Running title: Alzheimer's disease, insulin resistance, CSF biomarkers

Insulin resistance is associated with increased Alzheimer disease pathology and reduced memory function in at risk healthy middle-aged adults

Siobhan M. Hoscheidt^{*1}, Erika J. Starks¹, Jennifer M. Oh¹, Henrik Zetterberg^{3,4}, Kaj Blennow³, Rachel Krause¹, Carey E. Gleason^{1,2}, Luigi Puglielli^{1,2}, Craig S. Atwood^{1,2}, Ozioma C. Okonkwo^{1,5}, Cynthia M. Carlsson^{1,2,5}, Sanjay Asthana^{1,2,5}, Sterling C. Johnson^{1,2,5}, Barbara B. Bendlin^{1,5}

¹Wisconsin Alzheimer's Disease Research Center, University of Wisconsin School of Medicine and Public Health, Madison, WI, USA

² Geriatric Research Education and Clinical Center, Wm. S. Middleton Veterans Hospital, Madison, WI, USA

³Clinical Neurochemistry Laboratory, Department of Psychiatry and Neurochemistry, Institute

of Neuroscience and Physiology, The Sahlgrenska Academy at the University of Gothenburg, Sweden

⁴Department of Molecular Neuroscience, UCL Institute of Neurology, Queen Square, London, UK

⁵Wisconsin Alzheimer's Institute, University of Wisconsin School of Medicine and Public Health, Madison, WI, USA

Correspondence:

Barbara B. Bendlin, PhD J5/1M, Clinical Science Center 600 Highland Ave Madison, WI 53792 Phone: (608) 265-2483 E-mail: bbb@medicine.wisc.edu

Figures: 4 Total word count: 1 Abstract

2 **Background:** Metabolic disorders in midlife increase risk for Alzheimer's disease (AD). 3 Dysfunction in insulin signaling is suspected to increase the formation of A β_{42} peptides and is 4 thought to be associated with impaired memory function. Insulin resistance (IR) observed in 5 midlife prior to indication of neurodegeneration and cognitive decline is a candidate risk factor 6 that can be modified by targeted interventions. Regulation of normal insulin function, as a 7 modifiable risk factor, may be important in reducing the prevalence of AD, particularly in 8 individuals who harbor a genetic risk or have a parental family history of AD. The relationship 9 between IR and AD pathology remains poorly understood, particularly at the preclinical stage of 10 the disease.

11

Objective: The objective of the current study was to examine whether IR is associated with
increased AD pathology and decreased memory performance in asymptomatic middle-aged
adults enriched for AD (i.e. with a parental family history of sporadic late-onset AD). We
postulated that IR would be associated with greater AD pathology, particularly in carriers of the *APOE* ε4 allele, and poorer memory performance.

17

18 **Method:** Asymptomatic middle-aged adults with a parental family history of AD (N=70, mean 19 age=57.7 years) from the Wisconsin Alzheimer's Disease Research Center underwent lumbar 20 puncture, blood draw and neuropsychological testing. Cerebrospinal fluid (CSF) samples were 21 assayed for AD related biomarkers including soluble amyloid precursor protein β (sAPP- β), 22 abeta₄₂ (A β_{42}) and phosphorylated tau (P-tau₁₈₁). The ratio P-tau₁₈₁/A β_{42} was also examined as a 23 more sensitive measure of AD pathology with respect to APOE E4 status, IR and memory 24 function. IR was indexed by the Homeostatic Model Assessment for Insulin Resistance (HOMA-25 IR). Data were analyzed using linear regression models to determine significant effects of IR and 26 APOE E4 on CSF biomarkers of AD and memory performance.

27

28 **Results:** Both APOE ε 4 carriage and higher IR were associated with higher sAPP- β . APOE ε 4

29 carriers were observed to have significantly higher levels of AD pathology (P-tau₁₈₁/A β_{42})

30 compared to non-carriers. Investigation of memory performance revealed that higher IR and the

 $31 \quad \mbox{concurrent presence of AD pathology (P-tau_{181}/A\beta_{42}) were associated with worse delayed \\ memory performance.$

34	Conclusion: The current study provides evidence that IR in middle age is associated with higher
35	sAPP- β , a soluble marker of amyloidogenic APP-processing. Further, these results reveal that IR
36	and AD pathology concomitantly result in subclinical memory impairment in asymptomatic
37	middle-aged individuals enriched for AD. Lastly, these findings converge with prior studies
38	indicating that APOE E4 carriage is associated with increased AD pathology that can be observed
39	in early aging prior to the onset of clinical memory impairment. This study has implications for
40	development of interventions targeted at reducing the prevalence of IR through modifiable
41	lifestyle factors, particularly in individuals who harbor the $\varepsilon 4$ allele.
42	
43	Keywords: insulin resistance, CSF AD biomarkers, APOE e4, memory function
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	
61	

62 1. Introduction

63 Epidemiological studies have provided mounting evidence that insulin resistance (IR) is 64 associated with an increased risk for Alzheimer's disease (AD) (Arvanitakis et al., 2004; Ott et 65 al., 1999; Peila et al., 2002; Schrijvers et al., 2010). Over the past fifty years, the rate of type 2 66 diabetes mellitus (a non-insulin dependent form of diabetes mellitus) has increased alarmingly 67 with approximately 387 million individuals diagnosed with the disease as of 2014. Recent 68 evidence indicates that cognitively healthy individuals in late-middle life with higher IR harbor 69 more AD-like neuropathology (Starks et al., 2014, Willette et al., 2015). Such studies provide 70 support that IR may be an underlying mechanism driving an increased risk for development of 71 AD pathology and that neurodegenerative changes can be observed prior to the onset of marked 72 cognitive or behavioral impairment. While several animal studies suggest a mechanistic link 73 between IR and neuropathology (for review see de la Monte and Wands, 2005), the effect of IR 74 on the development of AD pathology among humans remains poorly understood, particularly at 75 the preclinical stage of the disease. Identification of modifiable risk factors in midlife may be 76 particularly relevant to individuals who harbor known genetic risk factors for AD (e.g., APOE 77 ɛ4) or who have parental family history of AD. Provided the increasing prevalence of Type 2 78 diabetes and a growing incidence of dementia, elucidation of neuropathologic processes that 79 arise from IR that may be associated with increased risk of AD is of paramount importance.

80

81 Efficient utilization of insulin by neurons underlies normal cellular and cognitive function. 82 Insulin signaling regulates processes critical for neuronal survival and synaptic plasticity 83 moderating processes such as glucose uptake and energy production through glucose oxidation 84 (Cersosimo and DeFronzo, 2006). A growing body of evidence has shown that insulin plays an 85 integral role in learning and memory by modulating excitatory and inhibitory receptors such as 86 glutamate and GABA receptors, respectively (see Zhao et al., 2004a). Insulin homeostasis is 87 believed to be an instrumental component in signal transduction cascades that underlie memory 88 consolidation and long-term memory function (Cardoso et al., 2009; Zhao et al., 2004a; Zhao 89 and Aklon 2001). It is suspected that brain regions dense with insulin receptors, such as the 90 medial temporal and frontal lobes, are preferentially sensitive to insulin signaling. Interestingly, 91 frontal and medial temporal lobe regions, particularly the hippocampus, are some of the first to 92 be adversely affected in AD (Craft & Watson, 2004; Henneberg & Hoyer, 1995; Zhao et al.,

93 2004). Cognitively healthy individuals with reduced peripheral insulin function show subtle 94 cognitive deficits and reduced cerebral glucose metabolism and diminished cerebrocortical 95 activity (Baker et al., 2011; Tschritter et al., 2006). Among cognitively intact older adults, 96 reduced glucose tolerance has been associated with poor memory performance and hippocampal 97 atrophy (Convit et al., 2003). Further, abnormal glucose metabolism-believed to arise from 98 desensitization of cerebral insulin receptors—is a core feature of sporadic late-onset AD 99 (Henneberg & Hoyer, 1995) and a number of human studies have provided evidence for a 100 linkage between Type 2 diabetes and greater risk for developing AD (Li et al., 2015; Peila et al., 101 2002). Histology from human postmortem studies has provided evidence that insulin pathways 102 are severely degraded in AD brains showing a vast reduction in insulin receptor expression and 103 insulin receptor binding (de la Monte & Tong, 2014). These findings are suggestive of an 104 integral role of insulin dysfunction in the pathogenesis of AD. 105 106 The proteolytic processing of amyloid precursor protein (APP) is believed to be one of many 107 neurobiological processes negatively affected by IR that may contribute to AD pathology. APP is 108 processed by two competing pathways: the amyloidogenic β-secretase-mediated and the 109 nonamyloidogenic α -secretase-mediated pathways. Cleavage of APP by α -secretase is thought to 110 mitigate the formation of extracellular amyloid plaques by preventing the formation of amyloid 111 beta (Aβ). APP processed through the α -secretase pathway is cleaved within the Aβ sequence 112 producing soluble peptides (Nunan and Small, 2000). By contrast, APP cleavage by β-secretase 113 produces sAPP-β that consequently results in generation of Aβ peptides by endoproteolytic 114 processing (Vassar et al., 1999). In the β -secretase pathway cleavage by γ -secretase adjacent to 115 residue 42 is the last step in formation of the 42 amino acid species $A\beta_{42}$. Among the $A\beta$ 116 peptides, $A\beta_{42}$ is implicated in the production and proliferation of amyloid plaques that arise 117 from the aggregation and oligomerization of A β_{42} . Amyloid plaques are a hallmark of AD 118 pathology and amyloidogenic processes such as A β_{42} oligomerization is suspected to be involved in neuronal dysfunction and cell death. Over the past decade a number of studies have provided 119 120

- 120 evidence that $A\beta_{42}$ oligomers have neurotoxic effects that include degradation of synaptic
- 121 structure (Lacor et al., 2004, 2007) and disruption of molecular and cellular mechanisms integral
- 122 to memory formation such as synaptic transmission and plasticity (Rowan et al., 2003; Shankar
- 123 et al., 2008; Lambert et al., 1998). Transgenic mouse models provide evidence that IR promotes

activation of the amyloidogenic pathway via cleavage of amyloid precursor protein by β secretase (sAPP- β) (Farris et al., 2003; Gasparini et al., 2001) and that overproduction of sAPP- β is associated with disrupted synaptic transmission and plasticity in the hippocampus (Rowan et al., 2003). Congruent with these findings, a study of diet-induced IR in Tg2576 transgenic mice showed that IR promoted amyloidogenic A β production, increased AD-type amyloid plaque burden, and impaired performance on hippocampal-dependent memory tasks (i.e., allocentric spatial water maze) (Ho et al., 2004).

131

132 Other risk factors for AD include non-modifiable risk factors such as genetic risk, parental 133 family history of AD and age. It is known that carriage of one or two ɛ4 alleles of apolipoprotein 134 E (APOE E4) is a major genetic risk factor for AD. Several studies on asymptomatic APOE E4 135 carriers compared to non-carries have provided evidence that ɛ4 allele carriers harbor features 136 characteristic of AD pathology such as hypometabolism (Reiman et al., 2004, 2005; Small et al., 137 2000), higher amyloid burden (Corder et al., 1993) and gray matter atrophy in AD-sensitive 138 regions (Lemaitre el a., 2005). Animal studies have provided evidence that APOE E4 carriage 139 may be particularly deleterious to the hippocampus showing that APOE E4 mice are impaired on 140 hippocampal-dependent tasks such as the Morris Water Maze, object recognition and context 141 fear conditioning (Salomon-Zimri et al., 2013). Thus APOE £4 has been implicated in several 142 AD-relevant pathways, including amyloid accumulation, brain hypometabolism and gray matter 143 degradation.

144

145 Several studies have shown that asymptomatic late-midlife individuals with parental family

146 history of AD (FH+) show early pathological changes in AD-sensitive regions (Adluru et al.,

147 2014; Honea et al., 2010, 2011; Johnson et al., 2006, 2014; Mosconi et al., 2010; Okonkwo et al.,

148 2012). Neurodegeneration and neuropathology observed in these individuals is consistent with,

although substantially less severe, pathology observed in mild cognitive impairment (MCI) and

150 AD patients. As genetic and familial risk factors are invariable, modifiable behaviors that may

151 decrease the risk of developing AD are of clinical interest. Development of intervention-based

strategies aimed at preventing or reversing IR in midlife may help to significantly reduce one's

153 risk of incipient neuropathological changes that may underlie or contribute to the

to etiopathogenesis of AD. Midlife is a critical time period during which lifestyle factors that

155 confer risk for AD may be altered. While carriage of the ε4 allele and parental family history of

156 AD are known to increase Alzheimer's disease risk moderating effects of these factors on

157 molecular mechanisms alone are not sufficient to cause the disease.

158

159 To investigate the association between IR and AD pathology observed in healthy middle-aged 160 adults enriched for AD [i.e. parental family history of AD (FH+)], the current study examined 161 the effect of IR on cerebrospinal fluid (CSF) biomarkers of AD and a hippocampal-dependent 162 memory task (i.e., delayed recall). Specifically we examined the effects of IR and APOE E4 163 status on CSF sAPP- β , A β_{42} , P-tau_{181} and P-tau_{181}/A β_{42} , and performance on the Rev Auditory 164 Verbal Learning Test (RAVLT), delayed recall. APOE E4 status was also examined as an 165 invariable risk factor for AD pathology in midlife. We hypothesized that both IR and APOE E4 166 would be associated with 1) increased CSF biomarkers of neural injury and amyloid burden and 167 2) decreased memory performance. Given prior studies showing that the effects of IR may 168 depend on APOE £4 carriage, we also tested for interactions between IR and APOE £4 on CSF 169 biomarkers and memory function.

170

171 2. Methods and Materials

172 2.1. Participants

173 The current study examined 70 asymptomatic middle-aged adults (mean age= 57.7 years, SD= 174 5.11, range= 46-66, 78.6% female) from the Wisconsin Alzheimer's Disease Research Center 175 (ADRC) Investigating Memory in People At risk, Causes and Treatments (IMPACT) cohort 176 (Table 1). All participants had a parental family history of sporadic AD (FH+) defined as one or 177 both biological parents meeting AD clinical diagnosis criteria (McKhann et al., 1984, 2011). 178 Parental family history was determined by a validated interview reviewed by a multidisciplinary 179 diagnostic consensus panel (Kawas et al., 1994) or postmortem neuropathological diagnosis of 180 AD. Individuals underwent lumbar puncture, fasting blood draw and comprehensive 181 neuropsychological testing. Inclusion criteria entailed no history of a clinical diagnosis of 182 diabetes or current diabetes treatment. Fasting plasma glucose (FPG) was evaluated to exclude 183 individuals who met FPG criteria for diabetes diagnosis (i.e. FPG > 125 mg/dL). All participants 184 were required to have normal cognitive function, as determined by neuropsychological 185 evaluation and consensus review, and no current diagnosis of major psychiatric illness.

- 186 Participants were defined as *APOE* ε 4 positive if they harbored at least one copy of the ε 4 allele.
- 187 Hetero- and homozygous ɛ4 allele carriers constituted 47% of the sample.
- 188

189 2.2. Design and Procedure

190 To assess insulin resistance and biomarkers of CSF AD pathology, venous blood and CSF were 191 collected in the morning after a 12h fast. Blood samples were collected in 9ml polypropylene 192 tubes, allowed to clot for 30mins and centrifuged at 4*C at 3000 rpm for 10mins. Cell-free 193 plasma/serum was aliquoted into 1.5mL micro centrifuge tubes and frozen at -80 degrees 194 Celsius. Plasma and serum samples were analyzed at the University of Wisconsin Hospital and 195 Clinics Hospital Laboratory (Madison, WI). To assess fasting glucose, plasma was assayed using 196 hexokinase glucose method (Siemens Dimension Vista). To assess fasting insulin, serum was 197 assayed using chemiluminescent immunoassay on an ADVIA Centaur XP Immunoassay System 198 (Siemens Corporation, Washington DC, USA). Insulin resistance was calculated from fasting 199 serum insulin and fasting plasma glucose using the homeostatic model assessment of insulin 200 resistance (HOMA-IR) method (Matthews et al., 1985) calculated as HOMA-IR = Insulin 201 (mg/dL) x Glucose (uIU/mL) / 405.

202

203 CSF was collected through gentle extraction via lumbar puncture using a Sprotte 25- or 24-gauge 204 spinal needle at the L3/4 or L4/5 level of the spinal column. Approximately 22mL of CSF was 205 extracted, gently mixed and centrifuged at 2000g for 10mins. Supernatants were frozen in 206 polypropylene tubes in 0.5mL aliquots and stored at -80 degrees Celsius. CSF was assayed for P-207 tau₁₈₁ and A β_{42} using commercially available enzyme-linked immunosorbent assay (ELISA) 208 methods (INNOTEST assays, Fujiurebio, Ghent Belgium) as previously described in detail 209 (Palmqvist et al., 2014). CSF sAPP-β was measured using the MSD Multiplex Soluble APP 210 assay (Meso Scale Discovery, Rockville, MD), as described by the manufacturer. Board-certified 211 laboratory technicians blinded to clinical information analyzed all samples in accordance to 212 protocols approved by the Swedish Board of Accreditation and Conformity Assessment 213 (SWEDAC). One batch of reagents was used yielding intra-assay coefficients of <10 % 214 variation. The P-tau₁₈₁/A β_{42} ratio was examined as a marker of multiple pathological processes 215 that occur in AD.

217 APOE £4 status was determined using genetic testing by the Wisconsin Alzheimer's Disease 218 Research Center. Genotyping was performed using non-fasting blood sample collected at 219 baseline visit, using standard polymerase chain reaction (PCR) and deoxyribonucleic acid (DNA) 220 sequencing techniques. DNA extracted from whole blood was genotyped with the use of a 221 homogenous Florescent Resonance Energy Transfer technology coupled to competitive allele 222 specific PCR (LGC Genomics; Beverly, MA). The National Cell Repository for Alzheimer's 223 disease (NCRAD) also performed genotyping. There was 100% concordance for APOE 224 genotyping between these analyses.

225

226 Participants underwent a comprehensive neuropsychological battery that included the Rey

227 Auditory Verbal Learning Test (RAVLT) (Rey, 1941) a widely used and well-validated

assessment of memory function. The delayed component of the RAVLT is designed to examine

long-term memory function and is a well-known measure that assesses cognitive changes from

intact, to MCI to AD patients (Estevez-Gonzalez et al., 2003; Ewers et al., 2012). In the current

study the RAVLT delayed task was chosen as a focal index of long-term hippocampal-dependentmemory function.

233

234 2.7. Statistical Analyses

Linear regression analyses were conducted in SPSS (Version 21.0). Analyses tested main effects of IR as indexed by HOMA-IR and *APOE* ɛ4 status, and the interaction between HOMA-IR and *APOE*ɛ4 on CSF biomarkers of AD and memory performance. A secondary analysis tested the interaction between HOMA-IR and CSF biomarkers of AD on delayed memory performance. Age, sex and body mass index (BMI) were included as covariates in all analyses. Education was added as a covariate in analyses that included neuropsychological test measures (i.e., RAVLT delayed) as a dependent variable.

242

243 3. Results

244 3.1. HOMA-IR, APOE ε4 and CSF biomarkers of AD

245 Linear multiple regression analysis revealed that HOMA-IR and APOE ɛ4 status were significant

246 predictors of CSF sAPP-β. Higher HOMA-IR was significantly associated with increased levels

of CSF sAPP- β ($F_{11,631} = 4.21$, p = 0.044) (Figure 1). Further, carriage of the ϵ 4 allele significantly

- 248 predicted higher levels of CSF sAPP- β ($F_{[1,63]} = 7.74$, p = 0.007) (Figures 2). The interaction 249 between HOMA-IR and *APOE* ϵ 4 on CSF sAPP- β was not significant.
- 250 Significant relationships between CSF A β_{42} and predictor variables and CSF P-tau₁₈₁ and predictor
- 251 variables were not found, however, a significant effect of APOE ε4 status on the ratio of CSF P-
- tau₁₈₁/A β_{42} was observed ($F_{[1,63]} = 5.21$, p = 0.026). CSF P-tau₁₈₁/A β_{42} was significantly greater in
- 253 *APOE* ε4 carriers compared to non-carriers (Figure 3).
- 254

255 3.2. HOMA-IR, APOE ε4, CSF biomarkers of AD and Memory Performance

To investigate independent and concomitant effects of IR, genetic risk and AD pathology on memory function, HOMA-IR, *APOE* ε 4 and P-tau₁₈₁/A β ₄₂ were examined with respect to delayed memory score on the RAVLT. Analyses yielded a significant interaction between HOMA-IR and P-tau₁₈₁/A β ₄₂ on memory performance ($F_{[1,60]} = 6.14$, p = 0.016). Higher HOMA-IR and greater P-tau₁₈₁/A β ₄₂ predicted impaired delayed memory performance (Figure 4). No other significant interactions were observed.

262

263 4. Discussion

264 IR is associated with an increased relative risk for developing sporadic late-onset AD.

265 Dysregulation of insulin signaling is thought to contribute to a cascade of neuropathological

changes that promote amyloidogenic processing, neurotoxicity, and brain amyloidosis (Devi et

267 al., 2012; Henneberg and Hoyer, 1995; Ho et al., 2004; Willette et al., 2015; Zhao et al., 2009).

268 Brain changes associated with IR that may underlie or contribute to the pathogenesis of AD are

269 poorly understood, particularly in midlife prior to clinical disease onset. IR is a modifiable risk

270 factor and thus regulation of normal insulin signaling is an important target for early

intervention, one that may be particularly relevant to persons who harbor genetic risk or have a

- 272 parental family history of AD. The current study aimed to investigate the relationship between
- 273 IR, AD pathology and memory performance in middle-aged individuals enriched for AD. As a
- subsidiary aim, we examined APOEE4 genotype relative to IR, AD pathology and memory

function, as it is a well-established genetic risk factor for AD.

276

Our findings revealed that higher IR and carriage of the ε4 allele were both predictors of
increased sAPP-β in CSF, suggesting preferential cleavage of APP through the amyloidogenic β-

279 secretase pathway. Evidence from animal studies shows that IR promotes amyloidogenic β-280 amyloid peptide production and upregulated levels of β-site APP cleaving enzyme 1 (BACE1), a 281 process that generates A β_{42} (Devi, et al., 2012). Although higher IR was associated with an 282 increase in sAPP- β , a relationship between IR and A β_{42} was not observed. It is known that 283 cleavage of APP through the β -secretase pathway results in the formation of sAPP- β and the 284 eventual generation and deposition of A β . While the generation of A β_{42} is complex and the 285 relationship between sAPP- β and A β_{42} is not 1:1, a reasonable prediction would be that an 286 association between higher IR and increased levels of sAPP-β would yield an inverse 287 relationship between IR and A β_{42} Prior research from our group has shown that IR predicts brain 288 amyloid deposition in late middle-aged adults at risk for AD as indexed by [C-11] Pittsburg 289 compound B (PiB) positron emission tomography (Willette et al., 2015). Given that we did not 290 find an effect of IR on A β 42, one could postulate that in the case of IR, A β ₄₂ is overproduced and 291 begins depositing in the brain but mechanisms of clearance through the CSF remain intact and 292 appear to be within normal range during subclinical stages of the disease. The participants 293 studied here were also younger than in Willette et al (2015). Longitudinal follow-up on CSF and 294 amyloid-PET will provide further information for mapping out longitudinal trajectories of 295 amyloid deposition.

296

297 IR may also be linked to pathological processes via amyloidosis by decreasing availability of 298 insulin-degrading enzyme (IDE), a large zinc-binding protease that binds to and degrades insulin. 299 IDE preferentially binds to insulin but also has an affinity for A β proteins. In the absence of IR, 300 IDE is available to bind to and degrade A β proteins. In the case of IR, IDE preferentially binds to 301 the elevated levels of insulin as it aggregates in extracellular space, resulting in less availability 302 for IDE to be allocated toward degradation of A β proteins. Excess amyloid peptides 303 consequently lead to the formation of amyloid plaques, brain degeneration and neuronal 304 dysfunction. Animal studies have provided evidence that selective removal of the IDE gene 305 results in more than a 50% decrease in A β degradation and a significant increase in A β 306 deposition (Farris et al., 2003; Ho et al., 2004). Transgenic mouse models of IR have provided 307 evidence that IDE may not only be affected by IR through interference of IDE-mediated 308 degradation of A^β but may also decrease IDE expression and activity (Ho et al., 2004). Further, 309 recent research has provided evidence that IDE also degrades sAPP- β in the intracellular domain

310 (Edbauer et al., 2002) and that elevated levels of sAPP- β were observed in homozygous deletions 311 of the IDE gene (Farris et al., 2003). These results are aligned with the notion that IR may lead to

312 increased leaves of sAPP- β through mechanisms that involve diminished IDE availability and

313 reduced sAPP-β degradation.

314

315 Carriage of the ɛ4 allele of APOE was also a significant predictor of CSF markers of AD 316 pathology. APOE ε4 status showed a positive relationship with sAPP-β and the ratio of P-317 tau₁₈₁/A β_{42} although significant relationships between APOE ε 4 carriage and A β_{42} or P-tau₁₈₁ 318 were not observed. These results validate the notion that P-tau₁₈₁/A β_{42} may be a more sensitive 319 index of AD pathology in preclinical populations compared to a single marker of disease and 320 provide further evidence that multiple pathological processes associated with AD occurs at a 321 subclinical threshold in asymptomatic midlife persons, particularly $\varepsilon 4$ allele carriers. APOE $\varepsilon 4$ 322 status is a strong predictor of AD pathology and is believed to act on the AB pathway, leading to 323 a reduction in clearance and increased Aβ aggregation and deposition (Schmechel et al., 1993). 324 Our findings provide further support that ɛ4 allele carriage is associated with AD pathology, and 325 extends these findings to increased upstream amyloidogenic processing (i.e., sAPP- β) in healthy 326 middle-aged persons enriched for a family history of AD.

327

328 Interestingly, we found that high IR and P-tau₁₈₁/A β_{42} concomitantly act to impair memory 329 performance on the RAVLT delayed memory. Long-term memory tasks—such as the one 330 examined here—are dependent on the hippocampus, a medial temporal lobe structure negatively 331 affected by IR (Benedict et al., 2012; Rasgon et al., 2011; Stranahan et al., 2008) and among the 332 first regions to show structural and functional changes in AD (Convit et al., 2000; Du et al., 333 2001; Pennanen et al., 2004, Wang et al., 2006). Research from our group has shown that IR in 334 midlife is associated with hypometabolism in brain regions involved in episodic memory, 335 including the hippocampus (Willette et al., 2015). Taken together, our findings suggest that 336 dysregulation of the insulin system in midlife may have deleterious effects on functional 337 integrity of the hippocampus that precede or act in concert with early pathological AD changes 338 leading to subclinical memory dysfunction. It is worth noting that while we found that high IR 339 and AD pathology interacted to impair memory function, we did not observe an interaction with 340 ε4 allele carriage. This finding suggests that while AD pathology is more prominent in APOE ε4

- 341 carriers in midlife, the combined deleterious effects of IR and AD pathology on memory
- 342 performance is observed across carriers and non-carriers alike.
- 343 Overall, our findings provide evidence that pathological processes associated with AD are
- 344 observed early in aging, prior to the onset of memory difficulty or manifestation of clinically
- 345 significant symptoms of cognitive impairment. IR and carriage of the ɛ4 allele are predictors of
- 346 early changes in AD-related CSF biomarkers in persons enriched for parental history of late-
- 347 onset sporadic AD. We show that IR and AD pathology interact to impair long-term memory
- 348 function in middle age. IR is a modifiable risk factor that may be corrected with targeted
- 349 interventions. Insulin homeostasis plays an integral role in maintaining healthy neuronal
- 350 function, normal cerebral glucose metabolism and reducing or ameliorating pathological
- 351 processes associated with AD. Targeted interventions designed to maintain normal insulin
- 352 signally may be particularly affective in delaying or ameliorating AD disease onset in persons
- 353 with invariable risks for AD.

Acknowledgments

This research was supported by NIH grant P50 AG033514, University of Wisconsin Institute for Clinical and Translational Research, funded through a National Center for Research Resources/National Institutes of Health Clinical and Translational Science Award, 1UL1RR025011, a program of the National Center for Research Resources, United States National Institutes of Health, the Swedish Research Council, the Swedish Brain Foundation and Torsten Söderberg's Foundation to the University of Gothenburg. We want to thank the MRI staff at the Wisconsin Institute for Medical Research, Chuck Illingworth, the staff at the Wisconsin ADRC and above all, our dedicated participants. References

1. Arvanitakis, Z., Wilson, R.S., Bienias, J.L., Evans, D.A., Bennett, D.A. (2004). Diabetes mellitus and risk of Alzheimer disease and decline in cognitive function. *Arch Neurol*, *61*(5), 661-666.

2. Ott A, Stolk RP, van Harskamp F, Pols HA, Hofman A, Breteler MM. (1999). Diabetes mellitus and the risk of dementia: the Rotterdam Study. *Neurology*, *53*(9), 1937–1942.

3. Peila R, Rodriguez BL, Launer LJ. (2002). Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies The Honolulu-Asia Aging Study. *Diabetes*, *51*(4), 1256–1262.

4. Schrijvers, E. M. C. W., J.C.M.; Sijbrands, E.J.G.; Hofman, A.; Koudstaal, P.J.; Breteler, M.M.B. (2010). Insulin metabolism and the risk of Alzheimer disease The Rotterdam Study. *Neurology*, *75*, 1982–1987.

5. Starks E.J., O'Grady J.P., Hoscheidt S.M., et al. (2015). Insulin resistance is associated with higher cerebrospinal fluid tau levels in asymptomatic APOE ɛ4 carriers. *J Alzheimers Dis*. doi:10.3233/JAD-150065.

6. Willette, A. A., Bendlin, B. B., Starks, E. J., Birdsill, A. C., Johnson, S. C., Christian, B. T., . . . Asthana, S. (2015). Association of Insulin Resistance With Cerebral Glucose Uptake in Late Middle-Aged Adults at Risk for Alzheimer Disease. *JAMA Neurol*. doi:10.1001/jamaneurol.2015.0613

7. de la Monte, S. M. W., J. (2005). Review of insulin and insulin-like growth factor expression, signaling, and malfunction in the central nervous system: Relevance to Alzheimer's disease. *Journal of Alzheimer's Disease*, 7(1), 45-61.

8. Cersosimo, E., & DeFronzo, R. A. (2006). Insulin resistance and endothelial dysfunction: the road map to cardiovascular diseases. *Diabetes Metab Res Rev*, 22(6), 423-436. doi:10.1002/dmrr.634

9. Zhao, W. Q., Chen, H., Quon, M. J., & Alkon, D. L. (2004a). Insulin and the insulin receptor in experimental models of learning and memory. *Eur J Pharmacol*, 490(1-3), 71-81. doi:10.1016/j.ejphar.2004.02.045

10. Cardoso, S., Correia, S., Santos, R. X., Carvalho, C., Santos, M. S., Oliveira, C. R., . . . Moreira, P. I. (2009). Insulin is a two-edged knife on the brain. *J Alzheimers Dis*, 18(3), 483-507. doi:10.3233/JAD-2009-1155.

11. Zhao, W., Alkon, D.L. (2001). Role of insulin and insulin receptor in learning and memory. *Molecular and Cellular Endocrinology*, 177(1-2), 125-134. doi: 10.1016/S0303-7207(01)00455-5

12. Craft, S., Watson, G.S. (2004). Insulin and neurodegenerative disease: shared and specific mechanisms. Lancet Neurol, 3, 169-178.

13. Henneberg, N., Hoyer, S. (1995). Desensitization of the neuronal insulin receptor: a new approach in the etiopathogenesis of late-onset sporadic dementia of the Alzheimer type (SDAT)? *Archives of Gerontology and Geriatrics*, 21(1), 63-74. doi:http://dx.doi.org/10.1016/0167-4943(95)00646-3

14. Baker, L. D., Cross, D. J., Minoshima, S., Belongia, D., Watson, G. S., & Craft, S. (2011). Insulin resistance and Alzheimer-like reductions in regional cerebral glucose metabolism for cognitively normal adults with prediabetes or early type 2 diabetes. *Arch Neurol*, 68(1), 51-57. doi:10.1001/archneurol.2010.225

15. Tschritter, O., Preissl, H., Hennige, A.M., et al (2006). The cerebrocortical response to hyperinsulinemia is reduced in overweight humans: a magnetoencephalographic study. *Proc Natl Acad Sci U S A*, *103*(32), 12103–12108. doi:10.1073/pnas.0604404103

16. Convit, A., Wolf, O. T., Tarshish, C., & de Leon, M. J. (2003). Reduced glucose tolerance is associated with poor memory performance and hippocampal atrophy among normal elderly. *Proc Natl Acad Sci U S A*, *100*(4), 2019-2022. doi:10.1073/pnas.0336073100

17. Li, X., Song, D., & Leng, S. X. (2015). Link between type 2 diabetes and Alzheimer's disease: from epidemiology to mechanism and treatment. *Clin Interv Aging*, *10*, 549-560. doi:10.2147/CIA.S74042

18. de la Monte, S. M., & Tong, M. (2014). Brain metabolic dysfunction at the core of Alzheimer's disease. *Biochem Pharmacol*, *88*(4), 548-559. doi:10.1016/j.bcp.2013.12.012

19. Nunan, J. S., D.H. (2000). Regulation of APP cleavage by alpha-, beta- and gamma-secretases. *FEBS Letters*, 483(1), 6-10.

20. Vassar, R., Bennett, B.D., Babu-Khan, S., Kahn, S., Mendiaz, E.A., Denis, P., Teplow, D.B., Ross, S., Amarante, P., Loeloff, R., Luo, Y., Fisher, S., Fuller, J., Edenson, S., Lile, J., Jarosinski, M.A., Leona Biere, A., Curran, E., Burgess, T., Louis, J., Collins, F., Treanor, J., Rogers, G., Citron, M. (1999). β-Secretase Cleavage of AlzheimerŐs Amyloid Precursor Protein by the Transmembrane Aspartic Protease BACE. *Science*, *286*, 735-741.

21. Lacor, P. N., Buniel, M. C., Chang, L., Fernandez, S. J., Gong, Y., Viola, K. L., . . . Klein, W. L. (2004). Synaptic targeting by Alzheimer's-related amyloid beta oligomers. *J Neurosci*, *24*(45), 10191-10200. doi:10.1523/JNEUROSCI.3432-04.2004

22. Lacor, P. N., Buniel, M. C., Furlow, P. W., Clemente, A. S., Velasco, P. T., Wood, M., ... Klein, W. L. (2007). Abeta oligomer-induced aberrations in synapse composition, shape, and density provide a molecular basis for loss of connectivity in Alzheimer's disease. *J Neurosci*, 27(4), 796-807. doi:10.1523/JNEUROSCI.3501-06.2007 23. Rowan, M. J., Klyubin, I., Cullen, W. K., & Anwyl, R. (2003). Synaptic plasticity in animal models of early Alzheimer's disease. *Philos Trans R Soc Lond B Biol Sci*, *358*(1432), 821-828. doi:10.1098/rstb.2002.1240

24. Shankar, G. M., Li, S., Mehta, T. H., Garcia-Munoz, A., Shepardson, N. E., Smith, I., . . . Selkoe, D. J. (2008). Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat Med*, *14*(8), 837-842. doi:10.1038/nm1782

25. Lambert, M. P. B., A.K.; Chromy, B.A.; Edwards, C.; Freed, R.; Liosatos, M.; Morgan, T.E.; Rozovsky, I.; Trommer, B.; Viola, K.L.; Wals, P.; Zhang, C.; Finch, C.E.; Krafft, G.A.; Klein, W.L. (1998). Diffusible, nonfibrillar ligands derived from Ab1–42 are potent central nervous system neurotoxins. *Proc Natl Acad Sci USA*, *96*, 6448–6453.

26. Farris, W., Mansourian, S., Chang, Y., Lindsley, L., Eckman, E. A., Frosch, M. P., . . . Guenette, S. (2003). Insulin-degrading enzyme regulates the levels of insulin, amyloid beta-protein, and the beta-amyloid precursor protein intracellular domain in vivo. *Proc Natl Acad Sci U S A*, *100*(7), 4162-4167. doi:10.1073/pnas.0230450100

27. Gasparini L, G. G., Wang R, Gross RS, Beal MF, Greengard P, Xu H. (2001). Stimulation of beta-amyloid precursor protein trafficking by insulin reduces intraneuronal beta-amyloid and requires mitogen-activated protein kinase signaling. *J Neurosci, 21*(8), 2561-2570.

28. Ho, L., Qin, W., Pompl, P., Xiang, Z., Wang, J., Zhao, Z., Peng, Y., Cambareri, G., Rocher, A., Mobbs, C.V., Hof, P.R., Pasinetti, G.M. (2004). Diet-induced insulin resistance promotes amyloidosis in a transgenic mouse model of Alzheimer's disease. *FASEB J*.

29. Reiman EM,ChenK,AlexanderGE,etal. Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia. Proc Natl Acad Sci U S A. 2004;101(1):284-289.

30. Reiman EM,ChenK,AlexanderGE,etal. Correlations between apolipoprotein E ϵ 4 gene dose and brain-imaging measurements of regional hypometabolism. Proc Natl Acad Sci U S A. 2005; 102(23):8299-8302.

31. Small GW, Ercoli LM, Silverman DH, Huang SC, Komo S, Bookheimer SY, Lavretsky H, Miller K, Siddarth P, Rasgon NL, Mazziotta JC, Saxena S, Wu HM, Mega MS, Cummings JL, Saunders AM, Pericak- Vance MA, Roses AD, Barrio JR, Phelps ME (2000) Cerebral metabolic and cognitive decline in persons at genetic risk for Alzheimer's disease. *Proc Natl Acad Sci U S A*, *97*(11), 6037–6042.

32. Corder, E., Saunders, A., Strittmatter, W., Schmechel, D., Gaskell, P., Small, G., . . . Pericak-Vance, M. (1993). Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science*, *261*(5123), 921-923. doi:10.1126/science.8346443 33. Lemaitre H, Crivello F, Dufouil C, et al. (2005). No epsilon(4) gene dose effect on hippocampal atrophy in a large MRI database of healthy elderly subjects. *Neuroimage*, *24*, 1205–1213.

34. Salomon-Zimri, S., Boehm-Cagan, A., Liraz, O., Michaelson, D.M. (2003). Hippocampusrelated cognitive impairments in young apoE4 targeted replacement mice. *Neurodegener Dis*, *13*, 86-92. doi:10.1159/000354777

35. Adluru, N., Destiche, D.J., Lu, S., Doran, S.T., Birdsill, A.C., Melah, K.E., Okonkwo, O.C., Alexander, A.L., Dowling, N.M., Johnson, S.C., Sager, M.A., Bendlin, B.B. (2014). White matter microstructure in late middle-age: Effects of apolipoprotein E4 and parental family history of Alzheimer's disease. *NeuroImage:Clinical*, *4*, 730-742. doi:10.1016/j.nicl.2014.04.008

36. Honea, R. A. S., R.H.; Vidoni, E.D.; Goodwin, J.; Burns, J.M. (2010). Reduced gray matter volume in normal adults with a maternal family history of Alzheimer disease. *Neurology*, *74*, 113–120.

37. Honea, R. A. S., R.H.; Vidoni, E.D.; Burns, J.M. (2011). Progressive regional atrophy in normal adults with a maternal history of Alzheimer disease. *Neurology*, *76*, 822–829.

38. Johnson, S. C., Schmitz, T. W., Trivedi, M. A., Ries, M. L., Torgerson, B. M., Carlsson, C. M., . . . Sager, M. A. (2006). The influence of Alzheimer disease family history and apolipoprotein E epsilon4 on mesial temporal lobe activation. *J Neurosci, 26*(22), 6069-6076. doi:10.1523/JNEUROSCI.0959-06.2006

39. Johnson, S. C., Christian, B. T., Okonkwo, O. C., Oh, J. M., Harding, S., Xu, G., . . . Sager, M. A. (2014). Amyloid burden and neural function in people at risk for Alzheimer's Disease. *Neurobiol Aging*, *35*(3), 576-584. doi:10.1016/j.neurobiolaging.2013.09.028

40. Mosconi, L., Rinne, J. O., Tsui, W. H., Berti, V., Li, Y., Wang, H., . . . de Leon, M. J. (2010). Increased fibrillar amyloid-{beta} burden in normal individuals with a family history of late-onset Alzheimer's. *Proc Natl Acad Sci U S A*, *107*(13), 5949-5954. doi:10.1073/pnas.0914141107

41. Okonkwo, O. C. X., G.; Dowling, N.M.; Bendlin, B.B.; LaRue, A.; Hermann, B.P.; Koscik, R.; Jonaitis, E.; Rowley, H.A.; Carlsson, C.M.; Asthana, S.; Sager, M.A.; Johnson, S.C. (2012). Family history of Alizheimer disease predicts hippocampal atrophy in healthy middle-aged adults. *Neurology*, *78*, 1769-1776.

42. McKhann, G. M. D., D.; Folstein, M.; Katzman, R.; Price, D.; Stadlan, E.M. (1984). Clinical diagnosis of Alzheimer's disease: Report of the NINCDS-ADRDA Work Group* under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology*, *34*, 939-944.

43. McKhann, G. M., Knopman, D. S., Chertkow, H., Hyman, B. T., Jack, C. R., Jr., Kawas, C. H., . . . Phelps, C. H. (2011). The diagnosis of dementia due to Alzheimer's disease:

recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement*, 7(3), 263-269. doi:10.1016/j.jalz.2011.03.005

44. Kawas, C. H. S., J.; Stewart, W.F.; Corrada, M.; Thal, L.J. (1994). A validation study of the Dementia Questionnaire. *Arch Neurol*, *51*, 901-906.

45. Matthews, D. R. H., J.P.; Rudenski, A.S.; Naylor, B.A.; Treacher, D.F.; Turner, R.C. (1985). Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*, *28*, 412-419.

46. Palmqvist, S., Zetterberg, H., Blennow, K., Vestberg, S., Andreasson, U., Brooks, D. J., ... Hansson, O. (2014). Accuracy of brain amyloid detection in clinical practice using cerebrospinal fluid beta-amyloid 42: a cross-validation study against amyloid positron emission tomography. *JAMA Neurol*, *71*(10), 1282-1289. doi:10.1001/jamaneurol.2014.1358

47. Rey, A. (1941). L'examen psychologique dans les cas d'ence phalopathie traumatique. *Archives de Psychologie*, 28, 21

48. Estevez-Gonzalez, A., Kulisevsky, J., Boltes, A., Otermin, P., & Garcia-Sanchez, C. (2003). Rey Verbal Learning Test is a useful tool for differential diagnosis in the pre-clinical phase of Alzheimer's disease: Comparison with mild cognitive impairment in normal aging. *International Journal of Geriatric Psychiatry*, *18*, 1021–1028. doi:10.1002/gps.1010

49. Ewers, M., Walsh, C., Trojanowski, J. Q., Shaw, L. M., Petersen, R. C., Jack, C. R., Jr., . . . North American Alzheimer's Disease Neuroimaging, I. (2012). Prediction of conversion from mild cognitive impairment to Alzheimer's disease dementia based upon biomarkers and neuropsychological test performance. *Neurobiol Aging*, *33*(7), 1203-1214. doi:10.1016/j.neurobiolaging.2010.10.019

50. Devi, L., Alldred, M. J., Ginsberg, S. D., & Ohno, M. (2012). Mechanisms underlying insulin deficiency-induced acceleration of beta-amyloidosis in a mouse model of Alzheimer's disease. *PLoS One*, *7*(3), e32792. doi:10.1371/journal.pone.0032792

51. Willette, A. A., Bendlin, B. B., Starks, E. J., Birdsill, A. C., Johnson, S. C., Christian, B. T., . . . Asthana, S. (2015). Association of Insulin Resistance With Cerebral Glucose Uptake in Late Middle-Aged Adults at Risk for Alzheimer Disease. *JAMA Neurol.* doi:10.1001/jamaneurol.2015.0613

52. Zhao, W. Q., Lacor, P. N., Chen, H., Lambert, M. P., Quon, M. J., Krafft, G. A., & Klein, W. L. (2009). Insulin receptor dysfunction impairs cellular clearance of neurotoxic oligomeric a{beta}. *J Biol Chem*, 284(28), 18742-18753. doi:10.1074/jbc.M109.011015

53. Devi, L., Alldred, M. J., Ginsberg, S. D., & Ohno, M. (2012). Mechanisms underlying insulin deficiency-induced acceleration of beta-amyloidosis in a mouse model of Alzheimer's disease. *PLoS One*, *7*(3), e32792. doi:10.1371/journal.pone.0032792

54. Farris, W., Mansourian, S., Chang, Y., Lindsley, L., Eckman, E. A., Frosch, M. P., . . . Guenette, S. (2003). Insulin-degrading enzyme regulates the levels of insulin, amyloid beta-protein, and the beta-amyloid precursor protein intracellular domain in vivo. *Proc Natl Acad Sci U S A*, *100*(7), 4162-4167. doi:10.1073/pnas.0230450100

55. Ho, L., Qin, W., Pompl, P., Xiang, Z., Wang, J., Zhao, Z., Peng, Y., Cambareri, G., Rocher, A., Mobbs, C.V., Hof, P.R., Pasinetti, G.M. (2004). Diet-induced insulin resistance promotes amyloidosis in a transgenic mouse model of Alzheimer's disease. *FASEB J*.

56. Edbauer, D., Willem, M., Lammich, S., Steiner, H., & Haass, C. (2002). Insulin-degrading enzyme rapidly removes the beta-amyloid precursor protein intracellular domain (AICD). *J Biol Chem*, 277(16), 13389-13393. doi:10.1074/jbc.M111571200

57. Schmechel, D. E. S., A.M.; Strittmatter, W.J.; Crain, B.J.; Hulette, C.M.; Joo, S.H.; Pericak-Vance, M.A.; Goldgaber, D.; Roses, A.D. (1993). Increased amyloid beta-peptide deposition in cerebral cortex as a consequence of apolipoprotein E genotype in late-onset Alzheimer disease. *Proc Natl Acad Sci USA*, *90*, 9649-9653.

58. Benedict, C., Brooks, S. J., Kullberg, J., Burgos, J., Kempton, M. J., Nordenskjold, R., . . . Schioth, H. B. (2012). Impaired insulin sensitivity as indexed by the HOMA score is associated with deficits in verbal fluency and temporal lobe gray matter volume in the elderly. *Diabetes Care*, *35*(3), 488-494. doi:10.2337/dc11-2075

59. Rasgon, N. L., Kenna, H. A., Wroolie, T. E., Kelley, R., Silverman, D., Brooks, J., . . . Reiss, A. (2011). Insulin resistance and hippocampal volume in women at risk for Alzheimer's disease. *Neurobiol Aging*, *32*(11), 1942-1948. doi:10.1016/j.neurobiolaging.2009.12.005

60. Stranahan, A. M., Norman, E. D., Lee, K., Cutler, R. G., Telljohann, R. S., Egan, J. M., & Mattson, M. P. (2008). Diet-induced insulin resistance impairs hippocampal synaptic plasticity and cognition in middle-aged rats. *Hippocampus*, *18*(11), 1085-1088. doi:10.1002/hipo.20470

61. Convit, A. d. A., J.; de Leon, M.J.; Tarshish, C.Y., De Santi, S.; Rusinek, H. (2000). Atrophy of the medial occipitotemporal, inferior, and middle temporal gyri in non-demented elderly predict decline to Alzheimer's disease. *Neurobiol Aging*, *21*, 19-26.

62. Du, A. T. S., N.; Amend, D.; Laakso, M.P.; Hsu, Y.Y.; Jagust, W.J.; Yaffe, K.; Kramer, J.H.; Reed, B.; Norman, D.; Chui, H.C.; Weiner, M.W. (2001). Magnetic resonance imaging of the entorhinal cortex and hippocampus in mild cognitive impairment and Alzheimer's disease. *J Neurol Neurosurg Psychiatry*, *71*, 441-447.

63. Pennanen, C., Kivipelto, M., Tuomainen, S., Hartikainen, P., Hänninen, T., Laakso, M. P., . . . Soininen, H. (2004). Hippocampus and entorhinal cortex in mild cognitive impairment and early AD. *Neurobiology of Aging*, *25*(3), 303-310. doi:10.1016/s0197-4580(03)00084-8

64. Wang, L., Zang, Y., He, Y., Liang, M., Zhang, X., Tian, L., . . . Li, K. (2006). Changes in

hippocampal connectivity in the early stages of Alzheimer's disease: evidence from resting state fMRI. *Neuroimage*, *31*(2), 496-504. doi:10.1016/j.neuroimage.2005.12.033

Table 1

Demographic, glucoregulatory, and genetic data			
Sex	55 female, 15 male		
Age (years)	57.7 ± 5.1		
Education (years)	15.9 ± 2.5		
BMI (kg/m²)	28.66 ± 5.08		
HOMA-IR	2.26 ± 1.24		
Diabetes status			
Normoglycemic (fasting glucose < 100 mg/dL)	61		
Prediabetic (fasting glucose < 126 mg/dL)	9		
APOE genotype			
APOEε4 (hetero- or homozygous)	33 (47%)		

Include sAPP-beta, Abeta42 and P-tau data in this table, and perhaps also have another table with the same data comparing IR with non-IR groups?

Figures



Figure 1. Association between HOMA-IR (log) and CSF sAPP- β across all participants. Units are adjusted for age, sex and BMI.

Figure 2. Comparison of CSF sAPP- β in *APOE* ϵ 4 carriers compared to non-carriers. Error bars represent the standard error of the mean. *Significant at p < 0.01.



Figure 3. Comparison of CSF P-tau₁₈₁/A β_{42} in APOE ϵ 4 carriers compared to non-carriers. Error bars represent the standard error of the mean. *Significant at p < 0.01.



Figure 3. Comparison of CSF P-tau₁₈₁/A β_{42} in *APOE* ϵ 4 carriers compared to non-carriers. Error bars designate standard error of the mean. *Significant at p < 0.01. Figure 4. Interaction between HOMA-IR and CSF P-tau₁₈₁/A β_{42} on delayed memory performance. Memory performance was adjusted for sex, age and education. Shown as the median split of HOMA-IR(log). Individuals with lower IR are shown in blue. Individuals with

higher IR are shown in green.

