

A tour of contemporary color vision research

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Abstract

The study of color vision encompasses many disciplines, including art, biochemistry, biophysics, brain imaging, cognitive neuroscience, color preferences, colorimetry, computer modelling, design, electrophysiology, language and cognition, molecular genetics, neuroscience, physiological optics, psychophysics and physiological optics. Coupled with the elusive nature of the subjective experience of color, this wide range of disciplines makes the study of color as challenging as it is fascinating. This overview of the special issue Color: Cone Opponency and Beyond outlines the state of the science of color, and points to some of the many questions that remain to be answered in this exciting field.

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2 The study of color is perhaps the oldest discipline of psychology. For the ancient Greeks,
3 colors took fifth place after the classical elements of fire, air, water, and earth (see page 28 of
4 Kuehni & Schwarz, 2008). Helmholtz's observation that colors can be created by superposing
5 three primaries (von Helmholtz, 1867), and Maxwell's measurements of the proportions of
6 primary lights (positive and negative) required to make the match (Maxwell, 1860), paved the
7 way to our first scientific understanding of the psychological representation of color (Maxwell,
8 1860). Maxwell's work produced an objective set of rules that govern color mixture (Brainard &
9 Stockman, 2010). Eventually, with direct measurements of the three classes of cone
10 photoreceptors in the normal eye (Bowmaker & Dartnall, 1980; Dartnall, Bowmaker, & Mollon,
11 1983; Schnapf, Kraft, & Baylor, 1987; Roorda & Williams, 1999), and with advances afforded
12 by use of molecular genetics (Nathans, Thomas, & Hogness, 1986), the trichromatic theory of
13 color (Young, 1802; von Helmholtz, 1852; Maxwell, 1855) was firmly established.

14 Much of the spectral information contained in the patterns of photons arriving at the eye is
15 lost: the complex spectral power distribution entering the eye is reduced by the univariant
16 responses of the three classes of cone photoreceptor (Mitchell & Rushton, 1971). The
17 trichromatic code relayed by the relative responses in the three cone classes is the retinal basis
18 for color vision. We can now predict, with reasonable precision, how the three classes of cones
19 react to any given physical stimulus. Yet many mysteries about how that code leads to the
20 perception of color remain.

21 The trichromatic code is transformed at the first postreceptoral synapse between the
22 photoreceptor and bipolar cells aided by lateral feedback (and perhaps feedforward) signals
23 between cones from horizontal cells (Verweij, Hornstein, & Schnapf, 2003). It is at this and
24 subsequent stages that cone-opponency—in which signals are compared across different cone
25 types—comes into play. In this special issue, Segal & Perlman (2018) study the cone-opponent
26 properties of horizontal cells in turtle retina. They show that the strength and specificity of
27 horizontal cell feedback is increased by presence of the known neuromodulator nitric oxide,
28 perhaps as part of a mechanism for sharpening spectral discrimination as light levels increase.

29 It is important to keep in mind that *cone*-opponency – subtractive interactions between
30 cones of different types – is distinct from *color*-opponency, which relates to color appearance

1 and the phenomenological opponency of colors first described systematically by Hering (1878,
2 1920) (and reviewed by Mollon & Jordan, 1997; Shevell & Martin, 2017). *Cone*-opponency can,
3 in principle, occur without any *color* association (e.g., Stromeyer, Kronauer, Ryu, Chaparro, &
4 Eskew, 1995; Stockman, Henning, Anwar, Starba, & Rider, 2018). The same caution applies to
5 the nomenclature of the cones themselves. There has been a long tradition of applying color
6 names to the cone type (red, green, blue), but cones do not correspond to color percepts. For
7 example, the sensitivity peak of the so-called red cone is not in the part of the spectrum we call
8 red, but in the yellow-green. For these reasons, color scientists favor the cone names “L”, “M”,
9 and “S”, referring to the location in the spectrum where each cone type has its peak (long, near
10 566 nm; middle, near 541nm; and short, near 441 nm). The color categories red, green, and blue
11 are the output of the visual system, not the input; relating these categorical responses to natural
12 image statistics is considered by Milojevic et al. (2018). They showed that observers mainly use
13 the average color of a leaf photograph to assign it to a color category and use unique yellow as a
14 decision boundary

15 The two most important classes of opponent interactions in the retina occur between L-
16 cones and M-cones, and between the S-cones and some combination of the L- and M-cones. The
17 L-M signal is thought to be encoded by cone-opponent “midget” bipolar cells, which have
18 chromatic-opponency in the fovea by virtue of having single L-cone or M-cone inputs in the
19 centers of their receptive fields, but perhaps mixed L-cone and M-cone inhibitory surrounds
20 derived from horizontal cells. Midget cells project to the parvocellular layers of the lateral
21 geniculate nucleus (LGN) (reviewed by Solomon & Lennie, 2007; Lee, Martin, & Grünert,
22 2010). In this special issue, Lee, Cooper and Cao (2018) investigated the spatial structure of
23 cone-opponent receptive fields in macaque parafoveal retina and report responses of midget
24 ganglion cells to cone-isolating and red-green chromatic stimuli. Despite wide variation in the
25 relative size of center and surround in midget cells, they found most cells show good functional
26 segregation of L-cone and M-cone inputs (Verweij *et al.*, 2003; Crook, Manookin, Packer, &
27 Dacey, 2011).

28 The other main class of opponent interactions is between S cones and some combination of
29 L and M; this opponency is thought to be encoded by S-cone bipolar cells and the distinctive
30 bistratified “blue-yellow” ganglion cells (Kouyama & Mashak, 1992; Dacey & Packer, 2003)
31 that project to the koniocellular layers of the LGN (reviewed by Hendry & Reid, 2000; Martin &

1 Lee, 2014). Connections between S-cones and midget OFF bipolar cells (Field, Gauthier, Sher,
2 Greschner, Machado, Jepson, Shlens, Gunning, Mathieson, Dabrowski, Paninski, Litke, &
3 Chichilnisky, 2010) may form the basis for a similar but oppositely-signed “yellow-blue” signal.
4 In this issue, Eiber et al (2018) study opponent interactions in blue-on and blue-off cells in
5 marmoset LGN and show that both the timing and strength of opponent S and M/L cone inputs
6 are balanced, making these cells highly selective for color over brightness changes. This result
7 adds to other evidence that, unlike the midget-parvocellular system (which likely serves both
8 high-acuity vision and red-green color vision), the bistratified-koniocellular system is dedicated
9 to detecting color contrast.

10 How color is encoded in the cortex is still a major scientific question. Cataloguing the
11 physiological properties of cortical cells has been an enormous challenge, in part because of the
12 massive expansion in the number of neurons found at successive stages of visual processing.
13 There are about one million retinal ganglion cells in each eye, and a comparable number of cells
14 in each LGN, but over 100 million cells in each hemisphere in V1 the first cortical target of the
15 LGN (Curcio & Allen, 1990; Leuba & Kraftsik, 1994). Across all studies, we have far fewer
16 recordings of V1 color properties as a proportion of the population than we have in retina and
17 LGN, and the fraction drops precipitously for subsequent cortical areas (V2, V3, V4, and inferior
18 temporal cortex). It is tempting to bundle what little we know about the cortex into a tidy story.
19 But given how little data there are, the story is likely to be wrong. With that caveat, we will
20 attempt a summary of what we think we know about the physiology of color vision in the cortex,
21 and then return to the challenge of linking physiology with psychology.

22 The first single-cell recordings in V1 came to the surprising conclusion that in contrast to the
23 LGN, few neurons in V1 appeared to be overtly color tuned (Wiesel & Hubel, 1966; Hubel &
24 Wiesel, 1968). There is now a consensus that the population of cortical neurons in V1 (and
25 almost every other cortical area studied) have tunings that span a broad range of color directions
26 (reviewed by Gegenfurtner, 2003; Solomon & Lennie, 2007). The meaning of this multiplicity of
27 tunings with regards to color vision is not at all clear and may not even be relevant to color
28 perception (Eskew, 2009). A cortical neuron that receives equal input from L, M, and S cones
29 will respond best to achromatic modulations; any degree of imbalance of those three inputs will
30 cause the cell to respond best to some color direction other than the achromatic one.
31 Consequently, the mere fact that cortical cells are tuned to many color directions may simply

1 reflect the near-impossibility of precisely regulating the relative strengths and signs of the cone
2 inputs from retina to LGN to cortex. Meanwhile, among the population of V1 cells is a small
3 subset of neurons that show striking cone-opponency. These cells were originally overlooked
4 because the fraction seemed too small to be relevant (Lennie, Krauskopf, & Sclar, 1990). But
5 subsequent work has pinned down the spatial and temporal structure of the cone inputs of these
6 cells and shown many of them to have the elusive “double-opponent” structure first proposed by
7 Edwin Land in his color constancy models (Conway, 2001). Others of the cone-opponent cells in
8 V1 appear to be “single-opponent”, with simpler, and smaller receptive fields that look much like
9 parvocellular cells (Johnson, Hawken, & Shapley, 2004; Shapley & Hawken, 2011). Thus there
10 appear to be a range of color-responsive neurons in V1. This diversity is now widely
11 acknowledged, and improvements in theoretical and empirical methods of assessing color tuning,
12 as described in this issue by Weller and Horwitz (2018), are helping bridge the gap between
13 physiology and functional role. The diversity of color responses at various stages of the visual
14 cortex suggests that color plays a key role in many aspects of visual behavior. The importance of
15 color to visual computations is curiously recovered by an analysis of the layers in convolutional
16 networks, which shows surprising richness of color-tuned “neurons”, and provides clues for how
17 color information may be used at different stages of cortical (Rafegasa & Vanrell, 2018).

18 Because of technical limitations, a given study can only measure single-cell responses for a
19 tiny fraction of the neurons in a given area. The challenge has been how to relate the findings
20 across studies. The anatomical boundary that separates V1 from the rest of the cerebral cortex is
21 clear to the naked eye, which greatly promoted the study of V1. By contrast, physiological
22 studies outside of V1 were long hampered from a lack of anatomical or functional markers to
23 demarcate the areas (gross anatomical landmarks are not very useful because they vary across
24 individuals). Headway was made following the serendipitous discovery of subcompartments
25 within V1 and V2 using a stain for the enzyme cytochrome oxidase (Wong-Riley, 1979). In V1,
26 the stain uncovers blobs (or patches), each about 200 microns across (Horton & Hubel, 1981),
27 and in V2 it uncovers a system of thin and thick stripes. Within V1, color-tuned neurons are
28 often located within cytochrome oxidase blobs of V1 (Livingstone & Hubel, 1984), and the thin
29 stripes of V2 (summarized in Gegenfurtner, 2003). But the colocalization is not precise: many
30 color-tuned neurons are found outside cytochrome-oxidase regions, and many cells within
31 cytochrome-oxidase regions are not obviously color tuned (Leventhal, Thompson, Liu, Zhou, &

1 Ault, 1995). The cartoon that color is coded by blobs and thin stripes is clearly an over-
2 simplification that might be rather misleading. For example, there is a strong projection linking
3 blob cells with area MT (Boyd & Casagrande, 1999); MT is a region that is universally
4 recognized as specialized for motion processing, with limited role in color. And cats have clear
5 cytochrome oxidase blobs (Boyd & Matsubara, 1996), yet poor color vision. Cytochrome
6 oxidase blobs might be more associated with computations of surfaces, rather than color
7 specifically.

8 Pockets of color-tuned neurons outside of V1 and V2 were discovered first by Zeki (1973)
9 in V4, and then by Komatsu (1998) in inferior temporal cortex (IT), a very large swath of tissue
10 implicated in high-level object vision. However, progress was again hampered by the lack of
11 consistent functional or anatomical markers, and the tendency to extrapolate the function of an
12 entire brain region based on recordings from a very tiny fraction of the population of cells
13 housed there. The debate about the role of V4 in color is especially instructive: following Zeki's
14 initial work on V4, other studies failed to find clear evidence that V4 was any more involved in
15 color processing than V1 or V2 (Schein & Desimone, 1990). As we describe below, this
16 controversy was mostly resolved by experiments that combine fMRI and micro-electrode
17 recording.

18 The advent of fMRI, and its application in non-human primates as a tool for guiding micro-
19 electrode recording, has revolutionized the study of color circuits outside of V1 and V2. The
20 landmarks provided by fMRI relate directly to the computation under study (color), unlike
21 histochemical markers such as cytochrome-oxidase. Moreover, the combination of fMRI and
22 micro-electrode recording bridges the gap between gross cortical organization and microscopic
23 cellular resolution. Results using this approach reveal that area V4 region has a mesoscale
24 organization comprising millimeter-sized color-biased regions, dubbed globs, which are
25 massively enriched for color-tuned neurons, interspersed by inter-glob regions that show much
26 lower rates of color tuning (Conway, Moeller, & Tsao, 2007). These observations have been
27 confirmed by optical-imaging-guided microelectrode recording (Tanigawa, Lu, & Roe, 2010). In
28 retrospect, it seems likely that Zeki happened to target globs, while the later studies targeted
29 inter-globs. FMRI also shows at least four large color-biased domains within inferior temporal
30 cortex (Lafer-Sousa & Conway, 2013) one of which corresponds to the region discovered by
31 Komatsu. These and other fMRI studies of color provide insight into the fundamental organizing

1 principles of inferior temporal cortex, and also present clues to the neural basis for the role of
2 color in visual cognition (Bannert & Bartels, 2017). We are now at the threshold of making
3 questions of color cognition scientifically tractable at the circuit level.

4 A central question in color vision research is how different types of psychophysical color
5 measurements map onto different stages of the neural color processing stream. This is a difficult
6 question to answer, since psychophysical responses typically represent the output of the entire
7 neural system. Nevertheless, some psychophysical measurements can be reasonably linked to
8 particular stages of the processing stream. For example, color matching and the laws of color
9 matching (e.g., Krantz, 1975) can be straightforwardly linked to the univariant properties and
10 spectral sensitivities of the three cone types and color matching functions should be a linear
11 combination of the three cone spectral sensitivities (König & Dieterici, 1886; Brindley, 1960).
12 Moreover, color discrimination experiments can, in principle, be related to the cone-opponent
13 stages. For example, under some conditions an incremental L-cone light becomes more
14 detectable if it combined with a decremental M-cone light (Stromeyer, Cole, & Kronauer, 1985;
15 Cole, Stromeyer, & Kronauer, 1990), which implies a cone-opponent coding. Moreover, the
16 spectral sensitivities of these detection mechanisms are consistent with cone-opponency (i.e., L-
17 M) rather than red-green color opponency (Mollon & Cavonius, 1987). The simple picture of L-
18 M, S-(L+M) and L+M as the main second-stage mechanisms (Krauskopf, Williams, & Heeley,
19 1982) has become more complicated by the identification of additional psychophysical
20 mechanisms (Krauskopf, Williams, Mandler, & Brown, 1986; Stoughton, Lafer-Sousa, Gagin, &
21 Conway, 2012; Hansen & Gegenfurtner, 2013; T. G. Shepard, Swanson, McCarthy, & Eskew,
22 2016). Nevertheless, in one recent paper, Shepard, Lahlaf & Eskew (2017) were able to link six
23 distinct, categorical color percepts to specific detection mechanisms.

24 Although two different stimuli that cause the same triplet of activation in the three classes of
25 cones will appear the same in whatever context they are presented, we cannot predict the color of
26 that triplet, even though we know a fair amount about some of the intermediate stages between
27 cone quantal catch and ultimate appearance. The transformation of cone to cone-opponent
28 signals in the retina was foreshadowed by Hering (1878, 1920), who noted that color pairs such
29 as red versus green, and blue versus yellow are phenomenologically opposed in color perception
30 (there are no reddish greens). He postulated that color is represented by the responses of three
31 different mechanisms each driven by paired color-opponent processes (red/green, blue/yellow

1 and white/black). Hurvich and Jameson (1981) elaborated this observation using a
2 psychophysical procedure known as hue cancellation, from which they inferred spectral
3 sensitivities of red-green and blue-yellow color-opponent mechanisms. However, the spectral
4 sensitivities for these “valence” mechanisms, as they called them, are inconsistent with
5 physiological cone-opponent measurements in the retina and LGN, and with the color
6 discrimination measurements described above (e.g., Guth, 1991; De Valois & De Valois, 1993;
7 Webster, Miyahara, Malkoc, & Raker, 2000; Stockman & Brainard, 2010; Shevell & Martin,
8 2017). This inconsistency is also uncovered by measurements of the unique red, green, yellow
9 and blue colors, which do not appear to relate in a direct way to cone-opponent mechanisms
10 (Wuerger, Atkinson, & Cropper, 2005; Wool, Kombar, Kremkow, Jansen, Li, Alonso, & Zaidi,
11 2015). An important property of the color-opponent mechanisms (as opposed to cone-opponent
12 detection mechanisms) is that even the red-green mechanism has a pronounced S-cone input.
13 Another important property is that – despite the simple textbook picture – the two blue and
14 yellow ‘poles’ of the blue-yellow color-opponent mechanisms do not receive symmetric input
15 from the cones (Wuerger *et al.*, 2005). Thus, the loci of unique red and green (where the blue-
16 yellow response is zero) are not colinear in color space (Dimmick & Hubbard, 1939b, 1939a;
17 Burns, Elsner, Pokorny, & Smith, 1984; Wool *et al.*, 2015). Witzel *et al.* (2018) investigated
18 categorical perception for unique hues in order to test for the relationship between color
19 appearance, color discrimination, and low-level (second-stage) mechanisms. Their findings
20 underscore the point that the four chromatic unique hues cannot be related in a simple way to
21 cone-opponent stages of processing. In terms of psychophysical detection mechanisms, the S+
22 and S- mechanisms have different contrast response properties as demonstrated in this issue by
23 Gabree, Shepard and Eskew (2018).

24 A great deal is still uncertain about other aspects of color appearance. Although we have a
25 conviction that colors are ordered, we have little understanding of the rules that govern the
26 geometry of the space that represents color appearance. Colors can be defined by lightness
27 (value), color purity (saturation), and hue, but these properties interact. There is still no clear
28 empirical or theoretical way of relating these dimensions to each other. To address this
29 fundamental question, Schiller, Valsecchi and Gegenfurtner (2018) evaluated different measures
30 of color saturation. CIELUV, CIELAB, and CIECAM02 provided the best metrics of perception.
31 Many other aspects of color remain unexplained. One of the most pressing questions concerns

1 the relationship of color and shape processing. This question has been addressed by evaluating
2 how luminance edges influence color perception, such as in the watercolor effect, where colors
3 appear to spread from luminance boundaries into nearby regions (Pinna, Brelstaff, & Spillmann,
4 2001). Coia and Crognale (2018) show that contour adaptation reduces the spreading of edge
5 induced colors, which provides clues to the neural mechanisms that integrate color and shape
6 information.

7 Although we can measure neural responses to modulations in cone activity, we do not know
8 what stimuli to use to relate the neural activity to computational mechanisms (Sanada, Namima,
9 & Komatsu, 2016). There is even uncertainty at Marr-Level 1, regarding the computational goal
10 of color: why do we have color vision at all? Some evidence suggests that color may aid in
11 storing and remembering objects (Gegenfurtner & Rieger, 2000), although to a broad first-order
12 approximation color provides little benefit for object recognition. Indeed, most studies of object
13 vision use achromatic images. Perhaps the role of color extends beyond object recognition.
14 Science has largely overlooked one of the most important facts of all about color: color is
15 attractive. Our desire for color has occasionally fueled wars and continues to drive technological
16 developments such as those for high-gamut displays and LEDs. Color preferences are well
17 documented, but their origin remains a mystery. In order to address this broader question,
18 Schloss et al. (2018) modelled color preferences using a variety of color space metrics. They
19 found that the best model was one specified in CIELAB space that included 1st and 2nd
20 harmonics of hue. These results are interesting, in part because they relate to quantitative
21 analysis of color-tuning responses at mid-tier stages of visual processing. This analysis shows
22 that the V4 Complex encodes a uniform representation of color space best captured by CIELAB,
23 providing a link between physiology and perception (Bohon, Hermann, Hansen, & Conway,
24 2016), and now possibly preferences.

25 A plethora of other colorful topics graces this issue. Rucker et al. (2018) studied the role of
26 temporal contrast and blue light on emmetropization (the critical period of eye growth that
27 results in a focused retinal image). Dore et al. (2018) in a multi-faceted investigation examine
28 associations between global and local shape perception, colored backgrounds, color
29 discrimination, and non-verbal IQ. Koenderink, van Doorn and Gegenfurtner (2018)
30 investigated color weights in photometry by going beyond the simple heterochromatic methods,
31 the results of which comply with the CIE luminance, to tasks that involve mid-level and high-

1 level aspects of perception. These studies all help fill in gaps about the psychology of color, and
2 they will guide physiological investigations aimed at understanding the neural mechanisms that
3 support color behavior.

4 One theme that emerges from the collection of the papers in this special issue is that
5 studying color is not easy. Why is it so challenging to unlock color's secrets? One reason is that
6 color is a 'proper sensible' (Aristotle, 350 B.C.E.), meaning that we only have access to color
7 through our eyes. We can learn something about color through language, and blind people can
8 acquire a rather sophisticated knowledge of color relationships by listening to others describe
9 color (R. N. Shepard & Cooper, 1992). However, unlike object shape, which can be known by
10 both touch and sight, color can only be experienced through vision. This aspect of color
11 introduces a challenge because at a fundamental level color is a private experience. The
12 subjective nature of color can hinder scientific inquiry, but it adds much to its allure. And
13 through the study of color we hope to glean insight into fundamental aspects of human
14 experience.

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REFERENCES

- 1
2
3
4
5 Aristotle. (350 B.C.E.). *On Sense and the Sensible* (J. I. Beare, Trans.): Internet Classics
6 Archive.
- 7 Bannert, M. M., & Bartels, A. (2017). Invariance of surface color representations across
8 illuminant changes in the human cortex. *Neuroimage*, *158*, 356-370.
- 9 Bohon, K. S., Hermann, K. L., Hansen, T., & Conway, B. R. (2016). Representation of
10 Perceptual Color Space in Macaque Posterior Inferior Temporal Cortex (the V4
11 Complex). *eNeuro*, *3*(4).
- 12 Bowmaker, J. K., & Dartnall, H. J. A. (1980). Visual pigments of rods and cones in the human
13 retina. *Journal of Physiology*, *298*, 501-512.
- 14 Boyd, J. D., & Casagrande, V. A. (1999). Relationships between cytochrome oxidase (CO) blobs
15 in primate primary visual cortex (V1) and the distribution of neurons projecting to the
16 middle temporal area (MT). *Journal of Comparative Neurology*, *409*(4), 573-591.
- 17 Boyd, J. D., & Matsubara, J. A. (1996). Laminar and columnar patterns of geniculocortical
18 projections in the cat: relationship to cytochrome oxidase. *Journal of Comparative
19 Neurology*, *365*(4), 659-682.
- 20 Brainard, D. H., & Stockman, A. (2010). Colorimetry. In M. Bass, C. DeCusatis, J. Enoch, V.
21 Lakshminarayanan, G. Li, C. Macdonald, V. Mahajan & E. van Stryland (Eds.), *The
22 Optical Society of America Handbook of Optics, 3rd edition, Volume III: Vision and
23 Vision Optics* (pp. 10.11-10.56). New York: McGraw Hill.
- 24 Brindley, G. S. (1960). Two more visual theorems. *Quarterly Journal of Experimental
25 Psychology*, *12*, 110-112.
- 26 Burns, S. A., Elsner, A. E., Pokorny, J., & Smith, V. C. (1984). The Abney effect: chromaticity
27 coordinates of unique and other constant hues. *Vision Research*, *24*(5), 479-489.
- 28 Coia, A. J., & Crognale, M. A. (2018). Contour adaptation reduces the spreading of edge induced
29 colors. *Vision Research*, *XXX*, XXX-XXX.
- 30 Cole, G. R., Stromeyer, C. F., III, & Kronauer, R. E. (1990). Visual interactions with luminance
31 and chromatic stimuli. *Journal of the Optical Society of America A*, *7*(1), 128-140.

- 1 Conway, B. R. (2001). Spatial structure of cone inputs to color cells in alert macaque primary
2 visual cortex (V-1). *Journal of Neuroscience*, *21*(8), 2768-2783.
- 3 Conway, B. R., Moeller, S., & Tsao, D. Y. (2007). Specialized color modules in macaque
4 extrastriate cortex. *Neuron*, *56*(3), 560-573.
- 5 Crook, J. D., Manookin, M. B., Packer, O. S., & Dacey, D. M. (2011). Horizontal cell feedback
6 without cone type-selective inhibition mediates "red-green" color opponency in midget
7 ganglion cells of the primate retina. *Journal of Neuroscience*, *31*(5), 1762-1772.
- 8 Curcio, C. A., & Allen, K. A. (1990). Topography of ganglion cells in human retina. *Journal of*
9 *Comparative Neurology*, *300*(1), 5-25.
- 10 Dacey, D. M., & Packer, O. S. (2003). Colour coding in the primate retina: diverse cell types and
11 cone-specific circuitry. *Current Opinion in Neurobiology*, *13*(4), 421-427.
- 12 Dartnall, H. J. A., Bowmaker, J. K., & Mollon, J. D. (1983). Human visual pigments:
13 microspectrophotometric results from the eyes of seven persons. *Proceedings of the*
14 *Royal Society of London. Series B*, *220*(1218), 115-130.
- 15 De Valois, R. L., & De Valois, K. K. (1993). A multi-stage color model. *Vision Research*, *33*(8),
16 1053-1065.
- 17 Dimmick, F. L., & Hubbard, M. R. (1939a). The spectral location of psychologically unique red.
18 *American Journal of Psychology*, *52*, 348-353.
- 19 Dimmick, F. L., & Hubbard, M. R. (1939b). The spectral location of psychologically unique
20 yellow, green and blue. *American Journal of Psychology*, *52*, 242-254.
- 21 Dore, P., Dumani, A., Wyatt, G., & Shepherd, A. J. (2018). Links between global and local
22 shape perception, coloured backgrounds, colour discrimination, and non-verbal IQ.
23 *Vision Research*, *XXX*, XXX-XXX.
- 24 Eiber, C. D., Pietersen, A. N. J., Zeater, N., Solomon, S. G., & Martin, P. R. (2018). Chromatic
25 summation and receptive field properties of blue-on and blue-off cells in marmoset lateral
26 geniculate nucleus. *Vision Research*, *XXX*, XXX-XXX.
- 27 Eskew, R. T., Jr. (2009). Higher order color mechanisms: A critical review. *Vision Research*,
28 *49*(22), 2686-2704.
- 29 Field, G. D., Gauthier, J. L., Sher, A., Greschner, M., Machado, T. A., Jepson, L. H., Shlens, J.,
30 Gunning, D. E., Mathieson, K., Dabrowski, W., Paninski, L., Litke, A. M., &

- 1 Chichilnisky, E. J. (2010). Functional connectivity in the retina at the resolution of
2 photoreceptors. *Nature*, 467(7316), 673-677.
- 3 Gabree, S. H., Shepard, T. G., & Eskew, R. T., Jr. (2018). Asymmetric high-contrast masking in
4 S cone increment and decrement pathways. *Vision Research*, XXX, XXX-XXX.
- 5 Gegenfurtner, K. R. (2003). Cortical mechanisms of colour vision. *Nature Neuroscience*, 4(7),
6 563-572.
- 7 Gegenfurtner, K. R., & Rieger, J. (2000). Sensory and cognitive contributions of color to the
8 recognition of natural scenes. *Current Biology*, 10(13), 805-808.
- 9 Guth, S. L. (1991). A model for color and light adaptation. *Journal of the Optical Society of*
10 *America A*, 8(6), 976-993.
- 11 Hansen, T., & Gegenfurtner, K. R. (2013). Higher order color mechanisms: Evidence from
12 noise-masking experiments in cone contrast space. *Journal of Vision*, 13(1), 26.21-21.
- 13 Hendry, S. H. C., & Reid, R. C. (2000). The koniocellular pathway in primate vision. *Annual*
14 *Review of Neuroscience*, 23, 127-153.
- 15 Hering, E. (1878). *Zur Lehre vom Lichtsinne. Sechs Mittheilungen an die Kaiserliche Akademie*
16 *der Wissenschaften in Wien*. Wien: Carl Gerold's Sohn.
- 17 Hering, E. (1920). *Grundzüge der Lehre vom Lichtsinn*. Springer: Berlin.
- 18 Horton, J. C., & Hubel, D. H. (1981). Regular patchy distribution of cytochrome oxidase staining
19 in primary visual cortex of macaque monkey. *Nature*, 292(5825), 762-764.
- 20 Hubel, D. H., & Wiesel, T. N. (1968). Receptive fields and functional architecture of monkey
21 striate cortex. *Journal of Physiology*, 195(1), 215-243.
- 22 Hurvich, L. M. (1981). *Color Vision*. Sunderland, Massachusetts: Sinauer.
- 23 Johnson, E. N., Hawken, M. J., & Shapley, R. (2004). Cone inputs in macaque primary visual
24 cortex. *Journal of Neurophysiology*, 91(6), 2501-2514.
- 25 Koenderink, J., van Doorn, A., & Gegenfurtner, K. (2018). Color weight photometry. *Vision*
26 *Research*, XXX, XXX-XXX.
- 27 Komatsu, H. (1998). Mechanisms of central color vision. *Current Opinion in Neurobiology*, 8(4),
28 503-508.
- 29 König, A., & Dieterici, C. (1886). Die Grundempfindungen und ihre Intensitäts-Vertheilung im
30 Spectrum. *Sitzungsberichte Akademie der Wissenschaften in Berlin*, 1886, 805-829.

- 1 Kouyama, N., & Mashak, D. W. (1992). Bipolar cells specific for blue cones in the macaque
2 retina. *Journal of Neuroscience*, *12*(4), 1233-1252.
- 3 Krantz, D. H. (1975). Color measurement and color theory: I. Representation theorem for
4 Grassmann structures. *Journal of Mathematical Psychology*, *12*, 283-303.
- 5 Krauskopf, J., Williams, D. R., & Heeley, D. W. (1982). Cardinal directions of color space.
6 *Vision Research*, *22*(9), 1123-1131.
- 7 Krauskopf, J., Williams, D. R., Mandler, M. B., & Brown, A. M. (1986). Higher order color
8 mechanisms. *Vision Research*, *26*(1), 23-32.
- 9 Kuehni, R. G., & Schwarz, A. (2008). *Color ordered : a survey of color order systems from*
10 *antiquity to the present*. Oxford ; New York: Oxford University Press.
- 11 Lafer-Sousa, R., & Conway, B. R. (2013). Parallel, multi-stage processing of colors, faces and
12 shapes in macaque inferior temporal cortex. *Nature Neuroscience*, *16*(12), 1870-1878.
- 13 Lee, B. B., Cooper, B., & Cao, D. (2018). The spatial structure of cone-opponent receptive fields
14 in macaque retina. *Vision Research*, *XXX*, XXX-XXX.
- 15 Lee, B. B., Martin, P. R., & Grünert, U. (2010). Retinal connectivity and primate vision.
16 *Progress in Retinal and Eye Research*, *29*(6), 622-639.
- 17 Lennie, P., Krauskopf, J., & Sclar, G. (1990). Chromatic mechanisms in striate cortex of
18 macaque. *Journal of Neuroscience*, *10*(2), 649-669.
- 19 Leuba, G., & Kraftsik, R. (1994). Changes in volume, surface estimate, three-dimensional shape
20 and total number of neurons of the human primary visual cortex from midgestation until
21 old age. *Anatomy and Embryology*, *190*(4), 351-366.
- 22 Leventhal, A. G., Thompson, K. G., Liu, D., Zhou, Y., & Ault, S. J. (1995). Concomitant
23 sensitivity to orientation, direction, and color of cells in layers 2, 3, and 4 of monkey
24 striate cortex. *Journal of Neuroscience*, *15*(3 Pt 1), 1808-1818.
- 25 Livingstone, M. S., & Hubel, D. H. (1984). Anatomy and physiology of a color system in the
26 primate visual cortex. *Journal of Neuroscience*, *4*(1), 309-356.
- 27 Martin, P. R., & Lee, B. B. (2014). Distribution and specificity of S-cone (“blue cone”) signals in
28 subcortical visual pathways. *Visual Neuroscience*, *31*(Special Issue 02), 177-187.
- 29 Maxwell, J. C. (1855). Experiments on colours, as perceived by the eye, with remarks on colour-
30 blindness. *Transactions of the Royal Society of Edinburgh*, *21*(), 275-298.

- 1 Maxwell, J. C. (1860). On the theory of compound colours and the relations of the colours of the
2 spectrum. *Philosophical Transactions of the Royal Society of London*, 150, 57-84.
- 3 Milojevic, Z., Ennis, R., Toscani, M., & Gegenfurtner, K. R. (2018). Categorizing natural color
4 distributions. *Vision Research*, XXX, XXX-XXX.
- 5 Mitchell, D. E., & Rushton, W. A. H. (1971). Visual pigments in dichromats. *Vision Research*,
6 11(10), 1033-1043.
- 7 Mollon, J. D., & Cavonius, C. R. (1987). The chromatic antagonisms of opponent-process theory
8 are not the same as those revealed in studies of detection and discrimination. In G.
9 Verriest (Ed.), *Colour Deficiencies VIII, Documenta Ophthalmologica Proceedings*
10 *Series*, 46 (pp. 473-483). Dordrecht: Nijhoff-Junk.
- 11 Mollon, J. D., & Jordan, G. (1997). On the nature of unique hues. In C. M. Dickinson, I. J.
12 Murray & D. Carden (Eds.), *John Dalton's colour vision legacy* (pp. 381–392). London:
13 Taylor and Francis.
- 14 Nathans, J., Thomas, D., & Hogness, S. G. (1986). Molecular genetics of human color vision:
15 The genes encoding blue, green and red pigments. *Science*, 232(4747), 193-202.
- 16 Pinna, B., Brelstaff, G., & Spillmann, L. (2001). Surface color from boundaries: a new
17 'watercolor' illusion. *Vision Research*, 41(20), 2669-2676.
- 18 Rafegasa, I., & Vanrell, M. (2018). Color encoding in biologically-inspired convolutional neural
19 networks. *Vision Research*, XXX, XXX-XXX.
- 20 Roorda, A., & Williams, D. R. (1999). The arrangement of the three cone classes in the living
21 human eye. *Nature*, 397(6719), 520-522.
- 22 Rucker, F., Henriksen, M., Yanase, T., & Taylor, C. (2018). The role of temporal contrast and
23 blue light in emmetropization. *Vision Research*, XXX, XXX-XXX.
- 24 Sanada, T. M., Namima, T., & Komatsu, H. (2016). Comparison of the color selectivity of
25 macaque V4 neurons in different color spaces. *Journal of Neurophysiology*, 116(5),
26 2163-2172.
- 27 Schein, S. J., & Desimone, R. (1990). Spectral properties of V4 neurons in the macaque. *Journal*
28 *of Neuroscience*, 10(10), 3369-3389.
- 29 Schiller, F., Valsecchi, M., & Gegenfurtner, K. R. (2018). An evaluation of different measures of
30 color saturation. *Vision Research*, XXX, XXX-XXX.

- 1 Schloss, K. B., Lessard, L., Racey, C., & Hurlbert, A. C. (2018). Modeling color preference
2 using color space metrics. *Vision Research*, *XXX*, XXX-XXX.
- 3 Schnapf, J. L., Kraft, T. W., & Baylor, D. A. (1987). Spectral sensitivity of human cone
4 photoreceptors. *Nature*, *325*(6103), 439-441.
- 5 Segal, E., & Perlman, I. (2018). The role of nitric oxide in spectral information processing in the
6 distal turtle retina. *Vision Research*, *XXX*, XXX-XXX.
- 7 Shapley, R., & Hawken, M. J. (2011). Color in the Cortex: single- and double-opponent cells.
8 *Vision Research*, *51*(7), 701-717.
- 9 Shepard, R. N., & Cooper, L. A. (1992). Representation of Colors in the Blind, Color-Blind, and
10 Normally Sighted. *Psychological Science*, *3*(2), 97-104.
- 11 Shepard, T. G., Lahlaf, S. I., & Eskew, R. T., Jr. (2017). Labeling the lines: A test of a six-
12 mechanism model of chromatic detection. *Journal of Vision*, *17*(13), 9.1-9.18.
- 13 Shepard, T. G., Swanson, E. A., McCarthy, C. L., & Eskew, R. T., Jr. (2016). A model of
14 selective masking in chromatic detection. *Journal of Vision*, *16*(9), 3.1-3.17.
- 15 Shevell, S. K., & Martin, P. R. (2017). Color opponency: tutorial. *Journal of the Optical Society*
16 *of America A*, *34*(7), 1099-1108.
- 17 Solomon, S. G., & Lennie, P. (2007). The machinery of colour vision. *Nature Reviews*
18 *Neuroscience*, *8*(4), 276-286.
- 19 Stockman, A., & Brainard, D. H. (2010). Color vision mechanisms. In M. Bass, C. DeCusatis, J.
20 Enoch, V. Lakshminarayanan, G. Li, C. Macdonald, V. Mahajan & E. van Stryland
21 (Eds.), *The Optical Society of America Handbook of Optics, 3rd edition, Volume III:*
22 *Vision and Vision Optics* (pp. 11.11-11.104). New York: McGraw Hill.
- 23 Stockman, A., Henning, G. B., Anwar, S., Starba, R., & Rider, A. (2018). Delayed cone-
24 opponent signals in the luminance pathway. *Journal of Vision*, *18*(2), 6.1-6.35.
- 25 Stoughton, C. M., Lafer-Sousa, R., Gagin, G., & Conway, B. R. (2012). Psychophysical
26 chromatic mechanisms in macaque monkey. *Journal of Neuroscience*, *32*(43), 15216-
27 15226.
- 28 Stromeyer, C. F., III, Cole, G. R., & Kronauer, R. E. (1985). Second-site adaptation in the red-
29 green chromatic pathways. *Vision Research*, *25*(2), 219-237.

- 1 Stromeier, C. F., III, Kronauer, R. E., Ryu, A., Chaparro, A., & Eskew, R. T., Jr. (1995).
2 Contributions of human long-wave and middle-wave cones to motion detection. *Journal*
3 *of Physiology*, 485, 221-243.
- 4 Tanigawa, H., Lu, H. D., & Roe, A. W. (2010). Functional organization for color and orientation
5 in macaque V4. *Nature Neuroscience*, 13(12), 1542-1548.
- 6 Verweij, J., Hornstein, E. P., & Schnapf, J. L. (2003). Surround antagonism in macaque cone
7 photoreceptors. *Journal of Neuroscience*, 23(32), 10249-10257.
- 8 von Helmholtz, H. (1852). On the theory of compound colours. *Philosophical Magazine Series*
9 4, 4, 519-534.
- 10 von Helmholtz, H. (1867). *Handbuch der Physiologischen Optik*. Hamburg and Leipzig: Voss.
- 11 Webster, M. A., Miyahara, E., Malkoc, G., & Raker, V. E. (2000). Variations in normal color
12 vision. II. Unique hues. *Journal of the Optical Society of America A*, 17(9), 1545-1555.
- 13 Weller, J. B., & Horwitz, G. D. (2018). Measurements of neuronal color tuning: Procedures,
14 pitfalls, and alternatives. *Vision Research*, XXX, XXX-XXX.
- 15 Wiesel, T. N., & Hubel, D. (1966). Spatial and chromatic interactions in the lateral geniculate
16 body of the rhesus monkey. *Journal of Neurophysiology*, 29(6), 1115-1156.
- 17 Witzel, C., & Gegenfurtner, K. (2018). Are red, yellow, green, and blue perceptual categories?
18 *Vision Research*, XXX, XXX-XXX.
- 19 Wong-Riley, M. (1979). Columnar cortico-cortical interconnections within the visual system of
20 the squirrel and macaque monkeys. *Brain Research*, 162(2), 201-217.
- 21 Wool, L. E., Komban, S. J., Kremkow, J., Jansen, M., Li, X., Alonso, J. M., & Zaidi, Q. (2015).
22 Saliency of unique hues and implications for color theory. *Journal of Vision*, 15(2), 10,
23 11-11.
- 24 Wuerger, S. M., Atkinson, P., & Cropper, S. (2005). The cone inputs to the unique-hue
25 mechanisms. *Vision Research*, 45(25-26), 3210-3223.
- 26 Young, T. (1802). The Bakerian Lecture: On the theory of light and colours. *Philosophical*
27 *Transactions of the Royal Society of London*, 92, 12-48.
- 28 Zeki, S. M. (1973). Colour coding in rhesus monkey prestriate cortex. *Brain Research*, 53(2),
29 422-427.