

THE UNIVERSITY of EDINBURGH

Edinburgh Research Explorer

Thermochemical decomposition of coffee ground residues by TG-MS: a kinetic study

Citation for published version:

Fermoso, J & Masek, O 2017, 'Thermochemical decomposition of coffee ground residues by TG-MS: a kinetic study' Journal of Analytical and Applied Pyrolysis. DOI: 10.1016/j.jaap.2017.12.007

Digital Object Identifier (DOI):

10.1016/j.jaap.2017.12.007

Link:

Link to publication record in Edinburgh Research Explorer

Document Version: Peer reviewed version

Published In: Journal of Analytical and Applied Pyrolysis

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



Manuscript Details

Manuscript number	JAAP_2017_795_R1
Title	Thermochemical decomposition of coffee ground residues by TG-MS: a kinetic study
Article type	Research Paper

Abstract

Dynamic pyrolysis tests of coffee grounds residues (CGR) at heating rates in the range from 5 - 100 °C/min and at maximum temperature of 500 °C were carried out using a thermogravimetric analyser coupled to a mass spectrometer (TG-MS), for online evolved gas analysis, to determine kinetic parameters of thermochemical decomposition of CGR and its biopolymer constituents. During the pyrolysis, the maximum decomposition rate of each biomass component increased linearly with the heating rate. The slope increased with the biopolymer reactivity in the following sequence: hemicellulose > celluose > lignin. Main gases produced during the pyrolysis of CGR were oxygen containing species derived from parent biopolymers and primary and secondary vapours (250 - 425 °C), with H2O being the most important, followed by CO and CO2. The use of the Beta zeolite had only negligible effect on deoxygenation reactions, however it significantly promoted cracking reactions of pyrolysis vapours increasing the light hydrocarbons (C1-C2) formation with the subsequent improvement in the heating value of the pyrolysis gas. Kinetic parameters for any of the individual biopolymers in CGR were estimated using the model-free isoconversional dynamic methods: Kissinger-Akahira-Sunose (KAS) and Flynn-Wall-Ozawa (FWO) models. The average value for the apparent activation energy of the individual biopolymers (hemicellulose, cellulose and lignin) in CGR calculated by KAS and FWO methods: were estimated as 214, 241 and 266 kJ/mol, respectively; whilst for the CGR as a whole it was 242 kJ/mol. The two modelfree isoconversional dynamic methods have been shown to be useful tools for assessment of biomass pyrolysis kinetic parameters, as they can provide Ea values for use in reactor design models.

Keywords	coffee ground residues; TG-MS; catalytic pyrolysis; cracking; kinetics; isoconversional methods.
Corresponding Author	Javier Fermoso
Corresponding Author's Institution	IMDEA Energy
Order of Authors	Javier Fermoso, Ondrej Masek
Suggested reviewers	D Beneroso, Beatriz Fidalgo, Aimaro Sanna

Submission Files Included in this PDF

File Name [File Type]

Cover Letter - revised - 111217.docx [Cover Letter]

Respons to REVIEWERS - 111217.docx [Response to Reviewers]

Manuscript - revised - 111217.docx [Revised Manuscript with Changes Marked]

Highlights_revised - 111217.docx [Highlights]

Pyrolysis kinetics of coffee ground residues - final.docx [Manuscript File]

Figures - revised - 111217.docx [Figure]

Supporting information - 111217.docx [Figure]

Table 1.docx [Table]

Table 2.docx [Table]

To view all the submission files, including those not included in the PDF, click on the manuscript title on your EVISE Homepage, then click 'Download zip file'.

Dear Dr. Dufour

Enclosed you can find the electronic files of the revised version of our manuscript entitled "Thermochemical decomposition of coffee ground residues by TG-MS: a kinetic study", resubmitted to be considered for publication in *Journal of Analytical and Applied Pyrolysis*, which has been agreed by all authors.

As you may note we have carefully taken into account all the reviewer's comments making necessary changes where possible or providing suitable rebuttals where relevant.

Moreover, according to your recommendations, we have included more experimental results on MS analysis of the gaseous products (new Figs. 4 and 5) with the subsequent further improved discussion on the volatiles formation, thermally and catalytically. On the other hand, we have also moved some thermal modelling results to the new Supporting Information file (former Fig. 4 appears now as Fig. S1, and Table 3 appears now as Table S1).

Therefore, as you may note in the revised version of our manuscript it is devoted to study the thermochemical coffee ground residues decomposition by TG-MS. Main evolved gases online analysed by MS were oxygenates (H₂O, CO and CO₂), which overlapped with main CGR degradation regime (250 - 425 °C). Catalytic pyrolysis improved deoxygenation only negligibly but importantly enhanced vapours cracking increasing the light hydrocarbons (C₁-C₂) formation with the subsequent improvement in the heating value of the pyrolysis gas.

Kinetic parameters of the thermochemical decomposition of coffee ground residues (CGR) and those corresponding to the biopolymers comprising them were estimated by two model-free isoconversional dynamic methods: Kissinger-Akahira-Sunose (KAS) and Flynn-Wall-Ozawa (FWO) models. These, were satisfactorily employed resulting average apparent activation energy (E_a) values of 242 kJ/mol (for the entire CGR), and 214, 241 and 266 kJ/mol for its hemicellulose, cellulose and lignin components, respectively.

The use of model-free isoconversional dynamic methods proved to be valuable to assess the kinetic parameters of CGR as it is for other biomasses whatever their structural complexity. The E_a values for pyrolysis of CGR are important input parameters for modelling and design of reactors and their optimisation for production of biochar, fuels or high added-value chemicals.

Keywords: coffee ground residues; TG-MS; catalytic pyrolysis; cracking; kinetics; isoconversional methods.

Please, address your correspondence to the following author:

Dr. Javier Fermoso Domínguez Madrid Institute for Advanced Studies on Energy IMDEA Energy Institute Telephone: +34 917371120, FAX: +34 917371140 E-mail address: javier.fermoso@imdea.org

Looking forward to hearing from you,

Yours sincerely,

Dr. Javier Fermoso

Dear Dr Anthony Dufour,

We thank you and the reviewers for your helpful comments on our manuscript. We have carefully taken into account all of them making necessary changes where possible or providing suitable rebuttals where relevant. Below you can find our point-to-point response to the comments.

I hope that the revised manuscript can be accepted for publication.

Yours sincerely,

Dr. Javier Fermoso

Comments from the editors and reviewers:

* Reviewer 1

This manuscript deals with thermogravimetric analysis of coffee ground residues. The evolved gases were analysed by using mass spectrometry. The manuscript is well prepared, and the discussion and presentations are clear. However, this work lacks novelty or even interesting findings. Several similar articles can be found elsewhere. Therefore, the manuscript is not recommended to be published in the Journal of Analytical and Applied Pyrolysis.

Answer:

We thank Reviewer 1 for the time dedicated to reviewing our manuscript. However, we do not fully agree with the statement of lack of novelty or interesting findings. To start with, coffee is the second most traded commodity in the world after oil, and one of the most widely consumed beverages [1]; and therefore, we believe that the potential utilization of the generated residues for fuels or high value-added chemicals production by means of its pyrolysis can be an attractive solution for this residue, whose worldwide production is still increasing. Therefore, the first, but not last step for its utilization on the pyrolysis process would be its kinetics study to understand the complex reaction mechanism of its different biopolymers decomposition to help on the design of an adequate reactor.

Although this kind of kinetics study is not new, it does not mean that it is no longer necessary and interesting for the research community, which helps to understand the pyrolysis process itself, whose complex reaction mechanism is still under debate and remains unknown. The importance of such kinetic studies can be evidenced in the available literature, where the number of published studies on pyrolysis showed exponential growth in the past 15 years as shown in the following figure (number of publications from 2000 to present based on search results from SCOPUS on: "thermogravimetric analysis", "biomass pyrolysis" and "kinetics").



Number of annual publications from 2000 to present on: "thermogravimetric analysis", "biomass pyrolysis" and "kinetics" (based on search results from SCOPUS).

* Reviewer 2

This paper investigated the kinetic of coffee ground residues pyrolysis. But there are no any meanings about the 1-3 highlights, and distributed activation energy model (DAEM) is more suitable than Kissinger-Akahira-Sunose (KAS) and Flynn-Wall-Ozawa (FWO) models for coffee ground residues pyrolysis. The review could not identify the challenges of the present topic and position of this study. What's more, the review could not see sufficient discussions and useful conclusion.

Answer:

We thank Reviewer 2 for reviewing our manuscript.

Firstly, we consider the statement: "the distributed activation energy model (DAEM) is more suitable than those employed in our work, Kissinger–Akahira–Sunose (KAS) and Flynn–Wall– Ozawa (FWO) models" as a personal assessment from the reviewer. We believe that there are still a lot of controversy and contradictory information in the literature regarding which models fit better the pyrolysis kinetics of lignocellulosic biomass. There are several review works published during the last 5-6 years in which those methods and some others are described and compared without any resolute statement selecting one over the others [2–4]. As we have already discussed answering to the Reviewer 1, we think that the focus covered in this work is actually of great interest within the research community as demonstrate the previous figure that shows the annual number of publications on this topic.

On the other hand, more MS results and further discussion about the evolved gaseous species during the non-catalytic and catalytic pyrolysis of CGR have been included in the revised manuscript.

* Reviewer 3

A solid work on fundamental investigations concerning an interesting topic, the potential utilization of coffee ground residues for pyrolytic processes. I have only three points, which the authors should consider:

1) Figure 1B: The authors claim that there are "clearly three conversion peaks" in the DTG curve visible. However, the second and third "peak" are hardly visible as shoulders on the first larger peak. I also wonder why with lower heating rates the distinction between the

peaks is not getting better, i.e. at 5K/min it is even less pronounced than at 25 and 50 K/min, respectively.

Answer:

In accordance with the reviewer's statement, we have modified our claims in the revised version of the manuscript. So, where it said:"...clearly showing three conversion peaks corresponding to the well-established order of biopolymers decomposition: hemicellulose, cellulose and lignin [28–30]. As the figure shows, the three peaks can be clearly distinguished for all the heating rates, with the exception of 100 °C/min"; now it reads: "...showing a first large peak with two remarkable shoulders corresponding to the well-established order of biopolymers decomposition: hemicellulose, cellulose and lignin [28–30]. As the figure shows, the three peaks can be clearly distinguished for all the heating rates, with the exception of 100 °C/min"; now it reads: "...showing a first large peak with two remarkable shoulders corresponding to the well-established order of biopolymers decomposition: hemicellulose, cellulose and lignin [28–30]. As the figure shows, the peak with the subsequent shoulders can be clearly distinguished for all the heating rates, with the exception of 100 °C/min."

Regarding the second question, it is well documented that it is not always the case that the lower the heating rate the better the separation of biomass decomposition peaks. It depends on the biomass source and other experimental conditions, etc. This has been demonstrated in the literature, where different types of biomass were thermally decomposed at different heating rates [5–8]. In these papers it can be also observed that the best distinction of the decomposition peaks does not correspond to the lowest heating rate.

2) Figure 3: The temperature axis seems to be misplaced in this diagram, it should be on the x-axis parallel to the time axis, not to the MS-signal.

Answer:

We agree with this reviewer that the suggested way could also be used, however we believe that the way we used is advantageous as in this figure we want to show how the TGA (conversion and DTG curves) and MS (different gaseous evolved from the CGR decomposition results), but also the sample temperature progress with reaction time. As can be seen in that figure there is a non-isothermal step (during the heating up of the sample) but also a 30 min isothermal step which could not be properly plotted if temperature were on the X-axis parallel to the time axis.

3) It should be mentioned what type of ionization and mass analyser have been used for the MS analysis (I guess electron ionization and quadrupole). In this regard, have the authors considered to look deeper into the organic composition of the evolved gases by doing pyrolysis GC-MS for instance (or, if they have the opportunity, by coupling soft ionization MS or GC to the thermobalance)?

Answer:

In accordance with the reviewer's suggestion, we have included in the revised version of the manuscript the type of ionization source and mass analyzer that was used for the MS analysis. We thank the reviewer for the suggestions regarding further in-depth study of the pyrolysis gas and vapors and we will take these into consideration in future works focused on the bio-oil production from coffee ground residues.

* Reviewer 4

1. The apparent activation energy for the whole process was estimated as 244 and 241 kJ/mol for KAS and FWO methods, respectively. How this value was calculated from the activation energies of three components?

Answer:

These values of the apparent activation energy for the whole pyrolysis process for KAS and FWO methods were calculated as the average value of the activation energies obtained at conversion levels from 5 to 90 % as summarizes Table 3, in a similar way to other works found in the literature [9,10].

2. In "2.2. Analytical techniques," "asses" \rightarrow "assess"

Answer:

We have replaced "asses" by "assess" in the 2.2 Analytical techniques section according to the Reviewer comment.

References

- F. Girotto, A. Pivato, R. Cossu, G.E. Nkeng, M.C. Lavagnolo, The broad spectrum of possibilities for spent coffee grounds valorisation, J. Mater. Cycles Waste Manag. (2017) 1–7. doi:10.1007/s10163-017-0621-5.
- J.E. White, W.J. Catallo, B.L. Legendre, Biomass pyrolysis kinetics: A comparative critical review with relevant agricultural residue case studies, J. Anal. Appl. Pyrolysis. 91 (2011) 1–33. doi:10.1016/j.jaap.2011.01.004.
- [3] A. Sharma, V. Pareek, D. Zhang, Biomass pyrolysis—A review of modelling, process parameters and catalytic studies, Renew. Sustain. Energy Rev. 50 (2015) 1081–1096. doi:10.1016/j.rser.2015.04.193.
- [4] S. Wang, G. Dai, H. Yang, Z. Luo, Lignocellulosic biomass pyrolysis mechanism: A stateof-the-art review, Prog. Energy Combust. Sci. 62 (2017) 33–86. doi:10.1016/j.pecs.2017.05.004.
- [5] R. Soysa, S.K. Choi, Y.W. Jeong, S.J. Kim, Y.S. Choi, Pyrolysis of Douglas fir and coffee ground and product biocrude-oil characteristics, J. Anal. Appl. Pyrolysis. 115 (2015) 51–56. doi:10.1016/j.jaap.2015.07.002.
- P. Luangkiattikhun, C. Tangsathitkulchai, M. Tangsathitkulchai, Non-isothermal thermogravimetric analysis of oil-palm solid wastes, Bioresour. Technol. 99 (2008) 986–997. doi:10.1016/j.biortech.2007.03.001.
- [7] S. Wang, X.M. Jiang, N. Wang, L.J. Yu, Z. Li, P.M. He, Research on Pyrolysis Characteristics of Seaweed, Energy & Fuels. 21 (2007) 3723–3729. doi:10.1021/ef700214w.
- [8] G. Özsin, A.E. Pütün, Kinetics and evolved gas analysis for pyrolysis of food processing wastes using TGA/MS/FT-IR, Waste Manag. 64 (2017) 315–326. doi:10.1016/j.wasman.2017.03.020.
- [9] K. Slopiecka, P. Bartocci, F. Fantozzi, Thermogravimetric analysis and kinetic study of

poplar wood pyrolysis, Appl. Energy. 97 (2012) 491–497. doi:10.1016/j.apenergy.2011.12.056.

 Y. Liang, B. Cheng, Y. Si, D. Cao, H. Jiang, G. Han, X. Liu, Thermal decomposition kinetics and characteristics of Spartina alterniflora via thermogravimetric analysis, Renew. Energy. 68 (2014) 111–117. doi:10.1016/j.renene.2014.01.041.

Title:

Thermochemical decomposition of coffee ground residues by TG-MS: a kinetic study

Author names and affiliations

Javier Fermoso^{a,b,*}, Ondřej Mašek^b

^a Thermochemical Processes Unit, IMDEA Energy Institute, 28935, Móstoles, Madrid, Spain.

^b UK Biochar Research Centre, School of Geosciences, University of Edinburgh, Crew Building, Alexander Crum Brown Road, Edinburgh EH9 3FF, UK

Corresponding author*:

Tel.: +34 917371120. *E-mail address:* javier.fermoso@imdea.org (Javier Fermoso). Thermochemical Processes Unit, IMDEA Energy Institute, 28935, Móstoles, Madrid, Spain. Thermochemical decomposition of coffee ground residues by TG-MS: a kinetic study

Javier Fermoso^{a,b,*}, Ondřej Mašek^b

^a Thermochemical Processes Unit, IMDEA Energy Institute, 28935, Móstoles, Madrid, Spain.

^b UK Biochar Research Centre, School of Geosciences, University of Edinburgh, Crew Building, Alexander Crum Brown Road, Edinburgh EH9 3FF, UK

Abstract

Dynamic pyrolysis tests of coffee grounds residues (CGR) at heating rates in the range from 5 – 100 °C/min and at maximum temperature of 500 °C were carried out using a thermogravimetric analyser coupled to a mass spectrometer (TG-MS), for online evolved gas analysis, to determine kinetic parameters of thermochemical decomposition of CGR and its biopolymer constituents. During the pyrolysis, the maximum decomposition rate of each biomass component increased linearly with the heating rate. The slope increased with the biopolymer reactivity in the following sequence: hemicellulose > celluose > lignin.

Main gases produced during the pyrolysis of CGR were oxygen containing species derived from parent biopolymers and primary and secondary vapours (250 - 425 °C), primarily H₂O, followed by CO and CO₂. The use of the Beta zeolite had only negligible effect on deoxygenation reactions, however it significantly promoted cracking reactions of pyrolysis vapours increasing the light hydrocarbons (C₁-C₂) formation with the subsequent improvement in the heating value of the pyrolysis gas.

Kinetic parameters for any of the individual biopolymers in CGR were estimated using the model-free isoconversional dynamic methods: Kissinger–Akahira–Sunose (KAS) and Flynn–Wall–Ozawa (FWO) models. The average value for the apparent activation energy of the individual biopolymers (hemicellulose, cellulose and lignin) in CGR calculated by KAS and FWO methods: were estimated as 214, 241 and 266 kJ/mol, respectively; whilst for the CGR as a whole it was 242 kJ/mol. The two model-free isoconversional dynamic methods have been shown to be useful tools for assessment of biomass pyrolysis kinetic parameters, as they can provide E_a values for use in reactor design models.

Keywords: coffee ground residues; **TG-MS**; catalytic pyrolysis; cracking; kinetics; isoconversional methods.

1. Introduction

The depletion of fossil fuel reserves and the environmental problems derived from their utilization have made necessary the use of alternative fuels. Biomass is a clean and renewable energy source leading to environmental, technical and economic benefits. Residues from agricultural production and processing industries are readily available in large quantities. Coffee is the second most traded commodity in the world after oil, and one of the most widely consumed beverages in the world [1]. Moreover, the residues derived from its production are steadily increasing in proportion to the coffee consumption growth [1–3]. Coffee silverskin and spent coffee grounds are the main coffee industry residues [4]. The latter is a residue with fine particle size, high humidity (≈ 80 wt%), organic load and acidity, obtained during the treatment of raw coffee powder with hot water or steam for the instant coffee preparation. Therefore, this residue is generated in large amounts, with a worldwide annual generation of 8 million tons [3]. On an average one ton of green coffee ground are generated per kg of soluble coffee produced [5]. Coffee grounds are disposed as household waste that may be incinerated and/or

moved to landfill [4]. Some alternative applications for coffee ground residues are their utilisation to produce compost [6], deodorizer or adsorbents [7,8], as well as source of renewable energy or for the synthesis of high value-added chemicals [9,10]. In this latter regard, the pyrolysis is a thermochemical conversion process that, depending on the reaction conditions, can be used to transform biomass directly into solid, gaseous or liquid biofuels, being the latter also a promising feedstock for chemicals synthesis [11]. In this respect, a systematic understanding of pyrolysis kinetics is a key factor for the assessment of feasibility, design, and scale-up of such biomass conversion processes for energy applications [12]. Therefore, we believe that the potential utilization of the generated residues for fuels or high value-added chemicals production by means of its pyrolysis results at least an attractive and challenging solution for this residue, whose worldwide production is still increasing. Therefore, the first, but not last step for its utilization on the pyrolysis process would be its kinetics study to understand the complex reaction mechanism of its different biopolymers decomposition to help on the design of an adequate reactor. Therefore, thermogravimetric analysis (TGA) is the most commonly used technique to study the solid-phase thermal decomposition reactions [13]. Although it typically operates in a different form in comparison to a real reactor (pyrolyzer, gasifier or combustor), it provides an understanding of thermal degradation processes occurring during the fuel conversion. In addition, as this study is focused on slow pyrolysis conversion of coffee grounds to solid and gaseous products, the conditions in the TGA represented real reactor conditions much more realistically that would be the case for processes deploying high heating rates, such as fast pyrolysis and gasification.

The thermal decomposition of biomass proceeds via a very complex set of competitive and concurrent reactions and thus, the exact mechanism for biomass pyrolysis remains unknown. Each step likely has its own single apparent activation energy, and thus the use of an average, global apparent activation energy to define the kinetics of such processes could be interpreted as an inadequate simplification at best [14]. Furthermore, the DTG curves from these models may hide the true multi-stage character of pyrolytic reactions under a single peak [15].

During the second half of the 20th century several novel methods for determining Arrhenius parameters based on a single parameter emerged. These so-called "model-free" methods are founded on an isoconversional basis, wherein the degree of conversion, X, for a reaction was assumed to be constant and therefore the reaction rate, k, depended exclusively on the reaction temperature, T. By allowing apparent activation energy (E_a) to be calculated a priori, these approaches eliminate the need to initially hypothesize a form and rate order for the kinetic equation. Hence, isoconversional methods do not require previous knowledge of the reaction mechanism for biomass thermal degradation. Another advantage of such approaches is that the systematic error resulting from the kinetic analysis during the estimation of the Arrhenius parameters is eliminated [16]. Isoconversional models can follow either a differential or an integral approach to the treatment of TGA data. So, they are considered as a helpful solution for truly determining apparent activation energy [13].

During pyrolysis, due to the poor thermal conductivity of biomass, a temperature gradient is normally developed through the biomass sample between the external surface and the internal part. This gradient may be assumed to be proportional to the particle size, and decreases with reducing heating rate to the point that both, the external surface and the internal part of the biomass particles attains same temperature at a certain reaction time when appropriate time is given for heating [17]. Therefore, a defined particle size range was seleced in this study to avoid the effect of the particle size on the determination of kinetic parameters under dynamic conditions of different heating rates. Therefore, the main objectives of this study were, on the one hand to determine the kinetic parameters for the slow pyrolysis of such promising lignocellulosic residue, coffee ground residues, and its individual biopolymer components (cellulose, hemicellulose and lignin) employing isoconversional model-free dynamic methods: Kissinger-Akahira-Sunose (KAS) and Flynn-Wall-Ozawa (FWO), due to the advantages they offer in determining Arrhenius equation parameters without the need to make choices regarding kinetic models to be used [18]. The peak decomposition rates of biopolymers constituting the CGR and associated temperatures during pyrolysis process were evaluated. On the other hand, the non-condensable gases generated during the non-catalytic and catalytic tests were also monitored with reaction time and temperature by MS.

The obtained data and models provide important basis for design and operation of slow pyrolysis or estaged pyrolysis systems using coffee ground residues for production of biochar and biofuels or high value-added chemicals.

2. Materials and methods

2.1. Materials

The biomass employed in this study was coffee ground residue (CGR), collected from local canteen at the University, ensuring that one type of beans was used. As the pre-dried CGR still contained ≈ 15 wt% of moisture, the material was air dried for 24 hours at 105 °C in a laboratory oven before further use. To avoid the effect of the particle size on the kinetic parameters determination, the CGR was crushed and sieved to collect particles in the range of $250 - 500 \mu m$.

For the catalytic pyrolysis test, a Beta zeolite (Si/Al =150) in pellet form supplied by CLARIANT was employed. Prior to the catalytic pyrolysis test, the zeolite was crushed and sieved at same particle size as CGR sample.

2.2. Analytical techniques

The proximate analysis was determined according to European standards: moisture content (UNE-EN 14774-1:2010), ash content (UNE-EN 14775:2010), volatile matter (UNE-EN 15148:2010) and fixed carbon (determined by difference). A thermogravimetric analizer, TGA (Mettler-Toledo TGA/DSC1) equipped with automatic sample handling was employed to assess the volatile matter and the ash contents of the coffee ground residue. This is a well established thermoanalytical technique for thermal degradation studies of solid materials, such as biomass pyrolysis [13].

A quadrupole mass spectrometer, MS (HIDEN Analytical HPR-20) with an elecron ionisation source (70 eV) coupled to the TGA was employed for evolved gas analysis during the pyrolysis experiments. The ultimate analysis of feedstock and was carried out in a micro-elemental analyzer (Thermo Scientific) in order to determine content of C, H, N, S and O (by difference). The higher heating value (HHV) of CGR was calculated using the formula developed by Channiwala and Parikh [19]. The relative abundance of individual biopolymers (cellulose, hemicellulose and lignin) in CGR was determined by the sulfuric acid hydrolysis method [20].

2.3. Pyrolysis tests

The pyrolysis tests were performed in the same TGA previously mentioned, at atmospheric pressure. The CGR sample (around 15 mg) was deposited in an alumina crucible with a circular base and total volume of 150 μ l. In this work, all the experiments were performed under non-isothermal conditions at 500 °C and different heating rates (HR), 5–100 °C/min, with a nitrogen flow rate of 100 ml/min.

A zeolite to CGR ratio of 1:1 (g/g) was selected for the catalytic experiment, in which the Beta zeolite layer was deposited over the CGR sample in the crucible.

2.4. *Kinetic models*

The one-step global model assumes that the devolatilization phenomena proceeds as a single reaction.

$$Biomass \xrightarrow{k} Volatiles + Char \tag{1}$$

where *Volatiles* represents the sum of the gas and bio-oil, and *char* is the remaining unreacted solid. The fundamental rate of transformation from solid-state to volatiles is generally described by the following expression:

$$\frac{dX}{dt} = k(T)f(X) \tag{2}$$

where *X* is the degree of conversion of the fuel, *t* is the reaction time, k(T) is the reaction rate constant, and f(X) is a function that represents the reaction model.

The degree of conversion, X, is calculated as its relative weight loss as follows:

$$X = \frac{m_0 - m_t}{m_0 - m_f}$$
(3)

where m_0 , m_t and m_f represents the initial mass, the mass at time *t*, and the final residual mass of the sample, respectively.

The reaction rate constant, k, is temperature dependent, and it obeys the fundamental Arrhenius rate expression:

$$k = A \cdot e^{\frac{-E_a}{RT}} \tag{4}$$

where A is the pre-exponential factor (min⁻¹), E_a is the apparent activation energy (kJ/mol), T is the absolute temperature (K) and R is the gas constant (8.314 J/mol·K). Non-isothermal method employs a heating rate (β), normally linear, to raise the temperature. A linear heating program follows:

$$T = T_0 + \beta t \tag{5}$$

$$\beta = \frac{dT}{dt} \tag{6}$$

where T_0 is the starting temperature, β the constant heating rate (K/min), and T the temperature at time t. Then, substituting equations (4) and (6) in equation (2) gives:

$$\frac{dX}{dT} = \frac{A}{\beta} \cdot e^{\frac{-E_a}{RT}} f(X)$$
(7)

Equation (7) represents the differential form of the non-isothermal rate law.

Kinetics analysis is conventionally expected to produce a suitable kinetic description of the process in terms of the reaction model and the Arrhenius parameters. These three components, f(X), E_a , and A, are sometimes called the "kinetic triplet". There are many methods for analysing solid-state kinetic data [21]. These methods can be classified according to the experimental conditions and the mathematical analysis implemented. The mathematical approaches employed can be divided into model-fitting and isoconversional (model-free) methods. However, as discussed in the introduction section, in this work only the isoconversional model-free dynamic methods were used to calculate the kinetic parameters for the CGR pyrolysis, which require a set of experimental tests at different heating rates. These methods are the Kissinger, Kissinger-Akahira-Sunose (KAS) and Flynn-Wall-Ozawa (FWO) methods. The advantages of the model-free analysis are: its simplicity, and the avoidance of errors associated to choices of a kinetic model [18].

2.5. Model-free methods

Kissinger method

This method allows for the kinetic parameters of a solid-state reaction without prior knowledge of the reaction mechanism. Kissinger [22] developed a model-free non-isothermal method where E_a does not need to be calculated for each conversion value in order to evaluate kinetic parameters. The method equation is represented as follows:

$$Ln\left(\frac{\beta}{T_m^2}\right) = Ln\left(\frac{A \cdot R}{E_a}\right) - \frac{E_a}{RT_m}$$
(8)

According to Kissinger, in the differential thermogravimetric curve (DTG), the temperature at which the peak weight loss velocity (in %/min) occurs for a given heating rate is determined by both A and E_a . Then, changing the heating rate the peak temperature will change. Hence, plotting $Ln(\beta/T_m^2)$ versus $1/T_m$, should give a straight line of slope $-E_a/R$.

Flynn-Wall-Ozawa method (FWO)

The *FWO* method [23,24] is one of the most commonly accepted methods for the computation of kinetic parameters. It allows for the apparent activation energy to be obtained for each degree of conversion from the equation:

$$Ln(\beta) = Ln\left(\frac{A_X \cdot E_{a_X}}{R \cdot g(X)}\right) - 5.331 - 1.052 \frac{E_{a_X}}{R \cdot T_X}$$
(9)

where E_{aX} is the apparent activation energy for a fix degree of conversion X, and is calculated from the slope of the straight line obtained by plotting logarithm of heating rates, Ln β , versus $1/T_X$, where T_X is the reaction temperature at which this grade of conversion X is reached.

Kissinger-Akahira-Sunose method (KAS)

The *KAS* method [22,25] is an integral isoconversional technique based on the following expression:

$$Ln\left(\frac{\beta}{T_{X}^{2}}\right) = Ln\left(\frac{A_{X}\cdot R}{E_{a_{X}}\cdot g(X)}\right) - \frac{E_{a_{X}}}{R\cdot T_{X}}$$
(10)

where E_{aX} is the apparent activation energy for a fix degree of conversion X, and is calculated from the slope of the straight line obtained by plotting $\text{Ln}(\beta/T_X^2)$, versus $1/T_X$, where T_X is the reaction temperature at which this grade of conversion X is reached.

3. Results and discussion

3.1. Biomass analysis

Table 1 summarizes the proximate and ultimate analysis, as well as composition in terms of key biopolymers (hemicellulose, cellulose and lignin) of CGR. The pre-dried sample still showed 5 wt% of moisture because this material rapidly adsorbs moisture during transfer and storage. The table also shows that CGR contains more carbon and less oxygen (and therefore lower O/C ratio ≈ 0.66) than woody biomass and agricultural residues (O/C ≈ 0.8 -1.2) [26]. Such behaviour is attributed to the higher lignin contents in CGR, which reaches 40.6 wt% (whose O/C ratio $\approx 0.4 - 0.5$), while hemicellulose and cellulose account for 36.6 and 10.6 wt%, respectively. This composition is rather similar to that reviewed by Obruca et al. [27] in terms of holocellulose biopolymers but with larger content of lignin. Consequently, the high heating value (HHV) of CGR, 23.4 MJ/kg, is higher than that of most biomass (17-20 MJ/kg) [26]. However, ultimate analysis also showed that CGR contains more nitrogen (2.3 wt%) than other more commonly used lignocellulosic biomass (0.1-1.0 wt%) due to high protein and caffeine content [4].

3.2. CGR thermochemical decomposition (TGA)

Fig. 1 shows both the conversion curves as weight loss in wt% (A), and their first derivative curves with time (DTG) as wt%/min (B) of the thermochemical decomposition of CGR as a function of reaction temperature under nitrogen atmosphere at 500 °C and at five heating rates (HR): 5 - 100 °C/min. The conversion curves at all heating rates indicate that mass loss of CGR mainly occurred at temperatures ranging from 250 to 500 °C. The conversion curves shift to the right with increasing heating rate as can be observed in Fig. 1(A), which implies higher values of initial decomposition temperature (see Table 2). However, this representation of the conversion data makes it difficult to identify the changes in the slope at different temperatures and reaction rates for the thermal decomposition of the three biopolymers contained in CGR. Therefore, the DTG curves, as shown in Fig. 1(B) were derived from the TG data, showing a first large peak with two

remarkable shoulders corresponding to the well-established order of biopolymers decomposition: hemicellulose, cellulose and lignin [28–30]. As the figure shows, the peak with the subsequent shoulders can be clearly distinguished for all the heating rates, with the exception of 100 °C/min. Although the identification of the maximum degradation rate of lignin in biomass is not commonly reported in the literature, as it usually overlaps with the cellulose decomposition peak [13,31,32], in the case of CGR it was possible, due to the large difference in cellulose and lignin contents, \approx 11 and 42 wt%, respectively. Based on this initial observation it might be suggested that a more efficient staged pyrolysis process of these materials could be carried out to preferentially obtaining products of decomposition of individual biomass constituents separately when heating the material to different temperatures in stages.

On the other hand, it is evident from the results that increasing the heating rate from 5 to 100 °C/min resulted in a progressive rise in the total volatile matter released at 500 °C from 72.2 to 75.1 wt%, respectively as summarized in Table 2. This behaviour agrees with the theory that heating rate has an influence on the secondary reactions of the primary pyrolysis vapours. So, lower heating rates result in longer residence times of volatiles inside biomass particles and the reactor, favouring secondary reactions such as cracking, re-polymerization and re-condensation, which eventually lead to the char formation [13,17,33–35]. This is normally observed when comparing slow pyrolysis with fast/flash pyrolysis according to the product goal for the pyrolysis process; i.e., biochar production (for slow pyrolysis: 0.1-2 °C/s) or bio-oil production for fast (10-200 °C/s) and flash pyrolysis (>1000 °C/s) [36]. However, it is interesting to see this effect even in the heating rate range still corresponding to slow (5-50 °C/min) or at best intermediate (100 °C/min) pyrolysis here studied because it could be used to modulate the reaction to the desire products.

Fig. 2(A) depicts the temperatures of maximum decomposition rate of CGR biopolymers during pyrolysis as a function of the heating rate. Here it can be observed that these peak temperatures logarithmically depend on the heating rate for the three biopolymers; which implies that at low heating rates the mass transfer limitations are more important than at high heating rates, at which the maximum decomposition rate for the different bipolymers occurs at similar temperatures.

On the other hand, Fig. 2(B) shows that the maximum decomposition rate increased linearily with heating rate for all biopolymers in CGR. In addition, the observed difference in the slope of these lines suggests that the heating rate affected diverse biopolymers differently; thus, the more reactive the material (hemicellulose > celluose > lignin) the higher the slope. As lignin is the most stable and complex of biopolymers comprising biomass, its amount is assumed to be the main rate limiting factor in the thermochemical decomposition process of CGR.

Fig. 3 shows the TG-MS spectrum of the evolved gas species during the pyrolysis of CGR at 500 °C and 15 °C/min heating rate versus reaction time. This technique is the only one to simultaneously measure in real time the thermal decomposition and the gas product distribution of a very small sample. DTG curve and temperature profile are also plotted to show which gaseous compounds were evolved at each stage of the pyrolysis and at which temperatures. Water is the principal component, and it has two origins. Firstly, the physically adsorbed water, which is desorbed at \approx 90 °C; and secondly reaction water, produced at 250 – 400 °C, with a maximum production at 314 °C, originated from various dehydration reactions of the original CGR biopolymers and/or dehydration of the primary and secondary pyrolysis vapours from the removal of hydroxyl groups (–OH) overlapping with the main CGR degradation regime [37]. Fig. 4 displays the gases evolution trends at all the heating rates versus the reaction temperature. In this figure are also two water

peaks regardless the heating rate, 5 - 50 °C/min; though both peaks merged as a consequence of the fast heating at high heating rate. The other main gases, whose evolution also coincides with CGR decomposition rate profile are CO and CO₂. On the one hand, CO is mostly produced from the removal of carbonyl groups from biopolymers. While CO₂ is origninated from decarboxylation of –COOH and O–Acetyl groups from the original biopolymers, principally the hemicellulose. But also these oxygenates are originated from primary and secondary vapours; generated in the same range of temperatures, with maximum production shifted to a slightly higher temperature 330 °C. These three oxygen containing gases seem to originate from the thermal degradation of the three biopolymers that constitute the CGR (hemicellulose, cellulose and lignin), which is visible on the DTG curve. Moreover, this is also demonstrated by the two small shoulders to the right from the main peak of these evolved gases.

The removal of methoxyl groups ($-O-CH_3$) from the lignocellulosic structure is associated with the CH₄ production during pyrolysis. CH₄ is released from the biomass structure as a consequence of multi-step reactions, so the removal of methoxyl substituents of the hemicellulose and lignin and the conversion of the alkyl chain of the lignin are attributed to the CH₄ evolution during biomass pyrolysis [38–41]. The release of (C₁-C₂) and H₂ is mainly due to the instability of intermediate condensable species produced during primary degradation. Thus, as temperature increased, the C–C and C–H bonds break to form free radicals, which are recombined into small molecular compounds like light hydrocarbons C₂H₄ and C₂H₆, while H₂ was mainly produced from the breaking of C–H bonds. These species are evolved to a much lower extent, which might be related to the aromatization or the char structure during the secondary pyrolysis [13]. Fig. 4 also shows that the first appearance of the different gaseous species are shifted to lower reaction temperatures as the heating rate was raised. Fig. 5 displays de evolution with reaction time of the main evolved gases during a very preliminary catalytic pyrolysis test performed at 500 °C and 15 °C/min by placing a Beta zeolite layer over the CGR sample in the TGA crucible for comparison with its noncatalytic performance. In this figure can be appreciated that the progression of the oxygenated species, H₂O, CO and CO₂, during the catalytic test matched relatively well with that obtained without catalyst at temperatures below 400 °C, from which their production started to be slightly higher, especially CO. This means that the Beta zeolite employed herein shows certain deoxygenation activity for the pyrolysis primary vapors, but not much. However, is in the light hydrocarbons development where most significant differences are observed, reflecting a severe cracking activity of this zeolite over the pyrolysis primary and secondary vapours with temperature peaks around 475 – 500 °C [42]. On the one hand, the production of CH_4 and H_2 deviated from the non-catalytic behaviour above 405 $^{\circ}$ C, whilst the production of the C₂ hydrocarbons significantly increased from 330 to 500 °C, with peak productions of C₂H₄ and C₂H₆ at 470 amd 485 °C, respectively. The utilisation of this type of catalyst would significantly increase the heating value of the gas fraction due to its higher hydrocarbons content.

3.3. Kinetic analysis

The TGA experimental data were analyzed in order to obtain the kinetic parameters using three model-free methods. To avoid any influence of the physically bound moisture desorption from CGR sample, the conversion (in wt%) was calculated from the experimental data collected at temperatures between 150 and 500 °C, corresponding to the active pyrolysis stage where hemicellulose, cellulose and lignin decomposition occurs. Figs. S1(A – C) show the plots corresponding to the Kissinger, FWO and KAS models, respectively, used to calculate the kinetic parameters of the dynamic degradation of the biopolymers in CGR according to equations described in Section 2.3. Data in Fig.

S1(A) were used to calculate the A and E_a according to the Kissinger method. This method has the disadvantage that these parameters are calculated just from the temperature that corresponds to the highest weight loss rate (DTG_{max}). So, it means that this method should be employed simply for those samples showing a single DTG peak. In the case of CGR, these parameters would purely correspond to the hemicellulose degradation, which is the most reactive biopolymer in this biomass, in terms of degradation rate as shown in Fig. 2, disregarding the role of the other two biopolymers. However, the FWO and KAS methods calculate the kinetic parameters based on values of conversion from 5 to 90 wt%, with a 5 wt% step as shown in Figs. S1(B - C), which implies that the kinetic parameters for the individual biopolymers that comprise CGR may be estimated. These lines from linear fit at different conversion levels have fairly high linear correlation coefficients, > 0.995 (as summarized in Table S1), suggesting that the values of E_a and A satisfy accuracy requirements. This can be observed in Fig. 6, where the calculated apparent E_a is plotted as a function of the conversion level for FWO and KAS methods in comparison with that constant value obtained with Kissinger method. Here, it can be seen that E_a increases with temperature and conversion level. This is characteristic of processes with different reaction mechanisms. Even when each of the biopolymers that constitutes the CGR (hemicellulose, cellulose and lignin) has its own apparent activation energy, its thermal decomposition proceeds via a very complex set of competitive and concurrent reactions, due to the synergistic effects between its biopolymers. However, an approximate estimation of the E_a of these components can be assessed from the average values of different steps shown in Fig. 6. Thus, hemicellulose, cellulose and lignin would present values of 213, 240 and 265 kJ/mol, respectively according to FWO method; while 215, 242 and 268 kJ/mol, respectively for the KAS method as is summarized in Table S1.

The results show that the values estimated for the decomposition of hemicellulose are very close to those obtained for CGR using the Kissinger method (212 kJ/mol), which is in concordance as this method uses the maximum decomposition rate to calculate the kinetic parameters. When the conversion increases further than 80 wt%, the apparent activation energy increased sharply from 273 kJ/mol to 347 kJ/mol for FWO, and from 276 kJ/mol to 353 kJ/mol for KAS method as shown in Table S1, which could be due to the re-polymerization and re-condensation reactions leading to char formation. Then, taking into account all the steps in the pyrolysis process of the coffee ground residues, the apparent activation energy for the whole process was estimated as 244 and 241 kJ/mol for KAS and FWO methods, respectively. These E_a results are in correspondence with thermostability sequence analysis of these three CGR biopolymer components (hemicellulose, cellulose and lignin) [43]. Therefore, the lignin content would be the main controlling factor in biomass pyrolysis in industrial processes [44]. In this way, these kind of model-free isoconversional dynamic methods have been shown as a very useful tools to assess the kinetic parameters of CGR, as they can provide E_a values to be applied in models for designing reactors for its utilisation as fuel. Furthermore, due to the ability to obtain kinetic parameters for decomposition of individual biolopymenrs, the data could be used for designing a more efficient staged pyrolysis process allowing preferential recovery of decomposition products of individual biopolymers separately. Such process would enable more efficient production of high-value chemicals from biomass.

Conclusions

In this work, the utilisation of coffee ground residues, a lignocellulosic residue whose worldwide production is continuously increasing, has been explored for its pyrolysis application through the calculation of its kinetics parameters and those for its biopolymers constituents by thermogravimetric analysis. For that purpose, a thermogravimetric analyser coupled to a mass spectrometer (TG-MS) for the online detection of the evolved gases were employed to perform the pyrolysis tests at different heating rates (5 – 100 $^{\circ}$ C/min) and at maximum temperature of 500 $^{\circ}$ C. The results show that the heating rate significantly affected the thermal decomposition of coffee ground residues during pyrolysis. The maximum decomposition rate increased linearly with the heating rate; but also, the more reactive the material (hemicellulose > celluose > lignin) the higher the slope.

Main gases produced during the pyrolysis of CGR were oxygen containing species, and were evolved between 250 and 425 °C, with H₂O being the most important (from the removal of hydroxyl groups –OH), followed by CO (from decarbonylation reactions) and CO₂ (from decarboxylation reactions) of parent biopolymers and primary and secondary vapours. The use of the Beta zeolite had only negligible effect on deoxygenation reactions, however it significantly promoted cracking reactions of pyrolysis primary and secondary vapours giving rise to a significant increase of light hydrocarbons formation (C_1-C_2) with the subsequent improvement in the heating value of the pyrolysis gas. Kinetic parameters of the pyrolysis process were determined using isoconversional methods. While with the Kissinger method a single value for the apparent activation energy was obtained (212 kJ/mol), which correspond to the hemicellulose decomposition (as the most reactive component in this biomass), KAS and FWO methods showed that the E_a increases with the conversion level, revealing a complex set of competitive and concurrent reactions. The average value for the E_a of the hemicellulose, cellulose and lignin from these two methods were equal to 214, 241 and 266 kJ/mol, respectively. Whereas the apparent activation energy for the whole pyrolysis process would increase up to 242 kJ/mol. The use of model-free isoconversional dynamic methods proved to be valuable to assess the kinetic parameters of CGR. The E_a values for pyrolysis of CGR are

important input parameters for modelling and design of reactors and their optimisation for production of biochar, fuels or high added-value chemicals.

Acknowledgement

The authors gratefully acknowledge the experimental support provided by the UK Biochar Reseach Centre (University of Edinburgh). Dr Javier Fermoso also acknowledges the IMDEA Energy Institute for the financial support to accomplish the research stay at the UK Biochar Research Centre (University of Edinburgh).

References

- [1] F. Girotto, A. Pivato, R. Cossu, G.E. Nkeng, M.C. Lavagnolo, The broad spectrum of possibilities for spent coffee grounds valorisation, J. Mater. Cycles Waste Manag. (2017) 1–7. doi:10.1007/s10163-017-0621-5.
- [2] S.I. Mussatto, E.M.S. Machado, S. Martins, J.A. Teixeira, Production, Composition, and Application of Coffee and Its Industrial Residues, Food Bioprocess Technol. 4 (2011) 661–672.
- [3] R. Campos-Vega, G. Loarca-Piña, H.A. Vergara-Castañeda, B.D. Oomah, Spent coffee grounds: A review on current research and future prospects, Trends Food Sci. Technol. 45 (2015) 24–36.
- [4] S.I. Mussatto, E.M.S. Machado, S. Martins, J.A. Teixeira, Production, Composition, and Application of Coffee and Its Industrial Residues, Food Bioprocess Technol. 4 (2011) 661–672.
- [5] P.S. Murthy, M. Madhava Naidu, Sustainable management of coffee industry byproducts and value addition - A review, Resour. Conserv. Recycl. 66 (2012) 45– 58.
- [6] J.P. Bok, H.S. Choi, Y.S. Choi, H.C. Park, S.J. Kim, Fast pyrolysis of coffee grounds: Characteristics of product yields and biocrude oil quality, Energy. 47 (2012) 17–24.
- [7] T. Tokimoto, N. Kawasaki, T. Nakamura, J. Akutagawa, S. Tanada, Removal of lead ions in drinking water by coffee grounds as vegetable biomass, J. Colloid Interface Sci. 281 (2005) 56–61.
- [8] S. Rovani, M.T. Censi, S.L. Pedrotti, E.C. Lima, R. Cataluña, A.N. Fernandes, Development of a new adsorbent from agro-industrial waste and its potential use in endocrine disruptor compound removal, J. Hazard. Mater. 271 (2014) 311– 320.
- [9] O. Mašek, M. Konno, S. Hosokai, N. Sonoyama, K. Norinaga, J. ichiro Hayashi,

A study on pyrolytic gasification of coffee grounds and implications to allothermal gasification, Biomass and Bioenergy. 32 (2008) 78–89.

- [10] R. Soysa, S.K. Choi, Y.W. Jeong, S.J. Kim, Y.S. Choi, Pyrolysis of Douglas fir and coffee ground and product biocrude-oil characteristics, J. Anal. Appl. Pyrolysis. 115 (2015) 51–56.
- [11] J. Fermoso, P. Pizarro, J.M. Coronado, D.P. Serrano, Advanced biofuels production by upgrading of pyrolysis bio-oil, WIREs Energy Environ. 6 (2017) 1–18.
- [12] K. Raveendran, A. Ganesh, K. Khilar, Pyrolysis characteristics of biomass and biomass components, Fuel. 75 (1996) 987–998.
- [13] J.E. White, W.J. Catallo, B.L. Legendre, Biomass pyrolysis kinetics: A comparative critical review with relevant agricultural residue case studies, J. Anal. Appl. Pyrolysis. 91 (2011) 1–33.
- [14] B. Ramajo-Escalera, A. Espina, J.R. García, J.H. Sosa-Arnao, S.A. Nebra, Model-free kinetics applied to sugarcane bagasse combustion, Thermochim. Acta. 448 (2006) 111–116.
- [15] V. Mamleev, S. Bourbigot, Calculation of activation energies using the sinusoidally modulated temperature, J. Therm. Anal. Calorim. 70 (2006) 565– 579.
- [16] M.E. Brown, M. Maciejewski, S. Vyazovkin, R. Nomen, J. Sempere, A. Burnham, J. Opfermann, R. Strey, H.L. Anderson, A. Kemmler, R. Keuleers, J. Janssens, H.O. Desseyn, C.R. Li, T.B. Tang, B. Roduit, J. Malek, T. Mitsuhashi, Computational aspects of kinetic analysis Part A: The ICTAC Kinetics Projectdata, methods and results, Thermochim. Acta. 355 (2000) 125–143.
- [17] Z. Shuping, W. Yulong, Y. Mingde, L. Chun, T. Junmao, Pyrolysis characteristics and kinetics of the marine microalgae Dunaliella tertiolecta using thermogravimetric analyzer, Bioresour. Technol. 101 (2010) 359–365.
- [18] Y. Liang, B. Cheng, Y. Si, D. Cao, H. Jiang, G. Han, X. Liu, Thermal decomposition kinetics and characteristics of Spartina alterniflora via thermogravimetric analysis, Renew. Energy. 68 (2014) 111–117.
- [19] S.A. Channiwala, P.P. Parikh, A unified correlation for estimating HHV of solid, liquid and gaseous fuels, Fuel. 81 (2002) 1051–1063.
- [20] J.B. Sluiter, R.O. Ruiz, C.J. Scarlata, A.D. Sluiter, D.W. Templeton, Compositional analysis of lignocellulosic feedstocks. 1. Review and description of methods, J. Agric. Food Chem. 58 (2010) 9043–9053.
- [21] S. Vyazovkin, C. a. Wight, Isothermal and non-isothermal kinetics of thermally stimulated reactions of solids, Int. Rev. Phys. Chem. 17 (1998) 407–433.
- [22] H.E. Kissinger, Variation of peak temperature with heating rate in differential thermal analysis, J. Res. Natl. Bur. Stand. (1934). 57 (1956) 217–221.
- [23] J.H. Flynn, L.A. Wall, A quick, direct method for the determination of activation

energy from thermogravimetric data, J. Polym. Sci. Part C Polym. Lett. 4 (1966) 323–328.

- [24] T. Ozawa, A New Method of Analyzing Thermogravimetric Data A New Method of Analyzing Thermogravimetric Data By Takeo OZAWA of the curve. By these superpositions there is obtained curve, which is a standard and more accurate makes possible the mutual comparison of, Bull. Chem. Soc. Jpn. 38 (1965) 1881–1886.
- [25] T. Akahira, T. Sunose, Joint convention of four electrical institutes. Research report (Chiba Institute of Technology)., Sci. Technol. 16 (1971) 22–31.
- [26] J.J. Chew, V. Doshi, Recent advances in biomass pretreatment Torrefaction fundamentals and technology, Renew. Sustain. Energy Rev. 15 (2011) 4212– 4222.
- [27] S. Obruca, P. Benesova, D. Kucera, S. Petrik, I. Marova, Biotechnological conversion of spent coffee grounds into polyhydroxyalkanoates and carotenoids, N. Biotechnol. 32 (2015) 569–574.
- [28] M. Amutio, G. Lopez, R. Aguado, M. Artetxe, J. Bilbao, M. Olazar, Kinetic study of lignocellulosic biomass oxidative pyrolysis, Fuel. 95 (2012) 305–311.
- [29] H. Yang, R. Yan, H. Chen, C. Zheng, D.H. Lee, V. Uni, N.D. V, R. V March, V. Re, M. Recei, V. September, In-Depth Investigation of Biomass Pyrolysis Based on Three Major Components : Hemicellulose, Cellulose and Lignin, (2006) 388– 393.
- [30] H. Yang, R. Yan, H. Chen, D.H. Lee, C. Zheng, Characteristics of hemicellulose, cellulose and lignin pyrolysis, Fuel. 86 (2007) 1781–1788.
- [31] K. Slopiecka, P. Bartocci, F. Fantozzi, Thermogravimetric analysis and kinetic study of poplar wood pyrolysis, Appl. Energy. 97 (2012) 491–497.
- [32] E. Biagini, F. Barontini, L. Tognotti, Devolatilization of biomass fuels and biomass components studied by TG/FTIR technique, Ind. Eng. Chem. Res. 45 (2006) 4486–4493.
- [33] H. Haykiri-Acma, S. Yaman, S. Kucukbayrak, Effect of heating rate on the pyrolysis yields of rapeseed, Renew. Energy. 31 (2006) 803–810.
- [34] S. Maiti, S. Purakayastha, B. Ghosh, Thermal characterization of mustard straw and stalk in nitrogen at different heating rates, Fuel. 86 (2007) 1513–1518.
- [35] Y. Huang, S. Kudo, O. Masek, K. Norinaga, J.I. Hayashi, Simultaneous maximization of the char yield and volatility of oil from biomass pyrolysis, Energy and Fuels. 27 (2013) 247–254.
- [36] J. Fermoso, P. Pizarro, J.M. Coronado, D.P. Serrano, Transportation Biofuels via the Pyrolysis Pathway: Status and Prospects, in: R.A. Meyers (Ed.), Encycl. Sustain. Sci. Technol., Springer, New York, 2017: pp. 1–33.
- [37] J. Fermoso, H. Hernando, S. Jiménez-sánchez, A.A. Lappas, E. Heracleous, P. Pizarro, J.M. Coronado, D.P. Serrano, Bio-oil production by lignocellulose fast-

pyrolysis : Isolating and comparing the e ff ects of indigenous versus external catalysts, Fuel Process. Technol. 167 (2017) 563–574.

- [38] G. Özsin, A.E. Pütün, Kinetics and evolved gas analysis for pyrolysis of food processing wastes using TGA/MS/FT-IR, Waste Manag. 64 (2017) 315–326.
- [39] X. Yao, K. Xu, Y. Liang, Analytical Pyrolysis Study of Peanut Shells using TG-MS Technique and Characterization for the Waste Peanut Shell Ash, J. Residuals Sci. Technol. 13 (2016) 295–305.
- [40] L. Sanchez-Silva, D. López-González, J. Villaseñor, P. Sánchez, J.L. Valverde, Thermogravimetric-mass spectrometric analysis of lignocellulosic and marine biomass pyrolysis, Bioresour. Technol. 109 (2012) 163–172.
- [41] K. Werner, L. Pommer, M. Broström, Thermal decomposition of hemicelluloses, J. Anal. Appl. Pyrolysis. 110 (2014) 130–137.
- [42] H. Hernando, I. Moreno, J. Fermoso, C. Ochoa-Hernández, P. Pizarro, J.M. Coronado, J. Čejka, D.P. Serrano, Biomass catalytic fast pyrolysis over hierarchical ZSM-5 and Beta zeolites modified with Mg and Zn oxides, Biomass Convers. Biorefinery. 7 (2017) 289–304.
- [43] C. Quan, N. Gao, Q. Song, Pyrolysis of biomass components in a TGA and a fixed-bed reactor: Thermochemical behaviors, kinetics, and product characterization, J. Anal. Appl. Pyrolysis. 121 (2016) 84–92.
- [44] L. Burhenne, J. Messmer, T. Aicher, M.P. Laborie, The effect of the biomass components lignin, cellulose and hemicellulose on TGA and fixed bed pyrolysis, J. Anal. Appl. Pyrolysis. 101 (2013) 177–184.

Highlights

- Maximum decomposition rate of each biopolymer increased linearly with heating rate.

- Maximum biopolymers reactivity decreased in the order of: hemicellulose > cellulose > lignin.

- MS evolution of oxygenates (H₂O, CO and CO₂) overlaps with main CGR degradation regime (250 – 425 °C).

 Catalytic pyrolysis improved deoxygenation only negligibly but importantly enhanced vapours cracking.

- KAS and FWO kinetic methods satisfactorily gave rise to E_a values of CGR individual

biopolymers.

Title page

Title:

Thermochemical decomposition of coffee ground residues: a kinetic study

Author names and affiliations

Javier Fermoso^{a,b,*}, Ondřej Mašek^b

^a Thermochemical Processes Unit, IMDEA Energy Institute, 28935, Móstoles, Madrid, Spain.

^b UK Biochar Research Centre, School of Geosciences, University of Edinburgh, Crew Building, Alexander Crum Brown Road, Edinburgh EH9 3FF, UK

Corresponding author*:

Tel.: +34 917371120. *E-mail address:* javier.fermoso@imdea.org (Javier Fermoso). Thermochemical Processes Unit, IMDEA Energy Institute, 28935, Móstoles, Madrid, Spain.

Thermochemical decomposition of coffee ground residues: a kinetic study

Javier Fermoso^{a,b,*}, Ondřej Mašek^b

^a Thermochemical Processes Unit, IMDEA Energy Institute, 28935, Móstoles, Madrid, Spain.

^b UK Biochar Research Centre, School of Geosciences, University of Edinburgh, Crew Building, Alexander Crum Brown Road, Edinburgh EH9 3FF, UK

Abstract

A thermogravimetric analyser coupled to a mass spectrometer, for evolved gas analysis, were employed to perform pyrolysis tests at heating rates (5 - 100 °C/min) and at maximum temperature of 500 °C to determine kinetic parameters of thermochemical decomposition of the biopolymers comprising coffee ground residues. During the pyrolysis process, the maximum decomposition rate of each biomass component increased linearly with the heating rate used. The slope increased with the biopolymer reactivity in the following sequence: hemicellulose > celluose > lignin. Accordingly, kinetic parameters for any of these individual biopolymers in CGR were estimated using the model-free isoconversional dynamic methods: Kissinger-Akahira-Sunose (KAS) and Flynn–Wall–Ozawa (FWO) models. The average value for the apparent activation energy of the individual biopolymers (hemicellulose, cellulose and lignin) in CGR calculated by KAS and FWO methods: were estimated as 214, 241 and 266 kJ/mol, respectively; whilst for the CGR as a whole it was 242 kJ/mol. The two model-free isoconversional dynamic methods have been shown to be useful tools for assessment of biomass pyrolysis kinetic parameters, as they can provide E_a values for use in reactor design models.

Keywords: coffee ground residues; kinetics; isoconversional methods; apparent activation energy

1. Introduction

The depletion of fossil fuel reserves and the environmental problems derived from their utilization have made necessary the use of alternative fuels. Biomass is a clean and renewable energy source leading to environmental, technical and economic benefits. Residues from agricultural production and processing industries are readily available in large quantities. Coffee is the second most traded commodity in the world after oil, and one of the most widely consumed beverages in the world [1]. Moreover, the residues derived from its production are steadily increasing in proportion to the coffee consumption growth [1-3]. Coffee silverskin and spent coffee grounds are the main coffee industry residues [4]. The latter is a residue with fine particle size, high humidity (\approx 80 wt%), organic load and acidity, obtained during the treatment of raw coffee powder with hot water or steam for the instant coffee preparation. Therefore, this residue is generated in large amounts, with a worldwide annual generation of 8 million tons [3]. On an average one ton of green coffee generates about 650 kg of spent coffee ground and about 2 kg of wet spent coffee ground are generated per kg of soluble coffee produced [5]. Coffee grounds are disposed as household waste that may be incinerated and/or moved to landfill [4]. Some alternative applications for coffee ground residues are their utilisation to produce compost [6], deodorizer or adsorbents [7,8], as well as source of renewable energy [9,10]. In this latter regard, the pyrolysis is a thermochemical conversion process that, depending on the reaction conditions, can be used to transform biomass directly into liquid, solid or gaseous biofuels [11]. However, pyrolysis is also the first stage of other thermochemical processes, such as combustion and gasification. In this respect, a systematic understanding of pyrolysis kinetics is a key factor for the

assessment of feasibility, design, and scale-up of such biomass conversion processes for energy applications [12]. Thermogravimetric analysis (TGA) is the most commonly used technique to study the solid-phase thermal decomposition reactions [13]. Although it typically operates in a different form in comparison to a real reactor (pyrolyzer, gasifier or combustor), it provides an understanding of thermal degradation processes occurring during the fuel conversion. In addition, as this study is focused on slow pyrolysis conversion of coffee grounds to solid and gaseous products, the conditions in the TGA represented real reactor conditions much more realistically that would be the case for processes deploying high heating rates, such as fast pyrolysis and gasification.

The thermal decomposition of biomass proceeds via a very complex set of competitive and concurrent reactions and thus, the exact mechanism for biomass pyrolysis remains unknown. Each step likely has its own single apparent activation energy, and thus the use of an average, global apparent activation energy to define the kinetics of such processes could be interpreted as an inadequate simplification at best [14]. Furthermore, the DTG curves from these models may hide the true multi-stage character of pyrolytic reactions under a single peak [15].

During the second half of the 20th century several novel methods for determining Arrhenius parameters based on a single parameter emerged. These so-called "model-free" methods are founded on an isoconversional basis, wherein the degree of conversion, X, for a reaction was assumed to be constant and therefore the reaction rate, k, depended exclusively on the reaction temperature, T. By allowing apparent activation energy (E_a) to be calculated a priori, these approaches eliminate the need to initially hypothesize a form and rate order for the kinetic equation. Hence, isoconversional methods do not require previous knowledge of the reaction mechanism for biomass thermal degradation. Another advantage of such approaches is that the systematic error resulting from the

kinetic analysis during the estimation of the Arrhenius parameters is eliminated [16]. Isoconversional models can follow either a differential or an integral approach to the treatment of TGA data. So, they are considered as a helpful solution for truly determining apparent activation energy [13].

Therefore, the main objective of this study was to determine the kinetic parameters for the slow pyrolysis of coffee ground residues (CGR) and its individual biopolymer components (cellulose, hemicellulose and lignin) employing isoconversional model-free dynamic methods: Kissinger-Akahira-Sunose (KAS) and Flynn-Wall-Ozawa (FWO), due to the advantages they offer in determining Arrhenius equation parameters without the need to make choices regarding kinetic models to be used [17].

During pyrolysis, due to the poor thermal conductivity of biomass, a temperature gradient is normally developed through the biomass sample between the external surface and the internal part. This gradient may be assumed to be proportional to the particle size, and decreases with reducing heating rate to the point that both, the external surface and the internal part of the biomass particles attains same temperature at a certain reaction time when appropriate time is given for heating [18]. Therefore, a defined particle size range was seleced in this study to avoid the effect of the particle size on the determination of kinetic parameters under dynamic conditions of different heating rates.

The peak decomposition rates of biopolymers constituting the CGR and associated temperatures during pyrolysis process were evaluated. This was achieved by thermogravimetric analysis utilising different heating rates under inert (N_2) atmosphere. The obtained data and models provide important basis for design and operation of slow pyrolysis or estaged pyrolysis systems using coffee ground residues for production of biochar and chemicals.

2. Materials and methods

2.1. Materials

The biomass employed in this study was coffee ground residue (CGR), collected from local canteen at the University, ensuring that one type of beans was used. As the pre-dried CGR still contained ≈ 15 wt% of moisture, the material was air dried for 24 hours at 105 °C in a laboratory oven before further use. To avoid the effect of the particle size on the kinetic parameters determination, the CGR was crushed and sieved to collect particles in the range of $250 - 500 \mu m$.

2.2. Analytical techniques

The proximate analysis was determined according to European standards: moisture content (UNE-EN 14774-1:2010), ash content (UNE-EN 14775:2010), volatile matter (UNE-EN 15148:2010) and fixed carbon (determined by difference). A thermogravimetric analizer, TGA (Mettler-Toledo TGA/DSC1) equipped with automatic sample handling was employed to asses the volatile matter and the ash contents of the coffee ground residue. This is a well established thermoanalytical technique for thermal degradation studies of solid materials, such as biomass pyrolysis [13].

A mass spectrometer, MS (HIDEN Analytical HPR-20) coupled to the TGA was employed for evolved gas analysis during the pyrolysis experiments. The ultimate analysis of feedstock and was carried out in a micro-elemental analyzer (Thermo Scientific) in order to determine content of C, H, N, S and O (by difference). The higher heating value (HHV) of CGR was calculated using the formula developed by Channiwala and Parikh [19]. The relative abundance of individual biopolymers (cellulose, hemicellulose and lignin) in CGR was determined by the sulfuric acid hydrolysis method [20].

2.3. Pyrolysis tests

The pyrolysis tests were performed in the same TGA previously mentioned, at atmospheric pressure. The CGR sample (around 15 mg) was deposited in an alumina

crucible with a circular base and total volume of 150 μ l. In this work, all the experiments were performed under non-isothermal conditions at different heating rates (HR), 5–100°C/min, with a nitrogen flow rate of 100 ml/min.

2.4. Kinetic models

The one-step global model assumes that the devolatilization phenomena proceeds as a single reaction.

$$Biomass \xrightarrow{k} Volatiles + Char \tag{1}$$

where *Volatiles* represents the sum of the gas and bio-oil, and *char* is the remaining unreacted solid. The fundamental rate of transformation from solid-state to volatiles is generally described by the following expression:

$$\frac{dX}{dt} = k(T)f(X) \tag{2}$$

where *X* is the degree of conversion of the fuel, *t* is the reaction time, k(T) is the reaction rate constant, and f(X) is a function that represents the reaction model.

The degree of conversion, X, is calculated as its relative weight loss as follows:

$$X = \frac{m_0 - m_t}{m_0 - m_f}$$
(3)

where m_0 , m_t and m_f represents the initial mass, the mass at time *t*, and the final residual mass of the sample, respectively.

The reaction rate constant, k, is temperature dependent, and it obeys the fundamental Arrhenius rate expression:

$$k = A \cdot e^{\frac{-E_a}{RT}} \tag{4}$$

where A is the pre-exponential factor (min⁻¹), E_a is the apparent activation energy (kJ/mol), T is the absolute temperature (K) and R is the gas constant (8.314 J/mol·K). Non-isothermal method employs a heating rate (β), normally linear, to raise the temperature. A linear heating program follows:

$$T = T_0 + \beta t \tag{5}$$

$$\beta = \frac{dT}{dt} \tag{6}$$

where T_0 is the starting temperature, β the constant heating rate (K/min), and T the temperature at time t. Then, substituting equations (4) and (6) in equation (2) gives:

$$\frac{dX}{dT} = \frac{A}{\beta} \cdot e^{\frac{-E_a}{RT}} f(X)$$
(7)

Equation (7) represents the differential form of the non-isothermal rate law.

Kinetics analysis is conventionally expected to produce a suitable kinetic description of the process in terms of the reaction model and the Arrhenius parameters. These three components, f(X), E_a , and A, are sometimes called the "kinetic triplet". There are many methods for analysing solid-state kinetic data [21]. These methods can be classified according to the experimental conditions and the mathematical analysis implemented. The mathematical approaches employed can be divided into model-fitting and isoconversional (model-free) methods. However, as discussed in the introduction section, in this work only the isoconversional model-free dynamic methods were used to calculate the kinetic parameters for the CGR pyrolysis, which require a set of experimental tests at different heating rates. These methods are the Kissinger, Kissinger-Akahira-Sunose (KAS) and Flynn-Wall-Ozawa (FWO) methods. The advantages of the model-free analysis are: its simplicity, and the avoidance of errors associated to choices of a kinetic model [17].

2.5. Model-free methods

Kissinger method

This method allows for the kinetic parameters of a solid-state reaction without prior knowledge of the reaction mechanism. Kissinger [22] developed a model-free non-

isothermal method where E_a does not need to be calculated for each conversion value in order to evaluate kinetic parameters. The method equation is represented as follows:

$$Ln\left(\frac{\beta}{T_m^2}\right) = Ln\left(\frac{A \cdot R}{E_a}\right) - \frac{E_a}{RT_m}$$
(8)

According to Kissinger, in the differential thermogravimetric curve (DTG), the temperature at which the peak weight loss velocity (in %/min) occurs for a given heating rate is determined by both A and E_a . Then, changing the heating rate the peak temperature will change. Hence, plotting $Ln(\beta/T_m^2)$ versus $1/T_m$, should give a straight line of slope $-E_a/R$.

Flynn-Wall-Ozawa method (FWO)

The *FWO* method [23,24] is one of the most commonly accepted methods for the computation of kinetic parameters. It allows for the apparent activation energy to be obtained for each degree of conversion from the equation:

$$Ln(\beta) = Ln\left(\frac{A_X \cdot E_{a_X}}{R \cdot g(X)}\right) - 5.331 - 1.052 \frac{E_{a_X}}{R \cdot T_X}$$
(9)

where E_{aX} is the apparent activation energy for a fix degree of conversion X, and is calculated from the slope of the straight line obtained by plotting logarithm of heating rates, $\text{Ln}\beta$, versus $1/T_X$, where T_X is the reaction temperature at which this grade of conversion X is reached.

Kissinger-Akahira-Sunose method (KAS)

The *KAS* method [22,25] is an integral isoconversional technique based on the following expression:

$$Ln\left(\frac{\beta}{T_{X}^{2}}\right) = Ln\left(\frac{A_{X}\cdot R}{E_{a_{X}}\cdot g(X)}\right) - \frac{E_{a_{X}}}{R\cdot T_{X}}$$
(10)

where E_{aX} is the apparent activation energy for a fix degree of conversion *X*, and is calculated from the slope of the straight line obtained by plotting $\text{Ln}(\beta/T_X^2)$, versus $1/T_X$, where T_X is the reaction temperature at which this grade of conversion *X* is reached.

3. Results and discussion

3.1. Biomass analysis

Table 1 summarizes the proximate and ultimate analysis, as well as composition in terms of key biopolymers (hemicellulose, cellulose and lignin) of CGR. The pre-dried sample still showed 5 wt% of moisture because this material rapidly adsorbs moisture during transfer and storage. The table also shows that CGR contains more carbon and less oxygen (and therefore lower O/C ratio \approx 0.66) than woody biomass and agricultural residues (O/C \approx 0.8-1.2) [26]. Such behaviour is attributed to the higher lignin contents in CGR, which reaches 40.6 wt% (whose O/C ratio \approx 0.4 – 0.5), while hemicellulose and cellulose account for 36.6 and 10.6 wt%, respectively. This composition is rather similar to that reviewed by Obruca et al. [27] in terms of holocellulose biopolymers but with larger content of lignin. Consequently, the high heating value (HHV) of CGR, 23.4 MJ/kg, is higher than that of most biomass (17-20 MJ/kg) [26]. However, ultimate analysis also showed that CGR contains more nitrogen (2.3 wt%) than other more commonly used lignocellulosic biomass (0.1-1.0 wt%) due to high protein and caffeine content [4].

3.2. CGR thermochemical decomposition (TGA)

Fig. 1 shows both the conversion curves as weight loss in wt% (A), and their first derivative curves with time (DTG) as wt%/min (B) of the thermochemical decomposition of CGR as a function of reaction temperature under nitrogen atmosphere at 500 °C and at five heating rates (HR): 5 - 100 °C/min. The conversion curves at all heating rates indicate that mass loss of CGR mainly occurred at temperatures ranging from 250 to 500 °C. The conversion curves shift to the right with increasing heating rate as can be observed in Fig.

1(A), which implies higher values of initial decomposition temperature (see Table 2). However, this representation of the conversion data makes it difficult to identify the changes in the slope at different temperatures and reaction rates for the thermal decomposition of the three biopolymers contained in CGR. Therefore, the DTG curves, as shown in Fig. 1(B) were derived from the TG data, clearly showing three conversion peaks corresponding to the well established order of biopolymers decomposition: hemicellulose, cellulose and lignin [28–30]. As the figure shows, the three peaks can be clearly distinguished for all the heating rates, with the exception of 100 °C/min. Although the identification of the maximum degradation rate of lignin in biomass is not commonly reported in the literature, as it usually overlaps with the cellulose decomposition peak [13,31,32], in the case of CGR it was possible, due to the large difference in cellulose and lignin contents, ≈ 11 and 42 wt%, respectively. Based on this initial observation it might be suggested that a more efficient staged pyrolysis process of these materials could be carried out to preferentially obtaining products of decomposition of individual biomass constituents separately when heating the material to different temperatures in stages. On the other hand, it is evident from the results that increasing the heating rate from 5 to 100 °C/min resulted in a progressive rise in the total volatile matter released at 500 °C from 72.2 to 75.1 wt%, respectively as summarized in Table 2. This behaviour agrees with the theory that heating rate has an influence on the secondary reactions of the primary pyrolysis vapours. So, lower heating rates result in longer residence times of volatiles inside biomass particles and the reactor, favouring secondary reactions such as cracking, re-polymerization and re-condensation, which eventually lead to the char formation [13,18,33–35]. This is normally observed when comparing slow pyrolysis with fast/flash pyrolysis according to the product goal for the pyrolysis process; i.e., biochar production (for slow pyrolysis: 0.1-2 °C/s) or bio-oil production for fast (10-200 °C/s) and flash

pyrolysis (>1000 °C/s) [36]. However, it is interesting to see this effect even in the heating rate range still corresponding to slow (5-50 °C/min) or at best intermediate (100 °C/min) pyrolysis here studied because it could be used to modulate the reaction to the desire products.

Fig. 2(A) depicts the temperatures of maximum decomposition rate of CGR biopolymers during pyrolysis as a function of the heating rate. Here it can be observed that these peak temperatures logarithmically depend on the heating rate for the three biopolymers; which implies that at low heating rates the mass transfer limitations are more important than at high heating rates, at which the maximum decomposition rate for the different bipolymers occurs at similar temperatures.

On the other hand, Fig. 2(B) shows that the maximum decomposition rate increased linearily with heating rate for all biopolymers in CGR. In addition, the observed difference in the slope of these lines suggests that the heating rate affected diverse biopolymers differently; thus, the more reactive the material (hemicellulose > celluose > lignin) the higher the slope. As lignin is the most stable and complex of biopolymers comprising biomass, its amount is assumed to be the main rate limiting factor in the thermochemical decomposition process of CGR.

Fig. 3 shows the MS spectrum of the evolved gas species during the pyrolysis of CGR at 500 °C and 15 °C/min heating rate versus reaction time. DTG curve and temperature profile are also plotted to show which gaseous compounds were evolved at each stage of the pyrolysis and at which temperatures. Water is the principal component, and it has two origins. Firstly, the physically adsorbed water, which is desorbed at \approx 90 °C; and secondly reaction water, produced at 250 – 400 °C, with a maximum production at 314 °C, originated from various dehydration reactions of the original CGR biopolymers and/or dehydration of the primary and secondary pyrolysis vapours [37]. The other main gases

are CO and CO₂ produced from decarbonylation and decarboxylation reactions of same original biopolymers and primary and secondary vapours, respectively; generated in the same range of temperatures, with maximum production shifted to a slightly higher temperature 330 °C. These three oxygen containing gases seem to originate from the thermal degradation of the three biopolymers that constitute the CGR (hemicellulose, cellulose and lignin), which is visible on the DTG curve. Moreover, this is also demonstrated by the two small shoulders to the right from the main peak of these evolved gases. In addition, H₂ and light hydrocarbons (C₁-C₂) are evolved to a much lower extent, with two small peaks appearing at 330 and 480 °C, which might be related to the aromatization or the char structure during the secondary pyrolysis [13]. These gas evolution trends apply to all the heating rates investigated in this study.

3.3. Kinetic analysis

The TGA experimental data were analyzed in order to obtain the kinetic parameters using three model-free methods. To avoid any influence of the physically bound moisture desorption from CGR sample, the conversion (in wt%) was calculated from the experimental data collected at temperatures between 150 and 500 °C, corresponding to the active pyrolysis stage where hemicellulose, cellulose and lignin decomposition occurs. Figs. 4(A - C) show the plots corresponding to the Kissinger, FWO and KAS models, respectively, used to calculate the kinetic parameters of the dynamic degradation of the biopolymers in CGR according to equations described in Section 2.3. Data in Fig. 4(A) were used to calculate the *A* and E_a according to the Kissinger method. This method has the disadvantage that these parameters are calculated just from the temperature that corresponds to the highest weight loss rate (DTG_{max}). So, it means that this method should be employed simply for those samples showing a single DTG peak. In the case of CGR, these parameters would purely correspond to the hemicellulose degradation, which is the

most reactive biopolymer in this biomass, in terms of degradation rate as shown in Fig. 2, disregarding the role of the other two biopolymers. However, the FWO and KAS methods calculate the kinetic parameters based on values of conversion from 5 to 90 wt%, with a 5 wt% step as shown in Figs. 4(B - C), which implies that the kinetic parameters for the individual biopolymers that comprise CGR may be estimated. These lines from linear fit at different conversion levels have fairly high linear correlation coefficients, > 0.995 (as summarized in Table 3), suggesting that the values of E_a and A satisfy accuracy requirements. This can be observed in Fig. 5, where the calculated apparent E_a is plotted as a function of the conversion level for FWO and KAS methods in comparison with that constant value obtained with Kissinger method. Here, it can be seen that E_a increases with temperature and conversion level. This is characteristic of processes with different reaction mechanisms. Even when each of the biopolymers that constitutes the CGR (hemicellulose, cellulose and lignin) has its own apparent activation energy, its thermal decomposition proceeds via a very complex set of competitive and concurrent reactions, due to the synergistic effects between its biopolymers. However, an approximate estimation of the E_a of these components can be assessed from the average values of different steps shown in Fig. 5. Thus, hemicellulose, cellulose and lignin would present values of 213, 240 and 265 kJ/mol, respectively according to FWO method; while 215, 242 and 268 kJ/mol, respectively for the KAS method as is summarized in Table 3. The results show that the values estimated for the decomposition of hemicellulose are very close to those obtained for CGR using the Kissinger method (212 kJ/mol), which is in concordance as this method uses the maximum decomposition rate to calculate the kinetic parameters. When the conversion increases further than 80 wt%, the apparent activation energy increased sharply from 273 kJ/mol to 347 kJ/mol for FWO, and from 276 kJ/mol to 353 kJ/mol for KAS method as shown in Table 3, which could be due to

the re-polymerization and re-condensation reactions leading to char formation. Then, taking into account all the steps in the pyrolysis process of the coffee ground residues, the apparent activation energy for the whole process was estimated as 244 and 241 kJ/mol for KAS and FWO methods, respectively. These E_a results are in correspondence with thermostability sequence analysis of these three CGR biopolymer components (hemicellulose, cellulose and lignin) [38]. Therefore, the lignin content would be the main controlling factor in biomass pyrolysis in industrial processes [39]. In this way, these kind of model-free isoconversional dynamic methods have been shown as useful tools to assess the kinetic parameters of CGR, as they can provide E_a values to be applied in models for designing reactors for its utilisation as fuel. Furthermore, due to the ability to obtain kinetic parameters for decomposition of individual biolopymenrs, the data could be used for designing a more efficient staged pyrolysis process allowing preferential recovery of decomposition products of individual biopolymers separately. Such process would enable more efficient production of high-value chemicals from biomass.

Conclusions

In this work, the kinetic parameters of the thermochemical decomposition of biopolymer components of coffee ground residues were estimated by thermogravimetric analysis. For that purpose, a thermogravimetric analyser coupled to a mass spectrometer for the detection of the evolved gases were employed to perform the pyrolysis tests at different heating rates (5 - 100 °C/min) and at maximum temperature of 500 °C. The results shoed that the heating rate significantly affected the thermal decomposition of coffee ground residues during pyrolysis. The maximum decomposition rate increased linearly with the heating rate; but also, the more reactive the material (hemicellulose > celluose > lignin) the higher the slope.

Main gases produced during the pyrolysis of coffee ground residues were oxygen containing species, and were evolved between 250 and 425 °C, with H₂O being the most important, followed by CO (from decarbonylation reactions) and CO₂ (from decarboxylation reactions) of parent biopolymers and primary and secondary vapours. Kinetic parameters of the pyrolysis process were determined using isoconversional methods (Kissinger, KAS and FWO). The apparent activation energy (212 kJ/mol) obtained from the Kissinger method would simply correspond to that of the hemicellulose biopolymer in coffee residues, as the most reactive component in this biomass. However with the other two methods, KAS and FWO, the apparent activation energy and preexponential factors increase with the conversion level, revealing a complex set of competitive and concurrent reactions. The apparent activation energy estimated for the major constituents of coffee ground residues with the two isoconversional dynamic methods, FWO and KAS, were rather similar as shown in Fig. 5 and summarized in Table 3. The average value for the E_a of the hemicellulose, cellulose and lignin from these two methods were equal to 214, 241 and 266 kJ/mol, respectively. Whereas the apparent activation energy for the whole pyrolysis process would increase up to 242 kJ/mol using the same methods. Thus, these kind of model-free isoconversional dynamic methods have been demonstrated to be valuable tools to assess the kinetic parameters of CGR, as they can provide E_a values to be applied in models for designing reactors for its utilisation as fuel; but also, for a more efficient staged pyrolysis process.

Acknowledgement

The authors gratefully acknowledge the experimental support provided by the UK Biochar Reseach Centre (University of Edinburgh). Dr Javier Fermoso also

acknowledges the IMDEA Energy Institute for the financial support to accomplish the

research stay at the UK Biochar Research Centre (University of Edinburgh).

References

- [1] F. Girotto, A. Pivato, R. Cossu, G.E. Nkeng, M.C. Lavagnolo, The broad spectrum of possibilities for spent coffee grounds valorisation, J. Mater. Cycles Waste Manag. (2017) 1–7. doi:10.1007/s10163-017-0621-5.
- [2] S.I. Mussatto, E.M.S. Machado, S. Martins, J.A. Teixeira, Production, Composition, and Application of Coffee and Its Industrial Residues, Food Bioprocess Technol. 4 (2011) 661–672.
- [3] R. Campos-Vega, G. Loarca-Piña, H.A. Vergara-Castañeda, B.D. Oomah, Spent coffee grounds: A review on current research and future prospects, Trends Food Sci. Technol. 45 (2015) 24–36.
- [4] S.I. Mussatto, E.M.S. Machado, S. Martins, J.A. Teixeira, Production, Composition, and Application of Coffee and Its Industrial Residues, Food Bioprocess Technol. 4 (2011) 661–672.
- [5] P.S. Murthy, M. Madhava Naidu, Sustainable management of coffee industry byproducts and value addition - A review, Resour. Conserv. Recycl. 66 (2012) 45– 58.
- [6] J.P. Bok, H.S. Choi, Y.S. Choi, H.C. Park, S.J. Kim, Fast pyrolysis of coffee grounds: Characteristics of product yields and biocrude oil quality, Energy. 47 (2012) 17–24.
- [7] T. Tokimoto, N. Kawasaki, T. Nakamura, J. Akutagawa, S. Tanada, Removal of lead ions in drinking water by coffee grounds as vegetable biomass, J. Colloid Interface Sci. 281 (2005) 56–61.
- [8] S. Rovani, M.T. Censi, S.L. Pedrotti, E.C. Lima, R. Cataluña, A.N. Fernandes, Development of a new adsorbent from agro-industrial waste and its potential use in endocrine disruptor compound removal, J. Hazard. Mater. 271 (2014) 311– 320.
- [9] O. Mašek, M. Konno, S. Hosokai, N. Sonoyama, K. Norinaga, J. ichiro Hayashi, A study on pyrolytic gasification of coffee grounds and implications to allothermal gasification, Biomass and Bioenergy. 32 (2008) 78–89.
- [10] R. Soysa, S.K. Choi, Y.W. Jeong, S.J. Kim, Y.S. Choi, Pyrolysis of Douglas fir and coffee ground and product biocrude-oil characteristics, J. Anal. Appl. Pyrolysis. 115 (2015) 51–56.

1005		
1006	[11]	J Fermoso P Pizarro JM Coronado DP Serrano Advanced biofuels
1007	[]	production by upgrading of pyrolysis bio-oil WIREs Energy Environ 6 (2017)
1008		1–18
1009		1 10.
1010	[12]	K. Raveendran, A. Ganesh, K. Khilar, Pyrolysis characteristics of biomass and
1011		biomass components, Fuel. 75 (1996) 987–998.
1012		
1013	[13]	J.E. White, W.J. Catallo, B.L. Legendre, Biomass pyrolysis kinetics: A
1014		comparative critical review with relevant agricultural residue case studies, J.
1015		Anal. Appl. Pyrolysis. 91 (2011) 1–33.
1017	F1 43	
1018	[14]	B. Ramajo-Escalera, A. Espina, J.K. Garcia, J.H. Sosa-Arnao, S.A. Nebra,
1010		Model-free kinetics applied to sugarcane bagasse combustion, Thermochim.
1020		Acta. 448 (2006) 111–116.
1021	[15]	V. Mamloov, S. Pourbigot, Calculation of activation anargies using the
1022	[13]	v. Maineev, S. Bourdigor, Calculation of activation energies using the sinuscidally modulated temperature. J. Therm. Anal. Colorim. 70 (2006) 565
1023		570
1024		519.
1025	[16]	M E Brown M Maciejewski S Vyazovkin R Nomen I Semnere A
1026	[10]	Burnham I Opfermann R Strey H L Anderson A Kemmler R Keuleers I
1027		Janssens H.O. Dessevn C.R. Li T.B. Tang B. Roduit I. Malek T. Mitsuhashi
1028		Computational aspects of kinetic analysis Part A: The ICTAC Kinetics Project-
1029		data methods and results. Thermochim Acta 355 (2000) 125–143
1030		data, methods and results, methodmin. Acta. 555 (2000) 125-145.
1031	[17]	Y. Liang, B. Cheng, Y. Si, D. Cao, H. Jiang, G. Han, X. Liu, Thermal
1032		decomposition kinetics and characteristics of Spartina alterniflora via
1033		thermogravimetric analysis, Renew, Energy, 68 (2014) 111–117.
1034		
1035	[18]	Z. Shuping, W. Yulong, Y. Mingde, L. Chun, T. Junmao, Pyrolysis
1036		characteristics and kinetics of the marine microalgae Dunaliella tertiolecta using
1037		thermogravimetric analyzer, Bioresour. Technol. 101 (2010) 359-365.
1038		
1039	[19]	S.A. Channiwala, P.P. Parikh, A unified correlation for estimating HHV of solid,
1040		liquid and gaseous fuels, Fuel. 81 (2002) 1051–1063.
1041	[20]	I.B. Shuitar, B.O. Buiz, C.I. Saarlata, A.D. Shuitar, D.W. Tamplatan
1042	[20]	J.B. Slutter, R.O. Ruiz, C.J. Scarlata, A.D. Slutter, D.W. Templeton,
1043		Compositional analysis of lignocentriosic reedstocks. 1. Review and description
1044		of methods, J. Agric. Food Chem. 38 (2010) 9043–9053.
1046	[21]	S Vyazovkin C a Wight Isothermal and non-isothermal kinetics of thermally
1047	[21]	stimulated reactions of solids. Int. Rev. Phys. Chem. 17 (1998) 407–433
1048		
1049	[22]	H.E. Kissinger, Variation of peak temperature with heating rate in differential
1050		thermal analysis, J. Res. Natl. Bur. Stand. (1934). 57 (1956) 217–221.
1051		
1052	[23]	J.H. Flynn, L.A. Wall, A quick, direct method for the determination of activation
1053		energy from thermogravimetric data, J. Polym. Sci. Part C Polym. Lett. 4 (1966)
1054		323–328.
1055	50 (7	
1056	[24]	T. Ozawa, A New Method of Analyzing Thermogravimetric Data A New Method
1057		of Analyzing Thermogravimetric Data By Takeo OZAWA of the curve . By
1058		these superpositions there is obtained curve, which is a standard and more
1059		accurate makes possible the mutual comparison of, Bull. Chem. Soc. Jpn. 38
1060		
1061		

(1965) 1881-1886.

- [25] T. Akahira, T. Sunose, Joint convention of four electrical institutes. Research report (Chiba Institute of Technology)., Sci. Technol. 16 (1971) 22–31.
- [26] J.J. Chew, V. Doshi, Recent advances in biomass pretreatment Torrefaction fundamentals and technology, Renew. Sustain. Energy Rev. 15 (2011) 4212– 4222.
- [27] S. Obruca, P. Benesova, D. Kucera, S. Petrik, I. Marova, Biotechnological conversion of spent coffee grounds into polyhydroxyalkanoates and carotenoids, N. Biotechnol. 32 (2015) 569–574.
- [28] M. Amutio, G. Lopez, R. Aguado, M. Artetxe, J. Bilbao, M. Olazar, Kinetic study of lignocellulosic biomass oxidative pyrolysis, Fuel. 95 (2012) 305–311.
- [29] H. Yang, R. Yan, H. Chen, C. Zheng, D.H. Lee, V. Uni, N.D. V, R. V March, V. Re, M. Recei, V. September, In-Depth Investigation of Biomass Pyrolysis Based on Three Major Components : Hemicellulose, Cellulose and Lignin, (2006) 388– 393.
- [30] H. Yang, R. Yan, H. Chen, D.H. Lee, C. Zheng, Characteristics of hemicellulose, cellulose and lignin pyrolysis, Fuel. 86 (2007) 1781–1788.
- [31] K. Slopiecka, P. Bartocci, F. Fantozzi, Thermogravimetric analysis and kinetic study of poplar wood pyrolysis, Appl. Energy. 97 (2012) 491–497.
- [32] E. Biagini, F. Barontini, L. Tognotti, Devolatilization of biomass fuels and biomass components studied by TG/FTIR technique, Ind. Eng. Chem. Res. 45 (2006) 4486–4493.
- [33] H. Haykiri-Acma, S. Yaman, S. Kucukbayrak, Effect of heating rate on the pyrolysis yields of rapeseed, Renew. Energy. 31 (2006) 803–810.
- [34] S. Maiti, S. Purakayastha, B. Ghosh, Thermal characterization of mustard straw and stalk in nitrogen at different heating rates, Fuel. 86 (2007) 1513–1518.
- [35] Y. Huang, S. Kudo, O. Masek, K. Norinaga, J.I. Hayashi, Simultaneous maximization of the char yield and volatility of oil from biomass pyrolysis, Energy and Fuels. 27 (2013) 247–254.
- [36] J. Fermoso, P. Pizarro, J.M. Coronado, D.P. Serrano, Transportation Biofuels via the Pyrolysis Pathway: Status and Prospects, in: R.A. Meyers (Ed.), Encycl. Sustain. Sci. Technol., Springer, New York, 2017: pp. 1–33.
- [37] J. Fermoso, H. Hernando, S. Jiménez-sánchez, A.A. Lappas, E. Heracleous, P. Pizarro, J.M. Coronado, D.P. Serrano, Bio-oil production by lignocellulose fast-pyrolysis : Isolating and comparing the e ff ects of indigenous versus external catalysts, Fuel Process. Technol. 167 (2017) 563–574.
- [38] C. Quan, N. Gao, Q. Song, Pyrolysis of biomass components in a TGA and a fixed-bed reactor: Thermochemical behaviors, kinetics, and product characterization, J. Anal. Appl. Pyrolysis. 121 (2016) 84–92.

Figures

Figure Captions

Fig. 1. TG (A) and DTG (B) curves of the thermal decomposition of coffee ground residues (CGR) under N_2 atmosphere at 500 °C and at different heating rates (5 – 100 °C/min).

Fig. 2. Temperature of maximum DTG (A) and DTG_{max} values (B) corresponding to the peaks associated to the thermochemical decomposition of three biopolymers (hemicellulose, cellulose and lignin) of coffee ground residues (CGR) under N₂ atmosphere at 500 °C and at different heating rates (5 – 100 °C/min).

Fig. 3. Conversion and DTG curves, and evolved gaseous species (MS signal) versus reaction time during the pyrolysis of coffee ground residues (CGR) under N_2 atmosphere at 500 °C (heating rate: 15 °C/min).

Fig. 4. Evolution of the gaseous species (MS signal) versus reaction time during the pyrolysis of coffee ground residues (CGR) under N₂ atmosphere at 500 °C and different heating rates.

Fig. 5. Evolution of the gaseous species (MS signal) versus reaction time during the noncatalytic (red) and catalytic (blue) pyrolysis of coffee ground residues (CGR) under N_2 atmosphere at 500 °C (heating rate: 15 °C/min).

Fig. 6. Calculated apparent activation energy for pyrolysis of coffee ground residues (CGR) by the Kissinger, FWO and KAS kinetic methods for heating rates of between 5 and 50 °C/min.



Fig. 1. TG (A) and DTG (B) curves of the thermal decomposition of coffee ground residues (CGR) under N_2 atmosphere at 500 °C and at different heating rates (5 – 100 °C/min).



Fig. 2. Temperature of maximum DTG (A) and DTG_{max} values (B) corresponding to the peaks associated to the thermochemical decomposition of three biopolymers (hemicellulose, cellulose and lignin) of coffee ground residues (CGR) under N₂ atmosphere at 500 °C and at different heating rates (5 – 100 °C/min).



Fig. 3. Conversion and DTG curves, and evolved gaseous species (MS signal) versus reaction time during the pyrolysis of coffee ground residues (CGR) under N_2 atmosphere at 500 °C (heating rate: 15 °C/min).



Fig. 4. Evolution of the gaseous species (MS signal) versus reaction temperature during the pyrolysis of coffee ground residues (CGR) under N_2 atmosphere at 500 °C and different heating rates.



Fig. 5. Evolution of the gaseous species (MS signal) versus reaction time during the noncatalytic (red) and catalytic (blue) pyrolysis of coffee ground residues (CGR) under N_2 atmosphere at 500 °C (heating rate: 15 °C/min).



Fig. 6. Calculated apparent activation energy for pyrolysis of coffee ground residues (CGR) by the Kissinger, FWO and KAS kinetic methods for heating rates of between 5 and 50 °C/min.

SUPPORTING INFORMATION

Thermochemical decomposition of coffee ground residues by TG-MS: a kinetic study



Javier Fermoso, Ondřej Mašek

Fig. S1. Plots of the Kissinger (A), FWO (B) and KAS (C) kinetic methods of CGR thermal decomposition from 150 to 500 °C at different heating rates (5 – 50 °C/min) to the calculation of the apparent activation energy (E_a). Conversion values of between 5 and 90 wt% were used for FWO and KAS methods.

	KAS		FWO			
Conversion (wt%)	E _a (kJ mol ⁻¹)	A (min ⁻¹)	R ²	E _a (kJ mol ⁻¹)	A (min ⁻¹)	R ²
5	199.1	2.02E+19	0.9988	197.8	1.59E+19	0.9990
10	206.9	2.66E+19	0.9997	205.5	2.07E+19	0.9997
15	212.0	3.59E+19	0.9997	210.6	2.75E+19	0.9997
20	215.2	3.91E+19	0.9996	213.7	2.98E+19	0.9996
25	217.4	3.86E+19	0.9996	215.9	2.95E+19	0.9996
30	219.3	3.67E+19	0.9995	217.7	2.81E+19	0.9996
35	221.9	4.10E+19	0.9995	220.3	3.12E+19	0.9995
40	227.0	7.47E+19	0.9994	225.2	5.54E+19	0.9995
45	235.3	2.35E+20	0.9991	233.2	1.66E+20	0.9992
50	243.5	2.02E+19	0.9984	241.1	4.39E+20	0.9985
55	244.8	2.02E+19	0.9975	242.5	2.98E+20	0.9976
60	242.4	2.02E+19	0.9974	240.4	1.06E+20	0.9976
65	246.3	1.62E+20	0.9975	244.2	1.15E+20	0.9977
70	259.9	9.15E+20	0.9962	257.3	6.09E+20	0.9964
75	266.8	1.24E+21	0.9952	264.1	8.13E+20	0.9955
80	275.9	2.44E+21	0.9963	272.9	1.56E+21	0.9966
85	298.5	2.02E+19	0.9982	294.6	2.90E+22	0.9983
90	352.9	2.02E+19	0.9988	346.6	7.05E+25	0.9989
Estimated average values						
CGR	243.6	3.00E+20		241.3	3.92E+24	
Hemicellulose	214.8	3.91E+19		213.3	2.98E+19	
Cellulose	242.4	9.15E+19		240.3	2.25E+20	
Lignin	267.5	1.53E+21		264.8	9.94E+20	
KISSINGER	212.4	8.48E+18				

Table S1. Kinetic parameters of coffee ground residues (CGR) obtained from the Kissinger,

 KAS and FWO models

Analysis	Coffee ground residues (CGR)		
Moisture (wt%)	5.0		
Proximate analysis, db (wt%)			
Ash	0.9		
Volatile Matter	76.4		
Fixed Carbon	22.7		
Ultimate analysis, db (wt%)			
С	53.9		
Н	7.1		
Ν	2.3		
0	35.8		
HHV (MJ kg _{db} ⁻¹)	23.4		
Cellulose	10.6		
Hemicellulose	36.6		
Lignin	40.6		
Others*	12.2		

Table 1. Proximate, ultimate and biopolymers analyses of dry coffee ground residues

db: dry basis * Organic extractives unidentified compounds and ash determined by difference

Heating rate (°C min ⁻¹)	T _i (°C)	T _{DTGmax} (°C)	DTG _{max} (wt% min ⁻¹)	X _{max, 500°C} (wt%)
5	173	300	3.5	72.2
10	180	310	6.9	72.9
15	200	315	10.3	73.6
25	216	321	16.9	73.7
50	225	330	34.6	74.6
100	237	336	68.5	75.1

Table 2. Decomposition characteristic of coffee ground residues (CGR) at different heating rates