

Dispersed free phytosterols as structuring agents in lipid systems with reduced saturated fat

K.R.R. Godoi^{a,✉}, R.C. Basso^b, M.H.M. Buscato^c, L.P. Cardoso^d, T.G. Kieckbusch^c and A.P.B. Ribeiro^a

^aDepartment of Food Technology, School of Food Engineering, University of Campinas, Brazil.

^bInstitute of Science and Technology, Federal University of Alfenas, Brazil.

^cSchool of Chemical Engineering, University of Campinas, Brazil.

^dDepartment of Applied Physics, Institute of Physics, University of Campinas, Brazil.

✉Corresponding author: kamila.ramponi@hotmail.com

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SUMMARY: The negative effects of *trans* fatty acids and saturated fatty acids in food have been widely discussed and this has led to progressive changes in the legislation of many countries. The use of structuring agents or crystallization modifiers, as specific triacylglycerol and minor lipids have been indicated as the only viable alternative for obtaining low saturated fats with properties which are compatible with food application. In this context, phytosterols, natural products with hypocholesterolemic action, and hard fat-crystallization modulators, present a new option for structuring lipid matrices. This work characterized the effects of fully hydrogenated soybean oil and free phytosterols on the physical properties and crystallization behavior of palm oil and canola oil blends for the development of zero *trans*-fat bases with low levels of saturated fatty acids. The systems were evaluated for chemical composition, atherogenic index, solid fat profiles, microstructure, consistency, thermal behavior and polymorphism.

KEYWORDS: *Canola oil; Crystallization; Fully hydrogenated soybean oil; Lipid structuring; Palm oil; Phytosterols*

RESUMEN: *Fitosteroles libres dispersos como agentes estructurantes en sistemas lipídicos reducidos en grasa saturada.* Los efectos negativos de los ácidos grasos *trans* y los ácidos grasos saturados en los alimentos ha llevado a cambios progresivos en la legislación de muchos países. Se ha indicado que el uso de agentes estructurantes o modificadores de la cristalización, como triacilglicérolos específicos y componentes lipídicos menores, es la única alternativa viable para obtener grasas bajas saturadas que tienen propiedades compatibles con su aplicación en alimentos. En este contexto, los fitosteroles, productos naturales con acción hipocolesterolémica y moduladores de cristalización de la grasa saturada, se presentan como una reciente opción para estructurar matrices lipídicas. En este trabajo se realizó una caracterización del aceite de soja totalmente hidrogenado y fitoesteroles libres sobre las propiedades físicas y el comportamiento de cristalización de mezclas de aceite de palma/aceite de canola para el desarrollo de bases de grasas cero-*trans* con bajos niveles de ácidos grasos saturados. Se evaluó la composición química, el índice aterogénico, el perfil de grasa sólida, la microestructura, consistencia, y el comportamiento térmico y polimorfismo.

PALABRAS CLAVE: *Aceite de canola; Aceite de palma; Aceite de soja totalmente hidrogenado; Cristalización; Estructuración lipídica; Fitosteroles*

ORCID ID: Godoi KRR <http://orcid.org/0000-0003-4115-0710>, Basso RC <http://orcid.org/0000-0002-1714-8322>, Buscato MHM <http://orcid.org/0000-0002-9053-6110>, Cardoso LP <http://orcid.org/0000-0003-3910-2293>, Kieckbusch TG <http://orcid.org/0000-0003-3074-8581>, Ribeiro APB <http://orcid.org/0000-0002-6532-1265>

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1. INTRODUCTION

The increasing incidence of cardiovascular disease in the last century led to an endless search for risk factors related to its development. Several studies have found that a relatively high intake of fat, especially trans and saturated fat, is a major contributor to the high incidence of coronary heart disease (Zevenbergen *et al.*, 2009). Trans fatty acids are included among the dietary lipids that act as risk factors for coronary artery disease by modulating the synthesis of cholesterol and its fractions and acting on eicosanoids (Dhaka *et al.*, 2011). Saturated fat is one of the main causes of the increase in plasma cholesterol, whose reduction in the diet is widely supported for reducing the risk of cardiovascular disease (Wassell *et al.*, 2010).

Changing the composition of the lipid matrix in order to reduce the saturated fatty acid concentration and improve the lipid nutritional profile has become a strategic priority for food processing. For this purpose, the recent scientific literature indicates a potential alternative use of various components with structuring action and performance capacity in molecular or sub-micron levels in lipid matrices. These molecular agents may act on the process of structuring lipid systems as a whole, modulating properties such as thermal behavior, polymorphic stability and microstructure. Similarly, the effects of these modifiers on the macroscopic level, such as visual appearance, rheology and consistency has also been the subject of new studies (Rogers *et al.*, 2014; Smith *et al.*, 2011). In this context, different materials have been evaluated, such as trisaturated triacylglycerols, free fatty acids, partial glycerides, waxes, fatty alcohols, phospholipids, phytosterols and different classes of emulsifiers. However, a complete understanding of the effects of these modifiers in various lipid systems and their interactions, has not yet been completely established in the scientific literature (Ribeiro *et al.*, 2015).

Mechanisms that rely on the incorporation of a crystallization modifier compound into a mixture require a concentration from 0.5% to 5% of the additives. On the other hand, if the primary action type occurs during the nucleation step in crystal forming, concentrations lower than 0.1% may be sufficient for modifying the process. A concentration which is lower than the last one may be sufficient for crystallization modifying when the mechanism is based on the blocking of the growth points of crystals. The position of the hydroxyl group and the number of double bonds in the carbon cyclic chain affect the characteristics of the crystal lattice or hinder its formation (Herrera; Marquez Rocha, 1996; Perneti *et al.*, 2007b; Smith *et al.*, 2011).

The fatty acid composition of palm oil (PO) is characteristic and unique, with similar amounts of saturated and unsaturated fatty acids, mainly

palmitic and oleic acids. In addition, PO has from 10 to 16% saturated fatty acid in the sn-2 position of triacylglycerol molecules, resulting in specific crystallization properties (O'Brien, 2004).

Canola oil (CO) has, on average, 22% linoleic acid and a high content of oleic acid, about 60%, with a low content in saturated fatty acids (about 6%), resulting in a nutritionally favorable omega-3/omega-6 ratio. Total tocopherol content is 770 mg/kg, higher than sunflower and corn oils, and the total sterol content is about 0.7%, with β -sitosterol, campesterol and brassicasterol corresponding to 52.3%, 27.6% and 13.8%, respectively (Gunstone *et al.*, 2007; Przybylski *et al.*, 2005).

Soybean oil is composed mainly of fatty acids with 18 carbon atoms. The average contents of stearic acid, oleic acid, linoleic acid and linolenic acid are 3.9%, 23.9%, 52.1% and 7.8%, respectively (Dubois *et al.*, 2007). In its fully hydrogenated form, soybean oil has 87.7% stearic acid, a saturated fatty acid that has no adverse metabolic effects (Gebauer *et al.*, 2014). The addition of low contents of fully hydrogenated soybean oil (HSO) in some lipid matrices showed effective potential as a modifier of their crystallization and physical properties, as was reported in a study with cocoa butter (Ribeiro *et al.*, 2013).

Phytosterols and phytostanols, the hydrogenated form of phytosterols, have important effects on cholesterol inhibition in the body. This mechanism is still not completely understood, but there are some hypotheses: cholesterol absorption would be inhibited because of (1) micellar solubilization; (2) co-crystallization of cholesterol and phytosterols from the giant micelles; (3) inhibition of lipases, esterases, and other enzymes that are involved in the sterol absorption process; (4) effects on the activity of brush border transport (Rozner and Garti, 2006). Other beneficial effects of these compounds are the anti-carcinogenic and anti-polymerizing properties in oils rich in unsaturated fatty acids, such as canola oil (Moreau *et al.*, 2002). Despite their nutritional value, the addition of these compounds in food is restricted since in free form they are not miscible in water and have low solubility in fats and oils (Salo and Wester, 2005).

Phytosterols can be used as modifiers on the crystallization and structuring of lipids and to obtain lipid matrices with specific physical properties (Wassell *et al.*, 2010). These changes occur due to different mechanisms: covalent bonds, hydrogen bonds, Van der Waals forces or steric entanglement, causing changes in solid profiles, melting points, consistency and other physical properties in fatty systems (Perneti *et al.*, 2007). Several types of phytosterols have potential for structuring oils and fats to form a continuous crystal network, resulting in higher consistency for the lipid matrix (Perneti *et al.*, 2007). The esterification of phytosterols and stanols with long chain fatty acids increases

their solubility in the lipid phase and consequently facilitates their incorporation into foods, but this process adversely affects the cholesterol-lowering properties of phytosterols, which should preferably be used in their free form for the functional effect (Noakes *et al.*, 2005).

By dispersing phytosterols in the oil phase, it has been possible to maximize the concentration of phytosterols used and maintained in solution in oil over time, but also to achieve easily obtainable dispersion without the need for a step involving the melting of the phytosterols. The resulting products have acceptable organoleptic properties and are suitable for ingestion in the form of diverse oil-based nutritional and pharmaceutical formulations (Auriou, 2004).

PO is one of the most commonly used fats for food production due to its physical and chemical characteristics. This oil has a high content of palmitic acid, a saturated fatty acid responsible for cardiovascular diseases. With the aim of obtaining a lipid matrix with the required physical and chemical properties for use in fatty based products with hypocholesterolemic properties, a promising approach is the mixture of oils rich in monounsaturated fatty acids and/or other biofunctional compounds with PO.

The aim of this study was to develop a complete characterization of the effect of dispersed free phytosterols (FP) and HSO on the physical properties and crystallization behavior of PO/CO blends in order to obtain zero trans-fat lipid blends with reduced levels of saturated fatty acids for application in processed foods. It is possible that FP act as the main nuclei for crystallization and together with HSO can induce the crystallization process, modifying properties such as consistency, microstructure and polymorphism. The addition of CO in PO was used to reduce the amount of saturated fatty acids in the blends, producing healthy and applicable lipid systems. At the same time, phytosterols have a recognized hypocholesterolemic effect, which contributes to the reduction in the risk of cardiovascular diseases.

2. MATERIALS AND METHODS

2.1. Materials

Refined palm oil (PO), supplied by Agropalma (Brazil); Refined canola oil (CO) acquired in a local market; Fully hydrogenated soybean oil (HSO), available from Cargill Agricola (Brazil); Mixture of free phytosterols (FP), provided by the Cognis company (Brazil) with the following composition: beta-sitosterol (47.4%); campesterol (26.2%); stigmasterol (14.0%); brassicasterol (3.6%); delta 5-avenasterol (2.4%); cholesterol (0.4%); campestanol (0.6%); sitostanol (0.8%); delta 7-avenasterol (0.3%); delta 5-stigmasterol (0.5%); others (3.8%).

2.2. Preparation of blends

Mixtures of PO and CO were prepared as reference systems with PO:CO (w:w) ratios of 100:0, 80:20, 60:40, 40:60, 20:80 and 0:100; and identified respectively using the same numerical codes. Two sets of blends were prepared from the reference systems: i) a set added of 10% (w:w) dispersed FP, identified as 100:0-FP, 80:20-FP, 60:40-FP, 40:60-FP, 20:80-FP and 0:100-FP; ii) a set added with 10% (w:w) dispersed FP and 5% (w:w) HSO, identified as 100:0-FP-HSO, 80:20-FP-HSO, 60:40-FP-HSO, 40:60-FP-HSO, 20:80-FP-HSO and 0:100-FP-HSO. The oil mixtures were heated to 100 °C, and the structuring agents were added under magnetic stirring. The systems were maintained under agitation for 3 minutes for complete homogenization. This temperature was used because when free phytosterols and stanols are heated at higher temperatures, the formation of dimeric, oligomeric or other oxidation products may occur, according some studies (Raczyk *et al.* 2017; Senate Commission on Food Safety 2014).

2.3. Physical and chemical characterization

2.3.1. Fatty acid composition

The fatty acid composition was determined for PO, CO and HSO, using a gas chromatographic system, CGC Agilent Series 6850 (Santa Clara, California, USA) with FID detector; capillary column Agilent-DB-23 (50% cyanopropyl – methylpolysiloxane), length 60 m, internal diameter 0.25 mm and film thickness 0.25 µm. The operational conditions were: flow rate of 1.0 ml/min; linear velocity of 24 cm/s; injector temperature of 250 °C; detector temperature of 280 °C/min; oven temperature of 110 °C for 5 min, 110 – 215 °C at 5 °C/min, 215 °C for 34 min; helium carrier gas; injected volume of 1.0 µL/min and split of 1:50. Derivatization was performed by methylation using NH₄Cl in a methanol solution. Methyl esters were extracted using a NaCl saturated solution and petroleum ether, according to the methodology proposed by Hartman and Lago (1973). Individual fatty acid methyl esters were identified by comparison of retention times to commercial standards, and they were quantified based on relative peak areas. The fatty acid composition of the blends was calculated by weighing the composition PO and CO with their respective mass fractions in the mixtures.

2.3. Atherogenic index

The Atherogenic Index (AI) can be calculated for diets or foods according to fatty acid composition, the ratio between the sums of lauric, miristic and palmitic acids and unsaturated fatty acids present in the blends (Equation 1). The higher the value found

for this relationship, more atherogenic is the blend, with greater contribution to the development of cardiovascular diseases like atherosclerosis and thrombosis (Kim *et al.*, 2008; Ulbricht *et al.*, 1991).

$$AI = \frac{C12:0 + C14:0 + C16:0}{C18:1 + C18:2 + C18:3 + C16:1 + C20:1}$$

Equation 1

2.5. Triacylglycerol composition

This composition was determined for PO, CO and HSO using a gas chromatography CGC Agilent 6850 Series GC System (Santa Clara, California, USA). A capillary column Agilent - DB-17 HT (50% phenyl- 50% methylpolysiloxane) was used: length 15 m, internal diameter 0.25 mm and film thickness 0.15 μm . The operational conditions were: injector temperature 360 °C; detector temperature of 375 °C/min; oven temperature from 250°C to 350 °C at 5 °C/min, 350 °C for 20 min; helium carrier gas; injected volume of 1.0 μL /min and split of 1:100. Triacylglycerols were identified by comparison of retention times, and quantified based on relative peak areas (de Oliveira *et al.*, 2015; Ribeiro *et al.*, 2013). The triacylglycerol compositions of the blends were calculated by weighing the composition PO, CO and HSO with their respective mass fractions in the mixtures in terms of trissaturated (S_3), dissaturated (S_2U), monossaturated (SU_2) triunsaturated (U_3) triacylglycerols.

2.6. Thermal behavior

The thermal behavior of the blends was evaluated by crystallization curves. The equipment used was a differential scanning calorimeter TA Q 2000 Thermal Analyzer, with the data processing system Universal V4.7a coupled to a RCS90 Refrigerated Cooling System (TA Instruments, Waters LLC). The samples were subjected to the temperature program: 80 °C for 10 min, 80 °C to - 60 °C at 10 °C/min, - 60 °C for 30 min. (AOCS, 2009; Campos, 2005).

2.7. Consistency

The consistency of the blends was assessed using the texture analyzer TA-XT2i plus equipment (Stable Micro Systems). The samples were heated for complete melting and stored in 50 mL beakers. The conditioning of the samples was performed in an incubator using a pre-stabilization step of 24 h at 5 °C, and a stabilization step of 24 h at 15 °C. The tests were performed in quadruplicate at 15 °C, using a probe Plexiglas 60° angle cone with a non-truncated tip. The experiments were carried out at a distance of 10 mm, velocity of 2 mm/s and time

of 5 s (Campos, 2005; Oliveira *et al.*, 2015). The compression force was given in gf, and the penetration data were converted to yield value according to Haighton, 1959, as described by the equation 2:

$$YV = \frac{K \times W}{p^{1,6}}$$

Equation 2

where YV is yield value (gf/cm^2); K is a factor depending on the cone angle (2815 for 60° angle); W is the compression force (gf); p is penetration depth (0.1 mm).

2.8. Solid fat content

The solid fat content (SFC) as a function of temperature was determined by a low resolution NMR spectrometer (Bruker pc 120 Minispec). The temperature was controlled using high precision dry baths (TCO 2000, 0 - 70 °C). The analyses were carried out according to the AOCS Cd 16b-93 (AOCS, 2009) direct method, following sequential measurements at temperatures of 10, 15, 20, 25, 30, 35, 40, 45 and 50 °C.

2.9. Microstructure

The samples were heated to 80 °C, and a drop was placed on a pre-heated glass slide with the aid of a glass capillary tube covered with a pre-heated (80 °C) coverslip. Slides in triplicate were maintained at 25 °C for 2h in an incubator. After this period, the morphology of crystal agglomerates was assessed using light polarized microscopy (Olympus, model B50) coupled to a digital video camera (Media Cybernetics). The slides were transferred onto a temperature-controlled plate (25 °C). The images were taken from three different visual fields, at a magnification of 40 X, and then, a single representative image was selected to represent the systems (Campos, 2005). The software Image Pro Plus (version 7.0, Media Cybernetics) was used to take the images.

2.10. Polymorphism

The crystalline polymorphic form of the blends was determined according to the AOCS Cj 2-95 methodology (AOCS, 2009). The analyses were performed in a diffractometer (Philips PW 1710); Bragg-Brentano geometry ($\theta:2\theta$); Cu-K α radiation ($k = 1.54056 \text{ \AA}$; at 40 kV and 30 mA). The assays were carried out with a 0.02 step in 2θ , for an acquisition time of 2 seconds, over the range 5 - 40° (2 θ scale). The tests were performed at 15 °C, after melting and storage of the samples for 30 days at this temperature. The crystalline forms were identified from their characteristic short spacings, calculated using the Bragg Law:

$$\lambda = 2d \sin\theta$$

Where λ is the wavelength emitted X-ray, d is the short spacing and θ is the diffraction angle.

3. RESULTS AND DISCUSSION

3.1. Chemical composition

PO composition showed about 4% more unsaturated fatty acid compared to the saturated ones, while unsaturated fatty acids (62.8% monounsaturated) represented about 10.5 times more than saturated molecules in CO, as shown in Table 1. HSO did not present significant amounts of unsaturated fatty acids, with 87.2% total acid content composed for stearic acid due to full hydrogenation of oleic, linoleic and linolenic acids from natural soybean oil.

The saturated fatty acid contents of the control systems compared to those added with 5% HSO increased from 48.18 – 8.71% (100:0 – 0:100) to 50.77 – 13.27% (100:0-FP-HSO – 0:100-FP-HSO). Stearic acid content increased from 4.96 – 2.57% to 9.07 – 6.8%, while oleic acid decreased from 41.76% – 61.35% to 39.68 – 58.29%, respectively for 100:0 – 0:100 and 100:0-FP-HSO – 0:100-FP-HSO. The blends containing both PO and CO resulted in intermediate values of these compounds. In this way, the addition of HSO resulted in increases in stearic acid contents of 1.8 and 2.7 times, respectively, for the systems with no addition of CO and no addition of PO.

The results for fatty acid composition showed that it was possible to reduce the saturated fatty acids in the control blends by 39.7%; while there was an increase of 20.7% in monounsaturated fatty acids and 18.7% in polyunsaturated fatty acids. For blends containing HSO these reductions corresponded to 37.5%, 19.7% and 18.7%, respectively.

The systems containing 5% HSO showed a constant increase, as a function of the PO content, 2.05% of S_3 , as observed in Table 2. On the other hand, the same system showed a decrease in U_3 as a function of PO addition in the range of 13 – 14%. The mixtures added with HSO, had from about 3.3 to 1.5 times more S_3 , following the increase in palm oil addition, compared to those with no addition of HSO.

Systematic reviews indicate that Intake replacement of saturated fatty acids by unsaturated fatty acids may reduced the incidence of cardiovascular diseases (de Souza *et al.*, 2015) and the consumption of 2 g/day of phytosterols decreases LDL cholesterol concentration on average by 10% (Brufau *et al.*, 2008). Regarding the possible application of these blends, an effective portion to reduce cholesterol levels should contain 20 g/day of the blend. In margarines, for example, formed by 80% of the lipid phase, about 25g of the final product, equivalent to approximately one tea spoon should be consumed per day to ensure its functional effect.

The methodology used for evaluating chemical composition did not cover the investigation of minor lipids in the blends. However, CO and PO were refined and newly acquired for this study, thereby minimizing the presence of partial acylglycerols in the evaluated blends.

3.2. Atherogenic index

The intake of foods with large amounts of cholesterol and saturated fats favors the development of cardiovascular diseases. These considerations, along with an increased awareness among the public, led to the development of a single number to express the hypercholesterolaemic atherogenic potential of various foods through their fatty acid compositions (Connor *et al.*, 1986). This index allows a comparison of the influence of different oil/fat compositions on the development of coronary diseases. The AI can be calculated by the ratio between the sum of lauric, myristic and palmitic acids and the sum of linoleic, linolenic and monounsaturated acids. Higher values of AI indicate higher possibilities for the development of coronary diseases (Kim *et al.*, 2008; Ulbrich, Southgate., 1991). AI increased from 0.0509 to 0.8210 and from 0.0571 to 0.8318 as a result of the increase in PO contents for the systems with and without HSO, respectively, as shown in Table 3. AI increased from 1 to 12%, due to the addition of 5% (w/w) HSO to the blends, indicating a very low change in this parameter for systems with higher fractions of PO. The blends with higher contents of CO were characterized for low values of IA, indicating no negative impact on health.

3.3. Solid fat content

Figures 1 and 2 show the increase in the SFC of the systems over the whole temperature range evaluated, due to the addition of FP in the mixtures containing PO and CO and the addition of HSO to the mixtures containing PO, CO, and FP.

FP had a structuring effect on the lipid systems which allowed a similar or higher SFC for blends with lower saturated fat acid contents. Nutritionally, this is a very beneficial effect since saturated fats, presenting negative effects on human health, are substituted by a functional class of compounds with high nutritional value. Similarly, HSO had a positive effect on the increase in SFC for systems added with FP. Although HSO is composed of saturated fatty acids, the addition of small fractions of this component enabled the substitution of higher amounts of PO, which contains high amounts of palmitic acid, to obtain a similar solid profile and diminish the final content of saturated fatty acids in the blends.

The solid profile is directly related to the selection of lipid systems for use in lipid food based products. At low temperatures (4 – 10 °C), this parameter

TABLE 2. Triacylglycerol classes (%) of the blends.

Blends	S ₃	S ₂ U	SU ₂	U ₃
100:0	10.79	35.80	39.03	14.38
80:20	8.63	28.64	33.86	28.87
60:40	6.47	21.48	28.69	43.36
40:60	4.32	14.32	23.52	57.84
20:80	2.16	7.16	18.35	72.33
0:100	0.00	0.00	13.18	86.82
100:0-FP-HSO	15.25	34.01	37.08	13.66
80:20-FP-HSO	13.20	27.21	32.17	27.42
60:40-FP-HSO	11.15	20.41	27.26	41.19
40:60-FP-HSO	9.10	13.60	22.34	54.95
20:80-FP-HSO	7.05	6.80	17.43	68.72
0:100-FP-HSO	5.00	0.00	12.52	82.48

PO: Palm oil, CO: canola oil. FP: Free Phytosterols; HSO: Hardfat of soybean oil. Blends: PO:CO (w:w): 100:0, 80:20, 60:40, 40:60, 20:80 and 0:100, added 10% of FP denominated 100:0FP, 80:20FP, 60:40FP, 40:60FP, 20:80FP, 0:100FP; U₃: triunsaturated triacylglycerols; S₂U: dissaturated triacylglycerols; SU₂: monosaturated triacylglycerols; U₃: triunsaturated triacylglycerols.

TABLE 3. Atherogenic index calculated from the ratio between the sums of lauric, miristic and palmitic acids and unsaturated fatty acids present in the blends (Kim *et al.*, 2008; Ulbricht *et al.*, 1991).

Blends	AI
100:0	0.82
80:20	0.57
60:40	0.40
40:60	0.26
20:80	0.14
0:100	0.05
100:0-FP-HSO	0.83
80:20-FP-HSO	0.58
60:40-FP-HSO	0.41
40:60-FP-HSO	0.26
20:80-FP-HSO	0.15
0:100-FP-HSO	0.05

PO: Palm oil, CO: canola oil. FP: Free Phytosterols; HSO: Hard fat of Soybean Oil Blends: PO:CO (w:w): 100:0, 80:20, 60:40, 40:60, 20:80 and 0:100, 10% of FP added denominated 100:0FP, 80:20FP, 60:40FP, 40:60FP, 20:80FP, 0:100FP.

determines the use of these systems in products kept under refrigeration, in which the maximum value of SFC is 32% at 10 °C for proper spreadability (Wassel; Young, 2007). The blends used for this purpose could be 60:40 and 40:60, 60:40-FP and 40:60-FP and 60:40-FP-HSO and 40:60-FP-HSO. From 20 – 22 °C, SFC higher than 10% is essential to the physical stability of the products and to avoid oil

exudation (Wassel; Young, 2007). Thus the blends meeting this requirement could be 60:40-FP, 80:20-FP, 100:0-FP, 40:60-FP-HSO, 60:40-FP-HSO, 80:20-FP-HSO and 100:0-FP-HSO. Considering that SFC at temperatures around 35 °C has a significant effect on properties like flavor release, softness and thermal resistance, the blends 60:40-FP, 80:20-FP, 100:0-FP and all blends containing HSO had more than 4% SFC, in contrast to the blends composed only of CO and PO, in which SFC was even lower than this value (Ribeiro *et al.*, 2013).

For application in processed foods, the structured blends composed of PO/OC added with FP or HSO may be indicated for use in margarine and spreads even with 4% SFC, because this shows more than 30% SFC at 10 °C and less than 5% SFC at 30 °C. The curve characteristics of the decrease in solids shows a more plastic consistency with slow melting and no waxy sensation. Possible mouthfeel (bitterness, texture, grainy feel) related to phytosterols can be minimized in emulsified systems. Furthermore, in processed products the structured blends would be incorporated as ingredients of the formulation, effectively reducing the possibility of sensory defects (Metzroth, 2015).

3.4. Consistency

The yield value (YV) increased as a function of PO addition to the blends, as observed in Figure 3. For blends composed of only CO and PO, contents lower than 40% CO resulted in yield values lower than the texture analyzer sensibility due to high fractions in the liquid phase. The addition of 10% FP resulted in a high increase, from 1.5 to 4.3 times for YV, resulting in a value of 28.4g/cm² or a blend with 20% PO. The addition of 5% HSO decreased the YV for the blends added with FP containing 100% and 80% PO, causing a mechanical weakness in the crystal network structure formed.

Correlating the YV with the SFC of the blends, strong exponential correlations were observed. These correlations were described by $YV = 20.443^{0.1426 \cdot SFC}$ ($R^2 = 0.9788$), $YV = 19,393^{0.171 \cdot SFC}$ ($R^2 = 0.9080$) and $YV = 20,16^{0.1565 \cdot SFC}$ ($R^2 = 0.9054$), respectively, for blends the addition of FP and HSO, the addition of FP and composed of only PO and CO. This behavior indicates that the resistance of the structure formed in the blends is strongly affected by SFC.

It was possible to improve the consistency of the fatty systems by adding only FP, obtaining similar or better results compared to the addition of FP and HSO, as can be observed from the YV for both types of systems. The use of only FP is nutritionally advantageous since it replaces saturated fatty acids and presents a set of health benefits.

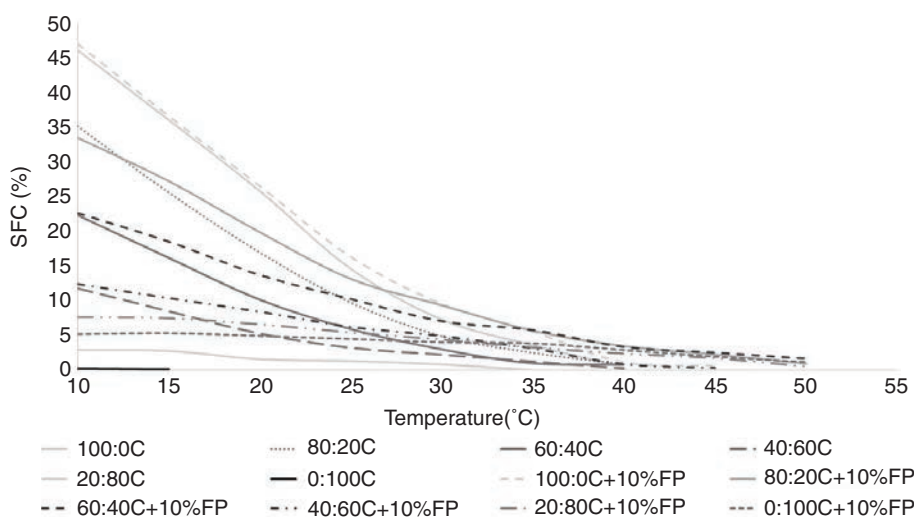


FIGURE 1. Solid fat content of lipid blends (100:0C, 80:20C, 60:40C, 40:60C, 20:80C, 0:100C) with the addition of phytosterols (100:0C+10%FL, 80:20C+10%FL, 60:40C+10%FL, 40:60C+10%FL, 20:80C+10%FL, 0:100C+10%FL) for temperatures from 10 °C to 50 °C (complete melting).

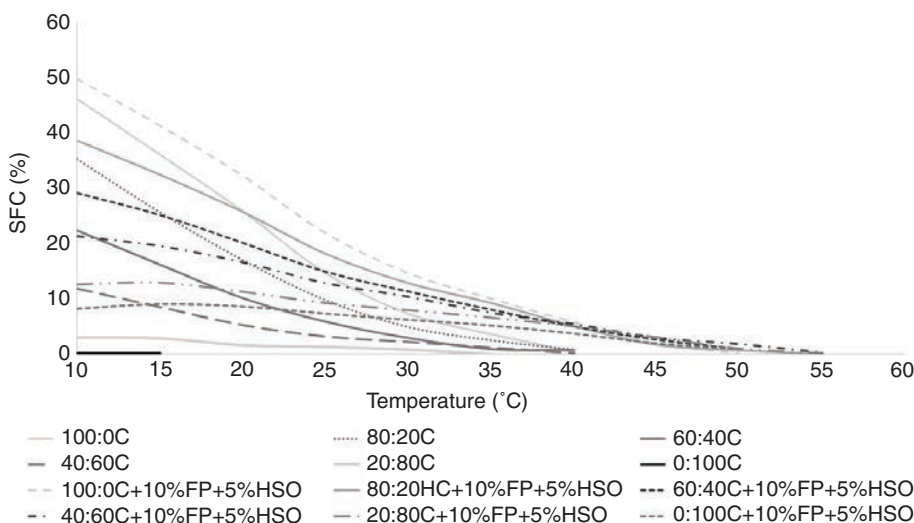


FIGURE 2. Solid fat content of lipid blends (100:0C, 80:20C, 60:40C, 40:60C, 20:80C, 0:100C) and addition of phytosterols and fully hydrogenated soybean oil (and addition of phytosterols (100:0C+10%FL, 80:20C+10%FL, 60:40C+10%FL, 40:60C+10%FL, 20:80C+10%FL, 0:100C+10%FL)) for temperatures from 10 °C to 50 °C (complete melting).

3.5. Microstructure

The formation of crystal structures was observed for all blends having a minimum PO content of 40%, as observed in Figure 4. A lower ratio between PO and CO resulted in the absence of crystallinity or in some few scattered structures, no formation of a crystal network. The microstructure of reference blends, composed of only PO and CO, showed crystal agglomerates with spherulitic form and increased crystal size as a function of the fraction of CO in the blends. On the other hand, the amount of crystals

was opposite to the unsaturated fatty acid content. Spherulites are formed due to crystal lamellae aggregation raising radially from the nuclei, with possible formation of branches with increasing size (Rousset, 2002). High size structures in systems containing high liquid fractions are the result of higher critical size of agglomerates since structures with lower sizes are not stable in this type of systems.

The FP addition resulted in the formation of a large amount of crystal structures with lower sizes than those in reference blends. The lower sizes are possibly related to the distribution of the

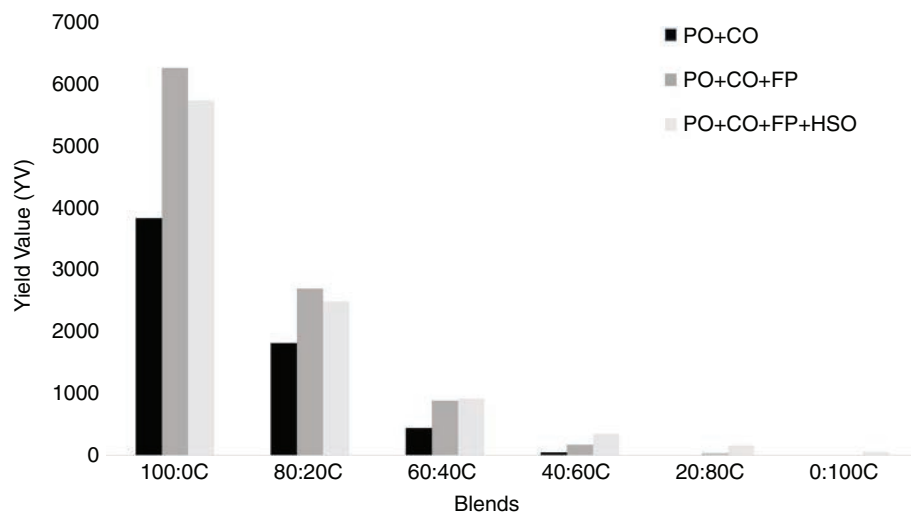


FIGURE 3. Consistency of the blends obtained through Campos methodology (2005) presented according to Yield value calculated by the Haighton equation (1959).

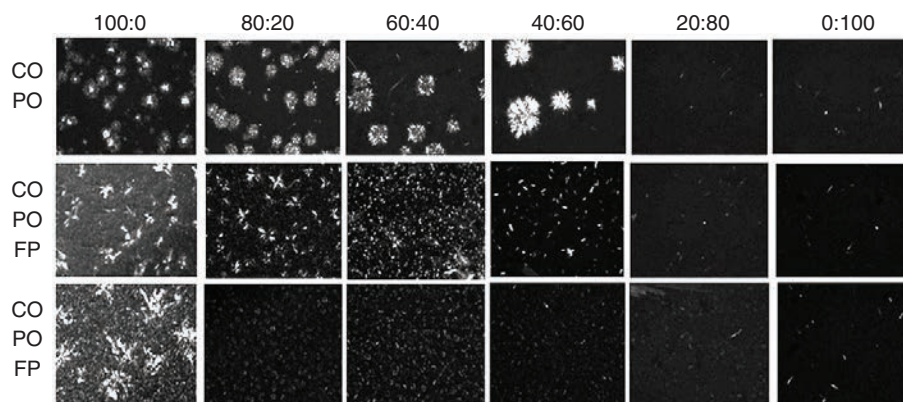


FIGURE 4. Crystalline networks of control blends (CO + PO), blends added with free phytosterols (CO + PO + FP) and free phytosterols and fully hydrogenated soybean oil (CO + PO + FP + HSO) obtained after maintaining at 25 °C for 2h at a magnification of 40 X (Campos, 2005)

triacylglycerols with higher melting points among the several crystal agglomerates formed by the FP added to the systems. By increasing the content of CO of the mixtures, lower melting point triacylglycerols are solubilized, leaving FP and S_3 triacylglycerols. The addition of HSO to the blends contributed to diminishing the crystal size of agglomerates and to raising the amount of crystal structures formed because high melting point material was introduced into the blends.

Sterols and sterol esters generally crystallize into bilayers roughly 4 nm thick. β -sitosterol, the major component of evaluated FP, has three known crystal forms: anhydrous crystals, hemi-hydrate crystals (1 mol of water to 2 mol of β -sitosterol) and mono-hydrate crystals (1 mol of water to 1 mol of β -sitosterol). These crystalline forms can change according to temperature, because the increase in

temperature results in a progressive dehydration of the crystals and the network is formed by fitting both molecules on top of each other, making it possible for the stacking to involve van der Waals interaction (Bot and Agterof, 2006; Co and Marangoni, 2012).

Both effects of FP and HSO on the microstructure of the systems are in agreement with the increase in YV of the blends. The large amount of agglomerates formed resulted in a dense crystal network with higher mechanical resistance, shown by higher consistency. Smaller agglomerates have greater superficial area, contributing to the formation of a large number of sites for binding among the crystals (Campos, 2005).

Despite being a study aimed at the application of lipid bases in foods, no evaluation of the impact of temperature variation on the microstructure and the formation of crystals was carried out.

3.6. Thermal behavior

The crystallization behavior of PO:CO blends was characterized by two main peaks (peak 1 and peak 2) as shown in Table 4 and Figures 5 to 10. The first one is a result of the high melting point

compounds, S_3 and SU_2 and the second one corresponds to a mixture of monounsaturated and S_2U . A third peak, mainly related to a fraction of U_3 , was observed at temperatures lower than -54°C . The diminishing of peak 1 and peak 2 as a function of CO addition is a result of the increase

TABLE 4. Thermal behavior parameters in crystallization obtained by differential scanning calorimeter (AOCS, 2009; Campos, 2005)

Samples	$T_{\text{initial}} (^{\circ}\text{C})$	$T_{\text{peak1}} (^{\circ}\text{C})$	$T_{\text{peak2}} (^{\circ}\text{C})$	$T_{\text{peak3}} (^{\circ}\text{C})$	$T_{\text{final}} (^{\circ}\text{C})$	$\Delta H_1 (\text{J/g})$	$\Delta H_2 (\text{J/g})$	$\Delta H_3 (\text{J/g})$
100:0C	19.38	18.11	1.31	-51.51	-59.12	9.91	46.25	1.93
20:80C	6.29	3.32	-12.87	-58.91	-59.55	2.14	11.28	1.66
40:60C	12.33	10.49	-7.22	-59.14	-59.69	4.01	16.83	3.86
60:40C	15.49	13.82	-4.11	-57.17	-59.55	6.02	24.7	3.59
80:20C	17.65	16.25	-1.11	-53.83	-59.26	8.03	34.82	2.35
0:100C	-13.26	-18.67	-59.02	-	-59.55	5.86	1.30	-
100:0FP	18.23	16.06	-1.84	-59.68	-60.13	7.41	26.57	0.44
20:80FP	8.31	3.56	-14.44	-59.66	-59.84	1.37	6.96	0.52
40:60FP	12.62	9.99	-10.53	-59.69	-59.98	3.08	14.30	0.43
60:40FP	16.36	12.07	-7.12	-59.88	-59.98	4.31	16.33	1.89
80:20FP	17.51	13.82	-5.11	-59.83	-59.25	6.24	21.01	1.21
0:100FP	-13.69	-19.31	-54.23	-	-59.00	0.87	5.05	-
100:0 FP – HSO	37.20	24.14	-0.95	-59.67	-59.98	11.68	26.56	0.44
20:80 FP – HSO	38.21	24.91	-13.61	-59.84	-60.13	5.86	7.44	1.49
40:60 FP – HSO	34.62	25.29	-9.58	-59.89	-60.15	9.04	11.66	1.70
60:40 FP – HSO	36.92	24.22	-6.16	-59.83	-60.13	9.89	16.69	1.18
80:20 FP – HSO	32.31	23.85	-3.91	-59.80	-59.98	10.64	20.83	1.09
0:100 FP – HSO	35.48	24.75	-38.18	-59.82	-60.11	5.31	2.86	1.80

Blends: PO:OC (w:w): 100:0, 80:20, 60:40, 40:60, 20:80 and 0:100, 10% added FP denominated 100:0FP, 80:20FP, 60:40FP, 40:60FP, 20:80FP, 0:100FP. PO: Palm oil, CO: canola oil. $T_{\text{initial}} (^{\circ}\text{C})$: Initial temperature of crystallization; $T_{\text{peak 1}} (^{\circ}\text{C})$: Temperature of peak 1; $T_{\text{peak 2}} (^{\circ}\text{C})$: Temperature of peak 2; $T_{\text{peak 3}} (^{\circ}\text{C})$: Temperature of peak 3; $T_{\text{final}} (^{\circ}\text{C})$: Final temperature of crystallization. $\Delta H_1 (\text{J/g})$: Enthalpy of Peak 1; $\Delta H_2 (\text{J/g})$: Enthalpy of peak 2; $\Delta H_3 (\text{J/g})$: Enthalpy of peak 3.

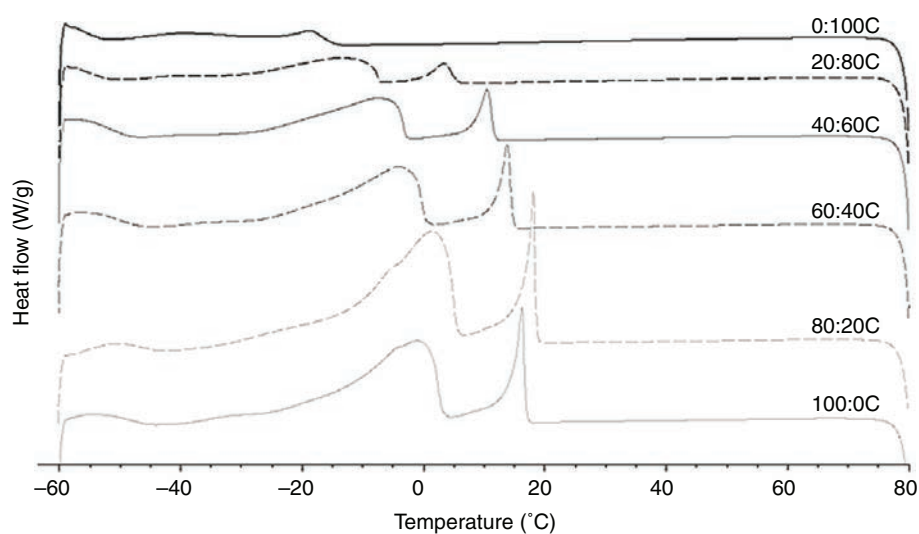


FIGURE 5. Crystallization behavior for the control blends (100:0C, 80:20C, 60:40C, 40:60C, 20:80C, 0:100C).

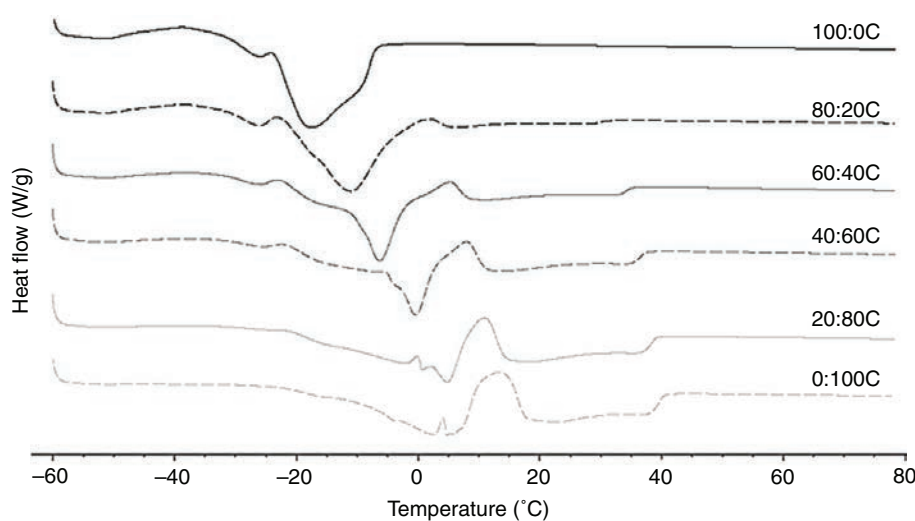


FIGURE 6. Melting behavior for the control blends (100:0C, 80:20C, 60:40C, 40:60C, 20:80C, 0:100C).

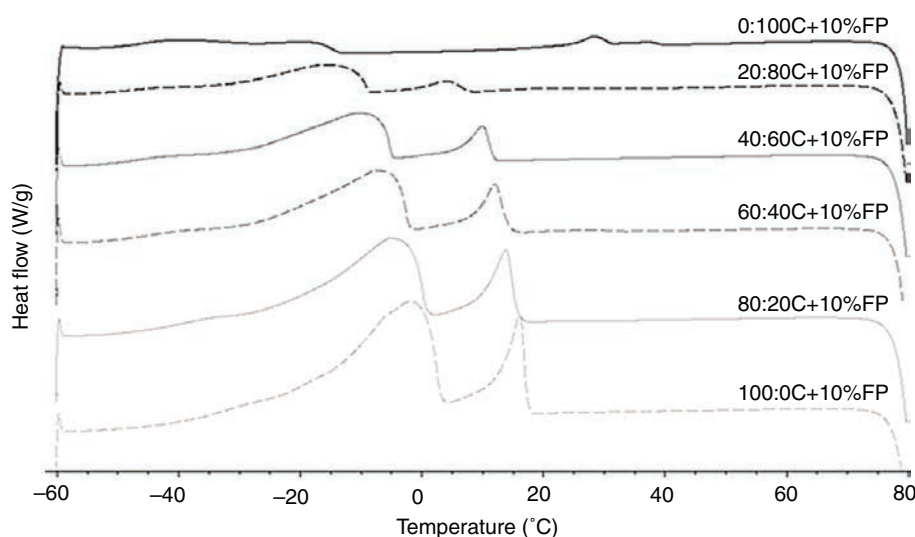


FIGURE 7. Crystallization behavior for the blends added with 10% free phytosterols (100:0C+10%FL, 80:20C+10%FL, 60:40C+10%FL, 40:60C+10%FL, 20:80C+10%FL, 0:100C+10%FL).

in U_3 and consequent decrease in the content of compounds crystallizing at this temperature range.

For the systems without PO, the FP addition caused a shifting of the peak 1 from -18.67°C (0:100) to 28.11°C (0:100-FP). Observing all other blends, the presence of FP, in general terms, did not modify their crystallization profile. The low effect on crystallization profile of these blends is due to the high melting point of FP, about 140°C for FP, and their low solubility in fatty systems, 2 – 5% ($25 - 50^\circ\text{C}$) in corn oil, according to data reported by Vaikousi

et al., 2007. Once the solubilized FP co-crystallize with triacylglycerols, and they are mainly responsible for the modification of the crystallization behavior of the systems. In these blends, a great fraction of these compounds was probably not solubilized, thus causing little effect on the thermal behavior of the fatty mixtures.

On the other hand, the addition of HSO modified the crystallization profile of all the blends. The initial crystallization temperatures and the temperatures of peak 1, different for each system with no addition of HSO, changed in about the same

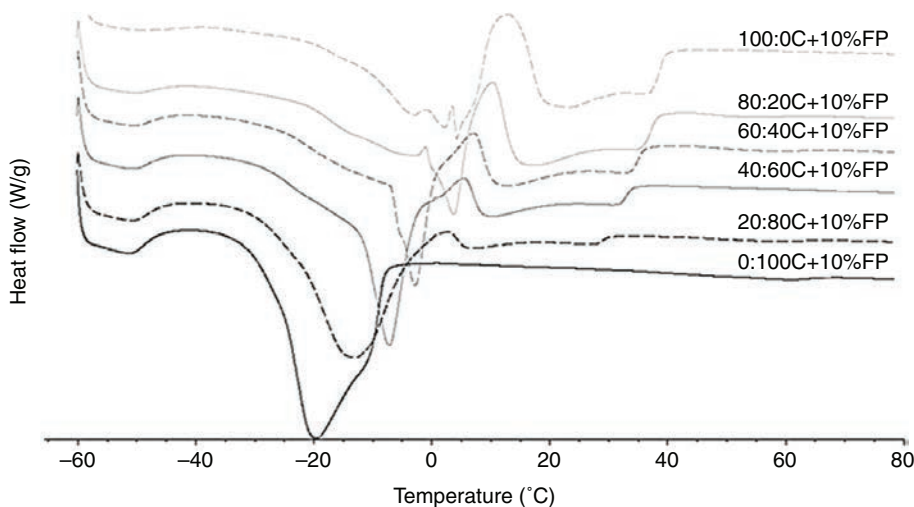


FIGURE 8. Melting behavior for the blends added with 10% free phytosterols (100:0C+10%FL, 80:20C+10%FL, 60:40C+10%FL, 40:60C+10%FL, 20:80C+10%FL, 0:100C+10%FL).

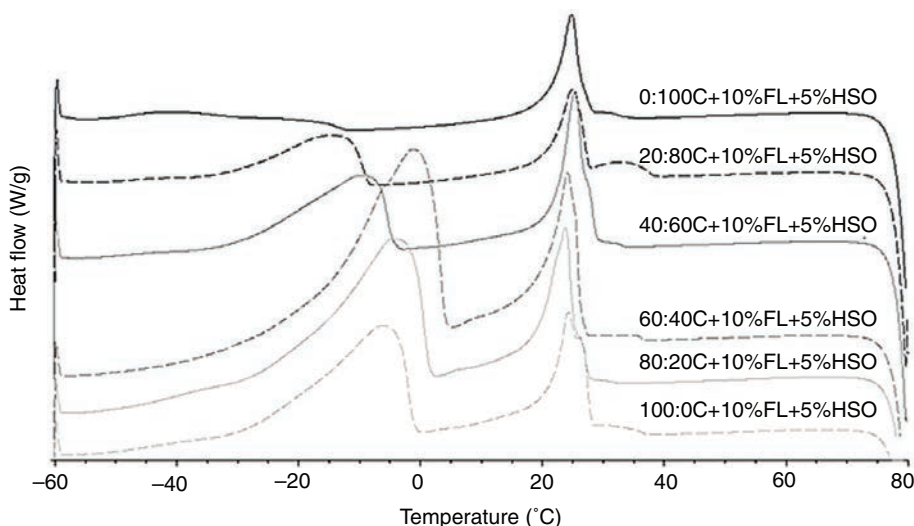


FIGURE 9. Crystallization behavior for the blends added with 10% free phytosterols and 5% hydrogenated soybean oil (100:0C + 10%FL + 5% HSO, 80:20C + 10%FL + 5% HSO, 60:40C + 10%FL + 5% HSO, 40:60C + 10%FL + 5% HSO, 20:80C + 10%FL + 5% HSO, 0:100C + 10%FL + 5% HSO)

temperature range, respectively, from 32.3 – 38.2 °C and 24.1 – 25.2 °C, which was determined by the melting temperature of the HSO. The ΔH for peak 1 also was higher than for the blends with no addition of HSO, since HSO added to the systems crystallizes at this temperature range.

In general, the addition of FP into the systems did not cause changes in the crystallization profile of the blends. On the other hand, HSO addition caused displacement at the end of the crystallization process to higher temperature ranges. In addition, a peak at a higher temperature range was observed even for blends with high contents of CO. These two

effects are the result of the introduction of S_3 with high melting points.

3.7. Polymorphism

The X-ray diffraction of the reference blends (with no addition of FP), presented characteristic β' -form profile, with short spacings of about 3.8 and 4.2 Å for blends containing from 100 to 60% PO, while for systems with higher contents of CO, no crystallinity was obtained, as showed in Table 5 and Figure 11. These polymorphs are result of the crystallization of high melting point

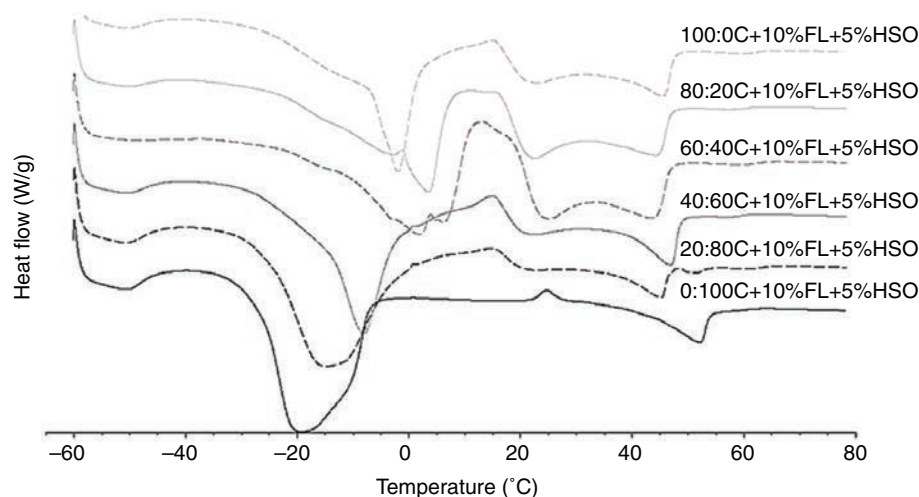
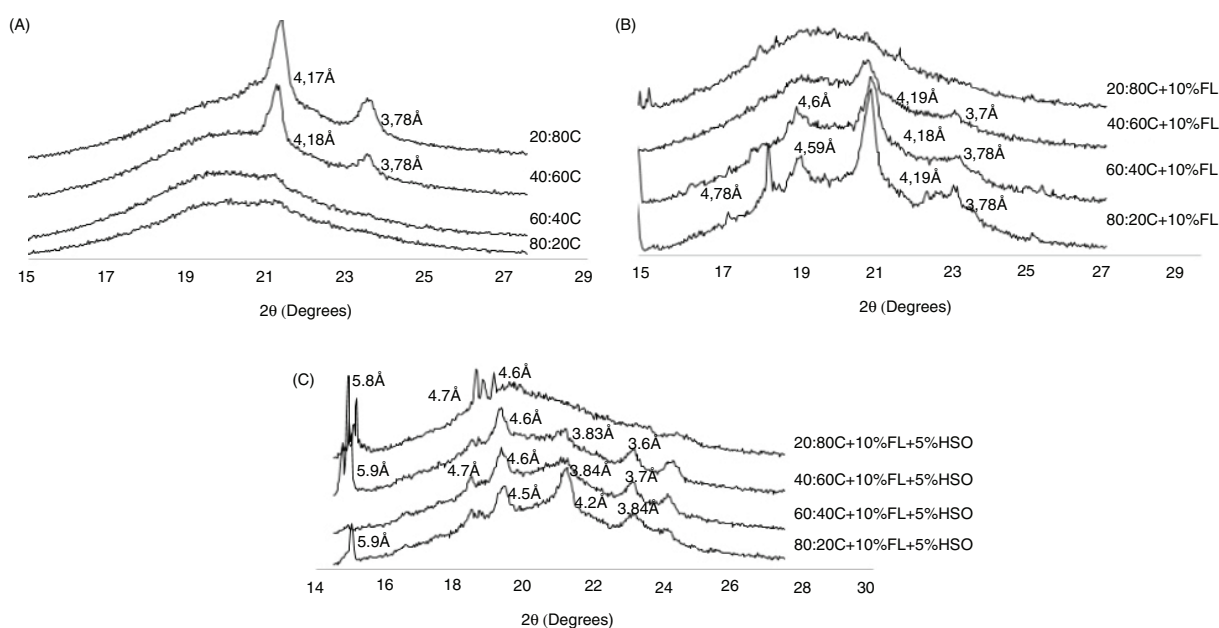


FIGURE 10. Melting behavior for the blends added with 10% free phytosterols and 5% hydrogenated soybean oil (100:0C + 10%FL + 5% HSO, 80:20C + 10%FL + 5% HSO, 60:40C + 10%FL + 5% HSO, 40:60C + 10%FL + 5% HSO, 20:80C + 10%FL + 5% HSO, 0:100C + 10%FL + 5% HSO) .



*The peaks that was not recognized as a polymorphic form is probably a free phytosterol peak.

FIGURE 11. Difratograms of blends; (A) Control blends; (B) Control blends added with 10%FP; (C) Control blends added with 10%FP and 5% HSO.

triacylglycerols from PO, since the most unsaturated triacylglycerols from CO are in liquid phase. The addition of FP to the reference blends (Figure 11, b) resulted on the formation of a β crystal polymorph, as observed from the rising of short spacing of about 4.6 Å, simultaneously to the presence of β' -form. In addition, the FP kept

the crystallinity of the blends up to 80% of CO. It was also observed for these blends a X-ray diffraction peak at 2θ of about 15.02° representing the crystal arrangement of the FP molecules. Vaikousi *et al.*, 2007 reported X-ray diffraction lines related to FP at 2θ of 4.6, 6.9, 9.35, 11.7, 14.85, 16.5, 18.4 and 21.9° .

TABLE 5. Characterization of polymorphic form in the samples through AOCS official methodology (2009)

Blends	Short spacings (Å)	Polymorphic form
100:0C	4.18 – 3.78	β'
80:20C	4.17 – 3.78	β'
60:40C	4.18 – 3.78	β'
40:60C	-	-
20:80C	-	-
0:100C	-	-
100:0FP	4.18 – 4.58 – 3.77	$\beta + \beta'$
80:20FP	4.19 – 4.59 – 3.78	$\beta + \beta'$
60:40FP	4.60 – 4.18 – 3.78	$\beta + \beta'$
40:60FP	4.60 – 4.19 – 3.78	$\beta + \beta'$
20:80FP	-	-
0:100FP	-	-
100:0 FP – HSO	4.59 – 4.19 – 3.84	$\beta + \beta'$
80:20 FP – HSO	4.59 – 4.19 – 3.84	$\beta + \beta'$
60:40 FP – HSO	4.59 – 3.84	$\beta + \beta'$
40:60 FP – HSO	4.58 – 3.83	$\beta + \beta'$
20:80 FP – HSO	4.60	β
0:100 FP – HSO	4.60	β

^a-. Sample liquids in the temperature of analysis.

The systems containing FP and HSO (Figure 11, c), presented β -form and β' -form, and the peak related to FP (2θ about 15°) for CO contents up to 60%. The diffractograms of systems 20:80-FP-HSO and 0:100-FP-HSO showed only crystal polymorph and the peak related to FP, without β' form. In these last two systems there are great amount of triacylglycerols in liquid phase, once high fractions of CO are present, solubilizing less stable polymorphs and facilitating the transition β' -form to β -form due to high molecular mobility for the crystal clusters. FP induce crystallization at β -form, simultaneously to a previous original β' -form, possibly working as impurities in which solid surface the triacylglycerols can bind, leading to a heterogeneous crystallization process. This mechanism probably changes the triacylglycerol arrangement in crystal structure, modifying the polymorphism of a fraction of the total triacylglycerols in the blends. In turn, HSO has high content of symmetric triacylglycerols, a type of molecule that promotes β polymorph, leading to formation of this kind of arrangement in fatty systems.

4. CONCLUSIONS

The addition of 10% FP into blends of PO:CO, and its association with low fractions of HSO shows promising results for the modification of the physical properties and crystallization behavior of these systems. The results of fatty acid composition show

that it was possible to reduce the saturated fatty acid contents in the control blends (reduction of 39.5%) and in the blends containing HSO (reduction of 37.5%), with respective increase in monounsaturated and polyunsaturated fatty acids. FP allows obtaining blends with lower amounts of PO and similar, or more stable in relation to temperature, consistency, solid fat content and thermal behavior. The addition of this compound improved the crystalline network of the systems, decreased the average crystal sizes and increased the crystallinity of the systems with high contents of CO. In addition, FP promoted changes in the polymorphism of the fatty systems, causing the rising β -form simultaneously to β' -form. HSO associated to FP, shifted the crystallization of the fatty mixtures to higher temperatures, increasing their SFC and the crystal networks, and decreasing crystal sizes. Additionally, HSO promote polymorphic transition β' -form to β -form for systems with higher contents of CO.

This study showed the effectiveness of the structuring of lipid blends for applications in foods. According to the solid fat content results, easily related to industrial application, the blends 60:40 and 40:60, 60:40-FP and 40:60-FP and 60:40-FP-HSO and 40:60-FP-HSO can be used in products which require good spreadability, such as margarine and spreads; and the blends 60:40-FP, 80:20-FP, 100:0-FP, 40:60-FP-HSO, 60:40-FP-HSO, 80:20-FP-HSO and 100:0-FP-HSO can be used in products to avoid oil exudation, as spreads at intermediate temperatures (25°C). Subsequent steps to this study involve the incorporation of the developed blends in food formulations, verifying their interaction with other ingredients and sensorial characteristics.

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REFERENCES

- AOCS. American Oil Chemists' Society, 2009. Official methods and recommended practices of the American Oil Chemists' Society, 6th Editio. ed. Champaign. <http://dx.doi.org/10.1002/0471740039.vec0152>
- Auriou N. 2004. Stabilized dispersion of phytosterol in oil. US 20040156887.
- Bot A, Agterof WGM. 2006. Structuring of edible oils by mixtures of γ -oryzanol with β -sitosterol or related phytosterols. *J. Am. Oil Chem. Soc.* **83**, 513–521.
- Campos R. 2005. Fat Crystal Networks, in: Fat Crystal Networks. Marcell Dekker Inc., New York, pp. 267–349.
- Cerdeira M, Martini S, Candal RJ, Herrera, ML, 2006. Polymorphism and growth behavior of low-trans fat blends formulated with and without emulsifiers. *J. Am. Oil Chem. Soc.* **83**, 489–496. <https://doi.org/10.1007/s11746-006-1231-x>
- Co ED, Marangoni AG. 2012. Organogels: An alternative edible oil-structuring method. *J. Am. Oil Chem. Soc.* **89**, 749–780. <http://doi.org/10.1007/s11746-012-2049-3>

- Connor SL, Gustafson JR, Flavell DP, Artaud-wild SM, Hatcher CJCLF, Connor WE. 1986. The cholesterol/saturated-fat index: An indication of the hypercholesterolaemic and atherogenic potential of food. *Lancet* **8492**, 1229–1232.
- Dhaka V, Gulia N, Ahlawat KS, Khatkar BS. 2011. Trans fats-sources, health risks and alternative approach - A review. *J. Food Sci. Technol.* **48**, 534–541. <https://doi.org/10.1007/s13197-010-0225-8>
- Dubois V, Breton S, Linder M, Fanni J, Parmentier M. 2007. Fatty acid profiles of 80 vegetable oils with regard to their nutritional potential. *Eur. J. Lipid Sci. Technol.* **109**, 710–732. <http://doi.org/10.1002/ejlt.200700040>
- Gebauer SK, Tracy RP, Baer DJ. 2014. Impact of stearic acid and oleic acid on hemostatic factors in the context of controlled diets consumed by healthy men. *Eur. J. Clin. Nutr.* **2600**, 1–3. <http://doi.org/10.1038/ejcn.2014.62>
- Gunstone FD, Harwood JL, Dijkstra AT. 2007. The Lipid Handbook, in: The Lipid Handbook. CRC Press, Boca Raton, pp. 37–141.
- Haighton AJ. 1959. The measurement of the hardness of margarine and fats with cone penetrometers. *J. Am. Oil Chem. Soc.* **36**, 345–348. <http://doi.org/10.1007/BF02640051>
- Hartman L, Lago RCA. 1973. No Title. Lab. Pract. **22**, 475–476.
- Herrera ML, Marquez Rocha FJ. 1996. Effects of sucrose ester on the kinetics of polymorphic transition in hydrogenated sunflower oil. *J. Am. Oil Chem. Soc.* **73**, 321–326. <http://doi.org/10.1007/BF02523425>
- Kim BH, Lumor SE, Akoh CC. 2008. trans -Free Margarines Prepared with Canola Oil / Palm Stearin / Palm Kernel Oil-Based Structured Lipids. *J. Agric. Food Chem.* **56**, 8195–8205.
- Lida HMDN, Ali RM. 1998. Physico-chemical characteristics of palm-based oil blends for the production of reduced fat spreads. *J. Am. Oil Chem. Soc.* **75**, 1625–1631. <http://doi.org/10.1007/s11746-998-0103-y>
- Metzroth DJ. 2015. Shortenings: Science and Technology, in: Bailey's Industrial Oil and Fat Products.
- Moreau RA, Whitaker BD, Hicks KB. 2002. Phytosterols, phytostanols, and their conjugates in foods: Structural diversity, quantitative analysis, and health-promoting uses. *Prog. Lipid Res.* **41**, 457–500. [http://doi.org/10.1016/S0163-7827\(02\)00006-1](http://doi.org/10.1016/S0163-7827(02)00006-1)
- Noakes M, Clifton PM, Doornbos AME, Trautwein EA. 2005. Plant sterol ester-enriched milk and yoghurt effectively reduce serum cholesterol in modestly hypercholesterolemic subjects. *Eur. J. Nutr.* **44**, 214–222. <http://doi.org/10.1007/s00394-004-0513-z>
- O'Brien RD. 2004. Fats and oils: formulating and processing for applications, New York.
- Oliveira GM, Badan Ribeiro AP, dos Santos AO, Cardoso LP, Kieckbusch TG. 2015. Hard fats as additives in palm oil and its relationships to crystallization process and polymorphism. *LWT - Food Sci. Technol.* **63**, 1163–1170. <http://doi.org/10.1016/j.lwt.2015.04.036>
- Pernetti M, van Malssen KF, Flöter E, Bot A. 2007. Structuring of edible oils by alternatives to crystalline fat. *Curr. Opin. Colloid Interface Sci.* **12**, 221–231. <http://doi.org/10.1016/j.cocis.2007.07.002>
- Przybylski R, Mag T, Eskin NM, McDonald BE. 2005. Canola Oil. *Bailey's Ind. Oil Fat Prod.* 61–121. <http://doi.org/10.1002/047167849X.bio004>
- Raczyk M, Kmieciak D, Przybylski R, Rudzińska M. 2017. Effect of Fatty Acid Unsaturation on Phytosterol Ester Degradation. *J. Am. Oil Chem. Soc.* **94**, 701–711. <http://doi.org/10.1007/s11746-017-2979-x>
- Ribeiro APB, Basso RC, dos Santos AO, Andrade GC, Cardoso LP, Kieckbusch TG. 2013. Hardfats as crystallization modifiers of cocoa butter. *Eur. J. Lipid Sci. Technol.* **115**, 1462–1473.
- Ribeiro APB, Masuchi MH, Miyasaki EK, Domingues MAF, Stroppa VLZ, de Oliveira GM, Kieckbusch TG. 2015. Crystallization modifiers in lipid systems. *J. Food Sci. Technol.* **52**, 3925–3946. <http://doi.org/10.1007/s13197-014-1587-0>
- Rogers MA, Strober T, Bot A, Toro-Vazquez JF, Stortz T, Marangoni AG. 2014. Edible oleogels in molecular gastronomy. *Int. J. Gastron. Food Sci.* **2**, 22–31. <http://doi.org/10.1016/j.ijgfs.2014.05.001>
- Roussel P. 2002. Physical Properties of Lipids, in: Physical Properties of Lipids. Marcell Dekker Inc., New York, pp. 1–36.
- Rozner S, Garti N. 2006. The activity and absorption relationship of cholesterol and phytosterols. *Colloids Surfaces A Physicochem. Eng. Asp.* **282–283**, 435–456. <http://doi.org/10.1016/j.colsurfa.2005.12.032>
- Salo P, Wester I. 2005. Low-fat formulations of plant stanols and sterols. *Am. J. Cardiol.* **96**, 10–13. <http://doi.org/10.1016/j.amjcard.2005.03.021>
- Senate Commission on Food Safety, 2014. Phytosterol oxidation products in foods: Analysis, occurrence, exposure and biological effects. *Dtsch. Forschungsgemeinschaft* **34**. <http://doi.org/10.13140/2.1.1369.4248>
- Smith KW, Bhagga K, Talbot G, Van Malssen K.F. 2011. Crystallization of fats: Influence of minor components and additives. *J. Am. Oil Chem. Soc.* **88**, 1085–1101. <http://doi.org/10.1007/s11746-011-1819-7>
- Timms RE. 1984. Phase behaviour of fats and their mixtures. *Prog. Lipid Res.* **23**, 1-38. [https://doi.org/10.1016/0163-7827\(84\)90004-3](https://doi.org/10.1016/0163-7827(84)90004-3)
- Ulbricht TLV, Southgate DT. 1991. Coronary heart disease: seven dietary factors. *Lancet* **338**, 985–992. [http://doi.org/10.1016/0140-6736\(91\)91846-M](http://doi.org/10.1016/0140-6736(91)91846-M)
- Vaikousi H, Lazaridou A, Biliaderis CG, Zawistowski J. 2007. Phase transitions, solubility, and crystallization kinetics of phytosterols and phytosterol-oil blends. *J. Agric. Food Chem.* **55**, 1790–1798. <http://doi.org/10.1021/jf0624289>
- Wassell P, Bonwick G, Smith CJ, Almiroin-Roig E, Young NWG. 2010. Towards a multidisciplinary approach to structuring in reduced saturated fat-based systems – a review. *Int. J. Food Sci. Technol.* **45**, 642–655.
- Wassell P, Young NWG. 2007. Food applications of trans fatty acid substitutes. *Int. J. Food Sci. Technol.* **42**, 503–517. <http://doi.org/10.1111/j.1365-2621.2007.01571.x>
- Yap P, Man J, Man L. 1989. Polymorphism of palm oil and palm oil products. *J. Am. Oil Chem. Soc.* **66**, 693–697. <https://doi.org/10.1007/BF02669954>
- Zevenbergen H, De Bree A, Zeelenberg M, Laitinen K, Van Duijn G, Flöter E. 2009. Foods with a high fat quality are essential for healthy diets. *Ann. Nutr. Metab.* **54**, 15–24. <http://doi.org/10.1159/000220823>