



University of Dundee

Mechanistic framework to link root growth models with weather and soil physical properties, including example applications to soybean growth in Brazil

De Moraes, Moacir Tuzzin; Bengough, A. Glyn; Debiasi, Henrique; Franchini, Julio Cezar; Levien, Renato; Schnepf, Andrea

Published in: Plant and Soil

DOI: 10.1007/s11104-018-3656-z

Publication date: 2018

Document Version Peer reviewed version

Link to publication in Discovery Research Portal

Citation for published version (APA):

De Moraes, M. T., Bengough, A. G., Debiasi, H., Franchini, J. C., Levien, R., Schnepf, A., & Leitner, D. (2018). Mechanistic framework to link root growth models with weather and soil physical properties, including example applications to soybean growth in Brazil. Plant and Soil, 428(1-2), 67-92. https://doi.org/10.1007/s11104-018-3656-z

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

• Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.

You may not further distribute the material or use it for any profit-making activity or commercial gain.
You may freely distribute the URL identifying the publication in the public portal.

Take down policy If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

1 Mechanistic framework to link root growth models with weather and soil 2 physical properties, including example applications to soybean growth in

3 4

5

6

7 8

9 10

Moacir Tuzzin de Moraes, A. Glyn Bengough, Henrique Debiasi, Julio Cezar Franchini, Renato Levien, Andrea Schnepf and Daniel Leitner

Keywords crop model; root growth model; soil compaction; soil strength; crop yield

Abstract

Brazil

11 12

Background and aims Root elongation is generally limited by a combination of mechanical impedance and water stress in most arable soils. However, dynamic changes of soil penetration resistance with soil water content are rarely included in models for predicting root growth. Better modelling frameworks are needed to understand root growth interactions between plant genotype, soil management, and climate. Aim of paper is to describe a new model of root elongation in relation to soil physical characteristics like penetration resistance, matric potential, and hypoxia.

Methods A new diagrammatic framework is proposed to illustrate the interaction between root
 elongation, soil management, and climatic conditions. The new model was written in Matlab®,
 using the root architecture model RootBox and a model that solves the 1D Richards equations
 for water flux in soil. *Inputs:* root architectural parameters for Soybean; soil hydraulic properties;
 root water uptake function in relation to matric flux potential; root elongation rate as a function of
 soil physical characteristics. *Simulation scenarios:* (a) compact soil layer at 16 to 20 cm; (b) test
 against a field experiment in Brazil during contrasting drought and normal rainfall seasons.

Results (a) Soil compaction substantially slowed root growth into and below the compact layer.
(b) Simulated root length density was very similar to field measurements, which was influenced greatly by drought. The main factor slowing root elongation in the simulations was evaluated using a stress reduction function.

31 Conclusion The proposed framework offers a way to explore the interaction between soil 32 physical properties, weather and root growth. It may be applied to most root elongation models, 33 and offers the potential to evaluate likely factors limiting root growth in different soils and tillage 34 regimes. 35

Introduction

36 37

38 39 Root growth, and root elongation in particular, can be limited by many factors in the soil 40 environment, including plant pathogens, mineral element toxicities or deficiencies (Foy 1992), 41 temperature (Licht and Al-Kaisi 2005), water availability (Bengough et al. 2011) aeration 42 (Valentine et al. 2012), and soil strength (Bengough 1997). In terms of physical limitations to 43 root growth, water stress (too little water for root growth), hypoxia or anoxia (too little or no 44 oxygen), and mechanical impedance (soil that is too hard for roots to penetrate rapidly) are 45 often major causes of poor root system growth and development (Bengough et al. 2011). 46 Various processes and mechanisms are involved in maintaining root elongation under water 47 stress, such as osmotic adjustment and enhanced cell wall loosening (Wu and Cosgrove 2000; 48 Schmidt et al. 2013). Penetrometer resistance in excess of 2 MPa is generally thought to 49 present a substantial limitation to root elongation rates, and has been used as a simple 50 threshold for characterising soil physical quality (Taylor et al. 1966; Silva et al. 1994; Lipiec et 51 al. 2012). Penetration resistances of >2 MPa, may occur even in relatively moist arable soils, 52 and frequently slow down root elongation to less than half of its unimpeded rate if continuous 53 cracks or macropores aren't available as low-resistance pathways (Bengough et al. 2011).

There is a need to develop predictive frameworks for understanding plant genotype, soil management, and climate interactions with root system growth: A major problem in applying laboratory-based understanding of root growth is that soil matric potential in the field changes constantly, and can vary spatially and temporally through the soil profile (Bengough 2006). Soil physical stresses and their degree of limitation to root elongation vary greatly between soil types, soil management regimes, and individual growth seasons – making very difficult to predict the effects of agronomic practices and climate on root growth, without using modelling tools.

62 Root water uptake is a major component of the terrestrial hydrological cycle. Macroscopic 63 models can estimate transpiration rates under limiting hydraulic conditions using spatial 64 averages of soil and root properties (de Jong van Lier et al. 2008). Empirical models have been 65 used to estimate the reduction function of water uptake due to the matric potential (Feddes et al. 66 1978) in many simulation models such as Soil-Water-Atmosphere-Plant (SWAP) (Kroes et al. 67 2008) and Hydrus (Simunek and Hopmans 2009). However, the macroscopic root water uptake 68 function should include the preferential uptake from wetter layers using root characteristics (root 69 length density and root diameter) and soil hydraulic status, specifically, the matric flux potential 70 (de Jong van Lier et al. 2008).

71 To simulate the root architecture distribution, many models still consider simple empirical 72 models (e.g. assuming an exponential root length distribution over depth, e.g. Jones et al. 73 (1991)), or density based root models (Kalogiros et al. 2016). Simplistic models that consider 74 only rooting depth to determine the root water uptake (Hartmann et al. 2017) are rarely related 75 to meaningful descriptions of the effect of soil properties on the root length density profile 76 though time. Thus, in order to improve the macroscopic root water uptake (de Jong van Lier et 77 al. 2013) in the soil water flux models (Tron et al. 2015), 3D root architectural models should be 78 used to take into account dynamic development of root structure (Leitner et al. 2010a; Schnepf 79 et al. 2017), and its interaction with soil properties.

80 There are many different root system models (Dunbabin et al. 2013), that can be divided 81 into pure root growth models (Hartmann and Šimunek 2016), which focus on describing the root 82 system's topology (Pagès et al. 2004), and more holistic models, which include several root-83 environment interaction processes (Javaux et al. 2008; Leitner et al. 2010a). In summary, the 84 most common and current models include RootTyp (Pagès et al. 2004), SimRoot (Lynch et al. 85 1997), Rootmap (Diggle 1988b, a), SPACSYS (Wu et al. 2007), R-SWMS (Javaux et al. 2008), 86 Archisimple (Pagès et al. 2014), OpenSimroot (Postma et al. 2017), RootBox (Leitner et al. 87 2010a, b), and CrootBox (Schnepf et al. 2017) which have been used for a range of root 88 modelling studies (Dunbabin et al. 2013).

The decision to use each root growth model should be related to computer power (Dupuy
et al. 2010), availability of input parameters (Bengough 1997) or requirement of the mechanistic
understanding of the soil-root interactions (Dunbabin et al. 2013). In most of the root growth
models soil strength dynamic effects on root elongation have not been considered.

93 Mathematical modelling continues to play an important role in our understanding of root 94 growth and plant water uptake (Schnepf et al. 2012), and further improvement of modelling soil 95 processes is necessary to predict effects on ecosystem services such as food production 96 (Vereecken et al. 2016). RootBox is a widely available dynamic 3D root architecture model 97 based on L-Systems¹ (Leitner et al. 2010b) in Matlab® code called RootBox. An advantage of 98 RootBox (Leitner et al. 2010a) over other models is that it is implemented in Matlab in a way 99 that keeps it open for any changes to the model structure (Dunbabin et al. 2013). This model 100 has been used to predict nutrient uptake (Leitner et al. 2010b; Schnepf et al. 2012), growth of 101 arbuscular mycorrhizal fungi (Schnepf et al. 2016), root-hydrology interactions (Tron et al. 2015) 102 and water stress tolerance (Leitner et al. 2014). RootBox has been used to simulate many types 103 of root system, e.g. to simulate 48 root architectures in 16 drought scenarios (Tron et al. 2015) 104 and to model structural attributes for root functional type in 288 simulated root systems (Bodner 105 et al. 2013). Visual comparisons of the root simulation with excavated roots from field conditions 106 are at least qualitatively promising for species including maize, Anagallis femina, and Brassica 107 napus (Leitner et al. 2010b).

108 This paper presents a new simple model for root elongation in relation to soil physical 109 characteristics (penetration resistance, water stress, hypoxia or anoxia), and implements the 110 model in RootBox for contrasting soil management regimes. Specifically, it:

- describes the overall model structure and its component sub-models (root architecture; soil water uptake and redistribution including evapotranspiration; root elongation in relation to soil physical properties);
- proposes a new diagrammatic framework to visualise the interactions between root growth, soil management, and weather conditions;
- calibrates the soil strength-water content relation with laboratory data, and then tests it against field experimental data.

¹ Lindenmayer's system for plant architecture modelling (Prusinkiewicz and Lindenmayer 1990).

118 Simulations are then run relating to 119 - a hypothetical soil with s

120

121

122 123

124

125 126

127

128

129

130

131

132 133 134

135

149

150

151

160 161

- a hypothetical soil with single compacted layer;
- a test of the model using two years of field data, incorporating a typical rainfall season, and a year of drought.

Material and Methods The Model: Modelling approach

A model (Fig. 1) was constructed that consisted of (a) the root architecture model RootBox; (b) a soil-water redistribution model using Richards' equation and a water uptake function; (c) a soil-strength function that relates soil strength to soil water status; and (d) a rootstress function to define root elongation rate as limited by soil physical conditions. The components of the model are described in the following sections, together with the input parameters used in a series of simulations scenarios and comparison with field tests.

Root architecture model (RootBox)

136 Three-dimensional root system architecture was generated using the RootBox model 137 (Leitner et al. 2010a). Root elongation was described as a negative exponential growth function, (Eq. 1), such that in the absence of stress root elongation follows a negative exponential 138 139 function of time until a predefined maximum root length (k) is reached (Pagès et al. 1989; 140 Leitner et al. 2010b). The maximum root length (k) is calculated for each individual root (Eq. 2) 141 (Leitner et al. 2014) as the sum of the length of basal and apical zones, plus the spacing and 142 number of lateral branches (Fig. 2). Basic rules are applied for simulating root growth, 143 branching, and different types of tropisms, e.g. gravitropism, exotropism, hydrotropism or 144 chemotropism as described by Tron et al. (2015).

The main equation of interest in this work describes the root elongation. Unimpeded root
elongation is given by Eq. (1).

148
$$RL = k \left(1 - exp \left(-\frac{re}{k} t \right) \right)$$

where RL is root length (cm), t is time (day), k the maximum root length (cm) and re the initial root elongation (cm day⁻¹).

(Eq. 1)

(Eq. 2)

152 $k = l_b + l_a + l_n \cdot (n_b - 1)$ 153 where k is the maximum root length (cm). l_b is the

where k is the maximum root length (cm), I_b is the basal zone (cm), I_a is the apical zone (cm), I_n is the inter-spaces between branching (cm), n_b is the maximal number of lateral branches (unit).

155 New branches emerge only after the distance between root tip and branch has reached 156 the required apical zone length, and they emerge at a user-defined axial branching angle Θ 157 (normal distribution β and random radial angle (drawn from a uniform distribution between $-\pi$ 158 and π).

Soil water redistribution, incorporating root water uptake

Water flux in the soil matrix was modelled in one dimension by solving the Richards'
equation (Eq. 3):

166
$$C(h)\frac{\partial h}{\partial t} = \frac{\partial}{\partial z} \left(K(h) \left(\frac{\partial h}{\partial z} + 1 \right) \right) - S(h)$$
 (Eq. 3)
167

where C is the differential water capacity $\left(\frac{\partial\theta}{\partial h}\right)$ (cm⁻¹), θ is the volumetric soil water content (cm³ cm⁻³), h is the soil water pressure head (cm), t is the time (d), K is the unsaturated hydraulic conductivity (cm d⁻¹), z is the depth (cm, positive upward), S is the root water extraction (cm³ cm⁻³ d⁻¹).

Eq. (3) was solved numerically by a combination of finite difference and finite element methods (Celia and Bouloutas 1990; van Dam and Feddes 2000), as described in van Dam and 175 Feddes (2000). The relations between θ, h and K (Mualem 1976; van Genuchten 1980) are 176 shown in Eq. 4 and Eq. 5;.

178
$$\Theta = \frac{\theta - \theta_r}{\theta_s - \theta_r} = (1 + |\alpha h|^n)^{\left(\frac{1}{n}\right) - 1},$$
(Eq. 4)

182 183

184

185

186

187

188

180
$$K = K_s \Theta^{\lambda} \left(1 - \left(1 - \Theta^{\frac{n}{(n-1)}} \right)^{1 - (\frac{1}{n})} \right)^2$$
,
181

(1)

in which Θ is the effective saturation; θ is the water content (cm³ cm⁻³); θ_s is the saturated water content (cm³ cm⁻³); θ_r is the residual water content (cm³ cm⁻³) that was estimated by regression for positive values; h is pressure head (cm); K is the hydraulic conductivity unsaturated (cm d⁻¹); Ks is the saturated hydraulic conductivity (cm d⁻¹); and α (cm⁻¹), n and λ are empirical parameters. The value of parameter λ (Eq. 5) was 0.5 (Mualem 1976; van Genuchten 1980).

(Eq. 5)

189 The initial condition is imposed by specifying the pressure head (h) at depth (z) at time 190 zero (t = 0). The boundary conditions can be of two types, Dirichlet boundary condition, i.e., 191 specification of the pressure head h, or Neumann boundary condition, i.e., specification of a flux 192 q through the boundaries (Feddes et al. 1978).

193 At the lower boundary we assumed that the water table was deep enough to not influence 194 soil water dynamics, setting the gradient of the water pressure head $\partial h/\partial z$ is equal to zero (Tron 195 et al. 2015). Thus, the flux was solely driven by gravity and is equal to the unsaturated hydraulic 196 conductivity calculated at this boundary.

197 At the top soil surface boundary (i.e. soil surface), water flux is solved by the 198 Richards equation (van Dam and Feddes 2000) and depends on crop development (water 199 uptake), meteorological data and soil conditions. The soil can lose water by evaporation or gain 200 water by infiltration. The potential of crop transpiration and soil water evaporation are estimated 201 with the dual crop coefficient (Allen et al. 1998, 2005). For this model, we consider only root 202 growth and neglect shoot development. However, the link between soil-plant-atmosphere is the 203 actual plant transpiration (de Jong van Lier et al. 2008). In case of evaporation, the potential 204 water flux from the soil surface only depends on atmospheric conditions, but the actual flux 205 would be restricted by availability of water in upper soil layers (Tron et al. 2015). If these soil 206 layers dry, the boundary condition will switch from flux-controlled, with q equal to the potential 207 evaporation (EP, cm day¹), to head-controlled, with $h = h_{atm}$. The parameter h_{atm} is the water 208 pressure head at the soil surface in equilibrium with the pressure head of the atmosphere 209 (Feddes et al. 1978). 210 211

In both cases, as described in Tron et al. (2015) the condition in the Eq. (6) must be respected. The model described by equations (1)-(6) was implemented in Matlab.

(Eq. 6)

212 $|q| \leq \left| -K(h) \left(\frac{\partial h}{\partial z} + 1 \right) \right|$ 213 214

215

217

218

219

220

221

222

216 First, the reference evapotranspiration (ETo) was determined with FAO Penman-Monteith equation (Allen et al. 1998). After that, the potential evapotranspiration (ETp) was estimated from reference evapotranspiration (ETo), using the dual crop coefficient approach (Allen et al. 1998), with one coefficient for crop transpiration (Kcb) and another coefficient for soil water evaporation (Ke). Crop potential transpiration was estimated for each day as a function of plant development, and soil water evaporation is a function of wetness and soil surface covering (Allen et al. 1998). Both, crop transpiration and soil water evaporation coefficients are estimated 223 224 for each crop stage (i.e. initial, crop development, mid-season, and late season) (Rosa et al. 2012).

225 Soil water evaporation was estimated as defined in (Allen et al. 1998, 2005) using the 226 227 parameters which allowed the computation of total and readily available soil water (TAW and RAW, mm), as well as the initial values for the total evaporable water (TEW, mm), readily 228 evaporable water (REW, mm), and thickness of the evaporation soil layer (Ze, m). Soil water 229 evaporation coefficient (Ke) represents the contribution of evaporation from soil to total 230 evapotranspiration (Pereira et al. 2015). Calculation of soil water evaporation coefficient (Ke)

231 uses a variation of the second stage of Ritchie's soil evaporation approach (Ritchie 1972) 232 described in details by Allen et al. (1998, 2005).

233 234 Root water uptake was simulated using the model proposed by de Jong van Lier et al. 235 (2008) (Eq. 7), that used the root length density and matric flux potential of each soil layer (Eq. 236 237 238 8). This model was coupled with the Richards' equation in one-dimensional (1D) by water uptake model (Eq. 7). The matric flux potential (M, cm² d⁻¹) is defined as the integral of unsaturated hydraulic conductivity, (K(h), cm d^{-1}), over pressure head (h); or equivalently as the 239 integral of diffusivity, ($D(\theta)$, cm² d⁻¹), over water content (θ , cm³ cm⁻³) (de Jong van Lier et al. 240 2008). The low bound of the integral is the permanent wilting point in terms of pressure head 241 (h_w, cm) or water content (θ_w , cm³ cm⁻³). 242

243
$$S_z = \rho_z (M(h)_z - M_{0,z})$$
 (Eq. 7)
244

245 where p_z (cm⁻²) is defined as weighting factor for matric-flux potential dependent of root uptake; 246 M(h) is the matric flux potential (cm^2 day⁻¹); M₀ is the matric flux potential at the root surface 247 (cm² day⁻¹). The matric flux popential is given by 248

249
$$M(h) = \int_{h_w}^{h} K(h) \partial h = \int_{\theta_w}^{\theta} D(\theta) \partial \theta,$$
 (Eq. 8)
250

251 where h is the pressure head (cm); h_w is the pressure head at the wilting point (cm); K is the 252 unsaturated hydraulic conductivity (cm day⁻¹); D is the diffusivity (cm² day⁻¹); θ is the soil water 253 content (cm³ cm⁻³); θ_w is the water content at the wilting point. 254

The procedure for calculating root water uptake (Eq. 7) was described by de Jong van Lier et al. (2008) and estimated the water uptake for each layer dependent on the matric flux potential (Eq. 8) with a reduction function for root length density, given by

$$\rho_z = \frac{4}{r_{o,z}^2 - a_z^2 r_{m,z}^2 + 2(r_{m,z}^2 + r_{o,z}^2) ln (a_z r_{m,z}/r_{o,z})},$$
(Eq. 9)

262 in which p_z (cm⁻²) is a weighting factor for matric-flux potential dependent of root uptake; a_z is a 263 constant equal to 0.53; z is the soil layer (cm); $r_{0,z}$ is the root radius (cm); and $r_{m,z}$ is the half-264 mean distance between roots (cm) which can be computed from the root length density 265 according to Eq. 10 (de Jong van Lier et al. 2008). 266

267
$$RLD = \frac{1}{\pi \cdot r_m^2}$$
 or $r_{m,z} = \sqrt{\frac{1}{\pi \cdot RLD}}$, (Eq. 10)
268

269 where RLD is the root length density (cm cm⁻³); r_m is the half-mean distance between roots 270 (cm); z is the each soil layer (cm). The half-mean distance between roots is a measure for the 271 soil volume each root can exploit to water uptake. 272

The root radius $(r_{0,z})$ and root length density are simulated from input parameters with the equations from the RootBox model (Leitner et al. 2010b).

275 Actual transpiration (T_a) cannot be higher than the potential transpiration (T_P , cm day⁻¹) of 276 the plant. Actual transpiration is given by the integral of water uptake (Eq. 7), resulting in Eq. (11). The matric flux potential M_0 is initially considered equal to zero, i.e., $h = h_w$ at the root 278 surface, but if the obtained transpiration is larger than potential transpiration, no water stress 279 occurs and thus M_0 is larger than zero and its value is obtained by setting $T = T_P$ (Tron et al. 280 2015). 281

$$T_a = \int_0^{z_{max}} S_{(z)} dz \tag{Eq. 11}$$

284 where Ta is actual transpiration (cm day-1); zmax is the maximum root depth (cm); S(z) is the water 285 uptake in each soil layer (cm day-1).

286 287

282 283

273

274

277

255

256

257

288 Calibration of soil strength function 289

Soil penetration resistance varies greatly with soil water status, and was modelled as a function of soil water content and bulk density using a non-linear model (Eq. 12) (Busscher 1990). In equation 12 the constants (*a*, *b*, and *c*) are found by empirically fitting to the experimental values of soil penetration resistance, water content and bulk density (Moraes et al. 2017). In this paper a soil penetration resistance function was used for a data set measured in the laboratory for a Rhodic Eutrudox (Ortigara et al. 2015). The relation between predicted and measured penetration resistances simulated are distributed closely around the one-to-one line in Fig. 3, with a correlation coefficient of 0.91. Thus, Busscher's model (Eq. 12) can be used to describe soil penetration resistance for this soil, given by

 $Q_p = a \gamma^b \theta^c$

290

291

292

293

294

295

296

297

298

299 300

301 302

303

304 305 306

307

(Eq. 12)

where Q_p (MPa) is the soil penetration resistance; γ (Mg m⁻³) is the dry bulk density; θ (cm³ cm⁻³) is volumetric soil water content and *a*, *b* and *c* are empirical parameters.

Root stress function: Root elongation as a function of soil physical stresses

Root elongation is a function of soil strength and matric potential (Fig. 4a,b), and for simplicity we assume that these stresses combine linearly i.e. the stresses act independently to decrease elongation rate. This results in a relation where root elongation rate is defined for all combinations of penetration resistance and matric potential, and so can be used to produce a heat-map where red indicates slow root elongation due to physical stress, and blue indicates unimpeded root growth (Fig. 4c,d).

314 The heat-map therefore represents the response of a particular plant genotype to soil 315 physical stresses. By plotting the penetration resistance as a function of soil matric potential on 316 the same diagram, the effect of soil physical stress on root elongation is represented for the 317 range of water contents that the plant may experience during the growth season (eg. see 318 plotted points on Fig. 4c, representing two soil compaction levels). If the soil is moist at the start 319 of the growth season, root elongation will be in the blue zone and relatively fast. As the soil 320 dries (e.g. during a period with little rain), the soil strength will increase and the elongation rate 321 will slow - much faster in the case of the upper strength characteristic curve (hollow circles in 322 Fig. 4c), than for the more benign soil (shaded squares in Fig. 4c). Thus, this heat-map diagram 323 can be used to capture many elements of the complex interactions that occur between 324 contrasting soil types, management regimes, climate and plant genotype. By interpreting this 325 diagram with weather data for a particular year, it can be used to compare stresses that limit 326 root elongation in different seasons.

One aspect that is difficult to represent is the increased root elongation due to continuous macropores. Whilst it is not possible to represent this phenomenon mechanistically without detailed root-tip scale modelling of the soil structure, we have sought to heuristically adjust the root growth function to permit faster elongation at the same penetrometer resistance in soil containing macropore channels (Fig. 4d). Our justification for this is that root growth has been observed in these Brazilian field soils with very large penetration resistances in a depth, where root growth would not normally be expected.

In the next two sections we describe the root model, the relation to the soil physical
stresses, i.e. water stress and hypoxia, and penetration resistance.

Root elongation under water stress and poor aeration

Using the Feddes concept for water uptake (Feddes et al. 1978), and adapting for root elongation yields

 $RE(h) = \alpha(h)RE_{max}$, (Eq. 13)

343 344 345

337 338

339 340

341

where RE(h) is the root elongation potential affected by matric potential (cm day⁻¹), α_h is a dimensionless prescribes function of soil water pressure head and, RE_{max} (cm day⁻¹) is the maximal possible root elongation without restrictions.

349 Under non-optimal conditions, i.e. either too dry (water deficit) or too wet (poor 350 aeration), the root elongation is reduced by means of the stress reduction factor ($\alpha(h)$) from 1 351 (maximum root elongation) to zero (no growth). The shape of this function for root elongation 352 follow the concept proposed to by Feddes et al. (1978) (Eq. 14). We used four limits of matric 353 potential (h), i.e., that there is a linear increment of root growth from h_1 (-0.1 kPa) to h_2 (-6 kPa), 354 and a linear reduction of the root elongation from h_3 (-10 kPa) to h_4 (-1000 kPa). The h_1 was 355 defined at the wet end and represents the start of water drainage and increase of soil aeration 356 (and oxygen concentration) necessary for root growth (Dresbøll et al. 2013). The h₂ and h₃ are 357 the values close to field capacity (lijima and Kato 2007), when there is no water stress and thus 358 root elongation rate is at its maximum. The value h₃ was defined as the limit of maximum growth 359 due to turgor pressure in the expanding cells of the root elongation zone, and is typically up to 1 360 MPa (Bengough et al. 2011). The equation is given by

- 361 362
- 363

$$\begin{array}{c|c} 368 \\ 369 \\ 370 \\ 371 \end{array} \quad \text{ where } \alpha(h) \text{ is the stress reduction factor of root elongation data} \end{array} \quad \text{if } |h_3| < |h| \le |h_4| \\ \text{if } |h| > |h_4| \\ \text{o the stress reduction factor of root elongation data} \end{array}$$

where $\alpha(h)$ is the stress reduction factor of root elongation due pressure head; |h| is the module of pressure head, and h_1 , h_2 , h_3 and h_4 are the limits of pressures head for root elongation (Fig. 4a). Root elongation below $|h_1|$ (critical respiratory oxygen pressure, with $|h_1|$ approaching to saturation (1 cm) (Saglio et al. 1984)) and above $|h_4|$ (maximum growth pressure, with $|h_4|$ approaching 1 MPa (Bengough et al. 2011)) is set equal to zero. Between $|h_2|$ and $|h_3|$ (reduction point, $|h_2|$ is 6 kPa, and $|h_3|$ is 10 kPa) root elongation is maximal. Between $|h_1|$ and $|h_2|$ and between $|h_3|$ and $|h_4|$ a linear variation is assumed.

Root elongation in relation to soil strength

381 The effects of water stress, poor aeration, and soil strength on root elongation (Eq. 15), 382 can be predicted from the stress reduction function Eq. (16) and is shown in Fig. 4c and Fig. 4d. 383 The root elongation rate can slow due to soil strength, with an exponential decrease for a soil 384 without continuous macropores (Eq. 17) (Bengough 1997). Thus, root elongation can be 385 represented as a function of soil strength (Q_p) and matric potential (h), at time (t) and depth (z) 386 (Eq. 15). The stress reduction function in a layer z, on day t shows the effect of field conditions 387 on root elongation (Eq. 16). Biopore effects on facilitating root elongation are included very 388 simply in the root model by changing the relationship between root elongation rate and soil 389 strength (Eq. 18), as indicated by the blue dashed line at the Fig. 4b (Bengough 2012; Jin et al. 390 2013). This favours faster root elongation, and changes the root response to soil strength. The 391 predicted rate of root elongation is therefore relatively faster in soil containing many biopores, 392 as compared with one containing few biopores, at the same penetration resistance (Fig. 4d 393 compared with Fig. 4c).

394 395

379

380

399 where $\operatorname{srf}(Q_p, h)_{t,z}$ is the total stress reduction function for root elongation due to mechanical 400 (Q_p) and hydric (h) stresses in each time (t) and depth (z); $\alpha(Q_p)$ is the stress reduction function 401 by soil strength and is given by Eq. (17) in a soil without continuous macropores or by Eq. (16) 402 for a soil with continuous macropores; $\alpha(h)$ is the stress reduction function by matric potential 403 (water and aeration stress) and, t is the time (day), z is the depth (cm); RE_{max} is the root elongation maximal possible without restrictions (cm day⁻¹), and RE is the root elongation (cm day⁻¹).
406

407	$\alpha(Q_p) = \exp(-0.4325 \mathrm{Q}_p),$	(Eq. 17)
408	$\alpha(Q_n) = \exp(-0.30 \mathrm{Q}_n),$	(Eq. 18)
409		

410 Note that we make the assumption that the combined effect (Eq. 16) of the two stresses (mechanical and hydric) is multiplicative for each time and depth. In addition, penetrometer resistance that depends on water content and bulk density is used as the measure of soil strength.

415 416 Coupling of the model

417 418 The link between root and soil water models is the water uptake (1D sink term) in 419 Richards' equation (Tron et al. 2015), and the link for soil physical conditions and root growth is 420 the stress reduction function for root elongation (Fig. 6). The sink term represents root water 421 uptake from each horizontal soil layer (1 cm thick), and soil physical conditions control the 422 stress reduction function that restricts root elongation and root system growth. The water uptake 423 term depends on two factors: root length density and availability of water in each soil layer (de 424 Jong van Lier et al. 2008). Thus, the model is dynamic: e.g. where there are more roots for 425 water uptake, the soil may become dry and hard, with greater restriction to root elongation. 426 However, the effects on root elongation act on each single root, which can ultimately influence 427 the root system architecture. 428

Implementation and parameterization

The root growth model was implemented by extending the L-system model for root growth, RootBox (Leitner et al. 2010a), and is written in Matlab®. The water flow in soil was implemented into Rootbox as described in Tron et al. (2015). Water flux, soil physical conditions, and the stress reduction function for root elongation and root growth are alternately computed at each time step, which was set to be 1 day. Each segment root was submitted to dynamic soil physical conditions in a specific day and position from a soil layer with 1 cm depth layer resolution.

Model parameters are: (1) parameters describing the soil (soil water retention curve, soil penetration resistance curve and bulk density) (e.g., Table 1), (2) parameters regarding climate (potential evaporation and transpiration, temperature, air humidity, rainfall and irrigation), and (3) root architecture parameters (i.e., initial root elongation, length of the apical and basal zone, spacing between branches, number of branches and insertion angle), type of tropism, growing period and limits for root elongation) (e.g., Table 2).

Model output parameters are related to the soil conditions (water balance, infiltration, runoff and deep drainage, actual evaporation rate, water content, matric potential, soil penetration resistance and unsaturated hydraulic conductivity) and crop root system structure, root length density, actual transpiration and water uptake at each layer.

450 451

429 430 431

432

452 Simulation scenarios

453 Calibration of soil strength function against field data of soil water content and soil 454 penetrometer resistance

455

The model was tested using soil physical parameters (Table 1), soil water content and soil penetration resistance of a Rhodic Eutrudox in a no-tillage system (Moraes et al. 2012, 2013). In this experiment, there was no plant growth, but the accuracy of the Richards' equation was evaluated. Data of water content and soil penetration resistance of two soil layers (0-10 cm and 10-20 cm depth) from a field experiment were obtained from 45 days in a wet-dry cycle on field conditions carried out at the Embrapa Soybean experimental station at Londrina, Brazil.

462 463 Simulation case study: effects of a compacted layer on root growth

464

We simulated the root growth of soybean for 87 days, from 10th October 2013 (sowing) to 5th January 2014 (sample roots). Two soil conditions were simulated in order to analyse the reduction of root elongation rate due to soil compaction, no-tillage without or with a compact soil layer from 16 to 20 cm depth. The bulk density in the soil profile (0-100 cm depth) for loose soil was 1.0 Mg m⁻³, using the same profile but with a compacted soil layer, from 16 to 20 cm of bulk density of 1.30 Mg m⁻³.

For the simulation we used the daily time series of climate data from Embrapa Soybean experimental station, in Londrina (latitude 23°11'S; longitude 51°11'W; and 620 m altitude), State of Paraná, southern Brazil (Fig. 5). The simulation was made using the soil physical properties (Table 1) of an Oxisol (Latossolo Vermelho Distroférrico, in Brazilian classification; and Rhodic Eutrudox, in USA classification) on no-tillage system (established in 1997) with 755 g kg⁻¹ of clay, 178 g kg⁻¹ of silt and 67 g kg⁻¹ of sand.

- 477
- 478 479

Field experiment comparison: typical versus drought seasons

480

481 For the simulation we used: the daily time series of climate data from two growing 482 seasons at Embrapa Soja, in Londrina (latitude 23º11'S; longitude 51º11'W; and 620 m 483 altitude), State of Paraná, southern Brazil (Fig. 11); the soil physical properties of an Oxisol 484 (Latossolo Vermelho Distroférrico, Brazilian classification: Rhodic Eutrudox, USA classification) 485 on a no-tillage system (established in 1997) with 755 g kg⁻¹ of clay, 178 g kg⁻¹ of silt and 67 g kg⁻¹ 486 ¹ of sand (Table 1). Two season growth conditions were used to compare the effects of weather, 487 a drought (2008/09) and a wet (2009/10) season on root development (Franchini et al. 2017). 488 The parameters used to calculated the plant transpiration, soil evaporation and crop growth 489 stages are summarized in Table 3.

490 Soybean root system development (Cultivar BRS-282) was simulated for 70 days, for 491 both wet and drought seasons. For the drought season, the soybean was seeded on 24th 492 November 2008, with roots sampled from the field on 2nd February 2009 (Franchini et al. 2017). 493 The main time with water stress in drought season was during the first 54 days after sowing, 494 that was used to identify the drought stress on root growth. In the wet season, soybean root 495 growth season was simulated from 12th November 2009 to 20th January 2010 (Franchini et al. 496 2017) for one individual plant. After that, the root length densities were converted to a per-area 497 basis and in 1D assuming the area of each plant with a population of 30 plants m⁻², with 0.45 cm 498 inter-rows, i.e., 7 cm inter-plants in the row. Soil water flux was simulated to 100 cm depth. Root 499 elongation was modelled daily incorporating the effects of soil and climate conditions. Root 500 length density and root system architecture were simulated for 70 days. Results were compared 501 with both root length density and an excavated profile wall in the field. 502

503 504 s

505

Statistical evaluation of model performance

The agreement between simulated and measured values was expressed by the mean absolute error (MAE) (Eq. 19) (Casaroli et al. 2010), the root mean squared error (RMSE) (Eq. 20) (de Jong van Lier et al. 2008), the coefficient of residual mass (CRM) (Eq. 21), the coefficient of correlation (r) (Eq. 22) (Bonfante et al. 2010), and the index of agreement (d) (Eq. 23) (Casaroli et al. 2010). Also, the modelling efficiency (EF) (Eq. 24) (Bonfante et al. 2010), and the one-to-one line were used as criteria to evaluate the model performance.

512

514

513 $MAE = \frac{1}{n} \sum_{i=1}^{n} |P_i - O_i|$

(Eq. 19)

where *n* is the total number of measurements, O_i and P_i are the measured and predicted values of the observation, respectively. The root mean square error (RMSE) has minimum and optimum value at 0. It is a difference-based measure of the model performance in a quadratic form, and it is sensitive to outliers.

520
$$RMSE = \sqrt{\frac{1}{n}\sum_{i=1}^{n}(P_i - O_i)^2}$$
 (Eq. 20)
521

522 The coefficient of residual mass (CRM), ranges between $-\infty$ and $+\infty$, with the optimum 523 equal to zero. Positive values indicates that the model underestimates the prediction, and 524 negative values indicates overestimation. When CRM are close to zero it indicates the absence 525 of trends.

526
527
$$CRM = \frac{\sum_{i=1}^{n} o_i - \sum_{i=1}^{n} P_i}{\sum_{i=1}^{n} o_i}$$
 (Eq. 21)
528

The optimum value of the coefficient of correlation (r) (Addiscott and Whitmore 1987) is equal to 1; zero means no correlation.

531
532
$$r = \frac{\sigma OP}{\sigma 0.\sigma P}$$
 (Eq. 22)
533

534 where σOP is the covariance between measured and estimated data and σO and σP are the 535 measured and estimated standard deviation, respectively. 536

The index of agreement of Willmott (d) is dimensionless, lies between -1.0 and 1.0, and is more related to model accuracy than other indices (Willmott et al. 2012).

540
$$d = 1 - \frac{\sum_{i=1}^{n} (P_i - O_i)^2}{\sum_{i=1}^{n} (|P_i - \bar{O}| + |O_i - \bar{O}|)^2}$$
 (Eq. 23)
541

Modelling efficiency (EF) (Greenwood et al. 1985) can get either positive or negative values, 1 being the upper limit, while negative infinity is the theoretical lower boundary. EF values lower than 0 result from a worse fit than the average of measurements.

$$EF = \frac{\sum_{i=1}^{n} (o_i - \bar{O})^2 - \sum_{i=1}^{n} (P_i - O_i)^2}{\sum_{i=1}^{n} (o_i - \bar{O})^2}$$
(Eq. 24)

Results

551 552

529

530

537

538

539

542

543

544

545 546

553 Soil water content and soil penetration resistance in a soil without roots

554

555 Simulated soil water content was plotted against measured soil water content at 8 cm and 556 16 cm depths in the Rhodic Eutrudox no-tillage treatment, in the absence of a crop during a 557 wet-dry cycle (Fig. 6a). Simulated penetrometer resistances were similarly compared with field 558 measurements at the same two depths (Fig. 6b). The simulations using weather and soil data 559 resulted in good prediction of soil water content and penetration resistance, with points lying 560 close to the 1:1 line (Fig. 6a and Fig. 6b). The linear regression lines for simulated and 561 measured data of soil water content or penetration resistance were not significantly different 562 from the 1:1 lines at 95% confidence level. The index of agreement (d) was 0.84 for water 563 content, with correlation coefficient (r) of 0.86 (Fig. 6a). Simulated and measured values of soil 564 penetration resistance were in very good agreement (index of agreement 0.92, correlation 565 coefficient 0.85; Fig. 6b). This suggests that the relevant processes were captured in the model. 566 Prediction of soil water content and penetrometer resistance was appropriate for Rhodic 567 Eutrudox soil.

568 The proposed model offers a useful framework to investigate the effects of soil physical 569 conditions on root growth and the stress reduction function can be used as input for other soil-570 plant models.

573 Simulation case study: effects of a compacted layer on root growth

574

The same climate data were used for the 87 day root growth simulation (see Supplementary videos S1 and S2). Total precipitation was 352 mm, during 31 days of rain. Simulations of the soybean root system are shown with (Fig. 7a and supplementary video S1) and without the presence of a compact layer (Fig. 7b and supplementary video S2). The stress reduction factor is plotted as a function of time and depth for these two soil conditions, adjacent to the root simulations (Fig. 7e and Fig. 7f): darker blue indicates more rapid root elongation, whilst red indicates a large decrease in root elongation due to soil physical stress.

The root distribution was much more uniformly tapering for the profile without a compact zone (Fig. 7a). Many main root axes penetrated below 16 cm depth, and lateral roots proliferated freely around these axes. The tap root spends 9 days more to cross compacted soil layer (16-20 cm depth) compared to the uncompacted soil (Fig. 7c,d). The stress reduction function indicated relatively little decrease in root elongation rates until day 60, when elongation slowed in the surface layers, the root system extended down to 70cm by day 70.

588 In the presence of a compact layer, the pattern of root growth was changed, with a 589 corresponding alteration of root distribution down the soil profile. Fewer main root axes crossed 590 the compact soil layer from 16 to 20 cm depth, and there was a noticeable gap in lateral root 591 proliferation in the compact zone. Root length density in the compacted soil layer (16-20 cm) 592 was reduced 83 %, from 0.90 cm cm⁻³ (compacted soil) to 0.15 cm cm⁻³ (uncompacted soil). 593 However, root length density in the layers (0-15 cm depth) above the compacted layer was 594 increased 25 % to 1.25 cm cm⁻³ (compacted soil) from 0.99 cm cm⁻³ (uncompacted soil) (Fig. 595 7c,d). The stress reduction function indicated that root elongation rates in the compact layer 596 were typically slowed to below 20% of the maximum root elongation rate - a very substantial 597 impediment to root elongation – although the main tap root still penetrated below 75 cm depth 598 by day 70.

599 The effect of a compact layer in the soil profile changed root architecture and root length 600 density distribution; however, rooting depth was similar in both soil conditions, to a maximum of 601 75 cm depth. The thin 5 cm compact layer substantially altered the water uptake pattern (Fig. 602 8b), mainly due to the restriction on root system development (Fig. 7b). Water uptake was 603 localised where the root length density was greatest adjacent to plant-available water, as 604 modelled by matric flux density (de Jong van Lier et al. 2008). In the compacted-layer scenario, 605 the root system (and water uptake) was restricted to shallower than 20 cm depth for 20 days 606 (Fig. 8b). However, by 20 days in the scenario with loose soil (free of compact layer), the root 607 system was already extracting water to 40 cm (Fig. 8a). The water uptake was generally higher 608 in the uncompacted scenario in the top 15 cm of soil until 65 days of plant growth, due to the 609 more superficial proliferation of the root system as compared with the uncompacted profile.

610 To understand which factors influenced the root elongation most, it is necessary to further 611 analyse the stress reduction factor parameter. Soil penetration resistance and water content 612 both varied substantially during the growth season (Fig. 9). Soil penetration resistance was 613 changed over time (Fig. 9a,b) due to water flux in the soil (Fig. 9c,d). Within the compacted soil 614 layer this lead to a soil penetration resistance that was higher than 4 MPa, acting as a limitation 615 to root elongation. Soil water content was changed during the growth season due to crop water 616 uptake or soil water movement (due to water evaporation, deep drainage, etc.). As expected, 617 the rooting depth developed faster in loose soil than in the soil including the compacted layer. 618 The root system under soil compaction was delayed, limiting the water available to root water 619 uptake from deeper layers and therefore plant transpiration. Faster root growth in loose soil 620 favours root system water uptake, and quickly depletes soil water over the soil profile. This 621 increases the penetration resistance which limits root elongation.

622 Fig 10. shows the effects on root elongation modelled by the stress reduction function for 623 the modal values of 0.73 in the loose soil and 0.14 in the soil with a compacted soil layer(Fig. 624 10a). The separation of the total stress reduction functions into two effects, one of penetration 625 resistance (Fig. 10b) and the other of matric potential (Fig. 10c), favour to analyse the relative 626 distribution of the stress reduction factor values that reduce the root growth of soybean. The 627 reduction factor associated with high soil penetration resistance was closer to zero (high 628 restriction) for the soil with a compact layer with modal value of 0.15 (Fig. 10b) in contrast to a 629 modal value of 0.76 in the uncompacted soil (Fig. 10b). Restrictions of root elongation due to 630 matric potential were similar for both soil conditions (Fig. 10c), with modal values of 0.93 (loose 631 soil) and 0.94 (soil with a compacted layer). This indicates that mechanical impedance (Fig. 632 10b) exerted a greater limitation to root elongation in soil with a compacted layer (Fig. 10a).

Field experiment comparison: typical versus drought seasons

In this section, comparisons between simulations and field experiments are reported
showing simulated root system architecture, root length density distribution, stress reduction
factor, and water uptake distribution. The simulations are then compared with measurements of
root length density distribution and excavated profile walls of soybean plants grown in the field
during the two seasons.

During the drought season 2008/09, it rained 306 mm during 70 days of root growth, with 32 of these days having rainfall (Fig. 11a). During the wetter season of 2009/10, there were 46 days with rainfall, giving a total of 515 mm rainfall (Fig. 11b). During the first 54 days in 2008/09, there was only 131 mm (2.4 mm day⁻¹) of rainfall compared to 354 mm (6.5 mm day⁻¹) of rainfall in the same period in 2009/10 (Fig. 11). The rainfall distribution was irregular in the drought season affecting root development, and was associated with large penetration resistance values in the field (Moraes et al. 2013).

649 The simulated stress reduction function limited root elongation in the dry season, with 650 corresponding changes in simulated root architecture and root length density distribution (Fig. 651 12 and see Supplementary videos S3 and S4). The simulated root system grew slower, 652 especially early in the dry season (see supplementary video S3). Reduced rainfall quantity in 653 2008/09 increased the hydric and mechanical stress to root growth. Rooting depth in 2008/09 654 was 52 cm after 54 days and 48% smaller than rooting depth in 2009/10 in the same period (77 655 cm depth). During this period the root length density was reduced for 46% (from 0.41 cm cm⁻³ to 656 0.28 cm cm⁻³, for 2009/10 and 2008/09 respectively) in the first 30 cm depth (Fig. 12). Both the 657 depth of and the volume of soil explored by the simulated root system during the drought 658 season (2008/09) was decreased in relation to the wet season (2009/10).

659 The distribution of the total stress reduction function differed between wet and dry 660 seasons (Fig. 13). The relative frequency of total stress indicates that 76 % of the values were 661 smaller than 0.4 (yielding a reduction of 60% in root elongation rate) in dry season. However, in 662 wet season it was only 36 % of total stress values. Stress from mechanical impedance 663 contributed to a reduction higher than 60 % of root elongation rate in 70 % of the cases. In 664 contrast, in the wet season root elongation was reduced only in 29 % of the cases. Stress from 665 matric potential to root elongation smaller than 0.90 was increased from 9 % to 42 % of values 666 in the wet and dry season, respectively (Fig. 13). The frequency of stress reduction over rooting 667 depth during the first 54 days (major rainfall deficit in 2008 season growth) showed that the 668 main stress limiting root elongation was due to soil penetration resistance in the wet season. In 669 the drought season, root elongation was also reduced due to hydric stress (42 % cases with 670 values smaller than 0.9), which was less in the wet season (9 % of the cases). The modelling 671 results suggest that in this soil during a dry season, mechanical impedance became a major 672 limitation to root elongation. In the drought season, roots experienced more days of soil physical 673 stress (Fig. 13a), with a combination of penetration resistance (stress reduction factor <0.4) 674 (Fig, 13b) and matric potential (stress reduction factor<0.9) limiting root elongation (Fig, 13c).

675 Water uptake rate and distribution differed substantially between wet and dry seasons 676 (Fig. 14). During the first 54 days in drought season (2008/09) soybean took up 51 mm (i.e. 677 denoted mm= litre m⁻²) of water depth, which is only 50% of the water uptake in the wet season 678 (2009/10), in the same period (see Fig. 14). In the dry season, the root system extended to 679 depth more slowly, resulting in a slower spread of the water extraction volume in depth. Plants 680 were submitted to greater stress under drought with an associated decrease in transpiration. 681 Simulated and measured root length density was very similar for all layers at the soil profile for 682 both weather conditions (Fig. 15 and Fig. 16). There was qualitative agreement between the model predictions and field observations of root length density for all soil profile (Fig. 16c). 683 684 Indicators describing the model quality were promising: modelling efficiency (EF) 0.87, index of 685 agreement (d) 0.97, RMSE 0.10, coefficient of residual mass 0.0061, coefficient of correlation 686 0.83, and means absolute error 0.08. The values indicate a good agreement between measured 687 and simulated values (Fig. 16c). 688

Discussion

689 690

691

633 634 635

692 Few studies have published comparisons between root elongation and soil physical 693 conditions (Bengough et al. 2011). Even in the most commonly studied crops, such as soybean 694 (Gregory 2006), maize (Schmidt et al. 2013), pea (lijima and Kato 2007) or cotton (Taylor and 695 Ratliff 1969), there is still a lack in investigating the combined effect of soil physical stresses of 696 hypoxia, water deficit and mechanical impedance (Bengough et al. 2011). In this paper, we 697 assumed that stresses from soil physical conditions reduce the root elongation as a combined 698 effect as described in Bengough et al. (2011). Root elongation parameters (Fig. 4) should be 699 similar for different soils or crops (lijima and Kato 2007), because they are a generic response 700 of root elongation to matric potential (Fig. 4a) and mechanical impedance (Fig. 4b). We used 701 the relationship of root elongation and penetrometer impedance from Bengough (1997). We 702 expect the same relationship between the stress reduction function (that is dimensionless 703 quantity, from zero to one) and the soil physical conditions (e.g. lijima and Kato (2007)), 704 furthermore the differences between crops should be only in the range of the elongation rate. 705 We presented the root system parameters used in our modelling case study (Table 2), including 706 the initial tip elongation rate (5.5 cm day¹) for unimpeded conditions. We calibrated the model 707 with field data from soybean root growth in no-tillage system at two weather conditions (Fig. 16). 708 We compared root length density modelled and measured in the field (Fig. 15 and 16) to include 709 the effect of continues pores and biopores (Fig. 4d) in no-tillage system (Moraes et al. 2016) to 710 reduce the mechanical impedance to root elongation (Bengough 2012; Jin et al. 2013) as 711 showed that changes in the relationship of root resistance and penetrometer resistance (Fig. 712 4b) due presence of biopores or crack in the soil profile (Bengough and Mullins 1991).

713 Root water uptake depends on soil water status, soil hydraulic properties, root length 714 density, and root radius (de Jong van Lier et al. 2013). In this work, we propose a new 715 modelling approach for root architecture development as affected by soil physical stresses and 716 its effect of root water uptake. We demonstrate that it well represents the root growth of 717 soybean growing in compacted soils. Soil strength and water availability can, independently, 718 reduce crop growth but there is no consensus on which of these stresses or combination of 719 stresses is the most important (Jin et al. 2013). By separating the total stress reduction function 720 for root elongation into two effects, one being the penetration resistance and the other one the 721 matric potential, we could analyse the relative contribution of the different stresses to the overall 722 reduction factor values. The effect of matric potential on root elongation restriction was similar in 723 two different soil conditions, with and without a compact layer. However, the relative frequency 724 of mechanical stress on root elongation was found from 0.70 to 0.90 in the loose soil, while the 725 stress values in the soil with a compact layer ranged from 0.00 to 0.30, indicating that 726 mechanical impedance exerted a greater limitation to root elongation in this case.

727 Rooting depth in both soil with or without a compacted layer were similar. However, root 728 system development was slower in the soil including the compact layer, leading to a reduced 729 root length density within this layer. Drought stress adversely affects plant growth by decreasing 730 the uptake of water and nutrients by plants (Miransari 2016a). Root length density was 731 decreased into the soil compacted layer due mechanical impedance (Bengough et al. 2011), 732 water stress (Benjamin and Nielsen 2006) and poor aeration (Valentine et al. 2012). The 733 strongest influence on root elongation in compacted soil is due to soil strength. Furthermore, the 734 combined effect of mechanical impedance and oxygen deficiency (hypoxia) impedes root 735 development (Valentine et al. 2012). Under compaction the root growth is adversely affected, as 736 the soil structure will not be suitable for root growth (Miransari 2016b).

737 Our new model considers the mechanical and hydric stresses for each root to elongation 738 over time (e.g. Fig. 12e and 12f) including root water uptake and soil water flux daily at soil 739 profile (e.g. Fig. 14). This was the first time that the soil physical conditions (mechanical and 740 hydric stresses to root elongation) were included into a root growth model; before that the 741 RootBox model (Fig. 12e and 12f) only predicted root growth due to time (root age) or due to 742 different types of tropisms such as chemotropism. In relation to other models, for example, 743 Hydrus (Hartmann and Šimunek 2016; Hartmann et al. 2017) or SWAP (Kroes et al. 2008) 744 models have a root growth package, however, those models do not consider the soil physical 745 conditions for each single root and do not consider the 3D root architecture.

In addition, there are few three-dimensional root architectural models actually in use, in
summary the most common and current models include RootTyp (Pagès et al. 2004), SimRoot
(Lynch et al. 1997), ROOTMAP (Diggle 1988b, a), SPACSYS (Wu et al. 2007), R-SWMS
(Javaux et al. 2008), Archisimple (Pagès et al. 2014), OpenSimroot (Postma et al. 2017),
RootBox (Leitner et al. 2010a, b), and CrootBox (Schnepf et al. 2017) which have been used for
a range of root modelling studies (Dunbabin et al. 2013). An advantage of RootBox (Leitner et

752 al. 2010a) over other models is that it is implemented in Matlab in a way that keeps it open for 753 any changes to the model structure (Dunbabin et al. 2013). Soil mechanic impedance effects on 754 root growth is used only in SPACSYS (Wu et al. 2007), R-SWMS (Clausnitzer and Hopmans 755 1994; Javaux et al. 2008), and HYDRUS (Hartmann et al. 2017); however, the strength of stress 756 is determined from empirical relationships between mechanical stress and bulk density, texture 757 (only sand content) and water content (Jones et al. 1991). Thus, for each soil we should fit the 758 relationship of soil penetration resistance, water content and bulk density (Busscher 1990) to 759 know the mechanical impedance over time. Those equations from Jones et al. (1991) do not 760 represent the correct relationship of soil penetration resistance with water content and bulk 761 density (Busscher 1990), especially when affected by soil structure (Moraes et al. 2017) in 762 clayey soils (Moraes et al. 2012), as described in this paper. In addition, the root growth module 763 of the HYDRUS model (Hartmann et al. 2017) uses environmental stresses such as 764 temperature, aeration, and chemical soil condition (Al toxicity and Ca deficiency) to reduce the 765 root system growth. Environmental stresses are calculated as a function of sand content and 766 soil moisture (Jones et al. 1991), and affect rooting depth and root proliferation within different 767 soil layers (Hartmann et al. 2017). Thus, that effect is very generic and does not consider each 768 individual root within the root system as detailed as in the RootBox model (Leitner et al. 2010a).

769 The new root architecture model can simulate the stress (mechanical and hydric) of each 770 individual root (tap root, lateral and secondary roots) in each position in the soil profile. We 771 include the root water uptake model due matric flux potential as a function of the distance to the 772 root (de Jong van Lier et al. 2008); the model includes compensation mechanisms such that 773 reductions in the uptake from dry layers are compensated by an increase in the uptake from 774 wetter layers. We are not creating a new water uptake model, but use a physically based root 775 water uptake model with an implicit compensation mechanism which has been validated by de 776 Jong van Lier et al. (2008). Also, this model has been tested in others studies (e.g. de Jong van 777 Lier et al. (2008), Casaroli et al. (2010) and Tron et al. (2015)). As described in the original 778 model, this water uptake model is based on an expression for the matric flux potential as a 779 function of the distance to the root, and assuming a depth-independent value of matric flux 780 potential at the root surface, uptake per layer (de Jong van Lier et al. 2008). The weighting 781 factor for root water uptake depth distribution that depends on root length density and root 782 radius (de Jong van Lier et al. 2008) is calculated by the RootBox model as result of soil 783 physical conditions.

784 Our soil-plant-atmosphere model simulated dynamic effects of soil stresses on root 785 growth and root water uptake by alternating the root growth and soil water flow model at a 786 coupling time step of 1 day. However, we could not sample daily fluctuation of root growth in the 787 field experiment; we sampled at the end of the crop season for measurement of total root 788 growth and stress limitation that the root system was submitted to during the cropping season. 789 The simulated root length density agreed with measured field data (Fig. 16). This indicates that 790 the model accurately simulated soybean root development considering soil physical limitation. 791 The results show that the root length density (Fig. 12) can be altered in the soil profile due to 792 rainfall deficit increasing the soil physical limitation to root growth. Mechanical impedance 793 caused the larger stress in both weather conditions, wet and dry seasons. Stress due to matric 794 potential was increased in a drought season favouring increment in total stress (Fig. 13). Root 795 growth modelling with soil physical limitation is fundamental to improve the understanding about 796 soybean response to drought stress, water use efficiency (Engels et al. 2017), evaporation and 797 effective water uptake by roots (Manavalan et al. 2009).

798 Root length density and rooting depth were limited by drought stress (Fig. 12). Rooting 799 depth was 25 cm deeper in wet season (75 cm depth) than in dry season (50 cm depth). This 800 indicates that during drought season increased mechanical and hydric stress strongly reduced 801 rooting depth. Roots grew deeper in conditions with higher water availability (wet season). This 802 shows that root elongation responds directly to mechanical and hydric stress during the growth 803 season. One of the major factors influencing soybean rooting depth is the taproot elongation 804 rate (Manavalan et al. 2009) especially its response to soil physical conditions. In soils that 805 impede root growth (e.g., because of a larger mechanical impedance), successive generations 806 of roots tend to reuse paths of least mechanical resistance (Pierret et al. 2007), such as pre-807 existing structural features like cracks and biopores (Jin et al. 2013). Hydric limitation to water 808 uptake and plant transpiration was increased in a drought season due smaller root system. Our 809 model includes a physically based description of macroscopic root water uptake with an implicit 810 compensation mechanism (de Jong van Lier et al. 2008). It is based on the matric flux potential 811 only; further steps will need to include hydraulic resistances along the soil-plant-atmosphere

812 continuum (de Jong van Lier et al. 2013, Javaux et al. 2013). In the drought season, the 813 combined effect of mechanical and hydric stress favours to reduce the root length density and 814 to increase the number of days where hydric stress due to water uptake appeared. Rainfall 815 deficit decreased the soil water content. This can increase the mechanical impedance 816 exponentially (Moraes et al. 2012), and represents the main stress to root growth. That can 817 favour a reduced plant transpiration due to physical barriers impeding root elongation (Tardieu 818 2013), decreasing the overall water flux from soil to rhizosphere (de Jong van Lier et al. 2013). 819 In drought season smaller root systems took up only 50% of water compared to the wet 820 season, and therefore acted as a major limitation to shoot development and grain yield 821 (Saikumar et al. 2016). Water stress during early reproductive growth (flowering and pod set) 822 reduces yield, usually as a result of fewer pods and seeds per unit area (Manavalan et al. 823 2009). For soybean the response of root growth to drought stress is controversial (Hirasawa et 824 al. 1994; Franchini et al. 2017). In some experiment no reduction of root length density due to 825 water stress during the vegetative growth was observed (Hirasawa et al. 1994). However, under 826 different field conditions, soybean root length density was reduced strongly due drought during 827 vegetative growth (Franchini et al. 2017). The consensus is that either way drought stress can 828 be a major limitation to the production and yield stability of soybean (Manavalan et al. 2009). 829 Therefore larger and deeper root systems are necessary to maintain water absorption (Lynch 830 2013; Tron et al. 2015) and plant transpiration for longer periods (Engels et al. 2017), with 831 interaction of hydraulic conductivity in the soil and in the plant (Tardieu et al. 2017). As a 832 consequence, optimum root systems for water uptake at a given time are not always those 833 associated with the best yields (Tardieu et al. 2017). One substantial issue that remains is how 834 to best simulate root growth in structured soils, where cracks and biopores offer low-resistance 835 channels for root growth. The field experiment was performed on the Rhodic Eutrudox soil in a 836 no-tillage system where many cracks and biopores were visible (Silva et al. 2014). By modifying 837 the stress reduction function for penetration resistance (Eq. 16), it was possible to obtain 838 qualitatively similar simulations of root growth (Fig. 15) comparable with field trench-wall root 839 distribution maps. However, the importance of the exact relation between root elongation rate 840 and penetration resistance, and how it might appropriately be modified to account for root 841 penetrable pore-space, requires considerable further investigation potentially with more detailed 842 simulation approaches such as explicit consideration of macropore geometry (e.g. see Landl et 843 al. (2017)).

844 Eco-hydrological and root architecture models are important paths to increase the 845 understanding of plant-environment interactions and plant physiological processes (Tron et al. 846 2015). Models of root functional architecture could also prove useful for crop improvement as 847 they can be used to derive robust biophysical indexes characteristic of some crop-848 environmental combinations, such as improved root sink terms for water uptake modelling 849 (Pierret et al. 2007). This work has considered the simulation of root growth and water uptake in 850 relation to soil physical conditions and weather. The development of functional-structural 851 models of root systems is a new way to account for root aging, in correlation with variation in 852 physiological properties and to study the influence of age on the uptake at the plant scale 853 (Vetterlein and Doussan 2016). Soil physical conditions affect shoot growth indirectly, by 854 reducing the size and extent of a root system and so restricting the uptake of water and 855 nutrients, if these are not abundant (Bengough 1997). They also may affect shoot growth 856 directly via root-shoot signalling mechanisms (e.g. Masle and Passioura (1987)). A further step 857 would be to consider how grain yield could be modelled from plant transpiration, although this 858 adds a further tier of assumptions and complication to the approach.

859 The main novelty of this model is the combination of mechanical and hydric stresses and 860 their application in a 3-D model of root growth. The response of root elongation to hydric and 861 mechanical stresses has already been known, however their combined effect has never been 862 applied in a model that considers individual stresses for each root (tap root, lateral and 863 secondary root) over a whole cropping season. Also, variation of soil penetration resistance, 864 water content and soil aeration (matric potential) has never been integrated into a root growth 865 model. Thus we created a simple way to consider water flux in the soil-plant-atmosphere 866 system, as well as the resulting variations in a stress reduction function (mechanical and hydric) 867 for root elongation for each root and soil layer.

This is a model that describes the theoretical and applied framework that scientists could
 use to link weather and soil physical conditions to plant growth. Application of this model was
 exemplified for one field site in Brazil, considering two years with contrasting weather

conditions. However, this is only an example of model application; the model can be calibratedand used for different pedoclimatic conditions around the world.

One conceptual advance, that may be of immediate practical application is the use of diagrams such as in Fig. 4c and Fig. 4d to qualitatively explain the way that soil physical properties, weather and management system interact to restrict root growth. In addition, examples of Fig. 10 and Fig 13 can help to understand which physical stresses contribute more to reduction of root elongation, i.e. mechanical or hydric stresses. This type of diagram (Fig. 1,4) may be of help in teaching or explaining the interaction between plant physiological responses, soil properties and weather to both scientists and land managers.

Conclusions

881 882

883 884

897 898

899 900

901 902

903

904

905

906

907

908

909

910

911

912

913

885 This model represents a relatively simple approach to modelling root growth under 886 different soil and weather conditions. The proposed framework with mechanical and hydric 887 stresses implemented into the RootBox model offers a way to explore the interaction between 888 soil physical properties, weather and root growth. In particular, it helps to explain the interaction 889 between plant physiological responses and individual soil physical stresses. This root growth 890 model separates the total stress reduction functions into two effects, one the penetration 891 resistance (mechanical stress) and the other one the matric potential (aeration and water 892 stress). The relative contribution of the stress reduction factor values favour understanding 893 which physical stress contribute more for to reducing the root growth. It may be applied to most 894 root elongation models, and offers the potential to evaluate likely factors limiting root growth in 895 different soils and tillage regimes. 896

References

- Addiscott TM, Whitmore AP (1987) Computer simulation of changes in soil mineral nitrogen and crop nitrogen during autumn, winter and spring. J Agric Sci 109:141 . doi: 10.1017/S0021859600081089
- Allen RG, Pereira LS, Raes D, Smith M (1998) Crop evapotranspiration: Guidelines for computing crop requirements. Irrig Drain Pap No 56, FAO 300
- Allen RG, Pereira LS, Smith M, et al (2005) Dual Crop Coef cient Method for Estimating Evaporation from Soil and Application Extensions. Irrig Drain 131:2–13 . doi: 10.1061/(ASCE)0733-9437(2005)131
- Bengough AG (2006) Root responses to soil physical conditions; growth dynamics from field to cell. J Exp Bot 57:437–447 . doi: 10.1093/jxb/erj003
- Bengough AG (2012) Root elongation is restricted by axial but not by radial pressures: So what happens in field soil? Plant Soil 360:15–18 . doi: 10.1007/s11104-012-1428-8
- Bengough AG (1997) Modelling Rooting Depth and Soil Strength in a Drying Soil Profile. J Theor Biol 186:327–338. doi: 10.1006/jtbi.1996.0367
- Bengough AG, McKenzie BM, Hallett PD, Valentine TA (2011) Root elongation, water stress,
 and mechanical impedance: A review of limiting stresses and beneficial root tip traits. J
 Exp Bot 62:59–68. doi: 10.1093/jxb/erq350
- Bengough AG, Mullins CE (1991) Penetrometer resistance, root penetration resistance and root
 elongation rate in two sandy loam soils. Plant Soil 131:59–66. doi: 10.1007/BF00010420
- 919Benjamin JG, Nielsen DC (2006) Water deficit effects on root distribution of soybean, field pea920and chickpea. F Crop Res 97:248–253 . doi: 10.1016/j.fcr.2005.10.005
- 921Bodner G, Leitner D, Nakhforoosh A, et al (2013) A statistical approach to root system922classification. Front Plant Sci 4:1–16. doi: 10.3389/fpls.2013.00292
- Bonfante A, Basile A, Acutis M, et al (2010) SWAP, CropSyst and MACRO comparison in two
 contrasting soils cropped with maize in Northern Italy. Agric Water Manag 97:1051–1062.
 doi: 10.1016/j.agwat.2010.02.010
- 926 Busscher WJ (1990) Adjustment of flat-tipped penetrometer resistance data to a common water 927 content. Trans ASAE 33:0519–0524 . doi: 10.13031/2013.31360
- 928 Casaroli D, de Jong van Lier Q, Dourado Neto D (2010) Validation of a root water uptake model
 929 to estimate transpiration constraints. Agric Water Manag 97:1382–1388 . doi:
 930 10.1016/j.agwat.2010.04.004

931 Celia MA, Bouloutas ET (1990) A general Mass-Conservative Numerical solutuion for 932 Unsaturated flow equation. Water Resour Res 26:1483-1496

936

937

956

957

964

967

968

976

977

978

979

980

981

982

983

984

- 933 Clausnitzer V, Hopmans JW (1994) Simultaneous modeling of transient three-dimensional root 934 growth and soil water flow. Plant Soil 164:299-314 . doi: 10.1007/BF00010082 935
 - de Jong van Lier Q, van Dam JC, Durigon A, et al (2013) Modeling Water Potentials and Flows in the Soil–Plant System Comparing Hydraulic Resistances and Transpiration Reduction Functions. Vadose Zo J 12:1–20 . doi: 10.2136/vzj2013.02.0039
- 938 de Jong van Lier Q, van Dam JC, Metselaar K, et al (2008) Macroscopic Root Water Uptake 939 Distribution Using a Matric Flux Potential Approach. Vadose Zo J 7:1065 . doi: 940 10.2136/vzj2007.0083
- 941 Diggle AJ (1988b) Rootmap: a root growth model. Math Comput Simul 30:175-180 . doi: 942 10.1016/0378-4754(88)90121-8
- 943 Diggle AJ (1988a) ROOTMAP-a model in three-dimensional coordinates of the growth and 944 structure of fibrous root systems. Plant Soil 105:169-178 . doi: 10.1007/BF02376780
- 945 Dresbøll DB, Thorup-Kristensen K, McKenzie BM, et al (2013) Timelapse scanning reveals 946 spatial variation in tomato (Solanum lycopersicum L.) root elongation rates during partial 947 waterlogging. Plant Soil 369:467-477 . doi: 10.1007/s11104-013-1592-5
- 948 Dunbabin VM, Postma JA, Schnepf A, et al (2013) Modelling root-soil interactions using three-949 dimensional models of root growth, architecture and function. Plant Soil 372:93-124 . doi: 950 10.1007/s11104-013-1769-y
- 951 Dupuy L, Gregory PJ, Bengough AG (2010) Root growth models: Towards a new generation of 952 continuous approaches. J Exp Bot 61:2131-2143 . doi: 10.1093/jxb/erp389
- 953 Engels C, Rodrigues F, Ferreira A, et al (2017) Drought Effects on Soybean Cultivation - A 954 Review. Annu Res Rev Biol 16:1-13 . doi: 10.9734/ARRB/2017/35232 955
 - Feddes RA, Kowalik PJ, Zaradny H (1978) Simulation of field water use and crop yield. Pudoc, Wageningen, Netherlands
- Foy CD (1992) Soil Chemical Factors Limiting Plant Root Growth. In: Hatfield J. L, Stewart BA 958 (eds) Advances in Soil Science: Limitations to Plant Root Growth, volume 19. Springer 959 New York, New York, NY, pp 97-131
- 960 Franchini JC, Balbinot Junior AA, Debiasi H, et al (2017) Root growth of soybean cultivars 961 under different water availability conditions. Semin Ciências Agrárias 38:715-724 . doi: 962 10.5433/1679-0359.2017v38n2p715 963
 - Greenwood DJ, Neeteson JJ, Draycott A (1985) Response of potatoes to N fertilizer: Dynamic model. Plant Soil 85:185-203 . doi: 10.1007/BF02139623
- 965 Gregory PJ (2006) Plant Roots: Growth, Activity and Interaction with Soils. Blackwell Publishing 966 Ltd, Oxford, UK
 - Hartmann A, Šimunek J (2016) Hydrus: Root Growth module, Version 1. Department of Environmental Sciences, University of California Riverside, Riverside, California, USA
- 969 Hartmann A, Šimůnek J, Aidoo MK, et al (2017) Implementation and Application of a Root 970 Growth Module in HYDRUS. Vadose Zo J 0:0 . doi: 10.2136/vzj2017.02.0040
- 971 Hirasawa T, Tanaka K, Miyamoto D, et al (1994) Effects of Pre-Flowering Soil Moisture Deficits 972 on Dry Matter Production and Ecophysiological Characteristics in Soybean Plants under 973 Drought Conditions during Grain Filling. Japanese J Crop Sci 63:721-730 . doi: 974 10.1626/jcs.63.721 975
 - lijima M, Kato J (2007) Combined Soil Physical Stress of Soil Drying, Anaerobiosis and Mechanical Impedance to Seedling Root Growth of Four Crop Species. Plant Prod Sci 10:451-459 . doi: 10.1626/pps.10.451
 - Javaux M, Couvreur V, Vanderborght J, Vereecken H (2013) Root Water Uptake: From Three-Dimensional Biophysical Processes to Macroscopic Modeling Approaches. Vadose Zo Jounal 12:1-14 . doi: 10.2136/vzj2013.02.0042
 - Javaux M, Schröder T, Vanderborght J, Vereecken H (2008) Use of a Three-Dimensional Detailed Modeling Approach for Predicting Root Water Uptake. Vadose Zo Jounal 7:1079-1088 . doi: 10.2136/vzi2007.0115
 - Jin K, Shen J, Ashton RW, et al (2013) How do roots elongate in a structured soil? J Exp Bot 64:4761-4777 . doi: 10.1093/jxb/ert286
- 986 Jones CA, Bland WL, Ritchie JT, Williams JR (1991) Simulation of Root Growth. In: Hanks J, 987 Ritchie JT (eds) Modeling plant and soil systems, 31st edn. Agron. Monogr, ASA, CSSA, 988 SSSA, Madison, WI., pp 91–123
- 989 Kalogiros DI, Adu MO, White PJ, et al (2016) Analysis of root growth from a phenotyping data 990 set using a density-based model. J Exp Bot 67:1045-1058 . doi: 10.1093/jxb/erv573

- Kroes JG, Van Dam JC, Groenendijk P, et al (2008) SWAP version 3.2. Theory description and user manual. Alterra, Wageningen, Netherlands
- Landl M, Huber K, Schnepf A, et al (2017) A new model for root growth in soil with macropores.
 Plant Soil 415:99–116 . doi: 10.1007/s11104-016-3144-2
- Leitner D, Klepsch S, Bodner G, Schnepf A (2010a) A dynamic root system growth model
 based on L-Systems. Plant Soil 332:177–192 . doi: 10.1007/s11104-010-0284-7
- Leitner D, Klepsch S, Knieß A, Schnepf A (2010b) The algorithmic beauty of plant roots an L System model for dynamic root growth simulation. Math Comput Model Dyn Syst 16:575–
 587 . doi: 10.1080/13873954.2010.491360
- Leitner D, Meunier F, Bodner G, et al (2014) Impact of contrasted maize root traits at flowering
 on water stress tolerance A simulation study. F Crop Res 165:125–137. doi:
 10.1016/j.fcr.2014.05.009
 Licht MA, Al-Kaisi M (2005) Strip-tillage effect on seedbed soil temperature and other soil

1004

1015

1016

1017

1018

1019

1020

1021

1022

1023

1027

1028

1029

1030

1031

1032

1033

1034

1035

1036

1037

- Licht MA, Al-Kaisi M (2005) Strip-tillage effect on seedbed soil temperature and other soil physical properties. Soil Tillage Res 80:233–249 . doi: 10.1016/j.still.2004.03.017
- Lipiec J, Horn R, Pietrusiewicz J, Siczek A (2012) Effects of soil compaction on root elongation
 and anatomy of different cereal plant species. Soil Tillage Res 121:74–81. doi:
 10.1016/j.still.2012.01.013
- Lynch JP (2013) Steep, cheap and deep: an ideotype to optimize water and N acquisition by maize root systems. Ann Bot 112:347–357 . doi: 10.1093/aob/mcs293
- Lynch JP, Nielsen KL, Davis RD, Jablokow AG (1997) SimRoot: Modelling and visualization of root systems. Plant Soil 188:139–151. doi: 10.1023/A:1004276724310
- Manavalan LP, Guttikonda SK, Phan Tran L-S, Nguyen HT (2009) Physiological and Molecular
 Approaches to Improve Drought Resistance in Soybean. Plant Cell Physiol 50:1260–1276
 doi: 10.1093/pcp/pcp082
 - Masle J, Passioura J (1987) The Effect of Soil Strength on the Growth of Young Wheat Plants. Aust J Plant Physiol 14:643 . doi: 10.1071/PP9870643

Miransari M (2016a) Soybean Tillage Stress. In: Miransari M (ed) Environmental Stresses in Soybean Production, 1st Ed. Elsevier, Amsterdam, pp 41–60

- Miransari M (2016b) Soybean Production and Compaction Stress. In: Miransari M (ed) Environmental Stresses in Soybean Production, 1st Ed. Elsevier, Amsterdam, pp 251–271
- Moraes MT de, Debiasi H, Carlesso R, et al (2017) Age-hardening phenomena in an oxisol from the subtropical region of Brazil. Soil Tillage Res 170:27–37 . doi: 10.1016/j.still.2017.03.002
- Moraes MT de, Debiasi H, Carlesso R, et al (2016) Soil physical quality on tillage and cropping
 systems after two decades in the subtropical region of Brazil. Soil Tillage Res 155:351–
 doi: 10.1016/j.still.2015.07.015
 - Moraes MT de, Debiasi H, Franchini JC, Silva VR da (2012) Correction of resistance to penetration by pedofunctions and a reference soil water content. Rev Bras Ciência do Solo 36:1704–1713. doi: 10.1590/S0100-06832012000600004
 - Moraes MT de, Debiasi H, Franchini JC, Silva VR da (2013) Soil penetration resistance in a rhodic eutrudox affected by machinery traffic and soil water content. Eng Agrícola 33:748–757. doi: 10.1590/S0100-69162013000400014
 - Mualem Y (1976) A new model for predicting the hydraulic conductivity of unsaturated porous media. Water Resour Res 12:513–522 . doi: 10.1029/WR012i003p00513
 - Ortigara C, Moraes MT de, Debiasi H, et al (2015) Modeling of soil load-bearing capacity as a function of soil mechanical resistance to penetration. Rev Bras Ciência do Solo 39:1036–1047. doi: 10.1590/01000683rbcs20140732
- Pagès L, Bécel C, Boukcim H, et al (2014) Calibration and evaluation of ArchiSimple, a simple
 model of root system architecture. Ecol Modell 290:76–84 . doi:
 10.1016/j.ecolmodel.2013.11.014
- 1041Pagès L, Jordan MO, Picard D (1989) A simulation model of the three-dimensional architecture
of the maize root system. Plant Soil 119:147–154 . doi: 10.1007/BF02370279
- 1043Pagès L, Vercambre G, Drouet J-L, et al (2004) Root Typ: a generic model to depict and1044analyse the root system architecture. Plant Soil 258:103–119 . doi:104510.1023/B:PLSO.0000016540.47134.03
- 1046Pereira LS, Allen RG, Smith M, Raes D (2015) Crop evapotranspiration estimation with FAO56:1047Past and future. Agric Water Manag 147:4–20 . doi: 10.1016/j.agwat.2014.07.031
- Pierret A, Doussan C, Capowiez Y, et al (2007) Root Functional Architecture: A Framework for
 Modeling the Interplay between Roots and Soil. Vadose Zo J 6:269–281. doi:
 10.2136/vzj2006.0067

- 1051Postma JA, Kuppe C, Owen MR, et al (2017) OpenSimRoot: widening the scope and1052application of root architectural models. New Phytol 215:1274–1286 . doi:105310.1111/nph.14641
- 1054 Prusinkiewicz P, Lindenmayer A (1990) The Algorithmic beauty of plants. Springer-Verlag, New 1055 York
- 1056Ritchie JT (1972) Model for predicting evaporation from a row crop with incomplete cover. Water1057Resour Res 8:1204–1213 . doi: 10.1029/WR008i005p01204
- Rosa RD, Paredes P, Rodrigues GC, et al (2012) Implementing the dual crop coefficient approach in interactive software. 1. Background and computational strategy. Agric Water Manag 103:8–24. doi: 10.1016/j.agwat.2011.10.013
- Saglio PH, Rancillac M, Bruzan F, Pradet A (1984) Critical Oxygen Pressure for Growth and
 Respiration of Excised and Intact Roots. PLANT Physiol 76:151–154 . doi:
 10.1104/pp.76.1.151
- Saikumar S, Varma CMK, Saiharini A, et al (2016) Grain yield responses to varied level of moisture stress at reproductive stage in an interspecific population derived from Swarna /O glaberrima introgression line. NJAS - Wageningen J Life Sci 78:111–122 . doi: 10.1016/j.njas.2016.05.005
- Schmidt S, Gregory PJ, Grinev D V, Bengough AG (2013) Root elongation rate is correlated
 with the length of the bare root apex of maize and lupin roots despite contrasting
 responses of root growth to compact and dry soils. Plant Soil 372:609–618. doi:
 10.1007/s11104-013-1766-1
- Schnepf A, Leitner D, Klepsch S (2012) Modeling Phosphorus Uptake by a Growing and
 Exuding Root System. Vadose Zo J 11: . doi: 10.2136/vzj2012.0001
- 1074 Schnepf A, Leitner Ď, Landl M, et al (2017) CRootBox: A structural-functional 1 modelling 1075 framework for root systems 2. Biorxiv 3:139980 . doi: 10.1101/139980
- Schnepf A, Leitner D, Schweiger PF, et al (2016) L-System model for the growth of arbuscular mycorrhizal fungi, both within and outside of their host roots. J R Soc Interface 13:1–11.
 doi: 10.1098/rsif.2016.0129
- Silva AP da, Babujia LC, Franchini JC, et al (2014) Soil structure and its influence on microbial biomass in different soil and crop management systems. Soil Tillage Res 142:42–53. doi: 10.1016/j.still.2014.04.006
- 1082
1083Silva AP da, Kay BD, Perfect E (1994) Characterization of the Least Limiting Water Range of
Soils. Soil Sci Soc Am J 58:1775 . doi: 10.2136/sssaj1994.03615995005800060028x
- 1084Šimůnek J, Hopmans JW (2009) Modeling compensated root water and nutrient uptake. Ecol1085Modell 220:505–521 . doi: 10.1016/j.ecolmodel.2008.11.004
- Tardieu F (2013) Plant response to environmental conditions: assessing potential production,
 water demand, and negative effects of water deficit. Front Physiol 4:1–11. doi:
 10.3389/fphys.2013.00017
- Tardieu F, Draye X, Javaux M (2017) Root Water Uptake and Ideotypes of the Root System:
 Whole-Plant Controls Matter. Vadose Zo J 16:0. doi: 10.2136/vzj2017.05.0107
 - Taylor HM, Ratliff LF (1969) Root elongation rates of cotton and peanuts as a function of soil strength and water content. Soil Sci 108:113–119

1091

- Taylor HM, Roberson GM, Parker JJ (1966) Soil strength-root penetration relations for mediumto coarse-textured soil materials. Soil Sci 102:18–22. doi: 10.1097/00010694-196607000-00002
- 1096Tron S, Bodner G, Laio F, et al (2015) Can diversity in root architecture explain plant water use1097efficiency? A modeling study. Ecol Modell 312:200–210 . doi:109810.1016/j.ecolmodel.2015.05.028
- Valentine TA, Hallett PD, Binnie K, et al (2012) Soil strength and macropore volume limit root
 elongation rates in many UK agricultural soils. Ann Bot 110:259–270. doi:
 10.1093/aob/mcs118
- van Dam JC, Feddes RA (2000) Numerical simulation of infiltration, evaporation and shallow groundwater levels with the Richards equation. J Hydrol 233:72–85. doi: 10.1016/S0022-1694(00)00227-4
- 1105van Genuchten MT (1980) A Closed-form Equation for Predicting the Hydraulic Conductivity of1106UnsaturatedSoils.SoilSciSocAmJ44:892–898doi:110710.2136/sssaj1980.03615995004400050002x
- 1108Vereecken H, Schnepf A, Hopmans JW, et al (2016) Modeling Soil Processes: Review, Key1109Challenges, and New Perspectives. Vadose Zo J 15:1–57 . doi: 10.2136/vzj2015.09.0131
- 1110 Vetterlein D, Doussan C (2016) Root age distribution: how does it matter in plant processes? A

- 1111 focus on water uptake. Plant Soil 407:145–160 . doi: 10.1007/s11104-016-2849-6
- 1112 Willmott CJ, Robeson SM, Matsuura K (2012) A refined index of model performance. Int J 1113 Climatol 32:2088–2094 . doi: 10.1002/joc.2419
- Wu L, McGechan MB, McRoberts N, et al (2007) SPACSYS: Integration of a 3D root architecture component to carbon, nitrogen and water cycling—Model description. Ecol Modell 200:343–359 . doi: 10.1016/j.ecolmodel.2006.08.010
- Wu Y, Cosgrove DJ (2000) Adaptation of roots to low water potentials by changes in cell wall
 extensibility and cell wall proteins. J Exp Bot 51:1543–1553 . doi:
 10.1093/jexbot/51.350.1543
- 1120

 Table 1
 Van Genuchten's parameter of a Rhodic Eutrudox under no-tillage system.

Depth	θs	θr	А	n	Ks	Bulk density
cm	cm ³ cm ⁻³	cm ³ cm ⁻³	cm⁻¹		cm day⁻¹	Mg m ⁻³
0-10	0.555	0.198	0.0892	1.1848	39.36	1.21
10-20	0.537	0.200	0.0822	1.1503	39.36	1.26
20-30	0.539	0.200	0.0756	1.1407	54.15	1.26
30-40	0.539	0.200	0.0756	1.1407	54.15	1.16
40-50	0.539	0.200	0.0756	1.1407	54.15	1.10
50-60	0.539	0.200	0.0756	1.1407	54.15	1.08
60-80	0.539	0.200	0.0756	1.1407	54.15	1.06
80-100	0.539	0.200	0.0756	1.1407	54.15	1.05

* θ r, θ s, α , and n are van Genuchten's parameters; Ks: hydraulic conductivity saturated;

Table 2 Root architectural	parameters of sov	ybean (<i>Glycine max</i>).
		, , , , , ,

Symbol	Parameter name	units	Values [mean, s.d.]
Tap root			
re	Initial tip elongation rate	cm day⁻¹	[5.5, 0]
а	Root radius	cm	[0.2, 0]
la	Length of apical zone	cm	[2.0, 0]
l _b	Length basal zone	cm	[1.0, 0]
In	Internodal distance	cm	[0.65, 0]
Nb	Maximum number of branches	-	[300, 0]
σ	Expected change of root tip heading	rad cm ⁻¹	0.4
type	Type of tropism	-	1
Ň	Strength of tropism	-	1.5
dx	Spatial resolution along root axis	cm	0.25
First-order laterals			
ſe	Initial tip elongation rate	cm dav⁻¹	[1.5. 0]
a	Root radius	cm	[0.05. 0]
Õ	Insertion angle	rad	[1.2217. 0]
a	Length of apical zone	cm	[3, 0]
b	l ength basal zone	cm	[3, 0]
	Internodal distance	cm	[0,7, 0]
n _b	Maximum number of branches	-	[60, 0]
σ	Expected change of root tip heading	rad cm ⁻¹	0.3
type	Type of tropism	-	1
N	Strength of tropism	-	0.1
dx	Spatial resolution along root axis	cm	0.25
Second-order laterals	oparial recordion along reet axie	onn	0.20
	Initial tip elongation rate	cm dav⁻¹	[1 0]
а	Root radius	cm	[0, 03, 0]
Ð	Insertion angle	rad	[1 22173 0]
K C	Maximal root length	cm	[2 0]
σ	Expected change of root tip heading	rad cm ⁻¹	0.4
type	Type of tropism	-	1
N	Strength of tropism	-	0
dx	Spatial resolution along root axis	cm	0 25
Basal roots	opular resolution along root axis	om	0.20
r _o	Initial tip elongation rate	cm dav ⁻¹	[2 0]
2	Root radius	cm	
e e	Insertion angle	rad	[1 39626 0]
	l ength of anical zone	cm	[1.55020, 0]
ia L	Length basal zone	cm	[10, 0]
	Internodal distance	cm	[2, 0] [2, 0]
n D-	Maximum number of branches	-	[2, 0]
α	Expected change of root tip heading	rad cm ⁻¹	[+0, 0] 0 1
type	Type of tropism		1
N	Strength of tropism	-	0.5
dv.	Spatial resolution along root avia	- Cm	0.0
un l	opanal resolution along root alls		0.20

s.d. is the standard deviation.

season (2009/2010).			
Parameters	Value	2008/2009	2009/2010
Crop transpiration*			
Kcb ini (20 days)	0-0.15	24/11 – 13/12	11/11 – 30/11
Kcb dev (35 days)	0.15-1.10	14/12 – 06/01	01/12 – 24/12
Kcb mid (40 days)	1.10	07/01 – 04/03	25/12 – 19/02
Kcb end (30 days)	1.10-0.30	05/03 - 29/03	20/02 - 16/03
Root sampling date		02/02/2009	20/01/2010
Soil evaporation			
REW (mm)		45	45
TEW (mm)		13.5	13.5
FC (m ³ m ⁻³)		0.35	0.35
WP (m ³ m ⁻³)		0.25	0.25
Ze (m)		0.10	0.10

Table 3. Soybean growth stage date and crop and soil evaporation parameters for estimating evapotranspiration using the dual crop coefficient approach for a drought (2008/2009) and a wet season (2009/2010).

*Basal crop coefficients (Kcb) were calculated during the crop growing season for initial (ini); crop development (Kcb dev); midseason (Kcb mid); and end season (Kcb end). TEW: total evaporable water; REW: readily evaporable water; FC: field capacity; WP: wilting point; Ze: thickness of the evaporation soil layer.



Fig. 1 Schematic representation of the model coupling in soil-root-plant-atmosphere relationship.







Fig. 3 Measured vs. calculated values of soil penetration resistance (Q_P) for a Rhodic Eutrudox, very clayed. Dashed line represents a one-to-one relationship. The Busscher's parameters were a=0.00587; b=8.0772; c=-4.65. The data set are from Ortigara et al. (2015). MAE: means absolute error; RMSE: root mean squared error; CRM: coefficient of residual mass; r: coefficient of correlation; EF: modelling efficiency; and d: index of agreement.



Fig. 4 Root elongation (RE) parameter as a function of matric potential (adapted from Feddes et al. (1978)) (a) and soil penetration resistance with presence (dashed blue line) or absence (red line adapted from Bengough (1997)) of continuous pores (b), and the total stress reduction function (srf) in a colour map for a soil without (c) or with (d) continuous pores. Root elongation parameter is expressed as an index from 1 (maximum root elongation) to 0 (no root growth). I: data set for a typical agricultural soil; II: data set for a compacted soil.



Fig. 5 Rainfall, reference evapotranspiration (ETo), potential transpiration (Tp) and actual transpiration (Ta) during soybean season growth in a loose soil and in a soil with a compact layer (data from Londrina, Brazil).



Fig. 6 Simulated and field measured values for water content (a) and soil penetration resistance (b) at 8 cm and 16 cm depths: points show average measured values under field conditions in Londrina/Brazil during a wet-dry cycle.





Fig. 7 Simulations of soybean root system distribution (a,b) and stress reduction function (c,d) and root length density over time (e,f) in soil without (a,c,e) or with (b,d,f) a compact layer from 16 to 20 cm. Timelapse video of root growth can seen at the supplementary material S1 (profile without soil compaction) and S1 (profile with a soil compacted layer).



Fig. 8 Water uptake versus depth and time in (a) a soil with no compact layer, or (b) with a compact layer at 16-20 cm depth.



Fig. 9 Soil penetration resistance (a,b) and soil volumetric water content (c,d) in soil without compact layer (a,c), or in a soil with a compact layer (b,d).



Fig. 10 Relative frequency of total stress reduction function (srf) (a,b) and effect from soil penetration resistance (c,d) or matric potential (e,f) and in the loose soil (a,c,e) or the soil with a compacted layer (b,d,f) from 16 to 20 cm depth.



Fig. 11 Rainfall, reference evapotranspiration (Eto), potential (Tp) and actual transpiration (Ta) during soybean season growth in a drought – 2008/2009 (a) or a wetter season – 2009/2010 (b).



Fig. 11 Rainfall, reference evapotranspiration (Eto), potential (Tp) and actual transpiration (Ta) during soybean season growth in a drought – 2008/2009 (a) or a wetter season – 2009/2010 (b).



Fig. 12 Simulated soybean root system (a,d), root length density (RLD) (b,e) and stress reduction function (srf) (c,f) in the drier season (a,b,c) or wetter season (d,e,f) growth. Timelapse video of root growth can seen at the supplementary material S3 (drier season) and S4 (wetter season).



Class of stress reduction function

Fig. 13 Frequency of the total stress (a) or soil resistance to penetration (b) and matric potential (c) for root elongation in all rooting depth during the first 54 days of soybean growth in a Rhodic Eutrudox. Values close to zero correspond to the absence of growth, while values close to 1 mean maximum potential of root elongation.



Fig. 14 Water uptake rate (mm per day) for a dry season (a) or a wet season (b) growth.







Fig. 16 Root length density measured on field and simulated for a dry season (a) and wet season (b) and relation 1 to 1 (c). *RMSE: root mean squared error; CRM: coefficient of residual mass; r:

coefficient of correlation; EF: modelling efficiency; d: index of agreement; MAE: means absolute error.