

## Research Article

# Identification of Novel Vaccine Candidates against *Campylobacter* through Reverse Vaccinology

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Campylobacteriosis is the most prevalent bacterial foodborne gastroenteritis affecting humans in the European Union. Human cases are mainly due to *Campylobacter jejuni* or *Campylobacter coli*, and contamination is associated with the handling and/or consumption of poultry meat. In fact, poultry constitutes the bacteria's main reservoir. A promising way of decreasing the incidence of campylobacteriosis in humans would be to decrease avian colonization. Poultry vaccination is of potential for this purpose. However, despite many studies, there is currently no vaccine available on the market to reduce the intestinal *Campylobacter* load in chickens. It is essential to identify and characterize new vaccine antigens. This study applied the reverse vaccinology approach to detect new vaccine candidates. The main criteria used to select immune proteins were localization, antigenicity, and number of B-epitopes. Fourteen proteins were identified as potential vaccine antigens. *In vitro* and *in vivo* experiments now need to be performed to validate the immune and protective power of these newly identified antigens.

## 1. Introduction

*Campylobacter* is the leading cause of human bacterial gastroenteritis in Europe [1]. It has been estimated that 9 million people are affected each year, costing around €2.4 billion. *C. jejuni* is responsible for approximately 90% of cases, and *C. coli* is responsible for 10%. Other species can cause human campylobacteriosis but are more rarely involved [2]. Most infections are not severe, leading to gastroenteritis symptoms, but they can cause extraintestinal manifestations such as reactive arthritis, Guillain-Barré syndrome (GBS), or inflammatory bowel disease (IBD) [3]. In some cases, infection can even lead to death. Human contaminations are mainly associated with handling and/or consuming poultry meat [1]. Domestic and wild birds constitute the bacteria's main reservoir, carrying up to  $10^9$  CFU·g<sup>-1</sup> of *Campylobacter* intestinally. In poultry flocks, natural colonization occurs in 2- to

3-week-old chicks by horizontal contamination from the environment [4], and birds remain infected until slaughter.

Decreasing avian colonization would appear to be an effective strategy for reducing the incidence of human campylobacteriosis. In 2013, Romero-Barríos et al. estimated that a reduction in *Campylobacter* cecal colonization from 2 to 3 log<sub>10</sub> units could lead to a 100% reduction in the risk of human disease [5]. Along with the implementation of biosecurity, hygiene, and nutritional measures in flocks, poultry vaccination is one way of reducing avian intestine colonization by *Campylobacter* [6]. Several vaccine prototypes have already been tested with variable results. These include whole-cell, subunit, or microorganism-vectored vaccines. Globally, whole-cell vaccines have not been efficient in decreasing *Campylobacter* intestinal loads despite the induction of a specific immune response [7–10]. Among subunit vaccines, flagellin—described as the immunodominant antigen of

*Campylobacter*—has been tested and proved to be able to induce an immune response but this was not necessarily correlated with any decrease in chicken gut colonization [9, 11–13]. Furthermore, because of its weak homology across *Campylobacter* strains, flagellin-based vaccines do not induce cross-protection, making these vaccines inefficient in combatting all *C. jejuni* strains [14]. Other antigens such as CjaA [15]—a periplasmic protein—or CadF, FlpA, CmeC [16], and Dsp proteins [17] involved in *Campylobacter* adherence during colonization have also been trialed as subunit vaccines. In the same way, total outer membrane proteins [18] or fusion proteins [16] have also been tested. Another strategy is to deliver vaccine antigens by vectors such as attenuated bacterial strains. *Salmonella enterica* serovar Typhimurium [15, 19–21] and *Eimeria tenella* [22] have been evaluated as a vector for *C. jejuni* CjaA delivery. For example, in 2004, Wyszynska et al. [19] indicated that chickens orally immunized with a virulent *Salmonella* strain carrying the *Campylobacter* CjaA gene develop a strong specific antibody response, and birds were protected from colonization after a homologous *C. jejuni* challenge. Recently, the same team was unable to confirm these results [21]. Other antigens were tested in the same way, including Omp18/CjaD, ACE393 [20], Dsp [17], Peb1A, GlnH, and ChuA [15]. Some of these experimental studies gave promising results, combining both the induction of a humoral immune response and a decrease in *Campylobacter* intestinal colonization in poultry, but experimentation has not yet been followed up. So, despite much research, no anti-*Campylobacter* vaccine aiming to reduce bacterial colonization in the poultry gut is yet available.

Identifying new potential vaccine antigens is one way of speeding up the development of new vaccines. Reverse vaccinology—a recent approach first described by Rappuoli in the early 2000s [23]—is used to predict antigens through the development of genomics and bioinformatic tools such as genome sequencing. This strategy is different from Hoppe et al. approach, where they identified novel immunodominant proteins by *in vitro* screening of mRNA of *C. jejuni* [24]. The following selection criteria are of particular importance for the reverse vaccinology approach. To be potentially good candidates, the selected proteins must be surface-exposed and able to be recognized by the immune system. Proteins with adhesin capacities are known to be involved in bacterial pathogenicity and invasion, so adhesins or adhesin-like proteins appear as good vaccine targets. The transmembrane helix number is also an important criterion. Indeed, it is difficult to purify proteins with more than one transmembrane helix, and it seems wise to exclude these proteins from the selection process [25]. Individual antigenicity and B-epitope density (the ratio between the number of B-epitopes and the protein length) need to be assessed as described by Oprea's study aimed at developing a vaccine against *S. aureus* endocarditis [26]. Although a few studies are describing innate intestinal inflammations and gut mucosa lesions upon *Campylobacter jejuni* infection (like in [27]), these bacteria are mostly described as a commensal organism for poultry [28]. In the avian intestinal tract, intensive *Campylobacter* multiplication occurs in the mucus layer of the epithelial cells. In this way, antigens need to induce a humoral immune

response to neutralize and eliminate *Campylobacter* from the avian intestinal gut. The induction of a cytotoxic cellular response may not be a selection criterion since *Campylobacter* multiplication in intestinal epithelial cells of chickens was not clearly highlighted [28]. Anyway, bioinformatic tools aiming at predicting T epitopes for avian vaccines are still poorly developed, limiting the reverse vaccinology analysis in this goal. Finally, to provide cross-protection and avoid autoimmune response, it is essential that vaccine candidates are common to many pathogenic strains and do not mimic host proteins [25].

Our research identifies new potential vaccine antigens against *Campylobacter* using the reverse vaccinology strategy to develop an avian vaccine which could impact the incidence of human campylobacteriosis.

## 2. Material and Methods

**2.1. Bacterial Strain.** The highly virulent *Campylobacter jejuni* subsp. *jejuni* 81-176 strain was chosen for this *in silico* analysis. Its genome is available on the NCBI website under accession number CP000538.1 and listed in the Vaxign program used below.

**2.2. OMP and Extracellular Protein Preselection.** Vaxign (<http://www.violinet.org/vaxign/index.php>) was used to shortlist proteins with potential as vaccine candidates due to their cellular localization, probability of having adhesin-like characteristics, and number of transmembrane helices [25]. Vaxign is a web-based pipeline dedicated to vaccine design and integrating several bioinformatic programs. Subcellular localization is predicted using PSORTb2.0 [29]. The probability of adhesin characteristics is predicted by SPAAN software [30] and the transmembrane helix topology is predicted by HMMTOP [31] using a hidden Markov model.

*Campylobacter jejuni* subsp. *jejuni* 81-176 is available in the Vaxign database of over 350 listed genomes, along with nine other *Campylobacter* genomes. Extracellular and outer membrane proteins having an adhesin probability score > 0.51 and either 1 or 0 transmembrane helices were preselected.

**2.3. Protein Antigenicity.** VaxiJen v2.0 (<http://www.ddg-pharmfac.net/vaxijen/VaxiJen/VaxiJen.html>) was used to predict protein antigenicity. This software uses the physicochemical properties of proteins to predict their antigenicity from FASTA-submitted amino acid sequences. This feature is characterized according to an antigenic score. Proteins with an antigenic score > 0.5 were selected as described by Doytchinova and Flower [32].

**2.4. Epitope B Prediction.** BCPreds software (<http://ailab.ist.psu.edu/bcpred/>) was used to identify B-cell epitopes in FASTA-submitted amino acid sequences. This program provides two methods based on different algorithms: the amino acid pair (AAP) antigenicity method [33] and the BCPreds method using string kernels [34]. These methods predict antigenic linear nonoverlapping 20-mer epitopes from the whole antigen. Each preselected protein was analyzed and B-cell epitopes with a score >0.8 were accepted (specificity > 80%).

The selected epitopes were again submitted to VaxiJen software to check their individual antigenicity and those having an antigenic score  $>0.5$  were selected. Furthermore, for each protein and each algorithm, the ratio of B-epitopes to protein length was calculated to assess B-epitope density.

**2.5. BLAST.** In order to assess conservation of the selected proteins in the different *Campylobacter* strains, tblastn analyses were performed for each amino acid sequence against both *C. jejuni* and *C. coli* whole genomes available on the NCBI site on the day of analysis (February 9, 2016): 93 for *C. jejuni* and nine for *C. coli*. The identity percentage was set to 80% and the minimum query coverage was set to 50%. The amount and percentage of sharing among the available genomes were determined. The proteins with a sharing percentage lower than 80% (i.e., about the value for the flagellin) were eliminated from the protein shortlist.

A blastp analysis was also performed to ensure that the host *Gallus gallus* does not express the selected proteins. The identity percentage was set to 50% and the minimum query coverage was set to 50%.

### 3. Results

The reverse vaccinology protocol applied here and results are summarized in Figure 1.

**3.1. Protein Preselection.** The Vaxign server was used to preselect vaccine candidates. Of the 1758 ORFs encoded by the *C. jejuni* 81-176 genome, only 24 were identified as potential vaccine antigens according to the applied criteria (localization, adhesion features, and number of transmembrane helices) (Table 1). Of these 24 identified ORFs, we found the two known flagellins A and B, which means that 22 new potential antigens were selected at this step.

**3.2. Protein Selection according to Antigenicity and Number of B-Epitopes.** To refine the selection, the 22 preselected proteins were submitted to the VaxiJen server for antigenicity prediction. Antigenicity scores ranged from 0.4511 to 0.7827. This step allowed the elimination of two proteins with an antigenicity score lower than 0.5 (YP\_001000503.1 and YP\_001000297.1) (Table 1). The VaxiJen software indicated that all other candidates were antigenic (score  $> 0.5$ ).

Each antigenic protein was assessed in terms of B-epitopes using BCPreds and AAP algorithms, and each B-epitope was studied for its antigenicity. Table 1 summarizes the number of B-epitopes predicted for each protein and each algorithm as well as the ratio between the number of B-epitopes and protein length.

**3.3. Conservation of the Selected Proteins in the Sequenced *C. jejuni* and *C. coli* Strains.** tblastn analyses were performed in order to assess the individual sharing of the preselected proteins among *C. jejuni* and *C. coli* strains. As shown in Table 2, all the proteins were shared with available *C. coli* strains except YP\_001001027.1, which was also poorly shared with the available *C. jejuni* strains (6%). This protein was therefore removed from the list of potential vaccine antigens. Of

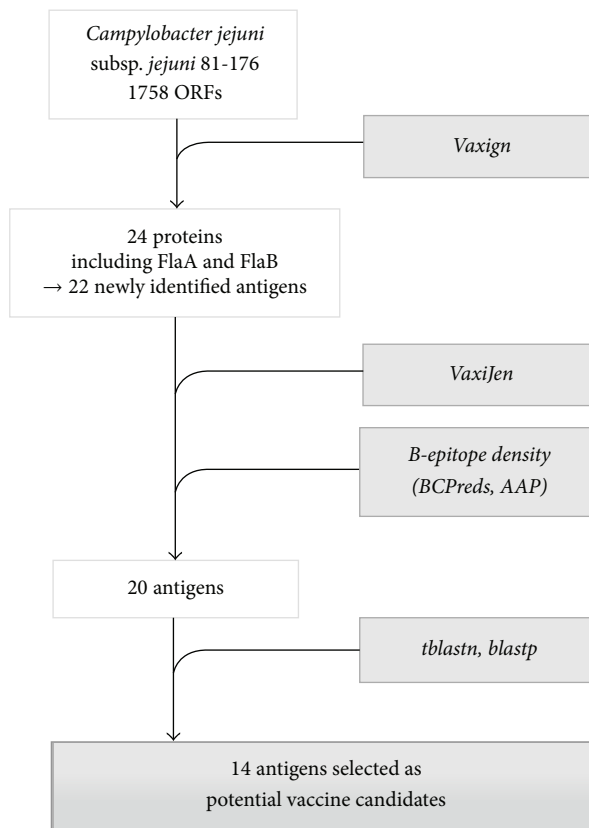


FIGURE 1: Summary of the reverse vaccinology protocol applied to *Campylobacter jejuni* for the selection of vaccine candidates.

the remaining 19 shortlisted proteins, five—YP\_001001371.1, YP\_001000248.1, YP\_001000204.1, YP\_001000654.1 and YP\_001000615.1—were removed from the candidate list because of poor sharing among *C. jejuni* strains ( $<80\%$ ).

Table 2 also shows that none of the proteins are expressed by *Gallus gallus*.

**3.4. Final Selection.** Table 3 shows the final selection of potential vaccine candidates after the whole bioinformatic analysis process. Fourteen candidates were selected. Of these, three are extracellular proteins whereas the others are outer membrane proteins. Four flagellar proteins were identified and several were not characterized and designated as hypothetical proteins.

### 4. Discussion

In the last decades, advances in genomics, genome sequencing, and annotation, coupled with the development of bioinformatic tools has revolutionized vaccine development strategy. Reverse vaccinology allows vaccines to be designed even for noncultivable pathogens; genome availability is the only factor enabling *in silico* analysis or not. All the proteins are targeted even if only transiently expressed or scarce during infection. Furthermore, this strategy considerably reduces the time needed to develop new vaccines [35]. Reverse vaccinology was first successfully applied to the development

TABLE 1: Potential vaccine candidates selected by the Vaxign program. Localization and length were obtained by the Vaxign program, antigenic score was obtained by the Vaxijen program, and the number of B-cell epitopes was obtained from both BCPreds and AAP methods. The calculated ratio between the number of B-epitopes and protein length is also shown. The two candidates eliminated because of a low antigenic score are shown in italics.

Protein accession	Description	ID	Localization	Length (aa)	Vaxijen score	BCPreds B-epitopes		AAP B-epitopes	
						N	Ratio N/length	N	Ratio N/length
YP_001000996.1	Flagellin B	FlaB	Extracellular	576	0.7650	10	0.017	11	0.019
YP_001000997.1	Flagellin A	FlaA	Extracellular	576	0.8185	11	0.019	12	0.021
YP_001001371.1	Flagellar hook protein	FlgE	Extracellular	838	0.7659	16	0.019	18	0.021
YP_001000562.1	Flagellin protein family		Extracellular	750	0.6965	15	0.020	15	0.020
YP_001000248.1	Flagellar capping protein	FlgD	Extracellular	642	0.7021	11	0.017	11	0.017
YP_001000204.1	Putative periplasmic protein		OMP	553	0.6702	11	0.021	11	0.021
YP_001000654.1	Putative periplasmic protein		OMP	553	0.6702	11	0.021	11	0.021
YP_999769.1	Flagellar hook protein		Extracellular	545	0.6567	10	0.018	12	0.022
YP_001001115.1	Flagellar hook-associated protein	FlgE-1	Extracellular	608	0.5836	11	0.018	10	0.016
YP_001000153.1	TonB-dependent receptor, putative, degenerate	FlgK	Extracellular	704	0.5437	12	0.017	9	0.013
YP_001000945.1	N-Acetylmuramoyl-L-alanine amidase		OMP	659	0.6475	9	0.014	11	0.017
YP_001001027.1	Serine protease		OMP	1121	0.5268	9	0.008	11	0.010
YP_001000437.1	Putative OMP		OMP	508	0.6122	9	0.018	6	0.012
YP_999838.1	Hypothetical protein		OMP	400	0.6809	5	0.013	9	0.023
YP_999817.1	Hypothetical protein		OMP	315	0.7827	6	0.019	7	0.022
YP_001000383.1	Flagellar basal body L-ring protein	FlgH	OMP	232	0.6978	6	0.026	4	0.017
YP_001000935.1	Major OMP	PorA	OMP	424	0.6051	5	0.012	5	0.012
YP_001001008.1	Phospholipase A	PldA	OMP	329	0.5819	4	0.012	3	0.009
YP_001001257.1	TonB-dependent heme receptor	ChuA	OMP	702	0.6213	7	0.010	5	0.007
YP_001000615.1	Hypothetical protein		Extracellular	294	0.5498	3	0.010	3	0.010
YP_001000663.1	Surface-exposed lipoprotein	JlpA	OMP	372	0.6642	2	0.005	3	0.008
YP_001000261.1	Hypothetical protein		OMP	309	0.5149	2	0.006	3	0.010
YP_001000503.1	<i>Hypothetical protein</i>		<i>Extracellular</i>	444	<i>0.4603</i>	<i>/</i>	<i>/</i>	<i>/</i>	<i>/</i>
YP_001000297.1	<i>Major antigenic peptide</i>	<i>PEB4</i>	<i>OMP</i>	273	<i>0.4511</i>	<i>/</i>	<i>/</i>	<i>/</i>	<i>/</i>

TABLE 2: Potential vaccine candidates selected after blast analysis. tblastn analyses were performed for each amino acid sequence against both *C. jejuni* and *C. coli* whole genomes available on the NCBI site. The amount and percentage of sharing among the available genomes were determined. A blastp analysis was also performed against the host *Gallus gallus*. The six candidates eliminated because of poor sharing among *Campylobacter* strains are shown in italics.

Protein accession	Description	ID	Sharing among <i>C. jejuni</i> strains N/93	%	Sharing among <i>C. coli</i> strains N/9	%	Similarity in <i>Gallus gallus</i>
YP_001000996.1	Flagellin B	FlaB	77	83	9	100	No
YP_001000997.1	Flagellin A	FlaA	75	81	9	100	No
YP_001001371.1	<i>Flagellar hook protein</i>	<i>FlgE</i>	15	16	8	89	No
YP_001000562.1	Flagellin protein family		93	100	9	100	No
YP_001000248.1	<i>Flagellar capping protein</i>	<i>FlhD</i>	38	41	9	100	No
YP_001000204.1	<i>Putative periplasmic protein</i>		2	2	9	100	No
YP_001000654.1	<i>Putative periplasmic protein</i>		2	2	9	100	No
YP_999769.1	Flagellar hook protein	FlgE-1	93	100	9	100	No
YP_001001115.1	Flagellar hook-associated protein	FlgK	93	100	9	100	No
YP_001000153.1	TonB-dependent receptor; putative, degenerate		90	97	9	100	No
YP_001000945.1	N-Acetylmuramoyl-L-alanine amidase		93	100	9	100	No
YP_001001027.1	<i>Serine protease</i>	<i>PEB4</i>	6	6	1	11	No
YP_001000437.1	Putative OMP		89	96	6	67	No
YP_999838.1	Hypothetical protein		93	100	9	100	No
YP_999817.1	Hypothetical protein		92	99	9	100	No
YP_001000383.1	Flagellar basal body L-ring protein	FlgH	93	100	9	100	No
YP_001000935.1	Major OMP	PorA	81	87	9	100	No
YP_001001008.1	Phospholipase A	PldA	92	99	9	100	No
YP_001001257.1	TonB-dependent heme receptor	ChuA	93	100	9	100	No
YP_001000615.1	<i>Hypothetical protein</i>		64	69	9	100	No
YP_001000663.1	Surface-exposed lipoprotein	JlpA	93	100	9	100	No
YP_001000261.1	Hypothetical protein		92	99	9	100	No

TABLE 3: Potential vaccine candidates selected after the whole bioinformatic analysis process including Vaxign and VaxiJen programs, BCPreds and AAP algorithms, and blast analyses. Of 1758 ORFs encoded by *C. jejuni*, strain 81-176 genome, 14 proteins were selected as vaccine candidates.

Protein accession	Description	Localization	ID
YP_001000562.1	Flagellin protein family	Extracellular	
YP_999769.1	Flagellar hook protein	Extracellular	FlgE-1
YP_001001115.1	Flagellar hook-associated protein	Extracellular	FlgK
YP_001000153.1	TonB-dependent receptor, putative, degenerate	OMP	
YP_001000945.1	N-Acetylmuramoyl-L-alanine amidase	OMP	
YP_001000437.1	Putative OMP	OMP	
YP_999838.1	Hypothetical protein	OMP	
YP_999817.1	Hypothetical protein	OMP	
YP_001000383.1	Flagellar basal body L-ring protein	OMP	FlgH
YP_001000935.1	Major OMP	OMP	PorA
YP_001001008.1	Phospholipase A	OMP	PldA
YP_001001257.1	TonB-dependent heme receptor	OMP	ChuA
YP_001000663.1	Surface-exposed lipoprotein	OMP	JlpA
YP_001000261.1	Hypothetical protein	OMP	

of a vaccine against B serogroup *Neisseria meningitidis* [36]. Despite available prophylactic vaccines based on capsular polysaccharides (CPS) for four *N. meningitidis* serogroups (A, C, W, and Y), the development of a capsular vaccine against serogroup B was not possible because of CPS mimicry of polysialic acid in human cells. *In silico* analysis identified three proteins (fHbp, NadA, and NHBA) which were combined with outer membrane vesicles containing known antigen PorA and led to the European licensure of the 4CMenB vaccine in 2013 [37]. This strategy was then applied to several other pathogens such as herpes simplex viruses using the Vaxign program [38], *Staphylococcus aureus* for the *in silico* characterization of ten surface-exposed proteins [26], *Mycobacterium tuberculosis* with the identification of six novel antigen candidates to improve the tuberculosis vaccine [39], or *Streptococcus pneumoniae* with the bioinformatic assessment of 13 protein targets [40]. The antigenicity and efficiency of the potential candidates selected in these last *in silico* studies have not yet been tested *in vitro* or *in vivo*.

Until now, and despite many studies, conventional development of a vaccine against *Campylobacter* in poultry has not led to an efficient vaccine in terms of immunogenicity and protection. Since 2005, *Campylobacter* has been and remains today the leading cause of bacterial foodborne gastroenteritis in Europe [1]. As poultry vaccination is one of the potential ways of reducing the incidence of human campylobacteriosis, it is important to pursue efforts to test new vaccine antigens. Reverse vaccinology is a suitable strategy to this end.

This *in silico* study predictively identified new vaccine antigens against *Campylobacter*. The reference *C. jejuni* ATCC (American Type Culture Collection) strain 81-176 was chosen for antigen prediction because of its high virulence in human diseases (namely, the chicken vaccine strategy is to prevent human infections). Even if this strain is not a good colonizer for chickens, this strain has been successfully used in several poultry experiments with high colonization levels [18]. Moreover, the reverse vaccinology aims to identify

shared proteins among many *Campylobacter* strains (here more than 100 strains). Thereby, other more avian colonizer *Campylobacter* strains should be used for *in vivo* challenge experiments to evaluate the effectiveness of the proteins found by the bioinformatics analysis of the *C. jejuni* 81-176 genome.

Based on their cellular localization, adhesin-like properties, antigenicity, B-epitope density, and conservation among *Campylobacter* strains, 14 proteins were selected. It was decided to eliminate proteins with a sharing percentage lower than the flagellin sharing percentage. It has already been observed that flagellin could not be used as a vaccine candidate because of poor sharing among *Campylobacter* strains and the lack of cross-protection [14]. The known vaccine antigens of flagellins A and B were also identified alongside potential antigens using the same criteria. This strengthens the validity of the bioinformatic protocol used, because the flagellin has already been described and used as the immune-dominant antigen of *Campylobacter* [9, 11, 12]. However, it is important to keep in mind that the identified proteins were selected on the basis of predictions by various algorithms. Only *in vitro* and more *in vivo* experiments will confirm or refute the proteins' immune power. In terms of antigen ranking, proteins with a high antigenicity score and B-epitope density seem to be the best vaccine candidates and should therefore be evaluated for *in vivo* immunogenicity as a priority. Indeed, it has already been demonstrated that a high epitope density significantly enhances antigenicity and immunogenicity [41]. This strategy, being based on genome analysis, does not take into consideration lipid and saccharide antigens, which could also have immune properties. Concerning *Campylobacter*, capsule polysaccharides are not targeted through the reverse vaccinology protocol, although they could be immunogenic [42].

Several of the identified proteins had already been characterized and were mainly associated with *Campylobacter* virulence [43]. This is the case for three selected flagellar

proteins—FlgE-1, FlgK, and FlgH—involved in *Campylobacter* motility, essential for bacteria survival in the gastrointestinal tract. These proteins were recently tested *in vitro* along with other flagellar proteins [44]. The first two were immunostained by more than 70% of tested sera from chickens older than 5-6 weeks; the third one was immunostained by 50% of the tested sera. The present *in silico* analysis is in line with these *in vitro* results, leading us to consider these three flagellar proteins as a potential vaccine antigen. However, no *in vivo* assessment is available yet. The FliD flagellar protein was similarly tested *in vitro* and was observed to react strongly to sera from chickens over 4 weeks of age [45]. In the present analysis, this flagellar protein was eliminated from the shortlist because of poor sharing with other *Campylobacter* strains (41%). Moreover, the ChuA protein—involved in the iron uptake system—had already been tested in an avian vaccine experiment using attenuated *Salmonella* as a vector [15] and did not significantly reduce cecal *Campylobacter* counts. Furthermore, the major PorA outer membrane protein was tested *in vivo* in a mouse model [46]. Mice vaccinations led to significantly higher antibody levels in serum and intestinal lavage fluids. A decrease in *C. jejuni* colonization levels was also observed after a heterologous challenge. Phospholipase A (PldA) and lipoprotein JlpA are involved in *Campylobacter* adhesion since it has been demonstrated that mutations of *pldA* impair the ability of *C. jejuni* to colonize cecum [47] and since Jin et al. highlighted the interaction of JlpA with a surface-exposed protein of epithelial cells [48].

To conclude, reverse vaccinology—a powerful tool for identifying new vaccine antigens—allowed 14 candidates to be selected for the development of a vaccine against *Campylobacter* in poultry. Several antigens identified as potential vaccine candidates are currently under *in vitro* and *in vivo* investigations to evaluate their immunogenicities and protective potentials against *Campylobacter* in chickens.

## Competing Interests

The authors declare that there is no conflict of interests regarding publication of this paper.

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## References

- [1] EFSA, "The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2013," *EFSA Journal*, vol. 13, no. 1, p. 3991, 2015.
- [2] I. A. Gillespie, S. J. O'Brien, J. A. Frost et al., "A case-case comparison of *Campylobacter coli* and *Campylobacter jejuni* infection: a tool for generating hypotheses," *Emerging Infectious Diseases*, vol. 8, no. 9, pp. 937–942, 2002.
- [3] R. Janssen, K. A. Krogfelt, S. A. Cawthraw, W. van Pelt, J. A. Wagenaar, and R. J. Owen, "Host-pathogen interactions in *Campylobacter* infections: the host perspective," *Clinical Microbiology Reviews*, vol. 21, no. 3, pp. 505–518, 2008.
- [4] O. Sahin, N. Luo, S. Huang, and Q. Zhang, "Effect of *Campylobacter*-specific maternal antibodies on *Campylobacter jejuni* colonization in young chickens," *Applied and Environmental Microbiology*, vol. 69, no. 9, pp. 5372–5379, 2003.
- [5] P. Romero-Barrios, M. Hempen, W. Messens, P. Stella, and M. Hugas, "Quantitative microbiological risk assessment (QMRA) of food-borne zoonoses at the European level," *Food Control*, vol. 29, no. 2, pp. 343–349, 2013.
- [6] M. Meunier, M. Guyard-Nicodème, D. Dory, and M. Chemaly, "Control strategies against *Campylobacter* at the poultry production level: biosecurity measures, feed additives and vaccination," *Journal of Applied Microbiology*, vol. 120, no. 5, pp. 1139–1173, 2016.
- [7] B. E. Rice, D. M. Rollins, E. T. Mallinson, L. Carr, and S. W. Joseph, "Campylobacter *jejuni* in broiler chickens colonization and humoral immunity following oral vaccination and experimental infection," *Vaccine*, vol. 15, no. 17-18, pp. 1922–1932, 1997.
- [8] G. Glünder, N. Spiering, and K. Hinz, "Investigations on parental immunization of chickens with a *Campylobacter* mineral oil vaccine," in *Proceedings of the International Congress of the World Veterinary Poultry Association*, B. Nagy and R. Mulder, Eds., pp. 247–253, Budapest, Hungary, 1997.
- [9] P. R. Widders, L. M. Thomas, K. A. Long, M. A. Tokhi, M. Panaccio, and E. Apos, "The specificity of antibody in chickens immunised to reduce intestinal colonisation with *Campylobacter jejuni*," *Veterinary Microbiology*, vol. 64, no. 1, pp. 39–50, 1998.
- [10] R. L. Ziprin, M. E. Hume, C. R. Young, and R. B. Harvey, "Inoculation of chicks with viable non-colonizing strains of *Campylobacter jejuni*: evaluation of protection against a colonizing strain," *Current Microbiology*, vol. 44, no. 3, pp. 221–223, 2002.
- [11] C. A. Khoury and R. J. Meinersmann, "A genetic hybrid of the *Campylobacter jejuni* *flaA* Gene with LT-B of *Escherichia coli* and assessment of the efficacy of the hybrid protein as an oral chicken vaccine," *Avian Diseases*, vol. 39, no. 4, pp. 812–820, 1995.
- [12] J.-L. Huang, Y.-X. Yin, Z.-M. Pan et al., "Intranasal immunization with chitosan/pCAGGS-*flaA* nanoparticles inhibits *Campylobacter jejuni* in a White Leghorn model," *Journal of Biomedicine and Biotechnology*, vol. 2010, Article ID 589476, 8 pages, 2010.
- [13] M. Meunier, M. Guyard-Nicodème, E. Vigouroux, T. Poezevara, and V. Beven, *Sequential Optimization of an Avian Vaccine Protocol Against Campylobacter*, 2015.
- [14] M. R. de Zoete, J. P. M. van Putten, and J. A. Wagenaar, "Vaccination of chickens against *Campylobacter*," *Vaccine*, vol. 25, no. 30, pp. 5548–5557, 2007.
- [15] A. M. Buckley, J. Wang, D. L. Hudson et al., "Evaluation of live-attenuated *Salmonella* vaccines expressing *Campylobacter* antigens for control of *C. jejuni* in poultry," *Vaccine*, vol. 28, no. 4, pp. 1094–1105, 2010.
- [16] J. M. Neal-McKinney, D. R. Samuelson, T. P. Eucker, M. S. Nissen, R. Crespo, and M. E. Konkel, "Reducing *Campylobacter jejuni* colonization of poultry via vaccination," *PLoS ONE*, vol. 9, no. 12, Article ID e114254, 2014.
- [17] J. R. Theoret, K. K. Cooper, B. Zekarias et al., "The *Campylobacter jejuni* *dps* homologue is important for *In vitro* biofilm

- formation and cecal colonization of poultry and may serve as a protective antigen for vaccination,” *Clinical and Vaccine Immunology*, vol. 19, no. 9, pp. 1426–1431, 2012.
- [18] T. Annamalai, R. Pina-Mimbela, A. Kumar et al., “Evaluation of nanoparticle-encapsulated outer membrane proteins for the control of *Campylobacter jejuni* colonization in chickens,” *Poultry Science*, vol. 92, no. 8, pp. 2201–2211, 2013.
- [19] A. Wysznińska, A. Raczko, M. Lis, and E. K. Jagusztyn-Krynicka, “Oral immunization of chickens with avirulent *Salmonella* vaccine strain carrying *C. jejuni* 72Dz/92 *cjaA* gene elicits specific humoral immune response associated with protection against challenge with wild-type *Campylobacter*,” *Vaccine*, vol. 22, no. 11–12, pp. 1379–1389, 2004.
- [20] S. L. Layton, M. J. Morgan, K. Cole et al., “Evaluation of *Salmonella*-vectored *Campylobacter* peptide epitopes for reduction of *Campylobacter jejuni* in broiler chickens,” *Clinical and Vaccine Immunology*, vol. 18, no. 3, pp. 449–454, 2011.
- [21] P. Laniewski, M. Kuczowski, K. Chrzastek et al., “Evaluation of the immunogenicity of *Campylobacter jejuni* CjaA protein delivered by *Salmonella enterica* sv. Typhimurium strain with regulated delayed attenuation in chickens,” *World Journal of Microbiology and Biotechnology*, vol. 30, no. 1, pp. 281–292, 2014.
- [22] J. D. Clark, R. D. Oakes, K. Redhead et al., “*Eimeria* species parasites as novel vaccine delivery vectors: anti-*Campylobacter jejuni* protective immunity induced by *Eimeria tenella*-delivered CjaA,” *Vaccine*, vol. 30, no. 16, pp. 2683–2688, 2012.
- [23] R. Rappuoli, “Reverse vaccinology,” *Current Opinion in Microbiology*, vol. 3, no. 5, pp. 445–450, 2000.
- [24] S. Hoppe, F. F. Bier, and M. von Nickisch-Roseneck, “Rapid identification of novel immunodominant proteins and characterization of a specific linear epitope of *Campylobacter jejuni*,” *PLoS ONE*, vol. 8, no. 5, Article ID e65837, 2013.
- [25] Y. He, Z. Xiang, and H. L. Mobley, “Vaxign: the first web-based vaccine design program for reverse vaccinology and applications for vaccine development,” *Journal of Biomedicine and Biotechnology*, vol. 2010, Article ID 297505, 15 pages, 2010.
- [26] M. Oprea and F. Antohe, “Reverse-vaccinology strategy for designing T-cell epitope candidates for *Staphylococcus aureus* endocarditis vaccine,” *Biologicals*, vol. 41, no. 3, pp. 148–153, 2013.
- [27] S. Humphrey, G. Chaloner, K. Kemmett et al., “*Campylobacter jejuni* is not merely a commensal in commercial broiler chickens and affects bird welfare,” *mBio*, vol. 5, no. 4, Article ID e01364–14, 2014.
- [28] K. Van Deun, F. Pasmans, R. Ducatelle et al., “Colonization strategy of *Campylobacter jejuni* results in persistent infection of the chicken gut,” *Veterinary Microbiology*, vol. 130, no. 3–4, pp. 285–297, 2008.
- [29] J. L. Gardy, M. R. Laird, F. Chen et al., “PSORTb v2.0: expanded prediction of bacterial protein subcellular localization and insights gained from comparative proteome analysis,” *Bioinformatics*, vol. 21, no. 5, pp. 617–623, 2005.
- [30] G. Sachdeva, K. Kumar, P. Jain, and S. Ramachandran, “SPAAN: a software program for prediction of adhesins and adhesin-like proteins using neural networks,” *Bioinformatics*, vol. 21, no. 4, pp. 483–491, 2005.
- [31] L. Käll, A. Krogh, and E. L. L. Sonnhammer, “Advantages of combined transmembrane topology and signal peptide prediction—the Phobius web server,” *Nucleic Acids Research*, vol. 35, no. 2, pp. W429–W432, 2007.
- [32] I. A. Doytchinova and D. R. Flower, “VaxiJen: a server for prediction of protective antigens, tumour antigens and subunit vaccines,” *BMC Bioinformatics*, vol. 8, article 4, 2007.
- [33] J. Chen, H. Liu, J. Yang, and K.-C. Chou, “Prediction of linear B-cell epitopes using amino acid pair antigenicity scale,” *Amino Acids*, vol. 33, no. 3, pp. 423–428, 2007.
- [34] Y. El-Manzalawy, D. Dobbs, and V. Honavar, “Predicting linear B-cell epitopes using string kernels,” *Journal of Molecular Recognition*, vol. 21, no. 4, pp. 243–255, 2008.
- [35] A. Sette and R. Rappuoli, “Reverse vaccinology: developing vaccines in the era of genomics,” *Immunity*, vol. 33, no. 4, pp. 530–541, 2010.
- [36] M. Pizza, V. Scarlato, V. Masignani et al., “Identification of vaccine candidates against serogroup B meningococcus by whole-genome sequencing,” *Science*, vol. 287, no. 5459, pp. 1816–1820, 2000.
- [37] S. M. Andrews and A. J. Pollard, “A vaccine against serogroup B *Neisseria meningitidis*: dealing with uncertainty,” *The Lancet Infectious Diseases*, vol. 14, no. 5, pp. 426–434, 2014.
- [38] Z. Xiang and Y. He, “Genome-wide prediction of vaccine targets for human herpes simplex viruses using Vaxign reverse vaccinology,” *BMC Bioinformatics*, vol. 14, supplement 4, article S2, 2013.
- [39] G. P. Monterrubio-López, J. A. González-Y-Merchand, and R. M. Ribas-Aparicio, “Identification of novel potential vaccine candidates against tuberculosis based on reverse vaccinology,” *BioMed Research International*, vol. 2015, Article ID 483150, 16 pages, 2015.
- [40] A. P. C. Argondizzo, F. F. da Mota, C. P. Pestana et al., “Identification of proteins in *Streptococcus pneumoniae* by reverse vaccinology and genetic diversity of these proteins in clinical isolates,” *Applied Biochemistry and Biotechnology*, vol. 175, no. 4, pp. 2124–2165, 2015.
- [41] W. Liu and Y.-H. Chen, “High epitope density in a single protein molecule significantly enhances antigenicity as well as immunogenicity: a novel strategy for modern vaccine development and a preliminary investigation about B cell discrimination of monomeric proteins,” *European Journal of Immunology*, vol. 35, no. 2, pp. 505–514, 2005.
- [42] P. Guerry, F. Poly, M. Riddle, A. C. Maue, Y.-H. Chen, and M. A. Monteiro, “*Campylobacter* polysaccharide capsules: virulence and vaccines,” *Frontiers in Cellular and Infection Microbiology*, vol. 2, p. 7, 2012.
- [43] D. J. Bolton, “*Campylobacter* virulence and survival factors,” *Food Microbiology*, vol. 48, pp. 99–108, 2015.
- [44] H.-Y. Yeh, K. L. Hiatt, and J. E. Line, “Reactions of chicken sera to recombinant *Campylobacter jejuni* flagellar proteins,” *Archives of Microbiology*, vol. 197, no. 2, pp. 353–358, 2015.
- [45] H.-Y. Yeh, K. L. Hiatt, J. E. Line, and B. S. Seal, “Characterization and antigenicity of recombinant *campylobacter jejuni* flagellar capping protein FliD,” *Journal of Medical Microbiology*, vol. 63, no. 4, pp. 602–609, 2014.
- [46] A. Islam, R. Raghupathy, and M. J. Albert, “Recombinant porA, the major outer membrane protein of *Campylobacter jejuni*, provides heterologous protection in an adult mouse intestinal colonization model,” *Clinical and Vaccine Immunology*, vol. 17, no. 11, pp. 1666–1671, 2010.



- [47] R. L. Ziprin, C. R. Young, J. A. Byrd et al., "Role of *Campylobacter jejuni* potential virulence genes in cecal colonization," *Avian Diseases*, vol. 45, no. 3, pp. 549–557, 2001.
- [48] S. Jin, Y. C. Song, A. Emili, P. M. Sherman, and V. L. Chan, "JlpA of *Campylobacter jejuni* interacts with surface-exposed heat shock protein 90 $\alpha$  and triggers signalling pathways leading to the activation of NF- $\kappa$ B and p38 MAP kinase in epithelial cells," *Cellular Microbiology*, vol. 5, no. 3, pp. 165–174, 2003.



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