Hindawi Publishing Corporation Oxidative Medicine and Cellular Longevity Volume 2016, Article ID 4529060, 8 pages http://dx.doi.org/10.1155/2016/4529060



# **Review Article Role of Endogenous Sulfur Dioxide in Regulating Vascular Structural Remodeling in Hypertension**

Jia Liu,<sup>1</sup> Yaqian Huang,<sup>1</sup> Selena Chen,<sup>2</sup> Chaoshu Tang,<sup>3,4</sup> Hongfang Jin,<sup>1</sup> and Junbao Du<sup>1</sup>

<sup>1</sup>Department of Pediatrics, Peking University First Hospital, Beijing 100034, China

<sup>2</sup>University of California, San Diego, La Jolla, CA 92093, USA

<sup>3</sup>Department of Physiology and Pathophysiology, Peking University Health Science Center, Beijing 100191, China <sup>4</sup>Key Lab of Molecular Cardiovascular Science of Ministry of Education, Beijing 100191, China

Correspondence should be addressed to Hongfang Jin; jinhongfang51@126.com and Junbao Du; junbaodu1@126.com

Received 22 March 2016; Accepted 21 July 2016

Academic Editor: Guangdong Yang

Copyright © 2016 Jia Liu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Sulfur dioxide (SO<sub>2</sub>), an emerging gasotransmitter, was discovered to be endogenously generated in the cardiovascular system. Recently, the physiological effects of endogenous SO<sub>2</sub> were confirmed. Vascular structural remodeling (VSR), an important pathological change in many cardiovascular diseases, plays a crucial role in the pathogenesis of the diseases. Here, the authors reviewed the research progress of endogenous SO<sub>2</sub> in regulating VSR by searching the relevant data from PubMed and Medline. In spontaneously hypertensive rats (SHRs) and pulmonary hypertensive rats, SO<sub>2</sub>/aspartate aminotransferase (AAT) pathway was significantly altered. SO<sub>2</sub> inhibited vascular smooth muscle cell (VSMC) proliferation, promoted apoptosis, inhibited the synthesis of extracellular collagen but promoted its degradation, and enhanced antioxidative capacity, thereby playing a significant role in attenuating VSR. However, the detailed mechanisms needed to be further explored. Further studies in this field would be important for the better understanding of the pathogenesis of systemic hypertension and pulmonary hypertension. Also, clinical trials are needed to demonstrate if SO<sub>2</sub> would be a potential therapeutic target in cardiovascular diseases.

### 1. Introduction

Since the 1980s, studies have shown that the endogenous gaseous molecules nitric oxide (NO), carbon monoxide (CO), and hydrogen sulfide (H<sub>2</sub>S) are endogenously produced and have a wide range of biological effects including vasodilation and inhibition of vascular smooth muscle cell (VSMC) proliferation and platelet aggregation. They have an important physiological and pathological significance. Gaseous signal molecules share the following characteristics [1]: (1) small gaseous molecules with low molecular weight; (2) freely passing through the cell membrane, independent of the specific cell receptor; (3) being endogenously generated under the enzyme catalysis and regulated by metabolic pathways; (4) having a clear specific function at physiological concentration; and (5) exerting biological effects, having specific cellular and molecular targets. Accordingly, gaseous signal molecule pathways and the physiologic and pathophysiologic

significance have become hot topic in cardiovascular system and other systems [2].

In recent years, studies showed endogenous  $SO_2$  pathways in tissues of rats including cardiovascular tissues [3, 4]. The physiological effects of  $SO_2$  were also confirmed, including vasodilation [4–6], negative regulation of cardiac function [7], and regulation of lipid metabolism [8–10]. Also, the pathophysiologic role of  $SO_2$  in the processes of vascular structural remodeling (VSR) [11, 12], inflammatory response [13], and oxidative response was indicated [12, 13]. In this review, the research progress of  $SO_2$  in regulating VSR was summarized.

## 2. The Property of SO<sub>2</sub>

2.1. General Physical and Chemical Properties of  $SO_2$ .  $SO_2$  is a colorless gas accompanied by a pungent odor [14]. The relative molecular mass of  $SO_2$  is 64 g/mol, and the ionic

charge of sulfur is 4+. Hence, it has both oxidative ability and reducibility. Reduced sulfur has a longer half-life in the body, and thereby it has a strong antioxidant effect [15]. In the atmosphere, however,  $SO_2$  is oxidized to sulfuric acid or sulfate aerosols which severely pollute the atmosphere [16].

2.2. Generation and Metabolism of Endogenous SO<sub>2</sub>. Endogenous SO<sub>2</sub> is produced during metabolism of sulfur-containing amino acids (Figure 1). Firstly, sulfur-containing amino acid is metabolized to L-cysteine that is then oxidized into Lcysteinesulfinate catalyzed by cysteine dioxygenase (CDO). L-Cysteinesulfinate, an analogue of L-asparaginic acid, can be transaminated into  $\beta$ -sulfinylpyruvate catalyzed by a spartate aminotransferase (AAT), which is then spontaneously decomposed into pyruvate and SO<sub>2</sub> [15, 17, 18]. In vivo,  $SO_2$  produces  $HSO_3^{-}/SO_3^{-2-}$  (molar ratio of 1:3) in water, which can be oxidated into  $SO_4^{2-}$  by sulfite oxidase and then excreted through the kidneys [19]. However, there are still some gaseous forms of SO<sub>2</sub> existing in the body. On the other hand, L-cysteinesulfinate can be also decarboxylated into CO<sub>2</sub> and hypotaurine by cysteinesulfinate decarboxylase (CSD). A large majority of hypotaurine can be further oxidized into taurine, which takes place during the metabolism of bile acids. Studies have found that taurine functions as an inhibitory neurotransmitter, membrane stabilizing factor, and broad spectrum cytoprotector, and so forth [20, 21].

In addition, endogenous  $SO_2$  can be generated by intracellular  $H_2S$ . Firstly,  $H_2S$  is oxidized to thiosulfate by heme compounds, metal-protein complexes, and ferritin. Thiosulfate reacts with reduced glutathione under the catalysis of thiosulfate reductase (TSR) to form sulfite or  $SO_2$  [19, 22]. Meanwhile, Mitsuhashi et al. [23] found that activated neutrophils in mammals could be catalyzed by NADPH to convert  $H_2S$  into sulfites through oxidative stress.

2.3. Detection Methods of SO<sub>2</sub>. In 2003, Balazy et al. [24] detected SO<sub>2</sub> in the coronary artery and myocardium using gas chromatography/mass spectrometry. In 2008, Du et al. [4] first found that there was an endogenous SO<sub>2</sub>/AAT pathway in cardiovascular system and other systems, with the highest SO<sub>2</sub> content in the arteries. In arteries, the highest SO<sub>2</sub> content was found in the aorta  $(5.55 \pm 0.35 \,\mu\text{mol/g protein})$ , followed by the pulmonary artery  $(3.27 \pm 0.21 \,\mu\text{mol/g protein})$ , mesenteric artery (2.67  $\pm$  0.17  $\mu$ mol/g protein), renal artery (2.50  $\pm$  0.20  $\mu$ mol/g protein), and the caudal artery (2.23  $\pm$  0.19  $\mu$ mol/g protein). The content in heart (1.78 $\pm$ 0.12  $\mu$ mol/g protein), liver (1.74  $\pm$  0.16  $\mu$ mol/g protein), lung (1.42  $\pm$  0.11  $\mu$ mol/g protein), and kidney (0.95 $\pm$ 0.11  $\mu$ mol/g protein) was relatively low. The content of SO<sub>2</sub> in plasma was 15.54  $\pm$  1.68  $\mu$ mol/L.

AAT, a key enzyme in the generation of endogenous SO<sub>2</sub>, is distributed in mammals [25]. There are two kinds of isoenzymes AAT1 and AAT2. AAT1 is mainly located in the cytoplasm, and AAT2 is mainly in mitochondria [26]. However, the activity of AAT in tissues was not consistent with SO<sub>2</sub> tissue level. AAT activity was highest in heart tissue (4469  $\pm$ 278 U/g), followed by liver (1328  $\pm$  198 U/g), kidney (381  $\pm$ 48 U/g), and lung tissue (175  $\pm$  38 U/g). AAT activity value in the arteries was (188 $\pm$ 30 U/g) in renal artery, (143 $\pm$ 36 U/g) in

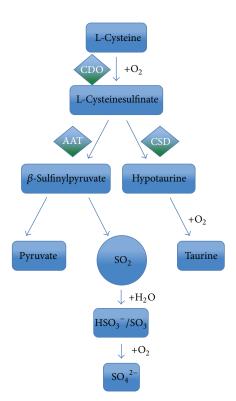


FIGURE 1: Endogenous generation of sulfur dioxide. Firstly, a sulfur-containing amino acid is metabolized to L-cysteine that is then oxidated into L-cysteinesulfinate under the role of CDO. L-Cysteinesulfinate, an analogue of L-asparaginic acid, can be transaminated into  $\beta$ -sulfinylpyruvate under the role of AAT, which is spontaneously decomposed into pyruvate and SO<sub>2</sub>. *In vivo*, SO<sub>2</sub> can produce HSO<sub>3</sub><sup>-/</sup>SO<sub>3</sub><sup>2-</sup> (molar ratio of 1:3) after combining with water, which can be oxidated into SO<sub>4</sub><sup>2-</sup> by sulfite oxidase and then excreted through the kidneys. On the other hand, L-cysteinesulfinate can be also decarboxylated into CO<sub>2</sub> and hypotaurine under the role of CSD. A large majority of hypotaurine can be further oxidized into taurine. SO<sub>2</sub>: sulfur dioxide; CDO: cysteine dioxygenase; CSD: cysteinesulfinate decarboxylase; AAT: aspartate aminotransferase.

caudal artery,  $(112\pm15 \text{ U/g})$  in mesenteric artery,  $(96\pm12 \text{ U/g})$  in pulmonary artery, and  $(88 \pm 11 \text{ U/g})$  in the aorta. Serum AAT activity was  $(87 \pm 18 \text{ U/g})$ . The distribution of AAT mRNA levels was consistent with AAT activity in rats. In heart, liver, kidney, and other tissues, AAT mRNA levels were significantly higher than those found in the artery [3].

The synthesis of SO<sub>2</sub> is regulated by a variety of factors. Balazy et al. [24] showed that acetylcholine (Ach) could cause an increased synthesis of SO<sub>2</sub> in porcine coronary artery. However, Meng et al. [27] found that endogenous Ach could promote the generation of SO<sub>2</sub> in rat vascular tissue in cultured vascular endothelial and VSMCs in a dose-dependent manner, but noradrenaline (NE) suppressed the generation of SO<sub>2</sub>. Factors regulating SO<sub>2</sub> generation need further studies.

#### 3. Vascular Structural Remodeling

In 1989, Baumbach and Heistad [28] first proposed the concept of "vascular structural remodeling" when studying

the changes in cerebral artery of rats with chronic hypertension. With the wide use of pathological morphology, microstructure morphologic metrology, vascular perfusion in vitro, and other technologies, the study on VSR has been deepened. VSR is defined as any structural changes of blood vessels [29]. The most typical characteristics of VSR include intimal endothelial cell swelling, medial SMC hyperplasia, and adventitia extracellular matrix (ECM) deposition [29]. Gibbons and Dzau [29] pointed out that VSR was a dynamic process; it relied on the interaction among haemodynamics, other mechanical stimuli with locally produced growth factors, including signal perception, signal transduction, and synthesis and release of regulatory factors. This eventually leads to changes in the structure of the vessel wall, simultaneously accompanied by decreased vascular wall compliance, changes in the release of vasoactive substances, and other functional disorders.

## 4. SO<sub>2</sub> and Vascular Remodeling in Hypertension

4.1. The Role of  $SO_2$  in the Development of Vascular Structural Remodeling in Systemic Hypertension. VSR is one of the important issues in pathogenesis of hypertension [30]. The characteristics of VSR in the process of hypertension include the hyperplastic medial SMCs, the swollen endothelial cells, and the increased ECM [31, 32]. Dzau et al. [33] summarized it up to four types: (1) both the media and intima are thickened; the inner diameter is shortened, resulting in an increased ratio of vessel wall thickness to inner diameter. This type of change is mainly due to the hypertrophy and proliferation of VSMCs. Or the proliferation may not be obvious; however, rearrangement of VSMCs and noncellular components may be significant; (2) both the inner and outer diameter are increased, suggesting that the hypertrophy of vessel wall is relatively mild. In this case, the ratio of vessel wall thickness to inner diameter is decreased. This change is due to the rearrangement of VSMCs and the proliferation is not obvious; (3) both the inner and outer diameter are decreased; and (4) the number of microcirculations is decreased.

 $SO_2$  plays an important role in the development of VSR in hypertension. In spontaneously hypertensive rats (SHRs), Zhao et al. [34] found that  $SO_2$  content and AAT activity in plasma and thoracic aorta were significantly decreased, but the ratio of vessel wall thickness and inner diameter was significantly increased, along with the accumulation of collagen I and collagen III in the aorta. However, intraperitoneal injection of  $SO_2$  derivatives (Na<sub>2</sub>SO<sub>3</sub>/NaHSO<sub>3</sub>, 0.54 mmol/kg: 0.18 mmol/kg, for five weeks) could significantly reduce the VSR and lower the blood pressure of SHRs. The results indicated that the downregulated endogenous  $SO_2$  pathway was involved in the VSR in hypertension.

Excessive proliferation of VSMCs is one of the key mechanisms involved in hypertensive VSR. Under the hypertensive condition, VSMCs switch from a contractile to a synthetic phenotype, with a high rate of proliferation. The abnormal proliferation of VSMCs leads to a variety of cytokines secretion and a marked ECM production, which finally results in a decrease of lumen ratio and an increase of media-lumen ratio

[35]. Zhao et al. [34] found that, in SHRs, aortic tunica media thickness and VSMCs proliferation index were significantly decreased after the exogenous supplement of SO<sub>2</sub> derivatives (0.54 mmol/kg: 0.18 mmol/kg, for five weeks). Liu et al. [36] found that SO<sub>2</sub> inhibited vascular proliferation by suppressing the progression of the cell cycle from G1 to S phase. Going further, AAT1 and AAT2 overexpression inhibited seruminduced VSMCs proliferation of rats, whereas AAT1 and AAT2 knockdown showed an opposite effect. These results showed that endogenous SO<sub>2</sub> had a negative effect on VSMCs proliferation. For the purpose of further exploring the potential mechanisms, a cell proliferation model stimulated by platelet derived growth factor-BB (PDGF-BB) was established. It was found that SO<sub>2</sub> dephosphorylated the active sites of ERK1/2, MAPK kinase 1/2, and c-Raf. Furthermore, SO<sub>2</sub> increased the AC activity of VSMCs of rats, thereby increasing the intracellular cAMP levels and activating PKA signaling molecules, which resulted in an increase in the Ser259 phosphorylation, an inhibitory site of Raf-1 molecule, thereby inhibiting the Raf-1 kinase activity and leading to the inactivation of the ERK/MAPK pathway [36]. The results suggested that SO<sub>2</sub> could attenuate VSR through suppressing proliferation of VSMCs. And cAMP/PKA/ERK/MAPK signaling was involved in the inhibitory effect of SO<sub>2</sub> on VSMC proliferation.

Numerous studies indicated that the VSMC apoptosis participated in the development of VSR. Zhao et al. [37, 38] found that, in SHRs, the exogenous supplement of SO<sub>2</sub> derivatives (0.54 mmol/kg: 0.18 mmol/kg, for five weeks) could promote apoptosis of VSMCs, and its mechanism might be related to inhibited Bcl-2 and activated Fas, ultimately inducing apoptosis through the molecule caspase-3. These data suggested that SO<sub>2</sub> inhibited VSR in hypertension in association with promoting VSMC apoptosis.

4.2. The Role of  $SO_2$  in the Development of Vascular Structural Remodeling in Pulmonary Artery Hypertension. Pulmonary artery hypertension (PAH) is a pathophysiologic syndrome that leads to pulmonary vascular bed obstruction and progressively increased pulmonary vascular resistance, ultimately resulting in right heart failure. It is well known that pulmonary VSR contributes to all types of pulmonary hypertension. The main pathologic characteristics of pulmonary VSR in pulmonary artery hypertension include thickening of the adventitial, medial and/or intimal layer of the pulmonary arteries, elevated stiffening of the elastic pulmonary arteries, vascular bed occlusive lesions, proliferation of pulmonary fibroblasts, SMCs and endothelial cells, and excessive accumulation of ECM [39]. Interestingly, SO<sub>2</sub> plays an important role in the development of pulmonary artery hypertension, including hypoxic pulmonary hypertension, high pulmonary blood flow-induced pulmonary hypertension, and monocrotaline-induced pulmonary hypertension.

4.2.1. Downregulation of Endogenous SO<sub>2</sub> Pathway Is Involved in the Development of Pulmonary VSR in Hypoxic Pulmonary Hypertension. Hypoxic pulmonary vasoconstriction caused by acute hypoxia and hypoxic pulmonary vascular structural

remodeling caused by chronic hypoxia are the main pathophysiological processes of hypoxic pulmonary hypertension [40]. In comparison to the response of systemic blood vessels, the response of pulmonary vasculature to hypoxia has its specific characteristics; that is, pulmonary vascular reactive constriction in hypoxic condition can help maintain arterial oxygen saturation, which has an important physiological significance. However, if hypoxia persists, pulmonary arteriole will develop structural remodeling and pulmonary artery pressure will stay at a high level. And pulmonary hypertension will lead to aggravated hypoxia, and form a vicious circle. Interestingly, SO<sub>2</sub> content in the plasma and lung tissues from rats with hypoxic pulmonary hypertension was significantly decreased, as well as AAT1 mRNA expression and AAT activity [11]. In order to examine the significance of downregulated SO<sub>2</sub> pathway in the development of hypoxic pulmonary hypertension, investigators administered SO<sub>2</sub> derivatives to the hypoxic rats (0.54 mmol/kg: 0.18 mmol/kg, for three weeks). The results showed that it could significantly reduce mean pulmonary arterial pressure of the hypoxic rat [11]. Meanwhile, SO<sub>2</sub> derivatives significantly alleviated hypoxic pulmonary VSR, as demonstrated by a reduced percentage of muscular arteries and an increased percentage of nonmuscular arteries. Moreover, the relative medial thickness and relative medial areas of muscular arteries were decreased in hypoxic rats after SO<sub>2</sub> treatment. The above results implied that the reduction of endogenous SO<sub>2</sub> was involved in the development of hypoxic pulmonary VSR.

The studies have shown the advances in the mechanisms by which SO<sub>2</sub> plays a crucial role in the development of hypoxic pulmonary VSR. The deposition of ECM, including collagen, elastin, proteoglycans, and glycoproteins, participates in pulmonary VSR. Collagen deposition plays a crucial role in VSR. Exogenous supplementation of SO<sub>2</sub> derivatives could alleviate pulmonary VSR and attenuate pulmonary hypertension. Furthermore, SO<sub>2</sub> could significantly reduce the mRNA expressions of procollagens I and III in hypoxic pulmonary hypertension rats [11]. On the contrary, the mRNA levels of procollagens I and III were markedly increased in hypoxic pulmonary hypertensive rats administrated with hydroxamate (HDX), an inhibitor of AAT. The results indicated that SO<sub>2</sub> inhibited hypoxic hypertensioninduced pulmonary VSR possibly by the suppression of collagen deposition. It is known that reduced collagen degradation has an important significance in pulmonary hypertensive VSR. It is regulated by the balance between matrix metalloproteinase (MMP) and the tissue inhibitor of metalloproteinase (TIMP). Zaidi et al. [41] found that MMPs activity was increased in the development of hypoxic pulmonary hypertension. Vieillard-Baron et al. [42] transferred the TIMP-1 gene into the lungs of rats by adenovirus, which successfully inhibited the expression and activity of MMPs, preventing from hypoxic pulmonary hypertension and pulmonary VSR. SO2 was found to increase the mRNA ratio of MMP-13/TIMP-1 in pulmonary arteries of hypoxic pulmonary hypertensive rats by increasing MMP-13 mRNA level and decreasing TIMP-1 mRNA expression [11]. These data suggested that SO<sub>2</sub> attenuated hypoxic pulmonary VSR at least partly by reducing collagen degradation.

Additionally, SO<sub>2</sub> inhibited pulmonary VSMCs proliferation in association with the suppression of Raf-1 protein and the downstream ERK/MAPK pathway in hypoxic pulmonary hypertensive rats. Moreover, Bai and Meng [43] found that the exogenous inhalation of SO<sub>2</sub> downregulated mRNA levels of Bcl-2, increased protein expressions of p53 and Bax, and enhanced caspase-3 activity in rat lung, suggesting that SO<sub>2</sub> alleviated hypoxic pulmonary VSR by maintaining the balance between pulmonary vascular cell proliferation and apoptosis.

4.2.2. Downregulated Endogenous SO<sub>2</sub> Pathway Is Involved in the Development of Pulmonary VSR in High Pulmonary Blood Flow-Induced Pulmonary Hypertension. Pulmonary hypertension induced by high pulmonary blood flow is one of the most common complications of left-to-right shunt congenital heart disease. Increased pulmonary blood flow could result in pulmonary VSR and finally the development of pulmonary hypertension. Pronounced medial thickening and increased collagen content in pulmonary arteries were observed in cases of congenital heart disease accompanied with pulmonary hypertension [44]. Luo et al. [45] developed the rat model of high pulmonary blood flow-induced pulmonary hypertension by systemic-pulmonary shunting. They found that the content of SO2, the mRNA, and protein expression of AAT2 as well as AAT activity in pulmonary vessels were decreased in pulmonary hypertensive rats. The administration of SO<sub>2</sub> derivatives (0.54 mmol/kg: 0.18 mmol/kg, for eight weeks) could significantly reduce the mean pulmonary artery pressure and improve the pulmonary vascular pathological changes of the rats, as demonstrated by the decreased percentage of muscularized pulmonary arteries. Furthermore, Liu et al. [46] found that SO<sub>2</sub> alleviated the protein expression of collagen I and collagen III.

However, the mechanisms by which SO<sub>2</sub> alleviates VSR are incompletely understood. It was suggested that endogenous SO<sub>2</sub> might alleviate pulmonary VSR via upregulating the reduced endogenous H<sub>2</sub>S pathway [45, 47, 48]. To further investigate the signaling pathway by which SO<sub>2</sub> alleviated pulmonary VSR, Liu et al. used a Flexcell Fx-5000 Tension System to establish a cell model to mimic the mechanical stretching of high blood flow on vascular wall in high blood flow induced pulmonary hypertension. They found that mechanical stretching could downregulate endogenous SO<sub>2</sub>/AAT1 pathway in pulmonary fibroblasts (PAFs) and then activate TGF- $\beta$ 1/Smad2/3 pathway, which ultimately resulted in an excessive collagen synthesis. Overexpression of AAT1, however, could antagonize the activation of TGF- $\beta$ 1/Smad2/3 pathway caused by mechanical stretching, which could be exaggerated by knockdown of AAT1. Furthermore, in the rat model of high pulmonary blood flow-induced pulmonary hypertension by surgical systemic-pulmonary shunting, the activated TGF- $\beta$ 1/Smad2/3 pathway could be inhibited by the administration of SO<sub>2</sub> derivatives [46]. The above results suggested that in high pulmonary blood flow-induced pulmonary hypertension the downregulated endogenous SO<sub>2</sub> activated TGF-β1/Smad2/3 pathway, which ultimately resulted in collagen remodeling.

4.2.3. Upregulated Endogenous SO<sub>2</sub> Pathway Is Involved in the Development of Pulmonary VSR in Monocrotaline-Induced Pulmonary Hypertension. Monocrotaline (MCT), a pyrrolizidine alkaloid, is metabolized into MCT pyrrole in the liver. The substance of MCT induces pulmonary hypertension in a rat model by causing pulmonary artery smooth muscle hypertrophy, inflammation, and endothelial cells injury. Mean pulmonary artery pressure (mPAP) and the ratio of right ventricle to left ventricle plus septum were increased markedly in the MCT-treated rats [12]. Meanwhile, pulmonary VSR developed accompanied with the increased SO<sub>2</sub> level, AAT activity, and mRNA expression. SO<sub>2</sub> administration significantly alleviated pulmonary VSR and reduced mPAP, while HDX aggravated them in the MCT-treated rats [12]. It suggested that endogenous SO<sub>2</sub> might play a protective role in the MCT-induced pulmonary VSR in pulmonary hypertensive rats.

Studies have suggested that enhanced oxidative stress by increasing superoxide anions and other reactive oxygen species production was involved in the pathophysiology of MCT-induced pulmonary hypertensive VSR [49, 50]. Superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), glutathione (GSH), and catalase (CAT) are antioxidant enzymes, and the malondialdehyde (MDA) is oxidation product. Although inhalation of SO<sub>2</sub> was considered to cause oxidative damage to mammals [51], the endogenous SO<sub>2</sub> at a low level was found to have an antioxidant effect in MCT-induced pulmonary hypertension. In MCT-induced pulmonary hypertensive rats [12], the activities of SOD, GSH-Px, GSH, CAT, and MDA were elevated in lung tissues in association with a protective upregulated SO<sub>2</sub> level. With supplement of SO<sub>2</sub> derivatives, the content of SOD, GSH-Px, and CAT went higher, while inhibition of endogenous SO<sub>2</sub> with HDX suppressed the activities of SOD and CAT [12]. These data indicate that upregulated SO<sub>2</sub> production plays a protective role in pulmonary VSR by promoting endogenous antioxidative capacity.

#### 5. Conclusion

 $SO_2$  was previously recognized as an industrial waste gas. Environmental  $SO_2$  can cause oxidative damage to the cardiovascular system, respiratory system, and other systems [51].  $SO_2$  and sulphites can cause DNA damage in mammalian cells, such as chromatin breakage, sister chromatid exchange, micronucleus formation, DNA-protein cross-linking, and other stages [52, 53]. In recent years, however,  $SO_2$  as discussed herein is found to have the characteristics of gaseous molecules such as endogenous continuous generation, fast transmission, extensive action, and low molecular weight and therefore play an important role in the physiology and pathophysiology of cardiovascular diseases.

VSR is an important pathogenic base of cardiovascular diseases, such as systemic hypertension and pulmonary hypertension. In the process of VSR in systemic hypertension or pulmonary hypertension, VSMC proliferation is excessive while apoptosis reduced, and extracellular collagen synthesis and degradation are imbalanced, leading to an excessive deposition of collagen in the vascular wall.

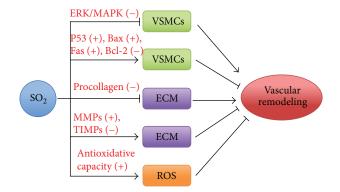


FIGURE 2: Mechanisms by which  $SO_2$  attenuates VSR.  $SO_2$  attenuates VSR by inhibiting proliferation of VSMCs, promoting apoptosis, inhibiting the synthesis of extracellular collagen, promoting the degradation of extracellular collagen, and increasing the antioxidative capacity.  $SO_2$ : sulfur dioxide; VSMCs: vascular smooth muscle cells; ECM: extracellular matrix; ROS: reactive oxygen species.

Although the increase in collagen content may help to resist the excessive stress in the vessel wall and maintain the integrity of blood vessels, the excessive collagen deposition can reduce the compliance and increase the resistance of the vascular wall, which has become an important factor in deterioration of systemic hypertension or pulmonary hypertension. Since VSR is a core pathological change of systemic hypertension and pulmonary hypertension, it is important to further explore its mechanisms in order to better understand its pathogenesis, which would ultimately provide scientific basis for potential therapeutic targets. In vitro and in vivo experiments demonstrated that downregulated SO<sub>2</sub>/AAT pathway was involved in VSR of systemic hypertension and pulmonary hypertension. Furthermore, SO<sub>2</sub> has been shown to play an important regulatory role in VSR by inhibiting VSMC proliferation and the synthesis of extracellular collagen and promoting VSMC apoptosis and collagen degradation. In addition, SO<sub>2</sub> can also enhance antioxidative capacity, thereby playing a significant role in alleviating VSR (Figure 2). Hence, the regulatory role of endogenous SO<sub>2</sub> in VSR would help to further understand the pathogenesis and the potential therapeutic targets of VSR in systemic hypertension and pulmonary hypertension.

However, in the present studies, there are still some limitations. (1) In some studies, investigators used exogenous  $SO_2$ derivatives to increase the level of  $SO_2$  in rats or cells. Increasing endogenous  $SO_2$  content by overexpressing AAT may be more in line with physiology. In addition, the AAT inhibitor HDX used to lower  $SO_2$  content was not specific. Knocking down AAT to reduce endogenous  $SO_2$  level together with a rescue experiment is likely a more convincing way to study the significance of endogenous  $SO_2$ . (2) As a gaseous molecule, the mechanisms by which  $SO_2$  exerts its function need to be further explained. (3) Previous studies showed that gaseous molecules, such as NO, CO, H<sub>2</sub>S, and  $SO_2$ , played an important role in the pathogenesis of VSR [54–57]. However, whether there are any interactions among these gaseous molecules when they function remains to be further studied. (4) Effective treatment for hypertension and pulmonary hypertension is the clinical problem to be solved. Since the regulatory role of VSR in systemic hypertension and pulmonary hypertension is gaining more and more interest, the treatment of VSR has become a new area of the diseases. The regulation of endogenous SO2 on VSR would likely become a new therapeutic target. Recently, benzothiazole sulfinate (BTS), a water-soluble SO<sub>2</sub> donor, was reported to exhibit slow and pH-dependent SO<sub>2</sub> release ability in aqueous solutions. And it also had SO<sub>2</sub>-like vasorelaxant effect on rat aorta rings [58]. Although the biological activities and validity of BTS still need more exploration, the discovery of potentially better donors would undoubtedly provide a great help and potential for the future clinical application of SO<sub>2</sub>. Meanwhile, although the in vivo experiments suggested that the exogenous supplement of SO<sub>2</sub> derivatives could play a protective role in hypertension and pulmonary hypertension, it still requires more validation in clinical trials to confirm its effects. Furthermore, there are still important things to note: (1) the suitable dosage. Although the current dosage of SO<sub>2</sub> derivatives (Na<sub>2</sub>SO<sub>3</sub>/NaHSO<sub>3</sub>) used in animal model is 0.54 mmol/kg: 0.18 mmol/kg, we should still take into account the difference between rats and human beings; (2) the potential biomarker. Since the studies suggest that the development of hypertension and pulmonary hypertension is associated with alteration of SO<sub>2</sub>/AAT pathway, it is speculated whether SO<sub>2</sub> or AAT would become a biomarker for the diseases. Of course, multiple clinical works need to be done to investigate the relationship between endogenous SO<sub>2</sub>/AAT pathway and the severity or survival rate of the diseases.

In summary, SO<sub>2</sub> inhibited VSMC proliferation, promoted VSMC apoptosis, inhibited the synthesis of extracellular collagen but promoted its degradation, and enhanced antioxidative capacity, thereby playing a significant role in attenuating VSR, which ultimately alleviated systemic hypertension and pulmonary hypertension. Therefore, the understanding of the relationship between endogenous SO<sub>2</sub> and VSR offers important insight into the pathogenesis of VSR and provides a potential therapeutic target of systemic hypertension and pulmonary hypertension. In the future, with the depth of the study, we are confident to believe that greater progress would be made in the field of SO<sub>2</sub> biology and medicine to improve the prognosis of disease and enhance the life quality of patients.

#### **Competing Interests**

The authors declare that they have no competing interests.

## **Authors' Contributions**

Jia Liu and Yaqian Huang contributed equally to this work.

#### Acknowledgments

This work was supported by the National Natural Science Foundation of China (81400311, 31440052, and 91439110).

#### References

- R. Wang, "The gasotransmitter role of hydrogen sulfide," Antioxidants and Redox Signaling, vol. 5, no. 4, pp. 493–501, 2003.
- [2] D. E. Koshland Jr., "The molecule of the year," *Science*, vol. 258, no. 5090, p. 1861, 1992.
- [3] L. Luo, S. Chen, H. Jin, C. Tang, and J. Du, "Endogenous generation of sulfur dioxide in rat tissues," *Biochemical and Biophysical Research Communications*, vol. 415, no. 1, pp. 61–67, 2011.
- [4] S.-X. Du, H.-F. Jin, D.-F. Bu et al., "Endogenously generated sulfur dioxide and its vasorelaxant effect in rats," *Acta Pharmacologica Sinica*, vol. 29, no. 8, pp. 923–930, 2008.
- [5] Z. Meng and H. Zhang, "The vasodilator effect and its mechanism of sulfur dioxide-derivatives on isolated aortic rings of rats," *Inhalation Toxicology*, vol. 19, no. 11, pp. 979–986, 2007.
- [6] Y. K. Wang, A. J. Ren, X. Q. Yang et al., "Sulfur dioxide relaxes rat aorta by endothelium-dependent and -independent mechanisms," *Physiological Research*, vol. 58, no. 4, pp. 521–527, 2009.
- [7] S. Zhang, J. Du, H. Jin et al., "Endogenous sulfur dioxide aggravates myocardial injury in isolated rat heart with ischemia and reperfusion," *Transplantation*, vol. 87, no. 4, pp. 517–524, 2009.
- [8] S. S. Haider, "Effects of exhaust pollutant sulfur dioxide on lipid metabolism of guinea pig organs," *Industrial Health*, vol. 23, no. 2, pp. 81–87, 1985.
- [9] M. R. Lovati, C. Manzoni, M. Daldossi, S. Spolti, and C. R. Sirtori, "Effects of sub-chronic exposure to SO<sub>2</sub> on lipid and carbohydrate metabolism in rats," *Archives of Toxicology*, vol. 70, no. 3-4, pp. 164–173, 1996.
- [10] W. Li, C. Tang, H. Jin, and J. Du, "Regulatory effects of sulfur dioxide on the development of atherosclerotic lesions and vascular hydrogen sulfide in atherosclerotic rats," *Atherosclerosis*, vol. 215, no. 2, pp. 323–330, 2011.
- [11] Y. Sun, Y. Tian, M. Prabha et al., "Effects of sulfur dioxide on hypoxic pulmonary vascular structural remodeling," *Laboratory Investigation*, vol. 90, no. 1, pp. 68–82, 2010.
- [12] H.-F. Jin, S.-X. Du, X. Zhao et al., "Effects of endogenous sulfur dioxide on monocrotaline-induced pulmonary hypertension in rats," *Acta Pharmacologica Sinica*, vol. 29, no. 10, pp. 1157–1166, 2008.
- [13] S. Chen, S. Zheng, Z. Liu et al., "Endogeous sulfur dioxide protects against oleic acid-induced acute lung injury in association with inhibition of oxidative stress in rats," *Laboratory Investigation*, vol. 95, no. 2, pp. 142–156, 2015.
- [14] T.-M. Chen, J. Gokhale, S. Shofer, and W. G. Kuschner, "Outdoor air pollution: nitrogen dioxide, sulfur dioxide, and carbon monoxide health effects," *The American Journal of the Medical Sciences*, vol. 333, no. 4, pp. 249–256, 2007.
- [15] M. H. Stipanuk, "Metabolism of sulfur-containing amino acids," Annual Review of Nutrition, vol. 6, pp. 179–209, 1986.
- [16] M. Bufalini, "Oxidation sulfur dioxide in polluted atmospheres. Review," *Environmental Science & Technology*, vol. 5, no. 8, pp. 685–700, 1971.
- [17] T. P. Singer and E. B. Kearney, "Intermediary metabolism of Lcysteinesulfinic acid in animal tissues," *Archives of Biochemistry* and Biophysics, vol. 61, no. 2, pp. 397–409, 1956.
- [18] O. W. Griffith, "Cysteinesulfinate metabolism: altered partitioning between transamination and decarboxylation following administration of β-methyleneaspartate," *The Journal of Biological Chemistry*, vol. 258, no. 3, pp. 1591–1598, 1983.

- [19] R. Shapiro, "Genetic effects of bisulfite (sulfur dioxide)," *Muta-tion Research*, vol. 39, no. 2, pp. 149–175, 1977.
- [20] J. Albrecht and A. Schousboe, "Taurine interaction with neurotransmitter receptors in the CNS: an update," *Neurochemical Research*, vol. 30, no. 12, pp. 1615–1621, 2005.
- [21] H. K. Kimelberg, N. B. Nestor, and P. J. Feustel, "Inhibition of release of taurine and excitatory amino acids in ischemia and neuroprotection," *Neurochemical Research*, vol. 29, no. 1, pp. 267–274, 2004.
- [22] P. Kamoun, "Endogenous production of hydrogen sulfide in mammals," *Amino Acids*, vol. 26, no. 3, pp. 243–254, 2004.
- [23] H. Mitsuhashi, S. Yamashita, H. Ikeuchi et al., "Oxidative stressdependent conversion of hydrogen sulfide to sulfite by activated neutrophils," *Shock*, vol. 24, no. 6, pp. 529–534, 2005.
- [24] M. Balazy, I. A. Abu-Yousef, D. N. Harpp, and J. Park, "Identification of carbonyl sulfide and sulfur dioxide in porcine coronary artery by gas chromatography/mass spectrometry, possible relevance to EDHF," *Biochemical and Biophysical Research Communications*, vol. 311, no. 3, pp. 728–734, 2003.
- [25] T. D. Scholz, S. L. Koppenhafer, C. J. TenEyck, and B. C. Schutte, "Ontogeny of malate-aspartate shuttle capacity and gene expression in cardiac mitochondria," *American Journal of Physiology -Cell Physiology*, vol. 274, no. 3, pp. C780–C788, 1998.
- [26] R. J. DeLorenzo and F. H. Ruddle, "Glutamate oxalate transaminase (GOT) genetics in Mus musculus: linkage, polymorphism, and phenotypes of the Got-2 and Got-1 loci," *Biochemical Genetics*, vol. 4, no. 2, pp. 259–273, 1970.
- [27] Z. Meng, J. Li, Q. Zhang et al., "Vasodilator effect of gaseous sulfur dioxide and regulation of its level by Ach in rat vascular tissues," *Inhalation Toxicology*, vol. 21, no. 14, pp. 1223–1228, 2009.
- [28] G. L. Baumbach and D. D. Heistad, "Remodeling of cerebral arterioles in chronic hypertension," *Hypertension*, vol. 13, no. 6, pp. 968–972, 1989.
- [29] G. H. Gibbons and V. J. Dzau, "The emerging concept of vascular remodeling," *The New England Journal of Medicine*, vol. 330, no. 2, pp. 1431–1438, 1994.
- [30] M. Ponticos and B. D. Smith, "Extracellular matrix synthesis in vascular disease: hypertension, and atherosclerosis," *Journal of Biomedical Research*, vol. 28, no. 1, pp. 25–39, 2014.
- [31] M. P. Jacob, C. Badier-Commander, V. Fontaine, Y. Benazzoug, L. Feldman, and J. B. Michel, "Extracellular matrix remodeling in the vascular wall," *Pathologie-Biologie*, vol. 49, no. 4, pp. 326– 332, 2001.
- [32] P. Brassard, F. Amiri, and E. L. Schiffrin, "Combined angiotensin II type 1 and type 2 receptor blockade on vascular remodeling and matrix metalloproteinases in resistance arteries," *Hypertension*, vol. 46, no. 3, pp. 598–606, 2005.
- [33] V. J. Dzau, G. H. Gibbons, R. Morishita, and R. E. Pratt, "New perspectives in hypertension research: potentials of vascular biology," *Hypertension*, vol. 23, no. 6, part 2, pp. 1132–1140, 1994.
- [34] X. Zhao, H.-F. Jin, C.-S. Tang, and J.-B. Du, "Effect of sulfur dioxide on vascular collagen remodeling in spontaneously hypertensive rats," *Zhonghua Er Ke Za Zhi*, vol. 46, no. 12, pp. 905–908, 2008.
- [35] K. R. Chien, Z. M. Liu, and D. F. Zhang, *Molecular Cardiology*, Public Health Publishing House, Beijing, China, 2002.
- [36] D. Liu, Y. Huang, D. Bu et al., "Sulfur dioxide inhibits vascular smooth muscle cell proliferation via suppressing the Erk/MAP kinase pathway mediated by cAMP/PKA signaling," *Cell Death* & Disease, vol. 5, Article ID e1251, 2014.

- [37] X. Zhao, H.-F. Jin, C.-S. Tang, and J.-B. Du, "Effects of sulfur dioxide, on the proliferation and apoptosis of aorta smooth muscle cells in hypertension: experiments with rats," *Zhonghua Yi Xue Za Zhi*, vol. 88, no. 18, pp. 1279–1283, 2008.
- [38] X. Zhao, H.-F. Jin, S.-X. Du, C.-S. Tang, and J.-B. Du, "The effect of sulfur dioxide on blood pressure and aortic structure of spontaneously hypertensive rat," *Chinese Pharmacological Bulletin*, vol. 24, no. 3, pp. 327–330, 2008.
- [39] L. A. Shimoda and S. S. Laurie, "Vascular remodeling in pulmonary hypertension," *Journal of Molecular Medicine*, vol. 91, no. 13, pp. 297–309, 2013.
- [40] R. L. Vender, "Chronic hypoxic pulmonary hypertension: cell biology to pathophysiology," *Chest*, vol. 106, no. 1, pp. 236–243, 1994.
- [41] S. H. E. Zaidi, X.-M. You, S. Ciura, M. Husain, and M. Rabinovitch, "Overexpression of the serine elastase inhibitor elafin protects transgenic mice from hypoxic pulmonary hypertension," *Circulation*, vol. 105, no. 4, pp. 516–521, 2002.
- [42] A. Vieillard-Baron, E. Frisdal, S. Eddahibi et al., "Inhibition of matrix metalloproteinases by lung TIMP-1 gene transfer or doxycycline aggravates pulmonary hypertension in rats," *Circulation Research*, vol. 87, no. 5, pp. 418–425, 2000.
- [43] J. Bai and Z. Meng, "Effects of sulfur dioxide on apoptosisrelated gene expressions in lungs from rats," *Regulatory Toxicology and Pharmacology*, vol. 43, no. 3, pp. 272–279, 2005.
- [44] E. S. Egito, V. D. Aiello, I. B. Bosisio et al., "Vascular remodeling process in reversibility of pulmonary arterial hypertension secondary to congenital heart disease," *Pathology, Research and Practice*, vol. 199, no. 8, pp. 521–532, 2003.
- [45] L. Luo, D. Liu, C. Tang et al., "Sulfur dioxide upregulates the inhibited endogenous hydrogen sulfide pathway in rats with pulmonary hypertension induced by high pulmonary blood flow," *Biochemical and Biophysical Research Communications*, vol. 433, no. 4, pp. 519–525, 2013.
- [46] J. Liu, W. Yu, Y. Liu et al., "Mechanical stretching stimulates collagen synthesis via down-regulating SO<sub>2</sub>/AAT1 pathway," *Scientific Reports*, vol. 6, article 21112, 2016.
- [47] X. Li, J. Du, H. Jin, B. Geng, and C. Tang, "Sodium hydrosulfide alleviates pulmonary artery collagen remodeling in rats with high pulmonary blood flow," *Heart and Vessels*, vol. 23, no. 6, pp. 409–419, 2008.
- [48] W. Li, H.-F. Jin, D. Liu et al., "Hydrogen sulfide induces apoptosis of pulmonary artery smooth muscle cell in rats with pulmonary hypertension induced by high pulmonary blood flow," *Chinese Medical Journal*, vol. 122, no. 24, pp. 3032–3038, 2009.
- [49] E. Hironaka, M. Hongo, A. Sakai et al., "Serotonin receptor antagonist inhibits monocrotaline-induced pulmonary hypertension and prolongs survival in rats," *Cardiovascular Research*, vol. 60, no. 3, pp. 692–699, 2003.
- [50] J. P. Versluis, J. W. Heslinga, P. Sipkema, and N. Westerhof, "Contractile reserve but not tension is reduced in monocrotaline-induced right ventricular hypertrophy," *American Journal of Physiology—Heart and Circulatory Physiology*, vol. 286, no. 3, pp. H979–H984, 2004.
- [51] Z. Meng, "Oxidative damage of sulfur dioxide on various organs of mice: sulfur dioxide is a systemic oxidative damage agent," *Inhalation Toxicology*, vol. 15, no. 2, pp. 181–195, 2003.
- [52] Z. Meng, G. Qin, B. Zhang, and J. Bai, "DNA damaging effects of sulfur dioxide derivatives in cells from various organs of mice," *Mutagenesis*, vol. 19, no. 6, pp. 465–468, 2004.

- [53] Z. Meng, G. Qin, and B. Zhang, "DNA damage in mice treated with sulfur dioxide by inhalation," *Environmental and Molecular Mutagenesis*, vol. 46, no. 3, pp. 150–155, 2005.
- [54] D. Junbao, Y. Hui, W. Bing, L. Jian, Q. Jianguang, and T. Chaoshu, "Effect of L-arginine on collagen of high flow-induced pulmonary arterial remodeling," *Circulation Journal*, vol. 69, no. 5, pp. 603–608, 2005.
- [55] H. Gompf, F. C. Luft, and I. Morano, "Nitric oxide synthase upregulation and the predelivery blood pressure decrease in spontaneously hypertensive rats," *Journal of Hypertension*, vol. 20, no. 2, pp. 255–261, 2002.
- [56] V. Hampl, J. Bíbová, A. Baňasová et al., "Pulmonary vascular iNOS induction participates in the onset of chronic hypoxic pulmonary hypertension," *American Journal of Physiology— Lung Cellular and Molecular Physiology*, vol. 290, no. 1, pp. L11– L20, 2006.
- [57] X. Zhao, L. K. Zhang, C. Y. Zhang et al., "Regulatory effect of hydrogen sulfide on vascular collagen content in spontaneously hypertensive rats," *Hypertension Research*, vol. 31, no. 8, pp. 1619–1630, 2008.
- [58] J. J. Day, Z. Yang, W. Chen, A. Pacheco, and M. Xian, "Benzothiazole sulfinate: a water-soluble and slow-releasing sulfur dioxide donor," ACS Chemical Biology, vol. 11, no. 6, pp. 1647– 1651, 2016.





**The Scientific** World Journal



Research and Practice









Computational and Mathematical Methods in Medicine

Behavioural Neurology





Oxidative Medicine and Cellular Longevity