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Cannabinoids on the Brain

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Cannabis has a long history of consumption both for recreational and medicinal uses. Recently there have been significant advances in our understanding of how cannabis and related compounds (cannabinoids) affect the brain and this review addresses the current state of knowledge of these effects. Cannabinoids act primarily via two types of receptor, CB₁ and CB₂, with CB₁ receptors mediating most of the central actions of cannabinoids. The presence of a new type of brain cannabinoid receptor is also indicated. Important advances have been made in our understanding of cannabinoid receptor signaling pathways, their modulation of synaptic transmission and plasticity, the cellular targets of cannabinoids in different central nervous system (CNS) regions and, in particular, the role of the endogenous brain cannabinoid (endocannabinoid) system. Cannabinoids have widespread actions in the brain: in the hippocampus they influence learning and memory; in the basal ganglia they modulate locomotor activity and reward pathways; in the hypothalamus they have a role in the control of appetite. Cannabinoids may also be protective against neurodegeneration and brain damage and exhibit anticonvulsant activity. Some of the analgesic effects of cannabinoids also appear to involve sites within the brain. These advances in our understanding of the actions of cannabinoids and the brain endocannabinoid system have led to important new insights into neuronal function which are likely to result in the development of new therapeutic strategies for the treatment of a number of key CNS disorders.

KEY WORDS: cannabis, cannabinoids, marijuana, CB₁, endocannabinoids, central nervous system (CNS), brain, synaptic transmission, synaptic plasticity, function, reward, cognitive function, attention, pain, neurodegeneration, epilepsy, appetite, therapeutic potential, review

DOMAINS: learning and memory, intercellular communication, neuroscience, signaling

INTRODUCTION

The hemp plant *Cannabis sativa*, often used in the herbal form (marijuana) or as resin (hashish), has a long history of consumption, both for recreational and medicinal uses; indeed, cannabis has widespread actions in the body, including the modulation of a number of key processes within the central nervous system (CNS). Some of these actions are described as undesirable or unpleasant whereas others have considerable therapeutic properties. Recently there have been significant advances in our understanding of how cannabis and related compounds (cannabinoids) affect the CNS.

The term *cannabinoid* originally described a family of chemically related 21-carbon alkaloids found uniquely in the cannabis plant, of which Δ^9 -tetrahydrocannabinol (Δ^9 -THC) is the principal active constituent (first isolated in 1964 by Gaoni and Mechoulam[1]); however, this term also now encompasses all those substances capable of activating cannabinoid receptors.

In animals, cannabinoid agonists produce a characteristic combination of four symptoms: hypothermia, analgesia, hypoactivity, and catalepsy[2]. In humans, where the majority of information relates to ingestion of the marijuana plant, the most familiar psychoactive effect is a state of mild euphoria (or "high"); however, cannabis also produces a wide range of other effects including an enhancement of sensory perception, tachycardia, antinociception, difficulties in concentration, suppression of nausea, and an increased appetite, together with impairment of linear thinking, memory, and locomotor and psychomotor skills (for detailed reviews see[3,4,5,6]). Adverse effects also include depersonalization and panic attacks, with frequent use by young adolescents contributing to an amotivational syndrome[3]. In terms of dependence, withdrawal symptoms from cannabis in man are thought to be mild and short lived; indeed, compared to cocaine, alcohol, opiates, and nicotine, marijuana has little addictive power, although dependence is observed in a significant minority of regular cannabis users. Chronic, high doses of cannabis may cause a subtle impairment of cognitive performance that can persist after cannabis use is discontinued, and these may be related to neurotoxic effects[7].

PHARMACOLOGY: RECEPTOR AND NONRECEPTOR-MEDIATED ACTIONS IN THE CNS

Two types of G-protein-linked cannabinoid receptors (CB₁ and CB₂) have been identified using molecular and pharmacological approaches[8], with CB₁ receptors expressed predominantly in neurons of the brain, spinal cord, and peripheral nervous system and CB₂ receptors present mainly on immune cells. A splice variant of the CB₁ receptor, CB_{1A}, has been identified in brain[9] and exhibits all the properties of CB₁, but to a slightly attenuated extent[10]; however, the functional significance of this splice variant is questionable as other groups have not found evidence for its expression. The CB₂ receptor has only 44% overall nucleotide sequence identity with the CB₁ receptor. Unlike the CB₁ receptors, there is no evidence for CB₂ receptor binding, protein, or mRNA in the brain[11] although there are some functional data pointing to the presence of CB₂ receptors in cultured cerebellar granule cells[12]. Advances in cannabinoid pharmacology have generated a number of selective agonists and antagonists for these receptor subtypes (reviewed in detail elsewhere)[8,13].

While the majority of the physiological actions of cannabinoids in the CNS are mediated by activation of CB_1 receptors, there are some notable exceptions. Recent studies with CB_1 receptor knockout mice have shown some residual CNS actions of cannabinoids on synaptic transmission[14] and point to the existence of a novel type of G-protein-coupled cannabinoid receptor[15]; but this result remains to be confirmed using molecular cloning approaches. The endogenous cannabinoid anandamide also has activity at vanilloid receptors (VR1)[16], blocks directly the background K⁺ channel TASK-1[17], and may directly interact with NMDA

receptors[18]. Likewise, cannabidiol activates VR1 receptors but also inhibits the cellular uptake and hydrolysis of anandamide[19]; indeed, some of the nonreceptor-mediated effects of cannabinoid compounds may be secondary to effects on metabolism of the active compounds. This distinction could be an important issue when working with cannabis extracts. Cannabinoids are reported to interact with 5HT-3 receptor channels at relatively low concentrations[20], and anandamide may directly interact with nicotinic acetylcholine receptors[21]. Anandamide also inhibits binding to the muscarinic acetylcholine receptors via a cannabinoid receptor-independent mechanism[22]. The synthetic, nonpsychoactive cannabinoid-like compound HU-211 directly antagonizes NMDA receptors but does not bind to cannabinoid receptors[23]. There is also evidence cardiovascular for nonreceptor-mediated neuroprotective (antioxidant) and effects of cannabinoids [24,25,26]. Nonreceptor-mediated responses to relatively low (up to 1 μM) concentrations of cannabinoid drugs include effects on neuronal uptake of certain neurotransmitters, inhibition of membrane-bound enzymes, and the perturbation of membrane phospholipids[27].

CELLULAR LOCALIZATION OF CB1 RECEPTORS

 CB_1 receptors display a unique CNS distribution and are present in the mammalian brain at higher levels than most known G-protein-coupled receptors (GPCRs). The patterns of distribution of CB₁ receptors have been studied mainly with radioligand binding and autoradiography and with immunohistochemical techniques. Tritiated cannabinoid receptor ligands that have been the most widely used in binding assays or for autoradiography are the CB₁-selective [³H]SR141716A, and [³H]CP55940, [³H]WIN55212-2, and [³H]HU-243. The latter three are agonists that bind equally well to both CB1 and CB2 receptors. Whole-brain cannabinoid receptor density rivals that of glutamate[28] and GABA[29]. In certain regions, cannabinoid receptor levels are similar to those for dopamine and greatly exceed those for neuropeptide receptors. Computer-assisted densitometric analyses carried out on film autoradiographs taken from dog, guinea pig, rat, monkey, and human brains[30] show the highest levels of binding associated with the globus pallidus and substantia nigra pars reticulata and the molecular layers of the cerebellum and the dentate gyrus of the hippocampus. Regions showing strong binding include other areas of the hippocampal formation, the cerebral cortex, and the striatum. The hypothalamus, basal ganglia, solitary tract nucleus, and spinal cord show moderate levels of radioligand binding, with slight differences between species[30].

Immunocytochemical studies indicate that CB₁ receptor distribution resembles that of radioligand binding sites for cannabinoids, with relatively high levels of expression in areas such as the olfactory system, hippocampal formation, neocortex, and cerebellum[31,32,33,34]. The heterogeneity of CB₁ receptor expression relates to areas associated with the characteristic effects of cannabinoid agonists on motor coordination and posture, hypothermia, learning and memory, appetite, nausea, and antinociception, supporting the hypothesis that cannabinoid effects in the brain are mediated via CB₁ receptors. At the cellular level, CB₁ receptor immunoreactivity is particularly associated with nerve fiber systems and axon terminals, but with little neuronal somatic expression, which is compatible with a presynaptic site of action. In the rat hippocampus, an area that has been studied in much detail, axonal CB_1 receptor immunoreactivity has been associated mainly with presynaptic GABAergic nerve endings of cholecystokinin-containing basket cells[35,36,37], and there is limited[32] but controversial[31,35,38] immunohistochemical evidence for the expression of CB_1 receptors on glutamatergic cells; however, in situ hybridization studies suggest that CB_1 receptors may be present, but at much lower levels than on GABAergic neurons[39]. Antibodies raised against extracellular epitopes of CB_1 receptors can be used to study their surface expression in living cells (Fig. 1). Cell surface CB₁ receptor immunolabeling in hippocampal neurons is also expressed at high levels at GABAergic synaptic terminals, with no detectable labeling of glutamatergic neurons[40].

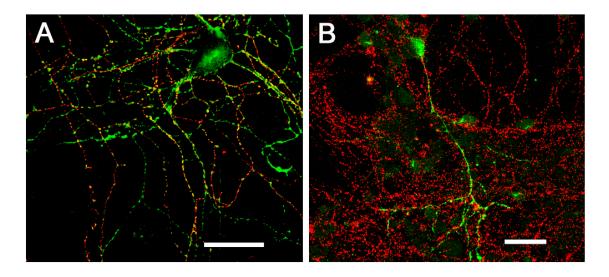


FIGURE 1. A comparison of cell surface CB_1 receptor immunoreactivity in cultured hippocampal neurons and cerebellar granule cells. (A,B) dual labeling of surface CB_1 receptors (red) and glutamate decarboxylase (GAD), a marker for inhibitory terminals/neurons (green). The extent of colocalization is shown by the yellow regions of overlap. Both cultures express strong, punctate CB_1 receptor immunoreactivity on fine fibers. Note the marked correspondence between CB_1 receptor label and clusters of GAD immunoreactivity in the cultured hippocampal neurons (A), but not in the cerebellar granule cells (B). An antibody raised against the N-terminus of the cloned rat CB_1 receptor was used to detect cell surface immunoreactivity. Scale bars, 20 μ m.

CANNABINOID SIGNAL TRANSDUCTION

Two of the most widely described intracellular effects of activation of CB₁ receptors in neurons are the inhibition of agonist-induced cyclic AMP (cAMP) formation[41] and blockade of inward Ca^{2+} currents mediated by N- and P/Q-type Ca^{2+} channels[42]. These characteristics are shared by a number of other G-protein-linked receptors (mGlu, muscarinic, and opioid) that modulate synaptic transmission. Moreover, these Ca²⁺ channels are chiefly located presynaptically and are required for evoked neurotransmitter release. Other electrophysiological effects of cannabinoids include activation of A-type, inwardly rectifying [43], and inhibition of M-type [44] K^+ channels. Some of these effects on K^+ channels may be secondary to the formation of cAMP[41,43]. The actions of cannabinoids on ion channels are usually blocked by pretreatment of cells with pertussis toxin, suggesting an involvement of $G_{i/0}$; however, in cultured striatal neurons, pertussis toxin treatment unmasks a cannabinoid receptor-mediated stimulatory effect on cAMP accumulation, suggesting the activation of a novel signaling pathway involving a G_s -type Gprotein[45]. Intracellular signaling pathways activated by cannabinoids in the CNS include the MAP kinase[46] and possibly c-Jun N-terminal kinase via a phosphoinositide 3'-kinasedependent process[47]. Additionally, Netzeband et al.[48] have shown that CB₁ receptors can interact with phospholipase C in cerebellar granule cells, leading to the mobilization of intracellular Ca²⁺.

The CB_1 like many other GPCRs, undergoes agonist-induced receptor, desensitization [49,50,51,52]. This process involves G-protein uncoupling in response to phosphorylation by G-protein-coupled receptor kinases (GRKs) and receptor internalization[51]. GRK-mediated receptor phosphorylation promotes the binding of beta-arrestins, which uncouple receptors from heterotrimeric G-proteins and target CB1 receptors for internalization in clathrincoated vesicles [50,51]. Studies of CB₁ receptor internalization have used transfected Chinese hamster ovary or AtT20 cells[49,50] and native receptors expressed on cultured hippocampal neurons or F11 cells[52]. Interestingly, the time course of internalization appears to be dependent on the cell type studied. The rate of internalization described for transfected cells or cell lines expressing CB_1 receptors is on the order of 10 to 30 min to achieve maximal levels, whereas it increases to between 5 and 16 h in hippocampal neurons maintained in primary culture[52]. This may reflect a difference in the internalization machinery between the cell types or between different subcellular compartments (axons vs. soma, for example).

Cell surface CB_1 receptor immunolabeling in hippocampal neurons is expressed at high levels at GABAergic synaptic terminals with no detectable labeling of somata[40]; however, GABAergic somata do express CB_1 receptor intracellular labeling in fixed and permeabilized cells, indicating that their site of synthesis is distant from their site of insertion into the plasma membrane. Likewise, in sensory neurons, cannabinoid CB_1 receptors are synthesized in somata and undergo axonal flow towards terminals, where they are inserted[53]. Following agonist exposure this situation is reversed with hippocampal CB_1 receptors being translocated back along axonal fibers towards the soma and proximal dendrites[52].

CELLULAR PHYSIOLOGY

Modulation of Synaptic Transmission

Cannabinoid actions can be classified into at least two types: (1) those mediated directly through cannabinoid receptors and (2) those mediated indirectly through other systems. Activation of cannabinoid CB₁ receptors modulates the release of a variety of neurotransmitters in the CNS (for a recent, detailed review, see[54]), including that of the principal excitatory and inhibitory neurotransmitters, glutamate and GABA, respectively, and also slower acting transmitters/neuromodulators such as the opioids[55,56], acetylcholine[57], dopamine[58], noradrenaline[59], and cholecystokinin[60]. The cellular mechanisms underlying these effects are thought to primarily reflect the actions of cannabinoids on presynaptic Ca²⁺ or K⁺ channels.

Cannabinoids modulate GABA release in a variety of CNS regions, including the hippocampus, basal ganglia, cerebellum, and brainstem. Studies have also shown inhibitory effects of Δ^9 -THC on GABA uptake in the hippocampus and basal ganglia[61,62]; however, electrophysiological investigations of GABAergic synaptic transmission have not confirmed this action. Anatomical invetigations have shown that high levels of CB₁ receptor immunoreactivity and mRNA are associated with GABAergic neurons[31,38,63]. In the hippocampus, neurochemical[38] and electrophysiological studies[37,64,65] have demonstrated that cannabinoids modulate GABA release from a subset of inhibitory neurons. In the basal ganglia (nucleus accumbens and striatum)[66,67,68,69,70], brainstem (rostral ventromedial medulla and periaqueductal gray)[71,72], and cerebellum[73,74], cannabinoids inhibit GABA release by a presynaptic mechanism. The majority of findings suggests that the actions of cannabinoids on GABA release are mediated via inhibition of presynaptic Ca²⁺ channels (N or N/PQ)[54,65], although in some cases there may be a direct action on the release process itself[40,54,72].

Glutamatergic synaptic transmission is also modulated by cannabinoids in a number of key pathways. CB₁ receptor labeling has been detected on excitatory terminals in cultured cerebellar granule cells[75]. There is also good electrophysiological evidence that cannabinoid receptor activation inhibits glutamate release at the Purkinje cell–parallel fiber synapse in cerebellar slices[73,76,77,78]. In the striatum, glutamate release from cortical afferents is inhibited by CB₁ receptor activation[79]. Furthermore, in the midbrain periaqueductal gray, glutamatergic synaptic transmission is inhibited by cannabinoids[71]. In the nucleus accumbens, cannabinoids may modulate glutamate release from glutamatergic cortical afferents via a mechanism involving modulation of presynaptic K⁺ channels[80] (but see[69]). Controversy exists with regard to excitatory synaptic transmission in the hippocampus with some studies showing an inhibition by cannabinoids[81,82] but others suggesting no effect[62,83]. These differences may reflect a developmental change in the sensitivity of glutamatergic synaptic transmission to cannabinoids[84] and/or species-dependent actions; thus, cannabinoids are

without effect on glutamatergic synaptic transmission in adult rat hippocampal slices[83], but inhibit this activity in adult mouse hippocampus[82], neonatal tissue from rat[84] or mouse[82], and in cultured rat hippocampal neurons[81]. There is also no consensus regarding the expression of CB₁ immunoreactivity on pyramidal neurons[31,32]. Of interest is the recent suggestion that in the brain there is a new type of cannabinoid receptor that modulates glutamatergic synaptic transmission[14]. It remains to be demonstrated whether this can account for the observed discrepancies between CB₁ receptor expression and function.

There is a growing body of evidence suggesting that cannabinoids can modulate postsynaptic responses to glutamate and glutamatergic agonists. In the nucleus accumbens medium spiny neurons, evoked glutamatergic excitatory postsynaptic currents are inhibited by a postsynaptic mechanism[69]. Activation of AMPA receptors is modulated by cannabinoids in a *Xenopus* oocyte expression system[85]. Studies by Hampson et al.[18] have shown that the endogenous cannabinoid ligand anandamide has a dual action on NMDA receptor–mediated responses in a variety of CNS regions. It directly potentiates electrophysiological responses to NMDA, but inhibits NMDA receptor–mediated Ca^{2+} elevations through a mechanism involving inhibition of voltage-gated Ca^{2+} channels. Recent work in cultured cerebellar granule suggests that cannabinoids can enhance NMDA receptor–mediated Ca^{2+} elevations via an indirect mechanism[48].

Synaptic Plasticity

Cannabinoids inhibit various forms of synaptic plasticity in the hippocampus, including long-term potentiation (LTP)[82,86,87] and long-term depression (LTD)[82] via a CB₁ receptor-dependent mechanism. Within the cortex[88] and striatum[89], synaptic plasticity is also compromised by cannabinoids. The mechanism underlying these effects remains controversial. Misner and Sullivan[82] have suggested that cannabinoid modulation of hippocampal synaptic plasticity is mediated by the inhibition of glutamate release and that this can be overcome under conditions where the synaptic activation of NMDA receptors is enhanced (Mg^{2+} -free medium or postsynaptic membrane depolarization during the LTP- or LTD-induction protocol); however, Paton et al.[83] suggest that in some cases, LTP is blocked where glutamatergic synaptic transmission is unaffected. The relationship between synaptic plasticity and the effects of cannabinoids on inhibitory synaptic transmission are likely to be subtle as they influence only a subpopulation of interneurons. This might explain why cannabinoids do not in fact enhance synaptic plasticity, as would be expected with a more general reduction in inhibitory drive. The actions of cannabinoids on inhibitory synaptic transmission could be important in controlling the power of network oscillations, which are thought to be critical for learning and memory[37]. Studies with CB_1 receptor knockout mice suggest that endocannabinoids exert a tonic inhibitory effect on synaptic plasticity and that anandamide could be an endogenous messenger involved in the modulation of memory processes[90,91].

ENDOCANNABINOIDS

The discovery of endogenous cannabinoid receptor ligands — anandamide (Narachidonylethanolamine) and 2-arachidonylglycerol (2-AG) and its ether form, noladin ether in the brain suggested the existence of a central endocannabinoid neuromodulatory system. Together with cannabinoid receptors, this cannabimimetic system is thought to have a widespread role in fine-tuning a variety of brain functions, including nociception, control of movement, memory, and neuroendocrine regulation. Recently it has also been suggested that this system is involved in brain development[92]. It is noteworthy that the highest levels of anandamide are expressed in the hippocampus[93]. Anandamide is released from neurons upon depolarization through a mechanism that requires Ca^{2+} -dependent cleavage from a phospholipid precursor in neuronal membranes[94]. The release of anandamide is followed by rapid uptake via a transporter into the cytosol and hydrolysis by the enzyme that inactivates endocannabinoids, fatty acid amide hydrolase (FAAH); indeed, the presence of CB₁ receptors/anandamide/FAAH in the thalamus, hippocampus, and cortex, or in the striatum, substantia nigra, and cerebellum, respectively, supports a role for the endogenous cannabinoid system in cognitive and motor responses. Studies with CB₁ receptor and FAAH knockout mice have also highlighted the importance of the endocannabinoid system in the control of normal CNS function, including retrograde synaptic inhibition in the hippocampus, long-term potentiation and memory, the development of opiate dependence, the control of appetite and pain perception[95,96].

Detailed neuroanatomical and electrophysiological analyses of mammalian nervous systems suggests that the CB_1 receptors are targeted to the presynaptic terminals of neurons; however, FAAH is preferentially located within the somatodendritic compartment of neurons that are postsynaptic to CB_1 -expressing axon terminals. This localization is consistent with a model of cannabinoid signaling in which anandamide is synthesized by postsynaptic cells and acts as a retrograde messenger molecule to modulate neurotransmitter release from presynaptic terminals[97] (Fig. 2). However, some of the actions of cannabinoids on synaptic transmission may also be postsynaptic[18,98]. Recent experimental work suggests that endogenous cannabinoids mediate signals retrogradely from depolarized postsynaptic neurons to presynaptic terminals to suppress subsequent neurotransmitter release, driving the synapse into an altered state. In hippocampal neurons[94], depolarization of postsynaptic neurons and resultant elevation of Ca^{2+} lead to a transient depolarization-induced suppression of inhibition (DSI). In cerebellar Purkinje cells, depolarization causes a transient suppression of both excitatory[78] and inhibitory[74] transmitter release from presynaptic terminals. The former is termed depolarization-induced suppression of excitation (DSE). Both DSI and DSE appear to share the same properties and may be a general and important mechanism by which the postsynaptic neuronal activity can influence the amount of transmitter release. Since it has been suggested that endocannabinoids released from a cell act over a short distance ($\sim 20 \text{ um}$)[94], only cells in close proximity will be influenced.

BEHAVIORAL EFFECTS AND THERAPEUTIC POTENTIAL

Attention and Perception

Anecdotal and experimental evidence in humans suggests that marijuana intoxication produces only minor distortions in sensory awareness, including some reports of heightened sensory perception[3]. In monkeys, acute or chronic exposure to marijuana smoke had no serious deleterious effects on simple visual discrimination tasks[99]; however, there are reports of significant effects of cannabinoids on attention processes in both humans and animals. Δ^9 -THC produced dose-dependent effects on both the accuracy and latency of the responses to differential tone discrimination[100,101] and on signal detection performance in rats[102]. The performance of monkeys, trained to respond in a choice reaction time task, was significantly disrupted by acute exposure to marijuana smoke[103]. In humans, marijuana intoxication produced detrimental effects on both attention span and divided-attention assignments[104]. These data suggest that cannabinoid receptor activation does not appear to affect the performance of tasks that do not require focused attention or persistent detailed perception. On the other hand, these same

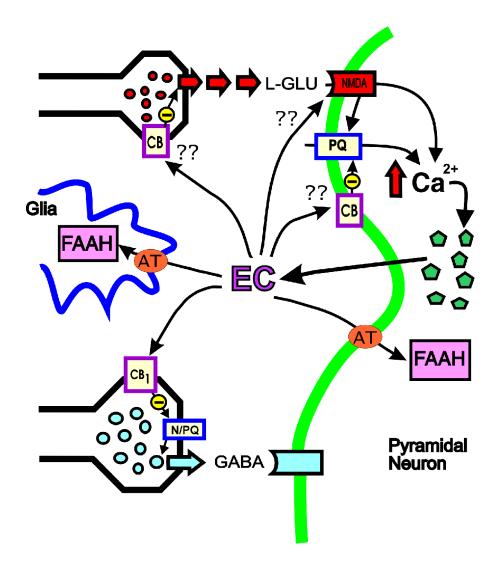


FIGURE 2. A model for the actions of endocannabinoids in the hippocampus. Strong excitation of pyramidal neurons via the release of L-glutamate (L-Glu) is very effective at increasing intracellular Ca^{2+} levels, involving both depolarization (N/PQ voltage-gated Ca^{2+} channels) and activation of NMDA receptors. It is suggested that this stimulates the release of endocannabinoids (EC), which act in a retrograde manner on both inhibitory (via CB₁ receptors) and excitatory terminals (possibly involving a novel cannabinoid receptor, CB). The postsynaptic activation of voltage-gated Ca^{2+} channels (PQ) and NMDA receptors may also be modulated. Endocannabinoids are removed by a transporter (anandamide transporter, AT), possibly into both neurons and glia, where they are metabolized by fatty acid amide hydrolase (FAAH). The net outcome will be a balance between these excitatory and inhibitory actions, with effects on excitatory transmission necessary to prevent the system from becoming unstable.

discriminatory processes may become susceptible to the influence of cannabinoid agonists when more sustained or divided attention is necessary. In general, in animal models the outcome of cannabinoid receptor activation on attention or perception tasks is thought to resemble that of hippocampal lesions.

Memory and Cognition

The main effects of cannabinoids on cognition in humans relate to the disruption of short-term memory[104,105], particularly with short-term amnesia and distortion of recently acquired information. These effects can be quite severe and dissociated from other effects of cannabinoids on cognition[106]. Experiments in animals also demonstrate cannabinoid-mediated memory

deficits, and these are related to impairment of the function of the hippocampus, a structure that is intimately involved in the processes that underlie learning and memory[107]. Studies have shown that activation of cannabinoid receptors produces memory deficits similar to those produced by neurochemical lesions of the hippocampus[108]. Such lesions impair performance in short-term spatial memory tasks learned prior to the lesion. In rats, Δ^9 -THC reduced exploratory parameters and motor activity and caused more errors in maze tests and problems with information retention. This was associated with a reduction in noradrenaline levels in the hippocampus and increased dopamine levels in the amygdala[109]; however, this situation is unclear since low doses of anandamide (1 µg/kg) were found to improve impaired maze performance and increase hippocampal dopamine and noradrenaline levels in food restricted mice[110]. In the zebra finch, a discrete neural network within the telencephalon controls song learning, and this area shows dense radioligand binding with the cannabinoid agonist $[^{3}H]$ -CP55940. The expression of CB₁ mRNA (5.5 kb) within the caudal telencephalon is altered during the course of vocal development and was shown by *in situ* hybridization to be expressed at high levels in areas involved in song learning and production, suggesting a potential role for cannabinoid signaling in zebra finch vocal development[111].

In conclusion, although there is little evidence of long-term memory disturbance, there is no definitive evidence that this, too, may not be affected. Some data suggest that there is no lasting or residual influence of cannabinoid exposure on memory processes and that the effects on short-term memory processes do not extend beyond the time that cannabinoids are present in the system[103,112]; nonetheless, cannabinoid neurotoxicity affecting the hippocampus may result in a more permanent impairment of cognitive performance[7].

Locomotor Actions

Central cannabinoid receptors are densely located in the output nuclei of the basal ganglia (globus pallidus and substantia nigra pars reticulata), indicating a possible role in the regulation of extrapyramidal motor activity[113,114,115]. Furthermore, in the striatum cannabinoids modulate synaptic transmission and plasticity[79,89]. The cerebellum is also involved in the fine tuning of movement, and in this region cannabinoids inhibit excitatory and inhibitory synaptic transmission onto Purkinje neurons[73,74,76,77,78]; thus, locomotor-related activity is likely to be compromised by cannabinoids at a number of different levels in the CNS. It is suggested that the endogenous cannabinoid tone participates in the control of movement and, therefore, the central endocannabinoid system might play a role in the pathophysiology of movement disorders[116,117]; thus, cannabinoids might be useful in the treatment of tics in Tourette's syndrome, in the reduction of levodopa-induced dyskinesia in Parkinson's disease, or spasm associated with multiple sclerosis, and in the treatment of chorea in Huntington's disease. The extent to which the beneficial actions of cannabinoids in motor disorders are due to psychoactive or analgesic effects, however, remains to be established.

Reward

Electrical or chemical stimulation of the medial forebrain bundle (MFB) can establish responses similar to those associated with natural rewards such as food or sexual contact, including intense self-stimulation in animals and euphoria in humans[118]. The reward/reinforcement circuitry of the mammalian brain consists of synaptically interconnected neurons associated with the MFB, linking the ventral tegmental area, nucleus accumbens, and ventral pallidum. Cannabinoids have been shown to increase the activity of dopaminergic neurons in the ventral tegmental area–mesolimbic pathway[118,119], circuits that are known to play a pivotal role in the neural substrates of drug addiction, withdrawal, and craving; however, these actions appear to be

indirect as they do not to directly affect dopamine release itself[54]. In the nucleus accumbens, medium spiny GABAergic projection neurons receive synaptic input from both intrinsic GABAergic interneurons and extrinsic glutamatergic sources[69]. Both GABAergic and glutamatergic synaptic transmissions onto medium spiny neurons are inhibited by cannabinoids as well as other drugs of abuse[69,70], suggesting that this action may contribute to their rewarding properties.

Appetite

Both marijuana and anandamide stimulate food intake and recent work is shedding light on the mechanism that underlies this effect. Endocannabinoids are implicated in the processes that regulate appetite in the hypothalamus[120], where they interact with leptin, a hormone critically involved in the central control of food intake and energy homeostasis. Following temporary food restriction, CB_1 receptor knockout mice eat less than their wild-type littermates, whereas the CB_1 antagonist SR141716A reduces food intake in wild-type, but not CB₁ knockout, mice[120]. The role of central cannabinoid systems in appetite regulation may also involve activation of reward systems and be mediated in part by opioidergic processes; thus, naloxone reduces the facilitory effect of Δ^9 -THC on feeding evoked by electrical stimulation of the lateral hypothalamus[121], and synergistic interactions between the effects of naloxone and SR141716A on nocturnal food (chow) intake tests have been documented [122]. From a therapeutic perspective, synthetic Δ^9 -THC (Marinol) has been used to stimulate the appetite of AIDS patients[4]. Cannabis may also be useful in the treatment of anorexia, but this action remains to be demonstrated in clinical trials. Clinical studies suggest SR141716A might be an effective antiobesity agent and has recently entered phase III clinical trials for this purpose (for more information see www.sanofisynthelabous.com/excellence/).

Pain

Analgesic effects of cannabis and cannabinoids have been demonstrated in most animal models of pain. These antinociceptive effects involve actions at a number of different levels, including peripheral sensory neurons, spinal cord, and central pathways[6,123]. In the brain and spinal cord a cannabinoid interaction with the opioidergic system may act to modulate the perception of painful stimuli[6]. Cannabinoids ameliorate pain by modulating rostral ventromedial medulla (RVM) neuronal activity in the brainstem in a manner similar to, but pharmacologically dissociable from, that of morphine[124]. The nucleus reticularis gigantocellularis pars alpha within the RVM is a major source of this descending modulation, and this system is activated in response to noxious stimulation[54,71]. Cannabinoids also inhibit synaptic transmission in the midbrain periaqueductal gray[72]. This area forms part of a descending antinociceptive pathway that, via the RVM, modulates nociceptive transmission at the level of the spinal cord[125]; thus, by acting at many different levels, cannabinoids may provide a useful alternative therapy for the treatment of pain, in particular in inflammatory and neuropathic pain[123], which do not respond well to conventional therapies.

Neurotoxicity

An exciting new area of cannabinoid research is the potential of these compounds to modulate neurotoxicity. In the CNS, cannabinoids may protect neurons from toxic insults such as glutamatergic overstimulation, ischemia, and oxidative damage. Cannabinoid receptor activation is neuroprotective in several experimental models of cerebral hypoxia/ischemia and seizure

activity[126,127]. A neuroprotective role for N-acylethanolamines, including anandamide, has also been suggested on the basis of their production at sites of neuronal damage or death[128]. In mice, the levels of endogenous 2-AG are also significantly elevated after closed head injury[129]; however, in neonatal rats, levels of 2-AG and other 2-monoacylglycerols are virtually unaffected following various models of *in vivo* neurodegeneration [128]. As a note of caution, relatively short periods of exposure to some cannabinoids, including Δ^9 -THC, are neurotoxic in cultured neurons, with shrinkage of neurons and DNA fragmentation in hippocampal tissue, an effect which is CB₁ receptor-mediated [7,130]. Chronic pretreatment with the cannabinoid agonist WIN 55212-2 can induce observable subcellular morphological changes, particularly in dendritic arrangements in the rat hippocampus[131]. There are also a number of unresolved issues that further complicate the role of cannabinoids in neuroprotection. First, the action of cannabinoids on glutamate release is controversial (see above). Second, some synthetic, nonpsychotropic cannabinoids such as HU-211 have NMDA receptor antagonist properties[23]. Third, cannabinoids have a multitude of seemingly contradictory actions; for example, activation of CB₁ receptors inhibits NMDAinduced intracellular Ca^{2+} rises in the cortex but enhances similar responses in the cerebellum[18,48]. Furthermore, anandamide enhances NMDA-receptor-mediated synaptic transmission in the hippocampus via a direct interaction with the NMDA receptor *per se*[18], and certain cannabinoids may possess intrinsic antioxidant activity, thereby limiting neuronal damage via a receptor-independent pathway[26]. Fourth, interactions between adenosine, a widely studied neuroprotective agent, and cannabinoids have also been described[132].

Epilepsy

Some data support the possible use of cannabis or cannabinoids in the treatment of epilepsy, but they can exert both convulsant and anticonvulsant effects[133,134]. The hippocampus is an area of the CNS particularly prone to the generation of seizures and is the subject of much study in this area. In the hippocampal slice preparation, anandamide, 2-AG, and WIN 55,212-2 attenuate epileptiform activity induced by exposure to Mg^{2+} -free medium or Mg^{2+} -free medium with elevated K⁺[135,136]. Low Mg^{2+} -induced aberrant synaptic activity in cultured hippocampal neurons is also blocked by cannabinoids[81]. These actions of cannabinoids are sensitive to SR141716A, suggesting an involvement of CB₁ receptors. Δ^9 -THC can exert both convulsant and anticonvulsant actions in a model of cortical focal epilepsy[134] and can lower the seizure threshold following withdrawal[137]. A CB₁ receptor-mediated reduction in excitatory drive in the hippocampus could provide a mechanism for the prevention of excessive excitability leading to seizures; however, an associated inhibition of GABAergic synaptic transmission in the hippocampus would oppose this effect and may underlie some of the proconvulsant actions observed with Δ^9 -THC.

The nonpsychotic cannabinoids cannabidiol and HU-211 also exhibit anticonvulsant activity [138,139]; however, these actions may be indirect via effects on anandamide metabolism and NMDA receptor function, respectively [23,139]. At least some of the anticonvulsant actions of cannabinoids thus may be independent of CB_1 receptors. In conclusion, the therapeutic potential of cannabis and cannabinoids as anticonvulsants requires clarification.

CONCLUSION

The role of cannabinoid systems in normal brain function and disease states is becoming clearer, and much detail has now been uncovered regarding their mechanisms of action. Considerable advances have been made in our understanding of cannabinoid receptor signaling pathways, their modulation of synaptic transmission and plasticity, the cellular targets of cannabinoids in different CNS regions and, in particular, the role of the endocannabinoid system; however, there are critical areas of controversy, including those related to the actions of cannabinoids in neurotoxicity and epilepsy. The presence of a new type of brain cannabinoid receptor is indicated and, if realized, will lead to both a further complexity in the actions of cannabinoids and to an exciting new therapeutic target. If all of the behavioral effects attributed to cannabinoids are mediated by CB_1 receptors, it will be more difficult to design clinical compounds that lack the undesirable psychotropic/cognitive/motor side effects. Alternative strategies using drugs that interfere with the synthesis, degradation, or uptake of endocannabinoids could prove to be useful therapeutically and would also help in our understanding of the brain endocannabinoid system. Given the current likelihood that cannabis will be legalized for medicinal use in the U.K. and possibly other countries, it is vital that we continue to unravel how its active constituents affect the CNS, both acutely and with sustained use.

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REFERENCES

- 1. Gaoni, Y. and Mechoulam, R. (1964) Isolation, structure, and partial synthesis of an active constituent of hashish. J. Am. Chem. Soc. **86**, 1646–1647.
- 2. Chaperon, F. and Thiebot, M.H. (1999) Behavioral effects of cannabinoid agents in animals. *Crit. Rev. Neurobiol.* **13**, 243–281.
- 3. Hollister, L.E. (1986) Health aspects of cannabis. *Pharmacol. Rev.* **38**, 1–20.
- 4. Mechoulam, R. (1999) Recent advances in cannabinoid research. Forsch. Komplementarmed. 6, 16–20.
- 5. Ameri, A. (1999) The effects of cannabinoids on the brain. *Prog. Neurobiol.* 58, 315–348.
- 6. Pertwee, R.G. (2001) Cannabinoid receptors and pain. *Prog. Neurobiol.* 63, 569–611.
- Chan, G.C., Hinds, T.R., Impey, S., and Storm, D.R. (1998) Hippocampal neurotoxicity of Δ⁹tetrahydrocannabinol. *J. Neurosci.* 18, 5322–5332.
- 8. Pertwee, R.G. (1997) Pharmacology of cannabinoid CB₁ and CB₂ receptors. *Pharmacol. Ther.* **74**, 129–180.
- 9. Shire, D., Carillon, C., Kaghad, M., Calandra, B., Rinaldi-Carmona, M., Le Fur, G., Caput, D., and Ferrara, P. (1995) An amino-terminal variant of the central cannabinoid receptor resulting from alternative splicing. *J. Biol. Chem.* **270**, 3726–3731.
- Rinaldi-Carmona, M., Calandra, B., Shire, D., Bouaboula, M., Oustric, D., Barth, F., Casellas, P., Ferrara. P., and Le Fur, G. (1996) Characterization of two cloned human CB₁ cannabinoid receptor isoforms. *J. Pharmacol. Exp. Ther.* 278, 871–878.
- Griffin, G., Wray, E.J., Tao, Q., McAllister, S.D., Rorrer, W.K., Aung, M.M., Martin, B.R., and Abood, M.E. (1999) Evaluation of the cannabinoid CB₂ receptor-selective antagonist, SR144528: further evidence for cannabinoid CB₂ receptor absence in the rat central nervous system. *Eur. J. Pharmacol.* 377, 117–125.
- Skaper, S.D., Buriani, A., Dal Toso, R., Petrelli, L., Romanello, S., Facci, L., and Leon, A. (1996) The ALIAmide palmitoylethanolamide and cannabinoids, but not anandamide, are protective in a delayed postglutamate paradigm of excitotoxic death in cerebellar granule neurons. *Proc. Natl. Acad. Sci. U. S. A.* 93, 3984–3989.
- 13. Howlett, A.C. (1995) Pharmacology of cannabinoid receptors. Annu. Rev. Pharmacol. 33, 607–634.
- 14. Hájos, N., Ledent, C., and Freund, T.F. (2001) Novel cannabinoid-sensitive receptor mediates inhibition of glutamatergic synaptic transmission in the hippocampus. *Neuroscience* **106**, 1–4.
- 15. Breivogel, C.S., Griffin, G., Di, M.V., and Martin, B.R. (2001) Evidence for a new G protein-coupled cannabinoid receptor in mouse brain. *Mol. Pharmacol.* **60**, 155–163.
- Zygmunt, P.M., Petersson, J., Andersson, D.A., Chuang, H., Sorgard, M., Di Marzo, V., Julius, D., and Hogestatt, E.D. (1999) Vanilloid receptors on sensory nerves mediate the vasodilator action of anandamide. *Nature* 400, 452–457.
- 17. Maingret, F., Patel, A.J., Lazdunski, M., and Honore, E. (2001) The endocannabinoid anandamide is a direct and selective blocker of the background K⁺ channel TASK-1. *EMBO J.* **20**, 47–54.
- Hampson, A.J., Bornheim, L.M., Scanziani, M., Yost, C.S., Gray, A.T., Hansen, B.M., Leonoudakis, D.J., and Bickler, P.E. (1998) Dual effects of anandamide on NMDA receptor-mediated responses and neurotransmission. *J. Neurochem.* **70**, 671–676.

- Bisogno, T., Hanus, L., De Petrocellis, L., Tchilibon, S., Ponde, D.E., Brandi, I., Moriello, A.S., Davis, J.B., Mechoulam, R., and Di Marzo, V. (2001) Molecular targets for cannabidiol and its synthetic analogues: effect on vanilloid VR₁ receptors and on the cellular uptake and enzymatic hydrolysis of anandamide. *Br. J. Pharmacol.* 134, 845–852.
- 20. Fan, P. (1995) Cannabinoid agonists inhibit the activation of 5-HT3 receptors in rat nodose ganglion neurons. *J. Neurophysiol.* **73**, 907–910.
- 21. Kobayashi, H., Suzuki, T., Kamata, R., Saito, S., Sato, I., Tsuda, S., and Matsusaka, N. (1999) Recent progress in the neurotoxicology of natural drugs associated with dependence or addiction, their endogenous agonists and receptors. *J. Toxicol. Sci.* **24**, 1–16.
- 22. Lagalwar, S., Bordayo, E.Z., Hoffmann, K.L, Fawcett, J.R., and Frey, W.H., II (1999) Anandamides inhibit binding to the muscarinic acetylcholine receptor. *J. Mol. Neurosci.* **13**, 55–61.
- 23. Feigenbaum, J.J., Bergmann, F., Richmond, S.A., Mechoulam, R., Nadler, V., Kloog, Y., and Sokolovsky, M. (1989) Nonpsychotropic cannabinoid acts as a functional N-methyl-D-aspartate receptor blocker. *Proc. Natl. Acad. Sci. U. S. A.* **86**, 9584–9587.
- 24. Nagayama, T., Sinor, A.D., Simon, R.P., Chen, J., Graham, S.H., Jin, K., and Greenberg, D.A. (1999) Cannabinoids and neuroprotection in global and focal cerebral ischemia and in neuronal cultures. *J. Neurosci.* **19**, 2987–2995.
- 25. Howlett, A.C. and Mukhopadhyay, S. (2000) Cellular signal transduction by anandamide and 2arachidonoylglycerol. *Chem. Phys. Lipids* **108**, 53–70.
- 26. Hampson, A.J., Grimaldi, M., Axelrod, J., and Wink, D. (1998) Cannabidiol and $(-)-\Delta^9$ -tetrahydrocannabinol are neuroprotective antioxidants. *Proc. Nat. Acad. Sci. U. S. A.* **95**, 8268–8273.
- Pertwee, R.G. (1990) The central neuropharmacology of psychotropic cannabinoids. In *Psychotropic Drugs of Abuse*. Balfour, D.J.K., Ed. Pergamon Press, New York. pp. 355–429.
- Greenamyre, J.T., Young, A.B., and Penney, J.B. (1984) Quantitative autoradiographic distribution of L-[3H]glutamate-binding sites in rat central nervous system. J. Neurosci. 4, 2133–2144.
- 29. Bowery, N.G., Hudson, A.L., and Price, G.W. (1987) GABA_A and GABA_B receptor site distribution in the rat central nervous system. *Neuroscience* **20**, 365–83.
- 30. Herkenham, M. (1995) Localization of cannabinoid receptors in brain and periphery. In *Cannabinoid Receptors (Neuroscience Perspectives)*. Pertwee, R.G., Ed. Academic Press, London. pp. 145–166.
- 31. Tsou, K., Brown, S., Sanudo-Pena, M.C., Mackie, K., and Walker, J.M. (1998) Immunohistochemical distribution of cannabinoid CB₁ receptors in the rat central nervous system. *Neuroscience* **83**, 393–411.
- 32. Pettit, D.A., Harrison, M.P., Olson, J.M., Spencer, R.F., and Cabral, G.A. (1998) Immunohistochemical localization of the neural cannabinoid receptor in rat brain. *J. Neurosci. Res.* **51**, 391–402.
- 33. Egertova, M. and Elphick, M.R. (2000) Localisation of cannabinoid receptors in the rat brain using antibodies to the intracellular C-terminal tail of CB₁. J. Comp. Neurol. **422**, 159–171.
- 34. Moldrich, G. and Wenger, T. (2000) Localization of the CB₁ cannabinoid receptor in the rat brain. An immunohistochemical study. *Peptides* **21**, 1735–1742.
- 35. Tsou, K., Mackie, K., Sanudo-Pena, M.C., and Walker, J.M. (1999) Cannabinoid CB₁ receptors are primarily localised on cholecystokinin-containing GABAergic interneurons in the rat hippocampal formation. *Neuroscience* **93**, 969–975.
- Katona, I., Sperlagh, B., Magloczky, Z., Santha, E., Kofalvi, A., Czirjak, S., Mackie, K., Vizi, E.S., and Freund, T.F. (2000) GABAergic interneurons are the targets of cannabinoid actions in the human hippocampus. *Neuroscience* 100, 797–804.
- Hájos, N., Katona, I., Naiem, S.S., Mackie, K., Ledent, C., Mody, I., and Freund, T. (2000) Cannabinoids inhibit hippocampal GABAergic transmission and network oscillations. *Eur. J. Neurosci.* 12, 3239–3249.
- Katona, I., Sperlagh, B., Sik, A., Kafalvi, A., Vizi, E.S., Mackie, K., and Freund, T.F. (1999) Presynaptically located CB₁ cannabinoid receptors regulate GABA release from axon terminals of specific hippocampal interneurons. *J. Neurosci.* 19, 4544–4558.
- 39. Marsicano, G. and Lutz, B. (1999) Expression of the cannabinoid receptor CB₁ in distinct neuronal subpopulations in the adult mouse brain. *Eur. J. Neurosci.* **11**, 4213–4225.
- Irving, A.J., Coutts, A.A., Harvey, J., Rae, M.G., Mackie, K., Bewick, G.S., and Pertwee, R.G. (2000) Functional expression of cell surface cannabinoid CB₁ receptors on presynaptic inhibitory terminals in cultured rat hippocampal neurons. *Neuroscience* 98, 253–262.
- 41. Childers, S.R. and Deadwyler, S.A. (1996) Role of cyclic AMP in the actions of cannabinoid receptors. *Biochem. Pharmacol.* **52**, 819–827.
- 42. Twitchell, W., Brown, S., and Mackie, K. (1997) Cannabinoids inhibit N- and P/Q-type Ca²⁺ channels in cultured rat hippocampal neurons. *J. Neurophysiol.* **78**, 43–50.
- 43. Mu, J., Zhuang, S.Y., Kirby, M.T., Hampson, R.E., and Deadwyler, S.A. (1999) Cannabinoid receptors differentially modulate potassium A and D currents in hippocampal neurons in culture. *J. Pharmacol. Exp. Ther.* **291**, 893–902.
- 44. Schweitzer, P. (2000) Cannabinoids decrease the K⁺ M-current in hippocampal CA1 neurons. J. Neurosci. **20**, 51–58.

- 45. Glass, M. and Felder, C.C. (1997) Concurrent stimulation of cannabinoid CB₁ and dopamine D₂ receptors augments cAMP accumulation in striatal neurons: evidence for a Gs linkage to the CB₁ receptor. *J. Neurosci.* **17**, 5327–5333.
- 46. Bouaboula, M., Poinot-Chazel, C., Bourrie, B., Canat, X., Calandra, B., Rinaldi-Carmona, M., Le Fur, G., and Casellas, P. (1995) Activation of mitogen-activated protein kinases by stimulation of the central cannabinoid receptor CB₁. *Biochem. J.* **312**, 637–641.
- 47. Rueda, D., Galve-Roperh, I., Haro, A., and Guzman, M. (2000) The CB₁ cannabinoid receptor is coupled to the activation of c-Jun N-terminal kinase. *Mol. Pharmacol.* **58**, 814–820.
- 48. Netzeband, J.G., Conroy, S.M., Parsons, K.L., and Gruol, D.L. (1999) Cannabinoids enhance NMDAelicited Ca²⁺ signals in cerebellar granule neurons in culture. *J. Neurosci.* **19**, 8765–8777.
- 49. Rinaldi-Carmona, M., Le Duigou, A., Oustric, D., Barth, F., Bouaboula, M., Carayon, P., Casellas, P., and Le Fur, G. (1998) Modulation of CB₁ cannabinoid receptor functions after a long-term exposure to agonist or inverse agonist in the Chinese hamster ovary cell expression system. *J. Pharmacol. Exp. Ther.* 287, 1038–1047.
- 50. Hsieh, C., Brown, S., Derleth, C., and Mackie, K. (1999) Internalization and recycling of the CB₁ cannabinoid receptor. *J. Neurochem.* **73**, 493–501.
- Jin, W., Brown, S., Roche, J.P., Hsieh, C., Celverm, J.P., Kovoor, A., Chavkinm, C., and Mackie, K. (1999) Distinct domains of the CB₁ cannabinoid receptor mediate desensitization and internalization. *J. Neurosci.* 19, 3773–3780.
- Coutts, A.A., Anavi-Goffer, S., Ross, R.A., MacEwan, D.J., Mackie, K., Pertwee, R.G., and Irving, A.J. (2001) Agonist-induced internalization and trafficking of cannabinoid CB₁ receptors in hippocampal neurons. *J. Neurosci.* 21, 2425–2433.
- 53. Hohmann, A.G. and Herkenham, M. (1999) Cannabinoid receptors undergo axonal flow in sensory nerves. *Neuroscience* **92**, 1171–1175.
- 54. Schlicker, E. and Kathmann, M. (2001) Modulation of transmitter release via presynaptic cannabinoid receptors. *Trends Pharmacol. Sci.* **22**, 565–572.
- 55. Houser, S.J., Eads, M., Embrey, J.P., and Welch, S.P. (2000) Dynorphin B and spinal analgesia: induction of antinociception by the cannabinoids CP55,940, Δ^9 -THC and anandamide. *Brain Res.* **857**, 337–342.
- 56. Valverde, O., Noble, F., Beslot, F., Dauge, V., Fournie-Zaluski, M.C., and Roques, B.P. (2001) Δ⁹-tetrahydrocannabinol releases and facilitates the effects of endogenous enkephalins: reduction in morphine withdrawal syndrome without change in rewarding effect. *Eur. J. Neurosci.* 13, 1816–1824.
- 57. Gifford, A.N. and Ashby, C.R., Jr. (1996) Electrically evoked acetylcholine release from hippocampal slices is inhibited by the cannabinoid agonist WIN 55212-2 and is potentiated by the cannabinoid agonist, SR141716A. *J. Pharmacol. Exp. Ther.* **277**, 1431–1436.
- 58. Cadogan, A.K., Alexander, S.P., Boyd, E.A., and Kendall, D.A. (1997) Influence of cannabinoids on electrically evoked dopamine release and cyclic AMP generation in the rat striatum. *J. Neurochem.* **69**, 1131–1137.
- 59. Schlicker, E., Timm, J., Zentner, J., and Gothert, M. (1997) Cannabinoid CB₁ receptor-mediated inhibition of noradrenaline release in the human and guinea-pig hippocampus. *Naunyn Schmiedebergs Arch Pharmacol.* **356**, 583–589.
- Beinfeld, M.C. and Connolly, K. (2001) Activation of CB₁ cannabinoid receptors in rat hippocampal slices inhibits potassium-evoked cholecystokinin release, a possible mechanism contributing to the spatial memory defects produced by cannabinoids. *Neurosci. Lett.* **301**, 69–71.
- 61. Maneuf, Y.P., Nash, J.E., Crossman, A.R., and Brotchie, J.M. (1996) Activation of the cannabinoid receptor by Δ^9 -tetrahydrocannabinol reduces gamma-aminobutyric acid uptake in the globus pallidus. *Eur. J. Pharmacol.* **308**, 161–164.
- 62. Coull, M.A., Johnston, A.T., Pertwee, R.G., and Davies, S.N. (1997) Action of Δ^9 -tetrahydrocannabinol on GABA_A receptor-mediated responses in a grease-gap recording preparation of the rat hippocampal slice. *Neuropharmacology* **36**, 1387–1392.
- 63. Herkenham, M., Lynn, A.B., Johnson, M.R., Melvin, L.S., de Costa, B.R., and Rice, K.C. (1991) Characterization and localization of cannabinoid receptors in rat brain: a quantitative *in vitro* autoradiographic study. *J. Neurosci*, **11**, 563–583.
- 64. Hoffman, A.F. and Lupica, C.R. (2000) Mechanisms of cannabinoid inhibition of GABA_A synaptic transmission in the hippocampus. *J. Neurosci.* **20**, 2470–2479.
- 65. Wilson, R.I., Kunos, G., and Nicoll, R.A. (2001) Presynaptic specificity of endocannabinoid signaling in the hippocampus. *Neuron* **31**, 453–462.
- 66. Miller, A.S. and Walker, J.M. (1995) Effects of a cannabinoid on spontaneous and evoked neuronal activity in the substantia nigra pars reticulata. *Eur. J. Pharmacol.* **279**, 179–185.
- 67. Chan, P.K.Y., Chan, S.C.Y., and Yung, W.H. (1998) Presynaptic inhibition of GABAergic inputs to rat substantia nigra pars reticulata neurons by a cannabinoid agonist. *Neuroreport* **9**, 671–675.

- 68. Szabo, B., Dörner, L., Pfreundtner, C., Nörenberg, W., and Starke, K. (1998) Inhibition of GABAergic inhibitory postsynaptic currents by cannabinoids in rat corpus striatum. *Neuroscience* **85**, 395–403.
- 69. Hoffman, A.F. and Lupica, C.R. (2001) Direct actions of cannabinoids on synaptic transmission in the nucleus accumbens: a comparison with opioids. *J. Neurophysiol.* **85**, 72–83.
- Manzoni, O.J. and Bockaert, J. (2001) Cannabinoids inhibit GABAergic synaptic transmission in mice nucleus accumbens. *Eur. J. Pharmacol.* 412, R3–R5.
- 71. Vaughan, C.W., McGregor, I.S., and Christie, M.J. (1999) Cannabinoid receptor activation inhibits GABAergic neurotransmission in rostral ventromedial medulla neurons *in vitro*. *Br. J. Pharmacol.* **127**, 935–940.
- 72. Vaughan, C.W., Connor, M., Bagley, E.E., and Christie, M.J. (2000) Actions of cannabinoids on membrane properties and synaptic transmission in rat periaqueductal gray neurons *in vitro*. *Mol. Pharmacol.* **57**, 288–295.
- 73. Takahashi, K.A. and Linden, D.J. (2000) Cannabinoid receptor modulation of synapses received by cerebellar Purkinje cells. *J. Neurophysiol.* **83**, 1167–1180.
- 74. Kreitzer, A.C. and Regehr, W.G. (2001) Cerebellar depolarization-induced suppression of inhibition is mediated by endogenous cannabinoids. *J. Neurosci.* **21**, RC174.
- 75. Caldwell, D., Coutts, A.A., Mackie K., and Irving, A.J. (1999) Cell surface CB₁ receptors are expressed at synaptic terminals in cultured rat cerebellar granule cells. *J. Physiol. (London)* **518**, 152P–153P.
- 76. Lévénés, C., Daniel, H., Soubrie, P., and Crepel, F. (1998) Cannabinoids decrease excitatory synaptic transmission and impair long-term depression in rat cerebellar Purkinje cells. *J. Physiol. (London)* **510**, 867–879.
- Maejima, T., Ohno-Shosaku, T., and Kano, M. (2001) Endogenous cannabinoid as a retrograde messenger from depolarized postsynaptic neurons to presynaptic terminals. *Neurosci. Res.* 40, 205–210.
- 78. Kreitzer, A.C. and Regehr, W.G. (2001) Retrograde inhibition of presynaptic Ca²⁺ influx by endogenous cannabinoids at excitatory synapses onto Purkinje cells. *Neuron* **29**, 717–727.
- 79. Huang, C.C., Lo, S.W., and Hsu, K.S. (2001) Presynaptic mechanisms underlying cannabinoid inhibition of excitatory synaptic transmission in rat striatal neurons. *J. Physiol (London)*. **532**, 731–748.
- 80. Robbe, D., Alonso, G., Duchamp, F., Bockaert, J., and Manzoni, O.J. (2001) Localization and mechanisms of action of cannabinoid receptors at the glutamatergic synapses of the mouse nucleus accumbens. *J. Neurosci.* **21**, 109–116.
- 81. Shen, M., Piser, T., Seybold, V.S., and Thayer, S.A. (1996) Cannabinoid receptor agonists inhibit glutamatergic synaptic transmission in rat hippocampal cultures. *J. Neurosci.* **16**, 4322–4334.
- Misner, D.L. and Sullivan, J.M. (1999) Mechanism of cannabinoid effects on long-term potentiation and depression in hippocampal CA1 neurons. J. Neurosci. 19, 6795–6805.
- Paton, G.S., Pertwee, R.G., and Davies S.N. (1998) Correlation between cannabinoid mediated effects on paired pulse depression and induction of long term potentiation in the rat hippocampal slice. *Neuropharmacology* 37, 1123–1130.
- 84. Al Hayani, A. and Davies, S.N. (2000) Cannabinoid receptor-mediated inhibition of excitatory synaptic transmission in the rat hippocampal slice is developmentally regulated. *Br. J. Pharmacol.* **131**, 663–665.
- 85. Akinshola, B.E., Taylor, R.E., Ogunseitan, A.B., and Onaivi, E.S. (1999) Anandamide inhibition of recombinant AMPA receptor subunits in *Xenopus* oocytes is increased by forskolin and 8-bromo-cyclic AMP. *Naunyn Schmiedebergs Arch. Pharmacol.* **360**, 242–248.
- 86. Collins, D.R., Pertwee, R.G., and Davies, S.N. (1994) The action of synthetic cannabinoids on the induction of long-term potentiation in the rat hippocampal slice. *Eur. J. Pharmacol.* **259**, R7–R8.
- Terranova, J.P., Michaud, J.C., Le Fur, G., and Soubrie, P. (1995) Inhibition of long-term potentiation in rat hippocampal slices by anandamide and WIN55212-2: reversal by SR141716A, a selective antagonist of CB₁ cannabinoid receptors. *Naunyn Schmiedebergs Arch. Pharmacol.* 352, 576–579.
- Auclair, N., Otani, S., Soubrie, P., and Crepel, F. (2000) Cannabinoids modulate synaptic strength and plasticity at glutamatergic synapses of rat prefrontal cortex pyramidal neurons. *J. Neurophysiol.* 83, 3287–3293.
- 89. Gerdeman, G. and Lovinger, D.M. (2001) CB₁ cannabinoid receptor inhibits synaptic release of glutamate in rat dorsolateral striatum. *J. Neurophysiol.* **85**, 468–471.
- 90. Bohme, G.A., Laville, M., Ledent, C., Parmentier, M., and Imperato, A. (2000) Enhanced long-term potentiation in mice lacking cannabinoid CB₁ receptors. *Neuroscience* **95**, 5–7.
- 91. Reibaud, M., Obinu, M.C., Ledent, C., Parmentier, M., Bohme, G.A., and Imperato, A. (1999) Enhancement of memory in cannabinoid CB₁ receptor knock-out mice. *Eur. J. Pharmacol.* **379**, R1–R2.
- 92. Fernandez-Ruiz, J., Berrendero, F., Hernandez, M.L., and Ramos, J.A. (2000) The endogenous cannabinoid system and brain development. *Trends Neurosci.* **23**, 14–20.
- 93. Felder, C.C., Nielsen, A., Briley, E.M., Palkovits, M., Priller, J., Axelrod, J., Nguyen, D.N., Richardson, J.M., Riggin, R.M., Koppel, G.A., Paul, S.M., and Becker, G.W. (1996) Isolation and measurement of the endogenous cannabinoid receptor agonist, anandamide, in brain and peripheral tissues of human and rat. *FEBS Lett.* **393**, 231–235.

- 94. Wilson, R.I. and Nicoll, R.A. (2001) Endogenous cannabinoids mediate retrograde signalling at hippocampal synapses. *Nature* **410**, 588–592.
- 95. Kunos, G. and Batkai, S. (2001) Novel physiologic functions of endocannabinoids as revealed through the use of mutant mice. *Neurochem. Res.* **26**, 1015–1021.
- 96. Cravatt, B.F., Demarest, K., Patricelli, M.P., Bracey, M.H., Giang, D.K., Martin, B.R., and Lichtman, A.H. (2001) Supersensitivity to anandamide and enhanced endogenous cannabinoid signaling in mice lacking fatty acid amide hydrolase. *Proc. Natl. Acad. Sci. U. S. A.* 98, 9371–9376.
- 97. Elphick, M.R. and Egertova, M. (2001) The neurobiology and evolution of cannabinoid signalling. *Philos. Trans. R. Soc. London B. Biol. Sci.* **356**, 381–408.
- 98. Ong, W.Y. and Mackie, K. (1999) A light and electron microscopic study of the CB₁ cannabinoid receptor in primate brain. *Neuroscience* **92**, 1177–1191.
- 99. Schultze, G.E., McMillan, D.E., Bailey, J.R., Scallet, A., Ali, S.F., Slikker, W., Jr., and Paule, M.G. (1988) Acute effects of Δ^9 -tetrahydrocannabinol in rhesus monkeys as measured by performance in a battery of complex operant tests. *J. Pharmacol. Exp. Ther.* **245**, 178–186.
- 100. Campbell, K.A., Foster, T.C., Hampson, R.E., and Deadwyler, S.A. (1986) Effects of Δ^9 -tetrahydrocannabinol on sensory-evoked discharges of granule cells in the dentate gyrus of behaving rats. *J. Pharmacol. Exp. Ther.* **239**, 941–945.
- 101. Campbell, K.A., Foster, T.C., Hampson, R.E., and Deadwyler, S.A. (1986) Δ^9 Tetrahydrocannabinol differentially affects sensory-evoked potentials in the rat dentate gyrus. J. Pharmacol. Exp. Ther. 239, 936–940.
- 102. Heyser, C.J., Hampson, R.E., and Deadwyler, S.A. (1989) Characteristics of hippocampal complex spike neurons during performance of a deleted match to sample task: selective influences of Δ^9 -THC. *Soc. Neurosci. Abstr.* **15**, 1170.
- 103. Paule, M.G., Allen, R.R., Bailey, J.R., Scallet, A.C., Ali, S.F., Brown, R.M., and Slikker, W., Jr. (1992) Chronic marijuana smoke exposure in the rhesus monkey. II. Effects on progressive ratio and conditioned position responding. J. Pharmacol. Exp. Ther. 260, 210–222.
- 104. Chait, L.D. and Perry, J.L. (1992) Effects of smoked marijuana on human performance: a critical review. In *Marijuana/Cannabinoids: Neurobiology and Neurophysiology*. Murphy, L. and Bartke, A., Eds. CRC Press, Boca Raton, FL. pp. 387–423.
- Miller, L.L. and Branconnier, R.J. (1983) Cannabis: effects on memory and the cholinergic limbic system. *Psychol. Bull.* 93, 441–456.
- Deadwyler, S.A., Hampson, R.E., and Childers, S.R. (1995) Functional significance of cannabinoid receptors in brain. In *Cannabinoid Receptors (Neuroscience Perspectives)*. Pertwee, R.G., Ed. Academic Press, London. pp. 206–231.
- 107. Sullivan, J.M. (2000) Cellular and molecular mechanisms underlying learning and memory impairments produced by cannabinoids. *Learn. Mem.* **7**, 132–139.
- 108. Hampson, R.E. and Deadwyler, S.A. (1998) Role of cannabinoid receptors in memory storage. *Neurobiol. Dis.* **5**, 474–482.
- 109. Hernandez-Tristan, R., Arevalo, C., Canals, S., and Leret, M.L. (2000) The effects of acute treatment with Δ^9 -THC on exploratory behaviour and memory in the rat. *J. Physiol. Biochem.* **56**, 17–24.
- 110. Hao, S., Avraham, Y., Mechoulam, R., and Berry, E.M. (2000) Low dose anandamide affects food intake, cognitive function, neurotransmitter and corticosterone levels in diet-restricted mice. *Eur. J. Pharmacol.* **392**, 147–156.
- Soderstrom, K. and Johnson, F. (2000) CB₁ cannabinoid receptor expression in brain regions associated with zebra finch song control. *Brain Res.* 857, 151–157.
- 112. Stiglick, A. and Kalant, H. (1985) Residual effects of chronic cannabis treatment on behavior in mature rats. *Psychopharmacology* (Berl). **85**, 436–439.
- 113. Herkenham, M. (1992) Cannabinoid receptor localization in brain: relationship to motor and reward systems. *Ann. N. Y. Acad. Sci.* **654**, 19–32.
- 114. Glass, M., Brotchie, J.M., and Maneuf, Y.P. (1997) Modulation of neurotransmission by cannabinoids in the basal ganglia. *Eur. J. Neurosci.* **9**, 199–203.
- 115. Sanudo-Pena, M.C., Tsou, K., and Walker, J.M. (1999) Motor actions of cannabinoids in the basal ganglia output nuclei. *Life Sci.* **65**, 703–713.
- 116. Giuffrida, A. and Piomelli, D. (2000) The endocannabinoid system: a physiological perspective on its role in psychomotor control. *Chem. Phys. Lipids.* **108**, 151–158.
- 117. Di Marzo, V., Hill, M.P., Bisogno, T., Crossman, A.R., and Brotchie, J.M. (2000) Enhanced levels of endogenous cannabinoids in the globus pallidus are associated with a reduction in movement in an animal model of Parkinson's disease. *FASEB J.* 14, 1432–1438.
- 118. Wise, R.A. (1996) Addictive drugs and brain stimulation reward. *Annu. Rev. Neurosci.* **19**, 319–340.
- 119. Gardner, E.L. and Vorel, S.R. (1998) Cannabinoid transmission and reward-related events. *Neurobiol. Dis.* **5**, 502–533.

- 120. Di Marzo, V., Goparaju, S.K., Wang, L., Liu, J., Batkai, S., Jarai, Z., Fezza, F., Miura, G.I., Palmiter, R.D., Sugiura, T., and Kunos, G. (2001) Leptin-regulated endocannabinoids are involved in maintaining food intake. *Nature* **410**, 822–825.
- 121. Trojniar, W. and Wise, R.A. (1991) Facilitory effect of Δ^9 -tetrahydrocannabinol on hypothalamically induced feeding. *Psychopharmacology (Berl.)* **103**, 172–176.
- 122. Kirkham, T.C. and Williams, C.M. (2001) Synergistic effects of opioid and cannabinoid antagonists on food intake. *Psychopharmacology (Berl.)* **153**, 267–270.
- 123. Rice, A.S. (2001) Cannabinoids and pain. Curr. Opin. Investig. Drugs 2, 399–414.
- 124. Meng, I.D., Manning, B.H., Martin, W.J., and Fields, H.L. (1998) An analgesia circuit activated by cannabinoids. *Nature* **395**, 381–383.
- 125. Fields, H.L., Heinricher, M.M., and Mason, P. (1991) Neurotransmitters in nociceptive modulatory circuits. *Annu. Rev. Neurosci.* 14, 219–245.
- 126. Guzman, M., Sanchez, C., and Galve-Roperh, I. (2001) Control of the cell survival/death decision by cannabinoids. *J. Mol. Med.* **78**, 613–625.
- 127. Lambert, D.M., Vandevoorde, S., Diependaele, G., Govaerts S.J., and Robert, A.R. (2001) Anticonvulsant activity of N-palmitoylethanolamide, a putative endocannabinoid, in mice. *Epilepsia* **42**, 321–327.
- 128. Hansen, H.H., Schmid, P.C., Bittigau, P., Lastres-Becker, I., Berrendero, F., Manzanares, J., Ikonomidou, C., Schmid, H.H., Fernandez-Ruiz, J.J., and Hansen, H.S. (2001) Anandamide, but not 2-arachidonoylglycerol, accumulates during *in vivo* neurodegeneration. *J. Neurochem.* 78, 1415–1427.
- Panikashvili, D., Simeonidou, C., Ben-Shabat, S., Hanus, L., Breuer, A., Mechoulam, R., and Shohami, E. (2001) An endogenous cannabinoid (2-AG) is neuroprotective after brain injury. *Nature* 413, 527–531.
- 130. Downer, E., Boland, B., Fogarty, M., and Campbell, V. (2001) Δ^9 -tetrahydrocannabinol induces the apoptotic pathway in cultured cortical neurones via activation of the CB₁ receptor. *Neuroreport* **12**, 3973–3978.
- Lawston, J., Borella, A., Robinson, J.K., and Whitaker-Azmitia, P.M. (2000) Changes in hippocampal morphology following chronic treatment with the synthetic cannabinoid WIN 55,212-2. *Brain Res.* 877, 407–410.
- 132. Dar, M.S. (2000) Cerebellar CB₁ receptor mediation of Δ^9 -THC-induced motor incoordination and its potentiation by ethanol and modulation by the cerebellar adenosinergic A₁ receptor in the mouse. *Brain Res.* **864**, 186–194.
- 133. Feeney, D.M. (1978) Marihuana and epilepsy: paradoxical anticonvulsant and convulsant effects. *Adv. Biosci.* **22/23**, 643–657.
- 134. Turkanis, S.A. and Karler, R. (1982) Central excitatory properties of delta 9-tetrahydrocannabinol and its metabolites in iron-induced epileptic rats. *Neuropharmacology* **21**, 7–13.
- 135. Ameri, A., Wilhelm, A., and Simmet, T. (1999) Effects of the endogeneous cannabinoid, anandamide, on neuronal activity in rat hippocampal slices. *Br. J. Pharmacol.* **126**, 1831–1839.
- 136. Ameri, A. and Simmet, T. (2000) Effects of 2-arachidonylglycerol, an endogenous cannabinoid, on neuronal activity in rat hippocampal slices. *Naunyn Schmiedebergs Arch. Pharmacol.* **361**, 265–272.
- 137. Karler, R. and Turkanis, S.A. (1980) Subacute cannabinoid treatment: anticonvulsant activity and withdrawal excitability in mice. *Br. J. Pharmacol.* **68**, 479–484
- 138. Cunha, J.M., Carlini, E.A., Pereira, A.E., Ramos, O.L., Pimentel, C., Gagliardi, R., Sanvito, W.L., Lander, N., and Mechoulam, R. (1980) Chronic administration of cannabidiol to healthy volunteers and epileptic patients. *Pharmacology* **21**, 175–185.
- 139. Filbert, M.G., Forster, J.S., Smith, C.D., and Ballough, G.P. (1999) Neuroprotective effects of HU-211 on brain damage resulting from soman-induced seizures. *Ann. N. Y. Acad. Sci.* **890**, 505–514.

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