

1 **Characterisation of organic inclusions in stalagmites using laser-ablation-**
2 **micropyrolysis gas chromatography-mass spectrometry**

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24 **Abstract**

25 Laser-micropyrolysis gas chromatography-mass spectrometry (La-Py-GC-MS) allows the
26 analysis of small targeted areas of organic material. In this proof of concept study a novel
27 application of the technique is demonstrated. Three types of organic matter preserved in
28 speleothems were analysed: dissolved organic matter **within the calcite crystal matrix**;
29 detrital organic inclusions; and lithified guano derived from birds and bats. The results
30 indicate that there is significant heterogeneity within each sample type, with guano samples
31 having the highest variability. However, there are also distinctive La-Py-GC-MS products that
32 allow separation of the sample types geochemically. These include the chain length
33 distribution within the longer chain *n*-alkanes ($>C_{20}$), with the guano sample having a
34 dominance of chain lengths below C_{27} , whilst the other two sample types are dominated by
35 *n*-alkanes of C_{27} and above. The detrital inclusion sample has a higher relative abundance of
36 pyrrole and **methylpyrroles**. A homologous series of longer chain **alkylbenzenes** is seen only
37 in the guano samples.

38 Our preliminary data indicate that fingerprints of La-Py-GC-MS analyses may be useful to
39 provide information on organic matter preserved in speleothems, but more work is required
40 to extend the technique to lower organic content samples, and to fully test the application
41 in a larger dataset of preserved organic matter types.

42 **Keywords**

43 Speleothem; laser micropyrolysis; organic inclusion, gas chromatography-mass
44 spectrometry

45 **1. Introduction**

46 Laser-micropyrolysis gas chromatography-mass spectrometry (La-Py-GC-MS) can yield
47 geochemical information from laser spots as small as ~20 µm spot size, and thus provides
48 the ability to provide reasonably detailed analyses of heterogeneous pyrolysable organic
49 matter at resolutions similar to that of microscopy (Greenwood et al., 1996). In general, La-
50 Py-GC-MS has been demonstrated to provide similar data to flash pyrolysis, but at the 10s of
51 microns scale (Yoshioka and Ishiwatari, 2002), and multiple applications have been
52 developed, especially within organic rich rocks. These include the analysis of individual
53 macerals, so as to ascertain the chemistry specific to each maceral type (Greenwood et al.,
54 1993; Stout, 1993; Greenwood et al., 1998; Greenwood et al., 2001), the characterisation of
55 solid bitumens at the molecular level (Greenwood et al., 2001; Sestak et al., 2009), and the
56 analysis of microfossils, such as Tasmanites algae (Greenwood et al., 1998; Greenwood et
57 al., 2000), acritarchs (Arouri et al., 1999; Arouri et al., 2000), chitinozoans (Jacob et al., 2007)
58 and pollen (Al Sandouk-Lincke et al., 2013). Oil-bearing fluid inclusions have also been
59 analysed by La-Py-GC-MS (Greenwood et al., 1998), although sensitivity and power of the
60 laser tends to lead to some laser artefacts that make comparison with other bulk analytical
61 methods difficult (George et al., 2012). Use of a pulsed femtosecond laser has allowed single
62 oil inclusions to be analysed successfully (Volk et al., 2010). Outside of geology, La-Py-GC-MS
63 has also found use in forensic studies such as analysis of paint flakes (Armitage et al., 2001;
64 Prati et al., 2014). A recent study comparing La-Py-GC-MS to Curie Point Pyrolysis GC-MS
65 (CP-Py-GC-MS) concluded that there were several advantages pertaining to the use of laser
66 pyrolysis, including the provision for smaller sample sizes and good spatial analytical control
67 (Al Sandouk-Lincke et al., 2014), but that the higher (estimated as > 1000 °C) and

68 unquantified temperature applied in laser pyrolysis will affect compound distributions. They
69 therefore concluded that the nature of the organic matter was important in assessing the
70 suitability of pyrolysis techniques, with more mature organic matter better suited to laser
71 work (Al Sandouk-Lincke et al., 2014).

72 Here, a laser pyrolysis instrument with an Nd-YAG infrared laser was used to characterise
73 organic matter preserved within speleothems. The analysis of organic matter in
74 speleothems is a subject of increasing research interest as it potentially provides new
75 palaeoenvironmental records focused on vegetation change (Xie et al., 2003; Blyth et al.,
76 2007; 2011; Rushdi et al. 2011; Li et al., 2014), soil dynamics (Xie et al., 2003; Blyth et al.,
77 2013b; Li et al., 2014), and temperature (Yang et al., 2011; Blyth & Schouten, 2013; Blyth et
78 al., 2014). However, the focus on biomarkers extractable by wet chemistry from the
79 carbonate means that most research to date has required relatively large sample sizes of
80 calcite (1 – 100 g) (Xie et al., 2003; Blyth et al., 2007; Yang et al., 2011; Blyth & Schouten
81 2013; Li et al., 2014), compared to established inorganic geochemical proxies. Selected
82 techniques published to date allow analysis of less than 1 g of calcite: thermochemolysis of
83 lignin (0.5 g; Blyth & Watson, 2009; Blyth et al., 2010); amino acid analysis (25–120 mg,
84 Lauritzen et al., 1998; Blyth et al., 2008); high performance liquid chromatography analysis
85 of short chain fatty acids (0.5–3.5 g, Bosle et al., 2014); and liquid chromatography-isotope
86 ratio mass spectrometry (LC-IRMS) analysis of bulk organic isotopes (0.2 g; Blyth et al.,
87 2013a,b). Of these, only LC-IRMS of bulk $\delta^{13}\text{C}$ has so far been fully applied in a multi-proxy
88 palaeoenvironmental reconstruction (Blyth et al., 2013b). The need for larger calcite
89 samples significantly limits the temporal resolution of the time-series recovered, and often
90 excludes the possibility of analysing individual organic inclusions or layers resulting from
91 biofilms or airborne, water-lain, or faunal detrital input. Alternative techniques such as

92 continuous fluorescence analysis have met with good success in providing high resolution
93 records of certain forms of dissolved organic matter (e.g. Baker et al., 1996; Baker et al.,
94 1998; Perrette et al., 2000; Baker & Bolton 2000; Perrette et al., 2005; Baker et al., 2008),
95 but are limited in the amount of molecular detail that can currently be recovered. The
96 development of new analytical techniques that can be applied at a high sample resolution
97 and with minimal wet chemical preparation is therefore desirable.

98 In this paper the La-Py-GC-MS technique was applied to three speleothem samples with
99 contrasting organic contents: (i) soil-derived dissolved organic matter distributed within the
100 crystalline calcite; (ii) detrital or *in-situ* particulate organic matter; and (iii) lithified guano of
101 a known faunal origin. Comparison of these samples and consideration of the characteristic
102 chemical differences provides an initial insight into the potential utility of the La-Py-GC-MS
103 technique, and possible future applications in speleothem geochemistry.

104

105 **2. Materials and methods**

106 **2.1 Samples**

107 Three speleothem samples were analysed in this preliminary study to represent three
108 preserved organic matter types. The first is an Australian stalagmite (370 FS03) of Plio-
109 Pleistocene age from the Nullarbor Plain, Australia. This sample has a black colouration
110 hypothesised to be due to a high proportion of humic acids entrapped in the crystalline
111 structure (Caldwell et al., 1982). Due to this abnormal organic content, it is the only
112 speleothem analysed where it was possible to pyrolyse the calcite using the La-Py-GC-MS
113 instrument as currently configured. The second sample is from an Ethiopian stalagmite

114 (Bero-1) from the Mechara region of Ethiopia, which formed between 10 ka and present in
115 an open cave, recessed into a cliff-face (Asrat et al., 2008). It contains laminae of visible
116 detrital organic matter, which may have been deposited by water, or due to the relatively
117 open nature of the cave, possibly by air or faunal input. The third sample (HV-1) is a crust of
118 lithified guano incorporated into a Peruvian stalagmite from Cueva de las Lechuzas in
119 Peruvian Amazonia, and is believed to be Holocene in age (H. Vonhof pers comm.). Based on
120 modern accumulations in the area, the guano is derived from a mixture of bats and oilbirds,
121 which both live in the cave.

122 A sample section approximately 5 mm x 5 mm was removed from each speleothem,
123 centring on visible organic inclusions in the case of Bero-1 and HV-1 guano. Prior to analysis,
124 the sample blocks were cleaned in dichloromethane to remove surface contamination.
125 Samples were thermally desorbed in the pyrolysis chamber of the instrument (see Section
126 2.2) at 100 °C for 10 minutes prior to analysis to reduce desorption artefacts, although the
127 latter were found to be significant only for the guano samples. To minimise these artefacts,
128 piece 2 of the guano was thermally extracted offline at 200 °C, and held at 100 °C in an oven
129 overnight prior to analysis.

130

131 **2.2 Analytical methods**

132 A detailed description of the instrument and methodology can be found elsewhere
133 (Greenwood et al., 1996; Greenwood et al., 1998; Bernard et al., 2010; Sandouk-Lincke et
134 al., 2013; Prati et al., 2014). Briefly, a continuous wave laser (Laser Applications 9500,
135 Nd:YAG laser, $\lambda = 1064$ nm) was tightly focused onto selected areas of the sample through
136 an Olympus BPX60M microscope equipped with reflected light illumination and using long

137 distance working objectives (20X/0.4 and 50X/0.5). The sample was located in a purpose-
138 built pyrolysis chamber (100-110°C, 100 mL helium flow) which was interfaced to a GC-MS
139 system (Hewlett Packard 6890 GC interfaced to a 5973 mass selective detector, electron
140 energy 70eV) via a gas inlet system designed for maximum transfer efficiency of the gaseous
141 products. The products of the pyrolysis process were cryogenically trapped in a coiled nickel
142 loop using a liquid nitrogen bath. After trapping the products, a 6 port transfer valve was
143 rotated to transfer 1 mL/min helium through the trap and the contents were then desorbed
144 by heating to 320 °C. The products were cryo-focused again in a loop of GC column
145 immersed in a liquid nitrogen bath. The full scan GC-MS analysis (m/z 50-550) of the
146 pyrolysates was performed on a DB-5MS column (J&W, 60 m, 0.25 mm I.D., 0.25 μ m film
147 thickness) with helium as carrier gas with constant pressure of 25 psi. The GC oven was
148 programmed for an initial temperature of 40 °C (2 min hold) followed by heating at 4 °C/
149 min to 310 °C (30 min hold).

150 The size of the laser crater was controlled by a combination of instrumental parameters
151 such as choosing the appropriate magnification depending on the spatial resolution
152 required for the sample, the laser power and time span of the laser event. The 370 FS03 and
153 Bero-1 stalagmite samples were pyrolysed using a 50x objective and applied power of 19.4
154 W for 1 s or 1.2 W for 0.2 s, respectively, yielding spot sizes of \sim 20 μ m. The lithified guano
155 sample was pyrolysed using a 20x objective and applied power of 1.9 W for 0.2 s, yielding
156 spot sizes of \sim 40-50 μ m. Between 80 and 127 shots into different parts of each sample were
157 aggregated in each run using a liquid nitrogen trap, so as to provide sufficient sensitivity for
158 compound identification. The analytical runs chosen for data analysis are those that
159 produced the largest compound yields for their sample type. Figure 1 shows SEM images of

160 the pyrolysis holes in Bero-1 and HV-1. Compounds were identified by comparison to mass
161 spectral libraries and gas chromatographic retention times (e.g. Eganhouse et al., 1993).

162

163 **3. Results and discussion**

164 **3.1 Compounds identified**

165 Figure 2 shows representative pyrograms for each of the three sample types, and
166 identified pyrolysis products are listed in Table 1. All the samples contain a homologous
167 series of *n*-alkanes (C₆ – C₃₁), *n*-alkenes (C₆-C₁₈) and a range of low molecular weight
168 aromatic compounds. In addition to the major peaks visible on the TIC pyrogram, other
169 peaks include several compounds of lower abundance that were quantified in some or all of
170 the samples, including *o*-xylene, *n*-propylbenzene, *iso*-propylbenzene, ethylmethylbenzenes,
171 trimethylbenzenes, methylindenes, pyrrole, methylpyrroles, phenol, a methylphenol, furan,
172 and methylfurans. In terms of organic matter fingerprinting, the main distinguishing
173 considerations to separate the three sample types were found to be the relative proportions
174 of compounds within the *n*-alkane series, and the presence or absence of some key
175 aromatic groups.

176

177 **3.2 *n*-Alkenes and *n*-alkanes**

178 Homologous series of *n*-alkenes and *n*-alkanes are frequently seen from the pyrolysis of
179 organic matter (Hatcher & Clifford 1994; Fezzey & Armitage, 2006; Al Sandouk-Lincke et al.,
180 2014), deriving from the pyrolysis breakdown of larger biopolymers. Their dominance in a
181 pyrogram has previously been associated with humic acids (Fezzey & Armitage, 2006),

182 although it has also been suggested that this dominance is a result of an inherent analytical
183 bias in favour of non-polar compounds (Hatcher & Clifford 1994). The results here indicate
184 that interrogation of the relative chain length distribution within each group (*n*-alkenes; *n*-
185 alkanes <C₂₀; *n*-alkanes >C₂₀) is informative in partially separating the three sample types.
186 Low and higher molecular weight *n*-alkanes are discussed separately due to the bimodality
187 of the chromatograms.

188 The *n*-alkene series runs from C₆-C₁₈ in Bero-1, C₇-C₁₆ in 370 FS03, and C₆-C₂₁ in the HV-1
189 guano samples (Fig. 3). Within the alkenes present, there is little variation between repeat
190 analyses of each sample type for Bero-1 and HV-1 ($r^2 = 0.90$ to 0.98 , $p = 0.0000$). For 370
191 FS03, only one analytical run (370 FS03 brown) produced a series of alkenes, with the
192 second run (370 FS03 black) containing only C₁₄ at a measurable abundance. When
193 considering the differences between sample types, the Bero-1 detritus and HV-1 guano
194 show a significant correlation (Fig. 4a; $r^2 = 0.83$, $p = 0.0000$), with decreasing abundance
195 with increasing chain length. In contrast the organic matter coating the crystals in 370 FS03
196 does not correlate with either of the other sample types, having a reduced number of
197 measurable alkenes, and three distinct maxima at C₈, C₁₄, and C₁₆.

198 For *n*-alkanes, different distributions and relationships are apparent between the three
199 sample types in both the low molecular weight (LMW, chain length C₁₉ and below) and high
200 molecular weight (HMW, chain length C₂₀ and above) components. Fig. 5 shows the LMW
201 groups. Variation within each sample type is greater than for the *n*-alkenes. Repeat analyses
202 for Bero-1 show a significant correlation ($r^2 = 0.73$, $p < 0.0001$), as do the two repeats on HV-1
203 1 piece 2 ($r^2 = 0.67$, $p < 0.0005$). However, there is no meaningful relationship between HV-1
204 piece 2 and HV-1 piece 1 ($r^2 = 0.16$, $p > 0.1$). This difference is most likely down to the
205 increased pre-extraction for piece 2, with piece 1 still showing alkane artefacts from

206 desorption. There is also no meaningful correlation between the two analyses of 370 FS03
207 ($r^2 = 0.06$, $p > 0.1$), partly because one repeat (370 FS03 black) did not contain measurable
208 alkanes at the lower end of the chain length range. However, the overall trend in this
209 sample, *n*-alkanes increasing in abundance with increasing chain length, is similar for both
210 analyses, and is contrary to that in Bero-1 and HV-1, both of which have a decrease in *n*-
211 alkane abundance with increasing chain length, but with different patterns. Therefore, as for
212 the *n*-alkenes, LMW *n*-alkanes seem most useful in separating 370 FS03 from the other two
213 sample types, based on the trend of the inverse correlations (Fig. 4b). **It is not certain at
214 present why the *n*-alkanes and *n*-alkenes exhibit different chain length patterns in these
215 results, and this is an issue that merits further research before definite environmental
216 interpretations are developed from this approach.**

217 The HMW group shows more distinct and useful patterns. In particular, the dominant
218 chain length in the distribution is different in each sample type, and consistent between
219 analyses of the same sample type, regardless of other variations in the composition. Bero-1
220 shows a maximum at C₂₉, 370 FS03 at C₂₇, and HV-1 at C₂₃ (Fig. 6). Over the whole
221 distribution there is very little variation between the two Bero-1 runs ($r^2 = 0.95$, $p = 0.0000$).
222 In the HV-1 guano samples, the two runs of piece 2 also have a good correlation ($r^2 = 0.79$, p
223 < 0.0001), but the relationship between the two pieces is much less significant ($r^2 = 0.53$, $p <$
224 0.1). The two runs of 370 FS03 also have a relatively weak correlation ($r^2 = 0.53$, $p < 0.1$).
225 When the average relative distributions are compared between sample type, there is a
226 reasonable correlation between Bero-1 and 370-FS03 ($r^2 = 0.70$, $p < 0.001$), but not between
227 either of these and HV-1. This is clearly shown in the % chart in Figure 7, where it can be
228 seen that the HV-1 guano is dominated by chain lengths between C₂₀ and C₂₃, whereas the
229 Bero-1 detritus and 370-FS03 organic matter are dominated by C₂₇-C₃₁.

230

231 3.3 Other compounds

232 A number of compound groups have been identified as being significant in natural
233 organic matter pyrolysis, including phenols, pyrroles, furans, and aromatic hydrocarbon
234 precursors such as indenes, methylbenzenes and naphthalenes (e.g. Wilson et al., 1983;
235 White et al., 2007). The relative abundance of these fractions has previously been suggested
236 as a method of discriminating between organic matter types in soils (White et al., 2007).
237 However, in this study, although there are some bulk differences between the three sample
238 types (Fig 8a), there are also good correlations, with p values under 0.05 (Fig 8b). This result
239 suggests that for these three samples, using this technique, the relative abundance of the
240 compounds listed above is not a good discriminator. However, two compound groups do
241 stand out as different. The pyrroles (nitrogen compounds most likely derived from proteins;
242 Wilson et al., 1983), have a much higher relative abundance in the Bero-1 detritus (Fig. 8),
243 and a set of longer-chain alkylbenzenes only occur in the HV-1 guano (Fig. 9). **The presence**
244 **of only a single isomer at each carbon number for these alkylbenzenes indicates a natural**
245 **source (Takada & Ishiwatari 1989).** The guano samples did not contain any of the
246 compounds previously identified as diagnostic pyrolysis markers for chitin (e.g.
247 acetylpyridones, acetamidofuran, etc; see Stankiewicz et al., 1996), indicating that the
248 guano may not have been derived from an insectivorous source, although further work on
249 modern cave guano samples would be required to demonstrate this.

250

251 **3.4 Future research directions**

252 Overall the results of this study indicate some intriguing differences between the
253 three sample types, but caution also needs to be applied in designating a parameter as due
254 to source organic matter variations, as some parameters (e.g. alkane chain length) have
255 been shown to have a relationship with pyrolysis temperature (Al Sandouk-Lincke et al.,
256 2014). Although laser pyrolysis temperatures will be uniformly high, they are not controlled
257 as exactly as in other pyrolysis techniques, **which means the precise range of pyrolysis**
258 **temperatures applied here cannot be quantified. The data here are preliminary, and the**
259 **abundances of compounds measured is relatively low, so** the sample range is not at this
260 stage sufficient to provide definitive identifications of the source of organic matter
261 inclusions in time-series records.

262 A limitation on this technique is the applicability only to speleothem samples with
263 discrete organic inclusions or a high visible organic content within the crystal matrix. The
264 early stage of this research trialed the technique on speleothems without visible organic
265 contents, but no data was recoverable, as the laser was not sufficient to pyrolyse the calcite.
266 Experimentation with the technique using alternate laser sources is therefore indicated to
267 attempt to expand the applicability, as this approach would be of particular potential use in
268 characterising organic matter at laminae level resolutions which are too small to be feasible
269 to study via the conventional wet chemistry methods currently used on speleothems. Such a
270 study would also be able to investigate the effect of calcite type and texture on organic
271 matter ablation.

272 A final issue that would benefit from further research is that of potential
273 contamination, as a recent study published after the completion of the experimental phase
274 of this research has suggested that some molecules in speleothem extractions may be

275 artefacts due to contamination (Wynn & Brocks 2014). The 2014 study only dealt with a
276 limited suite of biomarkers within speleothems, and not the organic inclusions and
277 macromolecules analysed in this paper. Precautions were taken in this work by solvent
278 cleaning and thermal desorption prior to laser pyrolysis to maximise the integrity of the
279 data. However, there is certainly scope for further research to consider the potential
280 changes in pyrolysis products with depth from the cut surface within a single inclusion or
281 laminae, and whether any such changes may relate to contamination or natural
282 heterogeneity.

283

284 4. Conclusions

- 285 • The Peruvian guano (HV-1) has two distinguishing features separating it from both
286 the other sample types: the dominance of the lower carbon numbers in the long
287 chain ($>C_{20}$) *n*-alkanes, and the presence of longer chain alkylbenzenes. These
288 features are consistent between both guano pieces, despite some other
289 differences in composition.
- 290 • The detrital inclusions in the Ethiopian sample Bero-1 contain higher proportions
291 of furans, pyrroles and phenols than the other samples.
- 292 • The DOM in the black Nullarbor Plain speleothem 370 FS03 differed from the
293 other samples in the trends of the *n*-alkenes and shorter chain *n*-alkanes, and in
294 the smaller range of aromatic and nitrogen compounds. This is consistent with
295 the proposed dominance of humic acids as the cause for the colouration in these
296 samples (Caldwell 1982), in the light of some research on pyrolysis of humic acids
297 (Fezzey & Armitage 2006). However, the absence of some compounds may be a

298 function of the low abundance of pyrolysable material in the sample, rather than
299 in the nature of that material, meaning that a direct comparison with other OM
300 types must be approached with caution.

- 301 • The results indicate that La-Py-GCMS may have the potential to **characterise**
302 detrital organic matter in speleothems, and further work on a greater range of
303 samples, and with alternative laser set-ups to allow the pyrolysis of the calcite is
304 indicated to test the true applicability of this approach

305

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315

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472

473 **Table and Figure captions**

474 Table 1. List of identified and quantified compounds

475 Figure 1. SEM images showing laser ablation holes (indicated with arrow) in a) a detrital
476 inclusion in Bero-1, and b) the lithified guano of HV-1.

477 Figure 2. Total ion chromatograms of La-Py-GC-MS analyses of a) Bero-1 debris run 1; b) 370
478 FS03 brown; c) HV-1 p2. 127 shots. Compound identifications are provided in Table
479 1.

480 Figure 3. Distribution of carbon chain lengths of *n*-alkenes in a) Bero-1 debris; b) 370 FS03; c)
481 HV-1. Scatter plots show the level of correlation between runs within each sample
482 type. C₁₁ is excluded from HV-1 piece 1, as it co-eluted with alkane desorption
483 artefacts.

484 Figure 4. Scatter plots showing the correlations between sample types for a) *n*-alkenes; b)
485 shorter chain *n*-alkanes (<C₂₀); c) longer chain *n*-alkanes (>C₂₀)

486 Figure 5. Distribution of carbon chain lengths of shorter chain *n*-alkanes (<C₂₀) in a) Bero-1
487 debris; b) 370 FS03; c) HV-1. Scatter plots show the level of correlation between runs
488 within each sample type. C₁₁ is excluded from HV-1 piece 1, as it co-eluted with
489 alkane desorption artefacts.

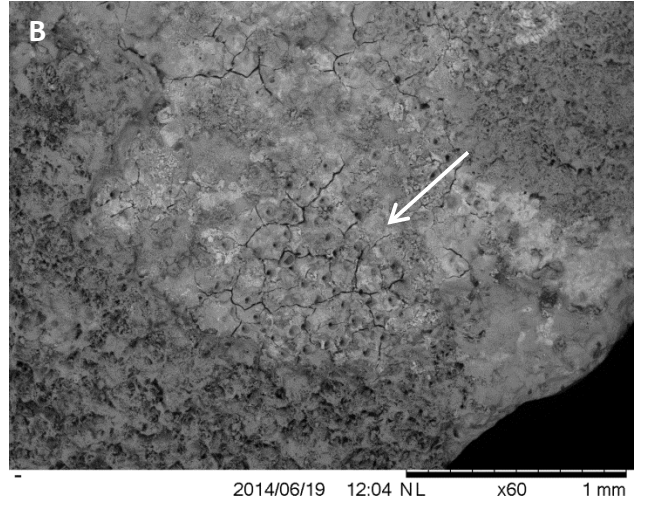
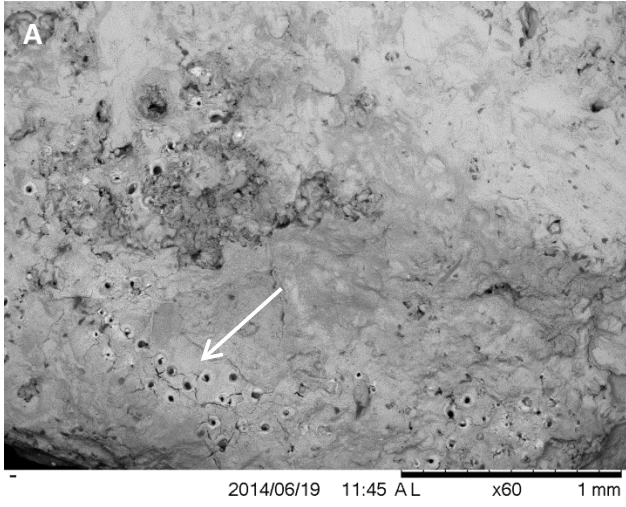
490 Figure 6. Distribution of carbon chain lengths of longer chain *n*-alkanes in a) Bero-1 debris;
491 b) 370 FS03; c) HV-1. Scatter plots show the level of correlation between runs within
492 each sample type.

493 Figure 7. A percentage bar chart showing the increased dominance of compounds below C₂₅
494 in HV-1 in comparison to increased longer chain compounds in Bero-1 and 370 FS03.

495 Figure 8. a) Pie charts showing the average relative abundances of aromatic compounds
496 within each sample type. Pyrroles are a major feature of Bero-1. b) Scatter plots
497 showing the correlation between the different sample types for the aromatic
498 compounds.

499 Figure 9. A percentage bar chart of propyl-, methyl- and alkyl benzenes in the three sample
500 types, showing the presence of longer chain alkyl benzenes in HV-1 only.

Figure 1



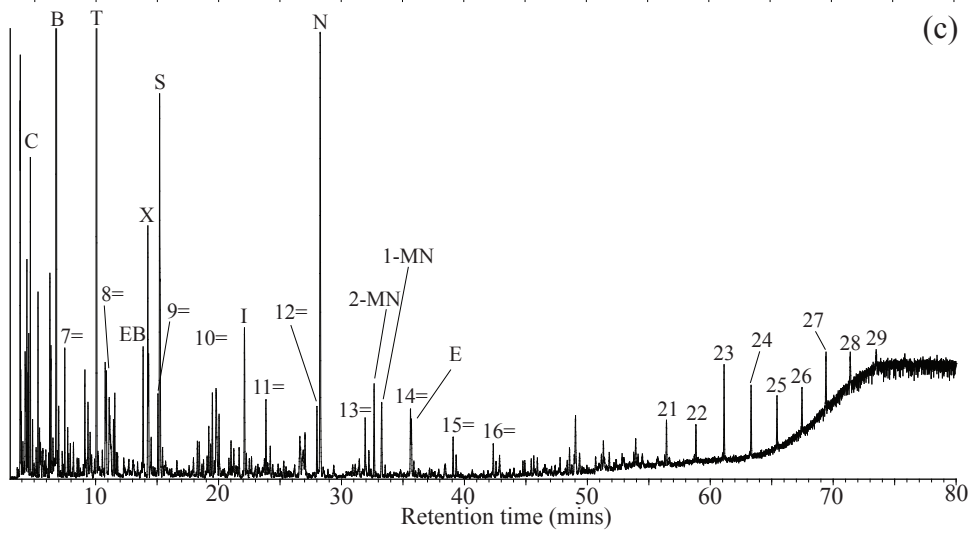
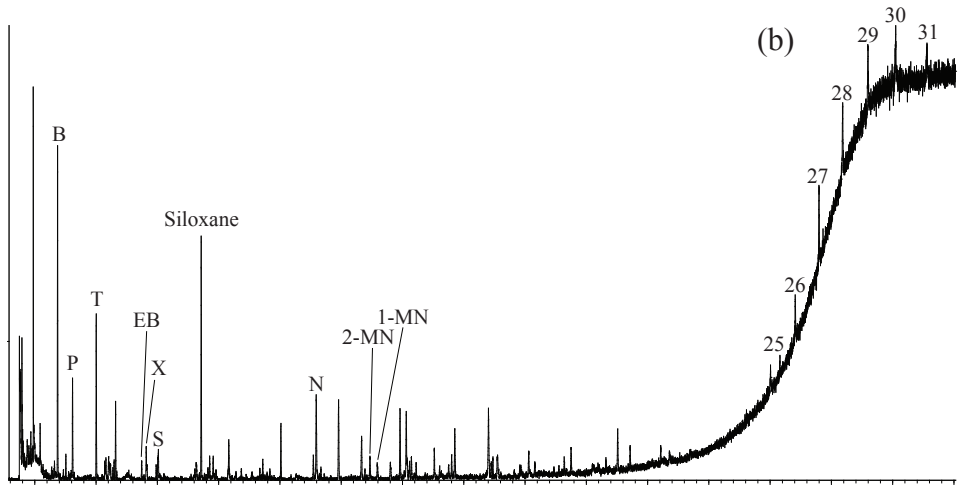
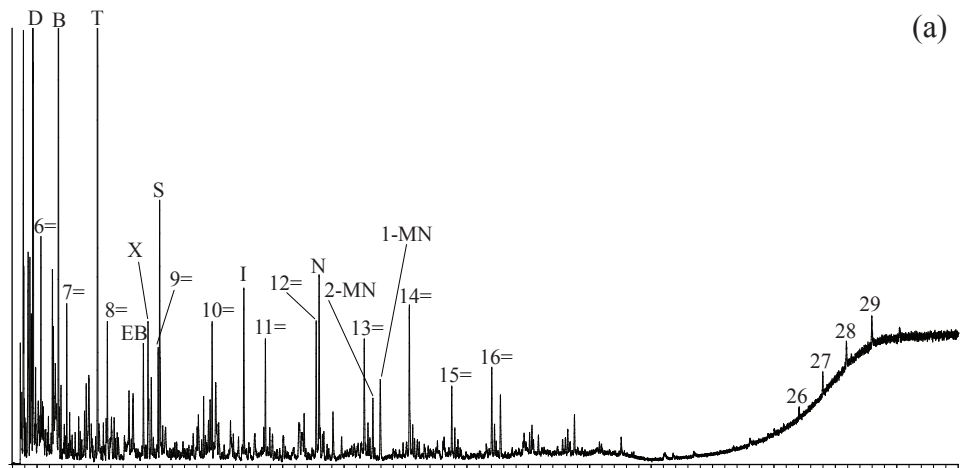
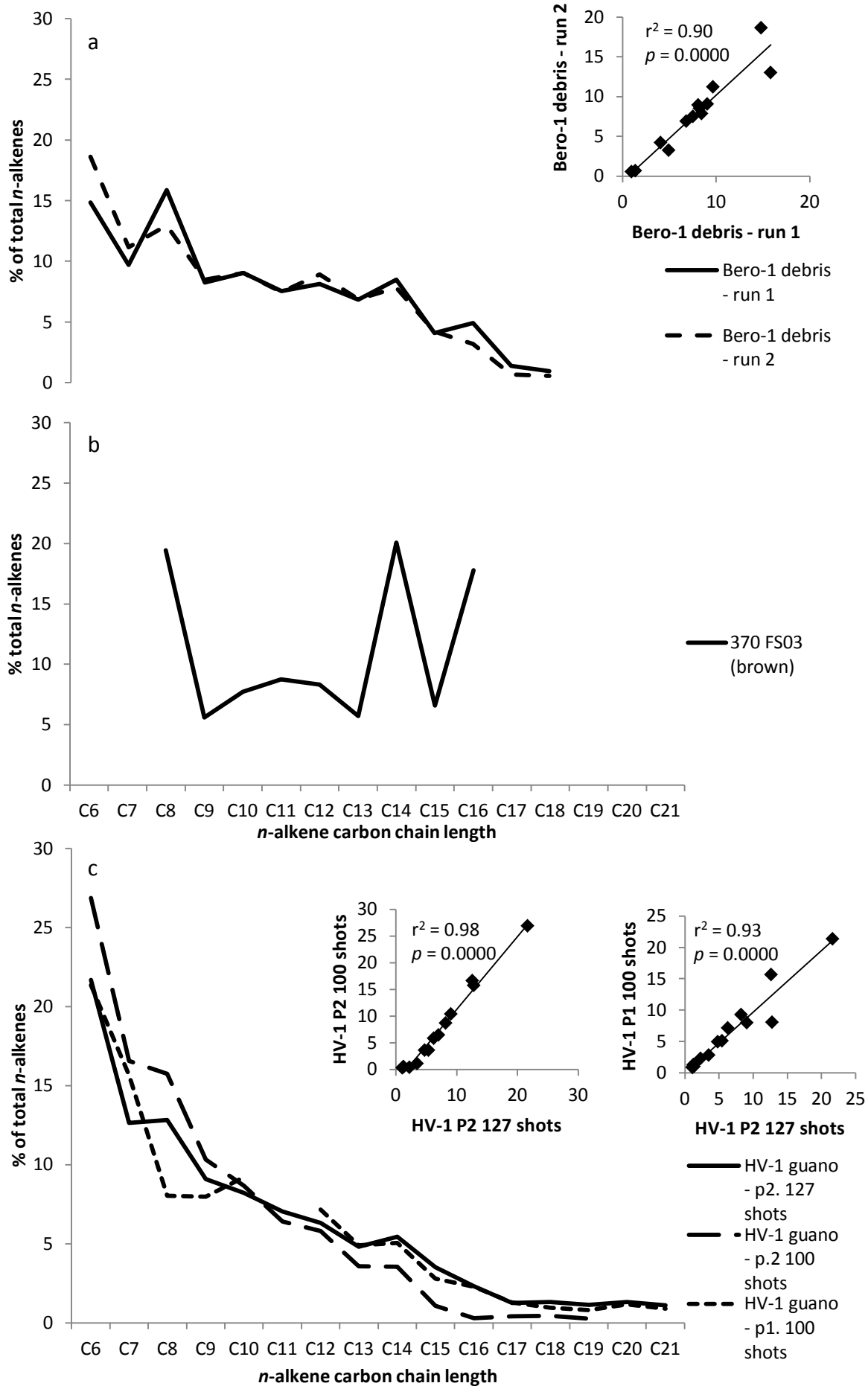
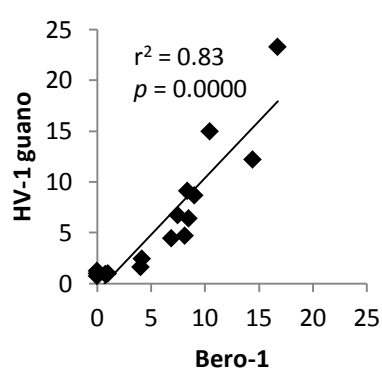
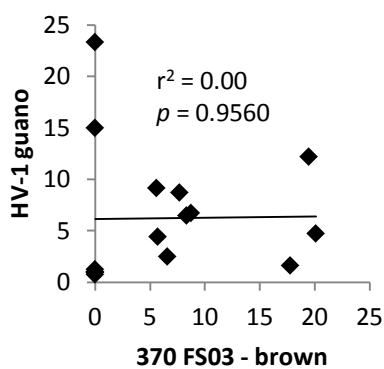
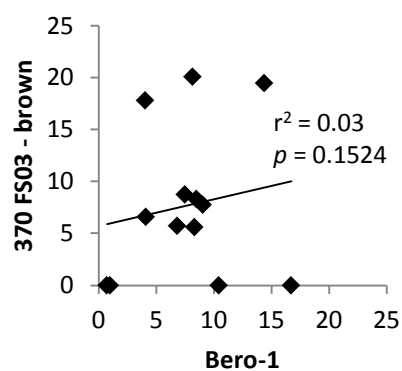


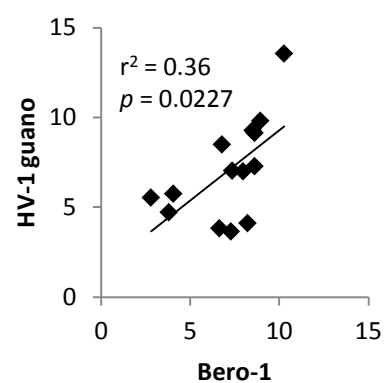
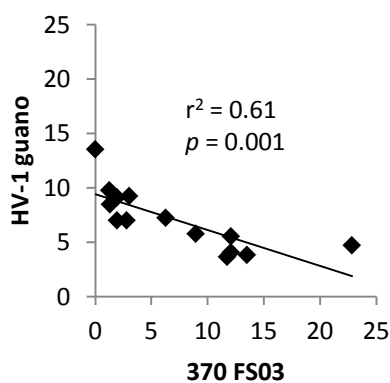
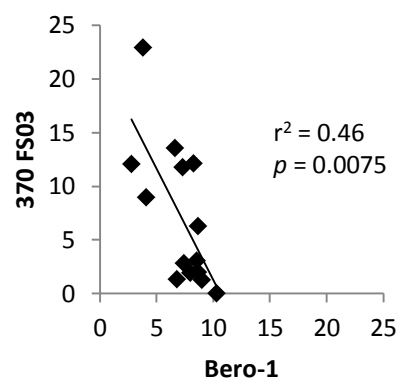
Figure 3.



a) *n*-alkenes



b) *n*-alkanes <C₂₀



c) *n*-alkanes >C₂₀

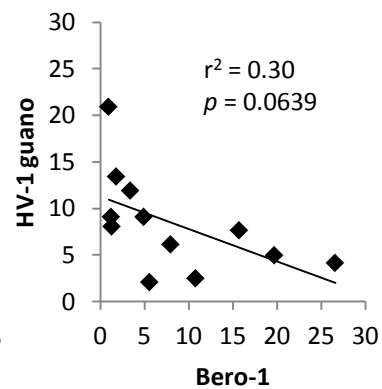
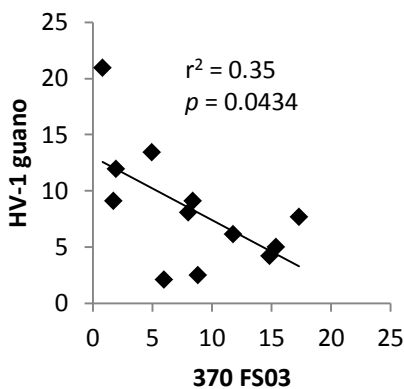
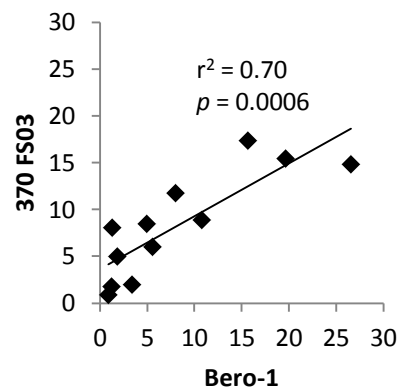


Figure 4.

Figure 5.

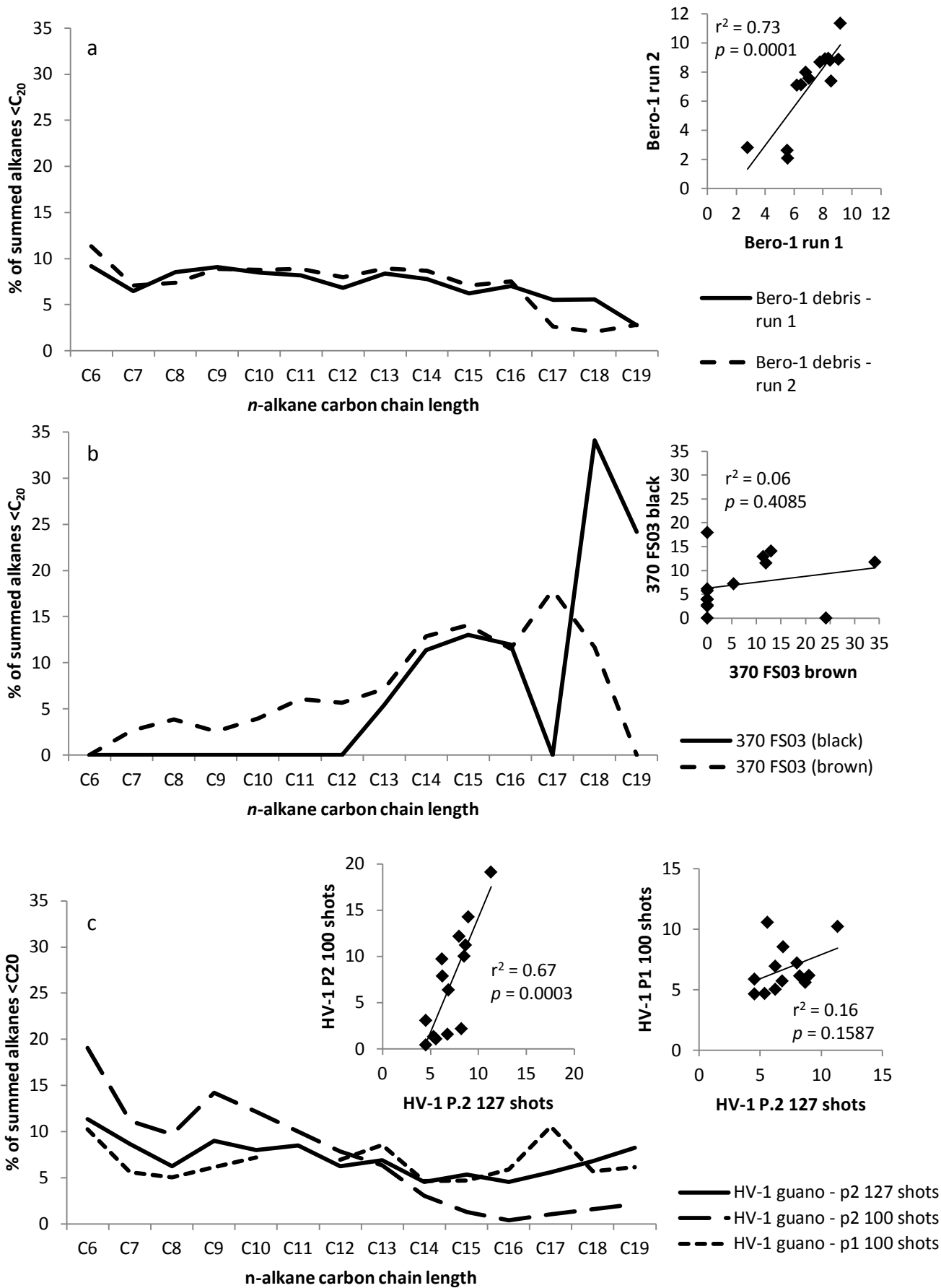


Figure 6.

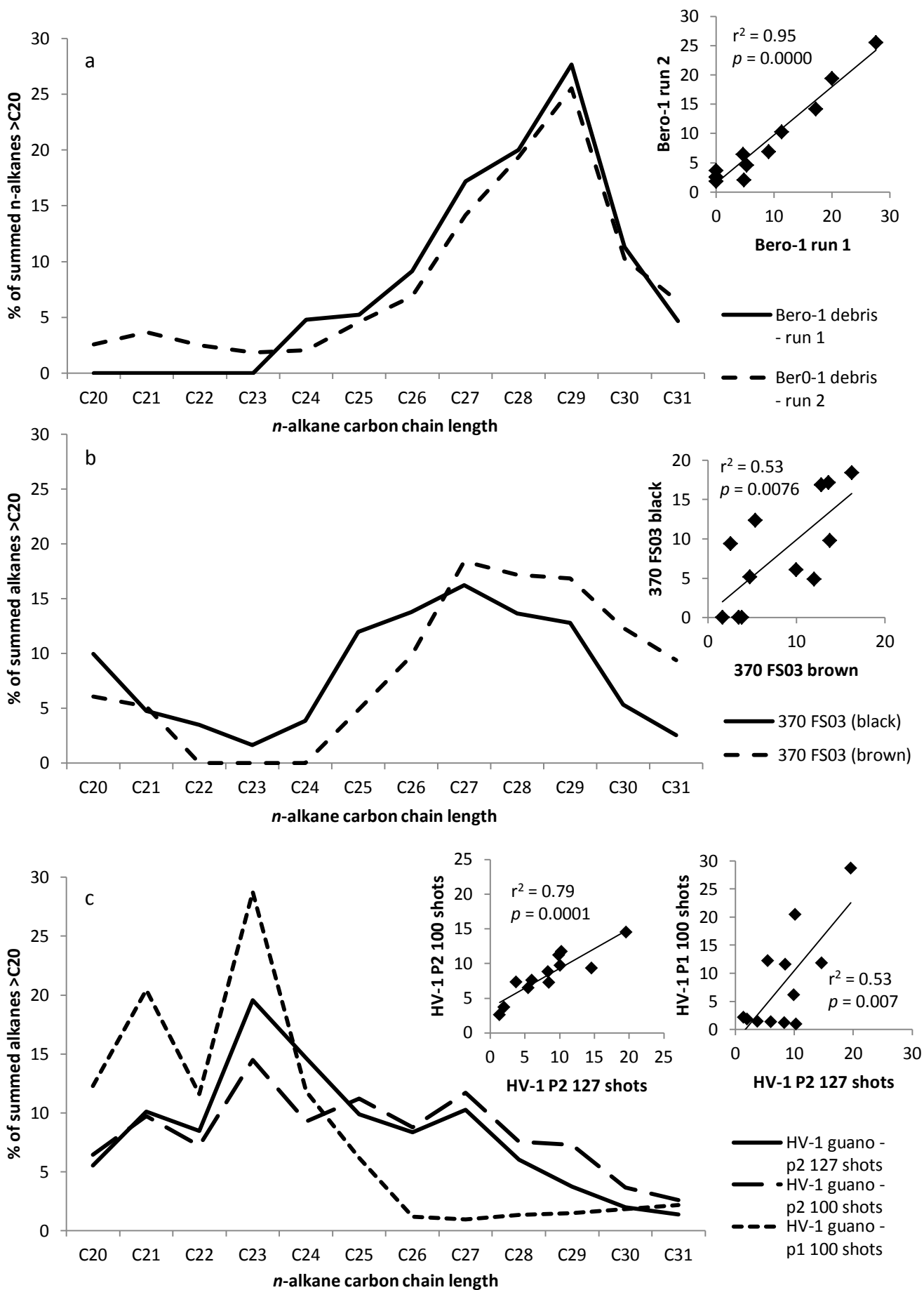


Figure 7.

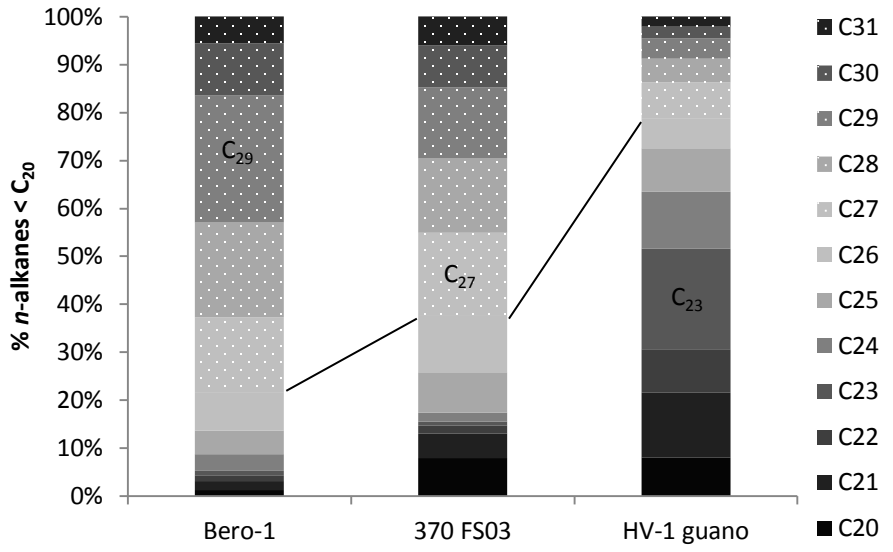
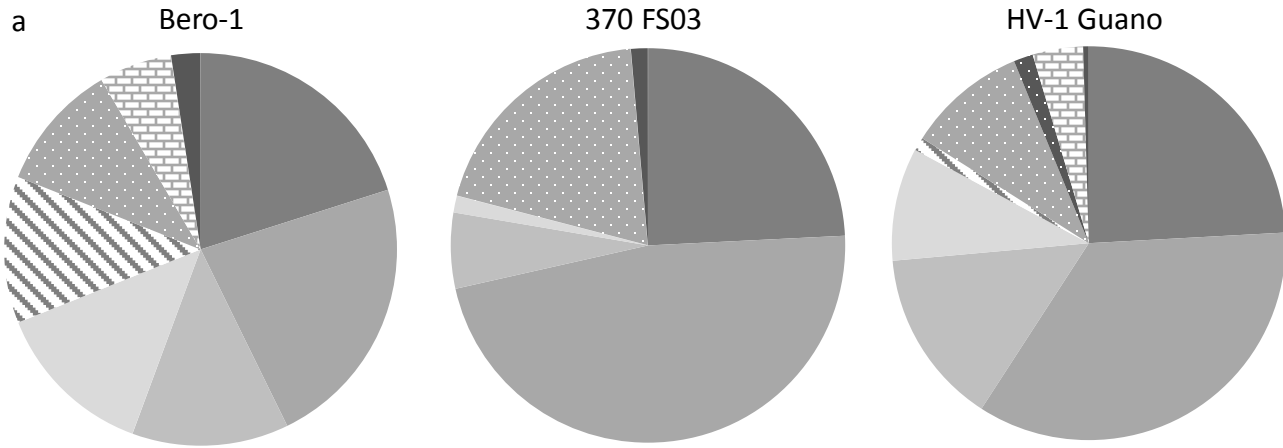


Figure 8.



- Xylenes & Ethylbenzenes
- Napthalenes
- Styrene
- 1-H-Indene & methyl
- ▨ Methylpyroles
- Methylbenzenes
- Long Chain Alkylbenzenes
- ▨ Furans & Methylfuran
- Phenol & Methylphenols

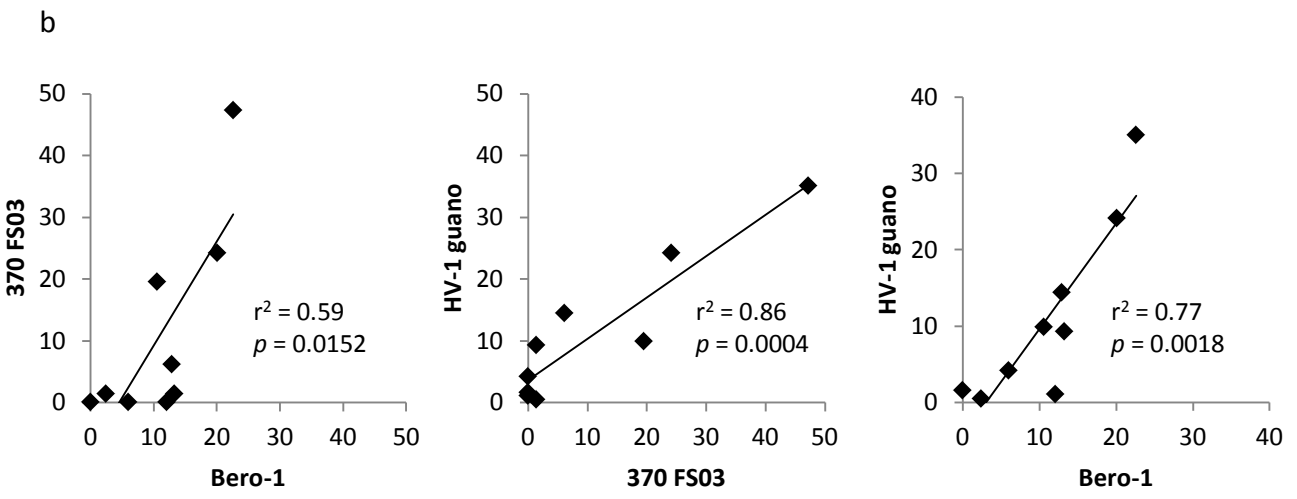


Figure 9.

