

## Research Article

# A Covalently Imprinted Photonic Crystal for Glucose Sensing

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We demonstrate a glucose-sensing material based on the combination of photonic crystal templating and a molecular-imprinting technique. In the presence of the target molecule glucose, poly(*N*-isopropylacrylamide-co-2-hydroxyethyl methacrylate) hydrogel with pendent phenylboronic acid groups was synthesized in the void of a poly(methyl methacrylate) (PMMA) colloidal crystal. After removing the PMMA photonic crystal template and glucose molecules, a 3D-ordered porous covalently imprinted photonic crystal (CIPC) hydrogel was created. The unique 3D-ordered porous hydrogel revealed optical changes in response to glucose concentration. At pH = 11 and 37°C, the diffraction of CIPC redshifted from 725 nm to 880 nm in response to 20 mmol L<sup>-1</sup> glucose. Due to the covalently imprinted recognizer, boronic acid group, the selectivity of the CIPC towards glucose over *D*-ribose and *L*-rhamnose was improved significantly.

## 1. Introduction

The prevalence of type 1 diabetes continues to increase worldwide. Real-time continuous glucose monitoring (RT-CGM) is crucial for diabetic care, which can lead to better treatment decisions, improve metabolic control, and decrease the incidence and progression of diabetic complications [1–6]. For this reason, many different analytical methods for RT-CGM have been developed, which are mainly electrochemical [7–9] and optical [3, 10, 11]. Photonic crystals are highly ordered materials with periods on the scale of visible light and exhibit unique structural color on the basis of Bragg diffraction and lattice spacing. It is found that the diffraction wavelength of photonic crystals is dependent on their lattices. Various approaches have been explored for making bioresponsive photonic crystal hydrogels for glucose sensing. Honda et al. developed a totally synthetic colorimetric glucose-sensing system composed of glucose-responsive hydrogel particles in an inverse opal polymer membrane [12]. The volume of the hydrogel particles can change reversibly as the glucose concentration varies and exhibits a reversible change in the color and the peak intensity of the reflection spectra. Asher's group has also made remarkable contributions to glucose-responsive photonic crystals. They incorporated a crystalline colloidal array (CCA) into a polyacrylamide [13]

or poly(vinyl alcohol) [14] hydrogel with pendent boronic acid groups, and the polymerized crystalline colloidal array (PCCA) diffraction wavelength was dependent on the hydrogel volume [15]. Bazin and Zhu prepared core-shell microspheres and assembled them into a crystalline colloidal array (CCA) [3]. The core-shell microspheres were functionalized with 3-aminophenylboronic acid to make them responsive to glucose. More recently, Ayyub et al. investigated one-dimensional photonic crystals as a sensitive carbohydrate sensor by modifying polystyrene-*b*-poly(2-vinyl pyridine) films with 2-(bromomethyl)phenylboronic acid [11]. Besides boronic acid, glucose oxidase (GOx) was also attached to the PCCA and glucose caused the PCCA to swell and redshift the diffraction wavelength [16]. Although photonic crystals now attract more and more attention, their selectivity is not promising. Immobilization of enzymes might improve the selectivity, but natural receptors usually deteriorate quickly due to the variation of temperature, pH, humidity, and toxic chemicals.

Molecular imprinting is a state-of-the-art technique to artificially mimic natural and biological receptors [17, 18]. Molecularly imprinted polymers (MIPs) have several advantages, including stability under harsh physical and chemical conditions, easy preparation, and more importantly the ability to be “tailored” recognition materials for the target

analytes. MIPs have attracted considerable attention as a recognition element with high selectivity for glucose sensors. Chen et al. reported an MIP that could be used to measure glucose concentrations in complex biological media. At alkaline pH, the metal-complexing polymer “catches” glucose and instantly releases protons in proportion to the glucose concentration over a clinically relevant range (0 to 25 mmol L<sup>-1</sup>) [19]. Malitesta et al. electropolymerized a glucose-imprinted poly(*O*-phenylenediamine) polymer, which was employed as the recognition element of a QCM biomimetic sensor for glucose [20]. Wu’s group designed a kind of molecularly imprinted hybrid microgels that can optically monitor glucose levels with high sensitivity and selectivity in complex media at physiological pH [21]. Recently, Li’s group coupled molecular imprinting with photonic crystal to create self-reporting specific sensors to detect protein, medicines and some biomarkers [22–25]. However, specificity cannot be guaranteed by traditional noncovalent imprinting protocol, and in most cases, the diffraction shifts are not remarkable. We here report on a covalently imprinted photonic crystal (CIPC) for glucose sensing. The use of boronic acid derivatives to sense diol-containing substances has been exploited for a long time. Boronic acid can bind cis diol-containing substances through covalent bonds. Our CIPC glucose-sensing hydrogel also utilized boronic acid groups as recognition elements. In order to improve the selectivity of photonic crystal sensor, covalently imprinting was introduced into the photonic crystal hydrogel. Boronic acid derivatives have been polymerized in the presence of glucose to create imprinted glucose-binding sites. Polymer hydrogel volume changes could be actuated by boronic acid-glucose complexation. The CIPC volume changes alter the photonic crystal spacing, which alters the Bragg diffraction condition.

## 2. Materials and Methods

**2.1. Materials.** Acrylamide (AM), acrylic acid (AA), 2,2'-azobisisobutyronitrile (ABVN), 2-hydroxyethyl methacrylate (HEMA), *N*-isopropylacrylamide (NIPA), 2,2-diethoxyacetophenone (DEAP), 4-ethylene phenylboronic acid (4-EPBA), and *N,N'*-methylenebisacrylamide (BIS) were all purchased from Tokyo Chemical Industry (Tokyo, Japan). Methyl methacrylate (MMA), glycine, glucose, *D*-ribose, *L*-rhamnose, acetonitrile, dimethyl sulfoxide (DMSO) and acetonitrile were all from local suppliers and were used directly.

**2.2. Instruments.** The diffraction spectra were detected using an Avaspec-2048TEC optical fiber spectrometer (Avantes, Netherlands). UV polymerization was carried out in a SCIENTZ D3-IIUV cross-linker (Ning Bo, China). The assembling of PMMA colloidal crystal was carried out in a Safe HWS-150 incubator (Ning Bo, China). SEM images were taken using a HITACHI S4800 field emission scanning electron microscope (Tokyo, Japan) with an accelerating voltage of 15 kV. A Shimadzu LC-20AT HPLC system on a Promosil silica column (4.6 × 250 mm, Agela Technologies, Tianjing, China) with an evaporative light-scattering detector

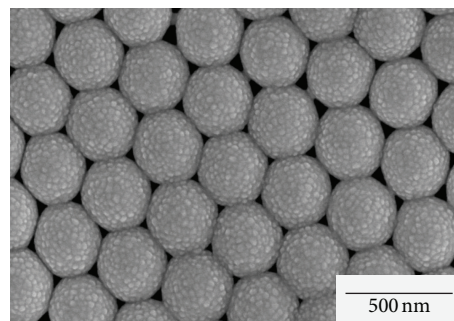


FIGURE 1: SEM image of the PMMA colloidal crystal.

(ELSD) was used to quantitatively characterize the imprinting effect of MIPs. The mobile phase was acetonitrile/water (2/1, v/v) with a flow rate of 1 mL min<sup>-1</sup>, and the injection volume was 20 μL.

**2.3. Preparation of PMMA Colloidal Crystal.** Poly(methyl methacrylate) (PMMA) monodispersed colloidal particles (280 nm) were synthesized using a modification of published methods [26, 27]. MMA (30 mL) and deionized water (250 mL) were mixed in a 4-neck flask at 80°C, stirred at 300 r min<sup>-1</sup>, and bubbled with nitrogen. After equilibrating to 80°C, 15 mL water containing 0.6 g potassium persulfate was added to the flask to initiate polymerization. Monodisperse PMMA particles with a diameter of approximately 280 nm were obtained after 40 minutes (Figure 1). The resulting particles were washed by centrifugation and rinsed at least three times with deionized water.

The PMMA colloids were self-assembled into colloidal crystals via a vertical deposition method as a template of the opal structure. Typically, glass slides were immersed in H<sub>2</sub>SO<sub>4</sub>/H<sub>2</sub>O<sub>2</sub> mixture (7:3, V/V) for 12 h, then rinsed with deionized water in ultrasonic bath three times, and dried before use. Mono-disperse PMMA particles were fully dispersed in deionized water (0.25%–0.35%, w/w), which were added to a clean Petri dish for the colloidal crystal formation. Clean glass slides were placed vertically into the Petri dish for colloidal crystal growth. After complete volatilization of the deionized water, a colloidal crystal of PMMA particles was formed on both sides of each glass slide.

**2.4. Preparation of CIPC Films.** Three MIPs pre-polymerization solutions, each with different recipes (see Table 1) were prepared. Take MIPs recipe (B), for instance, glucose (0.17 mmol), covalent functional monomer 4-APBA (0.17 mmol), backbone monomers HEMA (1.5 mmol), and NIPA (1.5 mmol) were dissolved into a mixture of 1.5 mL DMSO. Then, crosslinker BIS (0.254 mmol) and initiator ABVN (12.68 μmol) were added and the mixture was fully deoxygenated by nitrogen purging for 10 min. The prepolymerization solution was layered carefully on the PMMA photonic crystal template so that it can infiltrate into the CCA and then polymerize at 60°C to form hydrogel film. The film was immersed into acetone to remove the PMMA photonic crystal template, followed by the removal of glucose by acidic

TABLE 1: Composition of different MIPs prepolymerization solutions.

MIPs	Template (mmol)	Monomers (mmol)	Crosslinker (mmol)	Initiator (mmol)	Solvent (mL)	Polymerization
A	Glucose (0.7)	4-EPBA (0.7) + AM (1.4) + HEMA (1.4)	BIS (0.0162)	DEAP (0.05)	Acetonitrile (2)	UV irradiation 4°C
B	Glucose (0.17)	4-EPBA (0.17) + NIPA (1.5) + HEMA (1.5)	BIS (0.254)	ABVN (0.01)	DMSO (1)	60°C
C	Glucose (0.17)	4-EPBA (0.17) + HEMA (1.5)	BIS (0.317)	DEAP (0.5)	Acetonitrile (3)	UV irradiation 4°C

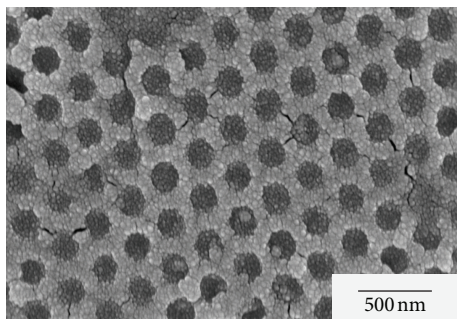


FIGURE 2: SEM image of the macroporous structure of the CIPC film.

solution to give the CIPC hydrogel. A nonimprinted photonic crystal (NIPC) was prepared in the same procedure in the absence of glucose.

The three CIPC films were dried in a vacuum oven over night and then were exposed to glucose to evaluate their adsorption capability and selectivity. Typically, glucose solutions ( $20 \text{ mmol L}^{-1}$ ), prepared in 1 mL methanol/deionized water (6:4, v/v), were incubated with 10 mg CIPC film for 1 h at room temperature. After incubation, the residual concentration of glucose in the solution was determined by HPLC.

### 3. Results and Discussion

Figure 1 shows the SEM images of the used PMMA colloidal crystal, in which the PMMA particle size is about 280 nm. From Figure 1, we can see that the PMMA colloidal crystal has high ordering. After removing the PMMA photonic crystal template, highly ordered macroporous structure could be observed using SEM (Figure 2). More importantly, the ordering of the macroporous structure was kept well. In order to optimize the recipe of covalently imprinted molecules, we tried three different recipes. The adsorption rates of the three kinds of MIPs in  $20 \text{ mmol L}^{-1}$  glucose were measured via HPLC. It was revealed in Figure 3 that even though recipe MIPs(A) and MIPs(C) had better adsorption rates, recipe MIPs(B) had better imprinting effect among the three MIPs. As a result, we chose recipe MIPs(B) to study the following absorption and sensing properties of the CIPC.

Boronic acid is a classic saccharide receptor [28, 29], which can form reversible covalent bonds with glucose. Herein, 4-EPBA which contains a boronic acid functional

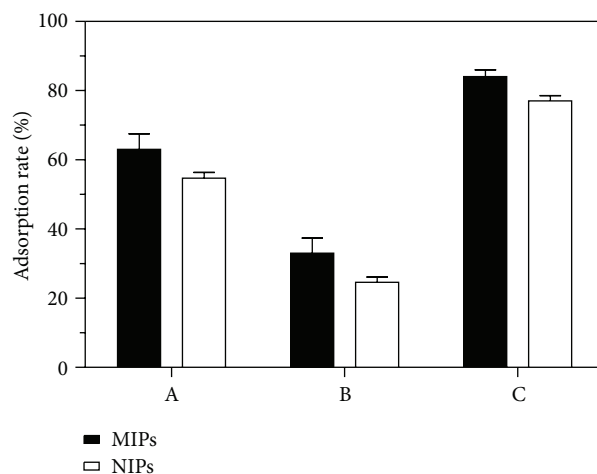
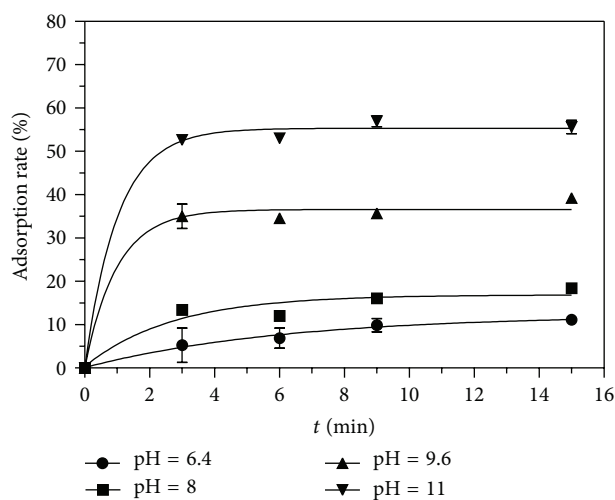


FIGURE 3: Adsorptivity of the MIPs of different recipes.

FIGURE 4: Adsorptivity of the CIPC towards  $10 \text{ mmol L}^{-1}$  glucose under different pH.

group was used as the covalent recognizer which forms boronate with glucose. The formation of boronate is dependent on pH, ionic strength, and temperature. Thus, a glucose sensor should work in an optimal environment. The adsorptivity of CIPC was measured by exposure to varying concentrations of glucose (0 to  $20 \text{ mmol L}^{-1}$ ), followed by the detection of glucose in the residual solution using HPLC.

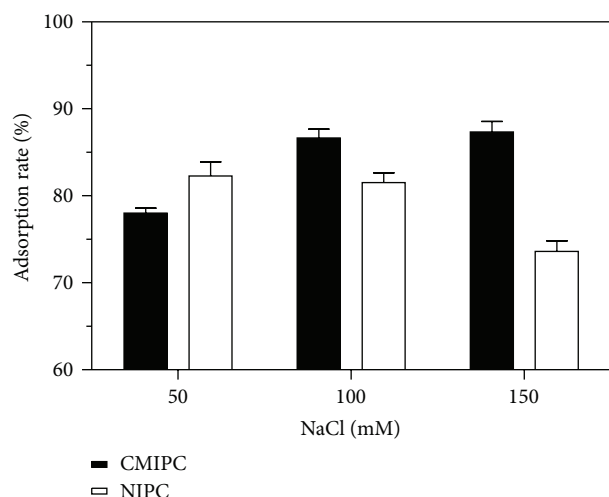


FIGURE 5: Adsorptivity of the CIPIC towards  $10 \text{ mmol L}^{-1}$  glucose under different ion strength.

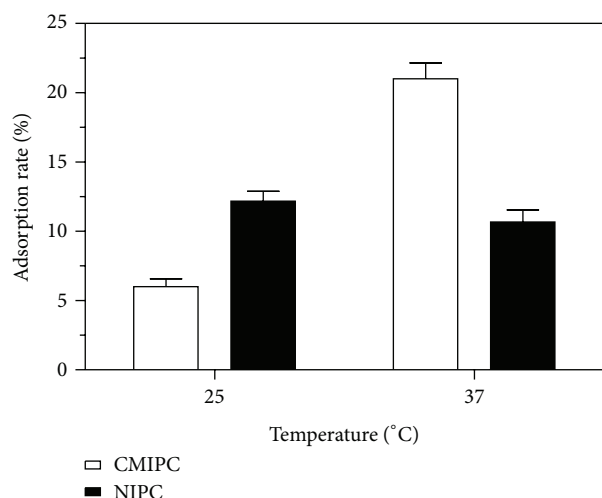


FIGURE 6: Adsorptivity of the CIPIC towards  $10 \text{ mmol L}^{-1}$  glucose at room temperature and body temperature.

Figure 4 shows that the adsorption kinetics of glucose on CIPIC in different pH condition. As shown in Figure 4, within about 3 minutes soaking in a solution of  $10 \text{ mmol L}^{-1}$  glucose the adsorption already reaches the adsorption equilibrium. The presence of macropores provides high specific surface area, more interaction sites, and can also offer increased mass transport. In addition, the response of the sensor to glucose decreases as the pH increases. The adsorptivity of  $10 \text{ mmol L}^{-1}$  glucose at pH 11 is 5 times higher than that of at pH 6.4. An alkaline environment is favorable for boronate formation between glucose and 4-EPBA.

The adsorptivity of glucose to the CIPIC also increases with increasing ionic strength (Figure 5). The effects of temperature on glucose sensing were also investigated by measuring the adsorptivity at room temperature and body temperature. The result in Figure 6 revealed that the CIPIC had better selectivity at  $37^\circ\text{C}$  than at  $25^\circ\text{C}$ . The sensitivity is

considerably better at body temperature, which is beneficial for the detection of glucose in bodily fluid. In short, the CIPIC can characteristically detect glucose under alkaline conditions, at high ionic strength, and human body temperature. With regard to the imprinting effect, even though the NIPC also contains substantial amount of 4-EPBA that enables the NIPC to absorb significant amount of glucose, the imprinting process gave molecular “memory” to the hydrogel matrix. This gives the CIPIC not only the 4-EPBA covalent recognizer but also the imprinted pockets that possess molecular “memory” for glucose. Therefore, the CIPIC absorbs more glucose than NIPC. For  $10 \text{ mmol L}^{-1}$  glucose at pH 11, the absorptivity of the CIPIC is more than 3 times higher than that of the NIPC.

The CIPIC swelled upon glucose adsorption at physiological ionic strengths. As seen in Figure 7(a), in the absence of glucose, the CIPIC diffracts visible light at  $725 \text{ nm}$ , and the diffraction redshifted gradually upon the addition of glucose. When the glucose concentration reached  $20 \text{ mmol L}^{-1}$ , which is clinically indicative of serious diabetes, the CIPIC diffracts visible light at  $880 \text{ nm}$ , giving a redshift of  $155 \text{ nm}$ . In the case of NIPC, a redshift of less than  $60 \text{ nm}$  was observed (Figure 7(b)). After adsorption of glucose, a charged complex forms between phenylboronic acid and the 1,2-*cis*-diol group of glucose, and the lattice of the CIPIC hydrogel swells, which results in the diffraction redshift. This swelling of the CIPIC is reversible. In addition to adsorptivity, the specificity of the hydrogel towards glucose was also improved significantly by imprinting. Compared with the NIPC which had ratios of redshift of glucose/ribose and glucose/rhamnose of 1.6, the corresponding ratios for CIPIC were 3.0 (Figure 8), suggesting that a higher selectivity towards glucose was achieved due to imprinting.

## 4. Conclusion

In summary, a glucose-responsive photonic crystal hydrogel was produced by synthesized hydrogel in the void of PMMA photonic crystal template followed by removing the template. Molecular imprinting improved the selectivity of the photonic crystal-responsive hydrogel towards glucose. The diffraction of the CIPIC red-shifted from  $725 \text{ nm}$  to  $880 \text{ nm}$  in response to  $20 \text{ mmol L}^{-1}$  glucose. Improvement of the current CIPIC technology is still underway, which aims to optimize the CIPIC composition to make the diffraction visible, thus allowing promising clinical application in noninvasive, real-time glucose monitoring.

## Conflict of Interests

None of the authors has ever had direct or indirect financial relations with the commercial identities mentioned in the paper, nor do they declare any other conflict of interests.

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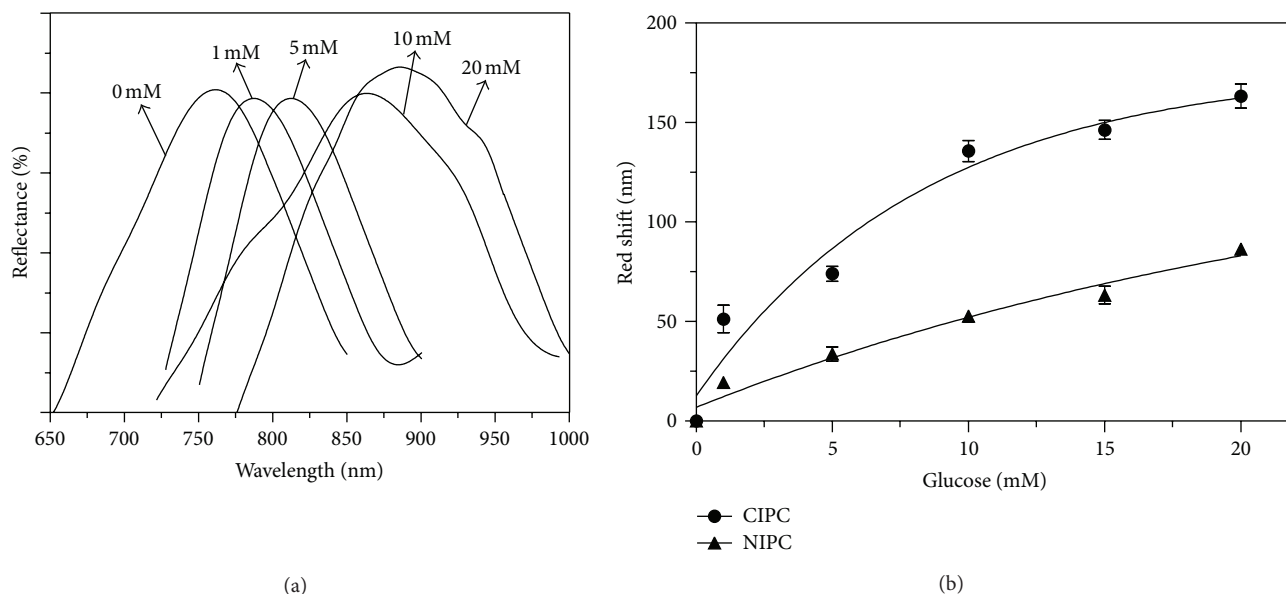


FIGURE 7: Redshift of CIPC (a) and the difference in redshift of CIPC and NIPC (b) in response to glucose. CIPC and NIPC were immersed in glucose solution of different concentration at pH 11, 150 mmol L<sup>-1</sup> ion strength, and 37°C.

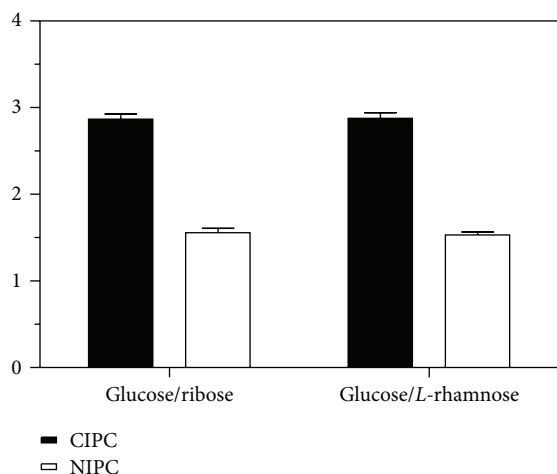


FIGURE 8: Ratio of the redshift between glucose and other saccharides for the CIPC and the NIPC.

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