Atlas of Genetics and Cytogenetics in Oncology and Haematology

OPEN ACCESS JOURNAL

Gene Section Review

HSPA8 (heat shock 70kDa protein 8)

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Published in Atlas Database: August 2013

Online updated version : http://AtlasGeneticsOncology.org/Genes/HSPA8ID40878ch11q24.html DOI: 10.4267/2042/53483

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Abstract

Review on HSPA8, with data on DNA/RNA, on the protein encoded and where the gene is implicated.

Identity

Other names: HSC54, HSC70, HSC71, HSP71, HSP73, HSPA10, LAP1, NIP71

HGNC (Hugo): HSPA8

Location: 11q24.1

DNA/RNA

Note

The human HSPA8 gene includes nine exons and eight introns. It is mapped into chromosome 11, 11q23.3-q25 (Sonna et al., 2002). Introns 5, 6 and 8 contain highly conserved repeats about 90 bp which code for U14 snoRNA (Chen et al., 1996). Exons 2, 3, 6, 7, and 9 code for the peptides of extremely uniform length, between 61 and 69 amino acids while exons 4, 8 and 5 code for peptides of 51, 78 and 185 amino acids, respectively. Two 150 bp direct repeats (nt 674 to nt 829 and nt 1783 to nt 1937) are 85% homologous to each other (Dworniczak and Mirault, 1987).

Description

The gene coding for HSC70, HSPA8 affects the posttranscriptional silencing mediated by RNAi and is a component of the RNAi pathway in Drosophila cultured cells (Dorner et al., 2006). The promoter region of the HSPa8 gene includes a TATA box, two CCAAT boxes, two SP1 elements and two sets

of heat shock response elements (HSE) where the heat shock transcriptional factors bind (Chen et al., 2002).

Protein

Note

The HSPA8 gene coding for the protein HSC70, also known as HSP73, is a 73 kDa heat shock cognate protein.

HSC70 is an ATP binding chaperone and has intrinsic ATPase activity which hydrolyzes ATP into ADP (Jakob et al., 1996).

HSC70 hydrolyzing ATP initiates the conformational change of HSC70 and further causes substrate binding by HSC70 (Sullivan and Pipas, 2002).

Description

HSC70, also called HSP73, is a 73 kDa heat shock cognate protein. The basic structure of human HSC70 includes three parts: a 44 kDa aminoterminal adenosine triphosphatase (ATPase) domain (residues 1-384), also known as the ATPbinding domain, an 18 kDa peptide (substrate) binding domain (residues 385-543), and a 10 kDa carboxyl-terminal domain (residues 544-646) which is also designated as the variable or "lid" domain (Smith et al., 1998; Tsukahara et al., 2000; Sullivan and Pipas, 2002).

The carboxyl-terminal amino acid sequence Glu-Glu-Val-Asp (EEVD motif), which is absolutely conserved in all eukaryotic HSC70 and HSP70 family members, is essential for association with some co-chaperones (Mosser et al., 2000).



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The model for HSC70 and substrates binding and releasing cycle. In the ATP-bound state, HSC70 has low affinity with the substrates. After hydrolysis of ATP with the ATPase activity, HSC70 in the ADP bound state binds with the substrates with high affinity. Some co-chaperones such as Dna J homologues enhance the ATPase activity of HSC70. Nucleotide exchange factors such as GrpE enhance the dissociation of bound ADP from HSC70 to allow the binding of ATP, resetting the cycle.

Two amino acid sequences have the characteristics of nuclear localization signals in human HSC70 which are involved in nuclear import of HSC70: DAKRL69-73 in the amino-terminal and KRKHKKDISENKRAVRR246-262 in the ATPase domain (Lamian et al., 1996; Tsukahara and Maru, 2004).

Expression

Various conditions and molecules can affect the expression of the HSPA8 gene.

The table below summarizes the factors which affect the expression of the HSPA8 gene (Liu et al., 2012).

Localisation

HSC70 is a major cytosolic molecular chaperone (Place and Hofmann, 2005).

However, HSC70 is also located in various cellular locations such as nuclear and close to cellular membrane (Arispe et al., 2002).

HSC70 can interact with the lipid bilayer in the cellular membrane directly and form ion-conductance channels allowing ion flow through the cellular membrane.

HSC70 is also identified as a tyrosinephosphorylated protein associated with the nuclear envelope. HSC70 supports the nuclear import of karyophilic proteins and may play a role in nucleocytoplasmic transport (Otto et al., 2001).

Function

1. As a molecular chaperone, HSC70 regulates protein folding, maintains protein normal structure and functions, and protects cells from physical and chemcial damage. HSC70 regulates protein maturation and interacts with nascent polypeptides in the process of new protein synthesis (Beckmann et al., 1990; Beckmann et al., 1992). HSC70 can also regulate the translocation of proteins into different cellular organelles such as endoplasmic reticulum and mitochondria (Chirico et al., 1998, Sheffield et al., 1990).

2. HSC70 was oginally characterized as an uncoating ATPase that dissociates clathrin triskelions from clathrin-coated vesicles. It catalyzes the ATP-dependent uncoating of clathrin-coated pits (chappell et al., 1986; Goldfarb et al., 2005).

3. HSC70 is involved in targeting protein to lysosomes for degradation (Terlecky et al., 1992) and to ubiquitin/proteasome machinery for degradation (Massyuki, 1994).

4. HSC70 regulates cellular signaling and functions such as steroid receptor maturation and Akt signaling pathway (Kimmins and MacRae, 2000; Shiota et al., 2010).

5. HSC70 is important in regulating apoptosis, embryonic development and aging (Beere, 2004; Sreedhar and Csermely, 2004; Kodiha et al., 2005).

NH2	44 kDa amino-terminal ATPase domain (also known as ATP binding domain)	18 kDa substrate- binding domain	10 kDa carboxyl- terminal domain (also know as "lid" domain)	— соон
	1	384	543	646

The structure of HSC70. The basic structure of human HSC70 includes three parts: a 44 kDa ATPase domain, an 18 kDa peptide (substrate) binding domain and a 10 kDa carboxyl-terminal domain, also known as "lid" domain.

Increase HSPA8 expression	Decrease HSPA8 expression	
Stress	Sodium 4-phenylbutyrate	
Heat	Butyrate	
Organochlorine	Glycerol	
Sodium arsenite	Quercetin	
Azetidine	Arginase	
Heave metals (nickel, cooper, lead, cadmium)	3,3',4,4',5-Pentachlorinateed biphenyls 126 (PCB126)	
Exercise	Mitotane (in mitochondria)	
Antimony containing drugs	Deoxyspergualin (DSG)	
Geranylgeranylacetone	cAMP	
Ethanol	D-galactosamine	
Phorbol 12-myristate 13- acetate	Lipopolysaccharide	
Estrogen	INF-gamma	
Progesterone	TNF-alpha	
IL-1 beta	Oxymatrine	
Krüppel-like factor 4	HSP70 overexpression	
Insulin		

Homology

HSC70 belongs to the HSP70 family in which includes other three members: the HSP70, the glucose-regulated protein 78 (GRP78), and HSP75. Human HSC70 shares 85% amino acid similarity with HSP70 (Ali et al., 2003).

They have similar functions and interact with each other in an ATP-dependent manner.

HSC70 can form a stable complex with newly synthesized HSP70 upon heat shock (Brown et al., 1993).

Implicated in

Various cancers

Note

The HSPA8 gene is higher expressed in some cancer cells such as human colon cancer (Kubota et al., 2000).

HSC70 regulates functions of various cancerrelated genes and proteins. HSC70 functions as a molecular stabilizer of nonphosphorylated retinoblastoma protein (pRb) by directly binding to it (Inoue et al., 1995).

HSC70 binds to a mutant form of p53 and p73 and links them for degradation.

HSPA8 gene can suppress oncogene (such as mutant p53 and Ras) mediated transformation (Yehiely and Oren, 1992; Gaiddon et al., 2001).

Cytogenetics

Intronic polymorphism (1541-1542delGT) of HSPA8 is associated with decrease of lung cancer risk (Rusin et al., 2004).

Cadiovascular diseases

Note

HSC70 plays a protective role in myocardial injuries. HSC70 is commonly found in atherosclerotic plaques during the atherogenesis and therosclerotic plaque progression and it appears to be a protective factor against cellular stress (Dupont et al., 2008).

HSC70 expression is significantly decreased in diabetic myocardium because of insulin deficiency.

Insulin directly increases the abundance of HSC70 in cultured cardiomyocytes and overexpression of HSC70 leads to protect against stress via suppression of apoptosis signalling (Chen et al., 2006).

Neurological diseases

Note

HSC70 is directly involved in cell survival during neurulation and HSC70 acts as an intrinsic protector of neuroepithelial and neural precursor cells (Rubio et al., 2002).

HSC70 mediates the chaperone mediated lysosomal degradation of α -synuclei which is accumulated in Parkingson's disease and other neurodegenerative diseases (Mak et al., 2010).

HSC70 facilitates degradation of the amyotrophic lateral sclerosis (ALS) -linked mutant SOD1 protein in an ubiquitination-dependent manner (Urushitani et al., 2004; Casoni et al., 2005). HSC70 mediates the autophagy for the degradation of mutant huntingtin protein which can cause Huntington's disease (Bauer et al., 2010).

Liver diseases

Note

HSC70 has been implicated in the pathogenesis and the pathophysiology of hepatic diseases such as hepatitis B and C, non-alcoholic steatohepatitis autoimmune hepatitis, and primary biliary cirrhosis. HSC70 plays important role in the replication of hepatitis B virus and hepatitis C virus.

HSC70 could be a novel molecular target for diagnosis and treatment of hepatitis B and C (Wang et al., 2010).

To be noted

Note

HSC70 interacts with many molecules and plays an important role in regulating cellualr functions. As a molecular chaperone, HSC70 interacts with cochaperones, also called co-factors. Those cochaperones include auxilin, BAG family members, Hip, Hop, HSPBP1 and CHIP (Liu et al., 2012). In addition to co-chaperones, HSC70 interacts with many other cellular molecules. The cooperation of HSC70 with HSP90 regulates the glucocorticoid receptor activation and signaling pathway (Furay et al., 2006). HSC70 facilitates the heat-shock factor1 mediated cell survival in response to cellular stress (Ahn et al., 2005). HSC70 interacts and forms a complex with histone deacetylase 3 (HDAC3). HSC70 interacts with newly synthesized cyclin D1 and mature cyclin D1/CDK4 holoenzyme complex, thus involved in cell cycle regulation (Diehl et al., 2003). HSC70 interacts genetically with target of rapamycin (TOR) and acts as a regulator of endocytosis (Hennig et al., 2006). HSC70 can regulate the synthesis of GABA by interacting with rate-limiting enzyme L-glutamic the acid decarboxylase (GAD).

Research has been conducted to use HSC70 as a drug target for cancer therapy. A recombinant vaccine composed of CD4+ and CD8+ T cell epitopes fused to HSC70 may prevent tumor growth and metastasis (Mizukami et al., 2008). Fusion the ATPase domain of HSC70 with tyrosinase-related protein-2 acts as a tumor vaccine against B16 melanoma (Zhang et al., 2006).

References

Sonna LA, Fujita J, Gaffin SL, Lilly CM. Invited review: Effects of heat and cold stress on mammalian gene expression. J Appl Physiol (1985). 2002 Apr;92(4):1725-42

Chappell TG, Welch WJ, Schlossman DM, Palter KB, Schlesinger MJ, Rothman JE. Uncoating ATPase is a member of the 70 kilodalton family of stress proteins. Cell. 1986 Apr 11;45(1):3-13

Dworniczak B, Mirault ME. Structure and expression of a human gene coding for a 71 kd heat shock 'cognate' protein. Nucleic Acids Res. 1987 Jul 10;15(13):5181-97

Chirico WJ, Waters MG, Blobel G. 70K heat shock related proteins stimulate protein translocation into microsomes. Nature. 1988 Apr 28;332(6167):805-10

Beckmann RP, Mizzen LE, Welch WJ. Interaction of Hsp 70 with newly synthesized proteins: implications for protein folding and assembly. Science. 1990 May 18;248(4957):850-4

Sheffield WP, Shore GC, Randall SK. Mitochondrial precursor protein. Effects of 70-kilodalton heat shock protein on polypeptide folding, aggregation, and import competence. J Biol Chem. 1990 Jul 5;265(19):11069-76

Beckmann RP, Lovett M, Welch WJ. Examining the function and regulation of hsp 70 in cells subjected to metabolic stress. J Cell Biol. 1992 Jun;117(6):1137-50

Terlecky SR, Chiang HL, Olson TS, Dice JF. Protein and peptide binding and stimulation of in vitro lysosomal proteolysis by the 73-kDa heat shock cognate protein. J Biol Chem. 1992 May 5;267(13):9202-9

Yehiely F, Oren M. The gene for the rat heat-shock cognate, hsc70, can suppress oncogene-mediated transformation. Cell Growth Differ. 1992 Nov;3(11):803-9

Brown CR, Martin RL, Hansen WJ, Beckmann RP, Welch WJ. The constitutive and stress inducible forms of hsp 70 exhibit functional similarities and interact with one another in an ATP-dependent fashion. J Cell Biol. 1993 Mar;120(5):1101-12

Ohba M. A 70-kDa heat shock cognate protein suppresses the defects caused by a proteasome mutation in Saccharomyces cerevisiae. FEBS Lett. 1994 Sep 5;351(2):263-6

Inoue A, Torigoe T, Sogahata K, Kamiguchi K, Takahashi S, Sawada Y, Saijo M, Taya Y, Ishii S, Sato N, Kikuchi K. 70-kDa heat shock cognate protein interacts directly with the N-terminal region of the retinoblastoma gene product pRb. Identification of a novel region of pRb-mediating protein interaction. J Biol Chem. 1995 Sep 22;270(38):22571-6

Chen MS, Featherstone T, Laszlo A. Amplification and altered expression of the hsc70/U14 snoRNA gene in a heat resistant Chinese hamster cell line. Cell Stress Chaperones. 1996 Apr;1(1):47-61

Jakob U, Scheibel T, Bose S, Reinstein J, Buchner J. Assessment of the ATP binding properties of Hsp90. J Biol Chem. 1996 Apr 26;271(17):10035-41

Lamian V, Small GM, Feldherr CM. Evidence for the existence of a novel mechanism for the nuclear import of Hsc70. Exp Cell Res. 1996 Oct 10;228(1):84-91

Smith DF, Whitesell L, Katsanis E. Molecular chaperones: biology and prospects for pharmacological intervention. Pharmacol Rev. 1998 Dec;50(4):493-514

Arispe N, De Maio A. ATP and ADP modulate a cation channel formed by Hsc70 in acidic phospholipid membranes. J Biol Chem. 2000 Oct 6;275(40):30839-43

Kimmins S, MacRae TH. Maturation of steroid receptors: an example of functional cooperation among molecular chaperones and their associated proteins. Cell Stress Chaperones. 2000 Apr;5(2):76-86

Mosser DD, Caron AW, Bourget L, Meriin AB, Sherman MY, Morimoto RI, Massie B. The chaperone function of hsp70 is required for protection against stress-induced apoptosis. Mol Cell Biol. 2000 Oct;20(19):7146-59

Tsukahara F, Yoshioka T, Muraki T. Molecular and functional characterization of HSC54, a novel variant of human heat-shock cognate protein 70. Mol Pharmacol. 2000 Dec;58(6):1257-63

Gaiddon C, Lokshin M, Ahn J, Zhang T, Prives C. A subset of tumor-derived mutant forms of p53 down-regulate p63 and p73 through a direct interaction with the p53 core domain. Mol Cell Biol. 2001 Mar;21(5):1874-87

Otto H, Dreger M, Bengtsson L, Hucho F. Identification of tyrosine-phosphorylated proteins associated with the nuclear envelope. Eur J Biochem. 2001 Jan;268(2):420-8

Chen MS, Goswami PC, Laszlo A. Differential accumulation of U14 snoRNA and hsc70 mRNA in Chinese hamster cells after exposure to various stress conditions. Cell Stress Chaperones. 2002 Jan;7(1):65-72

Rubio E, Valenciano AI, Segundo C, Sánchez N, de Pablo F, de la Rosa EJ. Programmed cell death in the neurulating embryo is prevented by the chaperone heat shock cognate 70. Eur J Neurosci. 2002 May;15(10):1646-54

Sullivan CS, Pipas JM. T antigens of simian virus 40: molecular chaperones for viral replication and tumorigenesis. Microbiol Mol Biol Rev. 2002 Jun;66(2):179-202

Ali KS, Dorgai L, Abrahám M, Hermesz E. Tissue- and stressor-specific differential expression of two hsc70 genes in carp. Biochem Biophys Res Commun. 2003 Aug 1;307(3):503-9

Diehl JA, Yang W, Rimerman RA, Xiao H, Emili A. Hsc70 regulates accumulation of cyclin D1 and cyclin D1dependent protein kinase. Mol Cell Biol. 2003 Mar;23(5):1764-74

Beere HM. "The stress of dying": the role of heat shock proteins in the regulation of apoptosis. J Cell Sci. 2004 Jun 1;117(Pt 13):2641-51

Rusin M, Zientek H, Krześniak M, Małusecka E, Zborek A, Krzyzowska-Gruca S, Butkiewicz D, Vaitiekunaite R, Lisowska K, Grzybowska E, Krawczyk Z. Intronic polymorphism (1541-1542delGT) of the constitutive heat shock protein 70 gene has functional significance and shows evidence of association with lung cancer risk. Mol Carcinog. 2004 Mar;39(3):155-63

Sreedhar AS, Csermely P. Heat shock proteins in the regulation of apoptosis: new strategies in tumor therapy: a comprehensive review. Pharmacol Ther. 2004 Mar;101(3):227-57

Tsukahara F, Maru Y. Identification of novel nuclear export and nuclear localization-related signals in human heat shock cognate protein 70. J Biol Chem. 2004 Mar 5;279(10):8867-72

Urushitani M, Kurisu J, Tateno M, Hatakeyama S, Nakayama K, Kato S, Takahashi R. CHIP promotes proteasomal degradation of familial ALS-linked mutant SOD1 by ubiquitinating Hsp/Hsc70. J Neurochem. 2004 Jul;90(1):231-44

Casoni F, Basso M, Massignan T, Gianazza E, Cheroni C, Salmona M, Bendotti C, Bonetto V. Protein nitration in a mouse model of familial amyotrophic lateral sclerosis: possible multifunctional role in the pathogenesis. J Biol Chem. 2005 Apr 22;280(16):16295-304

Kodiha M, Chu A, Lazrak O, Stochaj U. Stress inhibits nucleocytoplasmic shuttling of heat shock protein hsc70. Am J Physiol Cell Physiol. 2005 Oct;289(4):C1034-41

Place SP, Hofmann GE. Comparison of Hsc70 orthologs from polar and temperate notothenioid fishes: differences in prevention of aggregation and refolding of denatured proteins. Am J Physiol Regul Integr Comp Physiol. 2005 May;288(5):R1195-202

Chen HS, Jia J, Su HF, Lin HD, Chen JW, Lin SJ, Yang JY, Lai HC, Mestril R, Wang PH. Downregulation of the constitutively expressed Hsc70 in diabetic myocardium is

mediated by insulin deficiency. J Endocrinol. 2006 Aug;190(2):433-40

Dorner S, Lum L, Kim M, Paro R, Beachy PA, Green R. A genomewide screen for components of the RNAi pathway in Drosophila cultured cells. Proc Natl Acad Sci U S A. 2006 Aug 8;103(32):11880-5

Furay AR, Murphy EK, Mattson MP, Guo Z, Herman JP. Region-specific regulation of glucocorticoid receptor/HSP90 expression and interaction in brain. J Neurochem. 2006 Aug;98(4):1176-84

Goldfarb SB, Kashlan OB, Watkins JN, Suaud L, Yan W, Kleyman TR, Rubenstein RC. Differential effects of Hsc70 and Hsp70 on the intracellular trafficking and functional expression of epithelial sodium channels. Proc Natl Acad Sci U S A. 2006 Apr 11;103(15):5817-22

Hennig KM, Colombani J, Neufeld TP. TOR coordinates bulk and targeted endocytosis in the Drosophila melanogaster fat body to regulate cell growth. J Cell Biol. 2006 Jun 19;173(6):963-74

Zhang H, Wang W, Li Q, Huang W. Fusion protein of ATPase domain of Hsc70 with TRP2 acting as a tumor vaccine against B16 melanoma. Immunol Lett. 2006 Jun 15;105(2):167-73

Dupont A, Chwastyniak M, Beseme O, Guihot AL, Drobecq H, Amouyel P, Pinet F. Application of saturation dye 2D-DIGE proteomics to characterize proteins modulated by oxidized low density lipoprotein treatment of human macrophages. J Proteome Res. 2008 Aug;7(8):3572-82

Bauer PO, Goswami A, Wong HK, Okuno M, Kurosawa M, Yamada M, Miyazaki H, Matsumoto G, Kino Y, Nagai Y, Nukina N. Harnessing chaperone-mediated autophagy for the selective degradation of mutant huntingtin protein. Nat Biotechnol. 2010 Mar;28(3):256-63

Kubota H, Yamamoto S, Itoh E, Abe Y, Nakamura A, Izumi Y, Okada H, Iida M, Nanjo H, Itoh H, Yamamoto Y. Increased expression of co-chaperone HOP with HSP90 and HSC70 and complex formation in human colonic carcinoma. Cell Stress Chaperones. 2010 Nov;15(6):1003-11

Mak SK, McCormack AL, Manning-Bog AB, Cuervo AM, Di Monte DA. Lysosomal degradation of alpha-synuclein in vivo. J Biol Chem. 2010 Apr 30;285(18):13621-9

Shiota M, Kusakabe H, Izumi Y, Hikita Y, Nakao T, Funae Y, Miura K, Iwao H. Heat shock cognate protein 70 is essential for Akt signaling in endothelial function. Arterioscler Thromb Vasc Biol. 2010 Mar;30(3):491-7

Wang YP, Liu F, He HW, Han YX, Peng ZG, Li BW, You XF, Song DQ, Li ZR, Yu LY, Cen S, Hong B, Sun CH, Zhao LX, Kreiswirth B, Perlin D, Shao RG, Jiang JD. Heat stress cognate 70 host protein as a potential drug target against drug resistance in hepatitis B virus. Antimicrob Agents Chemother. 2010 May;54(5):2070-7

Liu T, Daniels CK, Cao S. Comprehensive review on the HSC70 functions, interactions with related molecules and involvement in clinical diseases and therapeutic potential. Pharmacol Ther. 2012 Dec;136(3):354-74

This article should be referenced as such:

Liu T, Cao S. HSPA8 (heat shock 70kDa protein 8). Atlas Genet Cytogenet Oncol Haematol. 2014; 18(3):169-173.