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Review Article

The Role of *H. pylori* CagA in Regulating Hormones of Functional Dyspepsia Patients

Wang-Ping Meng,¹ Zhong-Qiong Wang,¹ Jia-Qi Deng,² Yi Liu,¹ Ming-Ming Deng,¹ and Mu-Han Lü¹

¹Department of Gastroenterology, The Affiliated Hospital of Southwest Medical University, Luzhou, Sichuan Province 646000, China ²School of Foreign Languages, Southwest Medical University, Luzhou, Sichuan Province 646000, China

Correspondence should be addressed to Ming-Ming Deng; dengmm_ing@126.com and Mu-Han Lü; lvmuhan990@163.com

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Helicobacter pylori (H. pylori, Hp) colonizes the stomachs of approximately 20%–80% of humans throughout the world. The Word Healthy Organization (WHO) classified H. pylori as a group 1 carcinogenic factor in 1994. Recently, an increasing number of studies has shown an association between H. pylori infection and various extragastric diseases. Functional dyspepsia (FD) is considered a biopsychosocial disorder with multifactorial pathogenesis, and studies have shown that infection with CagA-positive H. pylori strains could explain some of the symptoms of functional dyspepsia. Moreover, CagA-positive H. pylori strains have been shown to affect the secretion of several hormones, including 5-HT, ghrelin, dopamine, and gastrin, and altered levels of these hormones might be the cause of the psychological disorders of functional dyspepsia patients. This review describes the mutual effects of H. pylori and hormones in functional dyspepsia and provides new insight into the pathogenesis of functional dyspepsia.

1. Introduction

Functional dyspepsia (FD), a very common condition that impairs quality of life, is a relapsing and remitting disorder with various chronic symptoms referable to the gastroduodenal region, including typical abdominal bloating or pain, early satiety, belching, heartburn, and nausea in the absence of organic or metabolic disease [1]. There are several diagnostic categories for FD based on the Rome III criteria, which are epigastric pain syndrome (EPS), postprandial discomfort syndrome (PDS), and a combination of these symptoms. The pathogenesis of FD is considered to be multifactorial or even a biopsychosocial disorder that causes abnormal gastrointestinal motility, visceral hypersensitivity, vagal dysfunction, and probable central nervous system disturbance [2]. Currently, Helicobacter pylori (H. pylori, Hp) infection is considered the major cause of the chronic gastric inflammation of FD patients [3]. Gastric inflammation has been shown to affect motor function and visceral sensitivity in experimental models. H. pylori strains that express CagA may be responsible for the FD associated with the more severe

forms of gastritis. It was reported that CagA-positive *H. pylori* strains induced more dyspeptic symptoms than CagA-negative or *H. pylori*-negative strains in patients with FD [4]. Some of these functional symptoms can be explained by a gut-driven brain disorder [5], and in vivo hormonal changes have also been implicated. However, no definite clinical manifestation is linked to CagA-positive *H. pylori* strains infection or fluctuating levels of hormones in FD patients. This review discusses the possible correlation between an infection with CagA-positive *H. pylori* strains and the levels of several hormones in FD patients.

2. CagA-Positive *H. pylori* Strains-Related Diseases (Figure 1)

2.1. Gastric Diseases. Infection with H. pylori is recognized as the greatest risk of chronic gastritis, peptic ulcers, mucosa-associated lymphoid tissue (MALT) lymphoma, and gastric adenocarcinoma. H. pylori secrete effector molecules to control the inflammatory, proliferative, and apoptotic processes

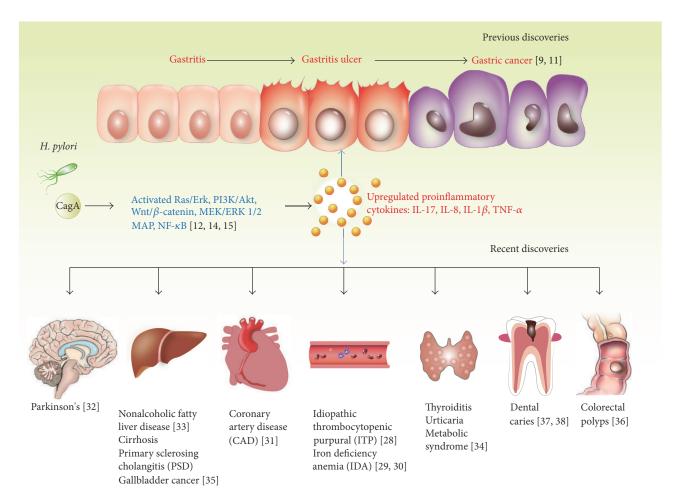


FIGURE 1: CagA-positive *H. pylori* strains-related diseases. According to previous research findings, infection with CagA-positive *H. pylori* strains is widely known as a higher risk of peptic ulcers and gastric adenocarcinoma. However, recent discoveries have shown that *H. pylori* is associated with some extragastric diseases, including cardiovascular, neurological, hematologic, metabolic, and dermatologic disorders. Inflammatory response and multiple signaling pathways might participate in mediating pathophysiological process.

of localized cells. Cytotoxin-associated protein A (CagA) is injected directly into the host epithelial cells via the type-four secretion system (T4SS), which is encoded by the Cag pathogenicity island (PAI) of *H. pylori* type I strains and associated with the development of gastric cancer. In the case of gastric MALT lymphoma, CagA was translocated into B-lymphoid cells and promoted their proliferation, possibly through the CagA-mediated proteins SHP-2, ERK, and MAPK, and increased the levels of Bcl-2 and Bcl-xL [6].

The relationship between infection with CagA-positive *H. pylori* strains and a higher risk of peptic ulcers and gastric adenocarcinoma in humans is widely known [7]. CagA is phosphorylated by host kinases, which alters cell signaling and various cellular responses involved in inflammation. Multiple oncogenic pathways were activated by CagA, such as the Ras/Erk, PI3K/Akt, and Wnt/beta-catenin pathways. Infection with CagA-positive *H. pylori* strains is the main factor driving the hyperactivity of the PI3K/Akt signaling pathway in gastric cancer, which is due to CagA-induced activation of the PI3K/Akt pathway, the representative downstream MEK/ERK pathway, and the nuclear factor-kappaB

(NF-kB) signaling pathway, which subsequently induces the nuclear translocation of beta-catenin [8]. As observed in human gastric mucosae infected by CagA-positive *H. pylori*, CagA activates the Wnt/beta-catenin signaling pathway and induces beta-catenin transcription [9]. These cellular responses and inflammation may be responsible for the increase in the level of p53. The level of this key tumor suppressor is increased following infection with CagA-positive *H. pylori* strains and decreased rapidly during *H. pylori* eradication [10], whereas a continuous bacterial infection caused a persistently high level of p53. This phenomenon may be driven by the DNA damage related to inflammatory processes [11].

In addition, *H. pylori* is known to activate the NF-kB signaling pathway. IkappaB kinase alpha (IKK alpha) is a critical regulator of NF-kB activity and *H. pylori* induces the nuclear translocation of IKK alpha, which is indispensable for an inflammatory response, through a Cag PAI-dependent manner [12]. A study explained that CagA could activate the NF-kB signaling pathway and induced the downstream release of IL-8 via the MEK/ERK signaling pathway [13].

Infection with CagA-positive H. pylori strains promotes inflammatory processes that result in neoplastic transformation [14]. The inflammatory response associated with CagA-positive H. pylori gastritis is due to the upregulated expression of proinflammatory cytokines, including tumor necrosis factor (TNF)- α , interferon gamma, interleukin- (IL-) 1beta and IL-8, and, particularly, IL-17, a key cell element in the inflammation caused by H. pylori, which mediates the activation of polymorphonuclear neutrophils [15]. Furthermore, IL-17 generally causes IL-8 secretion through the ERK 1/2 MAP kinase pathway [16]. Both CagA protein and Cag PAI have been shown to activate the NF-kB signaling pathway, which increased the level of IL-8 expression, but the effect of CagA in regulating NF-kB activation is still unclear. Recently, a study demonstrated that NF-kB activation and IL-8 release require the Cag PAI-encoded T4SS [17].

2.2. Extragastric Diseases. H. pylori has also been linked to some extragastric diseases, including cardiovascular, neurological, hematologic, metabolic, and dermatologic disorders, such as nonalcoholic fatty liver disease (NAFLD), gallbladder cancer, colorectal polyps, dental caries, metabolic syndrome, idiopathic thrombocytopenic purpura (ITP), iron deficiency anemia (IDA), coronary artery disease (CAD), and Parkinson's disease (PD) [18].

H. pylori eradication was shown to increase the platelet count in patients with ITP. The pathogenesis of ITP due to *H. pylori* infection is probably associated with variable host immune responses to VacA and CagA [19]. Recent guidelines indicated that IDA patients should be evaluated for an *H. pylori* type I strain infection because this bacterium can induce IDA through several mechanisms [20]. Active hemorrhaging caused by CagA-positive H. pylori gastritis or ulcers is known to be due to CagA increasing the level of transferrin, thus affecting iron acquisition [21]. Tamer et al. [22] have suggested that *H. pylori* may cause atherogenesis through persistent inflammation, and another possibility for this connection is the molecular mimicry of CagA. The persistence of serum CagA antibodies now appears to be predictive of Parkinson's disease with a poor prognosis; the proposed possible mechanism by which H. pylori causes this pathology is that it triggered mitochondrial damage and autoimmunity [23]. Moreover, H. pylori-like DNA is more commonly found in liver samples from chronic liver disease patients than from controls [24]. An H. pylori infection is positively correlated with metabolic syndrome and is inversely correlated with morbid obesity and type 2 diabetes mellitus (T2DM). The rate of seropositivity is higher in patients with metabolic syndrome than in healthy subjects [25]. Another study demonstrated that more than 75% of gallbladder cancer patients and 50% of chronic cholecystitis patients harbored H. pylori in the bile and gallbladder. An H. pylori infection was shown to aggravate gallbladder mucosal lesions and even lead to a potentially precancerous condition [26]. H. pyloribased gastritis is responsible for a higher risk of colorectal polyps, particularly dysplastic adenomas [27]. The findings showed that root canals may be a reservoir for H. pylori and a potential source for its transmission [28]. Whether

dental plaque is a primary source of *H. pylori* infection of the gastric mucosa of patients with poor oral hygiene needs to be confirmed [29].

The external manifestations of *H. pylori* infections suggest that the mechanism through which *H. pylori* CagA causes diseases is complex and diverse rather than a single pathophysiological mechanism.

- 2.3. Functional Dyspepsia. H. pylori is generally accepted as the main pathological agent for the occurrence of functional dyspepsia [30]. H. pylori CagA protein is associated with the development of functional gastrointestinal disorders (FGIDs). Although the role of CagA in peptic ulcer disease and gastritis is established, its role in functional dyspepsia is controversial. FD patients who are H. pylori positive have no clear clinical manifestations and the effect of H. pylori eradication is contradictory in these patients. A retrospective study showed that patients with CagA-positive H. pylori strains have a higher symptom score and more dyspeptic symptoms than patients with CagA-negative or H. pylorinegative functional dyspepsia [31].
- 2.4. Hormone Level Changes after H. pylori Colonization. An H. pylori infection could induce fluctuations in the levels of serotonin (5-HT), ghrelin, dopamine, cortisol, and other hormones in the circulatory system, resulting in damage to various systems, including the central nervous system, and the occurrence of the corresponding symptoms (Figure 2).
- 2.5. 5-Hydroxytryptamine (5-HT, Serotonin). Psychological disorders, such as anxiety or depression, have been reported to be associated with FD [32]. The results of a population-based investigation suggested that anxiety worsens the symptoms of FD [33]. A 12-year prospective population-based study found that people with higher levels of depression were significantly more likely to develop FD after 12 years [5]. Social anxiety disorder is associated with an overactive presynaptic serotonin system, increased serotonin synthesis, and increased transporter availability. The study by Harmer provided the insight that serotonin pathways may influence the mood of patients with depression by altering how the brain appraises emotional information at an implicit level [34].

Increasing evidence supports a close relationship between 5-hydroxytryptamine (5-HT, serotonin) and gastrointestinal motility and visceral hypersensitivity. Serotonin is synthesized through tryptophan hydroxylase-1 (TpH1) and TpH2, which are found in EC cells and neurons, respectively, and is inactivated by its uptake into enterocytes or neurons via the serotonin reuptake transporter (SERT). Approximately 90% of 5-HT in the body is synthesized in the gut, which has 14 different 5-HT receptor subtypes. The 5-HT 3A receptor and the 5-HT 2A receptor are associated with dyspeptic symptoms, while 5-HT4 receptor agonists may improve dyspeptic symptoms, particularly delayed gastric emptying [35]. In Japan, a 5-HT transporter gene polymorphism was found to be associated with dyspepsia [36].

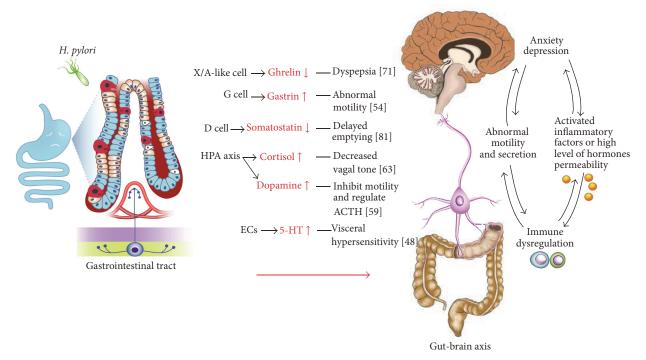


FIGURE 2: Effects of gastrointestinal hormones level to mental disorders after *H. pylori* colonization. CagA-positive *H. pylori* strains could induce fluctuations in the levels of serotonin (5-HT), ghrelin, dopamine, and cortisol, and might be the cause of some dyspepsia symptoms and mental disorders through blood circulation and brain-gut axis.

Abnormal levels of 5-HT have been reported in irritable bowel syndrome (IBS) patients. Raised plasma 5-HT levels were particularly found in female IBS-diarrhea patients, whereas reduced levels were found in IBS-constipation patients. Ahern [37] demonstrated that FD patients had significantly lower preprandial and postprandial plasma serotonin levels compared with those of healthy subjects. Decreased 5-HT levels may impair gastric accommodation or cause visceral hypersensitivity. A significant relationship between postprandial plasma serotonin levels and postprandial dyspepsia scores has been observed, which also indicated that serotonin plays a role in dyspeptic-symptom pathogenesis [37]. However, the role of 5-HT in regulating gastrointestinal tract function is imperfectly understood. Functional dyspepsia (FD) and IBS have been proposed to have a common pathogenesis. Nevertheless, little is known about the role of 5-HT in FD, which is largely due to the presence of various types of 5-HT receptors in the gastrointestinal tract and the absence of suitable and selective antagonists.

Serotonin has been demonstrated to affect many immunological processes and to increase or decrease the levels of proinflammatory cytokines [38]. Chemokines and signaling pathways active during inflammatory processes can affect the synthesis and degradation of serotonin. Stone and Darlington [39] discovered that NF-kB signaling pathway activation increased the rate of release of 5-HT via the phosphorylation of TPH1. It was known that IL-1, IL-2, IL-6, and IFN caused the degradation of the precursor of tryptophan (TRP) through activating an enzyme (indoleamine 2,3-dioxygenase,

IDO) and that a decreased level of TRP in the blood leads to a decreased level of 5-HT in the brain. The NF-kB signaling pathway that is involved in gene transcription participates in regulating inflammatory cytokines and plays a key role in the immune response, inflammation, and cell apoptosis in gastrointestinal diseases. Moreover, the increased expression of inflammatory cytokines in turn activates NF-kB. These results suggest a potential relationship between NF-kB activation and the 5-HT level in an infected host.

Because serotonin is a weak platelet agonist, a number of studies have shown an association between severe upper gastrointestinal bleeding with *H. pylori* infection and the levels of selective serotonin reuptake inhibitors (SSRIs). This effect depends on the release of 5-HT by platelets, which acquire serotonin from the blood. *H. pylori* eradication therapy of patients with upper gastrointestinal bleeding was reported to reduce their rebleeding rates [40]. Although 5-HT and *H. pylori* are related to gastric disease, this association requires further studies.

2.6. Ghrelin. Ghrelin, which is produced by gastric enteroendocrine X/A-like or G cells and is acylated by ghrelin O-acyltransferase (GOAT) before being released into the blood, plays a crucial role in gastric motility, appetite regulation, and acid secretion. Many gastrointestinal disorders involving inflammation, infection, and malignancy are also associated with altered ghrelin production and secretion. Age, lactation, sex hormones, and the expression of mRNA encoding GOAT, a critical enzyme for ghrelin activity, can impact ghrelin secretion.

A lower level of gastric ghrelin mRNA expression was observed in patients with an *H. pylori* infection compared to that in uninfected subjects [41]. Moreover, due to damage to the gastric X/A-like cells (which produce ghrelin), H. pyloriinfected patients also have a decreased serum ghrelin level. Inversely, the level of both serum ghrelin and ghrelin mRNA expression can rebound following H. pylori eradication therapy, with the consequent relief of dyspeptic symptoms [42, 43]. However, some researchers failed to find significant changes in ghrelin levels after *H. pylori* eradication [44]. They demonstrated that *H. pylori* infection of the stomach did not significantly affect the ghrelin levels. A study of mice found that the normal gut microbiota, independent of H. pylori, had a significant effect on ghrelin levels [45]. They also found that the presence of *H. pylori* did not upregulate leptin (a satiety hormone) production or decrease ghrelin secretion in the absence of the normal gastrointestinal microbiota, whereas when *H. pylori* colonized the gut together with the normal microbiota, the opposite result was observed.

An abnormally low level of ghrelin has been found in FD patients, particularly in those with PDS, compared with the level in healthy subjects. And the ghrelin plasma level is associated with the severity of the symptoms in patients with FD [46]. Paoluzi et al. [47] also reported recently that female patients with FD had lower fasting and postprandial ghrelin levels and that the abnormal ghrelin response was apparently involved in their meal-related symptoms. Although whether altered ghrelin levels are the cause or the result of dyspepsia is unclear, these data imply a possible role for ghrelin in the pathogenesis of FD patients with H. pylori positive. The gastritis induced by an H. pylori infection is predominantly related to T helper (Th)1/Th17 cell immunity. Ghrelin suppresses Th cell-dependent pathology. The downregulated level of ghrelin in the gastric mucosa of *H*. pylori-infected patients might promote an ongoing Th1-cell response and chronic active gastritis [48]. B. L. Slomiany and A. Slomiany [49] reported the role of phosphatidylinositol 3-kinase (PI3K) in digestive tract mucosa infected with H. pylori, demonstrating that the modulation of ghrelin as a gastric mucosal response to H. pylori depends on PI3K activation. The ghrelin receptor is highly specific, which means that only the acylated form of ghrelin can bind to it, and active ghrelin stimulates the appetite though neuropeptide Y (NPY). Because ghrelin receptor activation leads to the enhanced activity of the NPY pathway, activating the ghrelin receptor would be beneficial in abating early satiety and appetite loss. However, the mechanisms through which ghrelin is regulated in FD patients require more studies.

2.7. Gastrin and Somatostatin. Gastrin, CCK, and somatostatin are all sensitive to the stress of anxiety [50]. Gastrin, which is released by G cells in the pyloric mucosa, stimulates gastric acid secretion, promotes cell growth (increasing the rate of cell division and inhibiting apoptosis), and transacts with cholecystokinin-2 receptors (CCK2Rs). The CCK2Rs, mitogen-activated protein kinase 1 (MP1), and ERK1/ERK2 are reported to mediate the gastrin-induced growth of gastric adenocarcinomas [51]. Patients with chronic renal failure

have 2- to 3-fold higher serum gastrin levels because the kidneys are responsible for clearing gastrin [52]. Somatostatin is produced by D cells and neurons in the gastrointestinal tract and pancreas. In the stomach, the antrum mucosa secretes acid to stimulate the secretion of somatostatin and the latter inhibits gastrin secretion. That phenomenon explains why PPIs stimulate gastrin secretion by decreasing acid secretion, thus resulting in sustained hypergastrinemia, gastrinoma, and atrophic gastritis and possibly in gastric carcinoid tumors. It was shown that an acute H. pylori infection activates the sensory neurons associated with somatostatin stimulation [53]. H. pylori infection and a reduced somatostatin level have a complex etiological relationship in chronic gastritis. In H. pylori-positive patients, a decreased somatostatin content (likely mediated by proinflammatory cytokines) leads to increased gastrin secretion, perhaps due to the effect of H. pylori (a direct effect of CagL, a component of the T4SS for CagA) on G cells [54]. A study showed that Cag PAI-positive *H. pylori* strains or CagL activate the gastrin promoter, whereas Cag PAI-negative strains do not [55]. A further study indicated that gastrin expression stimulated by CagL is involved with epidermal growth factor receptor and MP1 signaling.

In investigating the relationship between the gastric motility disorder and the gastrointestinal hormone abnormality in the GI mucosa of FD patients, Van Oudenhove et al. [56] found that the levels of gastrin in the postprandial plasma and the gastric mucosa were significantly higher in FD patients with delayed emptying and suggested that the altered gastrin levels may play a role in the pathophysiology of the abnormal gastric motility of FD patients.

A study found that weight loss and symptom severity in FD patients were determined by somatization and depression [57]. A positive relationship was found between the degree of dyspeptic symptoms and the level of somatostatin [50]. The cited study also indicated that CCK and somatostatin might correlate certain psychological reactions with the pathophysiology of FD.

2.8. *Dopamine (DA)*. The homeostasis of the digestive system is dependent on the activity of aminergic mediators [such as 5-HT, noradrenaline (NA), and dopamine (DA)], which play crucial roles in the central or peripheral generation of gastrointestinal motility, secretion, and sensation. Dopamine is mainly produced by mesenteric organs (the GI tract, spleen, and pancreas) and is metabolized by monoamine oxidase and catechol-O-methyl-transferase (COMT). DA is known to modulate diverse physiological functions of the digestive system, such as acid and mucus secretion in the stomach and bicarbonate excretion in the duodenum. DA exerts its biological functions through two types of receptors, the D1-type receptors (D1 and D5) and D2-type receptors (D2, D3, and D4) receptors. In the digestive system, DA inhibits motility via the D1-type receptor present on smooth muscle and modulates the release of acetylcholine (ACTH) from myenteric neurons via D2-type receptors. Dopamine and its agonist have a protective effect on the development of various lesions in the gastroduodenum. It was also reported that dopamine

might protect the gastric mucosa against acidified ethanol through the activation of the $\alpha 2$ adrenoceptor, which leads to inhibition of gastric motility [58]. Recently, it was shown that dopamine acts as a strong antitumor/antiangiogenic factor by suppressing the expression of growth factors, such as vascular endothelium-derived growth factor (VEGF), to inhibit angiogenesis in malignancies [59]. However, another study suggested that dopamine is rapidly metabolized by COMT in the gastrointestinal tract; therefore, there would not be a sufficient effect on ulcer healing [60]. Giusti et al. [61] found that dopamine receptor antagonists or inhibitors, such as diazepam and other antipsychotic drugs, can stimulate the development of peptic ulcers and infections by *H. pylori*. However, Kirschbaum and Hellhammer [62] found the rate of H. pylori infection was increased after dopamine treatment in women with a high prolactin level. These studies may indicate a fuzzy relationship between dopamine and H. pylori infection, the pathological mechanism of which requires further study. Although DA receptors are considered to modulate GI motility and GI motor symptoms associated with FGIDs, their potential role in the pathophysiology of functional dyspepsia is still unknown.

2.9. Cortisol. Cortisol, which plays an important role in the defense of a host infected with bacteria, is secreted through the stress-mediated activation of the hypothalamic-pituitary-adrenal (HPA) axis. High levels of adrenocorticotropic hormone (ACTH) and cortisol are generally considered to be outcomes of an HPA-axis disorder. Moreover, the levels of the ACTH immunoreactive substance (IS) are regulated by negative feedback from neurogenic stimulation and plasma cortisol. On the other hand, leptin suppresses HPA-axis activity, whereas ghrelin stimulates neuropeptide Y and food intake, which induces high concentrations of plasma ACTH and cortisol. HPA-axis alterations are related to gut motor functions [63]. Moreover, serum-free cortisol fluctuations are associated with various symptoms of functional gastrointestinal disorders (FGIDs), including FD [64].

Some studies showed that an increased serum cortisol level promoted H. pylori colonization. Koşan et al. found that, compared with those of the healthy control group, the serum IGF-I and IGF-II concentrations were significantly decreased in *H. pylori*-positive patients, although their serum cortisol level was increased. These authors did not discuss the probable effect of cortisol on *H. pylori* infections [65]. In contrast, Katagiri et al. [66] reported that H. pylori-infected patients had significantly decreased cortisol levels than H. *pylori*-negative patients. They also demonstrated that cortisol prevents H. pylori colonization through strengthening the host defense mechanisms. Recently, studies have shown that drugs, such as cimetidine, reduce basal and stimulated cortisol synthesis through inhibiting enzymes. However, proton pump inhibitors, such as lansoprazole and rabeprazole, are thought to cause increased cortisol levels in a starvation condition due to their possible effects of stimulating the HPA axis and increasing the plasma ACTH-IS level [67].

However, few studies have investigated HPA-axis parameters in FD patients, and inconclusive results were obtained.

Adults with FD who have an autonomic nervous system disorder have been reported, and the most common finding in these patients is decreased vagal tone [68]. In PDS patients, mental stress before a meal increases the symptom severity through sympathetic hyperactivity and increased cortisol levels. Another study suggested that these neurohormonal responses to HPA-axis activation mainly affect gastric sensitivity [69]. A study in which the HPA-axis activity of FGID patients was inhibited showed that they displayed both salivary morning cortisol and diurnal cortisol levels that were significantly lower than those of controls [70]. De la Roca-Chiapas et al. [64] have reported that FGID patients with high cortisol levels complained of more depression than those with low or medium cortisol levels, whereas the latter described experiencing more pain. In contrast, another study did not find a tendency toward higher cortisol levels in FD patients and suggested that these patients have not had enough exposure to daily stress to activate the HPA axis. It is not impossible that both stress-induced anxiety and an altered neuroendocrine response could increase the severity of dyspeptic symptoms (Table 1).

2.10. Treatment of FD. The diverse clinical manifestations and the uncertain pathophysiological mechanisms make it difficult to select a therapeutic strategy to manage FD. There is currently no established treatment regimen. Updated guidelines for FD treatment have been published by academic gastroenterology organizations, such as the American Gastroenterological Association, promoting a comprehensive strategy that includes diet, behavior modification and cognitive therapy, psychological interventions, and drug therapy [71]. However, drug treatment is still the main form of practical FD treatment in China. H. pylori eradication and treatment with antacids, motility-regulatory drugs, and antidepressants are commonly recommended as an effective method to treat FD.

Patients in Asian countries receive a relatively higher benefit from H. pylori eradication. Researchers have shown that *H. pylori* eradication can result in a long-term remission of FD symptoms [72]. Some evidence suggested that H. pylori eradication significantly improved gastrointestinal symptoms in patients with EPS compared with its effects on patients with PDS [73]. H. pylori eradication certainly has a statistically significant but small benefit in patients with FD. It is likely that the results of *H. pylori* eradication reflect not only the effect of treating an H. pylori infection but also the effect on the gastrointestinal microbiome. The results of some randomized controlled trials (RCTs) support eradication therapy [74], whereas most other studies found no benefit. A Cochrane review reported no significant difference between patients who had received a placebo and those who had undergone H. pylori eradication. Furthermore, due to the wide use of antibiotics in H. pylori eradication regimens, drug-resistant H. pylori strains are arising. The success rate of triple-antibiotic eradication therapy is less than 80% throughout the world. This phenomenon makes it challenging for clinicians to manage infections with H. pylori strains that are resistant to antimicrobial agents, particularly those that are

		Relative enzymes	Possible signaling pathway	Altered hormones	Receptors	Consequence
		TPH1	NF-kB	5-HT	5-HTR 2A/3A	Dyspepsia
					5-HTR 4	Delayed gastric emptying
	To play a role in the	GOAT	P13K-Akt	Ghrelin	GHSR	Decreased motility Lower acid secretion Anorexia
	regulation of hormones	Monoamine oxidase	$cAMP \uparrow \rightarrow PKA \uparrow$	Dopamine	D1/D5	Psychotic
CagA (+) H. pylori strains	by certain factors such as CagA, CagL, MP1, IL-17, IL-8, or abnormal autonomic nervous system	or COMT	AC activation ↓		D2/D3/D4	Gastroduodenal lesion Tumor
		GC-GR compound	HPA axis	Cortisol	GR	Host defense mechanism recedes <i>H. pylori</i> colonization
		Cag PAI	MP1 signaling	Gastrin	CCK2Rs	Increased gastric acid Atrophy gastritis
						Decreased gastric

TABLE 1: Possible relationship between *H. pylori* and several hormones.

TPH1: tryptophan hydroxylase-1; GOAT: ghrelin O-acyltransferase; GHSR: growth hormone secretagogue receptor; COMT: catechol-O-methyl-transferase; AC: adenylate cyclase; GC: glucocorticoid; GR: glucocorticoid receptor.

HPA axis

Somatostatin

CCK

resistant to clarithromycin and metronidazole. As a secondline treatment, moxifloxacin has been investigated. Zhang et al. [75] suggest that moxifloxacin-based therapy is more effective than the standard triple or quadruple therapy for this disease. However, its adverse effects, such as tendonitis or a nervous system reaction, are a matter of concern. The diverse results of therapy may be due to the use of different trial designs, different methods of patient selection, or different H. pylori eradication strategies. Furthermore, accumulating evidence demonstrates that that the human stomach contains a complex microbial ecosystem that does not include H. pylori strains [76]. The balance of these communities is crucial for health maintenance, and their disturbance is considered to be involved in gastrointestinal diseases [77]. Andersson et al. [78] reported that the gastric mucosa displayed a diverse microbiota after *H. pylori* eradication, although it is difficult to discriminate whether the changes in the gastric microbiota were caused by H. pylori eradication or by antibiotic treatment.

Dyspepsia patients have significantly more health care consultations due to psychological distress. Chronic stress is considered a major risk factor for FD, possibly due to brain-gut axis dysregulation mediated by the hypothalamic-pituitary axis [79]. Antianxiety and antidepressant drugs have been reported to have positive effects on FD, particularly on intractable FD, with tricyclic antidepressants (TCAs) and small doses of selective serotonin uptake inhibitors (SSRIs)

most often mentioned as efficacious [72]. When these drugs are used to treat FD, clinicians should consider the direct effects of neurotransmitters on gastrointestinal disorders as well as their effects on mental disorders because they may affect the modulatory function of neurotransmitters (e.g., 5-HT) on gastrointestinal sensory, motor, and secretory functions. However, 5-HT receptors have been recognized as the target points for symptomatic improvement. 5-HT3 receptor antagonists and 5-HT4 receptor agonists have been selected to treat functional diarrhea or constipation. 5-HT4 receptor agonists cause the release of acetylcholine, which stimulates smooth muscle contraction, leading to accelerated gastric emptying [80].

SSTR

acid Chronic gastritis

Because the pathogenesis of FD is uncertain and most likely multifactorial, individualized treatment should be established based on the patients' chief complaints.

3. Summary

Whether there are distinct modes of pathogenesis of PDS and EPS remains controversial. Fang et al. [81] found that although PDS and EPS have some common risk factors, including a younger age, anxiety, and NSAID consumption, different risk factors appear to be associated with different FD subgroups; for example, *H. pylori* infection, an unmarried status, sleep disturbance, coffee consumption, and depression disorder are risk factors only for PDS but not for EPS. These

finding may indicate the distinct etiopathogenesis of the subgroups of FD.

CagA and 5-HT may participate in the pathogenesis of FD, but the specific underlying pathogenic mechanisms are still unclear. *H. pylori* infection and anxiety or depression are also significant factors in the pathogenesis of FD. Therefore, whether and how CagA and 5-HT affect the pathogenesis of FD (including PDS and EPS) are a topic worthy of study and more research is needed. Moreover, the wide distribution of 5-HT receptors and their respective roles in the pathogenesis of FD are of special significance and are worthy of further study.

Competing Interests

All authors declare no conflict of interests.

Authors' Contributions

Wang-Ping Meng and Zhong-Qiong Wang contributed equally to this work.

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References

- [1] N. J. Talley and A. C. Ford, "Functional dyspepsia," *The New England Journal of Medicine*, vol. 373, no. 19, pp. 1853–1863, 2015.
- [2] R. S. Choung and N. J. Talley, "Novel mechanisms in functional dyspepsia," *World Journal of Gastroenterology*, vol. 12, no. 5, pp. 673–677, 2006.
- [3] E. Turkkan, I. Uslan, G. Acarturk et al., "Does Helicobacter pylori-induced inflammation of gastric mucosa determine the severity of symptoms in functional dyspepsia?" *Journal of Gastroenterology*, vol. 44, no. 1, pp. 66–70, 2009.
- [4] R. J. L. F. Loffeld, B. F. M. Werdmuller, J. G. Kusters, and E. J. Kuipers, "Functional dyspepsia is associated with cagApositive Helicobacter pylori strains," *Scandinavian Journal of Gastroenterology*, vol. 36, no. 4, pp. 351–355, 2001.
- [5] N. A. Koloski, M. Jones, J. Kalantar, M. Weltman, J. Zaguirre, and N. J. Talley, "The brain—gut pathway in functional gastrointestinal disorders is bidirectional: a 12-year prospective population-based study," *Gut*, vol. 61, no. 9, pp. 1284–1290, 2012.
- [6] S.-H. Kuo, K.-H. Yeh, L.-T. Chen et al., "Helicobacter pylori CagA translocation is closely associated with the expression of caga-signaling molecules in low-grade gastric mucosaassociated lymphoid tissue lymphoma," *American Journal of Surgical Pathology*, vol. 39, no. 6, pp. 761–766, 2015.
- [7] M. Stein, P. Ruggiero, R. Rappuoli, and F. Bagnoli, "Helicobacter pylori CagA: from pathogenic mechanisms to its use as an anticancer vaccine," Frontiers in Immunology, vol. 4, article 328, 2013.

- [8] M. Suzuki, H. Mimuro, K. Kiga et al., "Helicobacter pylori CagA phosphorylation-independent function in epithelial proliferation and inflammation," Cell Host & Microbe, vol. 5, no. 1, pp. 23–34, 2009.
- [9] A. T. Franco, D. A. Israel, M. K. Washington et al., "Activation of β-catenin by carcinogenic Helicobacter pylori," Proceedings of the National Academy of Sciences of the United States of America, vol. 102, no. 30, pp. 10646–10651, 2005.
- [10] J. Wei, T. A. Nagy, A. Vilgelm et al., "Regulation of p53 tumor suppressor by *Helicobacter pylori* in gastric epithelial cells," *Gastroenterology*, vol. 139, no. 4, pp. 1333–1343, 2010.
- [11] X. Yong, B. Tang, B.-S. Li et al., "Helicobacter pylori virulence factor CagA promotes tumorigenesis of gastric cancer via multiple signaling pathways," Cell Communication and Signaling, vol. 13, article 30, 2015.
- [12] Y. Hirata, S. Maeda, T. Ohmae et al., "Helicobacter pylori induces $I\kappa B$ kinase α nuclear translocation and chemokine production in gastric epithelial cells," *Infection and Immunity*, vol. 74, no. 3, pp. 1452–1461, 2006.
- [13] D. W. Kang, W. C. Hwang, M. H. Park et al., "Rebamipide abolishes *Helicobacter pylori* CagA-induced phospholipase D1 expression via inhibition of NFκB and suppresses invasion of gastric cancer cells," *Oncogene*, vol. 32, no. 30, pp. 3531–3542, 2013.
- [14] N. Suzuki, N. Murata-Kamiya, K. Yanagiya et al., "Mutual reinforcement of inflammation and carcinogenesis by the *Heli-cobacter pylori* CagA oncoprotein," *Scientific Reports*, vol. 5, Article ID 10024, 2015.
- [15] R. M. Delahay and M. Rugge, "Pathogenesis of *Helicobacter pylori* infection," *Helicobacter*, vol. 17, no. 1, pp. 9–15, 2012.
- [16] N. Bagheri, F. Azadegan-Dehkordi, H. Shirzad, M. Rafieian-Kopaei, G. Rahimian, and A. Razavi, "The biological functions of IL-17 in different clinical expressions of *Helicobacter pylori-infection*," *Microbial Pathogenesis*, vol. 81, pp. 33–38, 2015.
- [17] O. Sokolova, M. Borgmann, C. Rieke, K. Schweitzer, H.-J. Rothkötter, and M. Naumann, "Helicobacter pylori induces type 4 secretion system-dependent, but CagA-independent activation of IκBs and NF-κB/RelA at early time points," *International Journal of Medical Microbiology*, vol. 303, no. 8, pp. 548–552, 2013.
- [18] F. Franceschi, A. Tortora, G. Gasbarrini, and A. Gasbarrini, "*Helicobacter pylori* and extragastric diseases," *Helicobacter*, vol. 19, supplement 1, pp. 52–58, 2014.
- [19] F. Franceschi, N. Christodoulides, M. H. Kroll, and R. M. Genta, "Helicobacter pylori and idiopathic thrombocytopenic purpura," *Annals of Internal Medicine*, vol. 140, no. 9, pp. 766–767, 2004.
- [20] F. Wong, E. Rayner-Hartley, and M. F. Byrne, "Extraintestinal manifestations of *Helicobacter pylori*: a concise review," *World Journal of Gastroenterology*, vol. 20, no. 34, pp. 11950–11961, 2014.
- [21] P. Papagiannakis, C. Michalopoulos, F. Papalexi, D. Dalam-poura, and M. D. Diamantidis, "The role of Helicobacter pylori infection in hematological disorders," *European Journal of Internal Medicine*, vol. 24, no. 8, pp. 685–690, 2013.
- [22] G. S. Tamer, I. Tengiz, E. Ercan, C. Duman, E. Alioglu, and U. O. Turk, "Helicobacter pylori seropositivity in patients with acute coronary syndromes," Digestive Diseases and Sciences, vol. 54, no. 6, pp. 1253–1256, 2009.
- [23] C. Weller, A. Charlett, N. L. Oxlade et al., "Role of chronic infection and inflammation in the gastrointestinal tract in the etiology and pathogenesis of idiopathic parkinsonism. Part 3:

- predicted probability and gradients of severity of idiopathic parkinsonism based on H. pylori antibody profile," *Helicobacter*, vol. 10, no. 4, pp. 288–297, 2005.
- [24] M. Kucukazman, B. Yavuz, M. Sacikara et al., "The relationship between updated Sydney system score and LDL cholesterol levels in patients infected with *Helicobacter pylori*," *Digestive Diseases and Sciences*, vol. 54, no. 3, pp. 604–607, 2009.
- [25] P. L. Lutsey, J. S. Pankow, A. G. Bertoni, M. Szklo, and A. R. Folsom, "Serological evidence of infections and Type 2 diabetes: the multiethnic study of atherosclerosis: original article: epidemiology," *Diabetic Medicine*, vol. 26, no. 2, pp. 149– 152, 2009.
- [26] E. H. Hassan, S. S. Gerges, K. A. El-Atrebi, and H. T. El-Bassyouni, "The role of *H. pylori* infection in gall bladder cancer: clinicopathological study," *Tumor Biology*, vol. 36, no. 9, pp. 7093–7098, 2015.
- [27] T. Tongtawee, S. Kaewpitoon, N. Kaewpitoon et al., "Helicobacter pylori associated gastritis increases risk of colorectal polyps: a hospital based-cross-sectional study in Nakhon Ratchasima Province, Northeastern Thailand," Asian Pacific Journal of Cancer Prevention, vol. 17, no. 1, pp. 341–345, 2016.
- [28] C. Hirsch, N. Tegtmeyer, M. Rohde, M. Rowland, O. A. Oyarzabal, and S. Backert, "Live Helicobacter pylori in the root canal of endodontic-infected deciduous teeth," *Journal of Gastroenterology*, vol. 47, no. 8, pp. 936–940, 2012.
- [29] C. M. Kilmartin, "Dental implications of Helicobacter pylori," Journal of Canadian Dental Association, vol. 68, no. 8, pp. 489–493, 2002.
- [30] K. Piriyapong, A. Tangaroonsanti, V. Mahachai, and R.-K. Vilaichone, "Helicobacter pylori infection impacts on functional dyspepsia in Thailand," Asian Pacific Journal of Cancer Prevention, vol. 15, no. 24, pp. 10887–10891, 2014.
- [31] Y. Li, Y. Nie, W. Sha, and H. Su, "The link between psychosocial factors and functional dyspepsia: an epidemiological study," *Chinese Medical Journal*, vol. 115, no. 7, pp. 1082–1084, 2002.
- [32] B. F. Filipović, T. Randjelovic, T. Ille et al., "Anxiety, personality traits and quality of life in functional dyspepsia-suffering patients," *European Journal of Internal Medicine*, vol. 24, no. 1, pp. 83–86, 2013.
- [33] A. Frick, F. Åhs, J. Engman et al., "Serotonin synthesis and reuptake in social anxiety disorder a positron emission tomography study," *JAMA Psychiatry*, vol. 72, no. 8, pp. 794–802, 2015.
- [34] C. K. Y. Cheung, Y. Y. Lee, Y. Chan et al., "Decreased basal and postprandial plasma serotonin levels in patients with functional dyspepsia," *Clinical Gastroenterology and Hepatology*, vol. 11, no. 9, pp. 1125–1129, 2013.
- [35] F. Toyoshima, T. Oshima, S. Nakajima et al., "Serotonin transporter gene polymorphism may be associated with functional dyspepsia in a Japanese population," *BMC Medical Genetics*, vol. 12, article 88, 2011.
- [36] W. Atkinson, S. Lockhart, P. J. Whorwell, B. Keevil, and L. A. Houghton, "Altered 5-hydroxytryptamine signaling in patients with constipation- and diarrhea-predominant irritable bowel syndrome," *Gastroenterology*, vol. 130, no. 1, pp. 34–43, 2006.
- [37] G. P. Ahern, "5-HT and the immune system," *Current Opinion in Pharmacology*, vol. 11, no. 1, pp. 29–33, 2011.
- [38] M. Haugen, R. Dammen, B. Svejda et al., "Differential signal pathway activation and 5-HT function: the role of gut enterochromaffin cells as oxygen sensors," *American Journal of Physiology—Gastrointestinal and Liver Physiology*, vol. 303, no. 10, pp. G1164–G1173, 2012.

- [39] T. W. Stone and L. G. Darlington, "Endogenous kynurenines as targets for drug discovery and development," *Nature Reviews Drug Discovery*, vol. 1, no. 8, pp. 609–620, 2002.
- [40] O. A. Paoluzi, D. V. G. Blanco, R. Caruso, I. Monteleone, G. Monteleone, and F. Pallone, "Impairment of ghrelin synthesis in *Helicobacter pylori*-colonized stomach: new clues for the pathogenesis of *H. pylori*-related gastric inflammation," *World Journal of Gastroenterology*, vol. 20, no. 3, pp. 639–646, 2014.
- [41] H. Suzuki and P. Moayyedi, "Helicobacter pylori infection in functional dyspepsia," Nature Reviews Gastroenterology & Hepatology, vol. 10, no. 3, pp. 168–174, 2013.
- [42] Y. J. Choi, N. Kim, H. Yoon et al., "Increase in plasma acyl ghrelin levels is associated with abatement of dyspepsia following Helicobacter pylori eradication," *Journal of Gastroenterology*, vol. 51, no. 6, pp. 548–559, 2016.
- [43] M. Eren, O. Colak, S. Işiksoy, and A. Yavuz, "Effect of H. pylori infection on gastrin, ghrelin, motilin, and gastroesophageal reflux," The Turkish Journal of Gastroenterology, vol. 26, no. 5, pp. 367–372, 2015.
- [44] Y. Khosravi, S. W. Seow, A. A. Amoyo et al., "Helicobacter pylori infection can affect energy modulating hormones and body weight in germ free mice," Scientific Reports, vol. 5, article 8731, 2015.
- [45] T. Shindo, S. Futagami, T. Hiratsuka et al., "Comparison of gastric emptying and plasma ghrelin levels in patients with functional dyspepsia and non-erosive reflux disease," *Digestion*, vol. 79, no. 2, pp. 65–72, 2009.
- [46] C. K. Cheung and J. C.-Y. Wu, "Role of ghrelin in the pathophysiology of gastrointestinal disease," *Gut and Liver*, vol. 7, no. 5, pp. 505–512, 2013.
- [47] O. A. Paoluzi, G. Del Vecchio Blanco, R. Caruso et al., "Helicobacter pylori infection associates with a mucosal downregulation of Ghrelin, negative regulator of Th1-cell responses," Helicobacter, vol. 18, no. 6, pp. 406–412, 2013.
- [48] B. L. Slomiany and A. Slomiany, "Role of ghrelin-induced phosphatidylinositol 3-kinase activation in modulation of gastric mucosal inflammatory responses to Helicobacter pylori," *Inflammopharmacology*, vol. 22, no. 3, pp. 169–177, 2014.
- [49] B. L. Slomiany and A. Slomiany, "Modulation of gastric mucosal inflammatory responses to *Helicobacter pylori* via ghrelininduced protein kinase Cδ tyrosine phosphorylation," *Inflam-mopharmacology*, vol. 22, no. 4, pp. 251–262, 2014.
- [50] T. S. Steigedal, W. S. Prestvik, L.-K. M. Selvik et al., "Gastrininduced proliferation involves MEK partner 1 (MP1)," In Vitro Cellular and Developmental Biology-Animal, vol. 49, no. 3, pp. 162–169, 2013.
- [51] A. Quattrone, B. Dewaele, A. Wozniak et al., "Promoting role of cholecystokinin 2 receptor (CCK2R) in gastrointestinal stromal tumour pathogenesis," *The Journal of Pathology*, vol. 228, no. 4, pp. 565–574, 2012.
- [52] M. Zaki, P. E. Coudron, R. W. McCuen, L. Harrington, S. Chu, and M. L. Schubert, "H. pylori acutely inhibits gastric secretion by activating CGRP sensory neurons coupled to stimulation of somatostatin and inhibition of histamine secretion," *American Journal of Physiology-Gastrointestinal and Liver Physiology*, vol. 304, no. 8, pp. G715–G722, 2013.
- [53] T. L. Cover, "Role of *Helicobacter pylori* CagL in modulating gastrin expression," *Gut*, vol. 61, no. 7, pp. 965–966, 2012.
- [54] T. Wiedemann, S. Hofbaur, N. Tegtmeyer et al., "Helicobacter pylori CagL dependent induction of gastrin expression via a novel $\alpha v \beta$ 5-integrin–integrin linked kinase signalling complex," *Gut*, vol. 61, no. 7, pp. 986–996, 2012.

- [55] M.-R. He, Y.-G. Song, and F.-C. Zhi, "Gastrointestinal hormone abnormalities and G and D cells in functional dyspepsia patients with gastric dysmotility," *World Journal of Gastroenterology*, vol. 11, no. 3, pp. 443–446, 2005.
- [56] L. Van Oudenhove, J. Vandenberghe, B. Geeraerts et al., "Determinants of symptoms in functional dyspepsia: gastric sensorimotor function, psychosocial factors or somatisation?" *Gut*, vol. 57, no. 12, pp. 1666–1673, 2008.
- [57] S. O'Mahony, T. G. Dinan, P. W. Keeling, and A. S. B. Chua, "Central serotonergic and noradrenergic receptors in functional dyspepsia," World Journal of Gastroenterology, vol. 12, no. 17, pp. 2681–2687, 2006.
- [58] D. Chakroborty, C. Sarkar, R. B. Mitra, S. Banerjee, P. S. Dasgupta, and S. Basu, "Depleted dopamine in gastric cancer tissues: dopamine treatment retards growth of gastric cancer by inhibiting angiogenesis," *Clinical Cancer Research*, vol. 10, no. 13, pp. 4349–4356, 2004.
- [59] K. Nishikawa, K. Amagase, and K. Takeuchi, "Effect of dopamine on the healing of acetic acid-induced gastric ulcers in rats," *Inflammopharmacology*, vol. 15, no. 5, pp. 209–213, 2007.
- [60] Y. Yilmaz, C. B. Gul, M. Arabul, and M. A. Eren, "Helicobacter pylori: a role in schizophrenia?" Medical Science Monitor, vol. 14, no. 7, pp. HY13–HY16, 2008.
- [61] M. Giusti, M. R. Falivene, L. Foppiani, V. Savarino, and A. Costa, "Increased prevalence of Helicobacter pylori infection in females treated with dopaminergic drugs for hyperprolactinaemia," *Clinical Endocrinology*, vol. 48, no. 3, pp. 373–374, 1998.
- [62] C. Kirschbaum and D. H. Hellhammer, "Salivary cortisol in psychoneuroendocrine research: recent developments and applications," *Psychoneuroendocrinology*, vol. 19, no. 4, pp. 313– 333, 1994.
- [63] U. Ehlert, U. M. Nater, and A. Böhmelt, "High and low unstimulated salivary cortisol levels correspond to different symptoms of functional gastrointestinal disorders," *Journal of Psychosomatic Research*, vol. 59, no. 1, pp. 7–10, 2005.
- [64] J. M. De la Roca-Chiapas, S. Solís-Ortiz, M. Fajardo-Araujo, M. Sosa, T. Córdova-Fraga, and A. Rosa-Zarate, "Stress profile, coping style, anxiety, depression, and gastric emptying as predictors of functional dyspepsia: a case-control study," *Journal* of *Psychosomatic Research*, vol. 68, no. 1, pp. 73–81, 2010.
- [65] B. Koşan, O. Yüksel, I. Üstün et al., "Role of endogenous cortisol on *Helicobacter pylori* colonization," *Clinical Biochemistry*, vol. 41, no. 10-11, pp. 917–919, 2008.
- [66] F. Katagiri, S. Inoue, Y. Sato, H. Itoh, and M. Takeyama, "Comparison of the effects of proton pump inhibitors on human plasma adrenocorticotropic hormone and cortisol levels under the starved condition," *Biomedicine & Pharmacotherapy*, vol. 60, no. 3, pp. 109–112, 2006.
- [67] S. L. Silva Lorena, M. J. De Oliveira Figueiredo, J. R. Souza Almeida, and M. A. Mesquita, "Autonomic function in patients with functional dyspepsia assessed by 24-hour heart rate variability," *Digestive Diseases and Sciences*, vol. 47, no. 1, pp. 27–31, 2002.
- [68] F. De Giorgi, G. Sarnelli, C. Cirillo et al., "Increased severity of dyspeptic symptoms related to mental stress is associated with sympathetic hyperactivity and enhanced endocrine response in patients with postprandial distress syndrome," *Neurogastroenterology and Motility*, vol. 25, no. 1, pp. 31.e1–31.e3, 2013.
- [69] C. Stasi, M. Bellini, F. Costa et al., "Neuroendocrine markers and psychological features in patients with irritable bowel syndrome," *International Journal of Colorectal Disease*, vol. 28, no. 9, pp. 1203–1208, 2013.

- [70] H. Miwa, U. C. Ghoshal, K. M. Fock et al., "Asian consensus report on functional dyspepsia," *Journal of Gastroenterology and Hepatology (Australia)*, vol. 27, no. 4, pp. 626–641, 2012.
- [71] A. C. Ford, "Eradicating Helicobacter pylori in functional dyspepsia," *Gastroenterology*, vol. 142, no. 7, pp. 1613–1614, 2012.
- [72] L. Lan, J. Yu, Y.-L. Chen et al., "Symptom-based tendencies of *Helicobacter pylori* eradication in patients with functional dyspepsia," *World Journal of Gastroenterology*, vol. 17, no. 27, pp. 3242–3247, 2011.
- [73] N. J. Talley, G. Holtmann, and M. M. Walker, "Therapeutic strategies for functional dyspepsia and irritable bowel syndrome based on pathophysiology," *Journal of Gastroenterology*, vol. 50, no. 6, pp. 601–613, 2015.
- [74] L. E. Mazzoleni, G. B. Sander, C. F. D. M. Francesconi et al., "Helicobacter pylori eradication in functional dyspepsia: HEROES trial," Archives of Internal Medicine, vol. 171, no. 21, pp. 1929–1936, 2011.
- [75] G. Zhang, J. Zou, F. Liu et al., "The efficacy of moxifloxacin-based triple therapy in treatment of Helicobacter pylori infection: a systematic review and meta-analysis of randomized clinical trials," *Brazilian Journal of Medical and Biological Research*, vol. 46, no. 7, pp. 607–613, 2013.
- [76] C. S. O. Eun, B. K. W. Kim, D. S. O. Han et al., "Differences in gastric mucosal microbiota profiling in patients with chronic gastritis, intestinal metaplasia, and gastric cancer using pyrosequencing methods," *Helicobacter*, vol. 19, no. 6, pp. 407–416, 2014.
- [77] K. Lertpiriyapong, M. T. Whary, S. Muthupalani et al., "Gastric colonisation with a restricted commensal microbiota replicates the promotion of neoplastic lesions by diverse intestinal microbiota in the *Helicobacter pylori* INS-GAS mouse model of gastric carcinogenesis," *Gut*, vol. 63, no. 1, pp. 54–63, 2014.
- [78] A. F. Andersson, M. Lindberg, H. Jakobsson, F. Bäckhed, P. Nyrén, and L. Engstrand, "Comparative analysis of human gut microbiota by barcoded pyrosequencing," *PLoS ONE*, vol. 3, no. 7, Article ID e2836, 2008.
- [79] C. Kiank, Y. Taché, and M. Larauche, "Stress-related modulation of inflammation in experimental models of bowel disease and post-infectious irritable bowel syndrome: role of corticotropinreleasing factor receptors," *Brain, Behavior, and Immunity*, vol. 24, no. 1, pp. 41–48, 2010.
- [80] G. M. Mawe and J. M. Hoffman, "Serotonin signalling in the gut-functions, dysfunctions and therapeutic targets," *Nature Reviews Gastroenterology and Hepatology*, vol. 10, no. 8, pp. 473–486, 2013.
- [81] Y.-J. Fang, J.-M. Liou, C.-C. Chen et al., "Distinct aetiopathogenesis in subgroups of functional dyspepsia according to the Rome III criteria," *Gut*, vol. 64, no. 10, pp. 1517–1528, 2015.

















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