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Review Article

Emerging PPAR*y***-Independent Role of PPAR***y* Ligands in Lung Diseases

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Peroxisome proliferator activated receptor (PPAR)- γ is a nuclear hormone receptor that is activated by multiple agonists including thiazolidinediones, prostaglandins, and synthetic oleanolic acids. Many PPAR γ ligands are under investigation as potential therapies for human diseases. These ligands modulate multiple cellular pathways via both PPAR γ -dependent and PPAR γ -independent mechanisms. Here, we review the role of PPAR γ and PPAR γ ligands in lung disease, with emphasis on PPAR γ -independent effects. PPAR γ ligands show great promise in moderating lung inflammation, as antiproliferative agents in combination to enhance standard chemotherapy in lung cancer and as treatments for pulmonary fibrosis, a progressive fatal disease with no effective therapy. Some of these effects occur when PPAR γ is pharmaceutically antagonized or genetically PPAR γ and are thus independent of classical PPAR γ -dependent transcriptional control. Many PPAR γ ligands demonstrate direct binding to transcription factors and other proteins, altering their function and contributing to PPAR γ -independent inhibition of disease phenotypes. These PPAR γ -independent mechanisms are of significant interest because they suggest new therapeutic uses for currently approved drugs and because they can be used as probes to identify novel proteins and pathways involved in the pathogenesis or treatment of disease, which can then be targeted for further investigation and drug development.

1. PPARs and PPAR Ligands

Peroxisome proliferator activated receptors (PPARs) are a family of ligand-binding nuclear hormone receptors that are involved in multiple important regulatory pathways including fat metabolism, cell differentiation, proliferation, apoptosis, and inflammation. The first to be identified was PPAR α , the transcription factor responsible for the upregulation of peroxisome proliferation by fatty acids and certain hypolipidemic fibrate drugs. Three PPARs have now been identified, α , β/δ , and γ , encoded by three separate

genes. PPAR*y* is expressed as two different isoforms, PPAR*y*1 and PPAR*y*2, with different transcription start sites. Their detailed domain structure, characteristics, and mode of action have been discussed in detail elsewhere [1, 2].

PPARs have two functional domains, a ligand-binding domain (LBD) and a DNA Binding domain (DBD). Both of these domains are highly conserved in all three receptors. The ligand-binding pocket of the LBD is relatively large and is assumed to bind to a wide range of ligands [2, 3]. Endogenous ligands of the PPARs include fatty acids and fatty acid metabolites; as a result, PPARs are believed to act

in part as lipid sensors. PPARs also bind leukotrienes and prostaglandins such as 15d-PGJ₂, which can have powerful regulatory effects on differentiation and immune responses [4]. At high enough concentration, the PPARs exhibit relatively nonselective binding for fatty acids. However, the significance of this is unclear because the concentration of any ligand in the cellular microenvironment has not been determined [2–5].

Synthetic ligands for all three receptors have been identified and developed. The thiazolidinediones (TZDs) (rosiglitazone, pioglitazone, troglitazone, and ciglitazone) were developed as hypoglycemic agents in the 1980s [6], and the first clinical trial of a TZD as an insulin sensitizer was reported in 1991 [7]. Only after the TZDs were in clinical use were they recognized as ligands of PPARy, activating gene transcription to upregulate expression of adipocyte genes and induce adipocyte differentiation [8]. Although TZDs bind mainly to PPARy, select TZDs activate other PPARs as well [2, 9, 10].

The fibrate drugs (clofibrate, fenofibrate, gemfibrozil, ciprofibrate) have been in clinical use since the 1930s as cholesterol-lowering agents, but it was not until 1995 that it was discovered that they acted via PPARs [11]. The fibrates act most strongly on PPAR α but have activity against PPAR β/δ and γ as well. For example, benzafibrate is a pan-agonist with similar activity against all three PPAR isoforms. Other dual or pan-PPAR ligands have been developed including KRP-297, a dual PPAR α/γ agonist. Some nonsteroidal anti-inflammatory drugs (NSAIDs) (ibuprofen, indomethacin, and fenoprofen) weakly activate both PPAR γ and α [12].

The triterpenoids are a newly recognized class of natural exogenous PPAR γ ligands initially identified in herbal medicines. These compounds are being studied in their native forms and as synthetic derivatives with increased stability, potency, and selectivity. Ursolic acid, a plant triterpenoid, activates PPAR α and has been shown to regulate hepatic lipid metabolism [13]. Oleanolic acid and its naturally occurring derivative 2-cyano-3,11-dioxo-18-olean-1,12-dien-30-oate (CDDO) are potent PPAR γ agonists [14, 15].

2. PPARy and Lung Disease

Beyond its classical role in regulating fat metabolism, PPARy plays a role in regulating cell differentiation and inflammation. It is thus of high interest as potential target for therapies for diseases involving dysregulated inflammation or differentiation. Chronic inflammation due to cigarette smoking and exposure to other environmental toxicants is a contributing factor to numerous lung diseases including COPD (emphysema and chronic bronchitis), asthma, lung cancer, and fibrosis. PPARy expression is dysregulated in patients with cystic fibrosis, sarcoidosis [16], COPD [17, 18], and acute lung injury [19]. Often, and particularly with respect to COPD and fibrosis, current therapies only treat the symptoms and do not modify the course of the underlying disease [17, 20]. Thus, there is great interest in understanding the molecular pathways of chronic lung disease and of using

PPAR*y* ligands both as tools to probe these pathways and as potential therapies.

The lung represents a particularly useful target for this research as (a) many lung diseases involve acute or chronic inflammation or cell proliferation, which are processes targeted by PPARy ligands, (b) drugs can be delivered by inhalation, possibly sparing systemic side effects like heart disease and weight gain, and (c) PPARy ligands appear particularly effective against lung fibrosis, a disease for which effective therapy can be challenging. There are numerous studies demonstrating the potential role of PPARy and PPARy agonists in regulating or treating lung diseases including cancer, fibrosis, and diseases of chronic inflammation. Surprisingly, many of the most interesting effects appear not to require PPARy itself as a transcription factor.

3. PPARy-Independent Effects of PPARy Ligands

Because they play important roles in multiple critical cellular pathways including differentiation, fat metabolism, proliferation, and control of inflammation, PPARy and its ligands have come under increasing scrutiny. However, some effects of PPARy ligands appear to be PPARy-independent that is, they do not require PPARy-dependent transcriptional activation and can occur when PPARy is absent or functionally inactivated (Figure 1). The independent effects of PPARy were first recognized in 2000 by Thieringer et al. [21] who reported that 15d-PGJ2 had anti-inflammatory activity on human peripheral blood monocytes in vitro that was mediated by a PPARy-independent mechanism. More recently, it was reported that 15d-PGJ₂ and ciglitazone induce apoptosis in both normal and malignant human B lymphocytes independent of PPARy activation [22]. It should be noted that PPARy-independent effects can be determined *in vitro* with relative confidence by using PPARyantagonists, gene deletions, overexpression of dominantnegative mutants, or siRNA. However, it is more difficult to discriminate between PPARy-dependent and -independent effects in vivo because complete PPARy deletion is embryonically lethal [14].

Emerging reports demonstrating PPARy-independent effects of PPARy ligands are igniting interest in understanding the mechanisms responsible for the PPARy-independent effects of these compounds. Although they were initially identified as ligands of PPARy, their off-target or direct effects are important as these ligands regulate numerous signaling components that are independent of the classical PPARy pathway. These effects are important for two reasons. First, as PPARy ligand-based therapies enter into clinical testing, it will be critical to understand both their PPARydependent and -independent effects, so that their mechanism of action and potential side effects can be evaluated [23]. Second, PPARy ligands with PPARy-independent effects can be used to identify novel metabolic and regulatory pathways that play a role in human disease and may lead directly to novel therapeutic uses of existing drugs, or to development

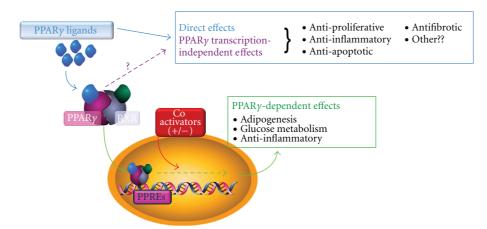


FIGURE 1: PPARy ligands have multiple PPARy-independent effects. In the classical PPARy-dependent pathway, ligand-bound PPARy forms a heterodimer with RXR and binds to PPARy-response elements (PPREs) which leads to modulation of transcription. However, PPARy ligands also exhibit direct effects that do not involve transcriptional activation by PPARy/RXR. These direct effects may involve PPARy protein interacting with PPARy ligands in a "non-classical manner" (not involving RXR or PPRE) or may be completely independent of PPARy (functioning even in the complete absence of PPARy protein, i.e., direct effects). PPARy-independent effects can alter multiple cellular programs including regulation of differentiation, inflammation, apoptosis and may be of significant therapeutic interest.

of new drugs that target these novel pathways uncovered by investigating PPAR-independent effects of these ligands.

4. Mechanism of PPARy-Independent Action of PPARy Ligands

A key question that must be answered as new PPARyligand-based therapies are developed is what the mechanisms and targets of the PPARy-independent effects of these compounds are. Current evidence suggests that the triterpenoids CDDO (and its derivatives) and the prostaglandin 15d-PGJ₂ contain electrophilic carbon atoms that mediate their effects. These carbons can form covalent bonds with free sulfhydryl groups in cellular proteins via the Michael addition reaction, which can then alter the function of the protein. For example, 15d-PGJ₂ can bind p50, which changes its DNA-binding activity [24]. CDDO is a strong electrophile that binds promiscuously to cellular proteins thus making its mechanism of action complex. A recent study in human embryonic kidney cells identified 577 cellular proteins that could bind to CDDO. The analysis was unable to discriminate between high abundance-low affinity targets and low abundance-high affinity targets, and many of these proteins may interact with CDDO without changing their function [25]. Nevertheless, direct protein binding by CDDO and other electrophilic PPARy ligands likely plays a key role in their function, and efforts are underway to identify specific binding partners in specific cell types and disease models.

One target of CDDO and 15d-PGJ₂ is cellular antioxidant defenses. Both CDDO and 15d-PGJ₂ bind glutathione, a key cellular antioxidant. By binding glutathione, CDDO and 15d-PGJ₂ appear to deplete cellular stores of free glutathione, leading to increased oxidative stress and upregulation of antioxidant genes [26–28]. In some but not all cell types,

CDDO also binds Keap-1, which then undocks from its binding partner, the transcription factor Nrf2. Nrf2 is then free to translocate to the nucleus where it is a master regulator of the oxidative stress response. Oral dosing with a CDDO Imidazole derivative prevented emphysema-like changes in lung structure in mice exposed to chronic cigarette smoke, and this effect was abrogated in Nrf2-knockout mice [29]. Activation of antioxidant defense systems may be one mechanism by which PPARy ligands can be used to prevent or treat diseases, especially cancer.

Glucocorticoid receptors (GRs) are also being explored as possible targets of PPARy-independent signaling mechanism. Initial work done by Ialenti et al. showed that rosiglitazone, and ciglitazone decrease IL-6 production in E8.2/GR3 and J774 cells, which is blocked by the GR inhibitor RU486 [30]. Rosiglitazone, pioglitazone and ciglitazone increased GR translocation to the nucleus in HeLa, A549, and PPARydeficient fibroblasts [31]. In addition, evidence was found for rosiglitazone-induced GR recruitment of the transcriptional coactivators SRC-1 and NCoR in HeLa cells [31]. It was also reported in human bronchial cells that the antiproliferative effects of rosiglitazone and troglitazone were mediated by calcium channel signaling through the GPR40 receptor [32].

It should also be noted that it is not always possible to determine whether an effect that is independent of PPARyregulated transcriptional control is also independent of the presence of PPARy protein (Figure 1). PPARy ligands may have "direct" effects that bypass classical RXR/PPRE interactions but which nevertheless require a functional PPARy protein acting in a nonclassical fashion (possibly as a result of covalent modification). Some studies make use of a small molecule inhibitor of PPARy, GW9662, that irreversibly binds to and inactivates the ligand-binding pocket of PPARy, while other studies have been performed in cells that are genetically deficient in PPARy. In these cases, it seems reasonable to conclude that the effects of the PPARy ligands are truly PPARy-independent. However, other studies utilize a dominant-negative PPARy construct that can bind ligands and the PPRE but cannot recruit transcriptional co-factors, or ligand derivatives that do not activate PPRE reporter constructs. In these cases, it can not be ruled out that PPARy ligands bind PPARy protein and carry out their function by a nonclassical pathway that bypasses RXR and the PPRE. Ligands may also act via different pathways in different cell types. More study is needed to understand these PPARyindependent mechanisms.

5. Inflammation

In the lung, PPARy is expressed in endothelial cells, fibroblasts, smooth muscle cells, airway epithelial cells, and resident alveolar macrophages, as well as infiltrating leukocytes including neutrophils, eosinophils, and mast cells [33, 34]. PPARy expression has been identified in multiple lung cell lines including A549, BEAS-2B, and NCI-H292 [35].

Clinical evidence suggests that PPARy and PPARy ligands are involved in regulating inflammatory responses in the lung [36, 37], and chronic or dysregulated inflammation is an important pathologic feature in many lung diseases including asthma, COPD, sarcoidosis, and pulmonary fibrosis. Expression of PPARy is increased in lung tissue from asthmatic patients, specifically localized in mucosal, epithelial, and smooth muscle cells and is associated with the severity of inflammation [34]. PPARy expression was decreased in patients with severe pulmonary hypertension, particularly around plexiform lesions [38].

Troglitazone, ciglitazone, and 15d-PGJ₂ strongly inhibited LPS-induced secretion of TNFa and IL-6 and downregulated expression of Cox-2 and iNos in mouse macrophages. PPARy-deficient macrophages responded similarly to wildtype control, suggesting a PPARy-independent signaling mechanism [39]. Troglitazone and 15d-PGJ₂ also activated PI3K signaling independently of PPARy in A549 cells (a human lung epithelial carcinoma line) via increased ERK phosphorylation and ERK/MAPK signaling, leading to increased production of Cox-2 and PGE₂ [40]. Troglitazone and rosiglitazone promoted proliferation in nonmalignant human bronchial epithelial cells by a PPARy-independent mechanism, while 15d-PGJ₂ blocked proliferation by a PPARy-dependent mechanism [32]. Recently, we reported that 15d-PGJ₂ and CDDO inhibited the silica-induced inflammatory response of primary human lung fibroblasts via a largely PPAR-independent pathway attributed to the electrophilic properties of non-TZD ligands of PPARy [41].

In light of their anti-inflammatory properties *in vitro*, animal models have been used to explore the potential of PPARy ligands as therapeutic agents in inflammatory lung diseases. Oral administration of ciglitazone decreased cell infiltration, epithelial dysplasia, and IL-2, IL-4, and IFNy production in a mouse model of allergic airway inflammation [42]. A similar effect was observed with intranasal administration of ciglitazone, with decreased cell infiltration in the lung, particularly that of eosinophils, and reduced airway remodeling [43]. Other TZDs such as rosiglitazone

also decreased neutrophil and eosinophil infiltration and reduced airway hyperresponsiveness to methacholine [44]. Ciglitazone also decreased local inflammation in the lung in a model of *S. pneumoniae*-induced pneumonia, with concurrent reductions in TNF α , IL-6, IL-12p70, and IFN γ secretion [45]. However, it is difficult to attribute the effects of the ligands in *in vivo* studies to PPAR γ -dependent or -independent mechanisms, and further study is needed.

6. Lung Cancer and PPARy Ligands

Lung cancer accounts for more cancer-related deaths than any other type of cancer worldwide [46]. In 2010, there were 157,300 lung cancer-related fatalities in the United States alone [47]. As a disease, lung cancer can be divided into two main categories, nonsmall cell lung cancer (NSCLC), which comprises around 85% of all lung malignancies, and small cell lung cancer (SCLC), which comprises the other 15% [48]. NSCLC can be further classified into three subgroups, large cell cancer, adenocarcinoma, and squamous cell carcinoma, depending upon cell type and histological classification. As lung cancer is a substantial cause of human suffering and death, a multitude of therapeutic approaches have been employed. Many therapeutic options are actively being pursued including several recent investigations that utilize PPARy ligands as treatments [49, 50]. While recent work demonstrates PPARy ligands have beneficial PPARy-dependent and -independent properties in treating malignancies, emerging insights into PPARyindependent functions in lung cancer are highlighted here.

6.1. In Vitro Models of Lung Cancer and PPARy Ligands. PPARy ligands including the TZDs, 15d-PGJ2, and triterpenoids (CDDO and its derivatives) have been investigated in lung cancer cells in vitro. Troglitazone dramatically reduces cell proliferation of A549 cells, a human lung adenocarcinoma cell line, at concentrations of 10 to $40 \,\mu$ M. Because PPARy activity, determined by reporter assays, reached a maximal level at $15 \,\mu\text{M}$ while the beneficial reductions in cell proliferation continued up to $40 \,\mu\text{M}$, the authors suggested that troglitazone had desirable PPARyindependent effects at elevated doses [51]. Troglitazone decreased expression of the cell cycle regulatory protein cyclin D1 and increased the number of cells that were growth-arrested in the G_1/G_0 phase of the cell cycle [51]. Recently, a troglitazone analog, delta2-Troglitazone, that does not stimulate PPARy-dependent transcription, was shown to increase proteasomal degradation of cyclin D1 in breast cancer cells [52]. This suggests that troglitazone, at least in part, affects cyclin D1 expression independent of PPARy. Cyclin D1 is highly expressed in many types of tumors, including the lung [53, 54]. Troglitazone also modifies ERK-1/2 signaling in NSCLC cells [51, 55-57] and triggers apoptosis in two lung cancer cell lines, but not in a control, nonmalignant line, underscoring the potential of TZDs to specifically target tumor cells [56]. In A549 cells, troglitazone directly interacts with the epidermal growth factor receptor (EGFR), binding the EGFR and augmenting its internalization into the lysosome where it is subsequently degraded [58]. The reduction in EGFR protein levels leads to a loss of EGFR signaling activity, which consequently halts cell proliferation. Interestingly, EGFR mutations are some of the most common mutations found in lung cancer patients [48].

Pioglitazone and rosiglitazone also show a number of beneficial effects in attenuating growth of lung cancer cells in vitro. In 2006, Han and Roman determined that rosiglitazone had significant PPARy-dependent and -independent effects on blunting NSCLC cell growth [59]. In this study, rosiglitazone impaired proliferation of NSCLC cells by increasing expression of the tumor suppressor phosphatase PTEN (phosphatase and tensin homolog), reducing Akt and p70S6K phosphorylation, and increasing phosphorylation of AMPK. The irreversible PPARy inhibitor GW9662 reversed the effects on PTEN expression and Akt phosphorylation but not AMPK and p70S6K phosphorylation, suggesting a mixture of PPARy-dependent and -independent effects. AMPK is a master regulator of energy metabolism and regulates mammalian target of rapamycin (mTOR) mediated cell growth, proliferation, and protein synthesis pathways. Phosphorylation of AMPK inhibits mTOR signaling to reduce proliferation and protein synthesis through a reduction of p70S6K phosphorylation. Interestingly, rosiglitazone enhanced the effects of rapamycin on halting NSCLC cell growth, accentuating the potential for combination therapies. The mechanism whereby rosiglitazone activates AMPK phosphorylation is still unclear.

Pioglitazone and rosiglitazone also decreased production of the prostaglandin PGE₂ in both A549 and A427 NSCLC cell lines. Increased levels of PGE2 stimulate proliferation and prosurvival pathways in numerous malignancies, including NSCLC, suggesting that decreasing PGE₂ production may reduce cancer cell growth. The effects of the agonists were not reversed by either GW9662 or by expression of a dominant-negative PPARy, demonstrating that the effects were PPARy-independent. Investigation into the mechanism determined a corresponding increase in the expression of 15-hydroxy prostaglandin dehydrogenase (15-PGDH), which converts PGE₂ into a biologically inactive 15-keto prostaglandin derivative [60]. Rosiglitazone also inhibited expression of the α 4 nicotinic acetylcholine receptor (α 4nAcR) in a PPARy-independent manner in three different NSCLC cell lines [61]. Nicotine stimulates proliferation of NSCLC cells through activation of α 4-nAcR. Rosiglitazone activated Erk-1/2 signaling to prompt the tumor suppressor protein p53 to inhibit α 4-nAcR expression. The effects of rosiglitazone treatment on α 4-nAcR expression were independent of PPARy as demonstrated by depletion of PPARy protein by PPARy siRNA as well as through the use of GW9662 [62]. Rosiglitazone and GW1929, another TZD, also inhibit fibronectin expression by tumor cells in vitro [63]. Fibronectin expression and extracellular deposition promote tumor cell interactions.

An interesting non-lung study demonstrated that TZDs reduce expression of the oncogenic transcription factor FoxM1 in hepatoma cells [64]. Regulation of FoxM1 expression by TZDs was independent of PPARy expression, as

expression of FoxM1 was reduced by TZDs even in the presence of PPAR γ siRNA. Because FoxM1 is often elevated in NSCLC, further investigation to see if TZDs can also reduce FoxM1 expression in lung cancer may be worthwhile.

The endogenous PPARy ligand, 15d-PGJ₂, has also been shown to have beneficial effects in treating lung cancer cells. 15d-PGJ₂ promotes apoptosis of A549 cells by activating caspase 3 and attenuating expression of cyclin D1 in a PPARy-independent manner [65]. 15d-PGJ₂ also increased production of reactive oxygen species (ROS) in lung cancer cells while reducing intracellular glutathione levels [66]. The effects could not be reversed by a PPARy siRNA or the antagonist GW9662 but was reversible with quercetin, a powerful antioxidant, suggesting that 15d-PGJ₂ modulated the redox state of the cell. Interestingly, 15d-PGJ₂ has been shown to covalently attach to glutathione and lead to its oxidation [67], which may account for the loss of glutathione seen in the studies.

The novel triterpenoid CDDO and its synthetic derivatives CDDO-Methyl ester (CDDO-Me) and CDDO-Imidazole (CDDO-Im) have also been investigated as chemotherapeutic agents in treating lung cancer. CDDO-Im promoted apoptosis in A549 and H358 human lung cancer cell lines in a dose-dependent manner with a rapid decrease in STAT5 phosphorylation [68]. STAT5, a phosphorylationdependent transcription factor in the JAK/STAT pathway, is involved in the transcription of proliferation and prosurvival genes. CDDO-Im induced expression of SOCS-1 (a STAT inhibitor) and SHP-1 (a phosphatase that targets STAT5) in as little as 30 minutes. Because SOCS-1 and SHP-1 are both regulated by the antioxidant response transcription factor, Nrf2, the authors speculate that CDDO-Im activates Nrf2 by binding to its regulatory partner Keap-1 [68].

6.2. Animal Models of Lung Cancer and PPARy Ligands. Although it has been difficult to dissect out the specific PPARy-independent functions of the PPARy ligands from their PPARy-dependent functions in vivo, the ability of PPARy ligands to inhibit tumor cell growth in vitro has led to a number of significant studies in animal models. Ciglitazone, which inhibits proliferation of A549 cells in vitro by a PPARy-independent mechanism as discussed above, also significantly reduced A549 tumor weights in the nude mouse xenograft model, with concomitant reduction in cyclin D1 and increased expression of the cell cycle inhibitor p21 [51]. The xenograft model was also used to demonstrate that daily treatment with 15d-PGJ₂ had potent antitumor effects against subcutaneous tumors comprised of A549 or H460 cells. Of note, 15d-PGJ₂ significantly enhanced the effects of the chemotherapeutic drug docetaxel [65].

In the lung, Girnun et al. utilized a transgenic mouse model containing inducible activating mutations in either KRAS or epidermal growth factor (EGFR), two genes commonly mutated in human lung cancer [69]. Rosiglitazone, in combination with the standard chemotherapeutic agent carboplatin, significantly reduced tumor size by increasing apoptosis and decreasing cell proliferation. Importantly, neither drug alone had significant effects, and rosiglitazone did not enhance the myelosuppressive effects of carboplatin, suggesting that combination therapy would be safe and effective. Addition of rosiglitazone to a chemotherapeutic cocktail of hydrazine, selenium, and phenylbutyrate significantly reduced lung hyperplasia and adenoma in a mouse carcinogen model compared to the cocktail alone [70]. In another carcinogen-induced lung cancer model in mice, pioglitazone treatment had a clear preventative effect [71]. Mice given oral doses of pioglitazone 8 weeks after initial injection of carcinogen had no change in number of adenocarcinomas but a 64% decrease in tumor volume compared to control, along with a 35% reduction in the number of squamous cell carcinoma.

The CDDO derivatives CDDO-Me and CDDO-ethyl amide also inhibit lung cancer in preclinical mouse studies. In one study, mice were fed the CDDO derivatives starting one week after treatment with vinyl carbamate, a carcinogen that induces adenocarcinoma of the lung. The CDDOtreated mice had a dramatically reduced tumor burden [72]. The authors suggest that CDDO prevented tumor formation through upregulation of the antioxidant heme-oxygenase-1 (HO-1) and through inhibition of STAT phosphorylation leading to increased apoptosis, as demonstrated in vitro. In another recent study using the transplantable tumor model, Lewis lung carcinoma (LLC), CDDO-Me drastically reduced tumor size and inhibited myeloid-derived suppressor cells that are involved in the immune suppressive response against tumors [73]. These data are very intriguing as they suggest CDDO-Me may augment cancer vaccine therapies.

7. PPAR-Independent Effects of PPAR Ligands in Pulmonary Fibrosis

Fibrotic remodeling following injury and repair occurs in many tissues including the kidney, heart, skin, and liver [74]. Pulmonary fibrosis is characterized by pathologic remodeling of the lung, with thickening of interstitial spaces, deposition of collagen and other matrix proteins, contraction and stiffening of lung tissue, loss of alveolar architecture, reduced gas exchange and ultimately respiratory failure. Pulmonary fibrosis can be caused by a variety of insults including chronic inflammation and inhalation of particulates like asbestos and silica, radiation, drugs, and as a squeal of connective tissue diseases. Idiopathic pulmonary fibrosis (IPF) is a severe form of pulmonary fibrosis with no current therapy. The only "cure" is a lung transplant. The disease is progressive and the median time from diagnosis to death at 2.9 years is shorter than that for lung cancer [15].

One of the key pathogenic processes in lung fibrosis is the differentiation and activation of fibroblasts and myofibroblasts to produce the components of scar tissue (hypercellularity, collagen, and other extracellular matrix proteins). The key pro fibrotic cytokine appears to be TGF β . Because of the central role of TGF β and myofibroblasts, there is intense interest in developing novel therapies that target these key players. Pirfenidone, a small orally active molecule that inhibits synthesis of TGF β and TNF α and attenuates growth of fibroblasts *in vitro*, was recently approved for clinical use in IPF in Europe, but not yet in the United States. The clinical benefits of pirfenidone appear to be small but significant in some studies [75]; no other therapies have been approved that specifically target the underlying cellular pathology of lung fibrosis.

7.1. In Vitro Models of Pulmonary Fibrosis and PPAR Ligands. There is a growing body of evidence that both natural and synthetic PPARy agonists have powerful antifibrotic effects *in vitro* [76], and these results are beginning to be translated to preclinical animal models. Typical experiments examine differentiation of human lung fibroblasts to myofibroblasts *in vitro* and associated changes in expression of profibrotic cytokines and matrix proteins. A variety of nontransformed human lung fibroblast (HLF) cell lines are used including fetal, neonatal, adult nonfibrotic, and adult fibrotic (derived from patient biopsies).

We and others reported that rosiglitazone and 15d-PGJ₂ inhibited TGF β -driven myofibroblast differentiation of primary HLFs [77–80]. Expression of a dominant PPARy was able to reverse the inhibitory effect of rosiglitazone more effectively than 15d-PGJ₂, suggesting that rosiglitazone can act via both PPARy-dependent and -independent mechanisms while 15d-PGJ₂ acts predominantly via an independent mechanism [77]. Since primary HLFs express abundant PPARy and RXR proteins and are capable of PPARy-dependent transcriptional regulation [77], this suggested that the antifibrotic effects of the PPARy agonists were mediated through both PPARy-dependent and -independent mechanisms. We also investigated CDDO, a triterpenoid originally identified in herbal preparations with anti-inflammatory properties. We determined that CDDO has an EC₅₀ for inhibition of myofibroblast differentiation that is 20-fold lower than 15d-PGJ₂ and 400-fold lower than rosiglitazone, and it acts independently of PPARy as confirmed by pharmacological and genetic approaches [78]. Recently, we reported that 15d-PGJ₂ and CDDO inhibit TGFβ-induced phosphorylation of phosphotidyl-inositol 3kinase-protein kinase B (PI3K-Akt) and focal adhesion kinase (FAK), but not TGF β -induced p38-MAPK phosphorylation, and that the mechanism was independent of PPARy [79]. We also noted that there is a strong correlation between the ability of a PPARy ligand to inhibit Akt phosphorylation with its ability to suppress myofibroblast differentiation (Figure 2). We find that rosiglitazone is a weak inhibitor of TGF β -induced Akt phosphorylation (Figure 2) and may be therefore a poor choice as an antifibrotic treatment [79].

It is essential to note that the three main mechanisms of activation of myofibroblast differentiation—TGF β , mechanical stress, or adhesion and integrin activation—act via a common pathway in which FAK is autophosphorylated, leading to increased levels of the active kinase (phospho-FAK). It is conceivable that once TGF β activates myofibroblast differentiation, the increased deposition of extracellular matrix proteins would cause additional mechanical stress at the cell surface leading to sustained and continual activation of FAK. Since FAK itself upregulates myofibroblast differentiation, once TGF β initiates this process, sustained activation of FAK

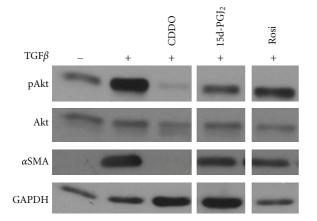


FIGURE 2: PPARy ligands inhibit TGF β -induced Akt phosphorylation and myofibroblast differentiation with varied potency. Primary human lung fibroblasts were treated with TGF β (5 ng/mL), alone or in combination with three PPARy ligands (CDDO (1 μ M), 15d-PGJ₂ (5 μ M) and rosiglitazone (20 μ M)). Protein lysates were electrophoretically separated on the same gel, and representative lanes from a single experiment are shown here. The potency of a PPARy ligand to inhibit Akt phosphorylation corresponds to its ability to inhibit myofibroblast differentiation. While CDDO inhibits both Akt phosphorylation and α SMA potently, rosiglitazone is a weak inhibitor of both.

would be able to perpetuate the fibrotic response even in the absence of additional external profibrotic signals.

Several other laboratories have also investigated PPARy ligands in lung fibroblast differentiation and activation. Lin et al. reported that rosiglitazone inhibited migration, proliferation, and phenotypic differentiation of cultured fetal human lung fibroblasts induced by fetal bovine serum albumin. The inhibitory effect of rosiglitazone on myofibroblast differentiation, determined by expression of α -smooth muscle actin, was also PPARy-independent [81]. Troglitazone and ciglitazone were evaluated against normal human fetal lung fibroblasts and fibroblasts isolated from patients with idiopathic interstitial pneumonias. Both ligands inhibited proliferative responses of normal and fibrotic lung fibroblasts to platelet-derived growth factor (PDGF) and TGF β in a dose- and time-dependent fashion and arrested cells at the G_1/G_0 stage. Troglitazone and ciglitazone inhibited expression of cyclin D1, similar to their activity in the A549 adenocarcinoma cell line (see above). Interestingly, they also found that both troglitazone and ciglitazone efficiently inhibited collagen expression by myofibroblasts and that collagen synthesis was also inhibited by overexpression of a constitutively active form of PPARy [80]. Since cyclin D1 is a known downstream target of PI3K-Akt pathway, it is conceivable that these and other ligands of PPARy (15d-PGJ₂ and CDDO) inhibit cyclin D1 and arrest fibroblast proliferation via a PPARy-independent mechanism involving Akt but inhibit collagen synthesis and other profibrotic functions by a PPARy-dependent mechanism.

Not all myofibroblasts in lung fibrosis are derived from preexisting lung resident fibroblasts. Some myofibroblasts may originate from lung epithelial cells via epithelial to mesenchymal transition (EMT). EMT is a process during which terminally differentiated epithelial cells transform into a mesenchymal phenotype and give rise to fibroblasts and subsequently to myofibroblasts. This process is believed to play an important role in normal repair following injury to alveolar epithelial cells (AECs) [74]. The in vivo significance of EMT is controversial, but it is well-described in vitro and is characterized by decreased expression of E-cadherin and increased expression of N-cadherin [82]. TGF β potently stimulates EMT and alters the morphology of human AECs. Tan et al. investigated whether PPARy ligands could inhibit TGF β -induced EMT in the A549 cell line, an adenocarcinoma of AEC origin. Rosiglitazone and ciglitazone rescued TGF β -mediated repression of E-cadherin via a PPARy-dependent mechanism but, at the same time, inhibited TGF β -induced increased N-cadherin expression independent of PPARy [82].

Myofibroblasts in lung fibrosis can also originate from circulating adult progenitor cells called fibrocytes. Fibrocytes are distinct from epithelial and endothelial cells, monocyte/macrophages, lymphocytes, dendritic cells, or tissue fibroblasts [83]. Work in the Strieter laboratory demonstrated that TGF β induces myofibroblast differentiation of fibrocytes harvested from human peripheral blood mononuclear cells as measured by α SMA expression [84]. Troglitazone inhibits TGF β -induced α SMA expression and myofibroblast differentiation, and this effect is not reversed by the PPARy inhibitor GW9662, indicating that the effects of troglitazone are PPARy-independent. $TGF\beta$ signaling activates the Smad and MAP kinases pathways in fibrocytes, and PPARy agonists negatively regulate this process through a PPARy-independent pathway. Of special note, troglitazone inhibited JNK activation, which both inhibited myofibroblast differentiation and stimulated differentiation of fibrocytes to adipocytes (fat storage cells) [84]. Thus, pharmacological intervention with PPARy ligands may alter the fate of circulating myofibroblast precursors before they ever reach the lung.

7.2. PPARy Ligands in Animal Models of Lung Fibrosis. There are currently only a few published reports investigating PPARy ligands in animal models of fibrosis, although additional studies are in progress. Troglitazone given orally (200 or 400 mg/kg/day) inhibited bleomycin-induced lung fibrosis in mice, with reductions in lung collagen content and TGF β levels [80]. Rosiglitazone and 15d-PGJ₂ were also tested in the mouse bleomycin model and reduced mortality, inflammation, cellular influx, and fibrosis [85]. To investigate the PPARy dependence of the effect, mice were cotreated with bisphenol A diglycidyl ether (BADGE), a PPARy antagonist. BADGE reversed the antifibrotic effects of rosiglitazone and 15d-PGJ₂, suggesting that at least some of the antifibrotic activity of these compounds is mediated by a PPARy-dependent mechanism in vivo [86]. However, while BADGE has been reported to be a PPARy antagonist in some studies [85], it activates PPARy in others [87]. Interestingly, the stimulation of heme-oxygenase (HO-1) by 15d-PGJ₂ in rat primary cortical neuron cultures was blocked by BADGE [88], but, in human lung fibroblasts, HO-1 induction was not blocked by another PPARy antagonist GW9662 [41]. This suggests that the same PPARy ligand may have dependent effects in some cell types and independent effects in others.

A different result was observed in a neonatal hyperoxia model. Premature infants requiring hyperoxia (100% O₂) during the first weeks of life suffer from a high rate of bronchopulmonary dysplasia (BPD), characterized by mild inflammation and fibrosis, and failed alveolarization. In neonatal mice, hyperoxia leads to upregulation of $TGF\beta$ and Wnt signaling and downregulation of PPARy signaling owing to reduction in both PPARy mRNA and protein [89, 90]. While rosiglitazone markedly prevented lung injury in hyperoxic neonates, the effect was reversed by administration of GW9662, suggesting a PPARy-dependent mechanism [89]. It should be noted that BPD is thought to represent a failure of the developmental program, and therefore the signals and mechanisms involved in neonatal hyperoxic fibrosis may be different than the signals involved in fibrosis of fully differentiated adult lungs, and it would be interesting to study PPARy-independent effects of electrophilic ligands of PPARy in the neonatal animal models [91].

Additional study is needed to characterize the *in vivo* mechanism of action of PPARy ligands. Because the bleomycin model is characterized by acute early inflammation, it is possible that the majority of the antifibrotic effects seen in these early studies are a result of PPARy-mediated inhibition of inflammation that precedes fibrosis. Therefore, it will be important to investigate the effects of PPARy ligands in fibrosis models that do not also have overt inflammation. CDDO and its derivatives have strong antifibrotic effects *in vitro* at much lower concentrations than TZDs but have not yet been investigated *in vivo* for lung fibrosis. However, it is clear that PPARy ligands have strong therapeutic potential in these studies regardless of whether the effects are dependent or independent of PPARy-mediated transcriptional regulation.

8. Other Pulmonary Diseases

Rosiglitazone attenuates pulmonary arterial hypertension in an ApoE null mouse model fed a high-fat diet [92]. This appears to be PPARy-dependent as mice with targeted deletion of the PPARy gene in smooth muscle cell tissue spontaneously developed pulmonary arterial hypertension [92]. Furthermore, under hypoxia-induced pulmonary hypertension conditions, rosiglitazone prevents and reverses pulmonary hypertension by downregulating Nox4 [93].

Acute promyelocytic leukemia (APL) is a subtype of acute myelocytic leukemia in which myeloid precursor cells grow aberrantly and can infiltrate into the pulmonary tract causing severe disease. APL is characterized by a genomic translocation fusing the PML tumor suppressor and the retinoic acid receptor α gene. While APL is normally treated with all-trans retinoic acid (ATRA), cells can become resistant to this therapy, thus novel drug cocktails are needed. CDDO has been shown to increase proapoptotic

and differentiating effects of ATRA in APL-derived NB4 cells and partially reverse ATRA resistance in ATRA-resistant NB4-derived cells [94]. In a mouse model of APL initiated by a PML-RAR α transgene, CDDO and ATRA cotreatment significantly increased animal survival rates by compared to ATRA treatment alone. The PPARy selective antagonist T0070907, or PPARy siRNA only partially impaired the effects of CDDO treatment, suggesting both PPARydependent and -independent effects of CDDO in APL cells. Another report suggested beneficial, proapoptotic effects of ciglitazone in APL [95]. Ciglitazone treatment caused a significant increase in APL cell apoptosis demonstrated by DNA fragmentation analysis, increased active caspase-3, cleaved PARP, and decreased expression of the X-linked inhibitor of apoptosis protein. Notably, the effects of ciglitazone were reversed by treatment with GW9662, suggesting a PPARydependent mechanism.

9. Potential Therapies

Animal models clearly show extraordinary translational potential for PPARy ligands to serve as either preventative therapies or cotreatments for lung malignancies. Recent clinical studies also suggest favorable outcomes in lung cancer patients treated with PPARy ligands. In a clinical study encompassing patients with diabetes, lung cancer occurrence in patients receiving TZD therapy was decreased by 33% compared to patients not taking a TZD [96]. Notably, the decrease in lung carcinomas was still evident after confounding factors (i.e., age and race/ethnicity) were accounted for. At the present time, clinical trials are underway to investigate the effectiveness of TZDs in the prevention and/or treatment of lung cancer, either alone, or in combination with other chemotherapies. It is important to note that TZDs show both PPARy-dependent and -independent modes of action and further exploration of their mechanism of action is needed to elucidate their specific targets. CDDO and its derivatives have proapoptotic and antiproliferative effects on cancer cells in vitro and are currently being investigated in phase 2 clinical trials for treatment of cancer [97]. Further studies are required to determine if CDDO-Me will also be useful as a myelosuppressive agent in combination with chemotherapies.

Currently, there are no PPARy ligands in clinical trial for fibrotic lung diseases. However, there is evidence in other organ systems suggesting that PPARy ligands may have antifibrotic potential in human lung fibrosis. Intriguingly, kidney remodeling is a frequent complication of diabetes, and improvements in renal function have been noted in patients with type II diabetes treated with TZDs [98, 99]. The use of TZDs in clinical therapy may be limited by undesirable side effects. However, unlike diabetes, which can often be managed with alternate therapies, there are no current treatments for IPF and the median survival time after diagnosis is only 2-3 years, which may shift the risk-benefit assessment in favor of use.

In addition to lung cancer and fibrosis, pioglitazone and rosiglitazone are in ongoing clinical trials for asthma (e.g., NCT01134835 and NCT00614874), cystic fibrosis (e.g., NCT00322868 and NCT01060566), pulmonary hypertension (e.g., NCT00825266 and NCT00006071), and pulmonary arterial disease (e.g., NCT00153166 and NCT00064727). These studies will evaluate whether the *in vitro* anti-inflammatory effects of PPARy ligands can be translated to improved patient outcomes.

Current PPARy ligands act via both PPARy-dependent and -independent mechanisms, and the impact of both pathways must be considered in evaluating their efficacy in animal models and clinical trials. However, the existence of PPARy-independent effects creates the potential for development of novel non-PPAR-based therapies that access the independent pathways uncovered by PPARy ligands. In this event, currently available PPARy ligands with strong PPAR-independent effects, such as 15d-PGJ₂ and CDDO, may play a more important role as probes for novel disease modifying pathways than as direct clinical treatments, paving the way for new molecules that target critical pathways (such as Akt, FAK, ERK, and JNK) without also activating PPARy.

It is interesting to note that some of the PPAR γ independent effects of PPARy ligands are mediated by modulating the phosphorylation status of key regulatory enzymes such as Akt, FAK, ERK, and JNK. Imatinib mesylate (Gleevec), a broad-spectrum tyrosine kinase inhibitor currently approved to treat some cancers, has been tested in several small open-label trials in systemic sclerosis, a fibrosing disease that often involves the lung. In some studies, imatinib improved lung function and reduced skin contraction, although the number of adverse effects was high [100, 101]. However, in a randomized, placebo-controlled trial of patients with mild to moderate IPF, imatinib did not significantly reduce disease progression [29]. These studies demonstrate the clinical potential as well as limitation of antikinase treatments in lung fibrosis and encourage further exploration of the non-PPAR molecular targets of PPARy ligands.

10. Conclusion

Discovered serendipitously, the off-target, or PPARyindependent effects of PPARy ligands may prove as interesting and therapeutically useful as their PPARy-dependent effects. PPARy agonists have potent PPARy-independent effects *in vitro* and *in vivo*, regulating proinflammatory responses and acting to promote apoptosis and inhibit differentiation, which may be beneficial in treating cancer and fibrosing diseases. Clinical trials are underway, investigating the currently approved TZDs in novel diseases, and investigating novel agonists such as CDDO and its derivatives. Significant clinical progress can be expected in the next few years.

The most complex problem associated with the use of PPARy agonists as disease therapies is indeed the combination of potent PPARy-dependent and -independent activities they exhibit. This presents both multiple challenges and multiple opportunities for translational investigation. The PPARy-dependent and independent effects can be decoded with specificity using *in vitro* models, but the results are

difficult to confirm *in vivo* because homozygous PPARy deletion is lethal. And, of course, patients will be subjected to both the dependent and independent effects of any new drugs, regardless of their primary mode of action *in vitro*. Thus, it will be critically important to carefully evaluate both the PPARy-dependent and -independent therapeutic effects and side effects of any new therapies.

Some PPARy ligands exhibit PPARy-dependent and -independent effects that act in the same direction, such as the antifibrotic activities of the TZDs. Unfortunately, the TZDs carry the risk of significant side effects including edema, weight gain, and cardiovascular effects. Troglitazone (Rezulin) has been withdrawn from the market due to hepatotoxicity [102], while rosiglitazone (Avandia) has been withdrawn in Europe and is under restriction in the US due to cardiovascular effects [103]. In the context of pulmonary disease, while there are alternative therapies for diabetes, there are no effective current therapies for lung fibrosis, and despite advances in chemotherapy, the five-year survival rate for NSCLC is only 15%. Thus, it is possible that these drugs will be revived for use in these deadly disease, either alone or in combination with other therapies. Alternatively, direct lung administration by inhalation may increase the effectiveness of the drugs while sparing the patient from systemic side effects, as is the case with inhaled steroids for asthma.

Also promising is the ability to use PPARy ligands as probes to uncover novel disease regulatory pathways which can then be targeted by new, specific therapies. Cyclin D1, Keap-1, Akt, and FAK are examples of disease targets that have been identified through the PPARy-independent effects of PPARy ligands. It is now possible, through mass spectrometry and other techniques, to determine exactly how these compounds bind to their targets and alter their function. This should allow the development of new compounds that have specific targeting activity against their new targets, while eliminating or reducing their affinity for PPARy and thus reducing or eliminating PPAR-dependent activity and its associated side effects. As these ligands enter clinical trials, there is an urgent need to understand their PPAR-dependent and -independent mechanisms of action for the future of targeted and personalized medicines.

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References

 B. Desvergne and W. Wahli, "Peroxisome proliferatoractivated receptors: nuclear control of metabolism," *Endocrine Reviews*, vol. 20, no. 5, pp. 649–688, 1999.

- [2] J. Berger and D. E. Moller, "The mechanisms of action of PPARs," *Annual Review of Medicine*, vol. 53, pp. 409–435, 2002.
- [3] M. Schupp and M. A. Lazar, "Endogenous ligands for nuclear receptors: digging deeper," *The Journal of Biological Chemistry*, vol. 285, no. 52, pp. 40409–40415, 2010.
- [4] N. Noy, "Ligand specificity of nuclear hormone receptors: sifting through promiscuity," *Biochemistry*, vol. 46, no. 47, pp. 13461–13467, 2007.
- [5] T. M. Willson and W. Wahli, "Peroxisome proliferatoractivated receptor agonists," *Current Opinion in Chemical Biology*, vol. 1, no. 2, pp. 235–241, 1997.
- [6] A. Y. Chang, B. M. Wyse, B. J. Gilchrist, T. Peterson, and A. R. Diani, "Ciglitazone, a new hypoglycemic agent. I. Studies in ob/ob and db/db mice, diabetic Chinese hamsters, and normal and streptozotocin-diabetic rats," *Diabetes*, vol. 32, no. 9, pp. 830–838, 1983.
- [7] T. Kuzuya, Y. Iwamoto, K. Kosaka et al., "A pilot clinical trial of a new oral hypoglycemic agent, CS-045, in patients with non-insulin dependent diabetes mellitus," *Diabetes Research* and Clinical Practice, vol. 11, no. 3, pp. 147–153, 1991.
- [8] C. Day, "Thiazolidinediones: a new class of antidiabetic drugs," *Diabetic Medicine*, vol. 16, no. 3, pp. 179–192, 1999.
- [9] M. Heald and M. A. Cawthorne, "Dual acting and Pan-PPAR activators as potential anti-diabetic therapies," *Handbook of Experimental Pharmacology*, vol. 203, pp. 35–51, 2011.
- [10] E. Adeghate, A. Adem, M. Y. Hasan, K. Tekes, and H. Kalasz, "Medicinal chemistry and actions of dual and pan PPAR modulators," *The Open Medicinal Chemistry Journal*, vol. 5, supplement 2, pp. 93–98, 2011.
- [11] S. S. Lee, T. Pineau, J. Drago et al., "Targeted disruption of the α isoform of the peroxisome proliferator-activated receptor gene in mice results in abolishment of the pleiotropic effects of peroxisome proliferators," *Molecular and Cellular Biology*, vol. 15, no. 6, pp. 3012–3022, 1995.
- [12] J. M. Lehmann, J. M. Lenhard, B. B. Oliver, G. M. Ringold, and S. A. Kliewer, "Peroxisome proliferator-activated receptors α and γ are activated by indomethacin and other nonsteroidal anti-inflammatory drugs," *The Journal of Biological Chemistry*, vol. 272, no. 6, pp. 3406–3410, 1997.
- [13] Y. Jia, M. J. Bhuiyan, H. J. Jun et al., "Ursolic acid is a PPARα agonist that regulates hepatic lipid metabolism," *Bioorganic* & *Medicinal Chemistry Letters*, vol. 21, no. 19, pp. 5876–5880, 2011.
- [14] Y. Wang, W. W. Porter, N. Suh et al., "A synthetic triterpenoid, 2-cyano-3,12-dioxooleana-1,9-dien-28-oic acid (CDDO), is a ligand for the peroxisome proliferator-activated receptor γ," *Molecular Endocrinology*, vol. 14, no. 10, pp. 1550–1556, 2000.
- [15] P. J. Sime, "The antifibrogenic potential of PPARy ligands in pulmonary fibrosis," *Journal of Investigative Medicine*, vol. 56, no. 2, pp. 534–538, 2008.
- [16] D. A. Culver, B. P. Barna, B. Raychaudhuri et al., "Peroxisome proliferator-activated receptor *y* activity is deficient in alveolar macrophages in pulmonary sarcoidosis," *American Journal of Respiratory Cell and Molecular Biology*, vol. 30, no. 1, pp. 1–5, 2004.
- [17] P. J. Barnes and B. R. Celli, "Systemic manifestations and comorbidities of COPD," *The European Respiratory Journal*, vol. 33, no. 5, pp. 1165–1185, 2009.
- [18] J. H. Lee, M. Hanaoka, Y. Kitaguchi et al., "Imbalance of apoptosis and cell proliferation contributes to the development and persistence of emphysema," *Lung*, vol. 190, no. 1, pp. 69–82, 2012.

- [19] H. Y. Cho, W. Gladwell, X. Wang et al., "Nrf2-regulated PPARy expression is critical to protection against acute lung injury in mice," *American Journal of Respiratory and Critical Care Medicine*, vol. 182, no. 2, pp. 170–182, 2010.
- [20] J. A. de Andrade and V. J. Thannickal, "Innovative approaches to the therapy of fibrosis," *Current Opinion in Rheumatology*, vol. 21, no. 6, pp. 649–655, 2009.
- [21] R. Thieringer, J. E. Fenyk-Melody, C. B. Le Grand et al., "Activation of peroxisome proliferator-activated receptor *y* does not inhibit IL-6 or TNF-*α* responses of macrophages to lipopolysaccharide in vitro or in vivo," *Journal of Immunology*, vol. 164, no. 2, pp. 1046–1054, 2000.
- [22] D. M. Ray, F. Akbiyik, and R. P. Phipps, "The peroxisome proliferator-activated receptor y (PPARy) ligands 15-deoxy- δ 12,14-prostaglandin J2 and ciglitazone induce human B lymphocyte and B cell lymphoma apoptosis by PPARy-independent mechanisms," *Journal of Immunology*, vol. 177, no. 8, pp. 5068–5076, 2006.
- [23] M. G. Belvisi, D. J. Hele, and M. A. Birrell, "Peroxisome proliferator-activated receptor gamma agonists as therapy for chronic airway inflammation," *European Journal of Pharmacology*, vol. 533, no. 1–3, pp. 101–109, 2006.
- [24] E. Cernuda-Morollon, E. Pineda-Molina, F. J. Canada, and D. Perez-Sala, "15-Deoxy-δ12,14-prostaglandin J2 inhibition of NF-κB-DNA binding through covalent modification of the p50 subunit," *The Journal of Biological Chemistry*, vol. 276, no. 38, pp. 35530–35536, 2001.
- [25] M. M. Yore, A. N. Kettenbach, M. B. Sporn, S. A. Gerber, and K. T. Liby, "Proteomic analysis shows synthetic oleanane triterpenoid binds to mTOR," *PLoS ONE*, vol. 6, no. 7, Article ID e22862, 2011.
- [26] P. Yue, Z. Zhou, F. R. Khuri, and S. Y. Sun, "Depletion of intracellular glutathione contributes to JNK-mediated death receptor 5 upregulation and apoptosis induction by the novel synthetic triterpenoid methyl-2-cyano-3, 12-dioxooleana-1, 9-dien-28-oate (CDDO-Me)," *Cancer Biology & Therapy*, vol. 5, no. 5, pp. 492–497, 2006.
- [27] A. L. Levonen, D. A. Dickinson, D. R. Moellering, R. T. Mulcahy, H. J. Forman, and V. M. Darley-Usmar, "Biphasic effects of 15-deoxy-δ12,14-prostaglandin J2 on glutathione induction and apoptosis in human endothelial cells," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 21, no. 11, pp. 1846–1851, 2001.
- [28] H. E. Ferguson, T. H. Thatcher, K. C. Olsen et al., "Peroxisome proliferator-activated receptor-y ligands induce heme oxygenase-1 in lung fibroblasts by a PPARy-independent, glutathione-dependent mechanism," *American Journal of Physiology*, vol. 297, no. 5, pp. L912–L919, 2009.
- [29] C. E. Daniels, J. A. Lasky, A. H. Limper et al., "Imatinib treatment for idiopathic pulmonary fibrosis: randomized placebo-controlled trial results," *American Journal of Respiratory and Critical Care Medicine*, vol. 181, no. 6, pp. 604–610, 2010.
- [30] A. Ialenti, G. Grassia, P. Di Meglio, P. Maffia, M. Di Rosa, and A. Ianaro, "Mechanism of the anti-inflammatory effect of thiazolidinediones: relationship with the glucocorticoid pathway," *Molecular Pharmacology*, vol. 67, no. 5, pp. 1620– 1628, 2005.
- [31] L. Matthews, A. Berry, M. Tersigni, F. D'Acquisto, A. Ianaro, and D. Ray, "Thiazolidinediones are partial agonists for the glucocorticoid receptor," *Endocrinology*, vol. 150, no. 1, pp. 75–86, 2009.
- [32] D. Gras, P. Chanez, V. Urbach, I. Vachier, P. Godard, and C. Bonnans, "Thiazolidinediones induce proliferation

of human bronchial epithelial cells through the GPR40 receptor," *American Journal of Physiology*, vol. 296, no. 6, pp. L970–L978, 2009.

- [33] G. M. Denning and L. L. Stoll, "Peroxisome proliferatoractivated receptors: potential therapeutic targets in lung disease?" *Pediatric Pulmonology*, vol. 41, no. 1, pp. 23–34, 2006.
- [34] L. Benayoun, S. Letuve, A. Druilhe et al., "Regulation of peroxisome proliferator-activated receptor y expression in human asthmatic airways: relationship with proliferation, apoptosis, and airway remodeling," *American Journal of Respiratory and Critical Care Medicine*, vol. 164, no. 8 I, pp. 1487–1494, 2001.
- [35] R. Pawliczak, C. Han, X. L. Huang, A. J. Demetris, J. H. Shelhamer, and T. Wu, "85-kDa cytosolic phospholipase A2 mediates peroxisome proliferator-activated receptor y activation in human lung epithelial cells," *The Journal of Biological Chemistry*, vol. 277, no. 36, pp. 33153–33163, 2002.
- [36] H. Duez, J. C. Fruchart, and B. Staels, "PPARs in inflammation, atherosclerosis and thrombosis," *Journal of Cardiovascular Risk*, vol. 8, no. 4, pp. 187–194, 2001.
- [37] S. L. Spinelli, J. J. O'Brien, S. Bancos et al., "The PPARplatelet connection: modulators of inflammation and potential cardiovascular effects," *PPAR Research*, vol. 2008, Article ID 328172, 16 pages, 2008.
- [38] S. Ameshima, H. Golpon, C. D. Cool et al., "Peroxisome proliferator-activated receptor gamma (PPARy) expression is decreased in pulmonary hypertension and affects endothelial cell growth," *Circulation Research*, vol. 92, no. 10, pp. 1162– 1169, 2003.
- [39] A. Chawla, Y. Barak, L. Nagy, D. Liao, P. Tontonoz, and R. M. Evans, "PPAR-y dependent and independent effects on macrophage-gene expression in lipid metabolism and inflammation," *Nature Medicine*, vol. 7, no. 1, pp. 48–52, 2001.
- [40] K. M. Patel, K. L. Wright, P. Whittaker, P. Chakravarty, M. L. Watson, and S. G. Ward, "Differential modulation of COX-2 expression in A549 airway epithelial cells by structurally distinct PPARy agonists: evidence for disparate functional effects which are independent of NF-κB and PPARy," *Cellular Signalling*, vol. 17, no. 9, pp. 1098–1110, 2005.
- [41] C. M. Hogan, T. H. Thatcher, R. E. Sapinoro et al., "Electrophilic PPARy ligands attenuate IL-1β and silicainduced inflammatory mediator production in human lung fibroblasts via a PPARy-independent mechanism," *PPAR Research*, vol. 2011, Article ID 318134, 11 pages, 2011.
- [42] C. Mueller, V. Weaver, J. P. Vanden Heuvel, A. August, and M. T. Cantorna, "Peroxisome proliferator-activated receptor *y* ligands attenuate immunological symptoms of experimental allergic asthma," *Archives of Biochemistry and Biophysics*, vol. 418, no. 2, pp. 186–196, 2003.
- [43] C. K. Rhee, S. Y. Lee, J. Y. Kang et al., "Effect of peroxisome proliferator-activated receptor-gamma on airway smooth muscle thickening in a murine model of chronic asthma," *International Archives of Allergy and Immunology*, vol. 148, no. 4, pp. 289–296, 2009.
- [44] J. E. Ward, D. J. Fernandes, C. C. Taylor, J. V. Bonacci, L. Quan, and A. G. Stewart, "The PPARy ligand, rosiglitazone, reduces airways hyperresponsiveness in a murine model of allergen-induced inflammation," *Pulmonary Pharmacology* and Therapeutics, vol. 19, no. 1, pp. 39–46, 2006.
- [45] M. E. Stegenga, S. Florquin, A. F. de Vos, and T. van der Poll, "The thiazolidinedione ciglitazone reduces bacterial outgrowth and early inflammation during streptococcus

pneumoniae pneumonia in mice," *Critical Care Medicine*, vol. 37, no. 2, pp. 614–618, 2009.

- [46] R. S. Herbst and S. M. Lippman, "Molecular signatures of lung cancer—toward personalized therapy," *The New England Journal of Medicine*, vol. 356, no. 1, pp. 76–78, 2007.
- [47] A. Jemal, R. Siegel, J. Xu, and E. Ward, "Cancer statistics, 2010," CA: A Cancer Journal for Clinicians, vol. 60, no. 5, pp. 277–300, 2010.
- [48] R. S. Herbst, J. V. Heymach, and S. M. Lippman, "Lung cancer," *The New England Journal of Medicine*, vol. 359, no. 13, pp. 1367–1380, 2008.
- [49] R. A. Nemenoff, M. Weiser-Evans, and R. A. Winn, "Activation and molecular targets of peroxisome proliferatoractivated receptor-γ ligands in lung cancer," *PPAR Research*, vol. 2008, Article ID 156875, 8 pages, 2008.
- [50] J. Roman, "Peroxisome proliferator-activated receptor y and lung cancer biology: implications for therapy," *Journal of Investigative Medicine*, vol. 56, no. 2, pp. 528–533, 2008.
- [51] V. G. Keshamouni, R. C. Reddy, D. A. Arenberg et al., "Peroxisome proliferator-activated receptor-*y* activation inhibits tumor progression in non-small-cell lung cancer," *Oncogene*, vol. 23, no. 1, pp. 100–108, 2004.
- [52] C. Colin, S. Salamone, I. Grillier-Vuissoz et al., "New troglitazone derivatives devoid of PPARy agonist activity display an increased antiproliferative effect in both hormonedependent and hormone-independent breast cancer cell lines," *Breast Cancer Research and Treatment*, vol. 124, no. 1, pp. 101–110, 2010.
- [53] O. Gautschi, D. Ratschiller, M. Gugger, D. C. Betticher, and J. Heighway, "Cyclin D1 in non-small cell lung cancer: a key driver of malignant transformation," *Lung Cancer*, vol. 55, no. 1, pp. 1–14, 2007.
- [54] W. I. Yang, K. Y. Chung, D. H. Shin, and Y. B. Kim, "Cyclin D1 protein expression in lung cancer," *Yonsei Medical Journal*, vol. 37, no. 2, pp. 142–150, 1996.
- [55] M. Li, T. W. Lee, A. P. Yim, T. S. Mok, and G. G. Chen, "Apoptosis induced by troglitazone is both peroxisome proliteratoractivated receptor-y- and ERK-dependent in human nonsmall lung cancer cells," *Journal of Cellular Physiology*, vol. 209, no. 2, pp. 428–438, 2006.
- [56] M. Li, T. W. Lee, T. S. Mok, T. D. Warner, A. P. Yim, and G. G. Chen, "Activation of peroxisome proliferator-activated receptor-*y* by troglitazone (TGZ) inhibits human lung cell growth," *Journal of Cellular Biochemistry*, vol. 96, no. 4, pp. 760–774, 2005.
- [57] R. A. Winn, M. Van Scoyk, M. Hammond et al., "Antitumorigenic effect of Wnt 7a and Fzd 9 in non-small cell lung cancer cells is mediated through ERK-5-dependent activation of peroxisome proliferator-activated receptor y," *The Journal of Biological Chemistry*, vol. 281, no. 37, pp. 26943–26950, 2006.
- [58] X. Li, X. Yang, Y. Xu et al., "Troglitazone inhibits cell proliferation by attenuation of epidermal growth factor receptor signaling independent of peroxisome proliferatoractivated receptor *y*," *Cell Research*, vol. 19, no. 6, pp. 720– 732, 2009.
- [59] S. Han and J. Roman, "Rosiglitazone suppresses human lung carcinoma cell growth through PPARy-dependent and PPARy-independent signal pathways," *Molecular Cancer Therapeutics*, vol. 5, no. 2, pp. 430–437, 2006.
- [60] S. Hazra, R. K. Batra, H. H. Tai, S. Sharma, X. Cui, and S. M. Dubinett, "Pioglitazone and rosiglitazone decrease prostaglandin E2 in non-small-cell lung cancer cells by upregulating 15-hydroxyprostaglandin dehydrogenase," *Molecular Pharmacology*, vol. 71, no. 6, pp. 1715–1720, 2007.

- [61] X. Sun, J. D. Ritzenthaler, X. Zhong, Y. Zheng, J. Roman, and S. Han, "Nicotine stimulates PPARβ/δ expression in human lung carcinoma cells through activation of P13K/mTOR and suppression of AP-2α," *Cancer Research*, vol. 69, no. 16, pp. 6445–6453, 2009.
- [62] X. Sun, J. D. Ritzenthaler, Y. Zheng, J. Roman, and S. Han, "Rosiglitazone inhibits α4 nicotinic acetylcholine receptor expression in human lung carcinoma cells through peroxisome proliferator-activated receptor *y*-independent signals," *Molecular Cancer Therapeutics*, vol. 8, no. 1, pp. 110–118, 2009.
- [63] S. Han, J. D. Ritzenthaler, H. N. Rivera, and J. Roman, "Peroxisome proliferator-activated receptor-y ligands suppress fibronectin gene expression in human lung carcinoma cells: involvement of both CRE and Sp1," *American Journal of Physiology*, vol. 289, no. 3, pp. L419–L428, 2005.
- [64] V. Petrovic, R. H. Costa, L. F. Lau, P. Raychaudhuri, and A. L. Tyner, "Negative regulation of the oncogenic transcription factor FoxM1 by thiazolidinediones and mithramycin," *Cancer Biology & Therapy*, vol. 9, no. 12, pp. 1008–1016, 2010.
- [65] S. V. Fulzele, A. Chatterjee, M. S. Shaik, T. Jackson, N. Ichite, and M. Singh, "15-Deoxy-δ12,14-prostaglandin J2 enhances docetaxel anti-tumor activity against A549 and H460 nonsmall-cell lung cancer cell lines and xenograft tumors," *Anti-Cancer Drugs*, vol. 18, no. 1, pp. 65–78, 2007.
- [66] J. J. Wang and O. T. Mak, "Induction of apoptosis by 15d-PGJ2 via ROS formation: an alternative pathway without PPARy activation in non-small cell lung carcinoma A549 cells," *Prostaglandins & Other Lipid Mediators*, vol. 94, no. 3-4, pp. 104–111, 2011.
- [67] C. M. Paumi, P. K. Smitherman, A. J. Townsend, and C. S. Morrow, "Glutathione S-transferases (GSTs) inhibit transcriptional activation by the peroxisomal proliferator-activated receptor *y* (PPARy) ligand, 15-Deoxyδ12,14Prostaglandin J2 (15-d-PGJ 2)," *Biochemistry*, vol. 43, no. 8, pp. 2345–2352, 2004.
- [68] K. Liby, N. Voong, C. R. Williams et al., "The synthetic triterpenoid CDDO-Imidazolide suppresses STAT phosphorylation and induces apoptosis in myeloma and lung cancer cells," *Clinical Cancer Research*, vol. 12, no. 14, pp. 4288– 4293, 2006.
- [69] G. D. Girnun, L. Chen, J. Silvaggi et al., "Regression of drugresistant lung cancer by the combination of rosiglitazone and carboplatin," *Clinical Cancer Research*, vol. 14, no. 20, pp. 6478–6486, 2008.
- [70] C. M. Lyon, D. M. Klinge, K. C. Do et al., "Rosiglitazone prevents the progression of preinvasive lung cancer in a murine model," *Carcinogenesis*, vol. 30, no. 12, pp. 2095– 2099, 2009.
- [71] Y. Wang, M. James, W. Wen et al., "Chemopreventive effects of pioglitazone on chemically induced lung carcinogenesis in mice," *Molecular Cancer Therapeutics*, vol. 9, no. 11, pp. 3074–3082, 2010.
- [72] K. Liby, D. B. Royce, C. R. Williams et al., "The synthetic triterpenoids CDDO-methyl ester and CDDO-ethyl amide prevent lung cancer induced by vinyl carbamate in A/J mice," *Cancer Research*, vol. 67, no. 6, pp. 2414–2419, 2007.
- [73] S. Nagaraj, J. I. Youn, H. Weber et al., "Anti-inflammatory triterpenoid blocks immune suppressive function of MDSCs and improves immune response in cancer," *Clinical Cancer Research*, vol. 16, no. 6, pp. 1812–1823, 2010.
- [74] T. A. Wynn, "Cellular and molecular mechanisms of fibrosis," *Journal of Pathology*, vol. 214, no. 2, pp. 199–210, 2008.

- [75] P. W. Noble, C. Albera, W. Z. Bradford et al., "Pirfenidone in patients with idiopathic pulmonary fibrosis (CAPACITY): two randomised trials," *The Lancet*, vol. 377, no. 9779, pp. 1760–1769, 2011.
- [76] C. M. Hart, J. Roman, R. Reddy, and P. J. Sime, "PPARy: a novel molecular target in lung disease," *Journal of Investigative Medicine*, vol. 56, no. 2, pp. 515–517, 2008.
- [77] H. A. Burgess, L. E. Daugherty, T. H. Thatcher et al., "PPARγ agonists inhibit TGF-β induced pulmonary myofibroblast differentiation and collagen production: implications for therapy of lung fibrosis," *American Journal of Physiology*, vol. 288, no. 6, pp. L1146–L1153, 2005.
- [78] H. E. Ferguson, A. Kulkarni, G. M. Lehmann et al., "Electrophilic peroxisome proliferator-activated receptor-y ligands have potent antifibrotic effects in human lung fibroblasts," *American Journal of Respiratory Cell and Molecular Biology*, vol. 41, no. 6, pp. 722–730, 2009.
- [79] A. A. Kulkarni, T. H. Thatcher, K. C. Olsen, S. B. Maggirwar, R. P. Phipps, and P. J. Sime, "PPAR-y ligands repress TGFβ-induced myofibroblast differentiation by targeting the PI3K/Akt pathway: implications for therapy of fibrosis," *PLoS ONE*, vol. 6, no. 1, Article ID e15909, 2011.
- [80] J. E. Milam, V. G. Keshamouni, S. H. Phan et al., "PPARy agonists inhibit profibrotic phenotypes in human lung fibroblasts and bleomycin-induced pulmonary fibrosis," *American Journal of Physiology*, vol. 294, no. 5, pp. L891– L901, 2008.
- [81] Q. Lin, L. P. Fang, W. W. Zhou, and X. M. Liu, "Rosiglitazone inhibits migration, proliferation, and phenotypic differentiation in cultured human lung fibroblasts," *Experimental Lung Research*, vol. 36, no. 2, pp. 120–128, 2010.
- [82] X. Tan, H. Dagher, C. A. Hutton, and J. E. Bourke, "Effects of PPARγ ligands on TGF-β1-induced epithelial-mesenchymal transition in alveolar epithelial cells," *Respiratory Research*, vol. 11, article 21, 2010.
- [83] R. M. Strieter, E. C. Keeley, M. D. Burdick, and B. Mehrad, "The role of circulating mesenchymal progenitor cells, fibrocytes, in promoting pulmonary fibrosis," *Transactions of the American Clinical and Climatological Association*, vol. 120, pp. 49–59, 2009.
- [84] K. M. Hong, J. A. Belperio, M. P. Keane, M. D. Burdick, and R. M. Strieter, "Differentiation of human circulating fibrocytes as mediated by transforming growth factor-β and peroxisome proliferator-activated receptor," *The Journal of Biological Chemistry*, vol. 282, no. 31, pp. 22910–22920, 2007.
- [85] H. M. Wright, C. B. Clish, T. Mikami et al., "A synthetic antagonist for the peroxisome proliferator-activated receptor *y* inhibits adipocyte differentiation," *The Journal of Biological Chemistry*, vol. 275, no. 3, pp. 1873–1877, 2000.
- [86] T. Genovese, S. Cuzzocrea, R. Di Paola et al., "Effect of rosiglitazone and 15-deoxy-δ12,14-prostaglandin J2 on bleomycin-induced lung injury," *The European Respiratory Journal*, vol. 25, no. 2, pp. 225–234, 2005.
- [87] D. Bishop-Bailey, T. Hla, and T. D. Warner, "Bisphenol A diglycidyl ether (BADGE) is a PPARy agonist in an ECV304 cell line," *British Journal of Pharmacology*, vol. 131, no. 4, pp. 651–654, 2000.
- [88] T. N. Lin, W. M. Cheung, J. S. Wu et al., "15d-prostaglandin J2 protects brain from ischemia-reperfusion injury," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 26, no. 3, pp. 481–487, 2006.
- [89] V. K. Rehan, Y. Wang, S. Patel, J. Santos, and J. S. Torday, "Rosiglitazone, a peroxisome proliferator-activated receptory agonist, prevents hyperoxia-induced neonatal rat lung

injury in vivo," *Pediatric Pulmonology*, vol. 41, no. 6, pp. 558–569, 2006.

- [90] C. Dasgupta, R. Sakurai, Y. Wang et al., "Hyperoxia-induced neonatal rat lung injury involves activation of TGF-β and Wnt signaling and is protected by rosiglitazone," *American Journal of Physiology*, vol. 296, no. 6, pp. L1031–L1041, 2009.
- [91] F. J. Auletta and A. P. Flint, "Mechanisms controlling corpus luteum function in sheep, cows, nonhuman primates, and women especially in relation to the time of luteolysis," *Endocrine Reviews*, vol. 9, no. 1, pp. 88–105, 1988.
- [92] G. Hansmann, R. A. Wagner, S. Schellong et al., "Pulmonary arterial hypertension is linked to insulin resistance and reversed by peroxisome proliferator-activated receptor-y activation," *Circulation*, vol. 115, no. 10, pp. 1275–1284, 2007.
- [93] R. E. Nisbet, J. M. Bland, D. J. Kleinhenz et al., "Rosiglitazone attenuates chronic hypoxia-induced pulmonary hypertension in a mouse model," *American Journal of Respiratory Cell* and Molecular Biology, vol. 42, no. 4, pp. 482–490, 2010.
- [94] Y. Tabe, M. Konopleva, Y. Kondo et al., "PPARy-active triterpenoid CDDO enhances ATRA-induced differentiation in APL," *Cancer Biology & Therapy*, vol. 6, no. 12, pp. 1967– 1977, 2007.
- [95] J. J. Liu, Y. W. Guo, Z. G. Fang et al., "Activation of peroxisome proliferator-activated receptor-y induces apoptosis on acute promyelocytic leukemia cells via downregulation of XIAP," *International Journal of Molecular Medicine*, vol. 24, no. 5, pp. 623–632, 2009.
- [96] R. Govindarajan, L. Ratnasinghe, D. L. Simmons et al., "Thiazolidinediones and the risk of lung, prostate, and colon cancer in patients with diabetes," *Journal of Clinical Oncology*, vol. 25, no. 12, pp. 1476–1481, 2007.
- [97] K. Liby, R. Risingsong, D. B. Royce et al., "Triterpenoids CDDO-methyl ester or CDDO-ethyl amide and rexinoids LG100268 or NRX194204 for prevention and treatment of lung cancer in mice," *Cancer Prevention Research*, vol. 2, no. 12, pp. 1050–1058, 2009.
- [98] R. E. Buckingham, "Thiazolidinediones: pleiotropic drugs with potent anti-inflammatory properties for tissue protection," *Hepatology Research*, vol. 33, no. 2, pp. 167–170, 2005.
- [99] M. C. Lansang, C. Coletti, S. Ahmed, M. S. Gordon, and N. K. Hollenberg, "Effects of the PPAR-y agonist rosiglitazone on renal haemodynamics and the renin-angiotensin system in diabetes," *Journal of the Renin-Angiotensin-Aldosterone System*, vol. 7, no. 3, pp. 175–180, 2006.
- [100] T. Philip, J. O. Armitage, G. Spitzer et al., "High-dose therapy and autologous bone marrow transplantation after failure of conventional chemotherapy in adults with intermediategrade or high-grade non-Hodgkin's lymphoma," *The New England Journal of Medicine*, vol. 316, no. 24, pp. 1493–1498, 1987.
- [101] Z. Tamaki, Y. Asano, M. Hatano et al., "Efficacy of low-dose imatinib mesylate for cutaneous involvement in systemic sclerosis: a preliminary report of three cases," *Modern Rheumatology*, vol. 22, no. 1, pp. 94–99, 2012.
- [102] A. C. Need, A. G. Motulsky, and D. B. Goldstein, "Priorities and standards in pharmacogenetic research," *Nature Genetics*, vol. 37, no. 7, pp. 671–681, 2005.
- [103] D. Cohen, "Insiders criticise FDA's decision not to withdraw rosiglitazone," *British Medical Journal*, vol. 341, Article ID c5333, 2010.



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