

LONDON
SCHOOL of
HYGIENE
& TROPICAL
MEDICINE



LSHTM Research Online

Kesdangakonwut, Sawang; Sommanustweechai, Angkana; Chaiprasert, Angkana; (2015) Disseminated Mycobacterium tuberculosis in Imported Sooty Mangabey, Thailand. EMERGING INFECTIOUS DISEASES, 21 (3). pp. 534-535. ISSN 1080-6040 DOI: <https://doi.org/10.3201/eid2103.131785>

Downloaded from: <http://researchonline.lshtm.ac.uk/4652260/>

DOI: <https://doi.org/10.3201/eid2103.131785>

Usage Guidelines:

Please refer to usage guidelines at <https://researchonline.lshtm.ac.uk/policies.html> or alternatively contact researchonline@lshtm.ac.uk.

Available under license: <http://creativecommons.org/licenses/by/2.5/>

<https://researchonline.lshtm.ac.uk>

3. Rojas-Solano JR, Villalobos-Vindas J. Ehrlichiosis granulocitotrópica humana. *Acta Med Costarric*. 2007;49:121–3 [cited 2013 Nov 25]. http://www.scielo.sa.cr/scielo.php?script=sci_arttext&pid=S0001-60022007000200012&lng=en&nrm=iso
4. Rojas A, Rojas D, Montenegro V, Gutiérrez R, Yasur-Landau D, Baneth G. Vector-borne pathogens in dogs from Costa Rica: first molecular description of *Babesia vogeli* and *Hepatozoon canis* infections with a high prevalence of monocytic ehrlichiosis and the manifestations of co-infection. *Vet Parasitol*. 2014;199:121–8. <http://dx.doi.org/10.1016/j.vetpar.2013.10.027>
5. Romero LE, Meneses AI, Salazar L, Jiménez M, Romero JJ, Aguiar DM, et al. First isolation and molecular characterization of *Ehrlichia canis* in Costa Rica, Central America. *Res Vet Sci*. 2011;91:95–7. <http://dx.doi.org/10.1016/j.rvsc.2010.07.021>
6. Dawson JE, Stallknecht DE, Howerth EW, Warner C, Biggie K, Davidson WR, et al. Susceptibility of white-tailed deer (*Odocoileus virginianus*) to infection with *Ehrlichia chaffeensis*, the etiologic agent of human ehrlichiosis. *J Clin Microbiol*. 1994;32:2725–8.
7. Kocan AA, Levesque GC, Whitworth LC, Murphy GL, Ewing SA, Barker RW. Naturally occurring *Ehrlichia chaffeensis* infection in coyotes from Oklahoma. *Emerg Infect Dis*. 2000;6:477–80. <http://dx.doi.org/10.3201/eid0605.000505>
8. Paddock CD, Sumner JW, Shore GM, Bartley DC, Elie RC, McQuade JG, et al. Isolation and characterization of *Ehrlichia chaffeensis* strains from patients with fatal ehrlichiosis. *J Clin Microbiol*. 1997;35:2496–502.
9. Breitschwerdt EB, Hegarty BC, Qurollo BA, Saito TB, Maggi RG, Blanton LS, et al. Intravascular persistence of *Anaplasma platys*, *Ehrlichia chaffeensis*, and *Ehrlichia ewingii* DNA in the blood of a dog and two family members. *Parasit Vectors*. 2014;7:298–305. <http://dx.doi.org/10.1186/1756-3305-7-298>

Address for correspondence: Norman Rojas, Centro de Investigación en Enfermedades Tropicales, Facultad de Microbiología, Universidad de Costa Rica, 2060 San Jose, Costa Rica; email: norman.rojas@ucr.ac.cr

Disseminated *Mycobacterium tuberculosis* in Imported Sooty Mangabey, Thailand

Sawang Kesdangakonwut,
Angkana Sommanustweechai,
Angkana Chairasert

Author affiliations: Chulalongkorn University, Bangkok, Thailand (S. Kesdangakonwut); Ministry of Public Health, Muang Nonthaburi, Thailand (A. Sommanustweechai); Mahidol University, Bangkok (A. Chairasert)

DOI: <http://dx.doi.org/10.3201/eid2103.131785>

To the Editor: Tuberculosis caused by bacteria of the *Mycobacterium tuberculosis* complex affects humans and various species of captive and free-living wildlife (1). In addition, *M. tuberculosis* has been used experimentally in many different species of Old World monkeys as part of the attempt to establish a suitable model for human tuberculosis (2). We report a case of disseminated tuberculosis

caused by *M. tuberculosis* Spoligotype International Type (SIT) 52 in a recently imported sooty mangabey (*Cercocebus atys*) from South Africa to Thailand.

A juvenile male sooty mangabey was imported from South Africa to Thailand in September 2009. Within 1 week, while in quarantine, convulsion and salivation developed in the mangabey, and it died suddenly. This animal, along with another mangabey and 4 mustached guenons (*Cercopithecus cephus*), was imported from its native Africa to Thailand for the pet trade. Complete histories of the second mangabey and the mustached guenons were not available.

A complete necropsy of the dead sooty mangabey was conducted, and full histopathologic and microbiological analysis was performed. At necropsy, the mangabey was emaciated, with no subcutaneous and abdominal fat tissues. Disseminated granulomas (up to 2 cm) were observed throughout the carcass, including the lungs, liver, spleen, kidneys, multiple lymph nodes (hilar, mediastinal, mesenteric, splenic, hepatic, renal, and pancreatic), and the ileum. The lung was also multifocally adhered to the thoracic wall and pleural diaphragm.

Histologically, the granulomas in all tissues examined demonstrated similar histopathologic features, characterized by a central core of caseous necrosis and surrounded by an unorganized rim of mixed inflammatory cells, including neutrophils, lymphocytes, plasma cells, and epithelioid macrophages. Numerous acid-fast bacilli were present in the cytoplasm of the epithelioid macrophages and in the necrotic area of all tissues. Acid-fast bacilli were isolated and classified as *M. tuberculosis* on the basis of 1-tube multiplex PCR (3) and sequencing of 16S rRNA gene results. Spoligotyping revealed that the *M. tuberculosis* isolate belonged to SIT 52.

The international wildlife trade had been reported to be a major source of imported zoonoses, particularly tuberculosis, in nonhuman primates (4–8). In the mangabey reported here, fulminant tuberculosis was diagnosed within 1 week after it arrived in Thailand, during the 21-day quarantine period. The granulomas were morphologically similar to the histopathologic description of tuberculosis lesions of experimentally infected cynomolgus macaques (*Macaca fascicularis*), which demonstrated lesions as early as 3 weeks after infection, with a gradual increase in severity (2). Previously, East African–Indian lineage (9) and Beijing spoligotype (SIT 1) accounted for most *M. tuberculosis* isolates in Thailand (10). In nonhuman primates in Thailand, *M. tuberculosis* complex had been detected at rates of up to 50% (5 positive samples from 10 test samples) by PCR from buccal swabs in long-tailed macaque (*Macaca fascicularis*) (1). *M. tuberculosis* belonging to SIT 52 observed in this case has been primarily isolated from countries in Africa (9). Only 1 case of

M. tuberculosis belonging to SIT 52 that caused tuberculous meningitis was reported in a human in Thailand (10), but that case was not related to the case reported here. Our finding of a relatively novel spoligotype of *M. tuberculosis* in an animal destined for the pet trade underscores the need for intensive testing of and extended quarantine for all imported nonhuman primates to prevent the spread of newly isolated *M. tuberculosis* (4,7,8).

Acknowledgments

We thank Areeya Disrattakit and Nampung Makao for their excellent technical assistance and Eric Lombardini and Roongroje Thanawongnuwech for their critical suggestions on this manuscript.

This report was financially supported by Grants for Development of New Faculty Staff, Ratchadaphiseksomphot Endowment Fund, Chulalongkorn University.

The authors declare no potential conflicts of interest with respect to the research, authorship, and/or publication of this article, and no competing financial interests exist.

References

1. Wilbur AK, Engel GA, Rompis A, Putra IG, Lee BP, Aggimarangsee N, et al. From the mouths of monkeys: detection of *Mycobacterium tuberculosis* complex DNA from buccal swabs of synanthropic macaques. *Am J Primatol*. 2012;74:676–86. <http://dx.doi.org/10.1002/ajp.22022>
2. Lin PL, Pawar S, Myers A, Pegu A, Fuhrman C, Reinhart TA, et al. Early events in *Mycobacterium tuberculosis* infection in cynomolgus macaques. *Infect Immun*. 2006;74:3790–803. <http://dx.doi.org/10.1128/IAI.00064-06>
3. Chaiprasert A, Prammananan T, Tingtoy N, Na-Ubol P, Srimuang S, Samerpitak K, et al. One-tube multiplex PCR method for rapid identification of *Mycobacterium tuberculosis*. *Southeast Asian J Trop Med Public Health*. 2006;37:494–502.
4. Panarella ML, Bimes RS. A naturally occurring outbreak of tuberculosis in a group of imported cynomolgus monkeys (*Macaca fascicularis*). *J Am Assoc Lab Anim Sci*. 2010;49:221–5.
5. Centers for Disease Control and Prevention. Tuberculosis in imported nonhuman primates—United States, June 1990–May 1993. *MMWR Morb Mortal Wkly Rep*. 1993;42:572–6.
6. Pavlin BI, Schloegel LM, Daszak P. Risk of importing zoonotic diseases through wildlife trade, United States. *Emerg Infect Dis*. 2009;15:1721–6. <http://dx.doi.org/10.3201/eid1511.090467>
7. Shipley ST, Coksaygan T, Johnson DK, McLeod CG Jr, DeTolla LJ. Diagnosis and prevention of dissemination of tuberculosis in a recently imported rhesus macaque (*Macaca mulatta*). *J Med Primatol*. 2008;37(Suppl 1):20–4. <http://dx.doi.org/10.1111/j.1600-0684.2007.00266.x>
8. Engel GA, Wilbur AK, Westmark A, Horn D, Johnson J, Jones-Engel L. Naturally acquired *Mycobacterium tuberculosis* complex in laboratory pig-tailed macaques. *Emerg Microb Infect*. 2012;1:e30. <http://dx.doi.org/10.1038/emi.2012.31>
9. Brudey K, Driscoll JR, Rigouts L, Prodinger WM, Gori A, Al-Hajj SA, et al. *Mycobacterium tuberculosis* complex genetic diversity: mining the fourth international spoligotyping database (SpolDB4) for classification, population genetics and epidemiology. *BMC Microbiol*. 2006;6:23. <http://dx.doi.org/10.1186/1471-2180-6-23>
10. Yorsangsukkamol J, Chaiprasert A, Prammananan T, Palittapongarnpim P, Limsoontarakul S, Prayoonwiwat N. Molecular analysis of *Mycobacterium tuberculosis* from tuberculous meningitis patients in Thailand. *Tuberculosis (Edinb)*. 2009;89:304–9. <http://dx.doi.org/10.1016/j.tube.2009.05.001>

Address for correspondence: Sawang Kesdangsakonwut, Department of Pathology, Faculty of Veterinary Science, Chulalongkorn University, Henri Dunant Rd, Pathumwan, Bangkok 10330, Thailand; email: sawang.k@chula.ac.th

Treatment of *Mycobacterium abscessus* subsp. *massiliense* Tricuspid Valve Endocarditis

R. Gordon Huth, Elizabeth Douglass, Kristin Mondy, Sruthi Vasireddy, Richard J. Wallace Jr.

Author affiliations: The University of Texas at Austin Dell Medical School Residency Programs, Austin, Texas, USA (R.G. Huth, E. Douglass, K. Mondy); University of Texas Health Science Center, Tyler, Texas, USA (S. Vasireddy, R.J. Wallace Jr.)

DOI: <http://dx.doi.org/10.3201/eid2103.140577>

To the Editor: *Mycobacterium abscessus* is a ubiquitous, rapidly growing mycobacteria (RGM) found in water supplies, soil, and dust. *M. abscessus* is considered the most pathogenic and difficult to treat of the RGM and is most often associated with pulmonary, skin, and soft tissue infections; it has also been reported to cause ocular infections, otitis, lymphadenitis, arthritis, osteomyelitis, disseminated disease, and prosthetic valve endocarditis (1,2). Most prosthetic valve endocarditis cases have been fatal.

M. abscessus subsp. *massiliense* is 1 of 3 subspecies of *M. abscessus*. *M. abscessus* subsp. *massiliense* has an identical 16S rRNA gene sequence to the other 2 subspecies, *Mycobacterium abscessus* subsp. *bolletii* and *Mycobacterium abscessus* subsp. *abscessus*, but can be differentiated by *rpoB* and *erm41* gene sequencing (3,4). *M. abscessus* subsp. *massiliense* grows readily in blood culture media and on sheep's blood agar within 2–4 days. Care should be taken in interpreting Gram staining of isolates because RGM is not identifiable by this method and could be mistaken for corynebacteria or diphtheroids (5,6). Such isolates could be further tested by acid-fast staining and, if positive, sent to a reference laboratory for definitive identification and susceptibility testing.

Five cases of *M. abscessus* native valve endocarditis have been reported; 4 were fatal and 1 was lost to follow-up (1,5–9). One of the 4 fatal cases also involved the tricuspid valve and was associated with intravenous heroin abuse