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RESEARCH NOTE

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A conditional Pax6 depletion study with no morphological effect on the adult mouse corneal epithelium

Natalie J. Dorà^{1,6}, Martine Manuel², Dirk-Jan Kleinjan^{3,7}, David J. Price², J. Martin Collinson⁴, Robert E. Hill³ and John D. West^{5*} 

Abstract

Objective: The corneas of heterozygous *Pax6*^{+/-} mice develop abnormally and deteriorate further after birth but it is not known whether the postnatal deterioration is predetermined by abnormal development. Our objective was to identify whether depletion of Pax6 in adult mice caused any corneal abnormalities, similar to those in *Pax6*^{+/-} mice, where Pax6 levels are low throughout development and adulthood. We used two tamoxifen-inducible, *Cre-loxP* experimental strategies to deplete Pax6 either ubiquitously or in a restricted range of cell types.

Results: In a preliminary study, ubiquitous depletion of Pax6 by tamoxifen treatment of E9.5 CAG-CreER^{Tg/-}; *Pax6*^{fl/fl} embryos affected eye development. Tamoxifen treatment of 12-week old, adult CAG-CreER^{Tg/-}; *Pax6*^{fl/+} and CAG-CreER^{Tg/-}; *Pax6*^{fl/fl} mice resulted in weak and/or patchy Pax6 immunostaining in the corneal epithelium but caused no corneal abnormalities. GFP staining in tamoxifen-treated CAG-CreER^{Tg/-}; RCE:*loxP* reporter mice was also patchy. We attribute patchy Pax6 staining to mosaic deletion of the *Pax6*^{fl} allele, probably caused by mosaic CAG-CreER^{Tg} expression. In a parallel study, we treated adult *Krt19*-CreER^{Tg/-}; *Pax6*^{fl/+} mice with tamoxifen to try to deplete Pax6 in limbal epithelial stem cells (LESCs) which replenish the corneal epithelium. However, Pax6 staining remained strong after a 12-week chase period so the *Krt19*-CreER^{Tg/-} transgene may have failed to target LESCs.

Keywords: Cornea, Corneal epithelium, *Cre-loxP*, CAG-CreER, *Krt19*-CreER, Pax6, Mouse, Mosaic transgene expression

Introduction

The mouse cornea comprises an outer epithelium of 5–6 cell layers, a thick stroma and an inner endothelium. The corneal epithelium is maintained by limbal epithelial stem cells (LESCs), in the ring-shaped limbus, which is a transition zone between the corneal epithelium and the conjunctiva. The LESCs replace themselves and produce transient (or transit) amplifying cells (TACs) that move centripetally across the basal corneal epithelium and produce more differentiated daughter cells, which move apically, through the epithelial layers, and are shed from the surface [1].

The *Pax6* gene, encoding the Pax6 transcription factor, is expressed in the brain, pancreas, olfactory system and several eye tissues, including the corneal and limbal epithelia, and is critical for eye development [2]. Low levels of Pax6 throughout development of heterozygous *Pax6*^{+/-} mice cause small eyes, aniridia plus lens and corneal defects [3–8]. The newborn *Pax6*^{+/-} cornea has a thin epithelium and the adult cornea deteriorates further, because the epithelium is poorly maintained and limbal blood vessels invade the stroma [6–8]. The adult *Pax6*^{+/-} corneal epithelium is thin and fragile, cell turnover is elevated, centripetal movement is disrupted and goblet cells accumulate. The expression of keratin 12 (K12), which is regulated by Pax6 [9], is delayed and immunostaining is weak and patchy [6–8, 10, 11]. Indirect evidence suggests that reduced Pax6 causes LESCS deficiency in both

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PAX6^{+/-} humans and *Pax6*^{+/-} mice and this may underlie adult corneal deterioration [1, 12].

It is not known if all corneal abnormalities in adult *Pax6*^{+/-} mice are predetermined by abnormal development or whether some are caused by reduced Pax6 in the adult. Our aim was to determine whether depletion of Pax6 in adult mice caused corneal abnormalities, comparable to those reported for *Pax6*^{+/-} mice. We used one experimental strategy to deplete Pax6 ubiquitously and another to deplete Pax6 in LESC.

Main text

Materials and methods

Mice

To deplete Pax6 ubiquitously, CAG-CreER^{Tg/-}; *Pax6*^{fl/+} mice were produced by crossing hemizygous CAG-CreER^{Tg/-} mice (formal transgene name: Tg(CAG-cre/Esr1*)5Amc) [13] to heterozygous *Pax6*^{fl/+} mice (formal name: *Pax6*^{tm1Ued/+}) [14]. Superscript symbols 'Tg/-' and '-/-' are used to distinguish hemizygous CAG-CreER^{Tg/-} mice and non-transgenic CAG-CreER^{-/-} siblings.

Keratin 19 (*Krt19* gene; K19 protein) is expressed in the basal epithelium of the mouse limbus and conjunctiva but not the cornea [15]. To try to target LESC in the limbal epithelium, *Krt19*-CreER^{Tg/-}; *Pax6*^{fl/+} mice were produced by crossing hemizygous *Krt19*-CreER^{Tg/-} mice (formal transgene name: *Krt19*^{tm1(cre/ERT)Ggu}) [16] to heterozygous *Pax6*^{fl/+} mice.

RCE:*loxP* mice have the R26R CAG-boosted EGFP (RCE) reporter allele with an upstream *loxP*-flanked STOP cassette (formal transgene name: *Gt(ROSA)26Sor*^{tm1.1(CAG-EGFP)Fsh}) [17]. CAG-CreER^{Tg/-};RCE:*loxP* mice with tamoxifen-inducible expression of GFP were bred by crossing CAG-CreER^{Tg/-} and RCE:*loxP* mice. *Krt19*-CreER^{Tg/-};RCE:*loxP* mice were bred by crossing *Krt19*-CreER^{Tg/-} and RCE:*loxP* mice.

Mice were maintained on a predominantly CBA/Ca genetic background and genotyped by PCR [13, 14, 16]. Some additional samples from mice on a CD-1 or (C57BL/6 × CBA/Ca)F1 genetic background from other studies [18, 19], were also analysed.

To activate CreER in adult mice, tamoxifen (Sigma-Aldrich) was freshly prepared in corn oil (25–40 mg/ml) by sonication in a 40 °C water bath and adjusted to 100 µg/g body weight in 0.1 ml. Mice of both sexes were injected intraperitoneally with tamoxifen at 12 weeks on 5 consecutive days and analysed 3 days later (no chase group) or after chase periods of 6 or 12 weeks. Control mice were injected with 0.1 ml corn oil. Mice were culled by cervical dislocation, following overdose of gaseous halothane, and eyes were enucleated. Procedures for the induction of Cre expression in embryos at embryonic day

(E) 9.5 and the subsequent collection of E13.5 fetal samples are described elsewhere [19]. Tamoxifen treatment causes CreER to move to the nucleus and recombine *loxP* sites to convert the functional *Pax6*^{fl} floxed allele to a *Pax6*^Δ null allele or express the GFP lineage marker in the target cells and their progeny. This should occur in all cell types in CAG-CreER^{Tg/-} mice, because CreER is expressed ubiquitously from the CAG promoter, but only in specific cell types in *Krt19*-CreER^{Tg/-} mice.

Analysis

Tissue samples were fixed in 4% paraformaldehyde overnight at 4 °C. Fetal heads were processed to OCT compound and stored frozen before cryosections were cut and stained with haematoxylin and eosin (H&E) [19, 20]. Adult eyes were processed to paraffin wax, then 7 µm sections were cut, mounted on glass slides and stained with H&E or periodic acid-Schiff (PAS) stain [20]. Stained sections were photographed and measured using a Zeiss Axioplan-2 microscope and calibrated Zeiss Axiovision 4.8 digital camera system. Numerical data are included in Additional file 1 and measurements were compared by Student's t-test.

Immunohistochemistry methods are described elsewhere [20]. Briefly, wax sections, mounted on glass slides were heat-treated to unmask antigens, then incubated with blocking serum, followed by primary antibody, biotinylated secondary antibody, avidin–biotin reagent and 3,3'-diaminobenzidine (DAB) stain. Sections were then lightly counterstained with haematoxylin, dehydrated and slides were mounted with DPX mounting medium under coverslips. Negative control slides were treated with blocking serum instead of primary antibody. The antibodies used for Pax6 and K12 immunostaining were as described elsewhere [20], except that the secondary antibody was biotinylated rabbit anti-mouse, diluted 1:200 (E0433 from Dako, Ely, UK). For GFP immunostaining, the primary antibody was rabbit anti-GFP diluted 1:500 (ab290 from Abcam, Cambridge, UK) and the secondary antibody was biotinylated goat anti-rabbit, diluted 1:200 (Sc-2012 from SantaCruz Biotechnology, Heidelberg, Germany).

Results

Ubiquitous depletion of Pax6 in embryos

For another study, CAG-CreER^{Tg/-}; *Pax6*^{fl/fl} and CAG-CreER^{Tg/-}; *Pax6*^{fl/+} embryos were exposed to tamoxifen at E9.5 and culled at E13.5 [19]. By E13.5 tamoxifen-treated CAG-CreER^{Tg/-}; *Pax6*^{fl/fl} fetuses (with two floxed *Pax6*^{fl} alleles) had smaller eyes and lenses than CAG-CreER^{Tg/-}; *Pax6*^{fl/+} fetuses (Additional file 1 and Additional file 2: Fig. S1). This showed that tamoxifen-mediated depletion of Pax6 could affect eye development.

Ubiquitous depletion of Pax6 in adults

The preliminary result with fetal eyes encouraged us to investigate whether tamoxifen-mediated, depletion of Pax6 in adults caused any corneal abnormalities, similar to those in *Pax6*^{+/-} mice, where Pax6 is low throughout development. Pax6-depletion in adults is unlikely to reproduce *Pax6*^{+/-} developmental defects but adult corneal deterioration could be mediated via Pax6-deficiency in adult LSCs, the LSC niche, the corneal epithelium or other ocular tissues [5, 10]. We compared the effects of tamoxifen treatment of CAG-CreER^{Tg/-};*Pax6*^{fl/+} mice to several genotype and treatment controls, which were included to control for any unexpected effects of the CAG-CreER transgene or the floxed *Pax6*^{fl} allele alone [20–24].

Following tamoxifen treatment at 12 weeks and a 6-week chase, Pax6 immunostaining was positive in the corneal epithelia of all the control combinations (Fig. 1a–g). Although immunohistochemistry was not quantified,

Pax6 appeared to be weak and/or patchy in the corneal epithelia of CAG-CreER^{Tg/-};*Pax6*^{fl/+} mice, treated with tamoxifen as adults (Fig. 1h–l). However, eye and corneal morphology appeared grossly normal (apart from some processing artefacts), with no blood vessels visible in the cornea (Fig. 2). PAS-positive goblet cells were not detected in the corneal epithelium (data not shown) and there was little or no effect on K12 staining (Fig. 1m–t). Similar results were obtained after a 12-week chase and, again, corneal morphology appeared grossly normal (Fig. 3). For comparison, previously published corneal histology and immunostaining are shown for wild-type *Pax6*^{+/+} and heterozygous *Pax6*^{+/-} eyes in Additional file 2: Fig. S2. GFP immunostaining of eyes from tamoxifen-treated CAG-CreER^{Tg/-};*RCE:loxP* reporter mice showed mosaic expression in the corneal epithelium (Additional file 2: Fig. S3).

We did not include CAG-CreER^{Tg/-};*Pax6*^{fl/fl} mice, with two floxed *Pax6*^{fl} alleles, in the main study because

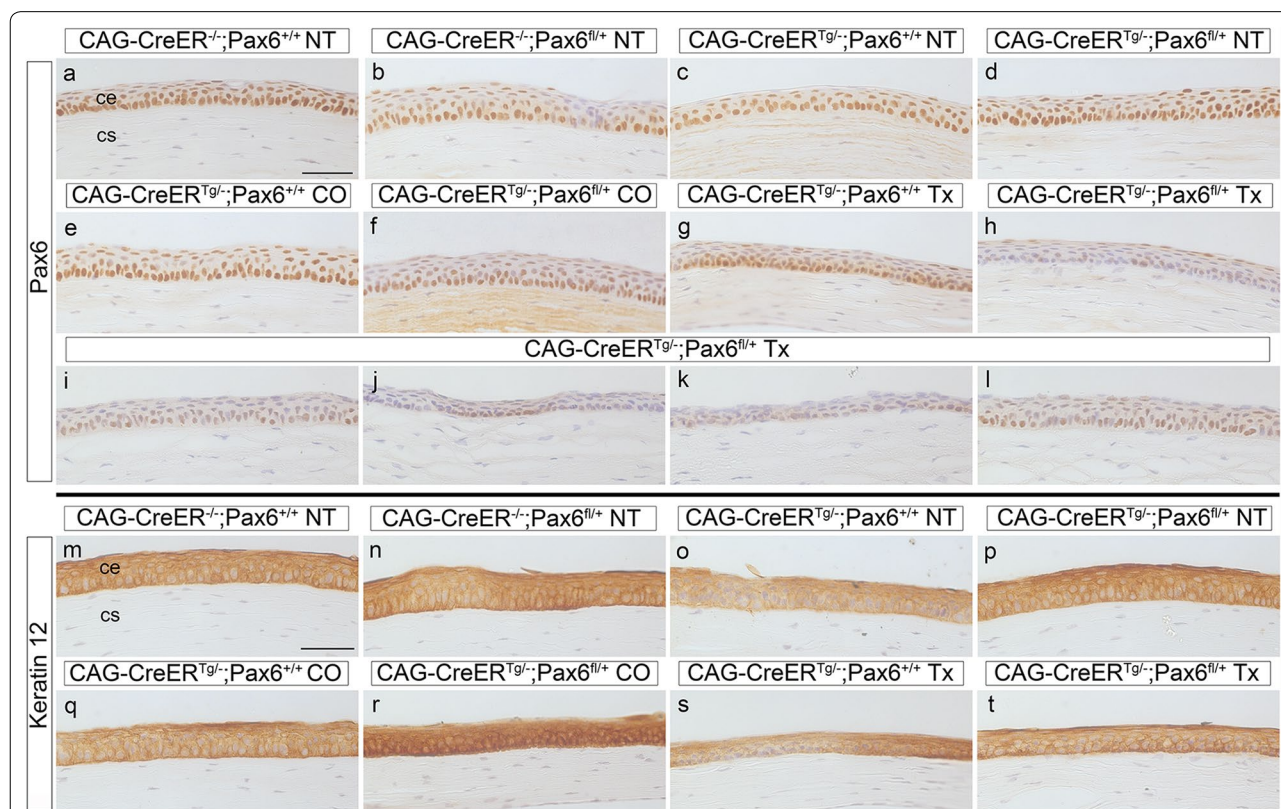
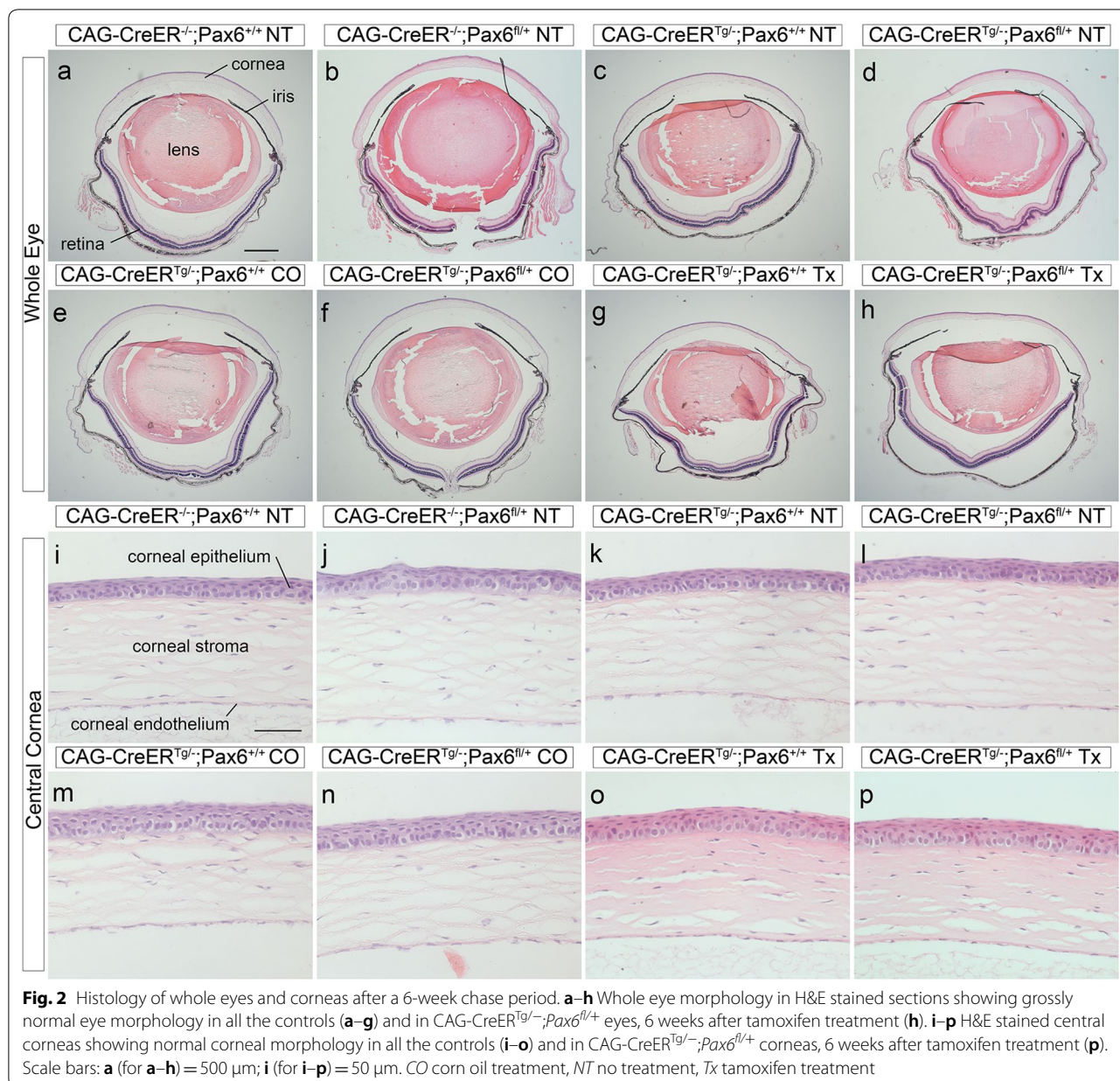


Fig. 1 Pax6 and keratin 12 immunohistochemistry of corneal epithelium after a 6-week chase period. **a–l** Central region of the adult corneal epithelium, immunostained for Pax6 (brown DAB endpoint) and counterstained with haematoxylin, showing various controls with different genotype and treatment combinations, 6 weeks after treatment (**a–g**), and several examples of weak and/or patchy staining of CAG-CreER^{Tg/-};*Pax6*^{fl/+} corneas, 6 weeks after treating with tamoxifen (**h–l**). **m–t** Central region of the adult corneal epithelium, immunostained for keratin 12 (brown DAB endpoint) and counterstained with haematoxylin, showing various controls with different genotype and treatment combinations, 6 weeks after treatment (**m–s**), and a CAG-CreER^{Tg/-};*Pax6*^{fl/+} cornea, 6 weeks after treating with tamoxifen (**t**). Scale bars = 50 μm. ce corneal epithelium, CO corn oil treatment, cs corneal stroma, NT no treatment, Tx tamoxifen treatment. All mice were on a predominantly CBA/Ca genetic background



severe, global depletion of Pax6 in these mice results in diabetes [18]. However, Pax6 immunostaining of corneas from CAG-CreER^{Tg/-};Pax6^{fl/fl} mice, produced for another study [18], showed that Pax6 protein was not eliminated following tamoxifen treatment and a 6-week chase period (Additional file 2: Fig. S4).

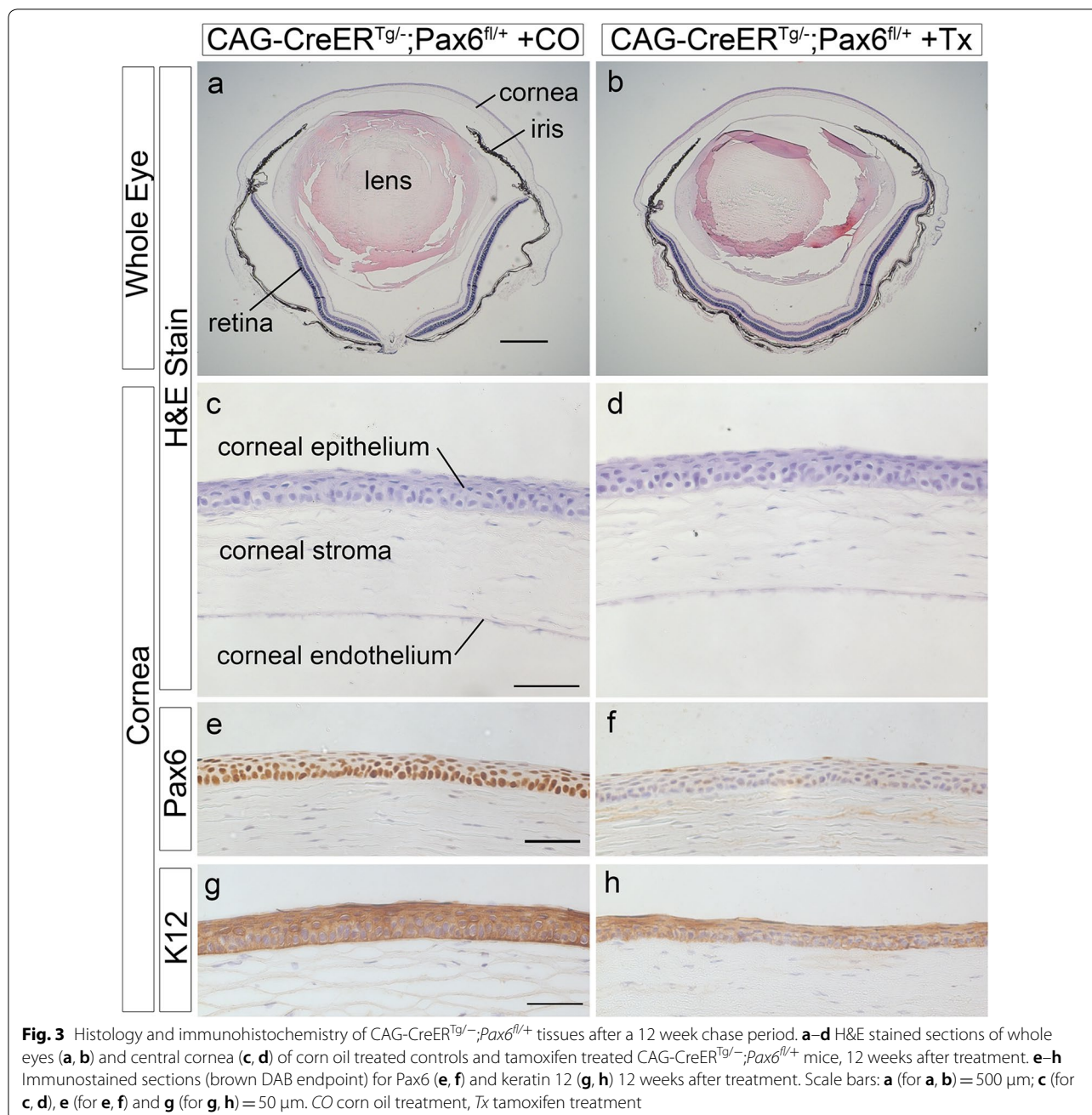
Targeting Pax6-depletion to LSCs

To try to deplete Pax6 in LSCs we treated Krt19-CreER^{Tg/-};Pax6^{fl/+} mice with tamoxifen at 12 weeks and analysed the results after a 12-week chase period, to allow sufficient time for treated LSCs to replenish the

corneal epithelium. Pax6 and K12 staining appeared normal and no corneal morphological abnormalities were seen (Additional file 2: Fig. S5). Tamoxifen-treated Krt19-CreER^{Tg/-};RCE:loxP reporter mice produced patchy GFP reporter immunostaining in the conjunctiva, some sparse staining in the limbus but no staining in the corneal epithelium (Additional file 2: Fig. S6).

Discussion

Tamoxifen treatment to deplete Pax6 in E9.5 CAG-CreER^{Tg/-};Pax6^{fl/fl} embryos affected eye morphology by E13.5 but did not prevent lens development. The lens



placode forms around E9.5 and is absent in Pax6^{-/-} homozygotes [25], so E9.5 tamoxifen-treatment was probably too late to prevent lens development in the CAG-CreER^{Tg/-};Pax6^{fl/fl} embryos but we did not investigate whether any Pax6 remained at E13.5.

As tamoxifen treatment of Krt19-CreER^{Tg/-};RCE:loxP mice did not produce any GFP-positive corneal epithelial cells after a 12-week chase, it is likely that this strategy failed to target LSCs. With hindsight, the Krt14-CreER^{Tg/-} mouse may have been more suitable for targeting

expression to LSCs, as this has been successful in lineage tracing experiments [26, 27].

The normal corneal morphology of the tamoxifen-treated adult CAG-CreER^{Tg/-};Pax6^{fl/+} mice is consistent with the possibility that normal Pax6 levels are not required in the adult to maintain the corneal epithelium. This would suggest that deterioration of the adult Pax6^{+/-} corneal epithelium was predetermined during development. However, we also need to consider technical explanations for our results, particularly as corneal

effects of Pax6-deficiency have now been corrected successfully in adult mice [28, 29].

One possibility is that our investigation was undermined by mosaic deletion of the *Pax6^{fl}* allele. The presence of some Pax6-positive cells in the corneal epithelia of tamoxifen-treated CAG-CreER^{Tg/-}; *Pax6^{fl/fl}* mice suggests that mosaicism occurred in these mice as well as CAG-CreER^{Tg/-}; RCE:*loxP* reporter mice. We, therefore, suggest that mosaic *Pax6^{fl}* deletion was caused by mosaic CAG-CreER^{Tg} transgene expression but recombination of *Pax6^{fl}* *loxP* sites could also be inefficient for other reasons. Mosaic *Pax6^{fl}* deletion would lead to a mixture of *Pax6^{Δ/Δ}*, *Pax6^{fl/Δ}* and *Pax6^{fl/fl}* cells in CAG-CreER^{Tg/-}; *Pax6^{fl/fl}* mice or a mixture of *Pax6^{Δ/+}* and *Pax6^{fl/+}* cells in CAG-CreER^{Tg/-}; *Pax6^{fl/+}* mice.

The effects of mixtures of wild-type and *Pax6^{-/-}* or *Pax6^{+/-}* cells, in ocular tissues, have been investigated using mouse chimaeras. Eye development was abnormal in *Pax6^{+/+}* ↔ *Pax6^{-/-}* chimaeras [30–33] but *Pax6^{+/+}* ↔ *Pax6^{+/-}* chimaeras had normal eyes [5, 10, 33] with normal corneal morphology [10]. It was suggested that unknown signals from wild-type cells in the cornea and/or other ocular tissues might rescue the *Pax6^{+/-}* cells [10]. This might also occur in tamoxifen-treated CAG-CreER^{Tg/-}; *Pax6^{fl/+}* corneas if the conditional *Pax6^{fl}* allele is not deleted in all cells.

Although the corneal epithelia of tamoxifen-treated CAG-CreER^{Tg/-}; *Pax6^{fl/fl}* mice contained many Pax6-positive cells (Additional file 2: Fig. S4), no Pax6 protein was detected by immunofluorescence in most pancreatic islet cells in equivalent mice from the same study [18]. Apparent differences in frequencies of Pax6-positive cells between these two tissues may reflect genuine biological differences rather than technical differences in detecting Pax6-positive cells. Mosaic CAG-CreER^{Tg} expression might be more common and/or recombination of *loxP* sites less efficient in the corneal epithelium than pancreatic islets, resulting in mosaic *Pax6^{fl}* deletion in the cornea.

Mosaic reporter expression also occurred in the corneal epithelium of CAG-CreER;R26R-*LacZ* and CAG-CreER;R26R-mT/mG reporter mice in a lineage-tracing experiment [34]. This lineage tracing experiment was undertaken after the present study and, with hindsight, it would be worth investigating whether mosaic transgene expression is more common for specific tissues and/or specific CAG-CreER;*loxP* combinations.

Limitations

We evaluated corneal histology, the absence of goblet cells and K12 immunohistochemistry. Future investigations could include additional endpoints and markers.

In lineage-tracing experiments, labelled cells produced by tamoxifen-treated LESC took at least 14 weeks to replace the whole corneal epithelium [26, 27, 34]. Thus, although our chase time of 12 weeks should have identified corneal defects attributable to Pax6-depletion in LESC or the niche, it might not have been sufficient to produce the maximum effects.

We did not investigate how effectively Pax6 protein was depleted in other CAG-CreER^{Tg/-}; *Pax6^{fl/+}* ocular tissues or whether mosaic transgene expression occurred in those tissues. Also, DNA or RNA methods were not used to confirm that at least some floxed *Pax6^{fl}* alleles were converted to *Pax6^Δ* in the corneal epithelium.

Additional files

Additional file 1. Fetal eye measurements.

Additional file 2: Fig S1. E13.5 fetal eye morphology following tamoxifen treatment at E9.5. **Fig. S2.** Previously published histology and immunohistochemistry of adult wild-type and heterozygous *Pax6^{+/-}* mouse eyes. **Fig. S3.** Expression of GFP reporter in corneal epithelium of CAG-CreER^{Tg/-}; RCE:*loxP* mice after tamoxifen treatment and different chase periods. **Fig. S4.** Pax6 immunohistochemistry of CAG-CreER^{Tg/-}; *Pax6^{fl/+}* and CAG-CreER^{Tg/-}; *Pax6^{fl/fl}* corneal epithelia after a 6-week chase period. **Fig. S5.** Histology and immunohistochemistry of *Krt19*-CreER^{Tg/-}; *Pax6^{fl/+}* tissues after a 12 week chase period. **Fig. S6.** Expression of GFP reporter in corneal epithelium of *Krt19*-CreER^{Tg/-}; RCE:*loxP* mice after tamoxifen treatment and different chase periods.

Abbreviations

DAB: 3,3'-diaminobenzidine; DPX: distyrene plasticizer xylene; E: embryonic day; EGFP: enhanced green fluorescent protein; GFP: green fluorescent protein; H&E: haematoxylin and eosin; K12: keratin 12; K19: keratin 19; LESC: limbal epithelial stem cell; OCT compound: optimal cutting temperature compound; PAS: periodic acid-Schiff; TAC: transient (or transit) amplifying cell.

Authors' contributions

NJD produced and analysed the results, produced the figures and wrote most of the first draft. MM and DJK provided samples, produced in other studies, for evaluation. DJP designed and supervised the study that provided tamoxifen-treated embryos. JMC and REH designed and supervised part of the main study. JDW designed and supervised part of the main study and wrote part of the manuscript. All authors read and approved the final manuscript.

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Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

The numerical data generated and analysed during this study are included in the additional information files of this published article.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Mice were bred and maintained at the University of Edinburgh, UK. Animal work was approved by the University of Edinburgh Ethical Review Committee and performed in accordance with UK Home Office Regulations under UK Home Office licences PPL 60/3635 and PPL 60/4302. Samples produced in other studies were from mice regulated by UK Home Office licences PPL 60/3785 and PPL 60/3913.

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References

- Mort RL, Douvaras P, Morley SD, Dorà N, Hill RE, Collinson JM, West JD. Stem cells and corneal epithelial maintenance: insights from the mouse and other animal models. In: Kubiak JZ, editor. *Results Probl Cell Differ*, 55, "Mouse development: from oocyte to stem cells". Berlin: Springer; 2012. p. 357–94.
- Shaham O, Menuchin Y, Farhy C, Ashery-Padan R. Pax6: a multi-level regulator of ocular development. *Prog Retin Eye Res*. 2012;31(5):351–76.
- Hill RE, Favor J, Hogan BLM, Ton CCT, Saunders GF, Hanson IM, Prosser J, Jordan T, Hastie ND, VanHeyningen V. Mouse small eye results from mutations in a paired-like homeobox-containing gene. *Nature*. 1991;354(6354):522–5.
- van Raamsdonk CD, Tilghman SM. Dosage requirement and allelic expression of *PAX6* during lens placode formation. *Development*. 2000;127(24):5439–48.
- Collinson JM, Quinn JC, Buchanan MA, Kaufman MH, Wedden SE, West JD, Hill RE. Primary defects in the lens underlie complex anterior segment abnormalities of the *Pax6* heterozygous eye. *Proc Natl Acad Sci USA*. 2001;98(17):9688–93.
- Davis J, Duncan MK, Robison WG, Piatigorsky J. Requirement for Pax6 in corneal morphogenesis: a role in adhesion. *J Cell Sci*. 2003;116(11):2157–67.
- Ramaesh T, Collinson JM, Ramaesh K, Kaufman MH, West JD, Dhillon B. Corneal abnormalities in *Pax6*^{+/-} small eye mice mimic human aniridia-related keratopathy. *Invest Ophthalmol Vis Sci*. 2003;44(5):1871–8.
- Ramaesh T, Ramaesh K, Collinson JM, Chanas SA, Dhillon B, West JD. Developmental and cellular factors underlying corneal epithelial dysgenesis in the *Pax6*^{+/-} mouse model of aniridia. *Exp Eye Res*. 2005;81(2):224–35.
- Liu JJ, Kao WWY, Wilson SE. Corneal epithelium-specific mouse keratin K12 promoter. *Exp Eye Res*. 1999;68(3):295–301.
- Collinson JM, Chanas SA, Hill RE, West JD. Corneal development, limbal stem cell function, and corneal epithelial cell migration in the *Pax6*^{+/-} mouse. *Invest Ophthalmol Vis Sci*. 2004;45(4):1101–8.
- Douvaras P, Mort RL, Edwards D, Ramaesh K, Dhillon B, Morley SD, Hill RE, West JD. Increased corneal epithelial turnover contributes to abnormal homeostasis in the *Pax6*^{+/-} mouse model of aniridia. *PLoS ONE*. 2013;8(8):e71117.
- Ahmad S. Concise review: limbal stem cell deficiency, dysfunction, and distress. *Stem Cells Transl Med*. 2012;1(2):110–5.
- Hayashi S, McMahon AP. Efficient recombination in diverse tissues by a tamoxifen-inducible form of Cre: a tool for temporally regulated gene activation/inactivation in the mouse. *Dev Biol*. 2002;244(2):305–18.
- Simpson TI, Pratt T, Mason JO, Price DJ. Normal ventral telencephalic expression of *Pax6* is required for normal development of thalamocortical axons in embryonic mice. *Neural Dev*. 2009;4:19.
- Yoshida S, Shimmura S, Kawakita T, Miyashita H, Den S, Shimazaki J, Tsubota K. Cytokeratin 15 can be used to identify the limbal phenotype in normal and diseased ocular surfaces. *Invest Ophthalmol Vis Sci*. 2006;47(11):4780–6.
- Means AL, Xu YW, Zhao AZ, Ray KC, Gu GQ. A CK19^{CreERT} knockin mouse line allows for conditional DNA recombination in epithelial cells in multiple endodermal organs. *Genesis*. 2008;46(6):318–23.
- Miyoshi G, Hjerling-Leffler J, Karayannis T, Sousa VH, Butt SJB, Battiste J, Johnson JE, Machold RP, Fishell G. Genetic fate mapping reveals that the caudal ganglionic eminence produces a large and diverse population of superficial cortical interneurons. *J Neurosci*. 2010;30(5):1582–94.
- Hart AW, Mella S, Mendrychowski J, van Heyningen V, Kleinjan DA. The developmental regulator Pax6 is essential for maintenance of islet cell function in the adult mouse pancreas. *PLoS ONE*. 2013;8(11):e54173.
- Quintana-Urzainqui I, Kozić Z, Mitra S, Tian T, Manuel M, Mason JO, Price DJ. Tissue-specific actions of Pax6 on proliferation-differentiation balance in the developing forebrain are Foxg1-dependent. *bioRxiv*. 2018. <https://doi.org/10.1101/374074>.
- Dorà NJ, Collinson JM, Hill RE, West JD. Hemizygous *Le-Cre* transgenic mice have severe eye abnormalities on some genetic backgrounds in the absence of *loxP* sites. *PLoS ONE*. 2014;9(10):e109193.
- Schmidt-Suppran M, Rajewsky K. Vagaries of conditional gene targeting. *Nat Immunol*. 2007;8(7):665–8.
- Harno E, Cottrell EC, White A. Metabolic pitfalls of CNS *Cre*-based technology. *Cell Metab*. 2013;18(1):21–8.
- Dora NJ, Crookshanks AJF, Leung KKY, Simpson TI, Mason JO, Price DJ, West JD. Analysis of compound heterozygotes reveals that the mouse floxed *Pax6*^{tm1Ued} allele produces abnormal eye phenotypes. *Transgenic Res*. 2016;25(5):679–92.
- Kristianto J, Johnson MG, Zastrow RK, Radcliff AB, Blank RD. Spontaneous recombinase activity of Cre-ERT2 in vivo. *Transgenic Res*. 2017;26(3):411–7.
- Grindley JC, Davidson DR, Hill RE. The role of *Pax-6* in eye and nasal development. *Development*. 1995;121:1433–42.
- Di Girolamo N, Bobba S, Raviraj V, Delic NC, Slapetova I, Nicovich PR, Halliday GM, Wakefield D, Whan R, Lyons JG. Tracing the fate of limbal epithelial progenitor cells in the murine cornea. *Stem Cells*. 2015;33(1):157–69.
- Amitai-Lange A, Altshuler A, Bubley J, Dbayat N, Tiosano B, Shalom-Feuerstein R. Lineage tracing of stem and progenitor cells of the murine corneal epithelium. *Stem Cells*. 2015;33(1):230–9.
- Gregory-Evans CY, Wang X, Wasan KM, Zhao J, Metcalfe AL, Gregory-Evans K. Postnatal manipulation of Pax6 dosage reverses congenital tissue malformation defects. *J Clin Invest*. 2014;124(1):111–6.
- Chen YT, Chen FYT, Vijmasi T, Stephens DN, Gallup M, McNamara NA. Pax6 downregulation mediates abnormal lineage commitment of the ocular surface epithelium in aqueous-deficient dry eye disease. *PLoS ONE*. 2013;8(10):e77286.
- Quinn JC, West JD, Hill RE. Multiple functions for *Pax6* in mouse eye and nasal development. *Genes Dev*. 1996;10(4):435–46.
- Collinson JM, Hill RE, West JD. Different roles for *Pax6* in the optic vesicle and facial epithelium mediate early morphogenesis of the murine eye. *Development*. 2000;127(5):945–56.
- Collinson JM, Quinn JC, Hill RE, West JD. The roles of *Pax6* in the cornea, retina, and olfactory epithelium of the developing mouse embryo. *Dev Biol*. 2003;255(2):303–12.

33. Li S, Goldowitz D, Swanson DJ. The requirement of Pax6 for postnatal eye development: evidence from experimental mouse chimeras. *Invest Ophthalmol Vis Sci.* 2007;48:3292–300.
34. Dorà NJ, Hill RE, Collinson JM, West JD. Lineage tracing in the adult mouse corneal epithelium supports the limbal epithelial stem cell

hypothesis with intermittent periods of stem cell quiescence. *Stem Cell Res.* 2015;15(3):665–77.

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