*}Cu-NOM chelation coefficient is dependent on a series of binding ligands varying in different NOM-containing water samples. They were not calculated by MINEQL in this study. Alternatively, the proportion of Cu-NOM was deducted from the difference between the total dissolved Cu(II) and the sum of inorganic copper compounds and free cupric ions.

	Range			
Units	μg/L			
MRW	200~1000	0.003	1.19	0.98
MWF	200~1000	0.003	1.16	0.99
MCW	200~400	0.003	1.17	0.99
NW	200~1000	0.0002	0.73	0.98

Note: Cuo: dissolved Cu(II) concentration at 0 mg-Fe(III)/L addition

C: Final dissolved Cu(II) concentration after ferric hydroxide flocs treatment

Freundlich isotherm: $Y = K_F C^{1/n}$

R²: coefficient of determination