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1	δD and $\delta^{13}C$ analyses of atmospheric volatile
2	organic compounds by thermal desorption gas
3	chromatography isotope ratio mass
4	spectrometry
5	
6	Authors:
7	Christiane Vitzthum von Eckstaedt ^a *
8	Kliti Grice ^b
9	Marisa Ioppolo-Armanios ^c
10	Geoff Chidlow ^d
11	Mark Jones ^e
12	
13	^a WA-Organic and Isotope Geochemistry Centre, The Institute for Geoscience Research, Department
14	of Chemistry, Curtin University, GPO Box U1987,- Perth -WA 6845,- Australia.
15	C.VitzthumVonEckstaedt@curtin.edu.au ; Tel: +61(0) 8 9266 7628 ; Fax: +61(0) 8 9266 2300
16	
17	^b WA-Organic and Isotope Geochemistry Centre, The Institute for Geoscience Research, Department
18	of Chemistry, Curtin University, GPO Box U1987, -Perth -WA 6845, -Australia.
19	K.Grice@curtin.edu.au; Tel: +61(0) 8 9266 2474-; -Fax: +61(0) 8 9266 2300
20	
21	^c Alcoa World Alumina, Australia, Technology Delivery Group, PO Box 161, Kwinana -WA 6167,
22	Australia. Marisa.Ioppolo-Armanios@alcoa.com.au
23	
24	^a WA-Organic and Isotope Geochemistry Centre, The Institute for Geoscience Research, Department
25	of Chemistry, Curtin University, GPO Box U1987, -Perth- WA 6845,- Australia.
26	G.Chidlow@curtin.edu.au
27	
28	Alcoa World Alumina, Australia, Technology Delivery Group, PO Box 161, Kwinana -WA 6167,
29	Austrana. Mark.Jones@alcoa.com.au

30 Contact author (during the review process):

von

Eckstaedt: C.v.Eckstaedt@gmx.de

33	Corresponding	Author (after publicatio	n):
32	C.VitzthumV	onEcksta	aedt@curtin.edu	ı.au
31	Christiane	D.	Vitzthum	,

Kliti Grice: K.Grice@curtin.edu.au

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Abstract 36

This paper describes the establishment of a robust method to determine compound specific δD and 37 δ^{13} C values of volatile organic compounds (VOCs) in a standard mixture ranging between C₆ - C₁₀-38 and was applied to various complex emission samples, e.g. from biomass combustion and car exhaust. 39 40 This was achieved by linking A thermal desorption (TD) unit was linked to a and gas chromatography 41 isotope ratio mass spectrometer (TD-GC-irMS), to enable compound specific isotope analysis (CSIA) of gaseous samples. Samples were adsorbed on TenaxTA was used as an adsorbent 42 43 material in-using stainless steel TD tubes. We determined instrument settings to achieve a minimal 44 water background level for reliable δD analysis and investigated the impact of storage time on δD and 45 δ^{13} C values of collected VOCs (176 days and 40 days of storage, respectively). Most of the standard compounds investigated showed standard deviations (SD) < 6 % (δ D) when stored for 148 days at 46 4°C. However, benzene revealed occasionally depleted 8D depleted values (21 % SD) for unknown 47 reasons. $\delta^{13}C$ analysis demonstrated that storage of 40 days had no effect on VOCs investigated. We 48 also showed that breakthrough (benzene and toluene, 37 % and 7 %, respectively) had only a 49 negligible effect (0.7 ‰ and 0.4 ‰, respectively) on δ^{13} C values of VOCs on the sample tube. We 50 established that the sample portion collected at the split flow effluent of the TD unit can be used as a 51 replicate sample for isotope analysis saving valuable sampling time and resources. We also applied 52 TD-GC-irMS to different emission samples (biomass combustion, petrol and diesel car engines 53 54 exhaust) and for the first time δD values of atmospheric VOCs in the above range are reported. Significant differences in δD of up to 130 % were observed between VOCs in emissions from petrol 55 56 car engine exhaust and biomass combustion (Karri tree). However, diesel car emissions showed a 57 high content of highly complex unresolved mixtures thus a baseline separation of VOCs was not

58 achieved for stable hydrogen isotope analysis. The ability to analyse δD by TD-GC-irMS

59 complements the characterisation of atmospheric VOCs and is maybe used for establishing further

60 source(s).

61 Keywords

62 volatile organic compounds

- 63 compound specific isotope analysis
- 64 BTEX
- 65 stable hydrogen isotopes
- 66 thermal desorption
- 67 active solid sorbent sampling

68 **1. Introduction**

Volatile organic compounds (VOCs) are ubiquitous in the environment (e.g. in soil, water and in the 69 atmosphere) emitted by anthropogenic or natural sources. The technique of compound specific isotope 70 71 analysis (CSIA) of organic compounds achieved by gas chromatography isotope ratio mass spectrometry (GC-irMS) has been often used in the fields of organic geochemistry, in food and in 72 73 forensic investigations (including the provenance of food and drugs) [1-4]. Techniques are currently available to measure $\delta^{13}C$ and δD of natural gas mixtures (e.g. methane to pentane), liquid 74 75 hydrocarbons (e.g. crude oil) and extracts from complex media (e.g. soil, sediments and modern 76 biological material) [5-8]. Thus, CSIA is useful to establish sources, processes, thermal history and 77 fractionations that occur by kinetic and exchange reactions. The analysis of atmospheric VOCs was previously mainly restricted to δ^{13} C analysis [9-12]. A technique for measuring δ D for a range of 78 79 VOCs in emissions to establish their sources is not currently available. Therefore this study 80 investigates a method to reliably measure the stable hydrogen isotope compositions of several VOCs 81 without significant isotopic fractionation. We combined a successful established technique for collecting VOCs (thermal desorption) with traditional gas chromatography isotope ratio mass 82

83	spectrometry (GC-irMS). Active solid sampling and thermal desorption (TD) in combination with	
84	TenaxTA as an adsorbent material are oftenregularly used for sampling VOCs [13-156]. The	
85	advantages of using TenaxTA and a TD unit with a split flow collection device (Markes Int. Ltd.)	
86	included This sampling method has the advantages of (i) being selectiveity for compounds of interest	
87	$(C_6 - C_{10})$, (ii) <u>its</u> economical maintenance <u>and</u> (iii) <u>the</u> convenient sample handling <u>in contrast to e.g.</u>	
88	whole air sampling procedures [16]. Previous studies could already demonstrate for stable carbon	
89	isotope analysis that TD-GC-irMS is a reliable technique to analyse $\delta_{\underline{13}}^{\underline{13}}$ C of some lowmolecular-	Formatted: Font color: Auto, Superscript
90	weight VOCs [17,19].	Catacate
91	and (iv) obtaining a replicate sample for further analyses. Previous studies could already demonstrate	
92	that TD-GC irMS is a reliable technique to analyse δ^{13} C of low molecular weight VOCs [17,18].	
93	We investigated the suitability of TD-GC-irMS for analysing compound specific δD in addition to	
94	expanding the range of VOCs (C_6 - C_{10}) for $\delta^{13}C$ and $\delta^{43}C$ of atmospheric VOCs ranging between C_6 -	
95	C_{10} -using TenaxTA as an adsorbent. Essential aspects of the technique were evaluated, e.g. influence	
96	of background water, sample breakthrough (BT) and storage behaviour. We also established that	
97	recollecting the split flow is-does not causing cause isotopic fractionation and thus can be used as a	
98	replicated sample for δD and $\delta^{13}C$ analyses. The application to various emissions (e.g. biomass	
99	combustion and car exhaust emissions) confirmed the robustness of TD-GC-irMS for stable isotope	
100	analysis.	

- 101 **2. Experimental section**
- 102 **2.1.** Chemicals, Materials and Gases

103 Standard mixtures contained VOCs ranging from $C_6 - C_{10}$. Compounds (benzene, toluene, 104 ethylbenzene, styrene, *m*-xylene, *o*-xylene, *p*-xylene, *n*-propylbenzene, cumene, 1,2,4-105 trimethylbenzene, butylbenzene) were purchased from Sigma Aldrich and Fluka with puritie 99.5 106 %. Standard mixtures were prepared gravimetrically in *n*-pentane (A.R.). *n*-Pentane was found to be 107 the most suitable solvent for the analyses of investigated VOCs by GC-MS and GC-irMS. Five 108 different standard concentrations were prepared for TD-GC-MS calibration (230 ng/ μ L, 170 ng/ μ L, 109 115 ng/ μ L and 34 ng/ μ L) from a stock solution of 340 ng/ μ L. For analyses by GC-irMS (i.e. without 110 a TD unit) standard solutions of 250 ng/ μ L were used. For analyses by TD-GC-irMS solutions

111 consisting of ca. 520 ng of each compound per μL were prepared.

112 Commercially available adsorbent materials for TD are suitable for a wide range of compounds and 113 different fields of application [14]. For the present study we chose TenaxTA due to its highly 114 hydrophobic characteristics and its selectivity for VOCs of interest (C_6 to C_{10} , boiling points between 115 80_°C and 220_°C). TenaxTA is a porous polymer resin on the basis of 2,6-diphenylene oxide and was 116 purchased from Agilent Technologies in mesh 60/80.

In this paper we compared the chromatographic performance of the VOCs on two GC stationary
phases (i) a polyethylene glycol (DB-WAXetr) and (ii) a non-polar phenyl-arylene polymer (DB5MS). Both columns were supplied from Agilent J&W with dimensions of 60 m length x 0.25 mm i.d.
and a 0.25 μm film thickness.

121 Reference gases (H₂ and CO₂) for CSIA stable isotope analyses were purchased from BOC Gases
122 Australia Ltd. in ultra high purity (99.999 %).

123

2.2. Preparation of the TD tubes

Tubes were manually packed with approximately 280 mg of TenaxTA with minimal compression.
Backpressure tests for each TD tube confirmed that packing was consistent (approximately -1.2 kPa).
TenaxTA was conditioned at 330_°C (4 hours) while a constant flow of pure helium (50 mL/min) was
passed through the tubes [2019].

Standard TD tubes were prepared in accordance to method TO-17 [2049]. An aliquot of 2 μL - 4 μL
of the standard mix was injected directly onto the sorbent bed while pumping air through the tube for
1 min at 50 mlmL/min. It is important to note that the TD-tube needs to be positioned vertically
during injection and sampling to avoid possible breakthrough_BT_by channelling. Thus a
breakthrough_BT_experiment was also performed (see 3.5). A standard solution of high concentration
(i.e. 800 ng per compound/µL) was injected (3 µL) onto a sample tube using a Calibration Solution
Loading Rig (CSLRTM) [2120]. The CSLRTM consisted of an unheated injector port with a controlled

carrier gas supply (i.e. helium). The standard mix was introduced through the injector septum using a
standard GC syringe. A helium flow of 200 mL/min was swept through the injection port for 40 min.
The solution vaporised in the flow of gas before it reached the sorbent bed in the TD tube.

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2.3. Collection of Emissions

Three different emissions were collected to investigate the applicability of TD-GC-irMS on real samples (biomass combustion, petrol and diesel car engine's exhaust). All samples were actively drawn through TenaxTA sample tubes using a compact Gilian LFS hand pump. Sampling volume was depending on the emission concentration to gain a sufficient amount of VOCs for stable isotope analysis. Sampling flow was adjusted accordingly but in the recommended range for active solid sampling [22,23]

(i) For the petrol engine exhaust <u>emissions emissions</u> samples were taken from a 1993 Mitsubishi
Magna Executive running on unleaded petrol (ULP). A sampling volume of 500 mL per sampling
tube was collected (at 50 mL/min).

(ii) For the diesel engine exhaust emissions samples were taken from a 2008 Toyota Land Cruiser
GXL. A sampling volume of 3L per sampling tube was collected (at 200 mL/min).

(iii) For emission sampling from biomass combustion approximately 1kg of Karri biomass
(*eucalyptus diversicolor*) was burnt in a 150 L drum with sufficient oxygen supply for a complete
combustion. A sampling volume of 3 L per sampling tube was collected (at 200 mL/min).

153 2.4. Instrumentation

154 Compound specific δD and $\delta^{13}C$ analyses of atmospheric VOCs were achieved by linking two 155 established techniques (i) thermal desorption (to collect and pre-concentrate the sample) and (ii) GC-156 irMS (to separate the compounds in the sample mixture and allowing accurate δD and $\delta^{13}C$ 157 measurements).

In addition to isotope analysis conventional TD-GC-MS analysis was used for method development,
 especially for compound identification in the samples and for evaluation of most suitable GC settings.

160 2.4.1 Thermal Desorption (TD) Unit

The TD unit (UNITY2TM, Markes International Limited) used was a single tube, two-stage desorber 161 162 (primary sample tube desorber and cold trap desorber). The desorption oven was suitable for stainless steel sample tubes (0.635 cm O.D. and 8.89 cm length). The installed cold trap contained also the 163 adsorbent material and was cooled by a 2-stage peltier cell to temperatures ranging between -10 °C 164 165 and 30_°C. The first step of the analyses was the thermal desorption of the sample tube at 300 °C for 5 min andat an adjustable but constant helium flow (see Table 1). transported the The desorbed 166 compounds were trapped onto the cold trap (at 10°C). In a second step the refocused sample was 167 rapidly desorbed from the cold trap at 100 °C/s to 300 °C and held for 1min. and The sample was then 168 169 transferred onto the GC-column through a heated deactivated fused-silica capillary (120°C) in thea helium carrier gas through a heated deactivated fused silica capillary (120°C) onto the GC column 170 171 (Fig. 1). The instant desorption process focused the sample on the GC-column to improve the 172 chromatographic resolution. The helium flow during trap desorption represents the sum of GC helium flow and split flow (when required) but needs a minimum flow of 2 mL/min for sufficient desorption. 173 174 Prior to each sample desorption the TD unit performed an automated leak test and sample tube and 175 cold trap were purged for 2 min and 1 min, respectively using helium to remove any residual oxygen. The purge flow used was equivalent to the desorption flow (Table 1). Sample tube and the subsequent 176 177 cold trap were desorbed at 300°C for 5 min and for 1min, respectively. During both desorption steps it 178 was possible to divert a split flow from the sample flow as required depending on the sample concentration. The actual split ratio is achieved by adjusting the volume of the split flow. A special 179 feature offered by UNITY2TM is a device for quantitative re-collection of the split flow on a separate 180 181 sample tube (SecureTD-Q) (Fig. 1). Thus the split sample provides a replicate of the injected sample. Detailed information about general instrument operation of the UNITY2TM is provided by Bates et al. 182 [<u>18</u>21]. 183

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184 2.4.2 Gas Chromatography - Mass Spectrometry (GC-MS)

The method development for TD-GC-MS analysis was carried out on an HP6890 GC linked to an HP5973 MS detector. The GC oven temperature was held isothermally at 50°C for 5 min and heated at 5°C/min to 80°C, held for 5 min and heated again at 12°C/min to a final temperature of 220°C for 2 min. The GC had to be set to splitless and constant pressure mode (108.3 kPa). Peak identification was performed in scan mode using m/z between 18 and 200.

190 2.4.3 Gas Chromatography - isotope ratio Mass Spectrometry (GC-irMS)

Compound specific isotope analysis (CSIA)_-was performed on a Micromass IsoPrime irMS 191 192 interfaced with an HP6890 GC, programmed with the same temperature settings as described above 193 for GC-MS analysis. An auto-sampler (HP6890) was used for direct injections of the sample mix 194 (split/splitless injector) in pulsed-splitless mode. Isotopic compositions of sample components are 195 given in the delta (δ) notation in per mil (∞) relative to the Vienna Standard Mean Ocean Water 196 (VSMOW) (hydrogen) or to the VPDB carbonate standard (carbon) (Equation 1). A reference gas 197 pulse of a known D/H or ${}^{13}C/{}^{12}C$ content was introduced into the isotope mass spectrometer at a time 198 different to the analyses. Stable isotopic ratios are reported relative to that of the reference gas which 199 was determined daily using organic standards (with known δ values) to monitor the instrument's 200 performance and reliability of the results.

201
$$\delta_{\text{sample}} = \left[\frac{R_{\text{sample}} - R_{\text{standard}}}{R_{\text{standard}}}\right] * 1000$$
 (1)

- 202 R is the ratio of heavy to light isotope (D/H, $^{13}C/^{12}C$)
- 203

The basic principles of δD analysis are shown in Fig. 2 and relevant calculations are previously described elsewhere [24-2722-25]. In brief, GC chromatographically separated compounds are pyrolysed at 1050 °C to H₂ +C +CO in a quartz furnace containing a sieved chromium catalyst. δD values were calculated by integrating the peaks of resulting ion currents of masses 2 and 3 (H₂ and HD). The results of the H³⁺ corrections (performed after Sessions et al. [2826]) were automatically included in software calculations. δ^{13} C values were calculated by integration of the masses 44, 45 and 46 ion currents of the peaks 211 produced by combustion (CuO reactor at 850°C) of the chromatographically separated compounds to $CO_2 + H_2O$ (see Fig. 3). The generated water was trapped at -100_°C to avoid the interference of HCO_2^+ (*m*/*z* 45).

For samples, average values of at least three analyses are reported and standard deviations (SD) are given (Table 4). For direct injections an internal standard mix of five compounds (*n*-undecane, *n*tridecane, *n*-tetradecane, *n*-heptadecane, *n*-octadecane, *n*-nonadecane, pentacosane) with known δD and $\delta^{13}C$ values was analysed after every 10th sample to monitor accuracy and precision of δ measurements. During the measurements, SDs of the internal standard for analyses were typically less than 4 ‰ (δD) and less than 0.3 ‰ ($\delta^{13}C$).

3. Results and Discussion

221 **3.1. Background Water**

The impact of water on results obtained by TD and the use of active solid sampling in its many forms 222 have been were previously discussed [22, 2927,28]. However, most published dataresearch refers to a 223 224 scan range above m/z 30 excluding the detection of water during measurements. The presence of water anywhere in the system will have especially an impact on δD analyses since pyrolysis will 225 transfer water to hydrogen gas (H₂, DH)-would be produced by pyrolysis. Therefore we monitored m/z226 227 ion 18 during TD-GC-MS preliminary tests on two different GC-columns (DB-5MS and DB-Waxetr) and noted an increased background and an additional peak in contrast to GC-MS analysis (i.e. without 228 229 TD unit). The mass spectrum indicated that the component was water. The occurrence of the peak and background elevation during D/H analysis using TD-GC-irMS verified the hydrogen content. Tests 230 231 with packed (adsorbent without a sample) and empty (unpacked) TD tubes showed no difference of 232 the background level. Efforts were made to minimise the water content by using a highly hydrophobic 233 adsorbent material (TenaxTA) and following the TD unit's water restriction measures as well as 234 applying standardised sampling methods (e.g. guidelines from U.S. EPA [2019]). Thorough leak

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235 checks on the TD-unit and all connections to the GC were carried out and various instrument 236 parameters were investigated. Despite these efforts, the elevated background level and peak due to water was still observed but the action taken reduced the water intake sufficiently for reliable results, 237 238 e.g. we found that the GC oven temperatures described above enabled the separation of toluene from the water peak (Fig. 4b). We we found also that by using cold trap temperatures of 10°C and by 239 240 maintaining a split flow of > $1_{\underline{L}}/min$ during trap desorption the background water was 241 significantly reduced (Fig. 4) allowing accurate δD measurements (see 3.3). Additionally the 242 determined GC oven temperatures (see 2.4.2) enabled the separation of toluene from the remaining 243 water peak (Fig. 4b).

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3.2. Comparison of GC-columns

The performance of a DB-Waxetr and a DB-5MS GC column was investigated for VOCs of the standard mixture analysed by TD-GC-MS. The DB-5MS is known to be less prone to damage by oxygen and generally has less column-bleed compared to the DB-Waxetr column. The advantage of the DB-Waxetr column over the DB-5MS is its higher polarity and thus its ability to separate *p*- and *m*-xylene isomers. For this study the column bleed had negligible impact on the compounds investigated. Therefore the DB-Waxetr was chosen for further analysis due to the efficient separation of all compounds of interest.

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3.3. Isotopic Fractionation

Previous work established that TD does not cause isotopic fractionation for δ^{13} C analysis of selected VOCs (C₂ - C₉) [17,<u>19</u>],<u>[18]</u>. In this study we investigated if any isotopic fractionation occurred for VOCs of the standard mixture during compound specific δ D analysis using TD. Therefore results achieved by TD-GC-irMS were compared to those obtained by GC-irMS (direct injection) analyses. The specifications of the irMS required sample sizes of >250 ng per compound on the GC column to measure δ D in the limits of instrument linearity. Due to the essential split flow setting of >1 mL¹/-min during trap desorption (see water restriction measures in <u>3.13.1</u>) samples in order >500 ng per compound needed to be collected on the adsorbent material. We could show repeatable results with SDs between 1-5 ‰ (average of 2‰). δ D values of most investigated compounds introduced by TD are slightly heavier (on average 4 ‰) compared to δ D values obtained by direct injection. However, the differences are still in range of instrument error (with the exception of 1,2,4-trimethylbenzene having a 11‰ difference between the two techniques) and therefore are deemed negligible. Thus we have shown that TD-GC-irMS is a reliable method for obtaining δ D results of VOCs ranging between C₆-C₁₀.

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3.4. Impact of Storage Time on δD and $\delta^{13}C$ of VOCs

The impact of storage time on δD and $\delta^{13}C$ values of VOCs was investigated for the VOCs in the 268 standard mix adsorbed on TenaxTA. For this experiment all All desorption tubes were stored at 4_°C 269 in an air tight jar containing a vial of activated charcoal. δ^{13} C results were determined after 8, 14, 16 270 and 40 days of storage and showed showing negligible differences (SD between 0.1 and 0.3 ‰) for all 271 VOCs (Table 3)Therefore sampling tubes for δ^{13} C measurements can be safely stored for 40 days. δ D 272 273 values were obtained during 176 days of storage (i.e. after 2, 4, 8, 24, 32, 50, 72, 240 hours and after 274 148 and 176 days) (Table 2). SDs of δD analyses were negligible for most compounds of the standard 275 mixture were negligible ranging between 3 - 6 ‰ during the first 148 days of storage. (with the 276 exception of 21 % for benzene). However, results obtained after 176 days of storage showed 277 generally more depleted δD values with differences of up to 44 % compared to between δD values 278 obtained on day 0. and day 176. As noted above benzene behaved differently. The acquired δD values 279 of benzene showed dissimilarities to other VOCs during the first 146 days of storage with a SD of 21 $\frac{9}{100}$, $\frac{9}{100}$, $\frac{9}{100}$, $\frac{1}{100}$, $\frac{1$ 280 0). (SD of 21 ‰) but itsThe variations in benzene's δD values show no clear trend with storage time 281 282 (Table 2), The majority of benzene's oD values vary less than 3 ‰ to the reference oD value (at day 0). Hhowever, a random volatility effect cannot be excluded [3029] possibly due to the lowest boiling 283 point among the VOCs of the standard mixture. Benzene could have been also subjected to hydrogen 284 exchange with a depleted hydrogen source. Although the exact cause is unclear; therefore it is 285

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286 suggested to use multiple sampling tubes for real samples to identify occasional shifts in δD values for

287 benzene during storage.

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3.5. Effects of Breakthrough

BT can occur during active solid sampling due to-e.g. specific sampling conditions or the nature of the 290 291 real sample [3130]. Therefore it is important to monitor any possible BT by installing a BT tube just 292 after the sample tube...-in the sampling train. In this study weWe evaluated whether any carbon 293 isotopic fractionation occurred during BT by analysing the standard mixture. An aliquot of the standard (3 µL) was introduced onto the sampling tube using a CSLRTM. The sampling tube and BT 294 295 tube were linked with a brass connector and purged with a helium flow of 200 mL/min (40 min) to 296 allow for BT of benzene and toluene based on parameters from the Health and Safety Executive regulations, UK [3231]. From this experiment it was establishedWe found that 37 % of benzene and 7 297 298 % of toluene broke through. Other compounds in the standard mix did not show any significant BT. Fig. 5 illustrates the δ^{13} C values of benzene and toluene obtained from the sample tube, from the BT 299 tube and from a reference analysis by TD-GC-irMS (sampled without BT occurrence) during standard 300 301 injection. The δ values from the BT tube were less depleted in ¹³C by 1.6 ‰ (benzene) and 3.3 ‰ (toluene) compared to those from the sample tube. However, comparing δ^{13} C of benzene and toluene 302 the results from sample tubes where BT occurred to those without BT δ^{13} C values showed no 303 significant differences in δ^{13} C-(0.7 ‰ and 0.4 ‰ for benzene and toluene, respectively) (Fig. 5). 304 305 Therefore it can be said that the impact of BT of benzene and toluene on their δ^{13} C is negligible. The 306 effect of BT on δD values for benzene and toluene was not investigated. It is assumed that BT of benzene and toluene would have also only a minimal effect on their δD values since stable isotopes of 307 308 C and H have similar physical properties in this regard.

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9 **3.6.** Usage of the Split Flow Sample

Having the opportunity to obtain a replicate sample from the split flow presents several advantages over traditional TD-units. These include (i) obtaining a duplicate sample allowing TD-GC-irMS measurements at different concentrations of the same sample,. This which is achieved by varying the
split ratio of the TD unit. (The merit of the split flow collection is demonstrated in its application to
emission samples (see 3.7).) (ii) oObtaining-a required replicate δ valuessample for stable isotope
analysis, (iii) obtaining a duplicate sample for identifying compounds by TD-GC-MS which leads to
(iv) saving important sampling time and resources. It reduces significantly the number of samples to
be taken in the field without compromising the need for number of replicate analyses.
The δD results obtained for VOCs from the split flow tube were similar/ identical to those obtained on

- the sample tube (Fig. 6). This is supported by results from multiple sample collection of the split flow effluent (circle of 6 split flow collections) with SD < 0.2 ‰ (δ^{13} C).
- 321 **3.7.** δD Analyses of VOCs in Different Emissions
- 322 δD analysies by TD-GC-irMS was applied to three emission sources (biomass combustion, petrol and
 323 diesel car engines).
- Table 4 shows the obtained δD values of collected VOCs. Most SDs for δD analyses of VOC in the
 emissions from a petrol engine and biomass combustion range between 1 and 5 ‰ and are well within
 the instrument precision. However, SDs are up to 14 ‰ for analyses of VOCs from the diesel engine
 emissions_are up to 14 ‰. The high SDs of VOCs in the diesel engine emission are which is related to
 a highly complex unresolved mixture of compounds. For CSIA it is essential to obtain baseline
 separation for compounds of interest. This was not the case for the diesel sample.
- 330 It was necessary to analyse the samples in To obtain reliable δD values for an array of compounds samples needed to be analysed at different concentrations (different split ratios) in order to be-assess 331 332 the data within the linear range of the instrument by using the replicate samples of the split flow 333 collection. to obtain reliable δD values for an array of compounds. This was possible due to the 334 ability of repeatedly using a sample by collecting the split flow effluent. The comparison of δD values 335 of benzene, toluene and m-xylene (Table 4) demonstrate significant differences (27 ‰, 37 ‰ and 130 336 ‰, respectively) between emissions from a petrol car engine and from biomass combustion (Karri). Thus clear source differentiation carries the potential to be used in source tracking of VOCs. 337

4. Conclusions

In this study we investigated compound specific δD and $\delta^{13}C$ analysis using thermal desorption to collect and concentrate atmospheric VOCs ($C_6 - C_{10}$) using TenaxTA as adsorbent material. We established a reliable method enabling reproducible results for δD and $\delta^{13}C$ analyses with negligible isotopic fractionation compared to direct standard analyses. We evaluated the impact of storage time on δD and $\delta^{13}C$ values and revealed that breakthrough has only minor impact on δ values. We also showed that the sample portion collected of the split flow effluent can be used as a replicate sample for isotope analysis saving valuable sampling time and reseources.

This paper presents for the first time δD values of VOCs in emission samples showing a clear
distinctions of δD between emissions from biomass combustion and car exhaust emissions (petroleum
engine). TD-GC-irMS is a reliable technique and maybe used for establishing further source(s).

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Fig. 1: Sequences of TD operation a) sample tube desorption and b) cold trap desorption. An optional
split flow is illustrated in a discontinuous line

447	Fig. 2: Principle of \deltaD analysis by TD-GC-irMS
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- 448 **Fig. 3**: Principle of δ^{13} C analysis by TD-GC-irMS
- 449 Fig. 4: TD-GC-MS analysis of a standard mix a) from all compounds 1-benzene, 2-toluene, 3-water,
- 450 4-ethylbenzene, 5-p-xylene, 6-m-xylene, 7-cumene, 8-o-xylene, 9- n-propylbenzene, 10-
- 451 styrene, 11-1,2,4-trimethylbenzene, 12-butylbenzene, respectively and b) its extracted ion
- 452 chromatogram for ion 78 (benzene), 91 (toluene) and 18 (water).
- **Fig. 5**: δ^{13} C values of benzene and toluene in the standard mix collected on a (•) sample tube and on the subsequent (**A**) breakthrough tube compared to a δ^{13} C value of a (\Box) reference analysis (adsorption without breakthrough)
- 456 Fig. 6: Differences between δD values of VOCs in the standard mix on the sample tube and on the
 457 associated split flow tube. (TMB = trimethylbenzene)

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- 459 Table 1: TD-unit flow settings for different tests
- 460**Table 2**: Results for δD [‰] analyses of VOCs (standard mixture) for up to 176 days of storage; mean461 $\delta D \pm SD$ [‰] over 146 days; shift in δD between day 0 to day 176
- **462** Table 3: Results for δ^{13} C [‰] analyses of VOCs in the standard mixture for up to 40 days of storage.
- 463 Table 4: δD values [‰] of VOCs in car exhaust emissions (petrol and diesel engine) and in biomass
- 464 combustion emissions (Karri *Eucalyptus diversicolor*) with related standard deviations (SD)
- 465 [‰]; n = number of analyses; $\delta_{K} = \delta D$ values Karri; $\delta_{p} = \delta D$ values petrol engine